


# Evaluation of diverse germplasm of cowpea [*Vigna unguiculata* (L.) Walp.] against bruchid [*Callosobruchus maculatus* (Fab.)] and correlation with physical and biochemical parameters of seed

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## Abstract

The current study was undertaken to identify the sources of tolerance to bruchid in cowpea, by screening a set of germplasm accessions as a source for natural resistance. A total of 103 diverse accessions of cowpea were evaluated for resistance to *Callosobruchus maculatus* Fab. under no-choice artificial infestation conditions. Significant differences among the cowpea accessions were observed for oviposition, adult emergence, exit holes and per cent seed weight loss (PSWL) caused by the bruchid infestation. The accessions showed variation in physical seed parameters viz., colour, shape, testa texture, length, width and seed hardness. Among the seed biochemical parameters studied, per cent sugar content ranged from 0.322 (IC330950) to 1.493 (IC249137), and per cent phenol content ranged from 0.0326 (EC390261) to 1.081 (EC528423). Correlation studies indicated that PSWL had significant positive correlation ( $r=0.335$ ) with exit holes, oviposition ( $r=0.219$ ), adult emergence ( $r=0.534$ ) and seed roundness ( $r=0.219$ ). Adult emergence had a significant negative correlation with seed hardness ( $r=-0.332$ ). Correlation with biochemical parameters indicated that PSWL had a significant positive correlation ( $r=0.231$ ) with sugar content and a significant negative correlation with phenol content ( $r=-0.219$ ). None of the accessions were found to be immune to bruchid infestation. However, out of studied accessions, EC528425 and EC528387 were identified as resistant based on PSWL and moderately resistant based on adult emergence. These resistance sources of cowpea germplasm can be used as potential donors for development of bruchid tolerant/resistant cultivars.

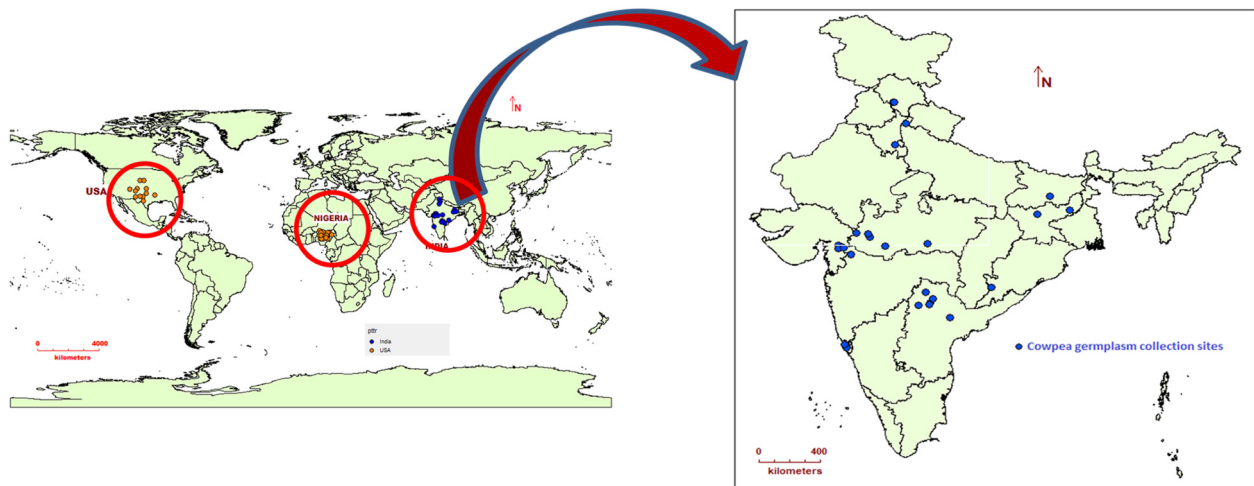
**Keywords:** bruchid, cowpea, germplasm, landraces, physical, resistance seed weight loss, *Vigna unguiculata*

## Introduction

Cowpea [*Vigna unguiculata* (L.) Walp.] is a versatile tropical legume grown throughout the world for pulse,

vegetable, fodder and green manure. In 2018, the world-wide production of dry cowpea amounted to 7.23 million tonnes, with Nigeria, Niger, Burkina Faso and Sudan as the leading cowpea-producing countries (FAOSTAT, 2020). The cowpea grain contains about 25% protein and 64% carbohydrate (Bressani, 1985). In the era of climate change, cowpea is referred to as food legume of the 21st

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**Fig. 1.** Cowpea germplasm collections sites.

century and is recognized as a smart food owing to its superior nutritional traits. But cowpea production is significantly hampered by substantial post-harvest losses. Storage losses are primarily caused by bruchids, *Callosobruchus maculatus* (Fab.) and *C. chinensis* (L.) (Coleoptera: Bruchidae). The initial damages caused by bruchids in pulses has been estimated at 30–40%, and subsequently, it can reach up to 100% (Mahendran and Mohan, 2002). Thus, irrespective of the level of initial infestation, the insect has a devastating effect on stored grains, and infested grain lots are rendered unfit for consumption. To curtail this huge loss, the most effective strategy is to develop resistant cultivars that can minimize insect infestation. In legumes, a genotype is designated as resistant if it records low per cent seed weight loss (PSWL) and as susceptible, if the seed weight loss is significantly high. Loss in seed weight is directly correlated with the feeding activity of the insect on the particular genotype, and it is in this context that the screening of germplasm for bruchid resistance has high significance.

Resistance of a host genotype against any insect pest is manifested through non-preference, antibiosis or tolerance and it is strongly correlated with the morphological, physiological and biochemical characteristics of the germplasm (Tripathi *et al.*, 2017). In legumes, physical seed parameters such as seed colour, texture, size and hardness are known to play a significant role in the resistance mechanism and functions synergistically with the biochemical factors, in rendering resistance against bruchids (Appleby and Credland, 2003). A combination of seed traits in cultivated genotypes has been reported to lower infestation levels and thereby contribute to bruchid resistance (Tripathi *et al.*, 2013). Cowpea germplasm collections are known to exhibit wide variability for seed traits (Lattanzio *et al.*, 2005), and screening of diverse germplasm accessions is

essential to identify novel sources of bruchid resistance (Carrillo-Perdomo *et al.*, 2019). However, only limited efforts have been made to screen germplasm collections for their trait variability. Consequently, there are very few breeding programmes on use of cowpea germplasm for introgression of bruchid resistance. Therefore, the current study was undertaken to identify bruchid (*C. maculatus*) resistant/tolerant sources of cowpea from a germplasm collection of 100 diverse accessions, using artificial seed infestation protocol.

