

## Review

**Cite this article:** Gowhari Shabgah A, Qasim MT, Mojtava Mostafavi S, Olegovna Zekiy A, Ezzatifar F, Ahmadi M, Mohammadian Haftcheshmeh S, Gholizadeh Navashenaq J (2021). CXC chemokine ligand 16: a Swiss army knife chemokine in cancer. *Expert Reviews in Molecular Medicine* **23**, e4, 1–11. <https://doi.org/10.1017/erm.2021.7>

Received: 5 January 2021

Revised: 20 March 2021

Accepted: 24 March 2021

### Key words:

Biomarker; cancer; cancer treatment; chemokine; CXCL16; CXCR6


### Abbreviations:

4E-BP1, 4E-binding protein 1; ABCG2, ATP-binding cassette super-family G member 2; ADAM10, a disintegrin and metalloproteinase domain-containing protein 10; AHNAK2, Ahnak nucleoprotein 2; AKT, protein kinase B; CAF, cancer-associated fibroblast; CCL, CC motif chemokine ligand; CRC, colorectal carcinoma; CXCL, CXC motif chemokine ligand; CXCR, CXC motif chemokine receptor; DC, dendritic cell; DLBCL, diffuse large B-cell lymphoma; ECM, extracellular matrix; ELR, glutamic acid-leucine-arginine; EMT, epithelial-mesenchymal transition; ERK1, extracellular signal-regulated kinase; FAK, focal adhesion kinase; GSK-3 $\beta$ , glycogen synthase kinase-3 $\beta$ ; HCC, hepatocellular carcinoma; HIF- $\alpha$ , hypoxia-inducible factor- $\alpha$ ; HL, Hodgkin lymphoma; IFN- $\gamma$ , interferon  $\gamma$ ; IRF8, interferon regulatory factor 8; KRAS, Kirsten rat sarcoma viral oncogene homolog; lncRNA, long non-coding RNA; MAPK, mitogen-activated protein kinase; MCA, muricholic acid; MDSC, myeloid-derived suppressor cell; MICR, minimally invasive colorectal resection; miRNA, microRNA; MMP, matrix metalloproteinase; MSC, mesenchymal stem cell; mTOR, mammalian target of rapamycin; NF- $\kappa$ B, nuclear factor  $\kappa$  B; NHL, non-Hodgkin lymphoma; NKT, natural killer T cell; NPC, nasopharyngeal carcinoma; NSCLC, non-small cell lung carcinoma; p70S6K, ribosomal protein S6 kinase  $\beta$ -1; PDAC, pancreatic ductal adenocarcinoma; PKC, protein kinase C; PI3K, phosphoinositide-3 kinases; pRCC, papillary renal cell carcinoma; PTC, papillary thyroid cancer; PTEN, phosphatase and tensin homolog; Ror1, receptor tyrosine kinase-like orphan receptor-1; S100A9, s100 calcium binding protein a9; SND1, staphylococcal nuclease domain-containing protein 1; sst2, somatostatin receptor subtype 2; STAT, signal transducer and activator of transcription; TAM, tumour-associated macrophage; TGF- $\beta$ , transforming growth factor- $\beta$ ; THBS2, thrombospondin-2; TIL, tumour-infiltrating lymphocyte; TNF $\alpha$ , tumour necrosis factor- $\alpha$ ; VEGF, vascular endothelial growth factor; YXPV, tyrosine-X-proline-valine

### Author for correspondence:

Jamshid Gholizadeh Navashenaq,  
E-mail: [jamshid.gholizadeh@gmail.com](mailto:jamshid.gholizadeh@gmail.com); [jamshid.gholizadeh@mubam.ac.ir](mailto:jamshid.gholizadeh@mubam.ac.ir)

# CXC chemokine ligand 16: a Swiss army knife chemokine in cancer

Arezoo Gowhari Shabgah<sup>1,2</sup>, Maytham T. Qasim<sup>3</sup>, Seyed Mojtava Mostafavi<sup>4</sup>, Angelina Olegovna Zekiy<sup>5</sup>, Fatemeh Ezzatifar<sup>6,7</sup>, Majid Ahmadi<sup>8</sup>, Saeed Mohammadian Haftcheshmeh<sup>9</sup> and Jamshid Gholizadeh Navashenaq<sup>10</sup> 

<sup>1</sup>School of Medicine, Bam University of Medical Sciences, Bam, Iran; <sup>2</sup>Student Research Committee, Bam University of Medical Sciences, Bam, Iran; <sup>3</sup>College of Health and Medical Technology, Al-Ayen University, Thi-Qar, Iraq; <sup>4</sup>HiTech Institute of Theoretical and Computational Chemistry, New Delhi, India; <sup>5</sup>Department of Prosthetic Dentistry, Sechenov First Moscow State Medical University, Moscow, Russia Federation; <sup>6</sup>Department of Immunology, Molecular and Cell Biology Research Center, School of Medicine, Mazandaran University of Medical Sciences, Sari, Iran; <sup>7</sup>Student Research Committee, Mazandaran University of Medical Sciences, Sari, Iran; <sup>8</sup>Stem Cell Research Center, Tabriz University of Medical Sciences, Tabriz, Iran; <sup>9</sup>Department of Basic Medical Sciences, Neyshabur University of Medical Sciences, Neyshabur, Iran and <sup>10</sup>Noncommunicable Diseases Research Center, Bam University of Medical Sciences, Bam, Iran

## Abstract

Today, cancer is one of the leading causes of death worldwide. Lately, cytokine and chemokine imbalances have gained attention amongst different involved pathways in cancer development and attracted much consideration in cancer research. CXCL16, as a member of the CXC subgroup of chemokines, has been attributed to be responsible for immune cell infiltration into the tumour microenvironment. The aberrant expression of CXCL16 has been observed in various cancers. This chemokine has been shown to play a conflicting role in tumour development through inducing pro-inflammatory conditions. The infiltration of various immune and non-immune cells such as lymphocytes, cancer-associated fibroblasts and myeloid-derived suppressor cells by CXCL16 into the tumour microenvironment has complicated the tumour fate. Given this diverse role of CXCL16 in cancer, a better understanding of its function might build-up our knowledge about tumour biology. Hence, this study aimed to review the impact of CXCL16 in cancer and explored its therapeutic application. Consideration of these findings might provide opportunities to achieve novel approaches in cancer treatment and its prognosis.

## Introduction

Chemokines are a large group of cytokines with a short sequence in length. Because of their ability to induce chemotaxis in receptive cells, the name of these factors is derived from *chemo*-tactic *cytokines*. Chemokines have a mass of 8–10 KD approximately and have four cysteine residues in conserved locations, which are important to form their 3D shape. Based on these four-cysteine residues and space between two cysteines in N-terminus, chemokines are divided into four subgroups. These subgroups include C (only one N-terminal cysteine), CC (two adjacent cysteines), CXC (separated by one amino acid) and CX3C (separated by three amino acids) (Ref. 1).

The expression of chemokines affects immune cell distribution in several tissues, organs and even the tumour microenvironment. Tumour cells and immune cells can express and secrete chemokines and their receptors in the tumour microenvironment. This process leads to the migration of various immune cells to the tumour microenvironment, thus regulating immune responses (Ref. 2). The CXC chemokines have attracted much interest among chemokine subgroups because they are involved in the regulation of tumour development, invasion and metastasis. Based on the presence of ELR (glutamic acid-leucine-arginine) motif immediately before the first cysteine, CXC chemokines *per se* are divided into two groups, including non-ELR and ELR CXC chemokines (Ref. 3).

## CXCL16 and its receptor CXCR6

CXCL16, as a member of CXC chemokines, is a non-ELR and considered a fairly novel chemokine. CXCL16, having 254 amino acids, is the largest one compared with other CXC chemokines. CXCL16 is one of two chemokines that have been found in soluble (sCXCL16) and transmembrane (TM-CXCL16) forms (Ref. 4). TM-CXCL16 has also been described as a scavenger receptor for oxidised low-density lipoprotein. Hence, it is also called SR-PSOX (scavenger receptor for phosphatidylserine and oxidised lipoprotein) (Ref. 5). CXCR6 (also named CD186) is the only known receptor for CXCL16. In addition to a receptor for CXCL16, CXCR6 was initially recognised as a co-receptor on CD4<sup>+</sup> and CD8<sup>+</sup> T cells for the human immunodeficiency virus (Ref. 6).

### Genomic characterisation and tissue distribution

For the first time, Matloubian *et al.* described CXCL16 in human cells. They have located the human CXCL16 on chromosome 17p13. CXCL16 is the first discovered transmembrane CXC chemokine and has 70% similarity to mice CXCL16. In terms of organ distribution, northern blot analysis has shown that CXCL16 mRNA is expressed in lymphoid (including spleen, lymph nodes, Peyer's patches, thymus but not bone marrow) and non-lymphoid organs (Ref. 7). The chemokine CXCL16 and its receptor CXCR6 have been shown to be expressed by a vast majority of immune cells. Although CXCL16 is mainly expressed by dendritic cells (DCs), macrophages, B-cells and monocytes, it is also constitutively expressed in bronchial epithelial cells, epidermal keratinocytes and renal podocytes (Ref. 8).

It has been identified three forms of CXCL16 in human and mice up to now. The first one is the complete form of CXCL16, which is also called transmembrane CXCL16 or TM-CXCL16. Structurally, TM-CXCL16 comprised of 254 amino acids and has three domains, including a spacer or transmembrane domain (rich in serine, threonine and proline, which resembles mucin structure), a cytoplasmic domain (contains YXPV motif and able to be phosphorylated with tyrosine kinase) and a chemokine domain (resembles CC chemokines and contains six cysteines) (Ref. 7). TM-CXCL16 is cleaved and is shed from cell surface by sheddase enzymes called ADAM10 and ADAM17 (a disintegrin and metalloproteinase domain-containing protein 10 and 17), producing secretory CXCL16 (sCXCL16) (Ref. 9). The last form is CXCL16v (variant CXCL16), lacking the transmembrane and cytoplasmic domains produced by the mRNA alternative splicing process. CXCL16v form is uniquely expressed by tissue-resident DCs (Ref. 10).

