

Appendages of an early Cambrian metadoxidid trilobite from Yunnan, SW China support mandibulate affinities of trilobites and artiopods

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Abstract – Appendage anatomy contributes crucial data for understanding the evolution and ecology of Euarthropoda. The Palaeozoic trilobites show a great diversity of exoskeletons in the fossil record. However, soft parts, especially appendages, have only been discovered from a few trilobite species. Here we report extraordinarily preserved appendages in the trilobite species *Hongshiyanspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980 (Redlichiida, Metadoxididae) from a single mudstone layer of the Xiazhuang fossil assemblage within the Hongjingshao Formation (Cambrian Series 2, Stage 3) near Kunming, Yunnan, SW China. The appendages exhibit the common architecture revealed by other trilobites and artiopods by consisting of a pair of uniramous antennae followed by a series of paired homonomous biramous limbs. The antennae in holaspid individuals comprise up to 27 spinous podomeres and their ontogeny occurs by lengthening of the podomeres. The post-antennal biramous limbs are similar to those in other polymerid trilobites and artiopods by having a single-segmented protopodite and an endopodite comprising seven segments, but possess a unique wide tripartite exopodite with long setae. Sophisticated appendage anatomy, including the body–limb junction, fine setae, putative muscle bundles and duct-type tissues, are also revealed. Appendages of trilobites, artiopods and other upper stem-group euarthropods are compared and summarized. The *H. yiliangensis* appendages highlight the high morphological disparity of exopodites and the conservativeness of endopodites in trilobites and artiopods. This morphological pattern, together with similar body patterning seen in crustaceans but not in chelicerates, supports the mandibulate affinities of trilobites and at least some artiopods.

Keywords: arthropods, trilobites, appendages, Mandibulata, Cambrian, soft-part preservation

1. Introduction

With biomineralized exoskeletons of more than 20 000 species discovered (only inferior to ostracods; Zhang, 2013), the trilobites (= Trilobita) are one of the most diverse extinct groups of Euarthropoda (e.g. Budd & Telford, 2009) that inhabited Palaeozoic seas from the Cambrian explosion (Hollingsworth, 2008) to the end-Permian mass extinction (Owens, 2003). However, in contrast to the megadiversity of exoskeletons, the soft-bodied anatomy of trilobites is poorly known, with only ~ 30 species primarily from Konservat-Lagerstätten showing soft-part preservation, especially appendages (Table 1; also see Hughes, 2003).

To date, all the reported appendages of polymerid trilobites consist of a pair of uniramous deutocerebral antennae and a series of homonomous biramous post-antennal limbs corresponding to each body segment (see Hughes, 2003; Scholtz & Edgecombe, 2005,

2006). It should be emphasized that the morphology of the post-antennal limbs of *Agnostus pisiformis* is distinct from that of polymerid trilobites (Müller & Walossek, 1987), which makes the supposed trilobite affinity of Agnostida problematic (Walossek & Müller, 1990; Fortey, 2001; Hughes, 2003). Although antennae of exactly similar uniramous multi-segmented architecture are preserved in ~ 20 of these polymerid trilobite species, complete post-antennal limbs have only been reconstructed in six species from five of the nine polymerid orders of Trilobita, including *Eoredlichia intermedia*, *Olenoides serratus*, *Triarthrus eatoni*, *Cryptolithus bellulus*, *Ceraurus pleurexanthemus* and *Chotecops ferdinandi* (see Table 1 for details; Fortey, 2001; Hughes, 2003, 2007). The known post-antennal limbs of these different polymerid trilobite species share a biramous architecture, with two rami (endopodite and exopodite) connected to a protopodite (e.g. Hughes, 2003). The protopodite consists of a single segment (e.g. Ramsköld & Edgecombe, 1996) and the endopodite is made up of seven segments (e.g. Hughes,

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Table 1. Updated summary of trilobites reported with preserved appendages, supplemented and modified from Hughes (2003)

Species	Order	Family	Soft parts				Age	Deposits	Preservation	Main references
			an	en	ex	ds				
<i>Eoredlichia intermedia</i>	Redlichiida	Redlichiidae	++	++	++	++	Cam. S3	Chengjiang	Burgess	Shu <i>et al.</i> 1995; Ramsköld & Edgecombe, 1996; Chen & Zhou, 1997; Hou & Bergström, 1997; Hou <i>et al.</i> 2008
<i>Yunnanocephalus yunnanensis</i>	Ptychopariida	Yunnanocephalidae	++	+	+	++	Cam. S3	Chengjiang	Burgess	This paper ; Zeng <i>et al.</i> 2014
<i>Kuanyangia pustulosa</i>	Redlichiida	Redlichiidae	++	+	+	++	Cam. S3	Chengjiang	Burgess	Hou <i>et al.</i> 2017
<i>Hongshiyanaspis yiliangensis</i>	Redlichiida	Metadoxididae	++	++	++	++	Cam. S3	Hongjingshao Fm.	Burgess	Dunbar, 1925
<i>Zhangshania typica</i>	Redlichiida	Gigantopygidae	++	o	o	o	Cam. S3	Hongjingshao Fm.	Burgess	Hu <i>et al.</i> 2013
<i>Olenellus getzi</i>	Redlichiida	Olenellidae	++	o	o	o	Cam. S3	Kinzers Fm.	Burgess	Hu <i>et al.</i> 2013
<i>Redlichia mansuyi</i>	Redlichiida	Redlichiidae	++	+	+	+	Cam. S4	Guanshan	Burgess	Hu <i>et al.</i> 2010, 2013
<i>Redlichia mai</i>	Redlichiida	Redlichiidae	+	+	+	+	Cam. S4	Guanshan	Burgess	Hu <i>et al.</i> 2013
<i>Palaeolenus douvillei</i>	Ptychopariida	Palaeolenidae	+	o	o	o	Cam. S4	Guanshan	Burgess	Hu <i>et al.</i> 2013
<i>Palaeolenus lantenoisi</i>	Ptychopariida	Palaeolenidae	++	+	+	++	Cam. S4	Guanshan	Burgess	Hu <i>et al.</i> 2013
<i>Megapalaeolenus deprati</i>	Ptychopariida	Palaeolenidae	++	o	o	o	Cam. S4	Guanshan	Burgess	Walcott, 1918, 1921; Raymond, 1920; Størmer, 1939; Whittington, 1975, 1980
<i>Redlichia takooensis</i>	Redlichiida	Redlichiidae	+	o	o	o	Cam. S4	Emu Bay Shale	Burgess	McHenry & Yates, 1993
<i>Olenoides serratus</i>	Corynexochida	Dorypygidae	++	++	++	+	Cam. S5	Burgess Shale	Burgess	
<i>Kootenia burgessensis</i>	Corynexochida	Dorypygidae	o	+	+	o	Cam. S5	Burgess Shale	Burgess	
<i>Elrathia permulta</i>	Ptychopariida	Alokistocaridae	++	o	o	o	Cam. S5	Burgess Shale	Burgess	
<i>Elrathina sp.</i>	Ptychopariida	Ptychopariidae	o	+	+	o	Cam. S5	Burgess Shale	Burgess	
<i>Elrathia kingi</i>	Ptychopariida	Alokistocaridae	o	+	o	+	Cam. Drumian	Wheeler Fm.	Burgess	Briggs <i>et al.</i> 2008
<i>Agnostus pisiformis</i>	Agnostida	Agnostidae	++	++	++	o	Cam. Guzhangia	Orsten	Phosphatic	Müller & Walossek, 1987
<i>Symphysurus sp.</i>	Asaphida	Nileidae	+	+	o	+	Ord. Tremadocian	Fezouata	Burgess	Van Roy <i>et al.</i> 2010
<i>Placoparia cambriensis</i>	Phacopida	Pliomeridae	++	o	o	+	Ord. Darrivilian	Lower Llanvirn	Mudstone mould	Whittington, 1993
<i>Triarthrus eatoni</i>	Ptychopariida	Olenidae	++	++	++	+	Ord. Katian	Beecher's-type beds	Pyritic	Cisne, 1975, 1981; Whittington & Almond, 1987; Farrell <i>et al.</i> 2009
<i>Cryptolithus bellulus</i>	Asaphida	Trinucleidae	+	+	+	o	Mid.-Upp. Ord.	Beecher's-type beds, Trenton Gr.	Pyritic, calcitic	Walcott, 1881, 1912; Beecher, 1895; Raymond, 1920; Størmer, 1939; Bergström, 1972; Farrell <i>et al.</i> 2009
<i>Primaspis trentonensis</i>	Odontopleurida	Odontopleuridae	o	+	+	o	Upp. Ord.	Trenton Gr.	Calcitic	Walcott, 1918, 1921; Raymond, 1920; Størmer, 1939, 1951
<i>Ceraurus pleurexanthemus</i>	Phacopida	Cheiruridae	o	++	++	o	Upp. Ord.	Trenton Gr.	Mudstone impression	Billings, 1870; Raymond, 1920
<i>Flexicalymene senaria</i>	Phacopida	Calymenidae	o	+	+	o	Upp. Ord.	Trenton Gr.	Mudstone impression	Walcott, 1884, 1918; Raymond, 1920
<i>Isotelus latus</i>	Asaphida	Asaphidae	o	+	o	o	Upp. Ord.	Richmond Gr.	Mudstone impression	Ross, 1979
<i>Isotelus maximus</i>	Asaphida	Asaphidae	o	+	o	o	Upp. Ord.	Richmond Gr.	Mudstone impression	Størmer, 1939; Stürmer & Bergström, 1973; Bergström & Brassel, 1984;
<i>Primaspis sp. X</i>	Odontopleurida	Odontopleuridae	o	+	o	o	Upp. Ord.	Clays Ferry Fm.	Silicified	Bartels, Briggs & Brassel, 1998; Bruton & Haas, 1999
<i>Rhenops cf. anserinus</i>	Phacopida	Acastidae	++	++	o	o	Dev. Emsian	Hunsrück Slate	Pyritic	
<i>Chotecops ferdinandi</i>	Phacopida	Phacopidae	++	++	++	+	Dev. Emsian	Hunsrück Slate	Pyritic	
<i>Asteropyge sp.</i>	Phacopida	Acastidae	++	+	+	o	Dev. Emsian	Hunsrück Slate	Pyritic	

Orders and families are mainly based on Harrington *et al.* (1959) and Whittington *et al.* (1997). Note that the former *Elrathina cordillerae* from the Burgess Shale is revised as an unnamed new species of *Elrathina* (Geyer & Peel, 2017). Preservation of complete, incomplete and absent anatomical structures are indicated by '++', '+', and 'o', respectively. Abbreviations: an – antenna; ds – digestive system; en – endopodite; ex – exopodite.

2003; Boxshall, 2004). Nevertheless, the exopodites show considerable morphological variations in different trilobite species (Müller & Walossek, 1987, fig. 27; Shu *et al.* 1995, fig. 21; also see Bruton & Haas, 1999, fig. 22 and Hou *et al.* 2008, fig. 14 for revised reconstructions for *C. ferdinandi* and *E. intermedia*, respectively). Other exceptional appendages of polymerid trilobites include a pair of antennae-like cerci that has only been found in the pygidium of *O. serratus* (e.g. Whittington, 1975, 1980).

The limited knowledge of the soft anatomy of trilobites and other stem-group euarthropods, especially appendages, has long constrained our understanding of the internal and external phylogenetic relationships of trilobites (see Scholtz & Edgecombe, 2005, 2006 and references therein). It was not until recently that phylogenetic analyses resolved the trilobites within the Artiopoda Hou & Bergström, 1997, as close relatives to several groups of soft-bodied ‘trilobitomorph’ euarthropods from Cambrian Lagerstätten (e.g. Edgecombe & Ramsköld, 1999; Ortega-Hernández, Legg & Braddy, 2013; Stein *et al.* 2013; Legg, Sutton & Edgecombe, 2013). These ‘trilobitomorphs’, including concilitergans (e.g. *Kuamaia* and *Saperion*), nektaspids (e.g. *Naraoia*, *Liwia* and *Emucaris*), xandarelids (e.g. *Xandarella* and *Cindarella*) and other problematic taxa, share the common appendage architecture composed of a pair of uniramous antennae and homonomous post-antennal biramous limbs, as well as other synapomorphies, with polymerid trilobites (e.g. Hou & Bergström, 1997; Edgecombe & Ramsköld, 1999; Zhang, Shu & Erwin, 2007; Ortega-Hernández, Legg & Braddy, 2013). Nevertheless, phylogenetic analyses have not reached a consensus on the sister group of trilobites and cannot determine whether the entire Artiopoda is closer to the Mandibulata or Chelicerata at present (e.g. Budd & Telford, 2009; Ortega-Hernández, Legg & Braddy, 2013; Stein *et al.* 2013; Legg, 2014). Therefore, further studies on trilobite soft anatomy are essential to deliver arguments for answers to these questions.

Here we describe exceptionally preserved appendages of the polymerid trilobite *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980 (Redlichiida, Metadoxididae) from the Xiazhuang fossil assemblage in the Hongjingshao Formation (Cambrian Series 2, Stage 3) near Kunming, Yunnan, SW China. The new material confirms the basic architecture of trilobite/artiopodan appendages, but also exhibits a morphological disparity in trilobite/artiopodan exopodites, providing new information for comparative anatomy and elucidating the affinities of trilobites and other related artiopods.

2. Geological setting

All specimens of *Hongshiyanaspis yiliangensis* were recovered from a single yellowish structureless claystone layer intercalated within sandstone layers from the lower part of the Hongjingshao Formation, the

lower fossil horizon yielding the Xiazhuang fossil assemblage at Xiazhuang, Chenggong, Kunming, eastern Yunnan, SW China (see Zeng *et al.* 2014 for detailed information on geography and stratigraphy). The soft-part preservation and various angles of burial of the fossils indicate that the fossiliferous layer was deposited rapidly, probably by a storm event (e.g. Zhu *et al.* 2001; Hu *et al.* 2010, 2013). Other euarthropods recovered from the same layer only include a large bivalve euarthropod *Jugatacaris?*, whose biramous limbs comprise more than 20 endopodite podomeres and are readily distinguishable from the trilobite limbs (Zeng *et al.* 2014). Although other trilobite species including *Yunnanocephalus yunnanensis*, *Malongocephalus yunnanensis* and *Kuanyangia (Sapushania) granulosa* were also found in the upper fossil horizon (Zeng *et al.* 2014), none of these species were discovered from the *Hongshiyanaspis* bed. The age of the Xiazhuang fossil assemblage is approximately identical to that of the lower part of the Xiaoshiba Lagerstätte (e.g. Hou *et al.* 2017) because both fossil assemblages are from the same stratigraphic interval in the lower part of the Hongjingshao Formation and from the same *Eoredlichia–Wutingaspis* Assemblage Zone of the regional Qiongzhusian Stage (Cambrian Stage 3). This fossil zone also yields the renowned Chengjiang biota from the underlying Yu’anshan Formation (Zhu *et al.* 2001). The similar age and faunal compositions suggests that these two fossil assemblages from the Hongjingshao Formation can be regarded as continuing the Chengjiang biota (Zeng *et al.* 2014). However, the upper part of the Xiaoshiba Lagerstätte extends into the regional Canglangpuan (Cambrian Stage 3) *Yiliangella* Assemblage Zone represented by *Zhangshania typica* (Hou *et al.* 2017), an interval which is absent in the section that contains the Xiazhuang fossil assemblage (Zeng *et al.* 2014).

3. Materials and methods

A total of 106 early to fully grown holaspid specimens of the trilobite *Hongshiyanaspis yiliangensis* were studied (Field IDs prefixed by HBHY; see online Supplementary Material available at <http://journals.cambridge.org/geo>). The majority of these specimens are dorsoventrally embedded, with only 11.3% laterally compressed. Nearly half of them (45.3%) exhibit preserved soft parts, including antennae, biramous limbs and parts of the digestive system. Owing to the different angles of burial, the shapes of original three-dimensional structures can vary, especially for the biramous limbs, but structures in various positions or on different levels can also reveal additional details of the morphology. Similar to soft parts of the Chengjiang fossils, the appendages of *H. yiliangensis* are preserved mainly as Fe-rich aluminosilicate films with limited organic ingredients (Zhu, Babcock & Steiner, 2005).

