

# Studies on the behaviour of peridomestic and endophagic M form *Anopheles gambiae* from a rice growing area of Ghana

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## Abstract

The ‘paddy paradox’, the occurrence of large populations of vectors but low amounts of malaria transmission where irrigated rice is grown, was investigated in a village in Ghana where M form *Anopheles gambiae* are common. Peridomestic and indoor host-seeking mosquitoes were collected in tent traps and light traps over 21 consecutive nights at the start of the rainy season in June 2009 when the population increased exponentially from less than 100 per night to over 1000. Infection rates in the overall mosquito population were 0.3% and in the estimated parous population were 1.9%. Numbers of *An. gambiae* in the tent trap peaked between midnight and 02:40 am. The majority of insects were taking their first blood meal, as virgins or shortly after mating. More than expected were collected in the light trap during a rainstorm at the start of the rains but overall numbers were not affected. Fewer than expected were collected after a subsequent storm. Recruitment to the adult population decreased over the following days. It is hypothesised that the ‘paddy paradox’ is due to young pre-gravid insects dispersing more widely than gravid ones, not necessarily to low survival in the mosquito.

**Keywords:** *Anopheles gambiae*, rainfall, behaviour, ‘paddy-paradox’, tent trap

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## Introduction

In order to properly understand the epidemiology of malaria, information on the ecology and bionomics of the local vector population is required. Information on factors such as population density, survival rate, infection rate, blood-feeding frequency, time and location of biting, as well as host preference, all affect the ability of a mosquito population to

transmit malaria. In West Africa, the most important vectors are the M and S forms of *Anopheles gambiae*. These are ‘incipient’ species in which hybrids are fertile and can be readily produced in the laboratory. Where they occur in sympatry in the wild, however, they often show low rates of hybridization. (Diabate *et al.*, 2009).

To paraphrase Voltaire, ‘If *Anopheles gambiae* did not exist, man would have created him’. Humans created the larval ecological niche, puddles in exposed forest, exploited by *An. gambiae*. We have also provided shelter and food for the adults since we first arrived on the planet but especially since the beginning of agriculture when the species went through a

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population expansion (Donnelly *et al.*, 2001). The creation of other larval habitats with expanding agriculture may be an evolutionary driving force in the *An. gambiae* complex in general and in *An. gambiae* s.s. in particular. The M and S forms differ in their most commonly used breeding sites. The pan-African S form predominates in temporary pools, the archetypal *An. gambiae* breeding site, whilst high population densities of M form *An. gambiae* are associated with rice cultivation. In many such irrigated sites, despite the large numbers of mosquitoes, transmission is low, leading to the so called 'paddy paradox' (Ijumba & Lindsay, 2001).

Examination of the behaviour of young mosquitoes has rarely been undertaken (Charlwood *et al.*, 2003a), and the reason for the relatively low survival and infection rates in mosquitoes from rice growing areas remains unknown. It may have something to do with their behaviour when newly emerged. From their emergence as adults to the time of their first oviposition, female *An. gambiae* undergo a number of physiological changes such as a rise in juvenile hormone (Noriega, 2004) and perform a number of 'one-off' behaviours, including mating and the taking of a pre-gravid blood meal, that are not repeated in subsequent oviposition cycles.

Should, as a result of these activities, they enhance the likelihood of their acquiring an infection, it may affect eventual transmission since, assuming age specific mortality rates, these are the mosquitoes that are most likely to survive to become vectors. In previous studies, parous/nulliparous rates, obtained by dissection at dispersed intervals, have been used to assess mosquito survival. More detailed dissections, undertaken on a nightly basis, provide more detailed information (Charlwood *et al.*, 1985; Holmes & Birley, 1987; Mutero & Birley, 1989).

Mosquitoes taking their first blood meal can be distinguished from other age groups by the early state of development of their ovaries in which follicles have little or no yolk and which are surrounded by tightly coiled trachea. They can, by examination of the reproductive tract, be further separated into virgins, insects that have recently mated (with a mating plug) and those that are mated but in which a mating plug is not seen. Whilst mating *per se* is unlikely to affect biting behaviour in anophelines (Klowden, 2001; Charlwood *et al.*, 2003b), information on the proportion of first-feeding insects that feed as virgins or that have recently mated can provide insights into what a female does following emergence.

Differences in behaviour between M and S form *An. gambiae* when newly emerged may affect their eventual vectorial capacity. Factors that might influence the likelihood of acquiring an infection are time and place that the mosquitoes feed, late biting endophagic insects being more likely to bite humans than exophagic early biting ones. The development of risk-free tent traps, such as the Furvela trap (Govella *et al.*, 2009), for the collection of biting mosquitoes before they enter houses enables a comparison between indoor and outdoor biting mosquitoes to be easily performed. The trap, which collected similar numbers of S form *An. gambiae* to the CDC light trap when tested in Tanzania (Govella *et al.*, 2009), has yet to be tested in West Africa where M form *An. gambiae* predominate.