### Signalling pathways

There are four proposed signalling pathways for CXCR6/CXCL16 complex in cancer until now (see Fig. 1). The ligation of CXCL16 to CXCR6 leads to intense phosphorylation of AKT. Activation of AKT initiates induction of p70S6K and 4E-BP1, as components of the mTOR signalling pathway, in prostate cancer. The activation of mTOR signalling pathway triggers the expression of CD44 and matrix metalloproteinases (MMPs), including MMP-2, 3 and 9. It has been shown that mTOR blocking by rapamycin inhibits CXCL16-dependent production of VEGF and IL-8 to induce angiogenesis, indicating activation of mTOR signalling pathway upon CXCR6 ligation (Ref. 11).

In the breast cancer model, it has been clarified that the interaction of CXCR6 and CXCL16 enhances the activity of the ERK1/2 pathway, which consecutively leads to small GTPase RhoA activation. RhoA activation results in actin cytoskeletal reorganisation and actomyosin-dependent contractility through indirect phosphorylation of cofilin and myosin light chain. This process eventually leads to the alteration of tight junction integrity and rearrangement of F-actin, which in turn mediates changes in the cytoskeletal system. Thereby, it has been concluded that high expression of CXCR6 in breast cancer cells has polarised cell morphology and enhanced cell mobility in these cells. Moreover, induction of high mobility and migratory properties in these cells has stimulated epithelial-mesenchymal transition (EMT) and metastasis (Ref. 12).

CXCL16 binding to CXCR6 also activates PI3K/AKT signalling to induce NF- $\kappa$ B translocation into the nucleus. The activation of NF- $\kappa$ B signalling also mediates the expression of IL-8, MMP2 and MMP9 to induce tumour progression (Ref. 13). Moreover, the activation of NF- $\kappa$ B upregulates cyclin A, B and D1 and downregulates P21cip/Waf1 and retinoblastoma expression. Consequently, this process promotes cell cycle transition from G<sub>1</sub> to S and induces cell proliferation in lung cancer cells

(Ref. 14). More recently, Ikeda *et al.* have introduced a new signalling pathway upon CXCL16 ligation on MKN45 cells. In this study, it has been demonstrated that CXCL16 induces STAT3 signalling (Ref. 15).

In addition to forward signalling through CXCR6, Wang *et al.* have elucidated that CXCL16 mediates reverse signalling in transmembrane form. The interaction of recombinant CXCR6 with CXCL16 on cell surface results in ERK1/2 signalling from the intracellular domain in CXCL16-expressing glioma cells. The fast migration of glioma cells has been attributed to this reverse signalling pathway *in vivo* (Ref. 16). Furthermore, TM-CXCL16 has also been shown to act as a receptor for sCXCL16. Hattermann *et al.* have indicated that in RNAi-dependent CXCR6-silenced meningioma cells, the interaction of sCXCL16 with TM-CXCL16 leads to reverse signalling from the TM-CXCL16 intracellular domain. Following this signal transduction, intracellular ERK1/2 and AKT are activated and mediated proliferation and rescued cultured meningioma cells from apoptosis (Ref. 17).

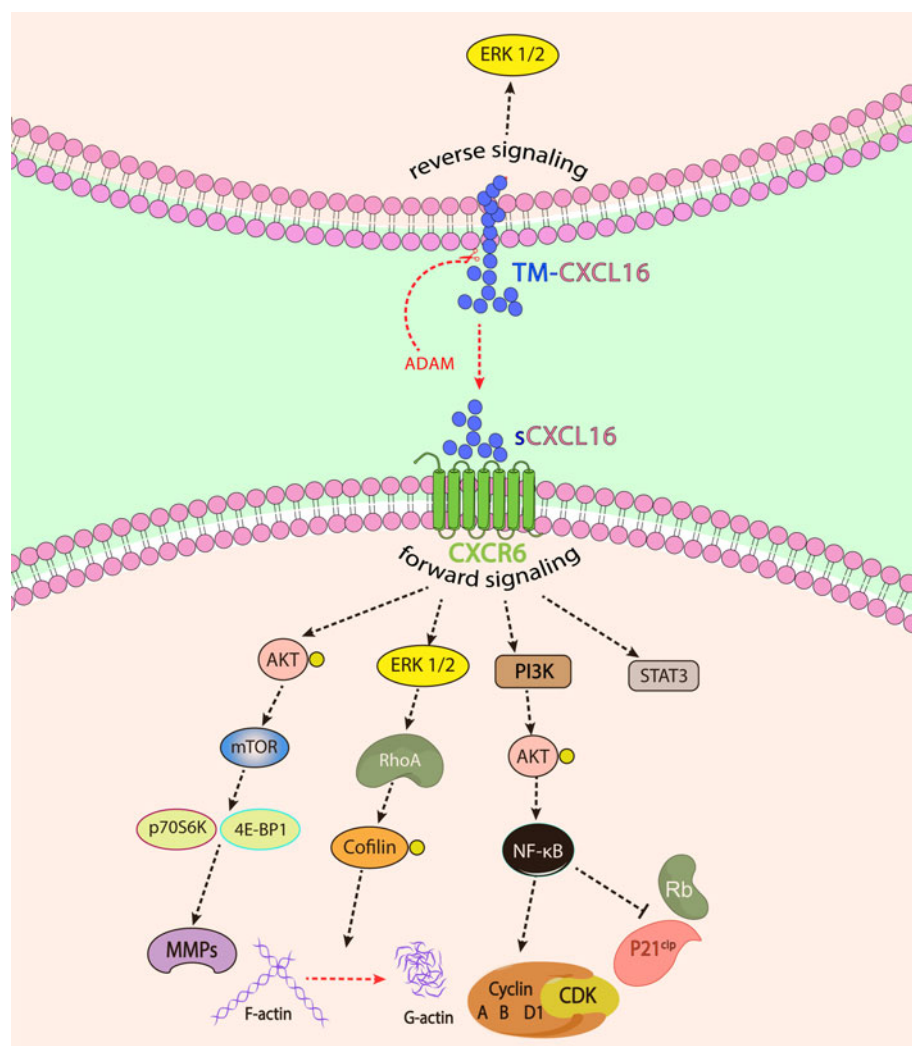
### CXCL16 expression in cancer

It has been shown that recurrent or chronic inflammatory conditions might predispose to various tumour types. Hence, lately, there has been growing attention to the relationship between chronic inflammation and tumour development (Ref. 18). Chemokines have been involved in tumour development processes, including transformation, cell proliferation, angiogenesis and lymphocyte recruitment to tumour sites (Ref. 19). Given the role in recruiting leukocytes and inflammation, it has been demonstrated that CXCL16 influences the tumour microenvironment (Ref. 8). The expression of CXCL16 and CXCR6 has been observed in numerous human cancers, including pancreatic cancer, thyroid carcinoma, breast cancer and so-forth.

Several factors influence the expression of CXCL16 in cancer (Fig. 2). Notch receptor signalling is a highly conserved pathway involved in cell proliferation, differentiation and apoptosis. Dysregulation of the Notch signalling pathway has been reported in the vast majority of human cancers. It has been shown that the expression of *Notch1* significantly upregulated in nasopharyngeal carcinoma (NPC) tissues and cell lines, and knockdown or downregulation of *Notch1* prevents NPC growth, decreases cell migration and stimulates apoptosis. Since *Notch1* knockdown inhibits CXCL16 expression in NPC, it has been concluded that CXCL16 might be a downstream target of Notch1 (Ref. 20).

The product of the mutated *KRAS* gene is considered an oncogene. Mechanistically, inflammatory mediators such as IL-6 and CXCL16 induce the expression of *KRAS* in pancreatic ductal adenocarcinoma (PDAC) cells. *KRAS* initiates PI3K/AKT/NF- $\kappa$ B signalling and subsequently induces the expression of CXCL16. The produced CXCL16 *per se* induces PI3K/AKT/NF- $\kappa$ B and establishes CXCL16-PI3K/AKT/NF- $\kappa$ B loop. In fact, mutated *KRAS* is the initiator of this loop. Further examination has shown that somatostatin receptor subtype 2 (*sst2*) can block this loop. On the contrary, a loss-of-function mutation in *sst2* has led to PDAC progression (Ref. 21).

Tumour necrosis factor-alpha (TNF- $\alpha$ ) is a multifunctional cytokine that plays pivotal roles in cell apoptosis and cell survival. Yang *et al.* have proposed a positive feedback loop between CXCL16 and TNF- $\alpha$ . In diffuse large B-cell lymphoma (DLBCL), administration of exogenous TNF- $\alpha$  induced secretion of sCXCL16 substantially through induction of *ADAM10* expression. In a positive feedback loop, sCXCL16 secretion promoted TNF- $\alpha$  production via the NF- $\kappa$ B signalling pathway in DLBCL cells (Ref. 22). In addition to TNF- $\alpha$ , it has also been shown



**Fig. 1.** The forward and reverse signalling pathways involved upon CXCL16 and CXCR6 interaction. In forward signalling, sCXCL16 or TM-CXCL16 binding to CXCR6 leads to signal transduction from CXCR6, whereas in reverse signalling, signal transduction is mediated by the intracellular domain of TM-CXCL16.

that IFN- $\gamma$  stimulated CXCL16 production from tumour cells in prostate cancer (Ref. 23).

