

Climatic and oceanic influences on the abundance of gelatinous zooplankton in the North Sea

C.P. LYNAM^{1,2}, M.J. ATTRILL³ AND M.D. SKOGEN^{4,5}

¹Marine Institute, Rinville, Oranmore, County Galway, Republic of Ireland, ²Present address: CEFAS, Lowestoft Laboratory, Pakefield Road, Lowestoft, Suffolk NR33 0HT, UK, ³Marine Biology & Ecology Research Centre, Marine Institute, University of Plymouth, Drake Circus, Plymouth, PL4 8AA, UK, ⁴Institute of Marine Research, PO Box 1870, Nordnes, N-5817 Bergen, Norway, ⁵Bjerknes Centre for Climate Research, Allegaten 55, N-5007 Bergen, Norway

Oceanographically based mechanisms are shown to explain the spatial variation in the climatic relationship between the abundance of medusae (Aurelia aurita and Cyanea spp. of the class Scyphozoa), in the North Sea between 1971 and 1986 during June–August, and the winter (December–March) North Atlantic Oscillation Index (NAOI). A scyphomedusa population to the west of Denmark shows a strong inverse relationship between medusa abundance and fluctuations in the NAOI; the NAOI correlates strongly ($P < 0.001$) with both annual sea surface temperature (SST) at 6.5°E 56.5°N (1950–2008) and with winter precipitation on the Danish coast at Nordby (1900–2008) suggesting a direct link between the influence of climate and medusae abundance. In contrast, scyphomedusa abundance and distribution in the northern North Sea appears to be influenced by oceanic and mixed water inflow, which may overwhelm or mask any direct climatic influence on jellyfish abundance. Similarly, advection can also explain much of the interannual variability (1959–2000) in the abundance of other gelatinous zooplankton taxa (Cnidaria, Ctenophora and Siphonophora) in the northern North Sea as identified by the capture of gelatinous tissue and nematocysts (stinging cells) in Continuous Plankton Recorder samples. Jellyfish (Scyphozoa) in the southern North Sea may benefit from low temperature anomalies and the long-term effects of global warming might suppress Aurelia aurita and Cyanea spp. populations there. However, the biological response to temperature is complex and future research is required in this area.

Keywords: jellyfish, climate, North Sea, inflow, temperature, North Atlantic Oscillation Index

Submitted 16 September 2008; accepted 6 May 2009; first published online 13 July 2009

INTRODUCTION

Many recent studies have shown links between the abundance of gelatinous zooplankton (Cnidaria and Ctenophora) and climatic fluctuations in diverse marine ecosystems ranging from the temperate Chesapeake Bay (USA) to the Ligurian Sea of the Mediterranean (Figure 1) and even a tropical lake in Palau, Micronesia (Molinero *et al.*, 2005; Purcell, 2005; Martin *et al.*, 2006; Gibbons & Richardson, 2009). In the northern Atlantic and its surrounding seas, the dominant atmospheric teleconnection pattern influencing marine ecosystems is the North Atlantic Oscillation (NAO) pattern (Drinkwater *et al.*, 2003; Hurrell *et al.*, 2003; Figure 1). This pattern of variability has been linked to interannual variability in gelatinous zooplankton in the North Sea (Lynam *et al.*, 2004, 2005; Attrill *et al.*, 2007; Attrill & Edwards, 2008; Gibbons & Richardson, 2009). In this study, we address the discord in the various findings, regarding the influence of the climate on gelatinous zooplankton in the North Sea, and we explore the likely mechanisms linking climate to jellyfish abundance.

The North Atlantic Oscillation (NAO) describes an atmospheric pressure dipole with two foci: a high-pressure

system in the south, centred on the Azores, and a low-pressure system over Iceland. The dipole is most pronounced in the winter months (December–March) and the normalized difference between the two pressures is an index used to measure the strength of the NAO. When the difference is greatest, the NAO is said to be in a high phase and the index is positive. In this circumstance, the pressure field results in strong westerly winds blowing warmer air towards northern Europe during the winter months, warming and mixing the North Sea surface waters. The associated increase in precipitation will result in increased river flows and run-off of nutrients to coastal areas. In the North Sea, a spatial pattern in the correlation between temperature and the NAO Index is evident, such that a positive NAO Index coincides with warm temperatures in the southwest and cool anomalies in the north-east (Lynam *et al.*, 2005). Oceanic inflows to the northern North Sea are also known to respond to a strong winter NAO pattern (greater inflow of surface water, 0–150 m deep, during a positive NAO Index; Figure 2) and even the most northerly extent of the Gulf Stream shows a lagged response (2–3 years) to the NAO Index (Planque & Taylor, 1998; Drinkwater *et al.*, 2003). Thus, the NAO has an underlying influence on the marine environment and interactions between species.