All figured specimens are housed at the Nanjing Institute of Geology and Palaeontology, Chinese

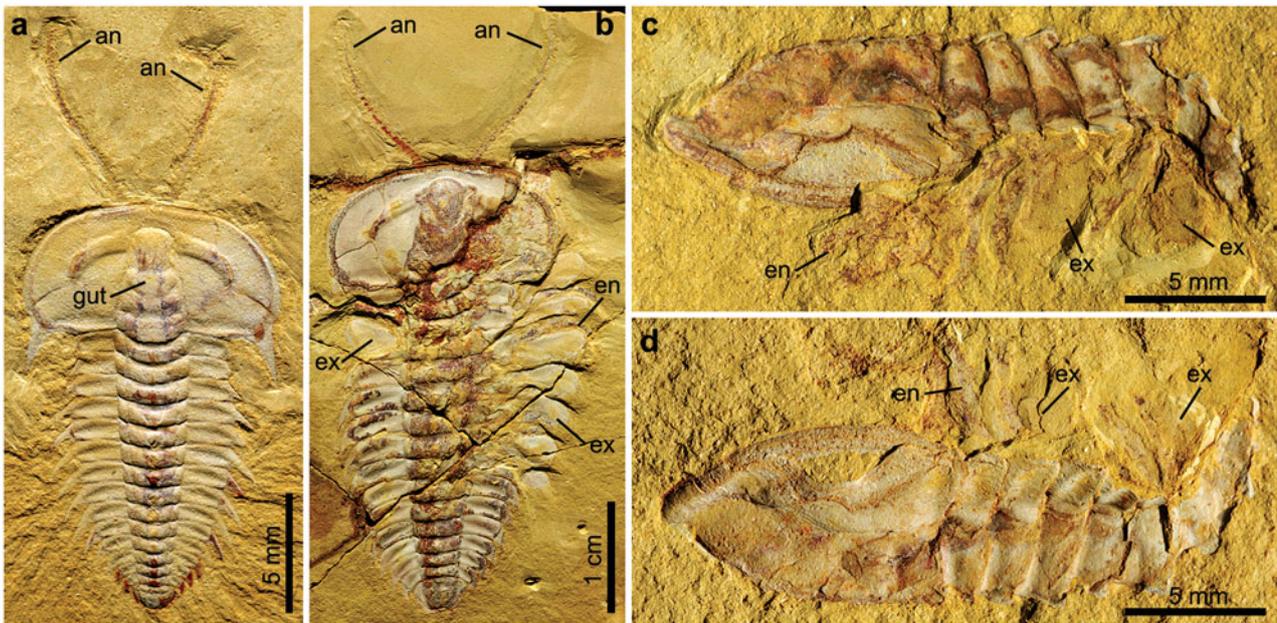


Figure 1. (Colour online) Complete specimens of *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980 preserved with appendages, from the Xiaozhuang fossil assemblage, Kunming, Yunnan, SW China. (a) Dorsoventrally compressed normal-sized holaspid with complete antennae, part, NIGPAS 164504A. (b) Dorsoventrally compressed fully grown holaspid with complete antennae and a series of post-antennal biramous limbs, part only, NIGPAS 164503. (c, d) Laterally compressed normal-sized holaspid with a series of post-antennal biramous limbs, NIGPAS 164505. (c) Part, NIGPAS 164505A. (d) Counterpart, NIGPAS 164505B. Abbreviations: an – antenna; en – endopodite; ex – exopodite; gut – gut of digestive tract.

Academy of Sciences (prefixed by NIGPAS). Appendages were prepared manually using blades. Photographs showing overall morphology were taken using a Nikon D300s digital camera with a Nikon AF-S VR105mm f/2.8G macro lens. Detailed anatomy was captured using a Carl Zeiss SterEO Discovery V12 microscope linked to an AxioCam HR3 digital microscope CCD camera. Illumination from various directions and angles was employed in order to show the three-dimensional structures. Line drawings were prepared on the basis of high-resolution pictures. Measurements were conducted on photographs within Adobe Photoshop™ CS6 and statistically analysed in Microsoft Office Excel™ 2013.

We follow most of the standard terminology for trilobites in Whittington *et al.* (1997), including the terms ‘antenna(e)’, ‘endopodite(s)’ and ‘exopodite(s)’, which are also the most commonly used terms in recent literature on fossil and extant euarthropods. However, the neutral term ‘protopodite’ is used rather than the term ‘basis’ or ‘basipodite’ (e.g. Boxshall, 2004), which is equal to the term ‘coxa’ or ‘coxopodite’ in earlier studies (e.g. Whittington *et al.* 1997). For the protopodite in post-antennal biramous limbs of trilobites and other arthropods, the term ‘basis’ was first introduced by Ramsköld & Edgecombe (1996). However, this term implies the evolutionary hypothesis that an undivided protopodite is homologous to the basis/basipodite in a multi-segmented protopodite with other more proximal podomeres such as the coxa or precoxa (see Boxshall, 2004 for discussion). The corresponding evolutionary scenario would be

that the origin of other non-basal podomeres occurred by addition of proximal podomeres (e.g. Walossek & Müller, 1998; Haug *et al.* 2013), which rejects an alternative by the subdivision of an originally undivided protopodite podomere (see Boxshall, 2004 for discussion).

4. Description of appendages

4.a. General arrangement of appendages

The appendages of a *Hongshiyanaspis yiliangensis* holaspid consist of a single pair of uniramous antennae (an) followed by a series of homonomous post-antennal biramous limbs (Figs 1–3). The preservation of incomplete cephalic biramous limbs (Figs 4c–e, 5c, d, 6a) and three paired digestive glands (gd) on the second and third glabellar lobes and the occipital lobe (Figs 1a, 2a, 4a, b, 5a, b) suggest the presence of three corresponding pairs of post-antennal limbs underneath the cephalon (ce). Each of the 14 thoracic segments (th1–th14) bears a single pair of biramous limbs (Figs 1b–d, 2b, 3, 4e, f, 6–8), as supported by limb fragments connected to the 14th thoracic segment (unfigured fragmentary specimen HBHY008). Fragments of limbs are connected to the first and only axial ring of the pygidium (pg) (unfigured fragmentary specimen HBHY008). It is unknown whether there are limbs corresponding to the terminal axial piece of the pygidium, including the cerci. Variations in shape, if there are any, are insignificant between the post-antennal biramous limbs.

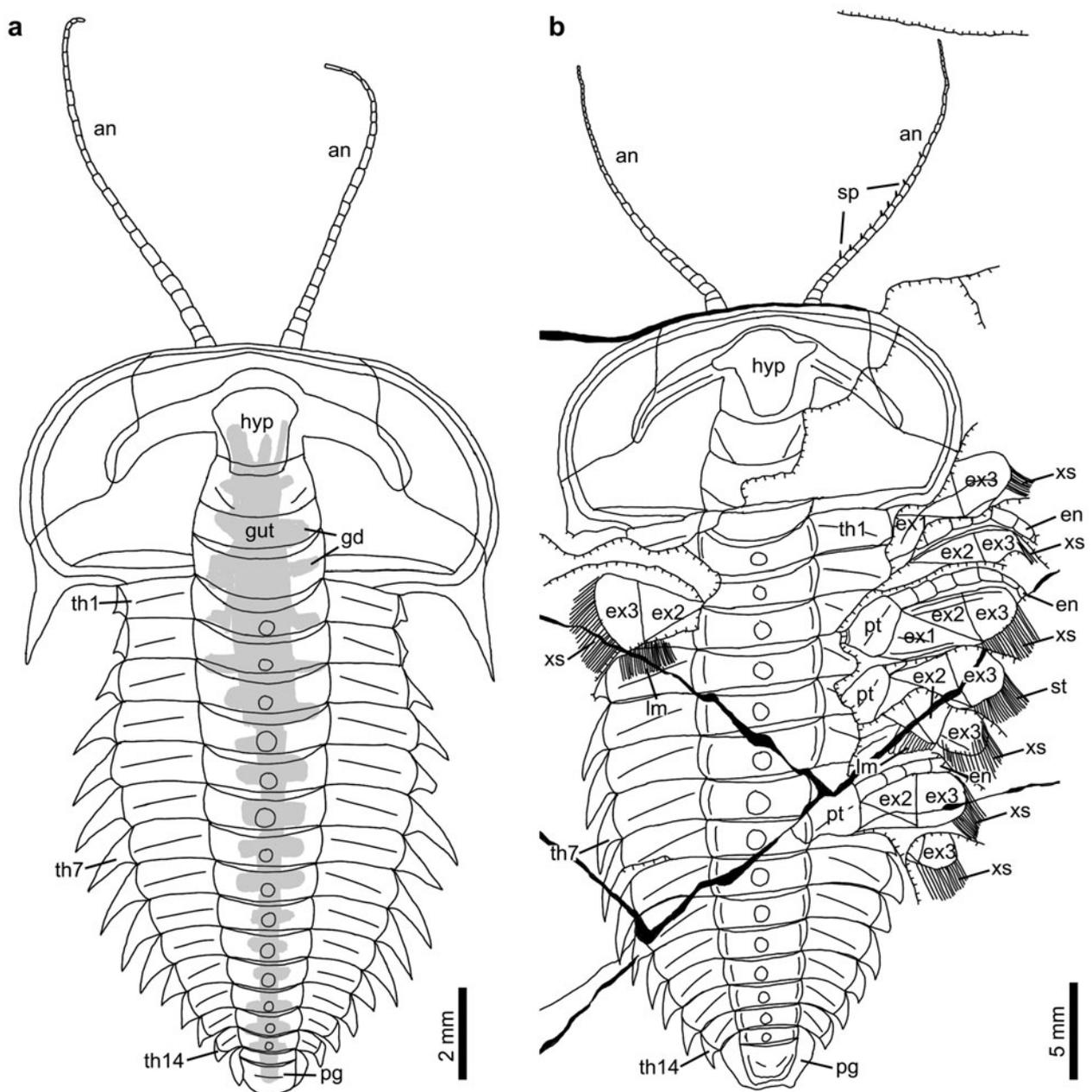


Figure 2. Line drawings of complete specimen of *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980 preserved with appendages, from the Xiaozhuang fossil assemblage, Kunming, Yunnan, SW China. (a) Part, NIGPAS 164504A, as in Figure 1a. (b) Part, NIGPAS 164503, as in Figure 1b. Additional abbreviations: ex1–ex3 – lobes of exopodites 1–3; hyp – hypostome; gd – digestive gland; lm – lamellae; pg – pygidium; pr – protopodite; sp – spines on antennae; th1–th14 – thoracic segments 1–14; xs – setae on exopodites. Grey areas indicate digestive system.

4.b. Antennae

The paired uniramous antennae are slender and flexible (Figs 1a, b, 2a, b). Each is attached to the corresponding side of the hypostome (hyp) (Figs 1a, b, 2, 4c, e, 5d, 6a) and emerge at the anterior rim of the cephalon ventrally as in the possible life position (Figs 1a, b, 2, 4a–c, e, 5a, b, d, 6a). The lengths of complete antennae exceed $\sim 50\%$ of the cephalon's length (Figs 1a, b, 2), and the relative proportion between the antennae and the complete body length decreases from 36% (Figs 1a, 2a) to 29% (Figs 1b, 2b) from the normal-sized to fully grown holaspis periods. Individual an-

tennae may be curved by up to 90° at their middle (Figs 4a, 5a) or terminal sections (Figs 1a, 2a). Both antennae may be stretched apart laterally with an intersection angle of up to 105° (Figs 4a, 5a).

The antennae are composed of up to 27 rectangular podomeres in fully grown holaspis (Figs 1b, 2b), while an approximately similar number of podomeres is also found in normal-sized holaspis (Figs 1a, 2a). Although antennae are also preserved in early holaspis (Figs 4a, 5a), their maximum numbers of podomeres cannot be determined owing to the difficulties in preparation. The most proximal podomeres are evidently stouter than the distal ones (Figs 1a, b, 2,

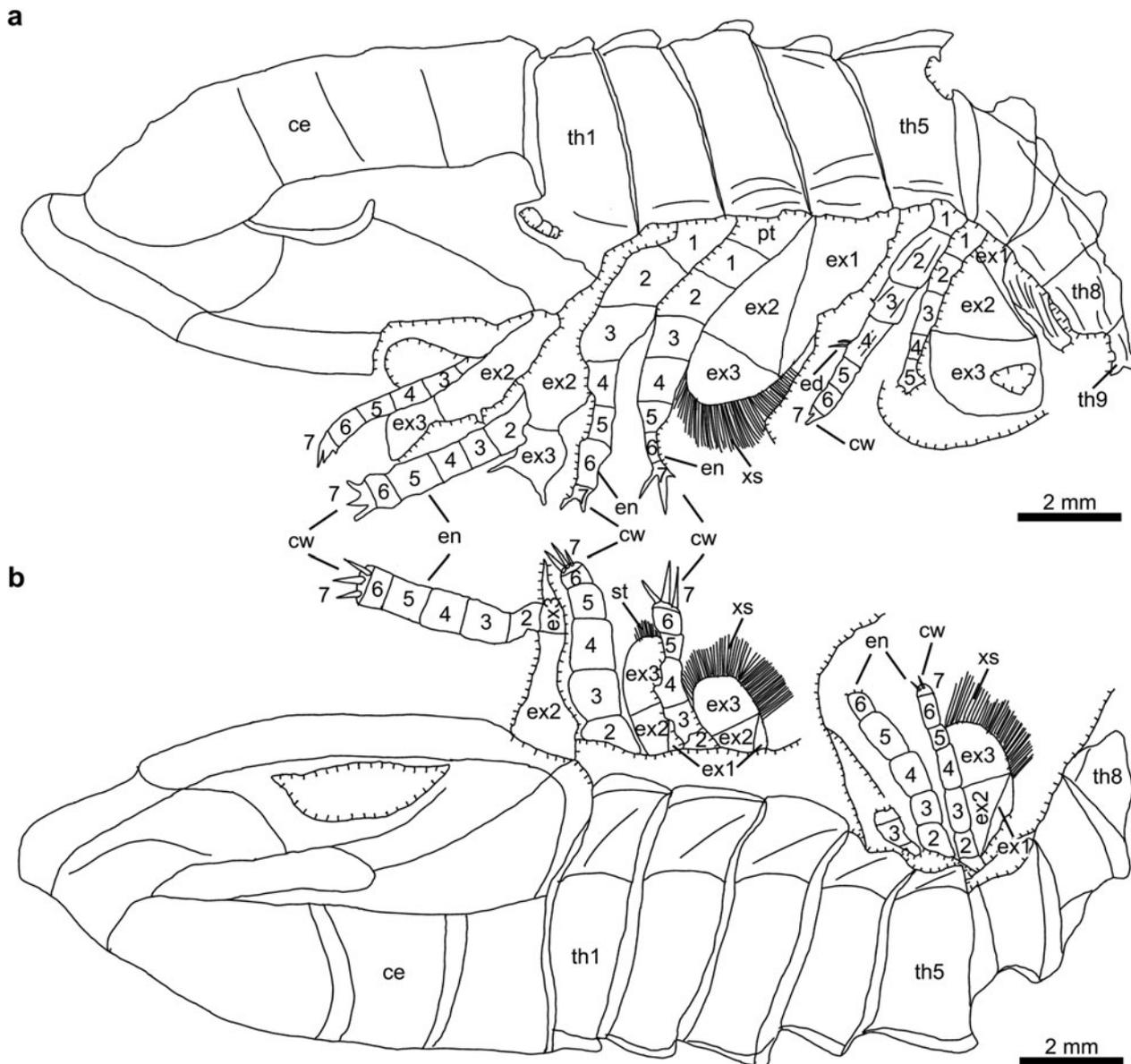


Figure 3. Line drawings of complete specimen of *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980 preserved with appendages, from the Xiaozhuang fossil assemblage, Kunming, Yunnan, SW China, NIGPAS 164505. (a) Part, NIGPAS 164505A, as in Figure 1c. (b) Counterpart, NIGPAS 164505B, as in Figure 1d. Additional abbreviations: ce – cephalon; cw – distal claws on the seventh podomere of the endopodite; ed – spinous endite; 1–7 – podomeres of the endopodite 1–7.

4a–c, e, 5a, b, d, 6a, 9a). Each podomere bears at least one sharp spine (sp) close to its distal arthroal membrane (Figs 1b, 2b, 4e, 6b, 9b). The length of the spine may reach one-third of the podomere length (Fig. 9b).

4.c. Post-antennal limbs

Each post-antennal limb is biramous and consists of a protopodite (pt) comprising a single segment, an endopodite (en) consisting of seven segments and a tripartite exopodite (ex) (Figs 1b–d, 2b, 3, 7a, b, 8a, b, 10, 11a, g, h, 12a–d, 13). The shapes of these limbs are homonomous, but their sizes decrease correspondingly to the sizes of the thoracic segments (Figs 1b–d, 2b, 3, 4e, f, 6–8, 10). The thoracic limbs are longer and wider than the thoracic exoskeleton so that they stretch

out from below the exoskeleton and are arranged in an imbricate series (Figs 1b–d, 2b, 3, 4e, f, 6–8, 10).

