The first Silurian chasmataspid, *Loganamaraspis* dunlopi gen. et sp. nov. (Chelicerata: Chasmataspidida) from Lesmahagow, Scotland, and its implications for eurypterid phylogeny

O. Erik Tetlie and Simon J. Braddy

ABSTRACT: A new chasmataspid (Chelicerata: Chasmataspidida) is described from the Early Silurian (Late Llandovery–Early Wenlock) of Lesmahagow, Scotland, as *Loganamaraspis dunlopi* gen. et sp. nov. It is distinguished from related forms by the low tapering ratio of the postabdomen, pediform sixth prosomal appendage and a heart-shaped metastoma. Additionally, a genital operculum with a genital appendage is preserved. The recognisable morphology of *L. dunlopi* gen. et sp. nov. bridges some of the gap between the Ordovician Chasmataspididae and the Devonian Diploaspididae. Traces of the gut are reported for the first time from a chasmataspid. Chasmataspids are regarded as sister group to the eurypterids: an anterior opercular plate anterior to the genital appendage in *L. dunlopi* gen. et sp. nov. is regarded as homologous to the anterior opercular plate of the three-segmented genital operculae of *Dolichopterus* and Stylonurina. This is considered a plesiomorphic character within Eurypterida, whilst the two-segmented genital operculum (with deltoid plates) of Eurypterina is considered apomorphic.



KEY WORDS: Arthropod, Diploaspididae, Eurypterida, genital operculum, metastoma.

Chasmataspids are a small and poorly known group of chelicerates that are distinguished from eurypterids by the segmentation pattern of their opisthosoma. They have a preabdomen of four and a postabdomen of nine segments, while eurypterids usually (i.e. excluding megalograptids) have a preabdomen of eight and a postabdomen of five segments. Prior to 1999, only three species of chasmataspids were known: Chasmataspis laurencii Caster & Brooks, 1956 from the Middle Ordovician (?Late Llandeilo) of Tennessee, USA; and Diploaspis casteri Størmer, 1972 and Heteroaspis novojilovi Størmer, 1972 from the Early Devonian (Emsian) of Alken, Germany. The number of described chasmataspids has more than doubled in recent years (Table 1), based mainly on specimens previously considered to be juvenile eurypterids. Chasmataspids are thus more common than previously thought and more discoveries may await in museum collections of 'juvenile

A restudy of C. laurencii Caster & Brooks, 1956 by Dunlop et al. (2004) has increased our understanding of early members of the group. A restudy of the original Alken material and some new specimens suggests that Heteroaspis novojilovi Størmer, 1972 is a preservational and/or ontogenetic junior synonym of Diploaspis casteri Størmer, 1972 (Dunlop et al. 2001). In addition, more material still awaits description or redescription: among these are Late Cambrian chasmataspid resting traces from Texas (Wahlman & Caster 1978; Dunlop et al. 2004), a new species of Diploaspis from the Early Devonian of Hombach, Germany (M. Poschmann, pers. comm.; Dunlop et al. 2001), and a specimen from the Early Devonian of Siberia, described by Novojilov (1959) as Eurypterus stoermeri Novojilov, 1959, that probably represents a chasmataspid (Table 1) based on its opisthosomal differentiation.

Morphologically, chasmataspids resemble both xiphosurans and eurypterids, but were originally referred to an order of Xiphosura: Chasmataspida (=Chasmataspidida) (Caster & Brooks, 1956). Bergström (1968) considered C. laurencii to be a synziphosurine and questioned the ordinal status of chasmataspids. Størmer (1972) supported the view of Caster & Brooks and regarded C. laurencii and his two new genera as belonging to the order Chasmataspidida within Xiphosura. Anderson & Selden (1997) excluded chasmataspids from Xiphosura, but left their status open. Dunlop & Selden (1997) raised Chasmataspida (=Chasmataspidida) to a separate taxon within Chelicerata on the basis of their differentiation of the opisthosoma into a four-segmented preabdomen (buckler) and a nine-segmented postabdomen, which is unique for this taxon. The first opisthosomal tergite of chasmataspids is reduced and can only be observed in Octoberaspis ushakovi Dunlop, 2002 and C. laurencii (Dunlop et al. 2004), confirming a total of 13 opisthosomal segments. Thirteen opisthosomal segments is a character also shared with eurypterids and scorpions (Dunlop & Webster 1997). Octoberaspis ushakovi possesses both a genital appendage and a metastoma, features previously thought to be autapomorphies for eurypterids only. This further provides strong evidence for a sister group relationship between Chasmataspidida and Eurypterida (see section 4).

When studying the eurypterid collections at the National Museums of Scotland (NMS), Chambers Street, Edinburgh, Scotland, the present authors and Dr Lyall I. Anderson (Invertebrate Curator at NMS) discovered a small specimen (NMS G. 1957.1.649) labelled as a 'young eurypterid'. They report here on this specimen, the first Silurian chasmataspid. Like *O. ushakovi*, it possesses fragmentary prosomal appendages, a genital appendage and a metastoma.