The abundance of scyphozoan medusae of: *Aurelia aurita* Linnaeus, 1758; *Cyanea lamarckii* Péron & Lesueur, 1809; and *Cyanea capillata* Linnaeus, 1758, in the North Sea

Corresponding author:

C.P. Lynam

Email: Chris.Lynam@cefas.co.uk

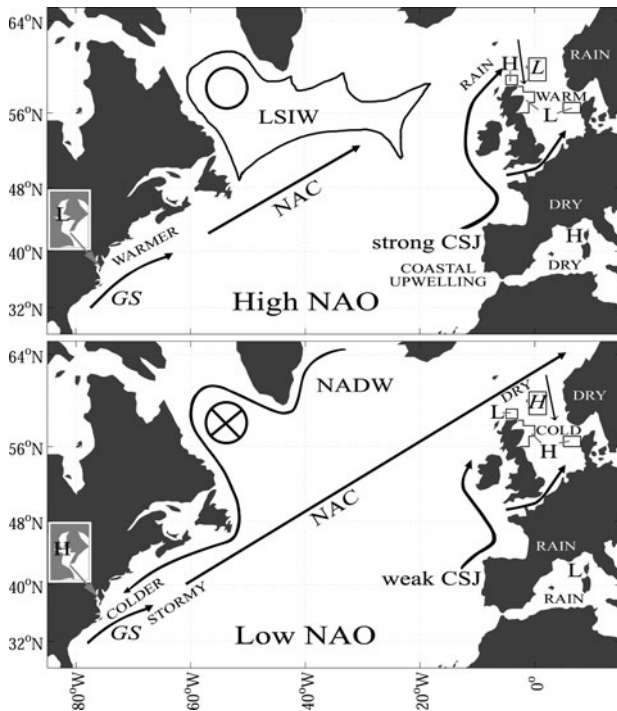


Fig. 1. Schematic diagram of North Atlantic Oscillation (NAO)-governed changes in the strength of Atlantic currents (proposed by Reid *et al.*, 1998) and the abundance of North Sea jellyfish. NAC, North Atlantic Current; GS, Gulf Stream; CSJ, Continental Shelf Jet; LSIW, Labrador Sea Intermediate Water; NADW, North Atlantic Deep Water. Note that the changes in CSJ, LSIW, and NADW may require a prolonged period of high/low NAO influence. H/L signifies relatively high/low jellyfish abundance in the defined rectangular regions, and italics indicate a two-year lag to NAO changes. Small arrows showing the shifting location of NAO-driven wintertime inflow to the northern North Sea are adapted from Planque & Taylor (1998). Chesapeake Bay relationship from Purcell & Decker (2005) and Mediterranean relationship from Molinero *et al.* (2005). Figure adapted from Lynam *et al.* (2005).

during June–August between 1971 and 1986 has been shown previously to be significantly related to the winter North Atlantic Oscillation Index (NAOI) (Lynam *et al.*, 2004). The relationship switches from strongly negative in the south-eastern North Sea, west of Denmark, through a weaker

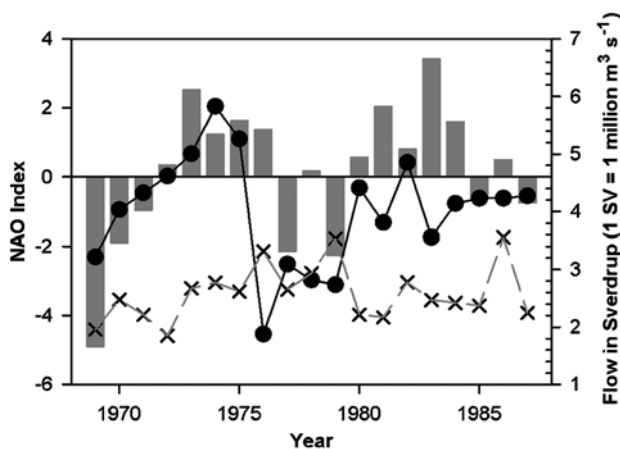


Fig. 2. Southward flow between Orkney and Norway of surface (<150 m deep) water during winter (December–February, solid black line and circles) and spring (March–May, dashed grey line and crosses) and for comparison the winter (December–March) NAOI (grey bars).

intermediate region east of Scotland to a positive relationship north of Scotland (Figure 1), mirroring the relationship between temperature and the NAOI (Lynam *et al.*, 2005). The North Sea regions are oceanographically distinct and therefore the spatial switch in the NAOI–medusa relationship could be due to differing local environmental processes and variable influence of inflow to the northern North Sea.