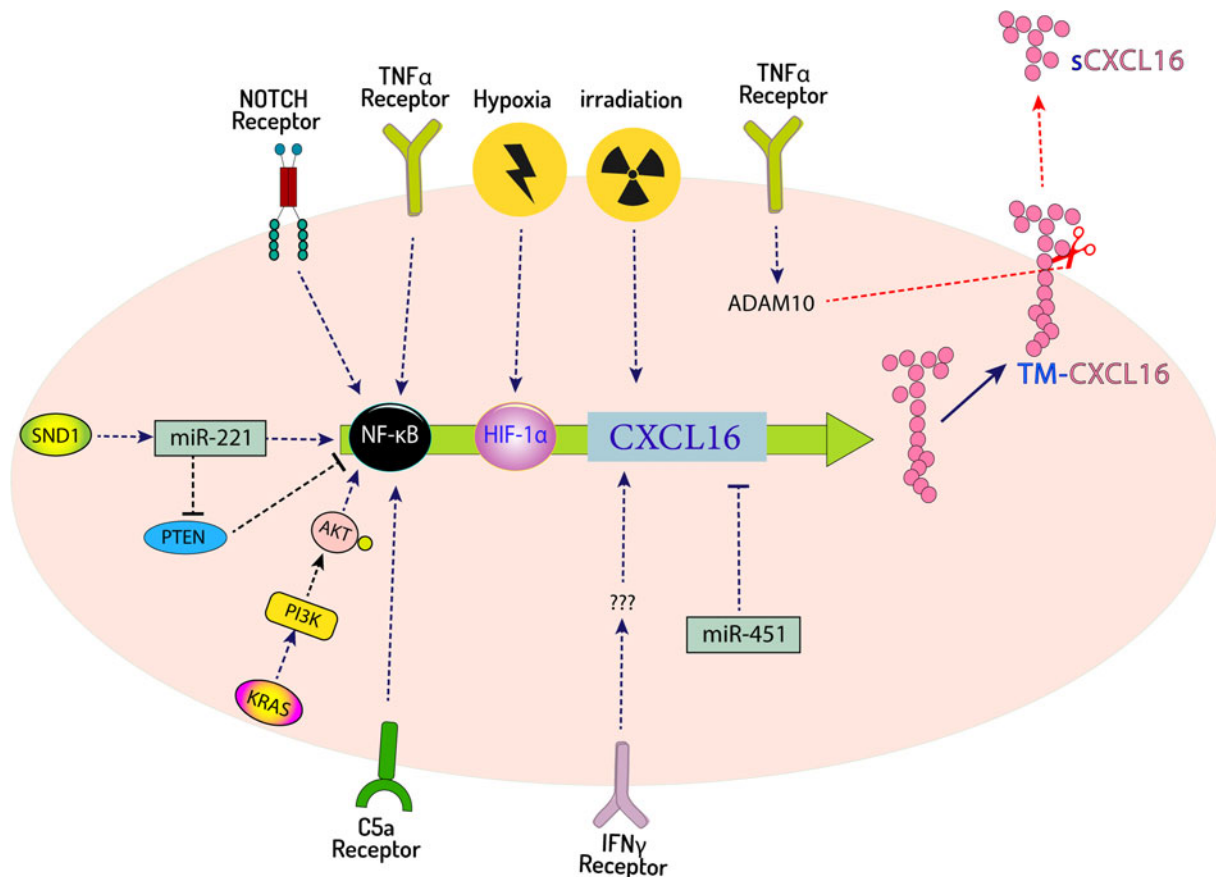
The binding of the cleaved C5 component of complement (C5a) to its receptor (C5aR1) initiates signalling responses, contributing to cancer progression. The expression of C5aR1 has been verified in various immune and non-immune cells, such as tumour cells. C5a/C5aR1 interaction impedes anti-tumour responses through multiple mechanisms, including the infiltration of myeloid-derived suppressor cells (MDSCs) into the tumour microenvironment. Besides, Ajona *et al.* have indicated another mechanism for C5a immunomodulatory properties in the tumour. They have shown that C5a/C5aR1 signalling induces the expression of CXCL16 in lung cancer, which causes inflammation in the tumour microenvironment (Ref. 24).

Non-coding RNAs, including miRNAs and lncRNAs, are other regulators of gene expression. The expression of miR-451 in osteosarcoma has been decreased. This downregulation led to tumour development and progression. In this regard, Zhang *et al.* showed that the mechanism of miR-451 function in osteosarcoma relies on CXCL16. They have found that miR-451 has targeted 3'UTR of CXCL16 gene and decreased its expression. Downregulation of miR-451 in osteosarcoma resulted in the unrestrained expression of CXCL16, cell proliferation and tumour invasion (Ref. 25).

miR-221 is another regulator of CXCL16 expression. Staphylococcal nuclease domain-containing protein 1 (SND1) is a multidisciplinary protein that is upregulated in numerous cancers, including hepatocellular carcinoma (HCC) (Ref. 26). SND1

is involved in cell signalling pathways (such as STAT, NF- $\kappa$ B and Wnt pathways), miRNA splicing process, gene silencing, cell viability and cell proliferation (Ref. 27). Santhekadur *et al.* have shown that SND1 promoted NF- $\kappa$ B activation to induce the expression of miR-221 (Ref. 28). The elevation of miR-221 was attributed to the expression of CXCL16 in HCC cells. In terms of mechanism, it has been speculated that miR-221 has targeted and inhibited phosphatase and tensin homolog (*PTEN*) and tissue inhibitor of metalloproteinases-3 (*TIMP3*). Inhibition of *PTEN* activates AKT signalling, and *TIMP3* inhibition leads to the activation of ADAM10 and ADAM17 for producing sCXCL16 (Ref. 29). In addition to miR-221 and miR-451, it has been shown that CXCL16 is the target gene of miR-873-5p. In papillary thyroid cancer (PTC), upregulation of miR-873-5p inhibited cell proliferation, migration and invasion, whereas these consequences were reversed upon treatment with CXCL16 (Ref. 30).

Hypoxia, radiation and gut microbiome are other CXCL16 regulators. As is well-known, a decreased oxygen level, called hypoxia, is a distinctive characteristic of most tumours. It has been shown that CXCL16 expression is upregulated in the hypoxic condition in the tumour microenvironment. Mechanistically, upregulation of transcription factor hypoxia-inducible factor- $\alpha$  (*HIF1 $\alpha$*  gene) might be involved in the overexpression of CXCL16 in triple-negative breast cancer (TNBC) (Ref. 31). In addition, it has been shown that irradiation of breast cancer cells upregulates CXCL16 production in these cells dose- and time-dependently (Ref. 32). Furthermore, studies have revealed that irradiation of



**Fig. 2.** The effect of various factors on the dysregulation of CXCL16 in cancer.

cells also influenced gene expression. In this regard, two independent studies showed that irradiation of breast cancer cells increased the expression of *CXCL16* gene in these cells, resulting in the recruitment of CXCR6<sup>+</sup> NK and T cells into tumour microenvironment (Refs 32, 33).

It has also been shown that the gut microbiome, through regulating the bile acid ratio, plays a vital role in *CXCL16* mRNA expression. The gut microbiome mediates the conversion of primary bile acid (such as  $\beta$ -muricholic acid ( $\beta$ -MCA)) into secondary bile acid (such as  $\omega$ -muricholic acid ( $\omega$ -MCA)). The elevation of  $\omega$ -MCA downregulated *CXCL16* expression, and  $\beta$ -MCA increased the expression of *CXCL16* in liver sinusoidal endothelial cells (Ref. 34).

#### The effect of CXCL16 on tumour development and metastasis

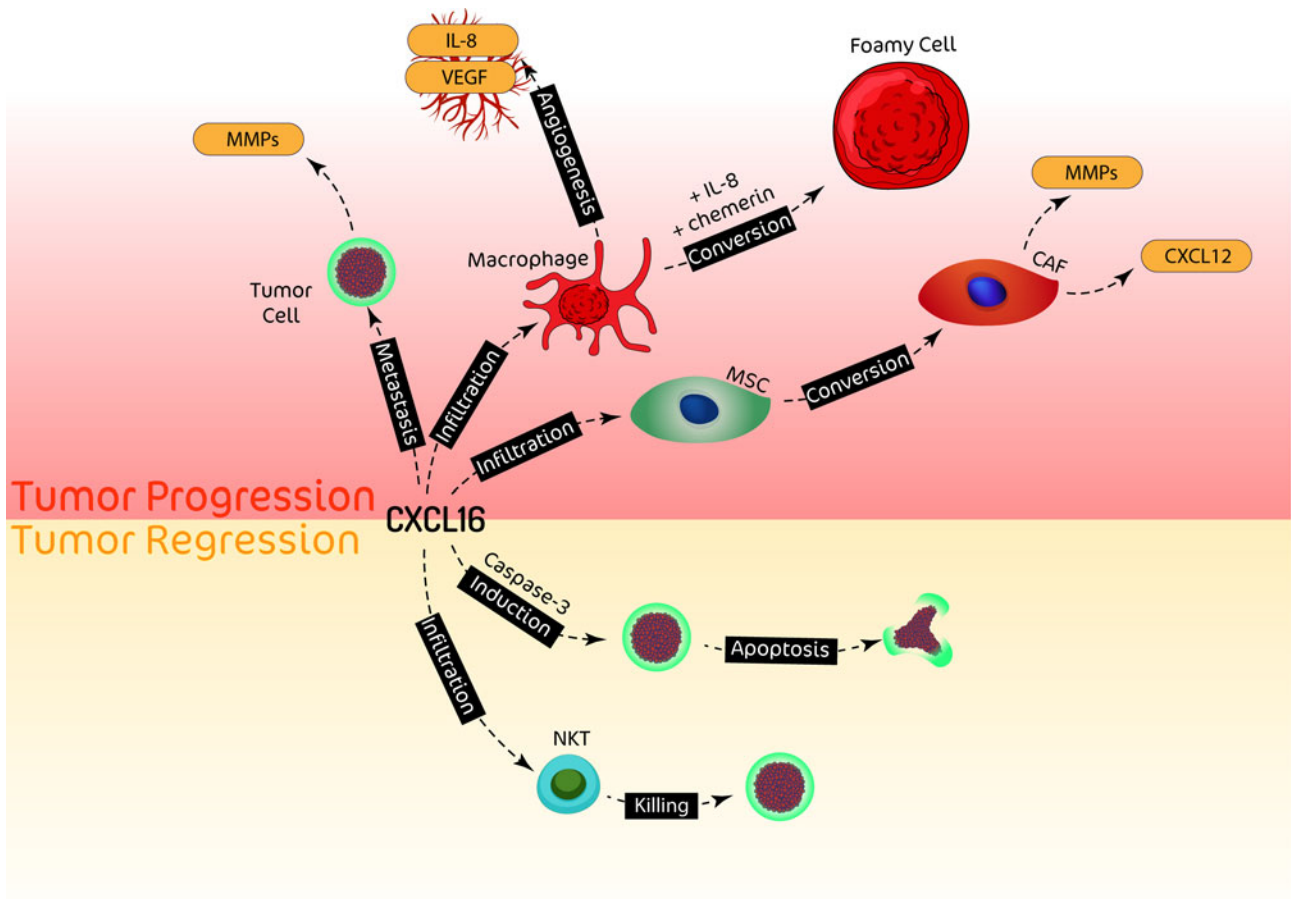
Chemokines and chemokine receptors have exerted a conflicting role in tumour development (Fig. 3). They mediate tumour progression directly via impacting survival, transformation, proliferation and metastasis of tumour cells or indirectly via enhancing leukocyte infiltration or angiogenesis (Ref. 35).