Table 1 Summary of chasmataspid occurrences in approximate stratigraphical order, including 'postabdominal tapering ratios' (i.e. the width of opisthosomal segment 5 divided by the width of segment 13) and inferred palaeoenvironment

Taxon	Age	Stratigraphy and locality	Palaeoenvironment	(W5/W13)	References		
'Chasmataspis-like resting trace'	Upper Cambrian	Riley Formation, Texas, USA	Shoreline or lacustrine	?	Wahlman & Caster 1978; Dunlop <i>et al.</i> 2004		
Chasmataspis laurencii	Middle Ordovician (Late Llandeilo)	?Five Oaks Formation, Douglas Dam, Tennessee, USA	Carbonate platform margin	1.5	Caster & Brooks 1956; Dunlop <i>et al.</i> 2004		
Loganamaraspis dunlopi	Late Llandovery or Early Wenlock	Patrick Burn Formation, Lesmahagow, Scotland	Non-marine or marginal marine	c. 2·5	This paper		
Forfarella mitchelli	Early Devonian	Dundee Formation, Angus Region, Midland Valley, Scotland	?Lacustrine	c. 5	Dunlop et al. 1999		
Octoberaspis ushakovi	Early Lochkovian	Severnaya Zemlya Formation, October Revolution Island, Russia	?Lagoonal	c. 3·5	Dunlop 2002		
Diploaspis n. sp.	Emsian (?)	?Hombach, Germany	Shallow deltaic	?	Poschmann (pers. comm.)		
Diploaspis casteri	Early Emsian	Nellenköpfchen Formation, Alken an der Mosel, Germany	?Shallow deltaic	4.1	Størmer 1972; Dunlop et al. 2001		
'Eurypterus' stoermeri	Emsian (?)	Zubova Suite Formation, Imaigda River, Norilsk, Russia	?Marginal marine	c. 3·4	Novojilov 1959; Plotnick 1999		
Achanarraspis reedi	Late Eifelian	Achanarras Limestone Formation, Caithness, Scotland	Lacustrine	3.4	Anderson et al. 2000		

1. Material and methods

The specimen, NMS G. 1957.1.649, was examined and drawn in a 70% alcohol solution using a binocular microscope with a drawing tube attachment. The photographs were taken under polarised light. Since no morphological standards have been proposed for chasmataspids, the terminology follows that of Tollerton (1989) for eurypterids. However, Tollerton (1989) did not propose any terminology for the segmentation of the genital operculum, and therefore, the present authors follow the nomenclature of Waterston (1979, fig. 4a); the segments anterior, lateral and posterior to the base of the genital appendage are the anterior, median and posterior opercular plates, respectively. All three plates are present only in chasmataspids, dolichopterid and stylonurid eurypterids. Eurypterina (sensu Novojilov 1962; appendage VI expanded into a paddle) have lost the anterior opercular plates. A structure dorsal to the metastoma, previously not reported from any chasmataspid or eurypterid, is termed the 'metastomal plate.'

1.1. Locality and geological setting

The specimen described here was collected by Robert Dunlop of Dunfermline, Scotland, and presented to the NMS by the Carnegie Dunfermline Trust in 1957. The specimen comes from the Patrick Burn Formation (lower part of Priesthill Group), from Dunside, Logan Water, Lesmahagow, Scotland. The formation is at least 850 m thick, and composed of grey and olive mudstones laminated with silty mudstones with laminae of dark grey carbonaceous siltstones (Paterson *et al.* 1998). The formation is regarded as late Llandovery (Paterson *et al.* 1998) or possibly early Wenlock (Plotnick 1999) in age. The precise bed that yielded the specimen is unknown, but based upon lithologic similarity, we suspect it is from the famous *Jamoytius* horizon, interpreted as non-marine or marginal marine (BA1) by Plotnick (1999), or the middle of a

constricted marine basin occasionally flushed by turbiditic flows by Paterson et al. (1998).

Although no other fossils are associated on the same slab, the lithology is the same as that yielding the eurypterids Slimonia acuminata Salter, 1855, Erettopterus bilobus Salter, 1855, Nanahughmilleria lanceolata Salter, 1855, ?Hardieopterus lanarkensis Waterston, 1979, the synziphosurids Cyamocephalus loganensis Currie, 1927 and an undescribed specimen of Pseudoniscus (Eldredge 1974), the phyllocarid Ceratiocaris papilio, the fish Jamoytius kerwoodi White, 1946, Logania scotica Traquair, 1899, and the ?thylacocephalan Ainiktozoon loganense Scourfield, 1937.

1.2. Preservation and taphonomy

NMS G. 1957.1.649 is preserved as black material and is partially flattened in a siltstone matrix. This preservation is typical for specimens (see above) from the Patrick Burn Formation. NMS G. 1957.1.649 b (Figs 1b, 2b) shows the ventral side of the animal, while NMS G. 1957.1.649 a (Figs 1a, 2a) shows superimposed dorsal (e.g. tergites), ventral (e.g. appendages) and internal features (i.e. traces of the gut, a feature never previously reported from any chasmataspid). The preservation of the appendages, metastoma and genital appendage is very faint.

Towards the posterior of the postabdomen, dark oval areas can be observed on the anterior parts of the segments, symmetrical around the midline. They are not close to the ventral segment margins on NMS G. 1957.1.649 b, but when superimposed with NMS G. 1957.1.649 a, it is clear that they are close to the dorsal anterior segment boundaries. The specimen has been obliquely flattened, and the oval areas represent impressions of the anterior and posterior openings of the flattened, ring-like postabdominal segments. The two posteriormost segments are preserved only as patches of cuticle, offering only indistinct segment boundaries.



Figure 1 Loganamaraspis dunlopi gen. et sp. nov.: (A) NMS G. 1957.1.649 a, showing mainly the dorsal surface of the holotype; and (B) NMS G. 1957.1.649 b, showing the ventral surface. Scale bars = 5 mm.