4.c.1. Protopodite and body–limb junction

The protopodite is robust and has a subrectangular outline, carrying the endopodite and the exopodite, respectively, at its dorsal and ventral distal margins (Figs 1b–d, 2b, 3, 10, 11a, g, h, 12a–d, 13). It is connected to the body by an arthroal membrane (am) as the body–limb junction (Figs 7c, 8c, 11a, b, i, 12a–g, 13). The body–limb arthroal membrane can be preserved as a section about half the width of the protopodite (Figs 11i, 12a–g, 13), or indicated by subparallel annulations (Figs 10, 11a, b). The attachment site of the protopodite to the body is the lateral side of the

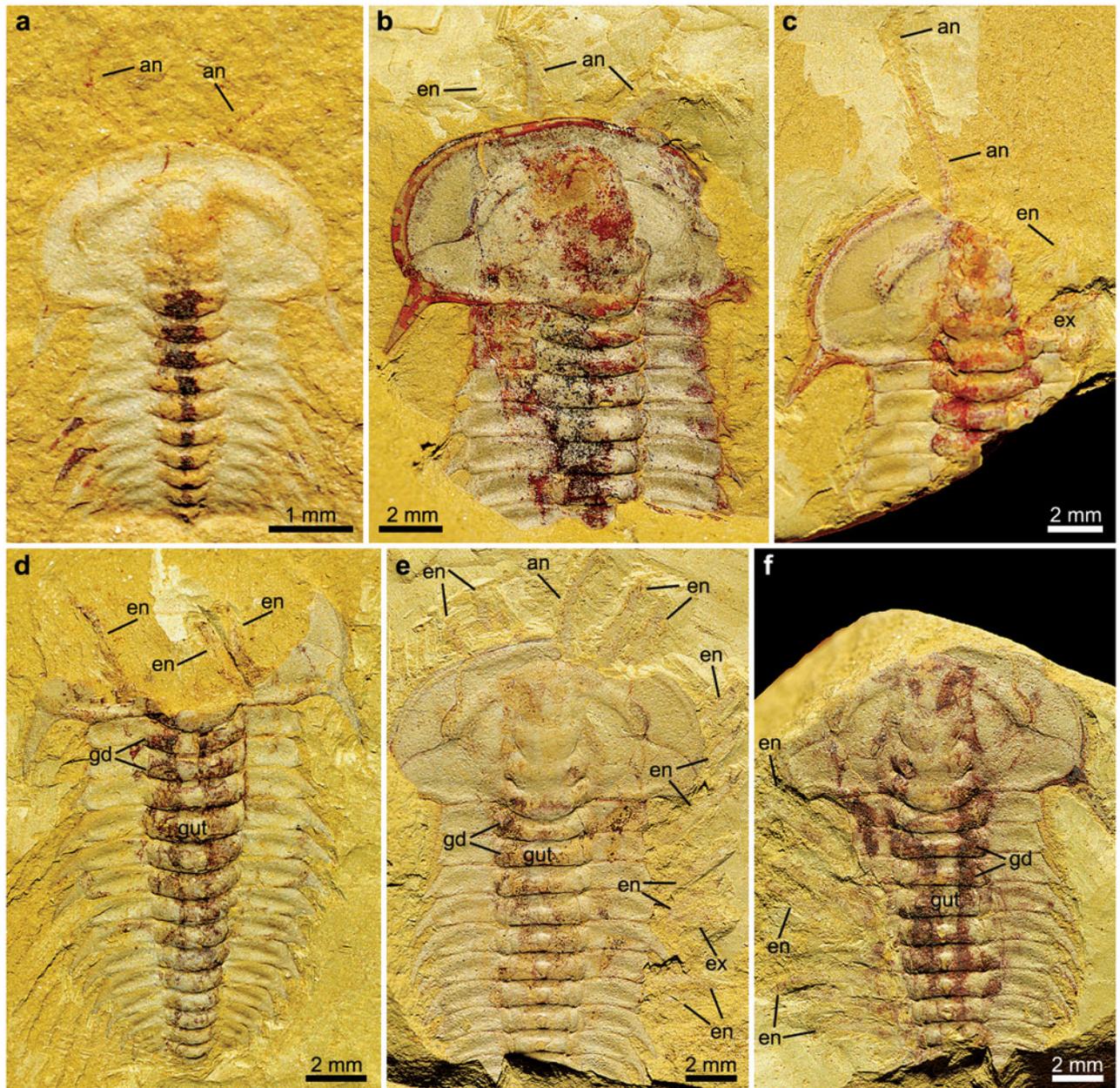


Figure 4. (Colour online) *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980 preserved with appendages, from the Xiaozhuang fossil assemblage, Kunming, Yunnan, SW China. (a) Dorsoventrally compressed early holaspid with antennae, counterpart only, NIGPAS 164514. (b) A dorsally compressed normal-sized holaspid with antennae and post-antennal biramous limbs, counterpart only, NIGPAS 164512. (c) Dorsoventrally compressed normal-sized holaspid with antennae and post-antennal biramous limbs, part, NIGPAS 164510. (d) Dorsoventrally compressed normal-sized holaspid with antennae and post-antennal biramous limbs, part, NIGPAS 164513. (e, f) Dorsoventrally compressed normal-sized holaspid with antennae and post-antennal biramous limbs, NIGPAS 164506. (e) Counterpart, NIGPAS 164506b. (f) Part, NIGPAS 164506a. Abbreviations as in Figures 1–3.

subrectangular to hourglass-shaped sternite (sn) (Figs 7c, 8c, 11i). Putative muscle bundles (ms) are preserved as reddish Fe-rich films along the boundary between the protopodite and the arthrodial membrane (Figs 12e, g, 13b). The protopodite extends significantly in a ventral direction, forming a stout endite (Fig. 12a–g). At least three clusters of robust spines (Figs 12a, b, e, f, 13), together with rows of non-clustered fine spines (Figs 12c, d, g, 13), develop along the ventral margin of the protopodite's endite and form a gnathobase (gs). Numerous fine setae (st) of a few hundred micrometres in length develop on the sur-

face of the protopodite (Figs 12e, f, 13). A shallow transverse furrow is also present on the protopodite (Figs 10, 11a, b). It is most likely to be a real anatomical structure rather than a taphonomic imprint because no corresponding structures are seen on the exoskeleton.

4.c.2. Endopodite

The endopodite composed of seven podomeres is attached to the distal margin of the protopodite by an arthrodial membrane (Figs 1b–d, 2b, 3, 4e, f, 6, 7a, b,

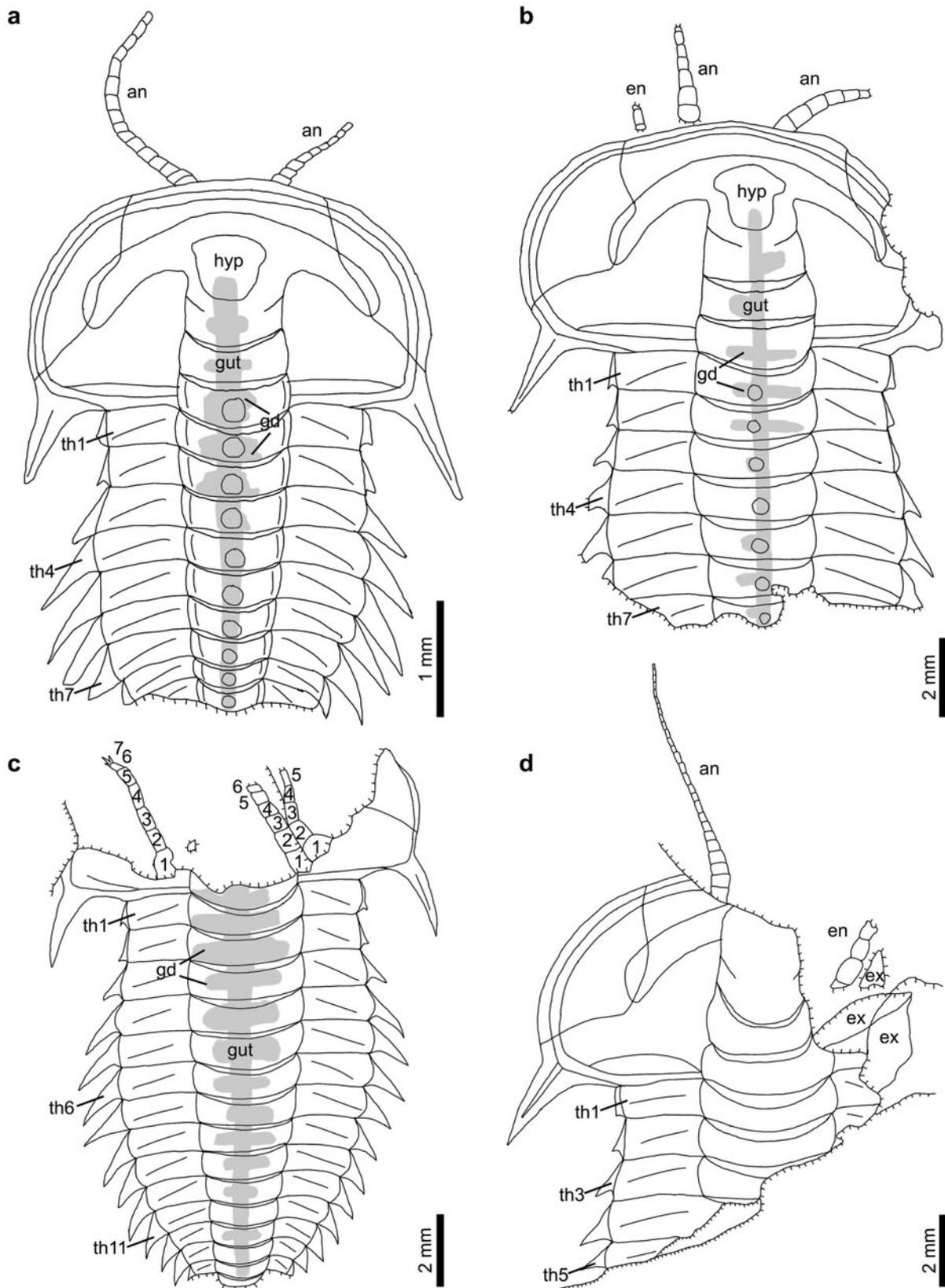


Figure 5. Line drawings of *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980 preserved with appendages, from the Xiaozhuang fossil assemblage, Kunming, Yunnan, SW China. (a) Counterpart only, NIGPAS 164514, as in Figure 4a. (b) Counterpart only, NIGPAS 164512, as in Figure 4b. (c) Part, NIGPAS 164513, as in Figure 4d. (d) Part, NIGPAS 164510, as in Figure 4c. Abbreviations as in Figures 1–3.

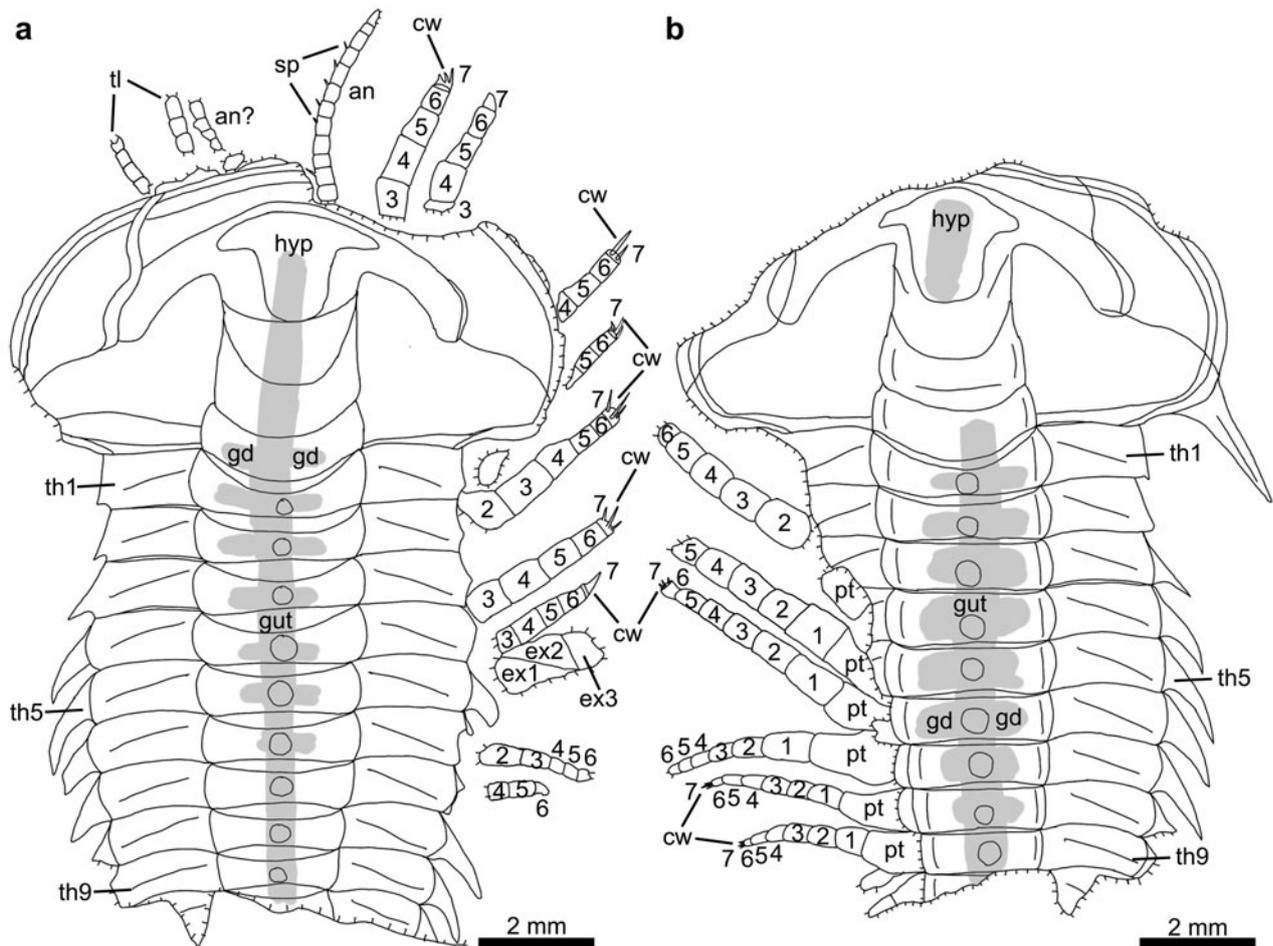


Figure 6. Line drawings of *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980 preserved with appendages, from the Xiaozhuang fossil assemblage, Kunming, Yunnan, SW China. (a) Counterpart, NIGPAS 164506b, as in Figure 4e. (b) Part, NIGPAS 164506a, as in Figure 4f. Grey areas indicate digestive system. Abbreviations as in Figures 1–3.

8a, b, 10, 11a, b, g, 12a–g, 13), which is also indicated by putative oblique muscle bundles (Figs 12c, g, 13b). The total length of the endopodite accounts for ~80% of that of the post-antennal limb (Figs 1b–d, 2b, 3, 10, 11a, g, h, 12d, 13a). The first six podomeres are subrectangular in outline, and their sizes decrease from proximal to distal (Figs 1b–d, 2b, 3, 4c–f, 5c, d, 6, 7a, b, 8a, b, 10, 11a, g, h). Each of these six podomeres bears an endite (ed) with thin spines (Figs 1c, 3a, 12a–d, g, h, 13). Tiny reddish Fe-rich dots interpreted as the bases of setae are present on these podomeres (Figs 12b–d, g, 13). The terminal, seventh podomere is extremely short but connected to the sixth podomere by an arthrodial membrane (Figs 1c, d, 3, 4e, f, 6a, b, 7a, 8a, 11e–h, 12d, h, 13a). Three highly sclerotized sharp claws (cw), one prominent in the middle and two subordinate lateral ones, are attached to the seventh podomere (Figs 11e, f, 12d, h, 13a).