As mentioned before, CXCL16 is a potent cell chemotactic factor. In a study by Jung *et al.*, it has been cleared that CXCL16 recruited CXCR6-expressing mesenchymal stem cells (MSCs) into the prostate cancer microenvironment. The signalling of CXCR6 in recruited MSCs leads to the conversion of MSCs into cancer-associated fibroblasts (CAFs), which these cells release stromal-derived factor-1 (also known as CXCL12) in tumour nest. The interaction of CXCL12 with its receptor (CXCR4) on tumour cells induces EMT and metastasis (Ref. 36). Foamy cells are other CXCL16-dependent recruited cells into the tumour microenvironment. These cells are a type of tumour-promoting M2

macrophages that generally are considered lipid-laden macrophages. In papillary renal cell carcinoma (pRCC), the presence of these cells is the hallmark of tumour aggressiveness, which almost neither chem- nor radiotherapy have any therapeutic effect following surgery. It has been defined that CXCL16 accompanied by IL-8 and adipokine chemerin are involved in the formation of foamy macrophages (Ref. 37).

Receptor tyrosine kinase-like orphan receptor-1 and -2 (Ror1/Ror2) are considered a heterodimer receptor for Wnt5a that mediate non-canonical Wnt signalling during tissue repair, developmental morphogenesis, and tumour progression. Wnt5a and Ror2 are strongly overexpressed in cancer cells, and their consecutive activation leads to tumour progression and poor prognosis in PDAC and ovarian cancer. Secretion of CXCL16 from MSCs and its ligation on CXCR6-expressing MKN45 cells (cells derived from gastric cancer) has resulted in upregulation of Ror1 through activation of STAT3. This process indicates the effect of MSC-released CXCL16 to induce Wnt signalling and subsequently tumour formation and development (Ref. 15). More interestingly, the Wnt5/Ror2 signalling augments *CXCL16* expression, indicating the reciprocal feedback loop between CXCL16 and Wnt5/Ror2 signalling (Ref. 38). The consequence of this loop is the promotion of tumour cell proliferation induced by MSCs.

MMPs are extracellular matrix digesting enzymes and contribute to tumour cell invasion, migration and metastasis to the other tissue. It has been shown that treatment of ovarian cancer cell lines (SKOV-3 and OVCAR-3) with CXCL16 induces the expression of MMP2, 3, 9, 13 in cancer cells (Ref. 39). Also, CXCL16 ligation on LNCap prostate cells promotes the expression of MMP2 and MMP9, resulting in prostate cancer cell metastasis to the bone (Ref. 40). Therefore, the expression of CXCL16/CXCR6



**Fig. 3.** Conflicting role of CXCL16 in cancer. The red area depicts the tumour-supportive role of CXCL16 in cancer, while the yellow area shows how CXCL16 can have a tumour-suppressive role.

and ADAM10 endows metastatic competencies to ovarian and prostate cancer cells (Refs 39, 40). Another study discussing the role of CXCL16 in ovarian cancer migration has shown that M2 macrophages induce CXCL16 production, and subsequently, its ligation to CXCR6 initiates PI3K/AKT pathway and induces cell migration and metastasis (Ref. 41).

MDSCs and CAFs are the other important tumour-supportive and immune-suppressive cells in the tumour microenvironment. In TNBC, myeloid cells are recruited into the tumour nest in response to CXCL16. The recruited MDSCs express *S100A9* and *CD163*, which in turn these markers stimulate fibroblast conversion into CAFs. Activated CAFs *per se* produce more CXCL16 in the tumour microenvironment and recruit more immature myeloid cells. These processes facilitate tumour stroma conversion into a much more active phenotype, which activates the tumour nest to become a high aggressive microenvironment (Ref. 42). In HCC, CAF-secreted CXCL16 triggers TGF- $\beta$  receptor (TGF $\beta$ R) signalling and activates phosphorylation of downstream proteins, including TGF $\beta$ R, Smad2 and Smad3, to mediate metastasis and EMT. However, the molecular mechanism of this process remains unclear (Ref. 43).

The primary step in cancer cell migration, invasion and metastasis is cytoskeletal rearrangement. Integrins, such as  $\alpha_v\beta_3$  and Ezrin, coordinate this process by regulating cytoskeletal polymerisation-depolymerisation and cellular adhesion. Chemokine receptor signalling has been shown to be involved in the regulation of cell migration and invasion. In prostate cancer, it has been established that aggressive tumour cells strongly express CXCR6, which is correlated to higher tumour invasiveness. The

interaction of CXCR6 with CXCL16 induces FAK/PI3K/PKC signalling pathway and modifies cellular adhesion and motility. Further examination has shown that CXCR6 signalling phosphorylates Ezrin and induces  $\alpha_v\beta_3$  integrin clustering. The phosphorylation of Ezrin forms pseudopod, and integrin clustering mediates tumour cell adhesion to ECM as well as *MMP1* and *MMP13* upregulation. Thereby, CXCL16/CXCR6 complex signalling leads to prostate cancer cell migration and invasion into other tissues (Ref. 44).

#### The role of CXCL16 in resistance to treatment in cancer

One of the fundamental complications in cancer is the presence of cells with the capability of self-renewing and sustained neoplastic growth. These cells, which are considered cancer stem cells, are highly proliferative and drug-resistant. It has been shown that melanoma-derived cancer stem cells express high levels of CXCR6 and ABCG2 (a transporter for drug efflux, mediating drug resistance in tumours), which correlates with asymmetric self-renewal phenotype in these cells (Ref. 45).

Several signalling pathways are involved in developing drug-resistance, including PI3K/Akt/mTOR, NF- $\kappa$ B, Src family kinases, PTEN and ERK1/2. The mediators involved in these pathways facilitate the expression and induction of several pro- and anti-apoptotic proteins (Ref. 46). Kapur *et al.* have delineated that docetaxel treatment increases the expression and activation of CXCL16 and CXCR6 in prostate cancer cells. Moreover, docetaxel activates CXCR6/NF- $\kappa$ B signalling pathway. The activation of this pathway leads to the induction of CXCL16 cleavage by ADAM10,

which consequently increases sCXCL16 production. The ligation of CXCL16 to CXCR6 following docetaxel administration phosphorylates NF- $\kappa$ B, GSK-3 $\beta$  and MAPK (ERK1/2) and subsequently promotes pro-survival signals. Since the administration of anti-CXCR6 antibody blocks docetaxel resistance, it is concluded that resistance to docetaxel in prostate cancer relies on CXCL16/CXCR6 signalling pathway (Ref. 46).

### CXCL16 and tumour angiogenesis

An alteration in cytokine secretion pattern is a hallmark of many malignancies, including prostate cancer. The development of new blood vessels (angiogenesis) within the tumour nest is crucial for its development, progression and metastasis. It is worth mentioning the CXCL16 tumour-supporting role is partly because of its ability to mediate angiogenesis. CXCL16 is also an effective pro-angiogenic factor other than being a tumour cell growth factor and metastasis mediator. Interestingly, CXCR6 signalling is able to control the expression of pro-angiogenic factors such as interleukin IL-8 or VEGF, which participate in regulating tumour angiogenesis (Ref. 11).

MDSCs are a group of immune-suppressive and tumour-supportive cells. These cells release a set of mediators that mediate tumour progression, metastasis and angiogenesis. It has been shown that supernatants from MDSC cultured media induce migration of human umbilical vein endothelial cells, form a capillary-like tube in these cells and eventually promote angiogenesis. Detailed information has indicated that the presence of CXCL16 and CCL2 in MDSC conditioned medium is responsible for angiogenesis so that neutralisation of these mediators suppresses angiogenesis (Ref. 47).

It has also been shown that CXCL16 induces the migration of monocytes to the tumour microenvironment. In the PTC microenvironment, monocyte polarisation to M2 macrophages is mediated by CXCL16. M2 macrophages are in a direct relationship with tumour angiogenesis. In fact, CXCL16 significantly upregulates M2 macrophage-related genes, including *IL10*, *CD163* and *CD206*. Recruitment and accumulation of M2 macrophages in tumour nest have led to an aggressive phenotype of the thyroid tumour, including lymph node metastasis, extrathyroidal extension, tall-cell variant and higher TNM staging (Ref. 48). Therefore, upregulation of pro-angiogenic factors, such as *VEGFA*, in CXCL16-induced M2 macrophages mediates angiogenesis (Ref. 49).