1.3. Abbreviations

The following abbreviations are used in the text and figures: (aop) anterior opercular plate; (ca) carapace; (do) doublure; (ga) genital appendage; (met) metastoma; (mp) 'metastomal plate'; (mt) microtergite; (op) operculum; (sp) spine; (tel) telson; and (ubl) unfused Blattfüsse. Prosomal appendages are labelled I–VI, coxae of prosomal appendages cIV–cVI, and opisthosomal segments 1–13.

2. Systematic palaeontology

Chelicerata Heymons, 1901 Chasmataspidida Caster & Brooks, 1956 Family Diploaspididae Størmer, 1972

Emended diagnosis. Small chasmataspids with semicircular, subrectangular or subquadrate carapace, preabdomen with curved segments, a tapering postabdomen and a short telson; no division of preabdominal segments into median and lateral plates (emended from Dunlop *et al.* 2001).

Remarks. The presence of a pediform sixth appendage in *L. dunlopi* gen. et sp. nov. might justify the erection of a new family, since all other members of Diploaspididae, where prosomal appendage VI is known, have an expanded swimming leg. Since the appendages in this specimen are so poorly preserved, the present authors choose not to erect a new family.

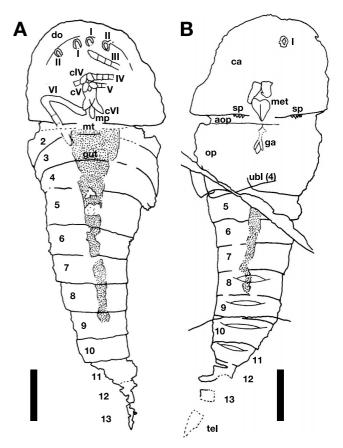


Figure 2 Loganamaraspis dunlopi gen. et sp. nov. Camera lucida drawings of the two specimens illustrated in Fig. 1. (A) NMS G. 1957.1.649 a; (B) NMS G. 1957.1.649 b. Scale bars = 5 mm.

Loganamaraspis gen. nov.

Diagnosis. Large diploaspid with posterolaterally expanded prosoma lacking genal spines; prosomal appendage VI, a pediform walking leg; metastoma cardioid (heartshaped). Postabdominal segments long with a tapering ratio (W5/W13) of c. 2·5.

Remarks. The tapering ratio (W5/W13) is low (c. 2.5) compared to other known Devonian diploaspid genera (3.4–c. 5.0; Table 1).

Type species. *Loganamaraspis dunlopi* gen. et sp. nov., by monotypy.

Etymology. The generic name is based on *Logan*, after Logan Water, where the specimen was discovered; *amare*, Latin for love, after the heart-shaped metastoma; and *aspis*, Greek for shield.

Loganamaraspis dunlopi gen. et sp. nov. (Figs 1–3)

Diagnosis. As for the genus.

Etymology. The species is named in joint honour of the discoverer of this specimen, Robert Dunlop of Dunfermline and Dr Jason A. Dunlop (Museum für Naturkunde, Berlin, Germany) for his valuable contributions to our rapidly expanding knowledge of chasmataspids.

Holotype. NMS G. 1957.1.649 a+b from the Patrick Burn Formation of the Priesthill Group, Dunside, Logan Water, Lesmahagow, Ayrshire, Scotland.

Additional material. None.

Description. Entire specimen approximately 35 mm long and devoid of any ornamentation. Prosoma subquadrate, posterolaterally expanded with no preserved genal spines.

Prosomal length 8·5 mm, width 11·4 mm, giving a L/W ratio of 0·75. Lateral angles are 90° on the right side and 100° on the left because of taphonomic distortion. Prosomal doublure (Fig. 2a, do) has even width anteriorly and laterally, but no sutures are evident. Posterior ventral margin of prosoma bears 0·25 mm long serrations on either side of the metastoma. Eyes and other dorsal structures are not preserved.

Two pairs of ring-like structures positioned anteriorly are interpreted as the cross-section of appendages I (chelicerae) and II, comparable in width to that of the posterior appendages, indicating that the chelicerae were very robust. Posterior to these appendages is a more slender appendage III (Fig. 2a, III), preserved length 3 mm, width 0.5 mm. It is possible to identify individual podomeres in this appendage; all are rectangular, 0.6-0.7 mm long, 0.5 mm wide. Both coxae of appendage IV are clearly present (Fig. 2a, cIV), but lack gnathobases; each coxa is 0.7 mm long and 0.5 mm wide. Proximal portions of both appendage IV preserved; individual podomeres slightly shorter (0.5 mm long) than those on appendage III. Two similar-sized coxae of appendage V are present (Fig. 2a, cV), lacking gnathobases; each coxa 0.7 mm long and 0.5 mm wide. On the left side, podomeres of a slender appendage V are preserved (Fig. 2a, V); individual podomeres are approximately the same size as appendage IV. The larger coxae (1.5 mm long, 0.8 mm wide) of both appendage VI (Fig. 2a, cVI) are preserved although no gnathobases are evident. From posterior right coxa, a long (total preserved length 8.5 mm) and slender appendage VI (constant width 0.6 mm) projects towards the prosomal rim, and bends posteriorly onto the opisthosoma. Individual podomeres cannot be distinguished, although they must have been longer than those of the anterior appendages since the appendage itself is longer than the anterior appendages.

Posteriormost on the prosoma, partially extending into the opisthosoma, a cardioid (heart-shaped) metastoma is evident (Fig. 2b, met) with a median suture. An oval plate overlies the metastoma posterior to coxae VI (Fig. 2a, mp).