4.c.3. Exopodite

The exopodite is oblong to subrectangular in outline and composed of a tripartite flattened flap (ex1–ex3) with setae (xs) and lamellae (lm) (Figs 1b–d, 2b, 3, 7, 8, 10, 11a–d, g, h, j, 12a–d, 13). The total length

of the exopodite, flap and setae included, is subequal to that of the endopodite (Figs 1b–d, 2b, 3, 10a, b, 11a, g, h, 12a, 13b). The flap is in addition ~50% wider than the sagittal length of the thoracic segment and almost double the maximum width of the endopodite. The exopodite flap is attached to the dorsal margin of the protopodite by an arthrodial membrane (Figs 7c, 8c, 10, 11a, b). Two joints, one transverse and the other oblique, divide the flattened flap into three parts (Figs 1b–d, 2b, 3, 4e, 6a, 7a, b, d, 8a, b, d, 10, 11a, g, h, j, 12a, c, d, 13). The transverse joint runs through about the distal third of the flap, separating the flap into a bell-shaped distal part (the third lobe, ex3) and a trapezoidal proximal part (Figs 1b–d, 2b, 3, 4e, 6a, 7a, b, d, 8a, b, d, 10, 11a, c, d, g, h, j, 12a, c, d, 13, 14). The oblique joint, which starts at the posterior end of the transverse joint and terminates at the distal end of the protopodite–exopodite junction, separates the proximal part of the flap into two subtriangular lobes (the first and second lobes, ex1 and ex2) (Figs 1b–d, 2b, 3, 4e, 6a, 7a, b, d, 8a, b, d, 10, 11a, c, d, g, h, j, 12a, c, d, 13). Up to 40 long, non-overlapping setae (xs) develop along the distal and posterior margins of the third lobe (Figs 1b–d, 2b, 3, 7a, b, 8a, b, 10, 11a, c, d, g, h, 12a, 13b, 14e–g). The maximum lengths

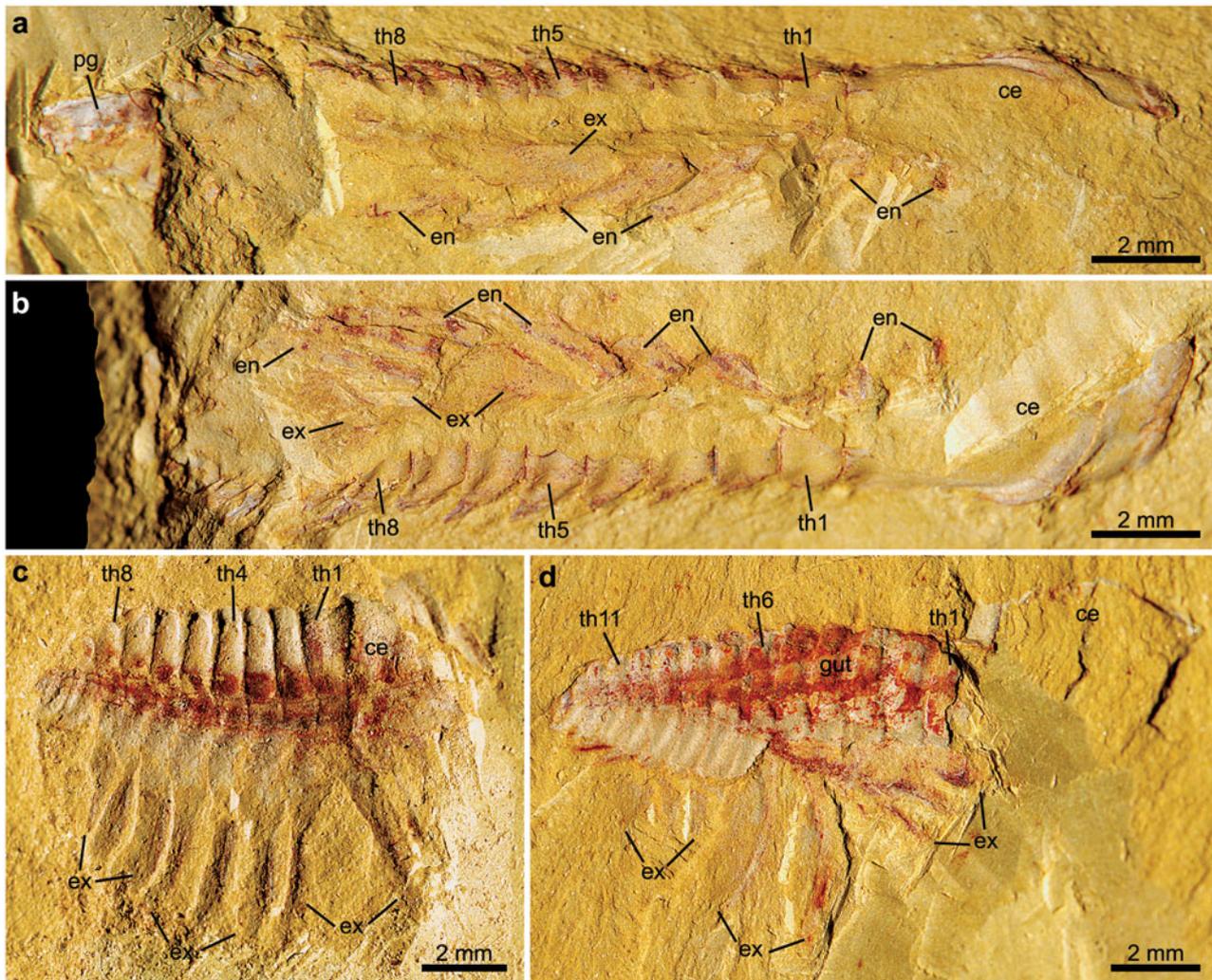


Figure 7. (Colour online) Laterally compressed normal-sized holaspids of *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980 preserved with post-antennal biramous limbs, from the Xiazhuang fossil assemblage, Kunming, Yunnan, SW China. (a) Part, NIGPAS 164507a. (b) Counterpart, NIGPAS 164507b. (c) Part only, NIGPAS 164509. (d) Counterpart, NIGPAS 164511b. Abbreviations as in Figures 1–3.

of the setae are approximately equal to the width of the flap. Flattened imbricate lamellae (lm) develop along the posterior margin of the second lobe (Figs 1b, 2b, 10, 11c, d). They are preserved fragmentarily (Figs 1b, 2b, 10, 11c) or as imprints on the exoskeleton's surface (Figs 1b, 2b, 11d). A marginal rim runs along the margin of the flap (Figs 10, 12a–d, 13, 14). The anterior section of the marginal rim (ar) (Figs 10, 12a–d, 13, 14a–d) is generally wider than its posterior and distal sections (pr and dr) (Figs 10, 12a–d, 13, 14e–g). The anterior sections of the marginal rims of different limbs are in addition inserted by duct-type soft tissues (dt) preserved as reddish mineral films that merge into a main stem connected to the body (Figs 7d, 8d, 11j).

5. Ontogeny of antennae in *Hongshiyanaspis yiliangensis*

The variations in the trilobite exoskeletons during ontogeny have been studied for a long time and in many taxa. However, little is known about the ontogenetic

pattern of trilobite appendages (see Hughes, 2003, 2007 and references therein). The growth of antennae during the ontogeny of trilobites can be performed via two theoretical models: (1) by addition of podomeres; or (2) by stretching of individual podomeres. In order to test these growth models, the number of podomeres, total lengths of the antennae and the average lengths of the podomeres were measured and analysed statistically from our new *Hongshiyanaspis yiliangensis* material, as well as the lengths of the cephalon as quantification of ontogenetic stage (Table 2). A significantly positive correlation ($R^2 = 0.9369$) is found between the average length of the podomeres and the length of the cephalon (Fig. 15). Combined with the similar number of podomeres in nearly complete antennae in normal-sized and fully grown holaspids (Table 2; Figs 1a, b, 2a, b), the growth of antennae is interpreted to occur predominantly by the lengthening of individual podomeres. This may suggest that the number of podomeres increases during the meraspid stages but remains constant during the holaspid

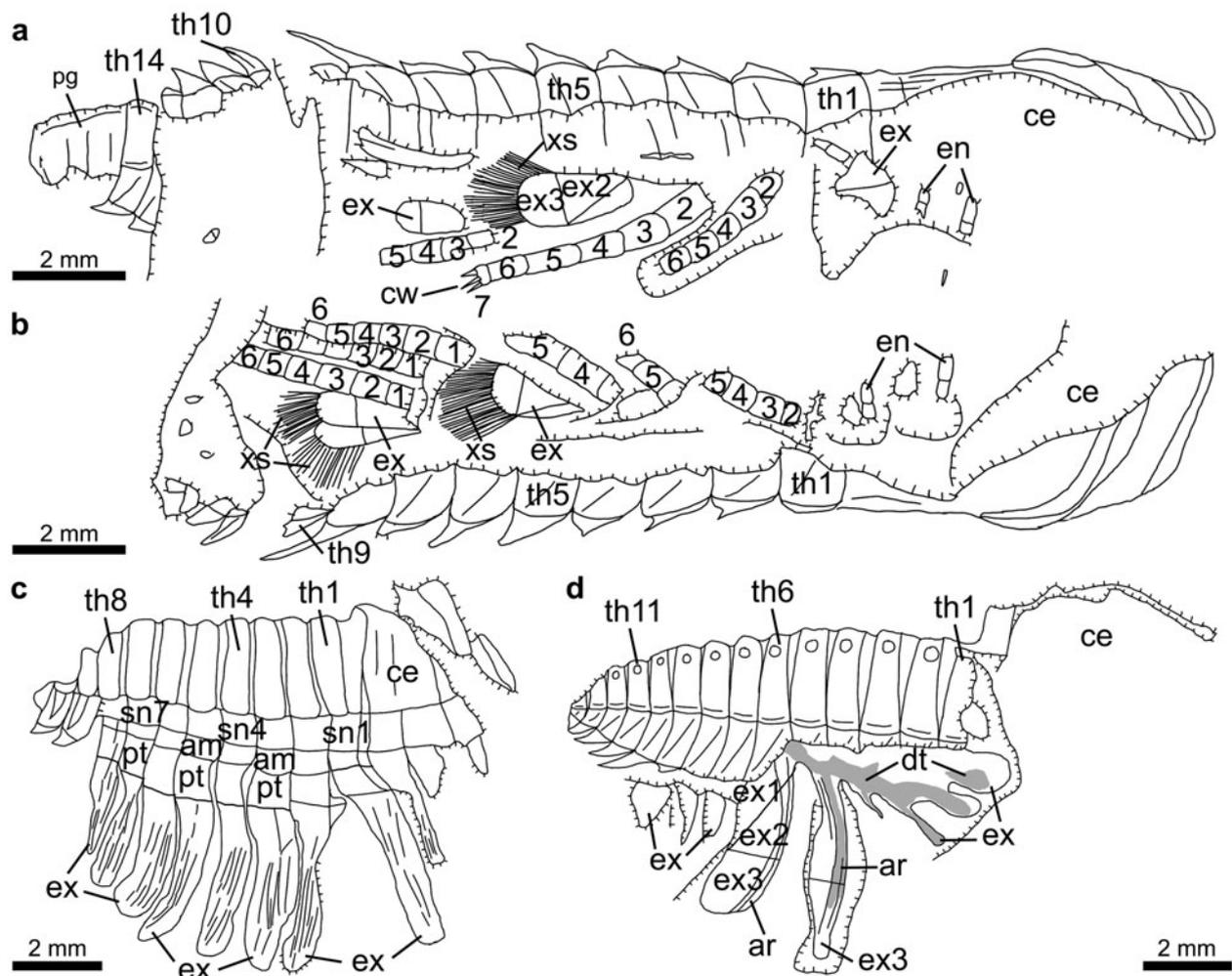


Figure 8. Line drawings of laterally compressed normal-sized holaspids of *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980 preserved with post-antennal biramous limbs, from the Xiaozhuang fossil assemblage, Kunming, Yunnan, SW China. (a) Part, NIGPAS 164507a, as in Figure 7a. (b) Counterpart, NIGPAS 164507b, as in Figure 7b. (c) Part only, NIGPAS 164509, as in Figure 7c. (d) Counterpart, NIGPAS 164511b, as in Figure 7d. Grey areas indicate duct-type soft tissues. Additional abbreviations: ar – anterior section of marginal rim of exopodite; dt – duct-type soft tissues; sn1–sn14 – sternite 1–14.

period, which is similar to the growth of thoracic segments.

6. Comparative anatomy of trilobites and arthropods

Before discussing the evolution of euarthropod limbs and trilobite affinities, it is necessary to make comprehensive anatomical comparisons between *Hongshiyanaspis yiliangensis* and other members of Redlichiida, Trilobita and Arthropoda. Detailed discussions organized by structure are given in the subsections below. The results are summarized in Table 3.

6.a. General arrangement of appendages

The appendages of *Hongshiyanaspis yiliangensis* conform to the basic architecture of polymerid trilobite appendages, which are developed as a pair of uniramous antennae followed by a series of homonomous biramous limbs, one pair at each segment (e.g. Hughes, 2003; Scholtz & Edgecombe, 2005, 2006). The attach-

ment sites of the antennae are close to the lateral margins of the hypostome in *H. yiliangensis*, as shown in other trilobite species (Stürmer & Bergström, 1973; Whittington, 1975, 1993; Whittington & Almond, 1987; Hou *et al.* 2008). The number of pairs of cephalic biramous limbs posterior to the antennae in *H. yiliangensis* is interpreted to be three, which is consistent with the situations in other well-documented trilobite species (e.g. Hughes, 2003), including *Eoredlichia intermedia* (Hou *et al.* 2008), *Olenoides serratus* (Whittington, 1975), *Triarthrus eatoni* (Cisne, 1975, 1981; Whittington & Almond, 1987), *Rhenops cf. anserinus* (Bartels, Briggs & Brassel, 1998) and *Chotecops ferdinandi* (Bruton & Haas, 1999). The claim of four pairs in *Ceraurus pleurexanthemus* (Størmer, 1939) or a fourth pair overlapping the cephalic/thoracic boundary in *Placoparia cambriensis* (Edgecombe & Ramsköld, 1999) requires further research (Hughes, 2003; Scholtz & Edgecombe, 2005).

The thoracic limbs of *H. yiliangensis* show no significant variation in shape, similar to other trilobites (see

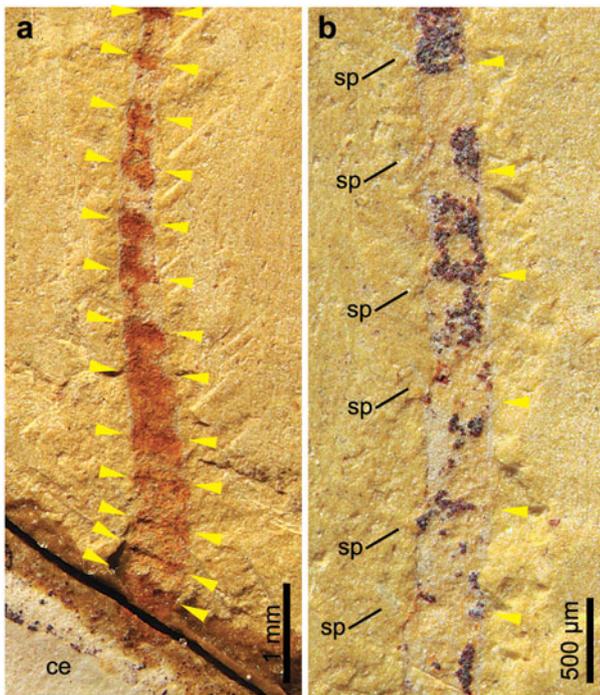


Figure 9. (Colour online) Details of antennae of *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980, from the Xiazhuang fossil assemblage, Kunming, Yunnan, SW China, part only, NIGPAS 164503, as in Figures 1b, 2b. (a) Proximal podomeres of the left antenna. (b) Middle podomeres bearing spines of the right antenna. Yellow arrows indicate arthroal membranes between podomeres. Abbreviations as in Figures 2, 3.

Hughes, 2003, pp. 189–90 for discussions) and arthropods (e.g. Hou & Bergström, 1997; Ortega-Hernández, Legg & Braddy, 2013). Numerous biramous limbs with decreasing sizes are known from subisopygous and isopygous taxa as well as the micropygous *T. eatoni* (see Hughes, 2003, pp. 191–2 for discussion), the number of which considerably exceeds the number of pygidial tergites (Whittington & Almond, 1987). Although *H. yiliangensis* shows a pair of limbs belonging to the only developed axial ring in the pygidium, and two small exopodite flaps are observed extending beyond the posterior margin of pygidium in *E. intermedia* (Hou *et al.* 2008, figs 2D, 5), the number and arrangement of pygidial limbs in the micropygous redlichiids are still uncertain.

6.b. Antennae

Like the antennae known from other trilobites, the antennae of *Hongshiyanaspis yiliangensis* comprise numerous podomeres with spines with putative sensory function (Whittington, 1975, 1993; Bruton & Haas, 1999; Hou *et al.* 2008). The proximal podomeres in *H. yiliangensis* are stouter than the distal podomeres, as seen in other trilobites (Raymond, 1920; Stürmer & Bergström, 1973; Whittington, 1975; Bergström & Brassel, 1984; Whittington & Almond, 1987; Shu

et al. 1995), probably providing a stronger mechanical force in the proximal section of the antenna to create effective swinging for the distal section. The maximum number of podomeres in *H. yiliangensis* antennae is 27, whereas more than 40 are found in *Chotecops ferdinandi* (Bruton & Haas, 1999), 45 in *Eoredlichia intermedia* (Hou *et al.* 2008), ~ 50 in *Olenoides serratus* (Whittington, 1975, fig. 3), > 30 in *Triarthrus eatoni* (Whittington & Almond, 1987) and ~ 30 in *Palaeolenus lantenoisi* (Hu *et al.* 2013). Thus, the numbers of podomeres in antennae are likely to vary among different trilobite species.