## Materials and methods

### Experimental materials

A diverse set of 100 cowpea germplasm accessions comprising landraces (52), elite lines (14), registered genetic stock (1), primitive cultivar (1) and accessions with unknown biological status (32), were obtained from the National Genebank, ICAR-National Bureau of Plant Genetic Resources (NBPGR), New Delhi, India (online Supplementary Table S1). Details of biological status and source of germplasm are available in the PGR portal developed by ICAR-NBPGR (NBPGR, 2020). Collection sites of cowpea accessions evaluated in the current study are depicted in Fig. 1. Along with the 100 accessions of cowpea, three checks viz. V-240, V-578 and Pusa Sukomal were used.

### Evaluation of insect parameters

The rearing of test insect, *C. maculatus*, was carried out on cowpea seeds of local variety at controlled temperature ( $28 \pm 1^\circ\text{C}$ ) and relative humidity ( $65 \pm 5\%$ ), in a Biological

Oxygen Demand incubator, following the procedure of Tripathi *et al.* (2013). The cowpea accessions were screened for their reaction to *C. maculatus* under no-choice artificial infestation conditions using a completely randomized design (Giga, 1995). For this purpose, 20 healthy and dried seeds of each accession were weighed and placed in glass bottles covered with perforated lids, to ensure aeration. Two pairs (male and female) of freshly emerged adults from the stock cultures were released in each container, for oviposition. Each accession (20 seeds/container) was replicated five times. The insect adults inoculated in each container were removed after 72 h. Observations were recorded on the number of eggs laid, adult emergence, exit holes and loss in seed weight due to larval feeding. The number of eggs laid by the bruchid females on seeds was counted to determine the level of oviposition on each accession. Adult emergence was initiated after 25 days of infestation. Observations for emergence were recorded at a regular interval of 24 h and continued until zero emergence was recorded. The experimental seeds were weighed (X1) before releasing the insects for egg-laying and were re-weighed after the emergence of adults (X2). The loss in seed weight as a result of feeding activity of the bruchid was calculated (X1 – X2) and expressed in percentage. Other insect-associated parameters viz., mean oviposition value and number of exit holes were also calculated.

### **Evaluation of physical parameters of seed**

Physical parameters of cowpea accessions were recorded using different descriptors such as colour (Mahajan *et al.*, 2000), shape and texture (IBPGR, 1983). For assessment of texture, the seeds were examined under a stereobinocular microscope and classified into six categories, namely (i) smooth, (ii) smooth to rough, (iii) rough-fine reticulation, (iv) rough to wrinkled, (v) wrinkled-coarse folds on the testa and (vi) smooth and shiny. Seed length and width were measured using a Vernier Callipers and expressed in millimetres (mm). Seed hardness was calculated by pressure exertion method using a Texture Analyzer at the Division of Post Harvest Technology, ICAR-Indian Agricultural Research Institute, New Delhi. The pressure was exerted on the individual grain until it cracked, and the reading at the cracking point was expressed in Newton. Seed weight was recorded by weighing 100 uniform seeds in an analytical balance and was measured in gram (g).

### **Estimation of total soluble sugar and total phenol**

Ethanollic extract was prepared for all accessions of cowpea. For this, 100 mg of dried and powdered seeds in

each sample was extracted with 5 ml of 80% (v/v) ethanol by continuous shaking for 30 min, at 60°C, in the dark. This was followed by centrifugation at 5000 rpm for 15 min, and the supernatant was collected. The process was repeated twice, using the same residue, and the supernatant was pooled and evaporated to dryness. Total soluble sugar content in the extract was determined using the anthrone reagent, with glucose as the standard (Roe, 1955). For this, 100 µl of ethanolic extract was evaporated to dryness in test tubes, on a water bath. Residue was dissolved in 1.0 ml of water, and 4.0 ml of anthrone reagent was added. Absorbance was read at 660 nm and corrected against the blank sample. Total phenols in the ethanolic extract were estimated using the Folin–Ciocalteu (FC) reagent method (Slinkard and Singleton, 1977). For this, the ethanolic extract was re-dissolved in 1000 µl of water. To this, 200 µl of 1 N FC reagent and 2.0 ml of sodium carbonate (7%, w/v) were added. Furthermore, the contents were mixed and allowed to stand for 30 min at room temperature (25 ± 1°C) in the dark. The absorbance was measured at 750 nm using an ultraviolet–visible spectrophotometer (Molecular Devices, USA), using gallic acid (0–100 µg/ml) as the standard. Total phenolic content was expressed as gallic acid equivalent (GAE) per 100 g dry sample.

### **Data analysis**

Statistical analysis was performed using Statistical Analysis software, Version 9.2 (SAS, 2009). Analysis of variance was carried out using PROC GLM to determine significant differences among the cowpea accessions for physical and biochemical parameters and to find out significant differences in infestation among the cowpea accessions. Simple linear correlation analysis was performed using PROC CORR to indicate the measure of correlation and strength of the relationship between physico-biochemical parameters of seed and specific life parameters of bruchids.

### **Results**

Experimental data revealed considerable variation among genotypes, in their reaction to *C. maculatus* under no-choice artificial infestation conditions (Table 1). There were significant differences among the accessions in terms of number of eggs laid, adult emergence, exit holes and PSWL. The number of eggs laid by *C. maculatus* ranged from 52.7 (EC528387) to 437 eggs/20 seeds (IC202931). Minimum number of eggs laid per 20 seeds was recorded in EC528387 (52.7) followed by IC421893 (59.3), V-578 (72) and EC332352 (72.7) indicating that these accessions were least preferred for oviposition. Most preferred accession for egg-laying per 20 seeds was IC202931 (437) followed by IC58905 (431). The mean

**Table 1.** Evaluation of cowpea germplasm for reaction to *C. maculatus* based on adult emergence (number of adults emerged)