Attrill & Edwards (2008) found a positive correlation for the period 1962–2000 between the NAOI and the ‘Nematocyst Index’, in one of the six standard North Sea Continuous Plankton Recorder (CPR) regions (C2) once corrected for a long term linearly increasing trend. The ‘Nematocyst Index’ is an annual index of frequency of occurrence of gelatinous tissue and nematocysts (stinging cells) and is an integrative measure of occurrence of Cnidaria, Ctenophora and Siphonophora (excluding separate recordings of rigid calyphoran siphonophores) in CPR samples. Since the CPR region C2 encompasses the east of Scotland region, investigated by Lynam *et al.* (2004), Attrill *et al.* (2007) suggested that the negative relationship found previously, between the abundance of medusae (Cnidaria and Scyphozoa) and the NAOI was merely a result of the use of a shorter time-series by Lynam *et al.* (2004). However, a recent study by Gibbons & Richardson (2009) noted that the CPR does not sample scyphozoan medusae preferentially to other gelatinous organisms and suggested that the comparison of the results of Attrill *et al.* (2007) to those of Lynam *et al.* (2004) was misleading. Nevertheless, Gibbons & Richardson (2009) also found positive correlations between CPR jellyfish frequency of occurrence and the NAOI in the western and north-western North Sea (CPR areas B2 and C2, once corrected for autocorrelation); the dataset used included Cnidaria, Ctenophora and all pelagic siphonophore data and thus differed to that of Attrill *et al.* (2007) and Attrill & Edwards (2008). On a wider scale, no further links to the NAO were found in an additional 35 CPR standard areas across the North Atlantic suggesting that the NAO influence is limited to jellyfish in the North Sea. In contrast, warm sea surface temperature and an abundance of prey (CPR total zooplankton) in oceanic areas were significant predictors of jellyfish frequency of occurrence (Gibbons & Richardson, 2009).

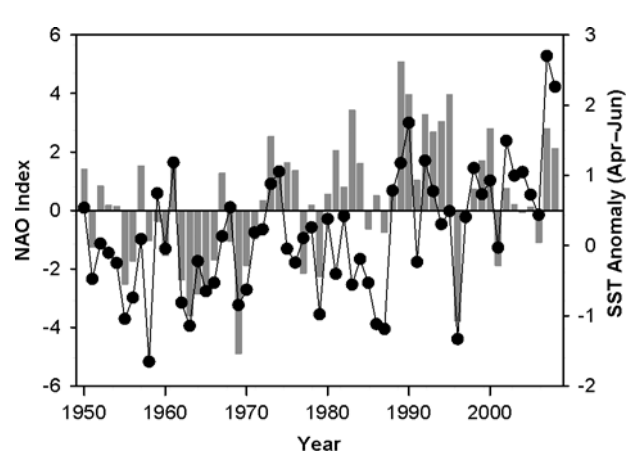


Fig. 3. Positive correlation ($R = 0.63$, $P < 0.001$, $N = 59$) between sea surface temperature (SST) to the west of northern Denmark (April–June, black line and circles) and the North Atlantic Oscillation (NAO) Index for the preceding winter (December–March, grey bars).

To determine the likely mechanisms linking the climate to jellyfish abundance in the North Sea we must explore the importance of the interaction, through wind-stress, between the NAO and the extent of seasonal oceanic water incursions into the northern North Sea. In fact, Attrill & Edwards (2008) found more and stronger correlations (at the 95% confidence level after detrending) between the Nematocyst Index and inflow (4 of 6 CPR regions: B2, C1, C2 and D1) than with the NAOI (C2 only). In this study, we attempt to determine whether oceanic inflow exerts a significant influence on the summertime distribution of scyphozoan medusae in the North Sea and, if so, whether the pattern in the scyphozoan medusae response is similar to that displayed by the gelatinous zooplankton community as sampled by the CPR. We also consider the biological responses of jellyfish to temperature change and we attempt to summarize the likely mechanisms linking regional scyphozoan jellyfish populations to interacting influences of oceanography and climate.

MATERIALS AND METHODS

The strength of the spatially varying association between the North Atlantic Oscillation and both sea surface temperature (SST) in the North Sea and precipitation was assessed through correlations between the station based NAOI (Hurrell *et al.*, 2003), spatially averaged SST anomalies determined from the HadSST2 1 degree dataset (Rayner *et al.*, 2006), and total rainfall at four coastal stations (Klein Tank *et al.*, 2002) (see Table 1 for locations and temporal periods). The effect of oceanic incursions on populations of Scyphozoa in the North Sea was explored using outputs from the Norwegian Ecological Model (NORWECOM); an hydrodynamically coupled 3-D primary production model (Skogen *et al.*, 1995; Skogen & Søiland, 1998) and the abundance of the dominant species of medusae found in each of the 4 regions used by Lynam *et al.* (2004; Figure 1). The scyphomedusae abundance data result from the ‘bycatch’ of whole animals during ICES International Young Gadoid Surveys of the North Sea; for details of the sampling methods see Hay *et al.* (1990) and Lynam *et al.* (2004). In order to reduce the number of correlations undertaken, and thus reduce the chances of committing a Type I error, the maximum regional abundance of those species suggested by Lynam *et al.* (2005) to be influenced by inflow of Atlantic

water was correlated here with modelled inflow. The biological response of medusae to changes in SST and precipitation are discussed as possible processes linking medusae to climatic fluctuations in conjunction with inflow effects.