6.c. Post-antennal biramous limbs

The post-antennal biramous limbs of *Hongshiyanaspis yiliangensis* are consistent with the basic architecture shown in other polymerid trilobites and arthropods, in having a protopodite composed of a single segment and an endopodite with seven segments composed of six endite-bearing podomeres and a claw-bearing terminal podomere (Fig. 16; Bergström, 1972; Whittington, 1975; Bergström & Brassel, 1984; Whittington & Almond, 1987; Bruton & Haas, 1999; Hou *et al.* 2008), but the exopodite is unique with its tripartite flap composition.

6.c.1. Protopodite and body–limb junction

The coexistence of both clustered and non-clustered spines on the gnathobase of the protopodite of *Hongshiyanaspis yiliangensis* (Fig. 16) suggests that reconstructions of gnathobases in various polymerid trilobites are possibly incomplete when they only show either clustered or non-clustered spines (e.g. Müller & Walossek, 1987, fig. 27). Putative muscle bundles around the arthroal membranes of the protopodite in trilobites have not been reported in former studies and provide new information on the musculature of trilobite limbs. Fine setae that are nicely preserved on the protopodite and probably also on the endopodite are most likely to have a sensory function, as suggested in other euarthropods (e.g. Strausfeld, 2016).

The body–limb junction formed by an arthroal membrane between the protopodite and each thoracic sternite shown in the *Hongshiyanaspis yiliangensis* material supplements the poor record of the body–limb junction in polymerid trilobites. Although the body–limb arthroal membrane has been shown in several Orsten stem-euarthropods (see Haug *et al.* 2013 and references therein), the most similar junction to the arthroal membrane and sternite as in *H. yiliangensis* is best documented in the nektaspid *Misszhouia longicaudata* (Ramsköld *et al.* 1996, fig. 2; Chen, Edgecombe & Ramsköld, 1997, figs 8a, 9a), where curved annulations of arthroal membrane and hourglass-shaped sternites are also visible. No indication for either a second proximal podomere or a proximal endite is present on the protopodite of *H. yiliangensis*.

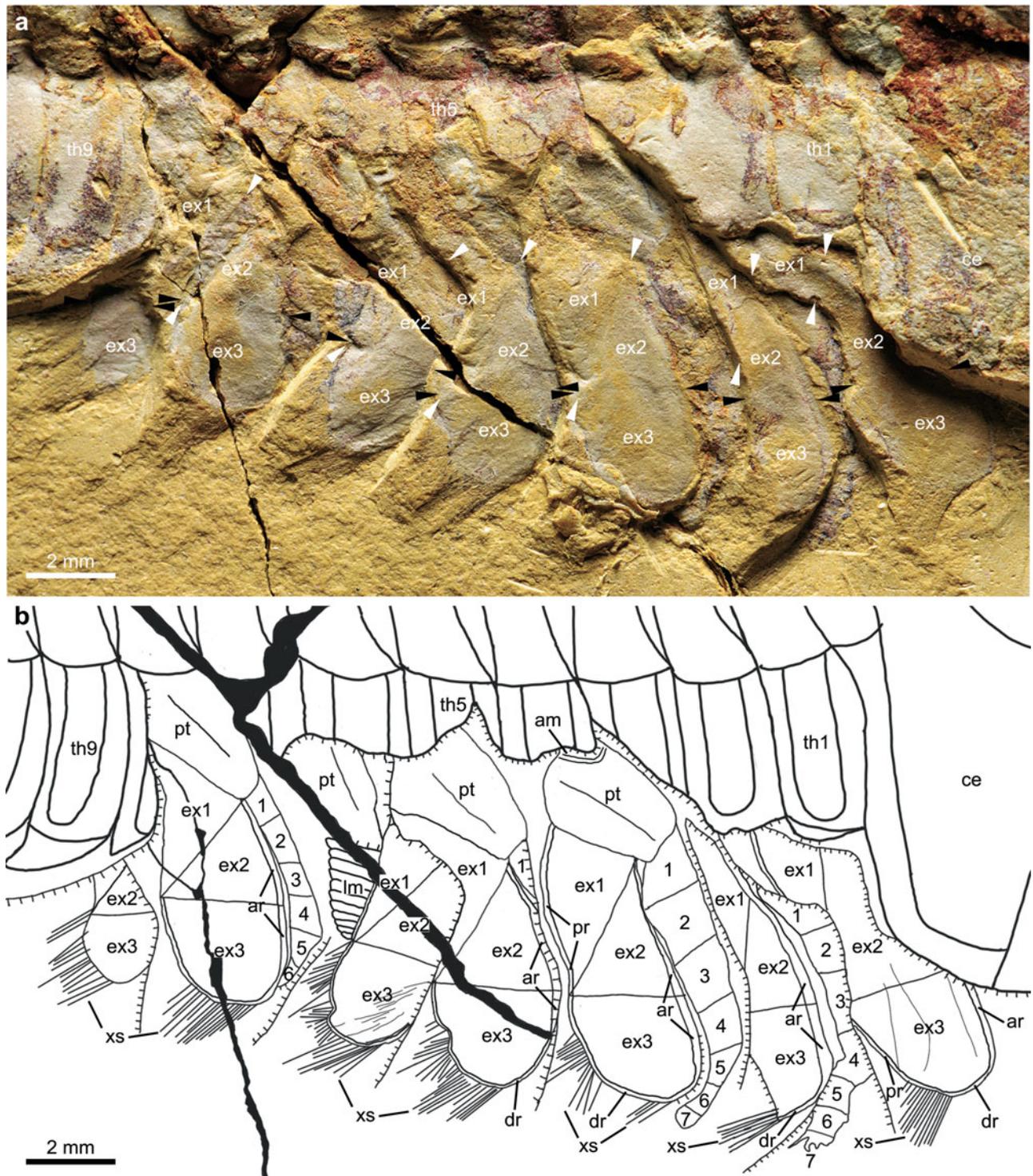


Figure 10. (Colour online) Right thoracic post-antennal biramous limbs of *Hongshiyanspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980, from the Xiaozhuang fossil assemblage, Kunming, Yunnan, SW China, part only, NIGPAS 164503, as in Figures 1b, 2b. (a) Photo. (b) Illustrative line drawing. Black and white arrows indicate the transverse and oblique joints separating the exopodite lobes. Additional abbreviations: dr – distal section of marginal rim of exopodite; pr – posterior section of marginal rim of exopodite.

6.c.2. Endopodite

As in other biramous trilobite limbs, the six proximal podomeres of the endopodite have differentiated shapes, especially in their spinous endites, and show a tendency to taper from the proximal towards the distal end (Fig. 16; Bergström, 1972; Whittington, 1975; Bergström & Brassel, 1984; Whittington &

Almond, 1987; Bruton & Haas, 1999; Hou *et al.* 2008). Rather than simply consisting of claws as indicated in other trilobite species, the seventh and terminal podomere has a short rigid base and is connected to the sixth podomere by an arthrodial membrane. Three distal claws are present, and their morphology and arrangements vary owing to the different angles of burial similar to the Hunsrück Shale species

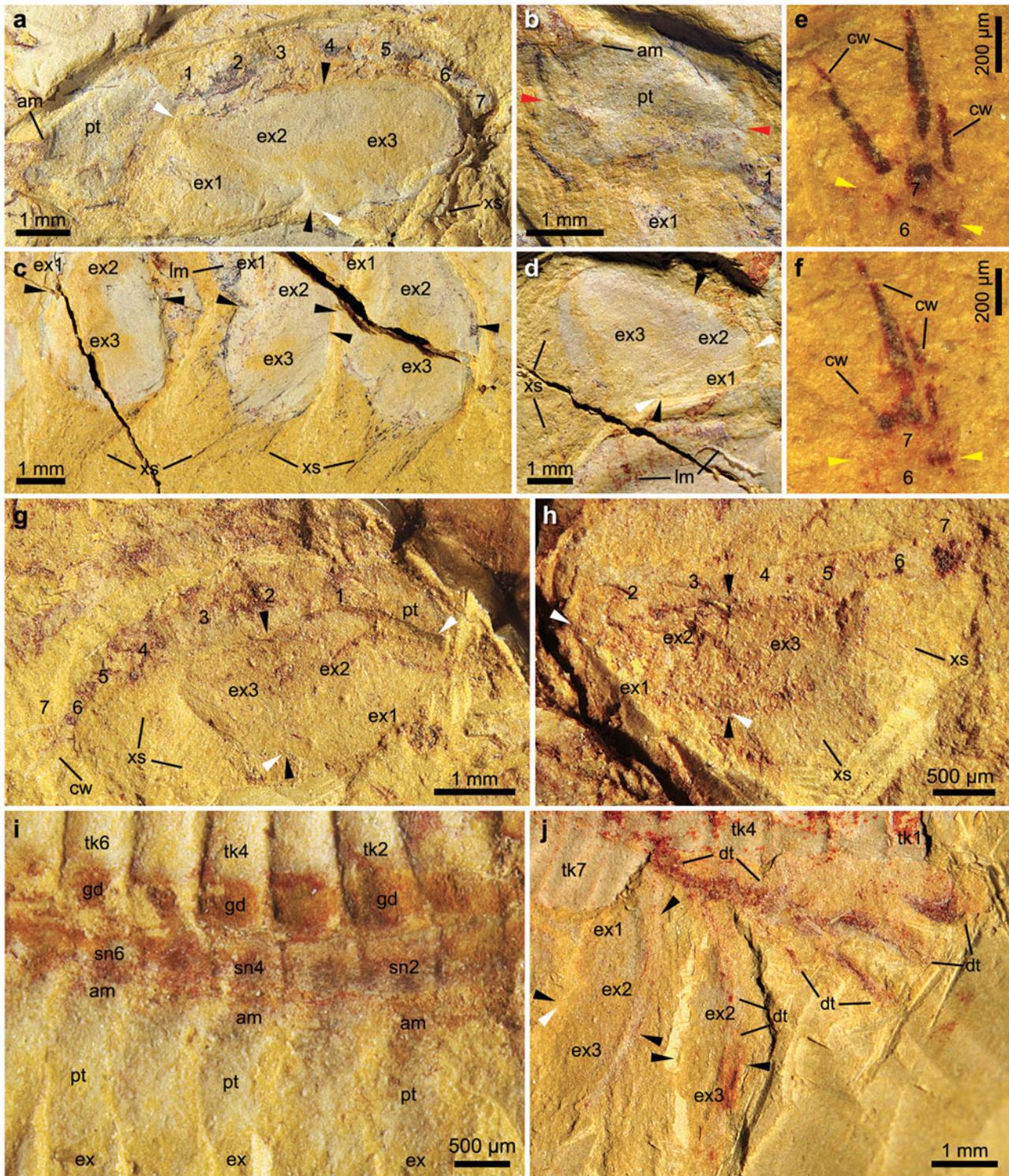


Figure 11. (Colour online) Details of post-antennal biramous limbs of *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980, from the Xiaozhuang fossil assemblage, Kunming, Yunnan, SW China. (a–d) Partial view, NIGPAS 164503. (a, b) Nearly complete fourth dextral thoracic limb, as in Figure 10a. (a) Image showing the protopodite, endopodite and exopodite. (b) Close-up of the protopodite in (a) showing an arthrodial membrane as the body–limb junction and a transverse furrow (red arrows). (c, d) Close-ups of exopodites. (e, f) Distal parts of exopodites of the fifth, sixth and eighth dextral thoracic limbs showing the setae along margins and the transverse joints. (d) Exopodite of the second sinistral thoracic limb, showing reddish imprints of lamellae on the exoskeleton. (e, f) Counterpart, NIGPAS 164506, close-ups of the seventh claw-bearing podomere of the endopodite, showing the arthrodial membrane between the sixth and seventh podomeres of the endopodites (yellow arrows) and the three distal claws, as in the first and second thoracic limbs of Figures 4e, 6a. (g, h) NIGPAS 164505, nearly complete thoracic limbs. (g) Part, NIGPAS 164505a, the third sinistral thoracic limb, as in Figures 1a, 3a. (h) Counterpart, NIGPAS 164505b, the sixth sinistral thoracic limb, as in Figures 1b, 3b. (i) Part only, NIGPAS 164509, close-up of the body–limb junctions, as in Figures 7c, 8c. (j) Counterpart, NIGPAS 164511b, close-up of the exopodites and duct-type soft tissues, as in Figures 7d, 8d. Black and white arrows indicate the transverse and oblique joints separating the exopodite lobes. Red arrows indicate the transverse furrow on the protopodite. Yellow arrows indicate the arthrodial membrane between the sixth and seventh podomeres of the endopodites. Abbreviations as in Figures 1–3, 8.

Table 2. Measurements of antennae and crania of holaspid specimens *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980

Specimen	Ontogenetic stage	Left/right antenna	Length of cephalon (mm)	Measurements of antennae		
				Total length (mm)	Number of podomeres	Average length of podomere (mm)
NIGPAS 164514	Early holaspid	Left	1.85	0.65	6	0.11
NIGPAS 164514	Early holaspid	Right	1.85	2.03	14	0.15
NIGPAS 164504	Normal-sized holaspid	Left	4.85	8.25	25	0.33
NIGPAS 164504	Normal-sized holaspid	Right	4.85	6.92	23	0.30
NIGPAS 164512	Normal-sized holaspid	Left	5.54	2.38	8	0.30
NIGPAS 164512	Normal-sized holaspid	Right	5.54	2.57	9	0.29
NIGPAS 164510	Normal-sized holaspid	Left	5.78	7.33	18	0.41
NIGPAS 164506	Normal-sized holaspid	Left	6.61	4.69	13	0.36
NIGPAS 164503	Fully grown holaspid	Left	9.40	13.39	23	0.58
NIGPAS 164503	Fully grown holaspid	Right	9.40	14.01	27	0.52

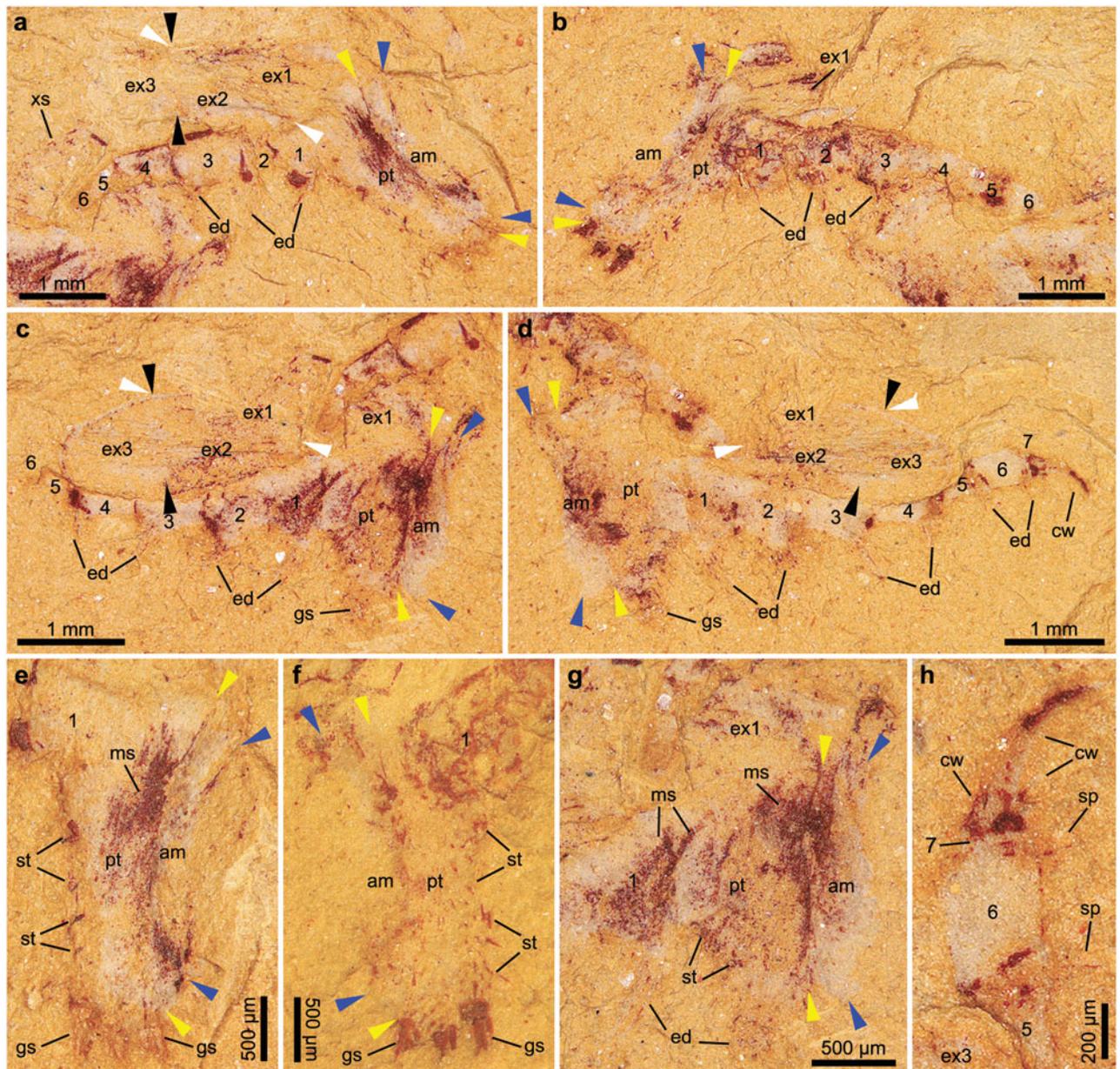


Figure 12. (Colour online) Two disarticulated post-antennal biramous limbs of *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980, from the Xiazhuang fossil assemblage, Kunming, Yunnan, SW China, NIGPAS 164508. (a, b) Limb A as part and counterpart, respectively. (c, d) Limb B as part and counterpart, respectively. (e, f) Close-ups of the protopodite of Limb A, as in (a) and (b), respectively. (g) Close-up of the protopodite of Limb B, as in (c). (h) Close-up of the distal podomeres of Limb B, as in (d). Blue and yellow arrows indicate the proximal and distal boundaries of the arthrodial membrane. Black and white arrows indicate the transverse and oblique joints separating the exopodite lobes. Abbreviations as in Figures 1–3, 8.