### The effect of CXCL16 on tumour regression

The analysis of CXCL16 expression in two breast cancer cell lines, MCF-7 and MDA-MB213, revealed a protective role of CXCL16 in breast tumorigenesis and displayed valuable pieces of evidence to better understanding the multifarious function of CXCR6/CXCL16 in cancers. The evaluation of CXCL16 in MCF-7 cells, a less aggressive breast tumour cells expressing both oestrogen and progesterone receptors, showed a high level of CXCL16 expression in these cells. In contrast, measurement of this chemokine in MDA-MB231 cells, a highly aggressive cell and triple-negative in terms of oestrogen, progesterone receptor and Her2, presented a lower level of CXCL16. This negative correlation between CXCL16 expression and tumour aggressiveness revealed that CXCL16 performs a tumour-suppressive role against the invasiveness and migration of breast cancer cells. Further evaluations have clarified that higher expression of CXCL16 in MDA-MB231 associates with upregulation of caspase-3 in this cell, which mediates apoptosis and growth inhibition (Ref. 50). In addition to apoptosis induction, it is speculated that TM-CXCL16 on tumour cells binds to CXCR6-expressing stroma

and prevents the detachment of individual tumour cells from the tumour mass similar to E-cadherin function, and eventually constrains cell invasion and migration (Ref. 50).

As mentioned before, there is a regulatory feedback loop between TNF- $\alpha$  and sCXCL16. Yang *et al.* have indicated that sCXCL16 reinforces cell proliferation inhibition and promotes TNF- $\alpha$ -dependent cell death and apoptosis in DLBCL cells. The activated apoptosis pathway induced by TNF- $\alpha$  is mediated by mitochondrial apoptotic pathway and Fas-associated death domain caspase signalling, which had been inhibited by anti-CXCR6 antibody (Ref. 22). Correspondingly, CXCL16 reinforces the activation of TNF $\alpha$ -induced apoptosis in colorectal cancer (CRC). In CRC, it has been shown that CXCL16 upregulates interferon regulatory factor 8 (*IRF8*; a transcription factor that regulates Fas-mediated apoptosis) expression and sensitises tumour cells to TNF- $\alpha$ -induced apoptosis. On the other hand, CXCL16 increases the infiltration of M1 macrophages into the tumour microenvironment and subsequently induces the expression of TNF- $\alpha$  in these cells (Ref. 51). Collectively, CXCL16 production in DLBCL and CRC mediates tumour inhibition through TNF- $\alpha$ -induced apoptosis.

Liver-resident natural killer T (NKT) cells are the high-expressing CXCR6 cells. These cells move towards the place with a high concentration of CXCL16. The recognition of tumour cells and antigens by NKT cells induces IFN- $\gamma$  to stimulate cytotoxic T cells and fight against tumour cells. Therefore, the infiltration and accumulation of NKT via CXCL16/CXCR6 interaction are favourable for tumour regression (Ref. 52). Moreover, the infiltration of NKT cells in the murine model of CRC suppresses the metastasis of these cells to liver tissue (Ref. 53). In another observation, alteration in the gut microbiome, through affecting CXCL16, influences the accumulation and infiltration of CXCR6<sup>+</sup> NKT cells and subsequently suppresses liver tumour (Ref. 34). In another study, the lower levels of mast cell proteases such as tryptase, carboxypeptidase A3 and chymase have decreased CXCL16 level in the tumour microenvironment, which in turn diminishes NKT cell responses against melanoma (Ref. 54). Investigation of Hojo *et al.* also has assumed that high expression of CXCL16 in CRC cell recruits and infiltrates CD8<sup>+</sup> and CD4<sup>+</sup> tumour-infiltrating lymphocytes (TILs) into the tumour microenvironment and represses tumour growth (Ref. 55).

### CXCL16/CXCR6 expression as diagnostic and prognostic markers in cancers

Cancer prognosis is an evaluation of the prospective course and outcome of the tumour. The prognosis of cancer-suffering patients is often regarded as the chance of efficacious cancer treatment and patient recovery. Many factors can impact the prognosis of cancer. The location and type, stage and grade of cancer are the most important cancer prognosis factors. Also, genetic properties and the biological factors expressed in the cancer cells are the other factors that affect cancer prognosis. Hence, defining a non-invasive biomarker for cancer diagnosis and prognosis is essential. There are many prognostic biomarkers identified in cancer. In the meantime, several studies have indicated that high expression of CXCL16 in cancer could be applied as a prognostic biomarker. However, the impact of CXCR6 and CXCL16 expression on patients survival differs noticeably among different malignancies. All studies investigating the effect of CXCL16 in cancer and prognosis have been summarised in Table 1.

There are studies indicating a negative correlation between high expression of CXCL16/CXCR6 and good prognosis in cancer. Since normal lung tissue expresses high endogenous CXCL16, it is suggested that dysregulation of CXCL16 is linked to cancer development (Ref. 82). Mir *et al.* have indicated that

**Table 1.** Altered expression of CXCL16 in various cancers

Cancer type	CXCL16 expression pattern	Role in cancer	Cancer prognosis with its expression	References
Brain cancer	Upregulated	Tumour-supportive	Poor	(Refs 16, 17, 56–60)
Breast cancer	Downregulated/ upregulated	Tumour-suppressive/ tumour-supportive	Good/poor	(Refs 12, 31, 32, 47, 50, 61)
Cervical cancer	Upregulated	Tumour-supportive	Poor	(Ref. 62)
Gastric cancer	Upregulated	Tumour-supportive	Poor	(Refs 15, 38, 63, 64)
Colorectal cancer	Downregulated/ upregulated	Tumour-suppressive/ tumour-supportive	Good/poor	(Refs 55, 65–67)
Thyroid cancer	Upregulated	Tumour-supportive	Poor	(Refs 48, 49)
Nasopharyngeal cancer	Upregulated	Tumour-supportive	Poor	(Refs 20, 68)
Lymphoma	Downregulated	Tumour-suppressive	Good	(Refs 22, 69)
Prostate cancer	Upregulated	Tumour-supportive	Poor	(Refs 11, 21, 23, 36, 40, 44, 46, 70–73)
Ovarian cancer	Upregulated	Tumour-supportive	Poor	(Refs 39, 41, 74, 75)
Liver cancer	Downregulated/ upregulated	Tumour-suppressive/ tumour-supportive	Good/poor	(Refs 17, 29, 34, 51, 53, 76–78)
Lung cancer	Downregulated/ upregulated	Tumour-suppressive/ tumour-supportive	Good/poor	(Refs 8, 14, 24, 79, 80)
Melanoma	Downregulated	Tumour-suppressive	Good	(Ref. 54)
Renal cancer	Upregulated/ downregulated	Tumour-supportive/ tumour-suppressive	Poor/good	(Refs 37, 81)

higher expression of CXCL16 supports lung cancer cell proliferation and metastatic process in NSCLC via enhancement of MMPs and ADAM10 expression. They also have shown that in two NSCLC subtypes, adenocarcinoma and squamous cell carcinoma, differential expression of CXCL16/CXCR6 could be correlated to prognostic differences (Ref. 79).

To distinguish PTC, an invasive type of thyroid cancer, from benign adenoma and normal thyroid tissue, CXCL16 has been proposed as a reliable marker. Immunohistochemistry on normal tissue, benign adenoma and PTC has shown that CXCL16 is highly expressed in PTC. Also, higher mRNA transcription of CXCL16 in PTC correlated to poor prognosis (Ref. 49). CXCL16 recruits M2 macrophages to the tumour microenvironment, which in turn exacerbates tumour outcome. Using CXCL16 in combination with the expression of *AHNAK2* and *THBS2* genes has also been proposed a prognostic biomarker panel for evaluating recurrence-free survival in PTC (Ref. 49).

In a large multicentre cohort study with 535 prostate cancer-suffering patients, it has been shown that the expression of CXCL16 and CXCR6 independently could be considered as predictors for a worse clinical outcome. High expression of CXCR6 (hazard ratio 2.29; 95% confidence interval 1.10–4.82;  $P$ -value = 0.028) and CXCL16 (hazard ratio 2.52; 95% confidence interval 1.12–5.68;  $P$ -value = 0.026) has been established as independent predictors for clinical failure. Co-expression of CXCR6 and CXCL16 (hazard ratio 5.1; 95% confidence interval 1–15.9;  $P$ -value = 0.05) was highly correlated to negative prognostic factors, such as Gleason score  $\geq 7$ , Gleason grade 4 + 3, positive surgical margins and vascular infiltration (Ref. 70).

Minimally invasive colorectal resection (MICR) is a common method to remove the diseased portion of the digestive tract in CRC. Since colon resection increases the risk of tumour establishment and metastasis after surgery, it is necessary to determine a suitable biomarker for prolonged follow-up of these patients regarding inflammation and tumour development after surgery. It has been shown that CXCL16, as a pro-angiogenic factor,

remains at a high level up to a month in the plasma. Therefore, elevated plasma levels of CXCL16 may encourage tumour angiogenesis in the first month after MICR and could be a useful marker for screening tumour recurrence after resection (Ref. 65). Also, Shibata *et al.* have identified that serum CXCL16 was a potential biomarker for the efficacy of therapy, including bevacizumab (i.e., anti-VEGF) in the treatment of advanced NSCLC. In this regard, pre- and post-chemotherapy assessment of serum CXCL16 levels showed that longer overall survival was observed in patients who had a significant decrease in serum CXCL16 levels (Ref. 83). Moreover, Farrow *et al.* reported that decreased expression of CXCL16 was associated with improved recurrence-free survival among the patients with resectable melanoma (Ref. 84).