Lynam *et al.* (2004, 2005) suggested that summertime inflow (independent of the winter NAO phase) via the Fair Isle current is likely to advect *Aurelia aurita* medusae from the North of Scotland region and aggregate them to the east of Scotland. Indices of inflow by this current during spring (March–May) and summer (June–August) were therefore studied. Medusae of *Cyanea capillata* were found regularly in the far north of the North Sea. The abundance of this species was compared to inflow by the Fair Isle current, and for total inflow to the North Sea (between the Orkneys and Norway), during spring and summer in the north of Scotland and east of Shetland regions respectively. Lynam *et al.* (2004) also suggested that the strong relationships found between the abundance of *A. aurita* and *C. lamarckii* and the NAOI in the relatively shallow region west of northern Denmark may be due to the relative isolation of this area from the effects of oceanic incursions and thus a strong influence on the population by direct interactions between the air–sea and land–sea interfaces. Considering the average current field in the North Sea and the generally coastal distribution of scyphozoan medusae, it is unlikely that medusae would be advected from either the east of Shetland region or the east of Scotland regions to the west of Denmark. However, it is possible that the current from the south and south-west may advect medusae towards Denmark, therefore the strength of the spring and summer north-eastward flow across a transect from Dogger Bank (3°E 55°N) to The Netherlands (5.5°E 53°N) was considered in this respect.

Attrill *et al.* (2007) suggested that the frequency of occurrence of gelatinous tissue and nematocysts in the CPR data overwhelmingly reflects the variability in the abundance of scyphozoans. While in contrast, Gibbons & Richardson (2009) tested the assumption and found no statistically significant correlations between the CPR jellyfish data (annual frequencies) and the species data of Lynam *et al.* (2004), individually or summed. A common weakness in these previous results is that the annual means have been calculated from the CPR data and then compared with summertime medusa abundance data from Lynam *et al.* (2004). So, correlations were made here between medusa abundance and the Nematocyst Index for the June–July period in order to

Table 1. Strength of associations between the station-based winter (December–March) North Atlantic Oscillation Index (NAOI) and both winter (December–February) rainfall, between 1900 and 2008, and spring (April–June) sea surface temperature (SST), between 1950 and 2008, for the 4 regions of interest in the North Sea.

Region	NAOI versus total rainfall in December–February				NAOI versus area averaged SST in April–June			
	Gauge location	N	R	R _{detrended}	Area co-ordinates	N	R	R _{detrended}
East of Shetland	Lerwick	62	0.43***	0.38***	59.5–60.5 N –0.5W–1.5E	59	0.29**	0.21
North of Scotland	Wick	76	0.06	0.18	58.5–59.5 N 3.5–4.5W	59	0.19	0.14
East of Scotland	Edinburgh	46	0.35**	0.25*	56.5–57.5 N 0.5–2.5W	59	0.34***	0.22*
West of Denmark	Nordby	106	0.32***	0.32***	56.5 N 5.5–7.5E	59	0.63***	0.56***

N, number of years; R, Pearson correlation coefficients; R_{detrended} Pearson correlation coefficients based on linearly detrended data; starred entries in bold indicate significant correlations at the *α = 0.1, **α = 0.05 and ***α = 0.01 significance levels.

verify, or otherwise, the suggestion that the CPR measures interannual variability in gelatinous zooplankton population abundance similarly to the scyphomedusa catches from trawl samples. The CPR device is a plankton sampling instrument, designed to be towed by merchant ships on their normal routes, which filters plankton from the water at a depth of about 10 m on a moving band of silk (270 micron mesh size) (for further details on the CPR sampling methodology regarding gelatinous zooplankton see Attrill *et al.*, 2007; Haddock, 2008; Gibbons & Richardson, 2009).

RESULTS

North Atlantic Oscillation: linkages to the marine environment

The significance of the relationships between the NAOI and both spring (April–June) temperature and winter (December–February) rainfall was tested for the four regions in the North Sea in which jellyfish data are available. Significant positive correlations were found in the average SST anomaly to the west of Denmark (5.5 to 7.5°E 56.5°N; Figure 3) and weak links to the east of Scotland and Shetland became non-significant, at the 95% confidence level, once detrended (Table 1). The pattern of significance was also shown in annual mean SSTs, albeit with lower correlation coefficients. A similar pattern was found with winter rainfall data, except that the correlation for the east of Shetland region remained significant once detrended ($P < 0.01$). There were no significant links between the NAOI and either SST or rainfall to the north of Scotland. Although winter inflows are known to be linked to the NAOI, the flows studied here in the spring and summer periods (see Table 2) for those years in which jellyfish abundance data are available (1971–1986) were not found to be correlated with the NAOI.

Possible inflow effects on jellyfish in the North Sea

The hypothesis that medusae may be aggregated to the east of Scotland region by the effects of current flow is supported

by a significant positive correlation between springtime (March–May) inflow by the Fair Isle current and the maximum abundance of *Aurelia aurita* east of Scotland (Table 2). In contrast, the summertime maximum abundance of *C. capillata* to the east of Shetland was negatively correlated with the summertime (June–August) inflow (largely independent of the winter NAOI) of oceanic waters to the North Sea, suggesting that medusae are typically transported out of the region by the inflow but the population is able to ‘bloom’ in the area if the flow weakens. Since there is no direct correlation between the NAOI and the abundance of this species to the east of Shetland, the advective influence appears to overwhelm any potential effect of fluctuations in the winter NAO on the medusa population. As expected, no significant relationships were found concerning the advection of *A. aurita* and *Cyanea lamarckii* toward the west of Denmark region; supporting the assumption that medusae in the region are not subject to strong oceanic influences (although the broadly negative correlations would suggest that weaker flow into the area coincides with higher abundance; Table 2).