Ventrally, the anterior opercular plate is evident anterior to the genital appendage (see sections 3 and 4). The genital appendage is 2.5 mm long (extending half-way down the preabdomen) and composed of two segments. The proximal segment of the genital appendage is split into two lobes, separated by an angle of approximately 35° (Fig. 2b, ga). Between these two lobes is the distal segment, widest at the point where the lobes of the proximal segment end and taper distally. The number of opercular plates is unclear since no sutures are evident on the operculum posterior to the base of the genital appendage. The presumed fourth segment appears to have been unfused at the midline since the left and right plates overlap in the posterior median portion of the preabdomen (Fig. 2b, ubl).

Dorsally, the three posterior segments of the preabdomen can be identified on NMS G. 1957.1.649 a, and although not complete laterally, have approximately the same width as the posterior portion of the prosoma. The posterior margin of these segments is strongly convex.

The first postabdominal segment is very long and is slightly disarticulated from the preabdomen (see the left side of Fig. 2b), but is easily distinguished from the preabdomen since it is narrower with an almost straight posterior segment margin. The succeeding postabdominal segments taper slightly. The postabdominal segments are approximately the same length except for the last two, which are shorter. The opisthosomal segments have the following preserved lengths and widths in millimetres (*denotes incomplete lateral preservation): (1) 1·5/?; (2) 0·5/11·0*; (3) 2·2/10·2*; (4) 2·4/9·5*; (5) 3·1/7·9; (6) 2·4/6·7; (7) 2·3/6·4; (8) 2·4/5·5; (9) 2·2/4·6*; and

(10) 1.9/4.3. The posterior structure (Fig. 2b, tel) is interpreted as a short telson.

A dark area, interpreted as the midgut and intestine, can be followed medially along the specimen from the posterior edge of the prosoma to the middle of the ninth segment (Fig. 2a, gut). The gut trace increases in width along the opisthosoma to a maximum width (4·1 mm) under the second and third segments, and decreases in width towards the posterior part of the abdomen.

3. Discussion

3.1. Prosoma and prosomal appendages

The appendages of L. dunlopi gen. et sp. nov. are very poorly preserved. The form of the distal parts (i.e. chelate as in xiphosurids and apparently C. laurencii, or non-chelate as in eurypterids and D. casteri) is unknown. Appendage VI is long and slender with a constant width and obviously used primarily for walking rather than swimming, as in the Devonian diploaspids and Eurypterina (sensu Novojilov 1962). This suggests that the swimming appendage in diploaspids and Eurypterina evolved independently (Fig. 4), an interpretation also supported by the different morphology of the swimming appendage in the two groups. The diploaspids appear to lack a small distal podomere (9) in appendage VI. More importantly, the small triangular podomere (7a in eurypterids) that occurs between the two largest podomeres of the swimming leg is situated posterolateral in eurypterids (e.g. Størmer 1955, figs 17 & 21–25), but anterolateral in diploaspids (Dunlop et al. 2001, fig. 8b; Dunlop 2002, fig. 9a).

The metastoma is cardioid (Tollerton 1989, fig. 5), but with a pointed posterior termination similar to that of *O. ushakovi* Dunlop, 2002. The suture along the midline suggests that the metastoma formed by the fusion of two structures, similar to the metastoma in many eurypterids, for example *Baltoeurypterus tetragonophthalmus* (Holm 1898, pl. 3, figs 19 & 21). The metastoma has previously been interpreted as homologous to the xiphosurid chilaria (Størmer 1934, 1970) and the fused coxae of the same (seventh) appendage of *Weinbergina opitzi* (Stürmer & Bergström 1981) or to the fused epicoxae of appendage VI (Braddy 1996). The oval plate observed between and posterior to coxae VI (Fig. 2a, mp) was probably connected to the dorsal side of the metastoma.

3.2. Genital appendages and sexual dimorphism

The genital appendage of *L. dunlopi* gen. et sp. nov. is composed of two segments. The proximal segment is divided into two lobes distally, and the smaller distal segment is a median lobe between these two distal lobes of the proximal segment. This is the same morphology as that in *O. ushakovi* (Dunlop 2002), although the lateral lobes are larger in *L. dunlopi* gen. et sp. nov. Dunlop (2002) hesitated to assign the metastoma and genital appendages in his material to chasmataspids, and suggested that they may represent eurypterids since the specimens showing these features had poorly preserved opisthosomal differentiation. However, the similarities between the present material and *O. ushakovi* remove any doubt that the structures described were truly features of chasmataspids, hence *O. ushakovi*.

Sexual dimorphism is poorly known in chasmataspids, although Dunlop (2002) reported two types of genital appendage in *O. ushakovi*. There are close similarities between the genital appendage of *L. dunlopi* gen. et sp. nov. and the two-segmented type A appendages (Størmer 1934) of *O. ushakovi* and *Dolichopterus jewetti* Caster & Kjellesvig-Waering (1952) (Fig. 4), suggesting that the holotype of

Table 2 Homology of opisthosomal segments for chasmataspids and eurypterids: ? unknown; (Mt) microtergite; (Con) concealed; (Ank) ankylosed; (B) Blattfüsse; and (T) tergite

Taxon	Opisthosomal segment									
	Aspect	1	2	3	4	5	6	7	8–13	Telson
Chasmataspis laurencii (Caster & Brooks)	Dorsal	Mt	Buckler		Ank	Ank	Ank	Ank	Long styliform	
	Ventral	?	Ventral buckler							
Loganamaraspis dunlopi	Dorsal	Mt	Buckler		Ank	Ank	Ank	Ank	Short styliform	
	Ventral	Aop	Operculum B		В					
Dolichopterus jewetti	Dorsal	Con?	Т	Т	Т	Т	Т	Т	Ank	Long styliform
	Ventral	Aop	Operculum l		В	В	В	В		
Baltoeurypterus tetragonophthalmus	Dorsal	Con	Т	Т	Т	Т	Т	Т	Ank	Long styliform
	Ventral	Lost	Operculum		В	В	В	В		

L. dunlopi gen. et sp. nov. could be a female (see also Braddy & Dunlop 1997). The proximal segment is divided into two lobes in L. dunlopi gen. et sp. nov. and O. ushakovi, but these lobes are fused along a midline in D. jewetti. The type B appendage assigned to O. ushakovi was short and ovate, as in most eurypterids, but is very poorly preserved.