Both M and S form *An. gambiae* at high densities have been described from Okyereko, a village in an irrigation scheme in Ghana (Yawson *et al.*, 2004, 2007; Okoye *et al.*, 2005). Temperature is one factor that appears to determine the distribution of M and S forms of *An. gambiae* in Ghana, the M form being apparently better adapted to higher temperatures than

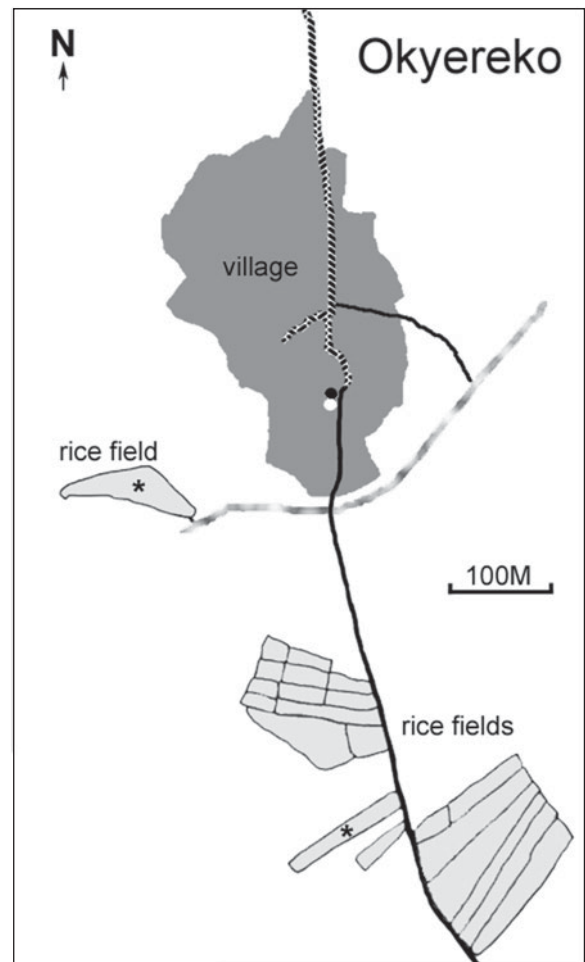


Fig. 1. Map of Okyereko showing the location of sample sites and a selection of fields. The other fields to the south, east and west of the village were not mapped. The rice paddies observed to contain large numbers of *Anopheles gambiae* larvae at the start and end of the study are indicated by a star (—, canal; - - - - - , road; ·····, footpath; ●, tent-trap; ○, light-trap; \*, larvae).

the S form (De Souza *et al.*, 2010). We therefore examined the population dynamics of both endophagic and exophagic mosquitoes at the start of the rains from Okyereko and applied a more detailed dissection than has previously been applied to these mosquitoes (Dzodzomenyo & Simonsen, 1999). A possible explanation for the paddy paradox is provided.

## Methods

Okyereko (5° 24.87'N, 0° 36.25'W), some 70 km to the west of Accra, consists of 80 relatively run-down cement houses, 5 km from the coast, and is bordered on two sides by extensive irrigated rice fields. According to Yawson *et al.* (2007), the village had 35% S to 65% M form *An. gambiae* whilst, according to Dzodzomenyo & Simonsen (1999), the proportion was 99% M form. During the study, some of the fields, each 100 × 50 m in size, were being harvested whilst others were recently planted. At the start of the study, one field in particular, 350 m to the south of the collection sites, with rice at an early stage of cultivation (fig. 1), contained large numbers of small pools,

Table 1. Total number of mosquitoes collected by species in tent and light traps, Okyereko, Ghana, June 2009.

	Anopheles				Culex			Mansonia
	<i>gambiae</i>	<i>funestus</i>	<i>pharoensis</i>	<i>zeimanni</i>	<i>quinquefasciatus</i>	<i>tritaeniorhynchus</i>	sp.	<i>africana</i>
Tent	5532	69	130	3	557	54	5	24
Light	7761	91	48	4	725	83	0	185

each with large numbers of larvae of *An. gambiae* s.l. By the end of the study, this was no longer suitable for mosquito larvae (the rice having grown and the water having largely dried up), but pools in another set of recently cultivated fields, 200 m to the west of the collection sites, contained mosquito larvae (fig. 1).