Gelatinous zooplankton samples in trawls versus the Continuous Plankton Recorder (CPR)

A significant correlation was found between summer (June–July) CPR Nematocyst Index and maximum abundance of *A. aurita* medusae in the CPR C2 region ($r = 0.72$ raw and 0.66 detrended, both $P < 0.05$, $N = 10$). No other significant correlations were found with *Aurelia* or *Cyanea* spp. in any region. Since the abundance of *A. aurita* east of Scotland is also linked to spring inflow, this significant correlation with the CPR data suggests that both measures record the effects of advection on their target organisms similarly. However, the summer CPR data in general does not correlate with the medusa abundance data (for all other regions $P > 0.05$), suggesting that a relatively small proportion of the gelatinous tissue and nematocysts collected by the CPR are from scyphomedusae. The positive correlations found by Attrill *et al.* (2007) between the annual Nematocyst Index data and the NAOI were not evident for the summer (June–July) period,

Table 2. Strength of associations between modelled seasonal inflow and maximum medusa abundance for the dominant species in four regions of the North Sea.

Species	Area	Inflow transect (season)	N	R	$R_{\text{detrended}}$	Expected link	Result in agreement with expectations?
<i>Cyanea capillata</i>	East of Shetland	Ork–Nor (0–150 m) (spr)	15	−0.15	−0.32	Negative	x
		Ork–Nor (0–150 m) (sum)		−0.64*	−0.64*	Negative	✓
<i>Cyanea capillata</i>	North of Scotland	Ork–Shet (spr)	11	0.29	0.42	Negative	x
		Ork–Shet (sum)		0.38	0.44	Negative	x
<i>Aurelia aurita</i>	North of Scotland	Ork–Shet (spr)	11	−0.31	0.15	Negative	x
		Ork–Shet (sum)		−0.39	0.11	Negative	x
<i>Aurelia aurita</i>	East of Scotland	Ork–Shet (spr)	13	0.61*	0.62*	Positive	✓
		Ork–Shet (sum)		−0.15	−0.09	Positive	x
<i>Aurelia aurita</i>	West of Denmark	Neth–Dogger (spr)	10	−0.16	−0.24	No link	✓
		Neth–Dogger (sum)		−0.44	−0.46	No link	✓
<i>Cyanea lamarckii</i>	West of Denmark	Neth–Dogger (spr)	10	−0.54	−0.62	No link	✓
		Neth–Dogger (sum)		0.45	0.40	No link	✓

N, number of years; R, Pearson correlation coefficients $R_{\text{detrended}}$ Pearson correlation coefficients based on linearly detrended data; asterisk entries in bold indicate significant correlations at $\alpha = 0.05$ significance levels after sequential Bonferroni correction for multiple comparisons; Ork, Orkney; Nor, Norway; spr, spring; sum, summer; Shet, Shetland; Neth, The Netherlands; Dogger, Dogger Bank.

reflecting the dominance of the annual index by autumn–winter, non-scyphozoan samples.

Summary

The distribution of gelatinous zooplankton (Cnidaria and Ctenophora) in the northern North Sea is linked to inflows of oceanic waters to the east of Shetland and of mixed waters between the Shetland and Orkney Isles. Population variability of medusae (Scyphozoa) in the southern North Sea appears strongly coupled to fluctuations in the winter NAOI, which itself is linked strongly to variability in temperature and rainfall.

DISCUSSION

Gelatinous zooplankton samples in trawls versus the Continuous Plankton Recorder (CPR)

The CPR is a very different sampling device (with an apparatus opening of only 1.61 cm²) to the pelagic trawl (mouth opening ~14 m² with mesh size tapering down from 10 cm in the wings of the net to 1 cm in the cod-end, Lynam *et al.*, 2004) from which large medusae were collected during ICES young gadoid surveys. Thus the two datasets do not measure similar zooplankton communities (see also Haddock, 2008). In addition, the annual CPR Nematocyst Index has a large component of samples collected during autumn–winter (Attrill *et al.*, 2007; Figure 1) and thus the annual index is unlikely to represent solely the interannual variation in the abundance of large medusae of the class Scyphozoa (i.e. scyphomedusae), which are predominantly present during the summer. We should perhaps expect a difference in the correlations between the NAOI and the abundance of scyphomedusae from pelagic trawls (Lynam *et al.*, 2005, 2005) and between the NAOI and the diverse gelatinous zooplankton groups sampled by the CPR that can differ in seasonality from that of the scyphomedusae. A further difference could be exhibited in the spatial distribution of these groups, while that of scyphomedusae is generally coastal (e.g. many were caught in the east of Scotland area but relatively few to the east of the box), the gelatinous tissue and nematocysts sampled by the CPR can occur in high frequency in the central areas of the North Sea. Nevertheless, Attrill *et al.* (2007) found that there was a similarity in the annual Nematocyst Index and the abundance of *Aurelia aurita* in one (C2, east of Scotland) of the CPR regions. This similarity is due to an external force (inflow) acting in parallel on differing gelatinous organisms measured by their respective datasets. The assertion that the CPR annual Nematocyst Index records jellyfish abundance similarly to the International Young Gadoid Pelagic Trawl data used by Lynam *et al.* (2004, 2005) is not supported as concluded by Gibbons & Richardson (2009).