3.3. Opisthosoma and gut

Devonian chasmataspids and *L. dunlopi* gen. et sp. nov. (i.e. diploaspids) can be separated from eurypterids and the Ordovician *C. laurencii* by the broadly convex tergite boundaries on the preabdomen. The present authors cannot demonstrate that the preabdominal segments are fused into one unit in *L. dunlopi* gen. et sp. nov., although this seems to be the case in morphologically better understood chasmataspids (e.g. *C. laurencii* and *D. casteri*), and is probably the explanation for the vague segment boundaries in the preabdomen of *L. dunlopi* gen. et sp. nov. Assuming the holotype of *L. dunlopi* gen. et sp. nov. has an untelescoped abdomen (cf. *O. ushakovi*), it is 35 mm long, and is only rivalled in size by *C. laurencii* among other chasmataspids.

Based on the observed morphology of L. dunlopi gen. et sp. nov. alone, the preabdomen apparently consists of a microtergite and three fused segments dorsally, and an anterior opercular plate, one fused posterior opercular plate and one pair of non-fused Blattfüsse (opisthosomal segment four) ventrally. However, based on a Late Cambrian chasmataspid resting trace (Wahlman & Caster 1978) and reinterpretation of the ventral morphology of C. laurencii (sensu Dunlop et al. 2004), the ventral morphology of L. dunlopi gen. et sp. nov. might have consisted of two Blattfüsse-like segments (2 and 3) of the operculum which are not preserved (or not present as the operculum is in its place). The fourth is apparent and overlapping (Fig. 2b, ubl). Dunlop et al. (2004) discussed the possibility that the 'ventral plate' (cf. Caster & Brooks 1956) or 'median plate' (cf. Størmer 1972) was a sclerotised layer above a series of gill-bearing opercula. This seems to have been the case for O. ushakovi. One problem with this interpretation in C. laurencii is that Caster & Brooks (1956, p. 165) listed the number of specimens examined: pieces of 50 dorsal bucklers and 49 'ventral bucklers' were found, and it may be expected that examples of the operculae, ventral to this plate, should also occur if they were sclerotised. An alternative explanation

could be that they were lost during transportation or decay. Babcock et al. (2000) reported that the book gills become detached during the first week of transport in Limulus polyphemus, much earlier than the prosomal appendages (which become detached after 10–18 days). However, if these operculae were very lightly sclerotised, they may never have been preserved. It is also possible that the Upper Cambrian animal producing the traces in Texas and the Ordovician C. laurencii had not developed sclerotised ventral operculae, but L. dunlopi gen. et sp. nov., the Devonian chasmataspids and the eurypterids had developed them, and the genital appendage was developed in response to the sclerotisation of the ventral side. An attempt to homologise the ventral and dorsal segments of chasmataspids and eurypterids is given in Table 2.

The dark area along the midline is interpreted as the outline of the midgut and intestine, rather than the midgut glands reported from the Burgess Shale arthropod Leanchoilia superlata by Butterfield (2002). This is supported by the regular outline of the area, lack of internal structure and visible extent of the area all the way to the ninth segment. A gut trace has never been reported previously from chasmataspids, but examples are known from several eurypterid genera (e.g. Ruedemann 1921; Kjellesvig-Waering 1958, 1963; Heubusch 1962; Waterston 1979). The widening of the dark patch in the second and third segments corresponds to the gut that lay protected under the preabdomen, and this widened area may represent the midgut. If so, this might suggest that chasmataspids only had two pairs of serially repeated midgut glands (assuming one pair of glands per somite, as in L. superlata), a number shared with xiphosurans (Fahrenbach 1999). Scorpions have five pairs of midgut glands (Farley 1999) and the number in spiders is variable (Foelix 1996), while the number in eurypterids is still not known.

4. Implications for chasmataspid and eurypterid phylogeny

4.1. Chelicerate phylogeny

Chelicerate phylogeny is problematic. The most comprehensive study including fossil taxa was by Dunlop & Selden (1997). They identified two clades of chelicerates: the first

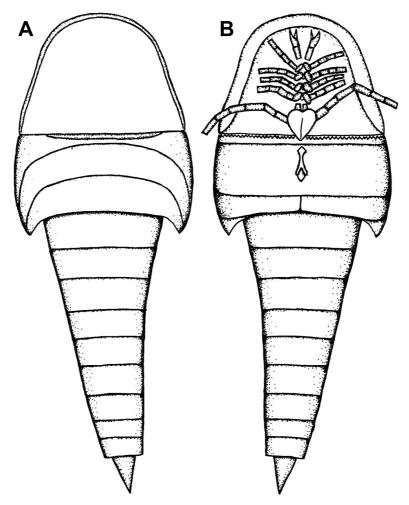


Figure 3 Morphological reconstruction of (A) the dorsal side of *Loganamaraspis dunlopi* gen. et sp. nov. and (B) the ventral side, illustrating the prosomal appendages, metastoma and genital operculum. The morphology of the prosomal appendages and telson is reconstructed according to Fig. 2, but distal parts of the appendages are not reconstructed since it is uncertain whether they are chelate. The postero-ventral serrated margin of the prosoma is reconstructed extending to the lateral margins. The anterior three segments on the postabdomen are not reconstructed with Blattfüsse since no evidence for this is provided by the specimen, although Blattfüsse on these segments are present in eurypterids.