Category	Range of adult emergence	Number of accessions	List of accessions
Immune (0)	–	Nil	–
Resistant (1–4)	–	Nil	–
Moderately resistant (4.1–9)	6.67–9.00	4	EC528425, EC528387, EC528689, EC528410
Moderately susceptible (9.1–14)	10.33–14.00	13	EC528700, EC4506, EC528388, EC528695, EC472276, EC528429, EC528405, Pusa Sukomal (Check), V-578 (Check), EC332352, EC528457, EC528691, IC371749
Susceptible (14.1–19)	14.33–19.00	54	IC397847, IC20584 EC528392, IC198327, EC390225, IC202777, EC390223, IC201098, IC198323, IC397349, IC396744, IC397455, EC528408, IC326718, IC 330950, IC398083, IC421893, EC472293, IC433467, EC240747, IC332240, IC397942, EC517129, EC528404, EC528397, IC398065, EC390244, IC202886, IC202931, IC326793, IC396667, IC397907, IC433448, EC528407, EC528415, EC528687, EC98442, V-240 (Check), IC202790, IC249137, IC249588, EC390254, EC514422, EC528386, IC385869, IC433465, IC398142, EC517139, IC394237, IC2946, IC219529, EC390261, EC528382, EC528423
Highly susceptible (>19)	19.33–30.00	32	IC202718, IC249586, IC347189, EC390257, EC496737, EC517131, EC528381, IC243313, IC331212, EC367713, EC390216, EC501045, EC528393, IC253277, IC326042, EC394779, EC472257, EC528383, IC433510, EC472273, IC253279, IC397618, IC 398828, IC322273, EC528402, EC390293, EC528406, IC219544, IC330968, IC201077, IC402162, IC58905

number of exit holes per 20 seeds on cowpea accessions ranged from 4 (EC528425) to 20 (EC390293). The mean adult emergence of *C. maculatus* ranged from 6.67 (EC528425) to 30 (IC58905). The minimum adult emergence was recorded in EC528425 (6.67) followed by EC528387 (7.67), EC528689 (8.67) and EC528410 (9) (Table 1).

The mean PSWL varied significantly among different accessions (Table 2). It was lowest in EC528425 (4.82%) followed by EC528387 (8.48%). The highest mean PSWL of 54.48% was observed in EC528423. Accessions showing maximum seed weight loss indicated that they were highly preferred (susceptible) for feeding and accessions that recorded minimum seed weight loss were the least preferred (resistant). Cowpea accessions were categorized into resistant, moderately resistant, moderately susceptible, susceptible and highly susceptible groups based on the parameters of PSWL and adult emergence. None of the cowpea accessions were found immune to bruchid attack. However, two accessions viz., EC528425 (Fig. 2) and EC528387 were identified as resistant, based on PSWL and moderately resistant based on the adult emergence. Scatter plot showing a linear trend between two key traits viz. PSWL and adult emergence is depicted in Fig. 2.

### Seed parameters (physical and biochemical)

Cowpea accessions varied in physical seed parameters viz., colour, shape, testa texture, length, width and hardness. The accessions were grouped into greyed purple, greyed orange, brown, yellow, white and black seed colour categories. Significant variation was observed for seed length, width and hardness. Seed length ranged from 6.67 mm (EC390254) to 15.65 mm (EC501045) and breadth ranged from 4.97 mm (EC390254 and EC528408) to 10.3 mm (EC501045). Seed roundness ranged from 0.4 mm in EC528429 to 0.89 mm (EC528382). Seed hardness ranged from 18.94 Newton (EC528429) to 149.87 Newton (Pusa Sukomal). Germplasm accessions exhibited wide variation in seed shape (kidney, globose, obtuse and round) and testa texture (smooth, smooth to rough, rough with fine reticulations and rough to wrinkled). Results indicated that the largest number of accessions were kidney-shaped (83); followed by globose and round (seven accessions each); six accessions had obtuse shape. Observation on testa texture showed that maximum accessions possessed rough texture with fine reticulations (41) followed by smooth (32), smooth to rough (24) and rough to wrinkled (6). Results of the biochemical parameters of seed indicated that per cent sugar content ranged

**Table 2.** Evaluation of cowpea germplasm for reaction to *C. maculatus* based on PSWL

Category	Range of PSWL	Number of accessions	List of accessions
Immune (0%)	–	Nil	–
Resistant (1–10%)	4.819–8.48	2	EC528425, EC528387
Moderately resistant (10.01–20%)	10.952–19.951	23	EC528405, EC332352, EC528700, EC528689, EC528388, EC528408, EC528410, EC472276, EC528392, V-578 (Check), EC390223, IC371749, IC198327, IC330950, IC397349, IC331212, EC472257, EC528457, EC528429, IC398083, EC528383, EC528691, EC528381
Moderately susceptible (20.01–30%)	20.110–29.763	51	EC528695, EC528415, EC390225, EC472273, IC421893, EC517139, EC472293, IC397847, IC198323, EC517129, EC528386, IC253277, IC398065, EC528382, IC202931, EC240747, IC433467, Pusa Sukomal (Check), IC253279, IC397455, IC202790, IC326718, IC396667, IC201098, IC332240, EC528393, IC398828, IC398142, EC528404, IC347189, EC528397, IC326042, EC4506, IC397942, EC528407, IC249137, IC330968, EC367713, EC 528402, V-240 (Check), EC390216, IC249586, IC402162, EC528687, EC98442, EC528406, IC394237, IC433448, IC202777, IC396744, IC2946
Susceptible (30.01–40%)	30.028–39.487	22	EC390257, IC433465, IC219529 EC390244, IC20584, EC390293, IC397618, EC390261, IC397907, EC501045, IC385869, IC322273, IC326793, EC496737, IC249588, IC219544, EC517131, EC514422, IC433510, EC390254, IC58905, IC243313
Highly susceptible (>40.01%)	40.348–54.4792	5	IC202718, IC202886, IC201077, EC394779, EC528423

from 0.322 (IC330950) to 1.493 (IC249137) and per cent phenol content ranged from 0.0326 (EC390261) to 1.081 (EC528423).

### Correlation studies

Correlation between PSWL with different growth parameters of bruchids and physico-biochemical parameters of cowpea accessions (Table 3) indicated that PSWL had significant positive correlation with exit holes ( $r=0.335$ ), oviposition ( $r=0.219$ ), adult emergence ( $r=0.534$ ) and seed roundness ( $r=0.219$ ). Correlation with biochemical parameters indicated that PSWL had a significant positive correlation ( $r=0.231$ ) with sugar content and significant negative correlation with phenol content ( $r=-0.219$ ). Adult emergence had significant positive correlation ( $r=0.135$ ) with sugar content while it was negatively correlated with phenol content ( $r=-0.021$ ). Adult emergence was negatively correlated with seed hardness ( $r=-0.332$ ).