Probable climatic and oceanic influences

There are two related abiotic influences on gelatinous zooplankton populations in the North Sea: advection by oceanic and mixed (coastal and shelf) water incursions into the

northern North Sea and direct climatic effects (warming of surface waters and precipitation); both of which are related to fluctuations in the atmospheric pressure field as encapsulated by the NAOI. A high NAOI is characterized by strong westerly winds during the winter and a greater wintertime inflow of surface waters (0–150 m deep) into the northern North Sea and a weaker wintertime inflow of deeper oceanic waters (150–300 m) (Planque & Taylor, 1998). These findings reflect the relative strengthening of the two pathways into the North Sea by the NAO during the winter, i.e. the Fair Isle current (on shelf, dominated by mixed surface water inflow) and the inflow East of Shetland (dominated by oceanic water inflow) (Figure 1). It appears that the NAOI is linked directly to the variability in the medusa populations to the west of Denmark, whereas, in the intermediate regions east of Scotland and east of Shetland, the effects of spring and summer advection are able to overwhelm or mask the influence of the winter NAO. To the east of Shetland, jellyfish appear to be advected out of the region by oceanic inflow during the summer. There is no evidence of a similar process to the north of Scotland, which sheds doubt on whether medusae are being advected to the east of Scotland from the north. However, the Fair Isle current is composed of a mixture of coastal water originating to the west of Scotland and Atlantic water coming on to the shelf west of Shetland (Turrell *et al.*, 1996), so it is possible that medusae to the north of Scotland are supplemented by individuals from the west. Alternatively, the increase in abundance of *Aurelia aurita* to the east of Scotland may be due to a number of causes that are linked to the increased inflow of the Fair Isle current, including but not limited to: an increase in nutrient input during the spring phytoplankton bloom; an increase in the abundance of oceanic copepods or other zooplankton prey; or a direct influence of salinity on strobilation.

An interesting comparison to this study is the recent finding that five scyphozoan species, *Aurelia aurita*, *Cyanea lamarckii*, *C. capillata*, *Chrysaora hysoscella* and *Rhizostoma octopus*, exhibit distinct species-specific distributions in the Celtic and Irish Seas (Doyle *et al.*, 2007). Over three years, *Cyanea* spp. generally dominated to the north of the Irish Sea (>53°N), while *Chrysaora hysoscella* dominated to the south of the seasonal Celtic Sea front. *Cyanea capillata* is also more abundant in the northern North Sea and its distribution may be linked in part to its thermal preferences for cool waters (Hay *et al.*, 1990; Doyle *et al.*, 2007). The influence of the current field in the northern Celtic Sea and Irish Sea alone does not explain the differing spatial distributions of these species suggesting that jellyfish, particularly the species with larger medusae (e.g. *R. octopus* and *C. capillata*), have habitat preferences, which, when coupled with behavioural mechanisms, may partly explain their distributions (Graham *et al.*, 2001).

Temperature effects on jellyfish population development

The spatial pattern in the relationship between the abundance of jellyfish and the NAOI is shown similarly in the relationship between sea surface temperature and the NAOI, suggesting that temperature is a possible direct route through which the climate can impact upon jellyfish. An indirect route through nutrient inflow (stimulated by high

precipitation, increased run-off from rivers and/or inflow of oceanic water to the North Sea) and elevated prey abundance is also conceivable (Lynam *et al.*, 2004, 2005). The effect of temperature on scyphozoan jellyfish is complicated by the unusual life cycle of these organisms, which involves sexual and asexual reproduction (Vagelli, 2007). The number of medusae produced in a particular year is due largely to the strobilation of the benthic scyphistoma, the 'polyp'. However, the number of polyps present is not measurable *in situ* due to their small size and unknown locations. Although polyps are able to survive over many years the mortality of this stage is presumably highly variable. New polyps may be created through the asexual budding of existent polyps or by larvae, which are released by the sexual medusa stage, settling on a substrate. Jellyfish populations in the North Sea display great interannual variability in the abundance of medusae, but it is not known if this fluctuating nature is due to variability in the mortality of polyps or in the number of ephyrae (young medusoids) produced per polyp, or both. Temperature is known to affect the processes of strobilation and budding, but the relationship is not clear. The onset of strobilation is stress-related and has been linked to the seasonal reduction in temperature, potentially with a threshold level above which strobilation may not occur (Russell, 1970, and references therein). A thorough investigation of *Aurelia aurita* from a tropical environment found that not only did warm temperatures (25°C and 30°C relative to 20°C) accelerate strobilation but that the mortality of scyphistomae also increased (Liu *et al.*, 2009). In contrast, budding rates increased at low temperatures and decreased at high temperatures. When scyphistomae were stressed, by reducing temperatures by 10°C, they began to strobilate and more ephyrae per polyp were produced at the higher temperature (30°C). However, the potential yield of ephyrae was shown to decrease in the high temperature group; where potential yield = (number of ephyrae produced) multiplied by (1 plus the number of new buds). The potential yield was also dependent on the light regime in which polyps were kept: no difference was found between the potential yield at 25°C in shade (56 lux) or dark (0 lux) conditions or between 20°C and 25°C in shade (polyps did not strobilate in the dark at 25°C). Interestingly in light (372 lux) conditions the potential yield was greater at 20°C relative to 25°C. So, although global warming may increase the rate of ephyral release, if temperatures warm too much, such that a possible winter minimum threshold is not met, strobilation may be suppressed and budding rates of new polyps might also decrease. Our results suggest that *in situ* jellyfish populations may benefit from low temperature anomalies, which are expected in the North Sea during low (negative index) NAO years. However, this response, as exemplified by *Aurelia aurita* and *Cyanea lamarckii* to the west of Denmark, is only evident when not compounded by the advective influence of currents on medusae that may serve to aggregate (east of Scotland) or disperse (east of Shetland) jellyfish.