composed of synziphosurines and xiphosurids (for a more comprehensive analysis of this clade, see Anderson & Selden 1997); and the second clade composed of chasmataspids, eurypterids, scorpions and non-scorpion arachnids, although concerns persist that Arachnida is paraphyletic (Dunlop & Webster 1999; Dunlop & Braddy 2001). This new chasmataspid casts no further light upon this question, but the presence of a metastoma and a genital appendage support a sister group relationship between eurypterids and chasmataspids.

4.2. Chasmataspid phylogeny

Concern that chasmataspids might be polyphyletic exists (Bergström 1979, 1980; R. E. Plotnick, pers. comm), in which case the Ordovician *Chasmataspis* would be separated from the Devonian forms based on its chelate appendages and a more complex, tripartite preabdomen. The present study of *L. dunlopi* reveals it to be morphologically intermediate in postabdominal tapering ratio and appendage morphology between the Ordovician and Devonian chasmataspids, although it is referred to the same family as the latter. However, the presence of a metastoma and a genital appendage in *O. ushakovi, L. dunlopi* gen. et sp. nov. and eurypterids combined with the apparent lack of these structures in *C.*

laurencii, renders support for a paraphyletic Chasmataspidida more likely (Fig. 4).

4.3. The anterior opercular plate and implications for eurypterid phylogeny

The anterior opercular plate in front of the genital appendage of L. dunlopi gen. et sp. nov. (Figs 2b, aop, & 3b) is unusual compared to eurypterids, but far from unique. The importance of this character has not been noted previously. Caster & Kjellesvig-Waering (1952) described Dolichopterus jewetti and D. stoermeri on the basis of two, type A operculae with a segment anterior to the deltoid plates (Fig. 4). An anterior third segment has also been noted several times in stylonurid eurypterids: Parastylonurus ornatus (Waterston 1979, fig. 4A, B), Stylonurella spinipes (see Kjellesvig-Waering in Waterston 1979, fig. 12D), Rhenopterus diensti (Størmer 1936, fig. 9), possibly Moselopterus ancylotelson and M. elongatus (Størmer 1974, figs 25 & 29), and finally, an operculum sp. II (Størmer 1934) that can be referred to Brachyopterella pentagonalis (Størmer, 1934) because of the character presented here, its size and provenance (OET pers. obs.). As mentioned earlier, this anterior segment probably represents the ventral expression of the dorsal microtergite or the appendages of this segment. The latter interpretation is problematic with respect to the origin of

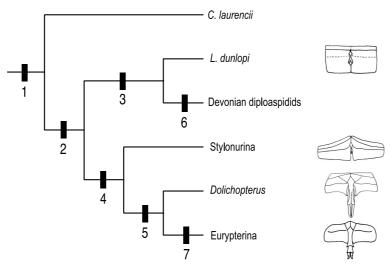


Figure 4 Chasmataspid phylogeny showing inferred position of Loganamaraspis dunlopi gen. et sp. nov. relative to other chasmataspids and the major groups of eurypterids based on the characters discussed herein. The figured genital operculae belong to L. dunlopi gen. et sp. nov., Stylonurella spinipes, Dolichopterus jewetti and Baltoeurypterus tetragonophthalmus. Character transformations occurring at the numbered nodes: (1) opisthosoma of 13 segments, 'ventral plate', chelate prosomal appendages; (2) non-chelate appendages, metastoma, operculum with genital appendage; (3) broadly curved preabdominal segments; (4) preabdomen of eight segments, microtergite concealed dorsally, 'ventral plate' lost; (5) deltoid plates on genital operculum, prosomal appendage VI expanded swimming leg with podomere 7a posteriorly; (6) prosomal appendage VI expanded swimming leg with podomere '7a' anteriorly; and (7) anterior opercular plate lost, prosomal appendage VI broad swimming leg with reduced podomere nine.

the metastoma; the anterior opercular plates in D. jewetti and L. dunlopi gen. et sp. nov. are here considered homologous, and both also have a metastoma. Therefore, whether the seventh pair of appendages in Weinbergina opitzi is homologous to the metastoma or the anterior opercular plate remains speculative. The anterior opercular plate is reduced in the suborder Eurypterina (sensu Novojilov 1962). Caster & Kjellesvig-Waering (1952, p. 26) suggested that a threesegmented operculum was the plesiomorphic state for eurypterids and this new evidence from L. dunlopi gen. et sp. nov. supports their view. Although a three-segmented operculum cannot be seen in L. dunlopi gen. et sp. nov., the homology of the anterior opercular segments of L. dunlop gen. et sp. nov., Dolichopterus and stylonurid eurypterids has wide implications for eurypterid phylogeny. It suggests that stylonurids are plesiomorphic with respect to this character, and Dolichopterus has a position intermediate between Stylonurina and Eurypterina. However, no stylonurids have deltoid plates which are present in Dolichopterus and the suborder Eurypterina (sensu Novojilov 1962). The phylogeny of the major clades of chasmataspids and eurypterids can be tentatively mapped from the morphology of appendage VI and the development of the genital operculum (Fig. 4).