### Discussion

Genebanks are vital for conserving germplasm and thereby, facilitating plant breeding programmes (Tanksley and

McCouch, 1997; Engels, 2002; FAO, 2010; Khoury et al., 2010; Diez et al., 2018; Mascher et al., 2019). However, one of the major obstacles in the use of Genebank accessions is the lack of adequate characterization and evaluation data associated with the conserved germplasm (Marshall, 1989; Hodgkin et al., 2003; Kell et al., 2017; Kehel et al., 2020). Hence, it is essential to evaluate accessions for the potential traits (de Carvalho et al., 2013; Anglin et al., 2018; Byrne et al., 2018). In legumes (including cowpea), information on bruchid resistance is crucial for the utilization of germplasm in crop improvement and screening of diverse germplasm is an important step to identify sources of bruchid resistance (Somta et al., 2008; Upadhyaya et al., 2011; Carrillo-Perdomo et al., 2019). The current study was undertaken to screen a diverse set of cowpea germplasm comprising landraces, elite lines, registered germplasm and primitive cultivars for *C. maculatus*. The accessions used in this study were diverse in terms of their geographical source of origin, as well as morphological parameters. Landraces represented 52% of the experimental material. Villa et al. (2005) defined a landrace as 'a dynamic population(s) of a cultivated plant that has historical origin, distinct identity and lacks formal crop improvement, as well as often being genetically diverse, locally adapted and associated with traditional farming systems'.

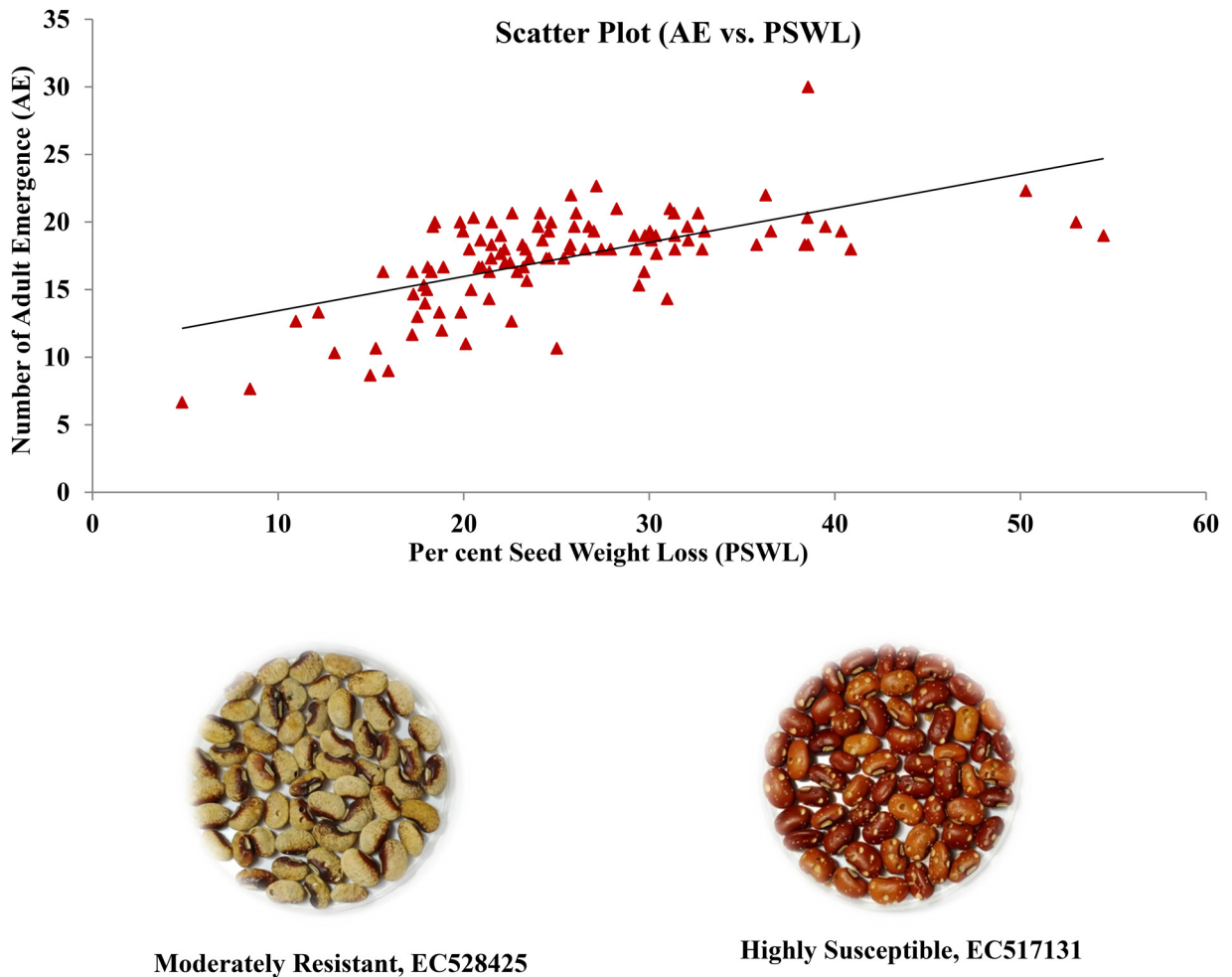


Fig. 2. Scatter plot showing two key traits viz. adult emergence (AE) and PSWL and representation of cowpea accessions.

Table 3. Correlation matrix of growth parameters (4) of bruchid and physico-biochemical parameters (6) of cowpea accessions

	PSWL	EH	OP	AE	SL	SW	SR	SH	SC	PC
PSWL	–	<b>0.335*</b>	<b>0.219*</b>	<b>0.534*</b>	–0.154	–0.071	<b>0.219*</b>	0.034	0.231*	–0.219*
EH		–	<b>0.199*</b>	<b>0.780*</b>	<b>–0.216*</b>	–0.098	0.162	–0.103	0.020	0.042
OP			–	<b>0.432*</b>	–0.135	–0.042	0.153	–0.028	–0.189	–0.194
AE				–	<b>–0.267*</b>	–0.178	<b>0.212*</b>	<b>–0.332*</b>	0.165*	–0.021*
SL					–	<b>0.688*</b>	<b>–0.643*</b>	0.164	0.131	<b>–0.210*</b>
SW						–	–0.115	0.109	0.170	–0.151
SR							–	–0.011	–0.011	<b>0.373*</b>
SH								–	–0.093	–0.177
SC									–	0.098
PC										–

PSWL, per cent seed weight loss; EH, exit hole; OP, oviposition; AE, adult emergence; SL, seed length; SW, seed width; SR, seed roundness; SH, seed hardness; SC, sugar content; PC, phenol content.