Table 3. Comparative anatomy between *Hongshiyanaspis yiliangensis* and other redlichiids, trilobites and artiopods

Body parts	Characters	<i>Hongshiyanaspis yiliangensis</i>	Redlichiida	Trilobita	Artiopoda
Arrangement of appendages	Overall morphology	Deutocerebral/hypostomal antennae and homonomous post-antennal limbs	Same	Same; unique paired cerci in <i>Olenoides</i>	Same
	Cephalon	Three pairs of biramous limbs	Same	Mostly three pairs; the claims of four pairs in <i>Ceraurus</i> and <i>Placoparia</i> are questionable	Mostly three pairs; those interpreted with more are open to question
	Thorax (trilobites) or trunk (artiopods)	One pair of biramous limbs per thoracic tergites	Same	Likely same; unknown for those with differentiated thorax	Various correspondences between numbers of limbs and tergites, extreme cases in those with fusion of trunk tergites, e.g. naraoiids and tegopeltid
	Pygidium (trilobites only)	Micropygous, one pair of biramous limbs in axial ring, multiple pairs in terminal axial piece	Same	Number of limbs almost doubles the number of pygidial tergites in <i>Triarthrus</i> ; unknown in others	
Antennae	Rami	Uniramous	Same	Same	Same
	Numbers of podomeres	Maximum ≥ 27	45 in <i>Eoredlichia</i>	Various	Various
	Shapes of podomeres	Thick in proximal, slender in distal	Same	Same	Mostly same; not obvious in some species
Post-antennal limbs	Spines	No less than one per podomere	Same	Multiple in <i>Olenoides</i>	Various
	Rami	Biramous	Same	Same	Same
	Body–limb junction	Arthrodistal membrane attached to sternite	Unknown	Unknown	Same in <i>Misszhouia</i> , <i>Sidneyia</i> ; unknown in others
Protopodite	Podomere(s)	Single, no differentiation	Same	Same	Same; except for the prebasal endite in <i>Sidneyia</i>
	Gnathobase	Clustered and non-clustered spines	Non-clustered only	Likely both types	Various
	Transverse furrow(s)	One	Unknown	Unknown; rejected in <i>Triarthrus</i>	Unknown
Endopodite	Musculature	Present across arthrodistal membrane	Unknown	Unknown	Unknown
	Setae	Sensory setae	Unknown	Putative sensory setae in <i>Olenoides</i>	Unknown in most species
	Numbers of podomeres	Seven	Same	Same	Questionable for those are not seven
	Shapes of podomeres	Differentiated	Same	Same	Same
	Endites	Spinous	Same	Same	Mostly same, a few can be smooth
	Claws	Three rooted on a very short podomere	Three	Three	Likely various
	Musculature	Oblique muscle bundles	Unknown	Unknown	Unknown
Exopodite	Setae/spines	Sensory setae	Unknown	Lateral spines in <i>Olenoides</i>	Mostly unknown, likely various
	Composition	A flap of three lobes	A flap of two lobes	Lobes or shafts with various numbers	Lobes or shafts with various numbers
	Numbers of joints	One transverse and one oblique	One transverse	Various	Various
	Lamellae	Imbricate	Same	Likely same if present	Likely same if present
	Distal setae	Non-overlapping	Same	Likely same if present	Likely same if present
	Marginal rims	Present on lobes	Unknown	Unknown	Unknown
	Duct-type structures	Present, merge into a main stem	Unknown	Unknown	Unknown

Note: the specific names of species are omitted for concision. Situations labelled as ‘same’ are compared with the adjacent column on the left side. The Redlichiida is represented by *Eoredlichia intermedia* (Hou *et al.* 2008; Shu *et al.* 1995). The Trilobita represent polymerid trilobites, excluding agnostids. See text for detailed discussions and references.

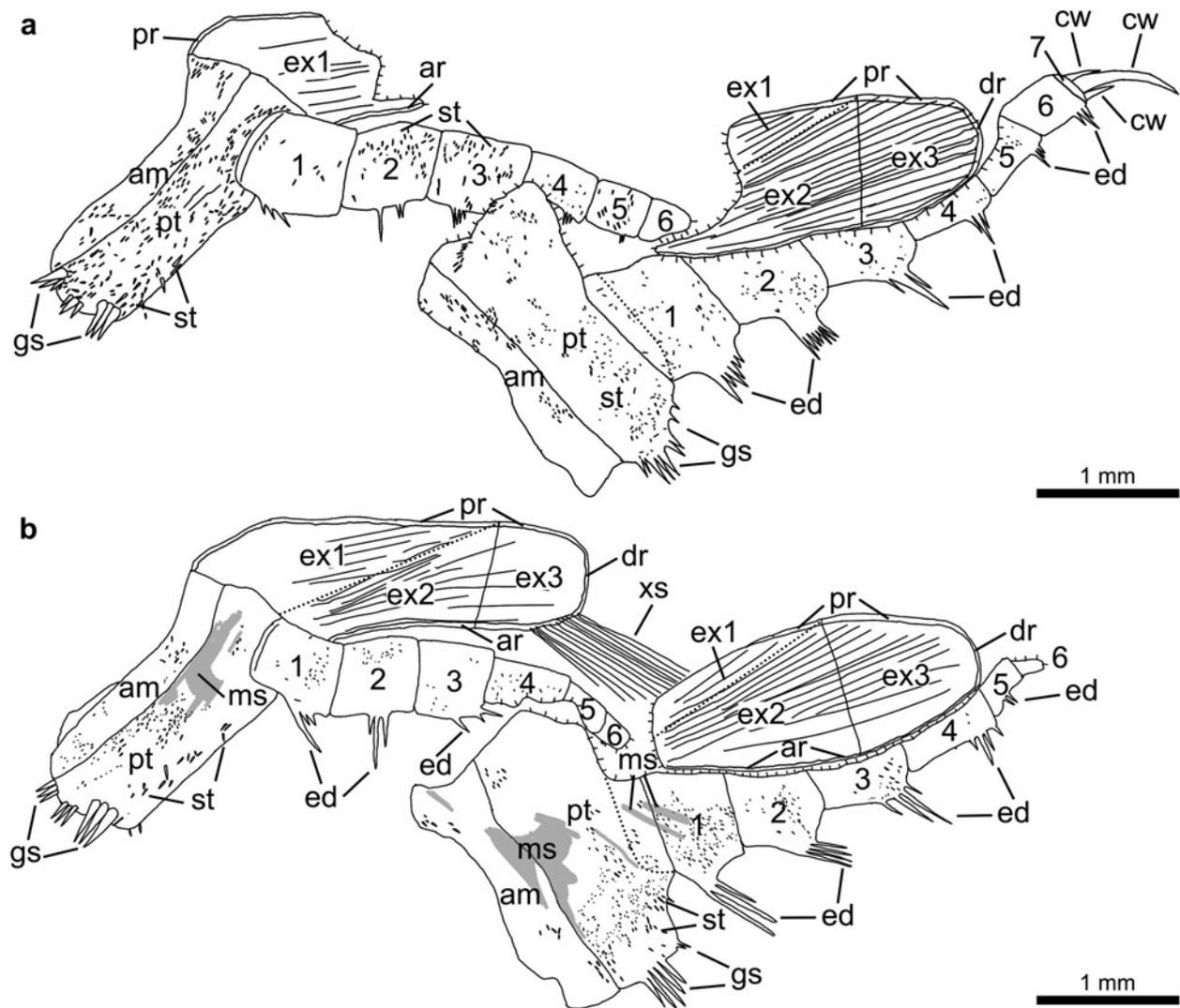


Figure 13. Interpretative line drawings of two disarticulated post-antennal biramous limbs of *Hongshiyanaspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980, from the Xiaozhuang fossil assemblage, Kunming, Yunnan, SW China, NIGPAS 164508. (a) Counterpart, NIGPAS 164508a, as in Figures 12b, d. (b) Part, NIGPAS 164508b, as in Figures 12a, c, but horizontally flipped. Abbreviations as in Figures 1–3, 8.

Chotecops ferdinandi (Bruton & Haas, 1999, text-fig. 19), whereas the three claws clearly do not merge together into a common base in *H. yiliangensis*. Taking into account the pyritized preservation, the common base of the claws shown in *C. ferdinandi* (Bruton & Haas, 1999, text-fig. 19) may be a taphonomic artefact.

6.c.3. Exopodite

Exopodites have been previously reported from several polymerid trilobites (Størmer, 1939, 1951; Bergström, 1972; Whittington, 1975; Whittington & Almond, 1987; Bruton & Haas, 1999; Hou *et al.* 2008). The morphology of the exopodite in *Hongshiyanaspis yiliangensis* is unique in possessing a tripartite flattened flap (Fig. 16). Although tripartite exopodite flaps are also found in several other artiopods, such as *Kua-maia lata* (Hou & Bergström, 1997), *Sidneyia inexpectans* (Stein, 2013), *Emeraldella brocki* (Stein &

Selden, 2012) and *Arthroaspis bergstroemi* (Stein *et al.* 2013), the flap in *H. yiliangensis* differs in having a transverse and an oblique joint, no sharp discontinuity of the margin at the endpoints of the joints, and lobes with subequal widths. The exopodite flap in *H. yiliangensis* is almost double the maximum width of the endopodite, which is distinct from those described from almost all other lobate exopodites of trilobites (Whittington, 1975; Bruton & Haas, 1999; Hou *et al.* 2008) but comparable to those in *K. lata* (Hou & Bergström, 1997), *Squamacula clypeata* (Zhang *et al.* 2004), *Kwan-yinaspis maotianshanensis* (Zhang & Shu, 2005) and *Naraoia spinosa* (Zhang, Shu & Erwin, 2007) (also see Ortega-Hernández, Legg & Braddy, 2013, fig. 4 for reconstructions). These wide exopodite flaps in *H. yiliangensis* may be an adaptation to powerful swimming and/or a more effective respiration, with the joints possibly directing water currents. Non-overlapping setae and imbricate lamellae develop along the margins of

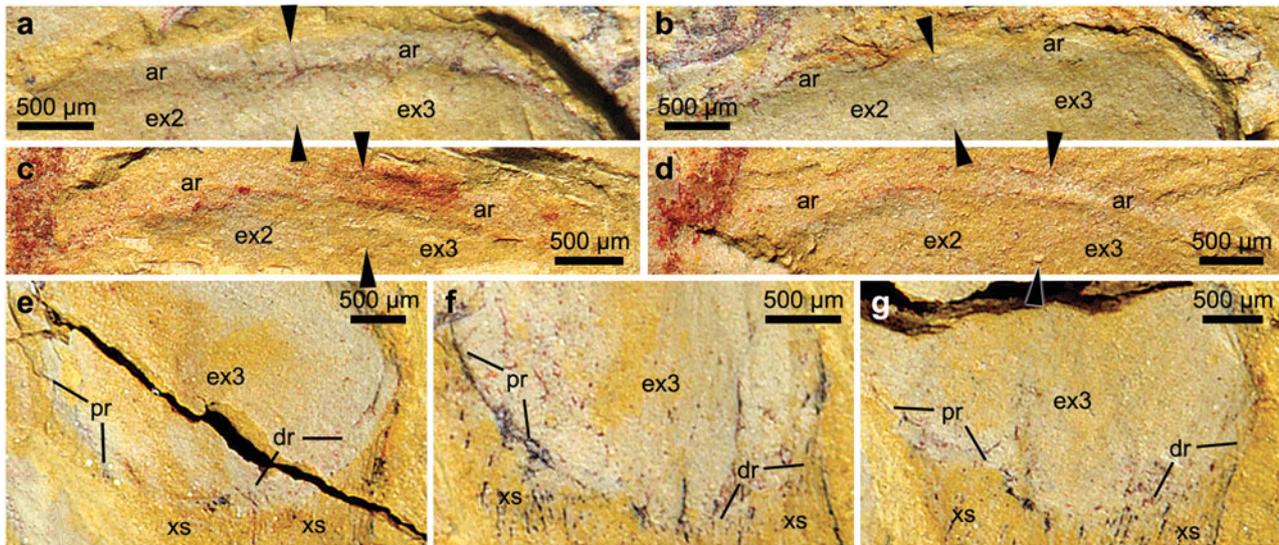


Figure 14. (Colour online) Marginal areas in the exopodites of post-antennal biramous limbs of *Hongshiyanspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980, from the Xiaozhuang fossil assemblage, Kunming, Yunnan, SW China. (a, b, e–g) NIGPAS 164503. (a, b) Anterior margins of the exopodites in the third and fourth thoracic limbs, respectively. (e–g) Distal and posterior margins of the exopodites in the eighth, sixth and fifth thoracic limbs, respectively. (c, d) NIGAPS 164511b, anterior margins of the exopodites in the fifth and sixth thoracic limbs. Black arrows indicate the transverse joints separating the exopodite lobes. Abbreviations as in Figures 1–3, 8, 10.

the distal (= third) and proximal (= first) lobes of the exopodite flap in *H. yiliangensis*, respectively. This situation is present not only in other polymerid trilobites such as the Cambrian Stage 3 *Eoredlichia intermedia* (Hou *et al.* 2008) and the Cambrian Stage 5 *Olenoides serratus* (Whittington, 1975), but also in a number of non-trilobite arthropods (see Zhang, Shu & Erwin, 2007; Ortega-Hernández, Legg & Braddy, 2013; Stein *et al.* 2013 and references therein). Despite the limited knowledge, the overall similarity between the exopodites of *O. serratus* (Order Corynexochida; Whittington, 1975) and *E. intermedia* (Order Redlichiida; Hou *et al.* 2008) as well as the differences between the almost contemporaneous *E. intermedia* and *H. yiliangensis* (Order Redlichiida) suggests that these anatomical variations are more likely the results of divergence owing to different ecological adaptations of the species rather than the different evolutionary tendencies of trilobite orders.