5. Acknowledgements

We thank Dr Lyall I. Anderson (National Museums of Scotland, Edinburgh) for providing information regarding the locality, and for helping locate the specimen. Simon Powell (University of Bristol) is thanked for preparing the photographs. Professor Derek E. G. Briggs (University of Yale) is thanked for valuable discussion. Professor Euan N. K. Clarkson (University of Edinburgh) provided information on the stratigraphy. Dr Jason A. Dunlop (Museum für Naturkunde, Berlin) and one anonymous referee are thanked for reviewing the manuscript. OET acknowledges grant 145565/432 into chelicerate phylogeny and evolution from Norges Forskningsråd.

7. References

Anderson, L. I., Dunlop, J. A. & Trewin, N. H. 2000. A Middle Devonian chasmataspid arthropod from Achanarras Quarry, Caithness, Scotland. Scottish Journal of Geology 36, 151–8.

Anderson, L. I. & Selden, P. A. 1997. Opisthosomal fusion and phylogeny of Palaeozoic Xiphosura. *Lethaia* 30, 19–31.

Babcock, L. E., Merriam, D. F. & West, R. R. 2000. *Paleolimulus*, an early limuline (Xiphosurida), from Pennsylvanian-Permian Lagerstätten of Kansas and taphonomic comparison with modern *Limulus*. *Lethaia* 33, 129–41.

Bergström, J. 1968. Eolimulus, a Lower Cambrian xiphosurid from Sweden. Geologiska Föreningens i Stockholm Förhandlingar 90, 489–503.

Bergström, J. 1979. Morphology of fossil arthropods as a guide to phylogenetic relationships. *In* Gupta, A. P. (ed.) *Arthropod phylogeny*, 3–56. New York, NY: Van Nostrand Reinhold Co.

Bergström, J. 1980. Morphology and systematics of early arthropods. In Kraus, O. (ed.) Arthropoden-Phylogenie. Abhandlungen des Naturwissenschaftlichen Vereins in Hamburg (NF) 23, 7–42.

Braddy, S. J. 1996. *Palaeobiology of the Eurypterida*. Unpublished PhD thesis, University of Manchester.

Braddy, S. J. & Dunlop, J. A. 1997. The functional morphology of mating in the Silurian eurypterid, *Baltoeurypterus tetragonoph-thalmus* (Fisher, 1839). *Zoological Journal of the Linnean Society* 121, 435–61.

Butterfield, N. J. 2002. *Leanchoilia* guts and the interpretation of three-dimensional structures in Burgess Shale-type fossils. *Paleobiology* **28**, 155–71.

Caster, K. E. & Brooks, H. K. 1956. New fossils from the Canadian-Chazyan (Ordovician) hiatus in Tennessee. *Bulletins of American Paleontology* 36, 157–99.

Caster, K. E. & Kjellesvig-Waering, E. N. 1952. Some notes on the genus Dolichopterus Hall. Journal of Paleontology 30, 19–28.

Currie, L. D. 1927. On *Cyamocephalus*, a new Synxiphosuran from the Upper Silurian of Lesmahago, Lanarkshire. *Geological Magazine* 64, 153–7.

Dunlop, J. A. 2002. Arthropods from the Lower Devonian Severnaya Zemlya Formation of October Revolution Island, Russia. *Geodiversitas* 24, 349–79.

Dunlop, J. A., Anderson L. I. & Braddy, S. J. 1999. A new chasmataspid (Chelicerata: Chasmataspida) from the Lower Devonian of the Midland Valley of Scotland. *Transactions of the Royal Society of Edinburgh: Earth Sciences* 89 (for 1998), 161–5.

Dunlop, J. A., Poschmann, M. & Anderson, L. I. 2001. On the Emsian (Early Devonian) arthropods in the Rhenish Slate Mountains: 3.