\*Significant at 5% level.

Plant breeders often utilize landrace diversity in the development of new cultivars (Frankel *et al.*, 1998; Casañas *et al.*, 2017). They have been recognized as an important source of genetic diversity for resistance to biotic and abiotic stresses (Allard, 1990; Brush, 1995; Frankel *et al.*, 1998; Hoisington *et al.*, 1999; Araújo and Nass, 2002; Scholten *et al.*, 2009; Maxted *et al.*, 2012, 2013).

In cowpea, global reports indicate the existence of very few landraces having superior bruchid resistance. Screening of more than 15,000 cowpea accessions at the International Institute of Tropical Agriculture (IITA), Nigeria, revealed only three landraces, TVu11952, Tvu11953 and Tvu2027 to be moderately resistant to *C. maculatus* (Srinives *et al.*, 2007). Out of thousands of *Vigna* accessions screened for bruchid resistance, V2709 and V2802 of green gram were moderately resistant to bruchid and VM2011 of black gram was resistant to bruchid infestation (Talekar and Lin, 1992; Tripathy, 2016). Similar results were recorded with regards to cowpea landrace *Goa local* having moderate resistance to *C. maculatus* (Nagaraja, 2006). In the current study, four landraces viz., IC330950, IC331212, IC371749 and IC397349 were found moderately resistant on the basis of PSWL. Though the number is not significant, these accessions merit utilization in breeding programmes since they would facilitate widening of genetic base in improved cultivars.

Earlier workers have also reported the differential reaction of various pulses to bruchid infestation. Tripathi *et al.* (2015) screened 52 accessions of cowpea to pulse beetle (*C. chinensis*) and observed significant differences among the accessions in terms of number of eggs laid, development period, adult emergence, number of emergence holes and seed weight loss. Kananji (2007) evaluated 42 genotypes of bean for resistance to Mexican bean weevil, *Zabrotes subfasciatus* (Boheman) and found significant differences in grain weight loss (%) and number of adults emerged.

In the current study, the ovipositional behaviour of *C. maculatus* varied significantly amongst accessions of cowpea. The differential responses of *C. maculatus* for oviposition might be due to odour of the seed emanating from its chemical constituents (Howe and Currie, 1964). Certain physical and chemical factors of the host seed play an essential role in regulating these responses (Gokhale and Srivastava, 1975). Bruchids are reported to be guided in their ovipositional preferences by seed surface, colour, texture, volume, curvature (Gokhale *et al.*, 1990) and the nutritional value of seed (Satya, 1980). Raina (1970) observed that the number of eggs laid on a single seed depends on the size of the host seed and the bruchid species involved. However, no correlation exists between the preferential oviposition for different seeds and the subsequent larval development.

Larval mortality is of considerable relevance in the host plant suitability, which is assessed on the basis of adult

emergence (Wiklund, 1973; Amusa *et al.*, 2018), manifested by the round exit holes carrying a 'flap' of seed coat formed during the exit of the insect. Manohar and Yadava (1990) studied the extent of damage by *C. maculatus* on 10 popular cultivars of cowpea. Of these, Udaipur-2 variety suffered the maximum loss of 44.97% in apparent weight, while Co-1 recorded the least loss of 16.25%. Obiadalla *et al.* (2007) screened 21 cultivars of cowpea for resistance to weevil, based on development assessment of various parameters. They classified them into three groups, sensitive, moderately tolerant and highly tolerant. The oviposition response and development of *C. chinensis* on different cowpea varieties revealed that pulse beetle preferred all the varieties for egg-laying, while differences in adult emergence and PSWL were observed (Singh and Sharma, 2003; Tripathi *et al.*, 2013).

The seed parameters analysed in this experiment exhibited significant variations. Seed hardness, small seed size, absence of nutritional factors and presence of toxic substances are known to affect bruchid damage in leguminous seeds (Kpoviessi *et al.*, 2019). Wrinkled seeds are not preferred for the growth and development of beetles. Cowpea weevil prefers smooth-coated seeds to wrinkled seeds for oviposition, and first instar larvae successfully penetrate the seed coat more in smooth seeds than in rough seeds (Nwanze and Horber, 1976). Erler *et al.* (2009) reported that rough (wrinkled) and thick seed coat of chickpea germplasm might be responsible for resistance to pulse beetle, *C. maculatus*. In our study, it was found that seed shape and testa texture of the two resistant accessions were kidney shape and rough texture, respectively. However, its contribution in resistance was not predictable as other accessions with kidney shape and with rough testa texture were susceptible/highly susceptible. In most of the cowpea accessions, the colour, texture and shape of seed had no direct influence on the resistance or susceptibility to beetles. Therefore, an absolute relationship between seed parameters and insect resistance could not be established. This might be due to the fact that the process of resistance involves morphological, physiological and biochemical mechanisms which range from simply minimizing the effect of insect attack to adversely affecting the insects' cellular processes, growth and development (Singh, 2002). Kapila and Pajni (1989) screened seeds of 17 cultivars of *Phaseolus vulgaris* for resistance to *Z. subfasciatus* and concluded that neither size nor colour of the seeds was important for susceptibility. Similarly, Hussain *et al.* (1997) observed that size, colour and protein content of the seeds have no influence on the susceptibility of green gram seeds to *C. chinensis*.