Concluding remarks

The similarity between the two studies (Lynam *et al.*, 2004; 2005; Attrill *et al.*, 2007; Attrill & Edwards, 2008) in relation to the climatic and oceanic influences on gelatinous zooplankton is a result of both groups (summer scyphozoan medusae abundance and the annual Nematocyst Index of gelatinous

zooplankton) responding to mixed-water incursions to the North Sea. Thus, the likely mechanism explaining the positive correlation between the NAOI and the annual Nematocyst Index by Attrill *et al.* (2007) is that of advection into the North Sea of organisms during the winter when the NAOI influence on inflow is greatest. The weakness in the relationship east of Scotland in the summer found by Lynam *et al.* (2004) between the abundance of medusae and the NAOI is likely due to the effects of currents (largely independent of the winter NAO) during the spring overpowering the winter NAO influence. The linkage between the NAO and jellyfish is displayed most strongly by the *A. aurita* and *C. lamarckii* populations west of Denmark and may be temperature-based. Although we have stressed the importance of current flow on the distribution and abundance of gelatinous zooplankton in the northern North Sea it should not be concluded that patterns in jellyfish distribution are driven solely by advection. The findings of this study demonstrate that the climate does appear to influence the abundance of medusae (Scyphozoa), but that there is a subtle interplay between abiotic factors (temperature, precipitation and currents) that impact on jellyfish populations. Thus, the direct and indirect processes linking jellyfish abundance to the climate should be further studied in areas, such as the waters to the west of northern Denmark, where the influence of current flows is minimal. This study also highlights the benefit of considering spatial scale when analysing biological data with respect to climatic and oceanographic variability.

ACKNOWLEDGEMENTS

We would like to acknowledge the efforts of M. Edwards, J. Wright and the entire CPR team at the Sir Alister Harvey Foundation for Ocean Science in collating the CPR gelatinous zooplankton data. The authors would also like to thank J. Hurrell for distributing the NAO Index data, S. Hay for supplying the scyphomedusae files and J. Kennedy for useful discussions regarding the Hadley Centre's HadSST2 dataset.

REFERENCES

- Attrill M.J., Wright J. and Edwards M. (2007) Climate-related increases in jellyfish frequency suggest a more gelatinous future for the North Sea. *Limnology and Oceanography* 52, 480–485.
- Attrill M.J. and Edwards M. (2008) Reply to Haddock S.H.D. reconsidering evidence for potential climate-related increases in jellyfish. *Limnology and Oceanography* 53, 2763–2766.
- Doyle T.K., Houghton J.D.R., Davenport J. and Hays G.C. (2007) The broad-scale distribution of five jellyfish species across a temperate coastal environment. *Hydrobiologia* 579, 29–39. DOI 10.1007/s10750-006-0362-2
- Drinkwater K.F., Belgrano A., Borja A., Conversi A., Edwards M., Greene C.H., Ottersen G., Pershing A.J. and Walker H. (2003) The response of marine ecosystems to climate variability associated with the North Atlantic Oscillation. In Hurrell J.W., Kushnir Y., Ottersen G. and Visbeck M. (eds) *The North Atlantic Oscillation: climatic significance and environmental impact*. Geophysical Monograph 134. American Geophysical Union, pp. 211–234.
- Gibbons M.J. and Richardson A.J. (2009) Patterns of pelagic cnidarian abundance in the North Atlantic. *Hydrobiologia* 616, 51–65.