- The chasmataspid *Diploaspis*. *Paläontologische Zeitschrift* **75** (2), 253–69.
- Dunlop, J. A., Anderson L. I. & Braddy, S. J. 2004. A redescription of *Chasmataspis laurencii* Caster & Brooks, 1956 (Chelicerata: Chasmataspidida) from the Middle Ordovician of Tennessee, USA, with remarks on chasmataspid phylogeny. *Transactions of the Royal Society of Edinburgh: Earth Sciences* 94 (for 2003), 207–25
- Dunlop, J. A. & Braddy, S. J. 2001. Scorpions and their sister-group relationships. *In* Fet, V. & Selden P. A. (eds) *Scorpions 2001*, In Memoriam Gary A. Polis. 1–24. Burnham Beeches: British Arachnological Society.
- Dunlop, J. A. & Selden, P. A. 1997. The early history and phylogeny of the chelicerates. In Fortey, R. A. & Thomas, R. H. (eds). Proceedings of the International Symposium on the Relationships of the Major Arthopod Groups, 221–35. London: Chapman and Hall.
- Dunlop, J. A. & Webster, M. 1999. Fossil evidence, terrestrialization and arachnid phylogeny. *Journal of Arachnology* **27**, 86–93.
- Eldredge, N. 1974. Revision of the Suborder Synziphosurina (Chelicerata, Merostomata), with remarks on merostome phylogeny. *American Museum Novitates* **2543**, 1–41.
- Fahrenbach, W. H. 1999. Merostomata. In Harrison, F. W. & Foelix,
 R. F. (eds) Microscopic anatomy of invertebrates, Vol. 8A.
 Chelicerate Arthropoda, 21–115. New York, NY: Wiley-Liss.
- Farley, R. D. 1999. Scorpiones. In Harrison, F. W. & R. F. Foelix (eds) Microscopic anatomy of invertebrates, Vol. 8A. Chelicerate Arthropoda, 117–222. New York, NY: Wiley-Liss.
- Foelix, R. F. 1996. *Biology of spiders*, 2nd edn. Oxford: Oxford University Press.
- Heubusch, C. A. 1962. Preservation of the intestine in three specimens of *Eurypterus*. *Journal of Paleontology* **36**, 222–4.
- Heymons, R. 1901. Die Entwicklungsgeschichte der Scolopender. *Zoologica*, Lief 2–5, 244 pp.
- Holm, G. 1898. Über die Organisation des Eurypterus fisheri Eichwald. Memoires l'Academie Imperale des Sciences de St.-Petersbourg, VII Serie, Tome 8, 1–57.
- Kjellesvig-Waering, E. N. 1958. Some previously unknown morphological features of *Carcinosoma newlini* (Claypole). *Journal of Paleontology* 32, 295–303.
- Kjellesvig-Waering, E. N. 1963. Pennsylvanian invertebrates of the Mazon Creek area; Illinois: Eurypterida. Fieldiana Geology 12, 169–97
- Novojilov, N. I. 1959. Mérostomes du Dévonien inférieur et moyen de Sibérie. *Annals de la Société Géologique du Nord* **78**, 241–8.
- Novojilov, N. I. 1962. Otrajd Eurypterida. *In* Orlov, J. A. (ed.) *Osnovy Paleontologii-Vlenistronogie, trahejnye I helicerovye*, 404–23. Moscow.
- Paterson, I. B., McAdam, A. D. & MacPherson, K. A. T. 1998. Geology of the Hamilton district. *Memoir for 1:50000 Geological Sheet 23W (Scotland)*. Keyworth, Nottingham: British Geological Survey.
- Plotnick, R. E. 1999. Habitat of Llandoverian-Lochkovian eurypterids. *In Boucot*, A. J. & Lawson, J. D. (eds) *Paleocommunities*:

- a case study from the Silurian and Lower Devonian, 106–31. Cambridge: Cambridge University Press.
- Ruedemann, R. 1921 Presence of alimentary canal in an eurypterid. New York State Museum Bulletin 228, 92–5.
- Salter, J. W. 1855. On some new Crustacea from the Uppermost Silurian Rocks. Quarterly Journal of the Geological Society 12, 26–34.
- Scourfield, D. J. 1937. An anomalous fossil organism, possibly a new type of chordate, from the Upper Silurian of Lesmahagow, Lanarkshire *Ainiktozoon loganense* gen. et sp. nov. *Proceedings of the Royal Society* **B121**, 533–47.
- Størmer, L. 1934. Merostomata from the Downtonian Sandstone of Ringerike, Norway. Skrifter utgitt av Det Norske Videnskapers Akademi i Oslo i Matematisk-Naturvidenskapelig Klasse, 1933 10, 1–125.
- Størmer, L. 1936. Eurypteriden aus dem Rheinischen Unterdevon. Abhandlungen der Preussischen geologischen Landesanstalt 175, 1–74.
- Størmer, L. 1955. Chelicerata. In Moore, R. C. (ed.) Treatise on invertebrate paleontology, (P) Arthropoda 2, 1-41. Boulder, Colorado and Lawrence, Kansas: Geological Survey of America and University of Kansas Press.
- Størmer, L. 1970. Arthropods from the Lower Devonian (Lower Emsian) of Alken an der Mosel, Germany. Part 1: Arachnida. *Senckenbergiana lethaea* **51**, 335–69.
- Størmer, L. 1972. Arthropods from the Lower Devonian (Lower Emsian) of Alken an der Mosel, Germany. Part 2: Xiphosura. *Senckenbergiana lethaea* 53, 1–29.
- Størmer, L. 1974. Arthropods from the Lower Devonian (Lower Emsian) of Alken an der Mosel, Germany. Part 4: Eurypterida, Drepanopteridae and other groups. Senckenbergiana lethaea 54, 359–451.
- Stürmer, W. & Bergström, J. 1981. Weinbergina, a xiphosuran arthropod from the Devonian Hunsrück Slate. Paläontologische Zeitschrift 55, 237–55.
- Tollerton, V. P., Jr. 1989. Morphology, taxonomy, and classification of the Order Eurypterida Burmeister, 1843. *Journal of Paleontology* **63**, 642–57.
- Traquair, R. H. 1899. Report on fossil fishes collected by the Geological Survey of Scotland in the Silurian rocks of the South of Scotland. *Transactions of the Royal Society of Edinburgh* 39, 827–64
- Waterston, C. D. 1979. Problems of functional morphology and classification of stylonuroid eurypterids (Chelicerata, Merostomata) with observations on the Scottish Silurian Stylonuoidea. *Transactions of the Royal Society of Edinburgh* **70**, 251–322.
- Wahlman, G. P. & Caster, K. E. 1978. Bearing of new Texas Upper Cambrian arthropods on merostome classification. Geological Society of America, Abstracts with Programs 10, 268.
- White, E. I. 1946. *Jamoytius kerwoodi* White, a new Chordate from the Silurian of Lanarkshire. *Geological Magazine* **83**, 89–97.

O. ERIK TETLIE and SIMON J. BRADDY, Department of Earth Sciences, University of Bristol, Wills Memorial Building, Queen's Road, Bristol, BS8 1RJ, UK e-mail: o.e.tetlie@bristol.ac.uk; s.j.braddy@bristol.ac.uk

MS received 20 January 2003. Accepted for publication 7 January 2004.