Although overexpression of CXCR6 and CXCL16 was observed in many cancers, and their increased expression has been related to a more aggressive phenotype and advanced tumour stage, some investigations have demonstrated that their increased expression is independently associated with good survival. For example, Hojo *et al.* have found that high levels of CXCL16 expression in CRC were a favourable prognostic factor, and it was associated with increased TIL levels (Ref. 55). Similarly, Xing *et al.* have shown that nuclear CXCL16 expression in gastric cancer is associated with improved survival and decreased cancer aggressiveness. This study has also conducted that patients with mixed-type carcinoma or with lymphatic invasion carcinoma showed lower CXCL16 concentrations than those with diffuse-type carcinoma or without lymphatic invasion. The serum concentration and expression of CXCL16 could specify the prognosis and aggressiveness of gastric carcinomas (Ref. 63).

The study of Gutwein *et al.* showed that the low expression of CXCL16 and CXCR6 in renal cell carcinoma is linked to decreased overall survival and proposed that the evaluation of CXCL16 in RCC could be used as an independent prognostic marker to determine better patient survival (Ref. 81). Moreover, Liu *et al.* have shown that the expression of CXCL16 was relatively

lower in non-Hodgkin lymphoma (NHL; a lymphoma with bad prognosis) than in Hodgkin lymphoma specimens (HL; a lymphoma with a fairly good prognosis), reflecting the different micro-environments of the two types of lymphoma and indicating that CXCL16 is a good prognostic biomarker in this type of cancer. Further examinations on the effect of CXCL16 on the fate of these two types of lymphoma have shown that CXCL16 induces proliferation of CD4<sup>+</sup> T cells to fight against HL cells rather than NHL cells (Ref. 69).

### Therapeutic approaches

CXCL16 is a transmembrane and secretory protein and hypothetically druggable so that neutralizing antibodies, aptamers and small inhibitors can be established to modulate its function. Owing to inadequate physiological data on the CXCL16 function, the potential adverse effects of CXCL16 antagonists are difficult to predict.

Escin, a natural mixture of triterpene saponins, is a Chinese herbal medicine and one of the major active compounds derived from *Aesculus hippocastanum* (horse chestnut) seed. It has been shown that Escin has anti-inflammatory, vascular protective, veinotonic and anti-cancer effects. In terms of the anti-cancer effect, Lee *et al.* have shown that Escin drastically decreased the secretory level of CXCL16 and migration of AGS cells (gastric adenocarcinoma cells). Further examination has shown that Escin inhibits FAK (a protein tyrosine kinase) and AKT phosphorylation in these cells and subsequently inhibits tumour growth (Ref. 64).

Blocking the expression of CXCL16 using shCXCL16 in PTC cell lines significantly decreased cell viability. Inoculation of CXCL16-silenced tumour cells into mice, as ectopic tumour model, showed significantly delayed tumour growths from day 10 to day 30 compared with CXCL16<sup>+</sup> tumour cells. The inhibitions of CXCL16 expression by targeting either the stromal cells or tumour cells could be a promising therapeutic strategy for an advanced and aggressive type of thyroid cancer (Ref. 49). Also, since CXCL16 is one of the important factors in forming foamy macrophages in pRCC, the blockade of CXCL16 could be promising for alleviating tumour severity when surgery and common therapeutic approaches fail (Ref. 37).

Inhibitors of signal transduction proteins gained much attention among investigations during the last decade. As mentioned above, ERK/MEK plays a pivotal role in the signalling of CXCR6 and reverse signalling from TM-CXCL16. U0126, as a MEK inhibitor, has been used to treat NPC cell lines. The results showed that the administration of U0126 inhibits the expression of CXCL16, cell proliferation and NPC cell metastasis into the bone, dose- and time-dependently. This idea has provided a novel therapeutic strategy for metastatic NPC by targeting the CXCL16/CXCR6 signalling pathway (Ref. 68).

As discussed above, KRAS mediates the expression of CXCL16 in PDAC and, on the contrary, sst2 blocks the activation of CXCL16 transcription. Using sst2 activators has been suggested as a therapeutic approach in KRAS-mutated PDAC. Octreotide, an octapeptide that acts as a new synthetic pharmacological analogue for natural somatostatin, has reversed sustained activation of KRAS/PI3K/AKT/NF- $\kappa$ B and proposed a promising approach for future consideration (Ref. 21).

Using monoclonal antibodies against tumour growth factors has attracted much focus among researchers. Since CXCL16 has been described as a tumour growth factor in several cancers, application of anti-CXCL16 antibody has been tested. In this regard, Kim *et al.* have declared that anti-CXCL16 antibodies in PTC murine model significantly reduced tumour volume (Refs 49, 80).

Since the transmembrane and secretory type of CXCL16 shows different functions in various tumours, these two variants have paradoxical performance. It has been concluded that the inhibition of the converting enzymes, including ADAM10 and ADAM17, might be promising in cancer treatment. In this regard, Gooden *et al.* have shown that the administration of ADAM10-specific inhibitor (GI254023x) or ADAM10/ADAM17 inhibitor (TAPI-2) reduced CXCL16 membrane shedding and significantly decreased migration and metastasis of ovarian cancer cells (Ref. 74). Other drugs or agents that can be mentioned for inhibiting or downregulation of CXCL16 expression in tumours is Namitecan (ST1968) (Ref. 85).

In the type of tumours in that CXCL16 has an anti-tumour function, the upregulation of this chemokine has clinical significance for cancer treatment. Radiotherapy is a widely-used and common treatment technique in cancer. In general, studies have confirmed that irradiation combined with chemotherapy plays a crucial role in tumour cell killing and is also known to be immune-supportive. Since Yoon *et al.* have shown that irradiation enhances the expression of CXCL16 and NK cell response against breast tumour cells, it is suggested that the optimal irradiation's dose and time might be promising in the treatment of those tumours that CXCL16 has tumour-suppressive activity (Ref. 32). In another study, ionizing radiation of TRAMP-C1, 67NR and 4T1 cells increased the expression of ADAM10 and ADAM17, which are responsible for the production of sCXCL16 and tumour inhibition (Ref. 61). Furthermore, Matsumura *et al.* have also shown that ionizing radiation therapy increases the activation of Th1 cells to induce cytotoxic T cells against breast tumour cells. This study also indicated that ionizing radiation elevates infiltration of CXCR6<sup>+</sup> T cells into the tumour microenvironment (Ref. 33). Overall, the investigations have emphasised the positive effect of irradiation on tumour cell inhibition mediated by the CXCR6/CXCL16 axis.

In some cancers such as RCC, it has been shown that the CXCL16 promoter is hypermethylated, and its re-expression inhibits tumour growth. Using methylation inhibitors or demethylating agents such as 5-azacytidine suppresses CXCL16 methylation and promotes tumour suppression (Ref. 86). Also, since Hojo *et al.* have emphasised the role of CXCL16 in CRC regression and TILs infiltration into tumour nest to fight against tumour cells, it can be assumed that using recombinant CXCL16 to induce T cell infiltration into tumour microenvironment might be promising in combination with other therapeutic approaches (Ref. 55).

Dormancy in cancer refers to a state in which the tumour cells cease proliferation but survive in a quiescent state and wait for suitable environmental situations to begin dividing once more. Dormant cancer cells are generally resistant to drugs because they are not proliferating, while chemotherapy is ideal for inhibiting rapid dividing cells. Hence, dormancy endows drug-resistance to tumour cells. The exit of dormancy relies on multiple mechanisms. One of the mechanisms that inhibit cell dormancy and promote cell proliferation is the ERK signalling pathway (Ref. 87). Since CXCL16 activates this signalling pathway, it is concluded that CXCL16 promotes cell proliferation. Inducing cell exit from dormancy by applying CXCL16 can be an approach in cancer therapy (Ref. 35).

### Future directions and concluding remarks

*In vitro* studies, murine models and clinical observations have clarified that inflammation can cause tumorigenesis. Among vital mediators for inflammation, chemokines and their receptors play a pivotal role in this process. Various functions have been defined for chemokines in cancer biology, including direct effects on cancer cells (e.g., cell proliferation, transformation and



survival) and indirect impacts (e.g., leukocyte infiltration, tumour protection and angiogenesis). Given the possible pro-tumorigenic role of chemokine in cancer, several studies have assumed that these cytokines, either produced by the cancer cells or cancer-associated leukocytes, irrespective of typical viewpoint, might contribute to tumour development through enhancement of leukocyte infiltration. However, some investigations believed that chemokines employ the immune system's anti-tumour arm, including NK, NKT, T cells, to fight against a tumour (Ref. 19).

During the decade, CXCL16 and its unique receptor CXCR6 have drawn attention to itself in cancer studies. It has been demonstrated that CXCL16 in cancer plays a multifarious role. Mainly, the overexpression of CXCL16 ligand and CXCR6 receptor in cancer correlates to lower survival rates and poorer prognostic profile. However, it has also been made known that CXCR/CXCL16 signalling has tumour-suppressive functions, proposing that this pathway is highly context-dependent. It seems that the cell origin of CXCL16 plays a pivotal role in determining the fate of the tumour. For instance, in breast and liver cancers in which CAFs are the origin of CXCL16, it has been shown that the secretion of CXCL16 is not favourable for a good prognosis (Refs 42, 43).

Since CXCL16 exists either in secretory or transmembrane forms, and the latter could be cleaved into the secretory form, it is hard to discriminate which form is being investigated in cancer studies. Hence, this is one of the problems in the study of CXCL16 in cancers. Another problem is that the origin of CXCL16 in the tumour microenvironment is not defined. Moreover, the difference in tumour stage investigating CXCL16 is almost not been considered. Therefore, future studies should focus on these issues. The *in vitro* and *in vivo* conditions were quite different from each other so that the model of tumour studies is affected by various mechanisms, including leukocyte infiltrating and the stromal cells. Therefore, *in vitro* studies cannot be generalised to the other forms of studies. Moreover, the different roles of transmembrane and secretory forms of CXCL16 during tumour progression have indicated that a high serum level of sCXCL16 is a signal of poor prognosis while TM-CXCL16 appears to be a signal of good prognosis. The different expression patterns of sCXCL16 or TM-CXCL16 in tumours cleared that the different forms of CXCL16 might be responsible for their conflicting roles in different types of cancers.