In bruchids, larva is the only feeding stage and is the most crucial stage determining the resistance/susceptibility of the cultivars. The intensity of larval feeding, measured through mean per cent loss in seed weight varied

significantly among different accessions, as elaborated above, and the result is also in conformation with earlier reports. The analysis of PSWL revealed significant positive correlation ( $r=0.335$ ) with exit holes, oviposition ( $r=0.219$ ), adult emergence ( $r=0.534$ ) and seed roundness ( $r=0.219$ ). In a previous study, Tripathi *et al.* (2012) reported that PSWL had a positive relationship with adult emergence of *C. maculatus*. Correlation with biochemical parameters indicated that PSWL had a significant positive correlation ( $r=0.231$ ) with sugar content and significant negative correlation with phenol content ( $r=-0.219$ ). Tripathi *et al.* (2013) reported that phenol content had a significant negative relationship with a growth index of *C. chinensis*. It is widely accepted that phenolics play a vital role in protecting plants from both insect and mammalian herbivory (Corcuera, 1993; Simmonds, 2003). The size of seeds is also critical in ascertaining the level of resistance. Small-sized grains are known to offer more resistance to pest attack than the larger grains because the latter supply more food and space for larval growth than small-sized grains (Singh *et al.*, 1974). However, this is not universally applicable, as per studies conducted by Lephale *et al.* (2012). Shangane (1.42 g) and Pan 311 (1.45 g) were among the cultivars with small seed size, yet were infested with a high number of bruchids. In contrast, Red Caloona, having relatively larger seed size, recorded minimal insect numbers. Talekar and Lin (1992) also investigated characteristics of resistance to *C. chinensis* in two green gram and one black gram accessions and concluded that smaller seed size of the accessions was not responsible for the resistance. Comparison of seed hardness with adult emergence revealed a significant negative correlation ( $r=-0.332$ ). Seed coat thickness also could not be concluded as a factor conferring resistance, as the grubs penetrated and reached the cotyledons in all the cases.

On contrary, one of the earliest studies on insect resistance by Southgate (1979) had demonstrated that size and hardness of the seeds influence the adult emergence and this was further supported by Semple (1992), wherein the size of insect population on a genotype was reported to be regulated by the reduced oviposition caused due to physical seed barriers. As per their report, the barrier may either limit access into the grain or make it unsuitable for oviposition and difficult for eggs to adhere to the seed or prevent the larva from penetrating the seed when they hatch. But, according to our study, the seed hardness or thickness, apparently do not serve the purpose of a physical barrier that can effectively prevent insect activity. It has also been reported by Pankaj and Singh (2011) that the seed morphological characters were not related to the ovipositional preference and host suitability of the insect pest, in different pulse seeds.

## Conclusion

Globally, cowpea serves as an affordable source of food and nutrition. But, storage losses have emerged as major constraints in its production. Control measures such as physical, chemical and cultural methods, may not adequately deal with the problem of bruchid damage. As a result, host plant resistance is one of the most effective and sustainable measures to limit the damage of bruchid. But, the sources of resistance are very few among the commercial varieties and a paradigm shift in breeding programme is needed to ensure greater use of landraces and local germplasm to find a durable source of bruchid resistance. In our study, cowpea accessions were categorized based on the parameters of per cent loss in seed weight and adult emergence. Accessions, EC528425 and EC528387, were identified as resistant and moderately resistant, based on parameters of PSWL and number of adult emergence respectively, and have the potential for use in conventional breeding programmes, to develop resistant cultivars. No strong correlation was observed between the seed parameters and insect infestation. However, the correlation between biochemical parameters and insect infestation indicated that bruchids have the least preference for accessions with less sugar content and high phenol content. The mechanism of resistance may be due to the effect of physico-biochemical characteristics of seeds that prevent females from laying eggs or the larvae from entering into the seeds. Conclusively, the current study indicated that huge variability exists in germplasm collections, with regards to bruchid response. A combination of factors plays a role in imparting resistance or susceptibility to the bruchid. It necessitates a systematic and effective evaluation of a large number of Genebank accessions to find sustainable and durable source of resistance in cowpea.

## Supplementary material

The supplementary material for this article can be found at <https://doi.org/10.1017/S1479262120000180>.

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## Conflict of interest

The authors declared that they have no conflict of interest in the content of the manuscript and study undertaken.