- Graham W.M., Pages F. and Hamner W.M.** (2001) A physical context for gelatinous zooplankton aggregations: a review. *Hydrobiologia* 451, 199–212.
- Haddock S.** (2008) Reconsidering evidence for potential climate-related increases in jellyfish. *Limnology and Oceanography* 53, 2759–2762.
- Hay S.J., Hislop J.R.G. and Shanks A.M.** (1990) North-Sea scyphomedusae—summer distribution, estimated biomass and significance particularly for O-group gadoid fish. *Netherlands Journal of Sea Research* 25, 113–130.
- Hurrell J.W., Kushnir Y., Ottersen G. and Visbeck M.** (2003) An overview of the North Atlantic Oscillation. In Hurrell J.W., Kushnir Y., Ottersen G. and Visbeck M. (eds) *The North Atlantic Oscillation: climatic significance and environmental impact*. Geophysical Monograph 134. American Geophysical Union, pp. 1–35.
- Klein Tank A.M.G., Wijngaard J.B., Konnen G.P., Bohm R., Demaree G., Gocheva A., Mileta M., Pashiardis S., Hejkrlik L., Kern-Hansen C., Heino R., Bessemoulin P., Muller-Westermeier G., Tzanakou M., Szalai S., Palsdottir T., Fitzgerald D., Rubin S., Capaldo M., Maugeri M., Leitass A., Bukantis A., Aberfeld R., Van Engelen A.F.V., Forland E., Miletus M., Coelho F., Mares C., Razuvaev V., Nieplova E., Cegnar T., Antonio Lopez J., Dahlstrom B., Moberg A., Kirchhofer W., Ceylan A., Pachaliuk O., Alexander L.V. and Petrovic P.** (2002) Daily dataset of 20th-century surface air temperature and precipitation series for the European Climate Assessment. *International Journal of Climatology* 22, 1441–1453.
- Liu W.C., Lo W.T., Purcell J.E. and Chang H.H.** (2009) Effects of temperature and light intensity on asexual reproduction of the scyphozoan, *Aurelia aurita* (L.) in Taiwan. *Hydrobiologia* 616, 247–258.
- Lynam C.P., Hay S.J. and Brierley A.S.** (2004) Interannual variability in abundance of North Sea jellyfish and links to the North Atlantic Oscillation. *Limnology and Oceanography* 49, 637–643.
- Lynam C.P., Hay S.J. and Brierley A.S.** (2005) Jellyfish abundance and climatic variation: contrasting responses in oceanographically distinct regions of the North Sea and possible implications for fisheries. *Journal of the Marine Biological Association of the United Kingdom* 85, 435–450. DOI: 10.1017/S0025315405011380
- Martin L.E., Dawson M.N., Bell L.J. and Colin P.L.** (2006) Marine lake ecosystem dynamics illustrate ENSO variation in the tropical western Pacific. *Biology Letters* 22, 144–147. DOI:10.1098/rsbl.2005.0382
- Molinero J.C., Ibanez F., Nival P., Buecher E. and Souissi S.** (2005) North Atlantic climate and northwestern Mediterranean plankton variability. *Limnology and Oceanography* 50, 1213–1220.
- Planque B. and Taylor A.H.** (1998) Long-term changes in zooplankton and the climate of the North Atlantic. *ICES Journal of Marine Science* 55, 644–654.
- Purcell J.E.** (2005) Climate effects on jellyfish and ctenophore blooms: a review. *Journal of the Marine Biological Association of the United Kingdom* 85, 461–476. DOI:10.1017/S0025315405011409
- Purcell J.E. and Decker M.B.** (2005) Effects of climate on relative predation by scyphomedusae and ctenophores on copepods in Chesapeake Bay during 1987–2000. *Limnology and Oceanography* 50, 376–387.
- Rayner N.A., Brohan P., Parker D.E., Folland C.K., Kennedy J.J., Vanicek M., Ansell T. and Tett S.F.B.** (2006) Improved analyses of changes and uncertainties in sea surface temperature measured *in situ* since the mid-nineteenth century: the HadSST2 data set. *Journal of Climate* 19, 446–469.
- Reid P.C., Planque B. and Edwards M.** (1998) Is observed variability in the long-term results of the Continuous Plankton Recorder survey a response to climate change? *Fisheries and Oceanography* 7, 282–288.
- Russell F.S.** (1970) *Pelagic Scyphozoa with a supplement to the first volume on hydromedusae. The medusae of the British Isles II*. Cambridge: Cambridge University Press, pp. 363–468.
- Skogen M.D., Svendsen E., Berntsen J., Aksnes D. and Ulvestad K.B.** (1995) Modelling the primary production in the North Sea using a coupled 3 dimensional Physical Chemical Biological Ocean model. *Estuarine, Coastal and Shelf Science* 41, 545–565.
- Skogen M.D. and Soiland H.** (1998) *A user's guide to NORWECOM v2.0. The NORwegian ECOlogical Model system*. Technical Report Fisker og Havet 18/98, Institute of Marine Research, Pb.1870, N-5024 Bergen, 42 pp.
- Turrell W.R., Slesser G., Payne R., Adams R.D. and Gillibrand P.A.** (1996) Hydrography of the East Shetland Basin in relation to decadal North Sea variability. *ICES Journal of Marine Science* 53, 899–916.
- and
- Vagelli A.A.** (2007) New observations on the asexual reproduction of *Aurelia aurita* (Cnidaria, Scyphozoa) with comments on its life cycle and adaptive significance. *Invertebrate Zoology* 4, 111–127.

Correspondence should be addressed to:

C.P. Lynam
 CEFAS, Lowestoft Laboratory
 Pakefield Road, Lowestoft, Suffolk NR33 0HT, UK
 email: Chris.Lynam@cefas.co.uk