The exopodite flaps in *Hongshiyanspis yiliangensis* exhibit duct-type soft tissues (Fig. 16). These tissues of the different limbs further merge into a main stem in the ventral soft parts of the body. The morphology and positions of these duct-type structures are comparable to those ‘invasive caeca’ or ‘triangular strips’ in a range of stem-group euarthropods from the Burgess Shale (see Aria & Caron, 2015, figs 2, 3, 5, 9). A digestive nature was suggested for those from the Burgess Shale because they are connected to the main alimentary canal (Aria & Caron, 2015). However, the duct-type structures here may be different tissues, since no connection to the gut is observable. Because exopodites have long been interpreted as respiratory organs like gills (e.g. Hou & Bergström, 1997), a putative circulatory nature for these duct-type structures is

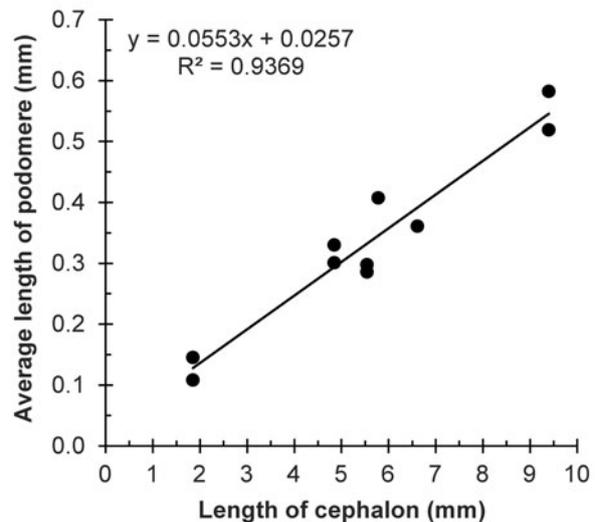


Figure 15. Scatter plot showing significant positive correlation between the length of the cephalon and the average length of the podomeres of the antenna in *Hongshiyanspis yiliangensis* Zhang & Lin in Zhang *et al.* 1980 holaspids from the Xiaozhuang fossil assemblage, Kunming, Yunnan, SW China.

proposed here. Still, further investigations are required to clarify these alternative possibilities.

6.d. Basic appendage morphology of trilobites and arthropods

By discussing comparative anatomy under the light of the new *Hongshiyanspis yiliangensis* appendages above, we can conclude in general that all trilobites and other arthropods share the same basic architecture of appendages comprising a pair of deutocerebral/

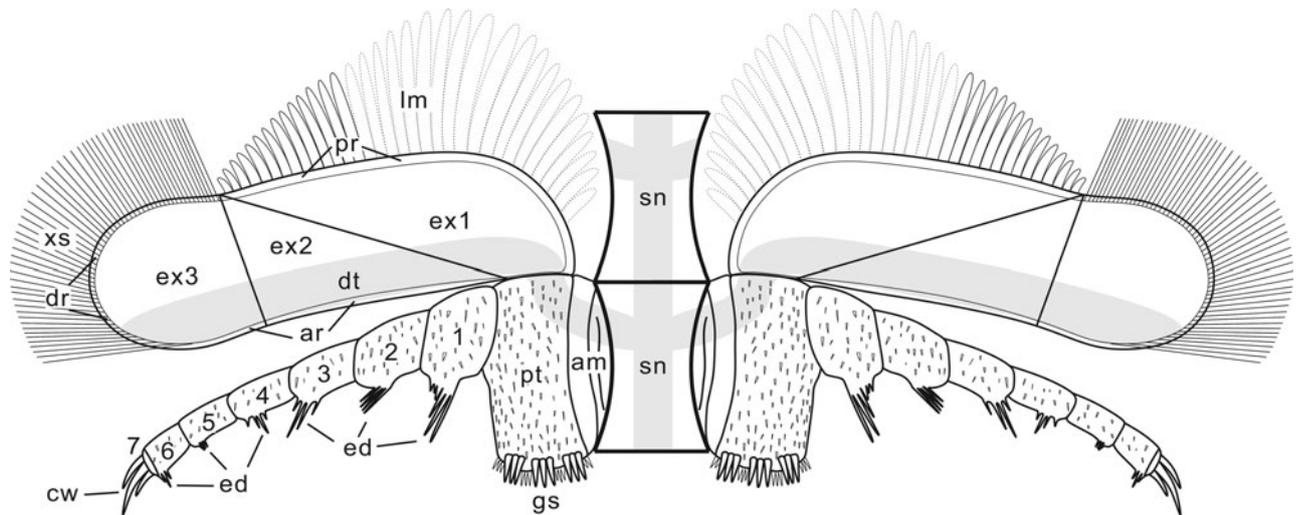


Figure 16. Reconstruction of paired post-antennal biramous limbs connected to sternites. Note that the structures are not in actual life positions, including the orientations of the protopodite–exopodite and body–limb junctions, but are flattened to exhibit the most detailed limb anatomy and their connections, as in the mainstream reconstructions of artiopodan limbs. The proximal lamellae are hypothetical and drawn as dashed lines, and the duct-type soft tissues are in grey. Abbreviations as in Figures 1–3, 8, 10.

hypostomal uniramous antennae and a series of paired homonomous biramous limbs (e.g. Hughes, 2003; Scholtz & Edgecombe, 2005, 2006). However, within ascending systematic hierarchies from Redlichiida, Trilobita to Artiopoda, their biramous limbs show a conserved morphology by consisting of seven podomeres in the endopodites and one single segment in the protopodite, but also considerable morphological disparity in the composition of the exopodites.

7. Affinities of trilobites: mandibulate or chelicerate?

Three alternative affinities for trilobites and other artiopods have been hypothesized: as stem-chelicerates, stem-mandibulates or a stem lineage of both Chelicerata and Mandibulata (Budd & Telford, 2009). Comparisons between major upper stem-group euarthropods and the stem and crown groups of Mandibulata and Chelicerata can reveal general evolutionary trends of euarthropod appendages in two aspects, i.e. the arrangement of appendages along the anterior–posterior main body axis, and the composition of limb rami. These trends can be essential for interpreting the affinities of trilobites and artiopods, which are discussed below and summarized in Table 4.

7.a. Arrangement of appendages

Considering the anteriormost (deutocerebral and tritocerebral) appendages, within upper stem-group euarthropods, trilobites and other artiopods are unique in lacking any specialized cephalic feeding appendages (Table 4), such as the tritocerebral specialized post-antennal appendages (SPAs) in large ‘bivalved’ stem-euarthropods (e.g. Legg *et al.* 2012) and fuxianhuiids (e.g. Yang *et al.* 2013), and the deutocerebral short great appendages (SGAs) in megacheirans (e.g. Tanaka *et al.* 2013). Nevertheless, they share the

multi-segmented deutocerebral ‘secondary antennae’ with large ‘bivalved’ stem-euarthropods (e.g. Legg *et al.* 2012), fuxianhuiids (e.g. Yang *et al.* 2013) and mandibulates (Scholtz & Edgecombe, 2005, 2006). In megacheirans and chelicerates, however, neither the SGAs nor the chelicerae are antenniform but raptorial, and consist of a limited number of podomeres (Chen, Waloszek & Maas, 2004). Therefore, the ‘secondary antennae’ have been regarded as strong evidence supporting the mandibulate affinities of trilobites and other artiopods (Scholtz & Edgecombe, 2005, 2006).

On other hand, all post-tritocerebral limbs are homonomous on both the cephalon and trunk of all the upper stem-group euarthropods mentioned above (e.g. Legg *et al.* 2012; Ortega-Hernández, Legg & Braddy, 2013; Yang *et al.* 2013; Aria, Caron & Gaines, 2015). In Mandibulata, although several anterior post-tritocerebral limbs are specialized for feeding and the rest can show various degrees of differentiation or tagmatization, continuous series of undifferentiated homonomous limbs are typically present on the trunk of myriapods and some crustaceans such as Anostraca and Remipedia. By contrast, in Chelicerata (except for pycnogonids), post-tritocerebral limbs on the anterior and posterior body tagma (prosoma and opisthosoma) are always differentiated. In Xiphosura, the post-tritocerebral limbs of the anterior and posterior tagma are differentiated into uniramous legs (except for the last leg with flabellum) and gills, respectively, whereas the posterior limbs have been reduced in other chelicerate crown groups. Therefore, the homonomous pattern of limbs shown in trilobites, artiopods and other upper stem-group euarthropods has been retained in Mandibulata, but was lost by the basic anterior and posterior tagmatization in Chelicerata, providing further evidence supporting the mandibulate concept.

Table 4. General comparisons of appendage arrangements and compositions of biramous limbs between upper stem groups of euarthropods, mandibulates (represented by crustaceans) and chelicerates

Euarthropod groups	Deuto- and trito-cerebral appendages	Post-tritocerebral limbs	Protopodites	Endopodites	Exopodites
Large 'bivalved' stem-euarthropods and fuxianhuidis	Antennae, SPAs	Homonomous	Single podomere	No less than ten podomeres, undifferentiated	One lobe
Megacheirans	Short great appendages, biramous limbs	Homonomous	Single podomere	Seven to nine podomeres, undifferentiated	Two lobes
Artiopods, including trilobites	Antennae, biramous limbs	Homonomous, excluding cerci	Single podomere	Seven podomeres, differentiated	One to three lobes, or one shaft
Stem- and crown-group mandibulates	(First) antennae, second antennae or absent	Homonomous, slightly differentiated, highly differentiated or specialized	At least one podomere	Generally no more than seven podomeres, mostly differentiated	Various numbers of lobes, or one shaft
Stem- and crown-group chelicerates	Chelicerae, pedipalps	Differentiated between anterior and posterior tagma, except for pycnogonids with possibly reduced segments	At least one podomere	Generally no more than seven podomeres, differentiated	?Gills in <i>Dibasterium</i> , <i>Offacolus</i> , and posterior appendages of xiphosurids

Note: in crown-group euarthropods, it is difficult to distinguish the protopodite and endopodite in post-antennal uniramous limbs, and that secondary resegmentation may generate more than seven podomeres in endopodites (Boxshall, 2004, 2013). See Briggs *et al.* (2012) for anatomy of *Dibasterium* and revisions on *Offacolus*. Also note that whether chelicerate gills are epipodites or exopodites is open to question. See text for detailed discussions and references.

7.b. Biramous limbs

Trilobites and other artiopods are also distinguished by the composition of their post-antennal biramous limbs. Despite that all of these limbs in the upper stem groups of Euarthropoda mentioned above are homonomous, the basic compositions of their endopodites and exopodites are different (Table 4). In fuxianhuidis (e.g. Yang *et al.* 2013) and large 'bivalved' stem-euarthropods (e.g. Legg *et al.* 2012), the podomeres of the endopodites show no differentiation in shape and their number exceeds ten, and their exopodites are undivided flaps. In megacheirans (e.g. Aria, Caron & Gaines, 2015), endopodite podomeres are also undifferentiated and are at least eight in number (including the distal claw), whereas their exopodites are flaps comprising two lobes. It is worth noting that the morphology of the exopodites is consistent in each of these groups, whereas the number of their endopodite podomeres varies. This indicates that the exopodites are more morphologically conserved than the endopodites in these upper stem-euarthropod groups. In trilobites and other artiopods, however, endopodites and exopodites exhibit different patterns of morphological disparity. For the endopodites, the number of podomeres is consistently seven (including distal claws) and the morphological differences of the podomeres are geometric rather than qualitative in trilobites and other artiopods with a few questionable exceptions. By contrast, the exopodites vary in composition between any two trilobite and artiopodan species. Their exopodites can be a flap comprising one to three lobes, or a shaft, showing a high morphological disparity. Within the two modern euarthropod

lineages Mandibulata and Chelicerata, this similar disparity of the exopodites is only exhibited in stem-group and crown-group crustaceans, whereas the exopodites have been reduced or lost in Chelicerata.

Developmental biology has shown that endopodites and exopodites are rami originating from the same main proximal–distal axis of the biramous limb bud (Wolff & Scholtz, 2008). However, the striking distinction in patterns of disparity between the endopodites and exopodites seen in stem-euarthropods suggests that their developmental regulatory machineries diverged early in the evolution of euarthropod biramous limbs (Davidson & Erwin, 2006). In trilobites, other artiopods and crustaceans, the developmental genetic regulatory programmes were relatively conserved in the endopodites, whereas they were much more flexible in the exopodites. Nevertheless, the situations are reversed in large 'bivalved' stem-euarthropods, fuxianhuidis and megacheirans, with higher conservativeness in the exopodites than in the endopodites. The chelicerate concept would thus require abandoning the highly evolved developmental machinery of the exopodites, which is an unlikely case.

Therefore, from a developmental perspective, the pattern of morphological disparity shown by the two rami also supports the mandibulate affinities of trilobites and other artiopods.

7.c. Evolutionary trends of euarthropod appendages

Most recent morphological and molecular phylogenetic frameworks of Euarthropoda have, respectively, revealed a successive appearance of euarthropod

characters in upper stem groups from ‘bivalved’ stem-euarthropods, fuxianhuids, megacheirans to artiopods, and the relationships of crown groups such as Chelicerata + Mandibulata (Myriapoda + Tetraconata/Pancrustacea) (e.g. Edgecombe & Legg, 2014). Within the upper stem groups, the early evolution of euarthropod appendages has undergone the origination of deutocerebral (antennae, SGAs) and tritocerebral (SPAs) specialized appendages (Edgecombe & Legg, 2014), reduction in the number of endopodite podomeres from more than ten to the ground plan of seven, and differentiation of endopodite podomeres (Boxshall, 2004, 2013). However, the subdivision of the protopodite, and the differentiation or specialization of post-tritocerebral or trunk appendages should occur during the early evolution of Mandibulata and Chelicerata (Boxshall, 2004, 2013).

7.d. Affinities of trilobites and artiopods

The chelicerate affinity hypothesis for trilobites and artiopods had been the mainstream for a long time until new evidence for mandibulate affinities and conflicts with the chelicerate concept were put forward by Scholtz & Edgecombe (2005, 2006). However, controversies have still been going on since then, as both the mandibulate and chelicerate concepts gained support from different recent phylogenetic analyses (e.g. Ortega-Hernández, Legg & Braddy, 2013; Legg, Sutton & Edgecombe, 2013). The evidence given by Scholtz & Edgecombe (2005, 2006), including the ‘second antennae’ and head segmentation, together with our new arguments based on the homonomous pattern and composition of the biramous limbs, supports the mandibulate concept of trilobites and at least some artiopods. Meanwhile, this evidence proposes the critical character transformations that are required to fit the chelicerate concept, including the loss of antennae, disappearance of delimited cephalon–trunk tagmatization, change of limb patterning along the main body axis and reorganization of developmental machineries in the limb rami. However, we cannot negate the possibility that with new data, some members of the current Artiopoda definition may be closer to the chelicerate lineage. If this is true, subdivision of artiopods would be essential.

8. Conclusion

The appendages of *Hongshiyanaspis yiliangensis* reinforce the idea that polymerid trilobites share a homonomous arrangement of biramous limbs as well as conserved anatomy in the protopodites and endopodites, but have significant inter-taxa differences in the exopodites. This appendage architecture of trilobites is highly comparable to that of other artiopods. Ontogeny of trilobite antennae is studied for the first time and shows a growth model of lengthening each podomere. By reinvestigating and comparing appendages in upper stem groups and crown groups of Eu-

arthropoda, we show similarities in the arrangement of homonomous limbs and patterns of morphological disparity in the endopodites and exopodites between artiopods (including trilobites) and mandibulates. Together with the shared ‘secondary antennae’ and head tagmosis, these new lines of evidence further support the mandibulate affinities of trilobites and at least some artiopods. However, more data on the appendages of trilobites and other stem-group euarthropods are still essential to resolve controversies surrounding the problem of trilobite affinities.