## References

- Allard RW (1990) The genetics of host-pathogen coevolution: implications for genetic resource conservation. *Journal of Heredity* 81: 1–6.
- Amusa OD, Ogunkanmi LA, Adetumbi JA, Akinyosoye ST and Ogunlape OT (2018) Genetics of bruchid (*Callosobruchus maculatus* Fab.) resistance in cowpea (*Vigna unguiculata* (L.) Walp.). *Journal of Stored Products Research* 75: 18–20.
- Anglin NL, Amri A, Kehel Z and Ellis D (2018) A case of need: linking traits to Genebank accessions. *Biopreservation and Biobanking* 16: 337–349.
- Appleby JH and Credland PF (2003) Variation in responses to susceptible and resistant cowpeas among West African populations of *Callosobruchus maculatus* (Coleoptera: Bruchidae). *Journal of Economic Entomology* 96: 489–502.
- Araújo PMD and Nass LL (2002) Caracterização e avaliação de populações de milho crioulo. *Scientia Agricola* 59: 589–593.
- Bressani R (1985) *Nutritive Value of Cowpeas*. New York: John Wiley & Sons, pp. 355–360.
- Brush SB (1995) *In situ* conservation of landraces in centers of crop diversity. *Crop Science* 35: 346–354.
- Byrne PF, Volk GM, Gardner C, Gore MA, Simon PW and Smith S (2018) Sustaining the future of plant breeding: the critical role of the USDA-ARS National Plant Germplasm System. *Crop Science* 58: 451–468.
- Carrillo-Perdomo E, Raffiot B, Ollivier D, Deulvot C, Magnin-Robert JB, Tayeh N and Marget P (2019) Identification of novel sources of resistance to seed weevils (*Bruchus* spp.) in a faba bean germplasm collection. *Frontiers in Plant Science* 9: 1914, doi:10.3389/fpls.2018.01914.
- Casañas F, Simó J, Casals J and Prohens J (2017) Toward an evolved concept of landrace. *Frontiers in Plant Science* 8: 145, doi:10.3389/fpls.2017.00145.
- Corcuera LJ (1993) Biochemical basis for the resistance of barley to aphids. *Phytochemistry* 33: 741–747.
- de Carvalho MAAP, Bebeli PJ, Bettencourt E, Costa G, Dias S, Dos Santos TM and Slaski JJ (2013) Cereal landraces genetic resources in worldwide GeneBanks. A review. *Agronomy for Sustainable Development* 33: 177–203.
- Díez MJ, De la Rosa L, Martín I, Guasch L, Cartea ME, Mallor C, Casals J, Simó J, Rivera A, Anastasio G and Prohens J (2018) Plant Genebanks: present situation and proposals for their improvement. The case of the Spanish network. *Frontiers in Plant Science* 9: 1794, doi:10.3389/fpls.2018.01794.
- Engels JMM (2002) Genebank management: an essential activity to link conservation and plant breeding. *Plant Genetic Resources Newsletter* 129: 17–24.
- Erler F, Ceylan F, Erdemir T and Tokar C (2009) Preliminary results on evaluation of chickpea, *Cicer arietinum*, genotypes for resistance to the pulse beetle, *Callosobruchus maculatus*. *Journal of Insect Science* 9: 58.
- FAO (2010) *The Second Report on the State of the World's Plant Genetic Resources for Food and Agriculture*. Rome: FAO. Available at [www.fao.org/agriculture/crops/thematic-sitemap/theme/seeds-pgr/sow/sow2/en/](http://www.fao.org/agriculture/crops/thematic-sitemap/theme/seeds-pgr/sow/sow2/en/). Accessed 5 June 2020.
- FAOSTAT (2020) FAOSTAT. Retrieved on 2 April 2020, from <http://www.fao.org/faostat/en/#data/QC>.
- Frankel OH, Brown AHD and Burdon JJ (1998) *The Conservation of Plant Biodiversity*, 2nd edn. Cambridge: Cambridge University Press, pp. 56–78.
- Giga D (1995) Selection of oviposition sites by cowpea weevils *Callosobruchus rhodesianus* (Pic.) and *Callosobruchus maculatus* (F.). *Insect Science and its Application* 16: 145–149.
- Gokhale VG and Srivastava BK (1975) Ovipositional behaviour of *C. maculatus* (Coleoptera: Bruchidae). I. Distribution of eggs and relative ovipositional preference on several leguminous seeds. *Indian Journal of Entomology* 37: 122–128.
- Gokhale VG, Honda H and Yamamoto I (1990) Role of physical and chemical stimuli of legume host seeds in comparative ovipositional behaviour of *C. maculatus* (Fab.) and *C. chinensis* (L.) (Coleoptera: Bruchidae). In: Fujii K, Gatehouse AMR, Johnson CD, Mitchel R and Yoshida T (eds) *Bruchids and Legumes: Economics, Ecology and Coevolution Series Entomologica*. Dordrecht: Springer, pp. 45–51.
- Hodgkin T, Rao VR, Cibrian-Jaramillo A and Gaiji S (2003) The use of ex situ conserved plant genetic resources. *Plant Genetic Resources* 1: 19–29.
- Hoisington D, Khairallah M, Reeves T, Ribaut JM, Skovmand B, Taba S and Warburton M (1999) Plant genetic resources: what can they contribute toward increased crop productivity? *Proceedings of the National Academy of Sciences* 96: 5937–5943.
- Howe RW and Currie JE (1964) Some laboratory observations on the rates of development, mortality and oviposition of several species of bruchidae breeding in stored pulses. *Bulletin of Entomological Research* 55: 437–477.
- Hussain M, Roy GC and Husain M (1997) Laboratory evaluation of some greengram strains for susceptibility to pulse beetle, *Callosobruchus chinensis* (L.). *Bangladesh Journal of Entomology* 7: 21–26.
- IBPGR (1983) International Board for Plant Genetic Resources AGPG: IBPGR/82/80 June, 1983.
- Kanarji GAD (2007) *Study of bruchid resistance and its inheritance in Malawian dry bean germplasm*. PhD Thesis, University of KwaZulu-Natal Republic of South Africa.
- Kapila R and Pajni HR (1989) Screening of different cultivars of *Phaseolus vulgaris* to the attack of *Zabrotes subfasciatus*. *Bulletin of Entomology* 28: 138–144.
- Kehel Z, Sanchez-Garcia M, El Baouchi A, Aberkane H, Tsivelikas A, Chen C and Amri A (2020) Predictive characterization for seed morphometric traits for genebank accessions using genomic selection. *Frontiers in Ecology and Evolution* 8: 32, doi:10.3389/fevo.2020.00032.
- Kell S, Marino M and Maxted N (2017) Bottlenecks in the PGRFA system: stakeholders' perspectives. *Euphytica* 213: 170.
- Khoury C, Laliberté B and Guarino L (2010) Trends in ex situ conservation of plant genetic resources: a review of global crop and regional conservation strategies. *Genetic Resources and Crop Evolution* 57: 625–639.
- Kpoviessi AD, Agbahounga S, Agoyi EE, Chougourou DC and Assogbadjo AE (2019) Resistance of cowpea to cowpea bruchid (*Callosobruchus maculatus* Fab.): knowledge level on the genetic advances. *Journal of Plant Breeding and Crop Science* 11: 185–195.
- Lattanzio V, Terzano R, Cicco N, Cardinali A, Di Venere D and Linsalata V (2005) Seed coat tannins and bruchid resistance in stored cowpea seeds. *Journal of the Science of Food and Agriculture* 85: 839–846.
- Lephale S, Abraham A-B and Victoria A (2012) Susceptibility of seven cowpea cultivars (*Vigna unguiculata*) to cowpea beetle (*Callosobruchus maculatus*). *Agricultural Science Research Journal* 2: 65–69.
- Mahajan R, Sapra R, Umesh S, Singh M and Sharma G (2000) *Minimal Descriptors for Characterization and Evaluation of Agri-Horticultural Crops (Part I)*. New Delhi: National Bureau of Plant Genetic Resources, p. 230.