By distinguishing the role of CXCL16 in the tumour, it is hoped that using CXCL16 would be promising in tumour therapeutic approaches and diagnosis.

**Acknowledgements.** None.

**Financial support.** None.

**Conflict of interest.** None.

## References

1. Griffith JW, Sokol CL and Luster AD (2014) Chemokines and chemokine receptors: positioning cells for host defense and immunity. *Annual Review of Immunology* **32**, 659–702.
2. Marcuzzi E *et al.* (2018) Chemokines and chemokine receptors: orchestrating tumor metastasization. *International Journal of Molecular Sciences* **20**. doi: 10.3390/ijms20010096.
3. Keeley EC, Mehrad B and Strieter RM (2010) CXC chemokines in cancer angiogenesis and metastases. *Advances in Cancer Research* **106**, 91–111.
4. Abel S *et al.* (2004) The transmembrane CXC-chemokine ligand 16 is induced by IFN- $\gamma$  and TNF- $\alpha$  and shed by the activity of the disintegrin-like metalloproteinase ADAM10. *The Journal of Immunology* **172**, 6362–6372.
5. Shimaoka T *et al.* (2000) Molecular cloning of a novel scavenger receptor for oxidized low density lipoprotein, SR-PSOX, on macrophages. *Journal of Biological Chemistry* **275**, 40663–40666.
6. Blaak H *et al.* (2005) CCR5, GPR15, and CXCR6 are major coreceptors of human immunodeficiency virus type 2 variants isolated from individuals with and without plasma viremia. *Journal of Virology* **79**, 1686–1700.
7. Matloubian M *et al.* (2000) A transmembrane CXC chemokine is a ligand for HIV-coreceptor Bonzo. *Nature Immunology* **1**, 298–304.
8. Hald SM *et al.* (2015) Prognostic impact of CXCL16 and CXCR6 in non-small cell lung cancer: combined high CXCL16 expression in tumor stroma and cancer cells yields improved survival. *BMC Cancer* **15**, 441–441.
9. Abel S *et al.* (2004) The transmembrane CXC-chemokine ligand 16 is induced by IFN-gamma and TNF-alpha and shed by the activity of the disintegrin-like metalloproteinase ADAM10. *Journal of Immunology* **172**, 6362–6372.
10. van der Voort R *et al.* (2010) An alternatively spliced CXCL16 isoform expressed by dendritic cells is a secreted chemoattractant for CXCR6+ cells. *Journal of Leukocyte Biology* **87**, 1029–1039.
11. Wang J *et al.* (2008) CXCR6 induces prostate cancer progression by the AKT/mammalian target of rapamycin signaling pathway. *Cancer Research* **68**, 10367–10376.
12. Xiao G *et al.* (2015) CXCL16/CXCR6 chemokine signaling mediates breast cancer progression by pERK1/2-dependent mechanisms. *Oncotarget* **6**, 14165–14178.
13. Wang Y-H *et al.* (2013) Vascular endothelial cells facilitated HCC invasion and metastasis through the Akt and NF- $\kappa$ B pathways induced by paracrine cytokines. *Journal of Experimental & Clinical Cancer Research: CR* **32**, 51–51.
14. Liang K *et al.* (2018) High CXC chemokine ligand 16 (CXCL16) expression promotes proliferation and metastasis of lung cancer via regulating the NF- $\kappa$ B pathway. *Medical Science Monitor: International Medical Journal of Experimental and Clinical Research* **24**, 405–411.
15. Ikeda T *et al.* (2020) Mesenchymal stem cell-derived CXCL16 promotes progression of gastric cancer cells by STAT3-mediated expression of Ror1. *Cancer Science* **111**, 1254–1265.
16. Adamski V *et al.* (2017) The chemokine receptor CXCR6 evokes reverse signaling via the transmembrane chemokine CXCL16. *International Journal of Molecular Sciences* **18**. doi: 10.3390/ijms18071468.
17. Hattermann K *et al.* (2016) 'Inverse signaling' of the transmembrane chemokine CXCL16 contributes to proliferative and anti-apoptotic effects in cultured human meningioma cells. *Cell Communication and Signaling: CCS* **14**, 26.
18. Greten FR and Grivnenkov SI (2019) Inflammation and cancer: triggers, mechanisms, and consequences. *Immunity* **51**, 27–41.
19. Chow MT and Luster AD (2014) Chemokines in cancer. *Cancer Immunology Research* **2**, 1125–1131.
20. Guo H *et al.* (2019) Knockdown of Notch1 inhibits nasopharyngeal carcinoma cell growth and metastasis via downregulation of CCL2, CXCL16, and uPA. *Molecular Carcinogenesis* **58**, 1886–1896.
21. Chalabi-Dchar M *et al.* (2015) Loss of somatostatin receptor subtype 2 promotes growth of KRAS-induced pancreatic tumors in mice by activating PI3K signaling and overexpression of CXCL16. *Gastroenterology* **148**, 1452–1465.
22. Yang H *et al.* (2019) Soluble CXCL16 promotes TNF- $\alpha$ -induced apoptosis in DLBCL via the AMAD10-NF- $\kappa$ B regulatory feedback loop. *Cell Biology International* **43**, 863–874.
23. Darash-Yahana M *et al.* (2009) The chemokine CXCL16 and its receptor, CXCR6, as markers and promoters of inflammation-associated cancers. *PLoS ONE* **4**, e6695.
24. Ajona D *et al.* (2018) Blockade of the complement C5a/C5aR1 axis impairs lung cancer bone metastasis by CXCL16-mediated effects. *American Journal of Respiratory and Critical Care Medicine* **197**, 1164–1176.
25. Zhang F *et al.* (2015) MiR-451 inhibits cell growth and invasion by targeting CXCL16 and is associated with prognosis of osteosarcoma patients. *Tumour Biology: The Journal of the International Society for Oncodevelopmental Biology and Medicine* **36**, 2041–2048.
26. Jariwala N *et al.* (2017) Oncogenic role of SND1 in development and progression of hepatocellular carcinoma. *Cancer Research* **77**, 3306.
27. Chidambaranathan-Reghupaty S *et al.* (2018) The multifaceted oncogene SND1 in cancer: focus on hepatocellular carcinoma. *Hepatoma Research* **4**, 32.
28. Santhekadur PK *et al.* (2012) Multifunction protein staphylococcal nuclease domain containing 1 (SND1) promotes tumor angiogenesis in human hepatocellular carcinoma through novel pathway that involves nuclear factor  $\kappa$ B and miR-221\*. *Journal of Biological Chemistry* **287**, 13952–13958.
29. Santhekadur PK *et al.* (2012) Multifunction protein staphylococcal nuclease domain containing 1 (SND1) promotes tumor angiogenesis in human