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Supplementary material

To view supplementary material for this article, please visit <https://doi.org/10.1017/S0016756817000279>

References

- ARIA, C. & CARON, J.-B. 2015. Cephalic and limb anatomy of a new isoxyid from the Burgess Shale and the role of “stem bivalved arthropods” in the disparity of the frontalmost appendage. *PLoS ONE* **10**, e0124979.
- ARIA, C., CARON, J.-B. & GAINES, R. 2015. A large new leanchoilid from the Burgess Shale and the influence of inapplicable states on stem arthropod phylogeny. *Palaentology* **58**, 629–60.
- BARTELS, C., BRIGGS, D. E. G. & BRASSEL, G. 1998. *The Fossils of the Hunsrück Slate: Marine Life in the Devonian*. Cambridge: Cambridge University Press, 324 pp.
- BEECHER, C. E. 1895. Structure and appendages of *Trinucleus*. *American Journal of Science* **3**, 307–11.
- BERGSTRÖM, J. 1972. Appendage morphology of the trilobite *Cryptolithus* and its implications. *Lethaia* **5**, 85–94.
- BERGSTRÖM, J. & BRASSEL, G. 1984. Legs in the trilobite *Rhenops* from the Lower Devonian Hunsrück Slate. *Lethaia* **17**, 67–72.
- BILLINGS, E. 1870. Notes on some specimens of Lower Silurian trilobites. *Quarterly Journal of the Geological Society* **26**, 479–86.
- BOXSHALL, G. A. 2004. The evolution of arthropod limbs. *Biological Reviews* **79**, 253–300.
- BOXSHALL, G. A. 2013. Arthropod limbs and their development. In *Arthropod Biology and Evolution: Molecules, Development, Morphology* (eds A. Minelli, G. A. Boxshall & G. Fusco), pp. 241–67. Berlin, Heidelberg: Springer.
- BRIGGS, D. E. G., LIEBERMAN, B. S., HENDRICKS, J. R., HALGEDAHL, S. L. & JARRARD, R. D. 2008. Middle Cambrian arthropods from Utah. *Journal of Paleontology* **82**, 238–54.
- BRIGGS, D. E. G., SIVETER, D. J., SIVETER, D. J., SUTTON, M. D., GARWOOD, R. J. & LEGG, D. A. 2012. Silurian

- horseshoe crab illuminates the evolution of arthropod limbs. *Proceedings of the National Academy of Sciences* **109**, 15702–5.
- BRUTON, D. L. & HAAS, W. 1999. The anatomy and functional morphology of *Phacops* (Trilobita) from the Hunsrück Slate (Devonian). *Palaeontographica Abteilung A* **253**, 29–75.
- BUDD, G. E. & TELFORD, M. J. 2009. The origin and evolution of arthropods. *Nature* **457**, 812–17.
- CHEN, J. Y., EDGEcombe, G. D. & RAMSKÖLD, L. 1997. Morphological and ecological disparity in naraoiids (Arthropoda) from the Early Cambrian Chengjiang fauna, China. *Records of the South Australian Museum* **49**, 1–24.
- CHEN, J. Y., WALOSZEK, D. & MAAS, A. 2004. A new ‘great appendage’ arthropod from the Lower Cambrian of China and homology of chelicerate chelicerae and raptorial antero ventral appendages. *Lethaia* **37**, 3–20.
- CHEN, J. Y. & ZHOU, G. Q. 1997. Biology of the Chengjiang fauna. *Bulletin of the National Museum of Natural Science* **10**, 11–105.
- CISNE, J. L. 1975. Anatomy of *Triarthrus* and the relationships of the Trilobita. *Fossils and Strata* **4**, 45–63.
- CISNE, J. L. 1981. *Triarthrus eatoni* (Trilobita), anatomy of its exoskeletal, skeletomuscular, and digestive systems. *Palaeontographia Americana* **9**, 99–142.
- DAVIDSON, E. H. & ERWIN, D. H. 2006. Gene regulatory networks and the evolution of animal body plans. *Science* **311**, 796–800.
- DUNBAR, C. O. 1925. Antennae in *Olenellus getzi* n. sp. *American Journal of Science* **5**, 303–8.
- EDGEcombe, G. D. & LEGG, D. A. 2014. Origins and early evolution of arthropods. *Palaeontology* **57**, 457–68.
- EDGEcombe, G. D. & RAMSKÖLD, L. 1999. Relationships of Cambrian Arachnata and the systematic position of Trilobita. *Journal of Paleontology* **73**, 263–87.
- FARRELL, Ú. C., MARTIN, M. J., HAGADORN, J. W., WHITELEY, T. & BRIGGS, D. E. G. 2009. Beyond Beecher’s Trilobite Bed: widespread pyritization of soft tissues in the Late Ordovician Taconic foreland basin. *Geology* **37**, 907–10.
- FORTEY, R. A. 2001. Trilobite systematics: the last 75 years. *Journal of Paleontology* **75**, 1141–51.
- GEYER, G. & PEEL, J. S. 2017. Middle Cambrian trilobites from the Ekspedition Bræ Formation of North Greenland, with a reappraisal of the genus *Elrathina*. *Journal of Paleontology* **91**, 265–93.
- HARRINGTON, H. J., HENNINGSMOEN, G., HOWELL, B. F., JAANUSSON, V., LOCHMAN-BALK, C., MOORE, R. C., POULSEN, C., RASETTI, F., RICHTER, E., RICHTER, R., SCHMIDT, H., SDZUY, K., STRUVE, W., STØRMER, L., STUBBLEFIELD, C. J., TRIPP, R., WELLER, J. M. & WHITTINGTON, H. B. 1959. *Part O; Arthropoda 1*. Boulder, Colorado: Geological Society of America; and Lawrence, Kansas: University of Kansas Press, 560 pp.
- HAUG, J. T., MAAS, A., HAUG, C. & WALOSZEK, D. 2013. Evolution of crustacean appendages. In *The Natural History of the Crustacea, Volume 1: Functional Morphology and Diversity* (eds L. Watling & M. Thiel), pp. 34–73. New York: Oxford University Press.
- HOLLINGSWORTH, J. S. 2008. The first trilobites in Laurentia and elsewhere. In *Advances in Trilobite Research* (eds I. Rábano, R. Gozalo & D. García-Bellido), pp. 171–75. Madrid: Instituto Geológico y Minero de España.
- HOU, X. G. & BERGSTRÖM, J. 1997. Arthropods of the lower Cambrian Chengjiang fauna, southwest China. *Fossils and Strata* **45**, 1–116.
- HOU, X. G., CLARKSON, E. N. K., YANG, J., ZHANG, X. G., WU, G. Q. & YUAN, Z. B. 2008. Appendages of early Cambrian *Eoredlichia* (Trilobita) from the Chengjiang biota, Yunnan, China. *Earth and Environmental Science Transactions of the Royal Society of Edinburgh* **99**, 213–23.
- HOU, J. B., HUGHES, N. C., YANG, J., LAN, T., ZHANG, X. G. & DOMINGUEZ, C. 2017. Ontogeny of the articulated yiliangelline trilobite *Zhangshania typica* from the lower Cambrian (Series 2, Stage 3) of southern China. *Journal of Paleontology* **91**, 86–99.
- HU, S. X., ZHU, M. Y., LUO, H. L., STEINER, M., ZHAO, F. C., LI, G. X., LIU, Q. & ZHANG, Z. F. 2013. *The Guanshan Biota*. Kunming: Yunnan Science and Technology Press, 204 pp.
- HU, S. X., ZHU, M. Y., STEINER, M., LUO, H. L., ZHAO, F. C. & LIU, Q. 2010. Biodiversity and taphonomy of the early Cambrian Guanshan biota, eastern Yunnan. *SCIENCE CHINA Earth Sciences* **53**, 1765–73.
- HUGHES, N. C. 2003. Trilobite tagmosis and body patterning from morphological and developmental perspectives. *Integrative and Comparative Biology* **43**, 185–206.
- HUGHES, N. C. 2007. The evolution of trilobite body patterning. *Annual Review of Earth and Planetary Sciences* **35**, 401–34.
- LEGG, D. A. 2014. *Sanctacaris uncata*: the oldest chelicerate (Arthropoda). *Naturwissenschaften* **101**, 1065–73.
- LEGG, D. A., SUTTON, M. D. & EDGEcombe, G. D. 2013. Arthropod fossil data increase congruence of morphological and molecular phylogenies. *Nature Communications* **4**, 2485. doi: [10.1038/ncomms3485](https://doi.org/10.1038/ncomms3485).
- LEGG, D. A., SUTTON, M. D., EDGEcombe, G. D. & CARON, J.-B. 2012. Cambrian bivalved arthropod reveals origin of arthropodization. *Proceedings of the Royal Society B: Biological Sciences* **279**, 4699–704.
- McHENRY, B. & YATES, A. 1993. First report of the enigmatic metazoan *Anomalocaris* from the southern hemisphere and a trilobite with preserved appendages from the Early Cambrian of Kangaroo Island, South Australia. *Records of the South Australian Museum* **26**, 77–86.
- MÜLLER, K. J. & WALOSZEK, D. 1987. Morphology, ontogeny, and life habit of *Agnostus pisiformis* from the Upper Cambrian of Sweden. *Fossils and Strata* **19**, 1–124.
- ORTEGA-HERNÁNDEZ, J., LEGG, D. A. & BRADY, S. J. 2013. The phylogeny of aglaspoid arthropods and the internal relationships within Artiopoda. *Cladistics* **29**, 15–45.
- OWENS, R. M. 2003. The stratigraphical distribution and extinctions of Permian trilobites. *Special Papers in Palaeontology* **70**, 377–97.
- RAMSKÖLD, L., CHEN, J. Y., EDGEcombe, G. D. & ZHOU, G. Q. 1996. Preservational folds simulating tergite junctions in tegopeltid and naraoiid arthropods. *Lethaia* **29**, 15–20.
- RAMSKÖLD, L. & EDGEcombe, G. D. 1996. Trilobite appendage structure – *Eoredlichia* reconsidered. *Alcheringa* **20**, 269–76.
- RAYMOND, P. E. 1920. The appendages, anatomy, and relationships of trilobites. *Memoirs of the Connecticut Academy of Arts and Sciences* **7**, 1–169.
- ROSS, R. J. 1979. Additional trilobites from the Ordovician of Kentucky. *United States Geological Survey Professional Paper* **1066D**, 1–13.
- SCHOLTZ, G. & EDGEcombe, G. D. 2005. Heads, Hox and the phylogenetic position of trilobites. *Crustacean Issues* **16**, 139–65.
- SCHOLTZ, G. & EDGEcombe, G. D. 2006. The evolution of arthropod heads: reconciling morphological,

- developmental and palaeontological evidence. *Development Genes and Evolution* **216**, 395–415.
- SHU, D. G., GEYER, G., CHEN, L. & ZHANG, X. L. 1995. Redlichiaean trilobites with preserved soft-parts from the lower Cambrian Chengjiang fauna (South China). *Beringeria Special Issue* **2**, 203–41.
- STEIN, M. 2013. Cephalic and appendage morphology of the Cambrian arthropod *Sidneyia inexpectans*. *Zoologischer Anzeiger* **253**, 164–78.
- STEIN, M., BUDD, G. E., PEEL, J. S. & HARPER, D. A. T. 2013. *Arthroaspis* n. gen., a common element of the Sirius Passet Lagerstätte (Cambrian, North Greenland), sheds light on trilobite ancestry. *BMC Evolutionary Biology* **13**. doi: 10.1186/1471-2148-13-99, 99 pp.
- STEIN, M. & SELDEN, P. A. 2012. A restudy of the Burgess Shale (Cambrian) arthropod *Emeraldella brocki* and reassessment of its affinities. *Journal of Systematic Palaeontology* **10**, 361–83.
- STÖRMER, L. 1939. Studies on trilobite morphology, Part I, The thoracic appendages and their phylogenetic significance. *Norsk Geologisk Tidsskrift* **19**, 143–273.
- STÖRMER, L. 1951. Studies on trilobite morphology, Part III, The ventral cephalic sutures with remarks on the zoological position of the trilobites. *Norsk Geologisk Tidsskrift* **29**, 108–58.
- STRAUSFELD, N. J. 2016. *Waptia* revisited: intimations of behaviors. *Arthropod Structure & Development* **45**, 173–84.
- STÜRMER, W. & BERGSTRÖM, J. 1973. New discoveries on trilobites by X-rays. *Paläontologische Zeitschrift* **47**, 104–41.
- TANAKA, G., HOU, X. G., MA, X. Y., EDGEcombe, G. D. & STRAUSFELD, N. J. 2013. Chelicerate neural ground pattern in a Cambrian great appendage arthropod. *Nature* **502**, 364–67.
- VAN ROY, P., ORR, P. J., BOTTING, J. P., MUIR, L. A., VINTHER, J., LEFEBVRE, B., HARIRI, K. E. & BRIGGS, D. E. G. 2010. Ordovician faunas of Burgess Shale type. *Nature* **465**, 215–8.
- WALCOTT, C. D. 1881. The trilobite, new and old evidence relating to its organization. *Bulletin of the Museum of Comparative Zoology at Harvard College* **8**, 190–242.
- WALCOTT, C. D. 1884. Appendages of the trilobite. *Science* **3**, 279–81.
- WALCOTT, C. D. 1912. Cambrian Geology and Paleontology II. No. 6. Middle Cambrian Branchiopoda, Malacostraca, Trilobita and Merostomata. *Smithsonian Miscellaneous Collections* **57**, 145–228.
- WALCOTT, C. D. 1918. Cambrian Geology and Paleontology IV. No. 4. Appendages of trilobites. *Smithsonian Miscellaneous Collections* **67**, 115–216.
- WALCOTT, C. D. 1921. Cambrian Geology and Paleontology IV. No. 7. Notes on structure of *Neolenus*. *Smithsonian Miscellaneous Collections* **67**, 365–456.
- WALOSSEK, D. & MÜLLER, K. J. 1990. Upper Cambrian stem-lineage crustaceans and their bearing upon the monophyletic origin of Crustacea and the position of *Agnostus*. *Lethaia* **23**, 409–27.
- WALOSSEK, D. & MÜLLER, K. J. 1998. Cambrian ‘Orsten’-type arthropods and the phylogeny of Crustacea. In *Arthropod Relationships* (eds R. A. Fortey & R. H. Thomas), pp. 139–53. London: Chapman & Hall.
- WHITTINGTON, H. B. 1975. Trilobites with appendages from the Middle Cambrian Burgess Shale, British Columbia. *Fossils and Strata* **4**, 97–136.
- WHITTINGTON, H. B. 1980. Exoskeleton, moult stage, appendage morphology, and habits of the Middle Cambrian trilobite *Olenoides serratus*. *Palaeontology* **23**, 171–204.
- WHITTINGTON, H. B. 1993. Anatomy of the Ordovician trilobite *Placoparia*. *Philosophical Transactions of the Royal Society of London, Series B: Biological Sciences* **339**, 109–18.
- WHITTINGTON, H. B. & ALMOND, J. E. 1987. Appendages and habits of the Upper Ordovician trilobite *Triarthrus eatoni*. *Philosophical Transactions of the Royal Society of London, Series B: Biological Sciences* **317**, 1–46.
- WHITTINGTON, H. B., CHATTERTON, B. D. E., SPEYER, S. E., FORTEY, R. A., OWENS, R. M., CHANG, W. T., DEAN, W. T., GEYER, G., JELL, P. A., LAURIE, J. R., PALMER, A. R., REPINA, L. N., RUSHTON, A. W. A., SHERGOLD, J. H., CLARKSON, E. N. K., WILMOT, N. V. & KELLY, S. R. A. 1997. *Part O Trilobita Revised Volume 1*. Boulder, Colorado: The Geological Society of America; and Lawrence, Kansas: University of Kansas Press, 530 pp.
- WOLFF, C. & SCHOLTZ, G. 2008. The clonal composition of biramous and uniramous arthropod limbs. *Proceedings of the Royal Society B: Biological Sciences* **275**, 1023–8.
- YANG, J., ORTEGA-HERNÁNDEZ, J., BUTTERFIELD, N. J. & ZHANG, X. G. 2013. Specialized appendages in fuxianhuids and the head organization of early euarthropods. *Nature* **494**, 468–71.
- ZENG, H., ZHAO, F. C., YIN, Z. J., LI, G. X. & ZHU, M. Y. 2014. A Chengjiang type fossil assemblage from the Hongjingshao Formation (Cambrian Stage 3) at Chenggong, Kunming, Yunnan. *Chinese Science Bulletin* **59**, 3169–75.
- ZHANG, Z. Q. 2013. Phylum arthropoda. In *Animal Biodiversity: An Outline of Higher-Level Classification and Survey of Taxonomic Richness (addenda 2013)* (ed. Z. Q. Zhang). *Zootaxa* **3703**, 17–26.
- ZHANG, X. L., HAN, J., ZHANG, Z. F., LIU, H. Q. & SHU, D. G. 2004. Redescription of the Chengjiang arthropod *Squamacula clypeata* Hou and Bergström, from the Lower Cambrian, south west China. *Palaeontology* **47**, 605–17.
- ZHANG, W. T., LU, Y. H., ZHU, Z. L., QIAN, Y. Y., LIN, H. L., ZHOU, Z. Y., ZHANG, S. G. & YUAN, J. L. 1980. *Cambrian Trilobite Faunas of Southwestern China*. Beijing: Science Press, 497 pp. (in Chinese with English summary).
- ZHANG, X. L. & SHU, D. G. 2005. A new arthropod from the Chengjiang Lagerstätte, Early Cambrian, southern China. *Alcheringa* **29**, 185–94.
- ZHANG, X. L., SHU, D. G. & ERWIN, D. H. 2007. Cambrian naraoiids (Arthropoda): morphology, ontogeny, systematics, and evolutionary relationships. *Journal of Paleontology* **81**, 1–52.
- ZHU, M. Y., LI, G. X., ZHANG, J. M., STEINER, M., QIAN, Y. & JIANG, Z. W. 2001. Early Cambrian stratigraphy of east Yunnan, southwestern China: a synthesis. *Acta Palaeontologica Sinica* **40**, 4–39.
- ZHU, M. Y., BABCOCK, L. E. & STEINER, M. 2005. Fossilization modes in the Chengjiang Lagerstätte (Cambrian of China): testing the roles of organic preservation and diagenetic alteration in exceptional preservation. *Palaeogeography, Palaeoclimatology, Palaeoecology* **220**, 31–46.