- Mahendran K and Mohan S (2002) Technology adoption, estimation of loss and farmers behavior in pulses storage. A study in Western Tamil Nadu. *Pestology* 26: 35–38.
- Manohar SS and Yadava SRS (1990) Laboratory observation on relative resistance and susceptibility of some cowpea cultivars to pulse beetle, *Callosobruchus maculatus* F. (Bruchidae: Coleoptera). *Indian Journal of Entomology* 52: 180–186.
- Marshall DR (1989) Limitations to the use of germplasm collections. In: Brown AHD, Frankel OH, Marshall DR and Williams JT (eds) *The Use of Plant Genetic Resources*. Cambridge: Cambridge University Press, pp. 105–122.
- Mascher M, Schreiber M, Scholz U, Graner A, Reif JC and Stein N (2019) Genebank genomics bridges the gap between the conservation of crop diversity and plant breeding. *Nature Genetics* 51: 1076–1081.
- Maxted N, Dulloo ME, Ford-Lloyd B V, Frese L, Iriondo J and de Carvalho MAP (2012) *Agrobiodiversity Conservation: Securing the Diversity of Crop Wild Relatives and Landraces*. UK: CABI.
- Maxted N, Magos BJ and Kell S (2013) *Resource Book for Preparation of National Conservation Plans for Crop Wild Relatives and Landraces*. UK: University of Birmingham.
- Nagaraja M (2006) *Evaluation of pigeonpea and cowpea genotypes for bruchid resistance (Bruchidae)*. MSc Thesis, University of Agricultural Sciences, Dharwad, India.
- NBPGR (2020) PGR Portal. Retrieved on 2 January 2020, from [http://www.nbpg.ernet.in:8080/PGRPortal/\(S\(nuqbopigblqmwv45rnce1b32\)\)/Default.aspx](http://www.nbpg.ernet.in:8080/PGRPortal/(S(nuqbopigblqmwv45rnce1b32))/Default.aspx).
- Nwanze K and Horber E (1976) Seed coats of cowpeas affect oviposition and larval development of *Callosobruchus maculatus*. *Environmental Entomology* 5: 213–218.
- Obiadalla-Ali HA, Salman AMA and Abd El-Hady MAH (2007) Screening some local and introduced cowpea cultivars for dry-seed yield and resistance to *C. maculatus* (F.). *Annals of Agricultural Sciences* 52: 197–212.
- Pankaj N and Singh HK (2011) Correlation of seed characters of pulses with host suitability and preference of *C. chinensis* (L.). *Indian Journal of Entomology* 73: 365–370.
- Raina AK (1970) *Callosobruchus* spp. infesting stored pulses (grain legumes) in India and comparative study of their biology. *Indian Journal of Entomology* 32: 303–310.
- Roe JH (1955) The determination of sugar in blood and spinal fluid with anthrone reagent. *Journal of Biological Chemistry* 212: 335–343.
- SAS (2009) *Statistical Analysis Software System, Version 9.3*. Cary, NC, USA: SAS Institute.
- Satya V (1980) Ovipositional response and development of *C. maculatus* (Fab.) on different varieties of cowpea. *Bulletin of Grain Technology* 18: 200–203.
- Scholten M, Green N, Campbell G, Maxted N, Ford-Lloyd B, Ambrose M and Spoor B (2009) Landrace inventory of the UK. In: Veteläinen M, Negri V, Maxted N (eds) *European Landraces: On-Farm Conservation, Management and Use*. Rome, Italy: Bioversity Technical Bulletin, No. 15, Bioversity International, pp. 161–170.
- Semple RL (1992) Host plant and varietal resistance to post-harvest insect attack. In: Semple RL, Hicks PA, Lozare JV and Casterman A (eds) *Towards Integrated Commodity and Pests Management in Grain Storage*. Manila: REGNET and NAPHIRE.
- Simmonds MSJ (2003) Flavonoid-insect interactions: recent advances in our knowledge. *Phytochemistry* 64: 21–30.
- Singh BD (2002) *Plant Breeding: Principles and Methods*. New Delhi, India: Kalyani Publishers.
- Singh S and Sharma G (2003) Preference of *Callosobruchus chinensis* in pea varieties. *Indian Journal of Entomology* 65: 277–280.
- Singh K, Agrawal NS and Girish GK (1974) The oviposition and the development of *Sitophilus oryzae* L. in different high yielding varieties of wheat. *Journal of Stored Products Research* 10: 105–111.
- Slinkard K and Singleton VL (1977) Total phenol analysis: automation and comparison with manual methods. *American Journal of Enology and Viticulture* 28: 49–55.
- Somta C, Somta P, Tomooka N, Ooi PC, Vaughan DA and Srinives P (2008) Characterization of new sources of mungbean (*Vigna radiata* (L.) Wilczek) resistance to bruchids, *Callosobruchus* spp. (Coleoptera: Bruchidae). *Journal of Stored Products Research* 44: 316–321.
- Southgate BJ (1979) Biology of the bruchidae. *Annual Review of Entomology* 24: 449–473.
- Srinives P, Somta P and Somta C (2007) Genetics and breeding of resistance to bruchids (*Callosobruchus* spp.) in *Vigna* Crops: a review. *NU. International Journal of Science* 4: 1–17.
- Talekar NS and Lin CP (1992) Characterization of *Callosobruchus chinensis* (Coleoptera: Bruchidae) resistance in green gram. *Journal of Economic Entomology* 85: 1150–1153.
- Tanksley SD and McCouch SR (1997) Seed banks and molecular maps: unlocking genetic potential from the wild. *Science (New York, N.Y.)* 277: 1063–1066.
- Tripathi K, Shashi B, Prasad TV and Kalyani S (2012) Differential reaction of cowpea (*Vigna unguiculata*) genotypes to pulse-beetle (*Callosobruchus maculatus*). *Vegetos (Bareilly, India)* 25: 367–374.
- Tripathi K, Bhalla S, Srinivasan K, Prasad TV and Gautam RD (2013) Physical and biochemical basis of resistance in cowpea [*Vigna unguiculata* (L.) Walp.] accessions to pulse beetle, *Callosobruchus chinensis* (L.). *Legume Research* 36: 457–466.
- Tripathi K, Chauhan SK, Gore PG, Prasad TV, Srinivasan K and Bhalla S (2015) Screening of cowpea [*Vigna unguiculata* (L.) Walp.] accessions against pulse-beetle, *Callosobruchus chinensis* (L.). *Legume Research* 38: 675–680.
- Tripathi K, Chauhan SK, Gore PG, Mehta PS, Bisht IS and Bhalla S (2017) Evaluation of wheat landraces of north-western Himalaya against rice weevil, *Sitophilus oryzae* L. vis-à-vis physical seed parameters. *Plant Genetic Resources* 15: 321–326.
- Tripathy SK (2016) Bruchid resistance in food legumes – an overview. *Research Journal of Biotechnology* 11: 97–105.
- Upadhyaya HD, Dwivedi SL, Ambrose M, Ellis N, Berger J, Smýkal P and Sharma SK (2011) Legume genetic resources: management, diversity assessment, and utilization in crop improvement. *Euphytica* 180: 27–47.
- Villa TCC, Maxted N, Scholten M and Ford-Lloyd B (2005) Defining and identifying crop landraces. *Plant Genetic Resources* 3: 373–384.
- Wiklund C (1973) Host plant suitability and the mechanism of selection in larvae of *Papilio machaon*. *Entomologia Experimentalis et Applicata* 16: 232–242.