- hepatocellular carcinoma through novel pathway that involves nuclear factor  $\kappa$ B and miR-221. *Journal of Biological Chemistry* **287**, 13952–13958.
30. Wang Z *et al.* (2020) miR-873-5p inhibits cell migration and invasion of papillary thyroid cancer via regulation of CXCL16. *OncoTargets and Therapy* **13**, 1037–1046.
  31. Chaturvedi P *et al.* (2014) Hypoxia-inducible factor-dependent signaling between triple-negative breast cancer cells and mesenchymal stem cells promotes macrophage recruitment. *Proceedings of the National Academy of Sciences of the USA* **111**, E2120–E2129.
  32. Yoon MS *et al.* (2016) Irradiation of breast cancer cells enhances CXCL16 ligand expression and induces the migration of natural killer cells expressing the CXCR6 receptor. *Cytotherapy* **18**, 1532–1542.
  33. Matsumura S *et al.* (2008) Radiation-induced CXCL16 release by breast cancer cells attracts effector T cells. *Journal of Immunology* **181**, 3099–3107.
  34. Ma C *et al.* (2018) Gut microbiome-mediated bile acid metabolism regulates liver cancer via NKT cells. *Science (New York, N.Y.)* **360**. doi: 10.1126/science.aan5931.
  35. Adamski V *et al.* (2020) Entry and exit of chemotherapeutically-promoted cellular dormancy in glioblastoma cells is differentially affected by the chemokines CXCL12, CXCL16, and CX3CL1. *Oncogene* **39**, 4421–4435.
  36. Jung Y *et al.* (2013) Recruitment of mesenchymal stem cells into prostate tumours promotes metastasis. *Nature Communications* **4**, 1795–1795.
  37. Krawczyk KM *et al.* (2017) Papillary renal cell carcinoma-derived chemerin, IL-8, and CXCL16 promote monocyte recruitment and differentiation into foam-cell macrophages. *Laboratory Investigation: A Journal of Technical Methods and Pathology* **97**, 1296–1305.
  38. Takiguchi G *et al.* (2016) Wnt5a-Ror2 signaling in mesenchymal stem cells promotes proliferation of gastric cancer cells by activating CXCL16-CXCR6 axis. *Cancer Science* **107**, 290–297.
  39. Mir H *et al.* (2019) Higher CXCL16 exodomain is associated with aggressive ovarian cancer and promotes the disease by CXCR6 activation and MMP modulation. *Scientific Reports* **9**, 2527.
  40. Hu W *et al.* (2008) CXCR6 is expressed in human prostate cancer in vivo and is involved in the in vitro invasion of PC3 and LNCap cells. *Cancer Science* **99**, 1362–1369.
  41. Hong L *et al.* (2018) Tumor-associated macrophages promote the metastasis of ovarian carcinoma cells by enhancing CXCL16/CXCR6 expression. *Pathology Research and Practice* **214**, 1345–1351.
  42. Allaoui R *et al.* (2016) Cancer-associated fibroblast-secreted CXCL16 attracts monocytes to promote stroma activation in triple-negative breast cancers. *Nature Communications* **7**, 13050.
  43. Liu J *et al.* (2016) Cancer-associated fibroblasts promote hepatocellular carcinoma metastasis through chemokine-activated hedgehog and TGF- $\beta$  pathways. *Cancer Letters* **379**, 49–59.
  44. Singh R *et al.* (2016) CXCR6-CXCL16 axis promotes prostate cancer by mediating cytoskeleton rearrangement via Ezrin activation and  $\alpha$ v $\beta$ 3 integrin clustering. *Oncotarget* **7**, 7343–7353.
  45. Taghizadeh R *et al.* (2010) CXCR6, a newly defined biomarker of tissue-specific stem cell asymmetric self-renewal, identifies more aggressive human melanoma cancer stem cells. *PLoS ONE* **5**, e15183.
  46. Kapur N *et al.* (2019) Prostate cancer cells hyper-activate CXCR6 signaling by cleaving CXCL16 to overcome effect of docetaxel. *Cancer Letters* **454**, 1–13.
  47. Han EC *et al.* (2014) Tumor-conditioned Gr-1(+)CD11b(+) myeloid cells induce angiogenesis through the synergistic action of CCL2 and CXCL16 in vitro. *Biochemical and Biophysical Research Communications* **443**, 1218–1225.
  48. Cho SW *et al.* (2016) CXCL16 signaling mediated macrophage effects on tumor invasion of papillary thyroid carcinoma. *Endocrine-Related Cancer* **23**, 113–124.
  49. Kim MJ *et al.* (2019) CXCL16 positively correlated with M2-macrophage infiltration, enhanced angiogenesis, and poor prognosis in thyroid cancer. *Scientific Reports* **9**, 13288.
  50. Fang Y *et al.* (2014) Chemokine CXCL16 expression suppresses migration and invasiveness and induces apoptosis in breast cancer cells. *Mediators of Inflammation* **2014**, 478641.
  51. Kee J-Y *et al.* (2014) CXCL16 suppresses liver metastasis of colorectal cancer by promoting TNF- $\alpha$ -induced apoptosis by tumor-associated macrophages. *BMC Cancer* **14**, 949–949.
  52. Cullen R *et al.* (2009) Enhanced tumor metastasis in response to blockade of the chemokine receptor CXCR6 is overcome by NKT cell activation. *Journal of Immunology* **183**, 5807–5815.
  53. Kee JY *et al.* (2013) Chemokine CXCL16 suppresses liver metastasis of colorectal cancer via augmentation of tumor-infiltrating natural killer T cells in a murine model. *Oncology Reports* **29**, 975–982.
  54. Grujic M *et al.* (2017) The combined action of mast cell chymase, tryptase and carboxypeptidase A3 protects against melanoma colonization of the lung. *Oncotarget* **8**, 25066–25079.
  55. Hojo S *et al.* (2007) High-level expression of chemokine CXCL16 by tumor cells correlates with a good prognosis and increased tumor-infiltrating lymphocytes in colorectal cancer. *Cancer Research* **67**, 4725–4731.
  56. Lepore F *et al.* (2018) CXCL16/CXCR6 axis drives microglia/macrophages phenotype in physiological conditions and plays a crucial role in glioma. *Frontiers in Immunology* **9**, 2750.
  57. Li G *et al.* (2013) The transmembrane chemokines CXCL16 and CX3CL1 and their receptors are expressed in human meningiomas. *Oncology Reports* **29**, 563–570.
  58. Held-Feindt J *et al.* (2008) Overexpression of CXCL16 and its receptor CXCR6/Bonzo promotes growth of human schwannomas. *Glia* **56**, 764–774.
  59. Airoidi I *et al.* (2008) CXCR5 may be involved in the attraction of human metastatic neuroblastoma cells to the bone marrow. *Cancer Immunology Immunotherapy* **57**, 541–548.
  60. Ludwig A *et al.* (2005) Enhanced expression and shedding of the transmembrane chemokine CXCL16 by reactive astrocytes and glioma cells. *Journal of Neurochemistry* **93**, 1293–1303.
  61. Matsumura S and Demaria S (2010) Up-regulation of the pro-inflammatory chemokine CXCL16 is a common response of tumor cells to ionizing radiation. *Radiation Research* **173**, 418–425.
  62. Huang Y *et al.* (2013) Expression of the CXCL12/CXCR4 and CXCL16/CXCR6 axes in cervical intraepithelial neoplasia and cervical cancer. *Chinese Journal of Cancer* **32**, 289–296.
  63. Xing YN *et al.* (2012) Role and clinicopathologic significance of CXC chemokine ligand 16 and chemokine (C-X-C motif) receptor 6 expression in gastric carcinomas. *Human Pathology* **43**, 2299–2307.
  64. Lee HS *et al.* (2014) Escin suppresses migration and invasion involving the alteration of CXCL16/CXCR6 axis in human gastric adenocarcinoma AGS cells. *Nutrition and Cancer* **66**, 938–945.
  65. Shantha Kumara HMC *et al.* (2018) Plasma levels of the proangiogenic protein CXCL16 remains elevated for 1 month after minimally invasive colorectal cancer resection. *World Journal of Surgical Oncology* **16**, 132.
  66. AbdelMageed M *et al.* (2019) The chemokine CXCL16 is a new biomarker for lymph node analysis of colon cancer outcome. *International Journal of Molecular Sciences* **20**. doi: 10.3390/ijms20225793.
  67. Chung S *et al.* (2015) Identification of novel biomarkers for metastatic colorectal cancer using angiogenesis-antibody array and intracellular signaling array. *PLoS ONE* **10**, e0134948.
  68. Zhu Y *et al.* (2015) MEK inhibitor diminishes nasopharyngeal carcinoma (NPC) cell growth and NPC-induced osteoclastogenesis via modulating CCL2 and CXCL16 expressions. *Tumour Biology: The Journal of the International Society for Oncodevelopmental Biology and Medicine* **36**, 8811–8818.
  69. Liu F *et al.* (2013) CXC chemokine ligand 16, inversely correlated with CD99 expression in Hodgkin Reed-Sternberg cells, is widely expressed in diverse types of lymphomas. *Oncology Reports* **30**, 783–792.
  70. Richardson E *et al.* (2015) The prognostic significance of CXCL16 and its receptor C-X-C chemokine receptor 6 in prostate cancer. *American Journal of Pathology* **185**, 2722–2730.
  71. Ha HK *et al.* (2011) Clinical significance of CXCL16/CXCR6 expression in patients with prostate cancer. *Molecular Medicine Reports* **4**, 419–424.
  72. Wente MN *et al.* (2008) Expression and potential function of the CXC chemokine CXCL16 in pancreatic ductal adenocarcinoma. *International Journal of Oncology* **33**, 297–308.
  73. Lu Y *et al.* (2008) CXCL16 functions as a novel chemotactic factor for prostate cancer cells in vitro. *Molecular Cancer Research* **6**, 546–554.
  74. Gooden MJ *et al.* (2014) Elevated serum CXCL16 is an independent predictor of poor survival in ovarian cancer and may reflect pro-metastatic ADAM protease activity. *British Journal of Cancer* **110**, 1535–1544.
  75. Guo L *et al.* (2011) Chemokine axes CXCL12/CXCR4 and CXCL16/CXCR6 correlate with lymph node metastasis in epithelial ovarian carcinoma. *Chinese Journal of Cancer* **30**, 336–343.
  76. Chew V *et al.* (2019) Immune activation underlies a sustained clinical response to Yttrium-90 radioembolisation in hepatocellular carcinoma. *Gut* **68**, 335–346.

77. **Gao Q et al.** (2012) CXCR6 upregulation contributes to a proinflammatory tumor microenvironment that drives metastasis and poor patient outcomes in hepatocellular carcinoma. *Cancer Research* **72**, 3546–3556.
78. **Matsushita K et al.** (2012) Soluble CXCL16 in preoperative serum is a novel prognostic marker and predicts recurrence of liver metastases in colorectal cancer patients. *Annals of Surgical Oncology* **19**, S518–S527.
79. **Mir H et al.** (2015) CXCR6 expression in non-small cell lung carcinoma supports metastatic process via modulating metalloproteinases. *Oncotarget* **6**, 9985–9998.
80. **Hu W et al.** (2014) CXCL16 and CXCR6 are coexpressed in human lung cancer in vivo and mediate the invasion of lung cancer cell lines in vitro. *PLoS ONE* **9**, e99056.
81. **Gutwein P et al.** (2009) Tumoural CXCL16 expression is a novel prognostic marker of longer survival times in renal cell cancer patients. *European Journal of Cancer* **45**, 478–489.
82. **Day C et al.** (2009) The chemokine CXCL16 is highly and constitutively expressed by human bronchial epithelial cells. *Experimental Lung Research* **35**, 272–283.
83. **Shibata Y et al.** (2020) The clinical significance of CXCL16 in the treatment of advanced non-small cell lung cancer. *Thoracic Cancer* **11**, 1258–1264.
84. **Farrow NE et al.** (2020) Characterization of sentinel lymph node immune signatures and implications for risk stratification for adjuvant therapy in melanoma. *Annals of Surgical Oncology*. doi: 10.1245/s10434-020-09277-w.
85. **Cassinelli G et al.** (2012) The curative efficacy of namitecan (ST1968) in preclinical models of pediatric sarcoma is associated with antiangiogenic effects. *Biochemical Pharmacology* **84**, 163–171.
86. **Morris MR et al.** (2008) Functional epigenomics approach to identify methylated candidate tumour suppressor genes in renal cell carcinoma. *British Journal of Cancer* **98**, 496–501.
87. **Aguirre-Ghiso JA et al.** (2003) ERK(MAPK) activity as a determinant of tumor growth and dormancy; regulation by p38(SAPK). *Cancer Research* **63**, 1684–1695.