Peridomestic and indoor biting mosquitoes were collected with a Furvela tent trap and a CDC light trap, respectively. The traps sample mosquitoes at slightly different phases of the hunting cycle. Tent traps catch mosquitoes that are attracted to odour (i.e. in the initial approach to a bait), rather like the Odour Baited Entry Trap (OBET) of Costantini *et al.* (1993), with the difference that no entry behaviour is required of the insect, whilst light traps catch hungry mosquitoes in the act of leaving a bedroom after an unsuccessful attempt to feed on a host protected by a mosquito bednet. Some species may also be attracted to light *per se*, which might bias the light trap results whilst the way the mosquitoes respond to the tent trap may reflect a similar approach to houses rather than a completely exposed host in the open. Nevertheless, previous results indicate that the Furvela tent trap and light traps have a very similar sampling efficiency (Govella *et al.*, 2009).

During the study, a CDC light trap was run on a daily basis inside the bedroom of a house close to the edge of the village. The trap was hung ~1.5 m from the floor, close to the end of the bed in which the householder slept under a mosquito net. The trap was run from 18:00 h to 06:30 h of the next day.

A slightly modified version of the Furvela tent trap described by Govella *et al.* (2009) for the collection of outdoor biting mosquitoes was also run on a daily basis. The trap differed in that wire, instead of string, was used to attach the trap to the tent. A three-person bell tent, with two adult hosts (JDC & EVET) inside it was used. On five nights, the collection bag on the tent trap was changed at three-hourly intervals, enabling an assessment of biting activity by time to be made.

Following identification and separation into species or species group, females of the *An. gambiae* complex were separated according to abdominal condition and dissected under a stereo-microscope using transmitted light to determine their gonotrophic age. Based on the appearance of their ovaries, female mosquitoes were separated into the following classes:

- (i) First blood-feeding
  - Virgin*: spermatheca empty, ovarioles Stage I.
  - Plug*: sperm in the spermatheca, a mating plug in the common oviduct, ovarioles Stage I.
  - Nulliparous I*: sperm in the spermatheca but without a mating plug ovarioles at Stage I.
- (ii) Second blood-feeding
  - Plug-blood*: mating plug present, old dark blood from previous feed in stomach.
  - Nulliparous II*: spermatheca with sperm, mating plug not present, ovarioles Stage II, i.e. yolk present in terminal ovariole.

- (ii) Subsequent blood-feeding

*Parous with sacs*: a sac with some distension still present, indicating that the mosquito had returned to feed shortly after oviposition.

*Parous without sacs*: the sac from the previous oviposition had contracted, indicating that there had been a delay between oviposition and re-feeding.

The presence of retained Stage V eggs in parous females, according to the sac stage, was also noted.

Estimated numbers of each age group collected were determined by multiplication of the number caught in a trap by the proportion in that category.

Samples of the *An. gambiae* complex were stored over silica gel for later species and form determination and for an assessment of infection with *Plasmodium falciparum*.

DNA extraction was performed individually following Collins *et al.* (1988). DNA samples were used to determine the molecular form by polymerase chain reaction (PCR)-restriction fragment length polymorphism (RFLP) (Fanello *et al.*, 2002).

The presence of circumsporozoite (CS) antigens of *P. falciparum* was determined using the sandwich enzyme-linked immunosorbent assay (ELISA) using the protocols of Wirtz *et al.* (1987).

Rainfall data was obtained from the National Meteorological Service at Winneba, 5 km to the west of Okyereko, and temperature data was obtained from Apam, 30 km to the east of Okyereko (no thermometer being available in Winneba).

The village boundaries, selected rice fields, and locations of tent and light traps were marked with a hand-held GPS unit (Garmin).

## Results

A total of 235 individual *An. gambiae* s.l. were used for species and molecular form identification, half of them were collected with the tent trap and the other half with a light trap. Of these, 234 were *An. gambiae* s.s., (230 (98%) being M molecular form, three S form, one an M/S hybrid and one was *An. melas*). All four S form specimens, including the hybrid, were collected indoors. Given their overwhelming frequency, in all further discussion, it is assumed that we were dealing only with M form *An. gambiae*.

The total number of mosquitoes collected in tent and light trap is shown in table 1. In both traps, *An. gambiae* was by far the most common mosquito collected.

The great majority of the mosquitoes collected were unfed. Only 3.4% of the mosquitoes in the tent trap (including an unspecified number of those seen to enter the tent when collection bags were being changed) and 2.8% in the light trap were either blood fed or part fed. A total of 2110 *An. gambiae* s.l. were dissected, 795 from the light trap and 1315 from the

Table 2. Age structure of unfed M form *An. gambiae* collected from tent and light traps, Okyereko, Ghana, June 2009.

	Virgin	Plug	Nulliparous Stage I	Nulliparous Stage II	Parous with sacs	Parous without sacs	Parous rate
Tent trap	203	334	216	153	136	269	0.33
Light trap	110	200	119	99	80	183	0.30

Fisher's exact test (two-tail) virgin/plug ratios tent/light trap  $P=0.507$ ; sac/no-sac tent/light trap  $P=0.399$ ; NI/NII tent/light trap  $P=0.38$ .

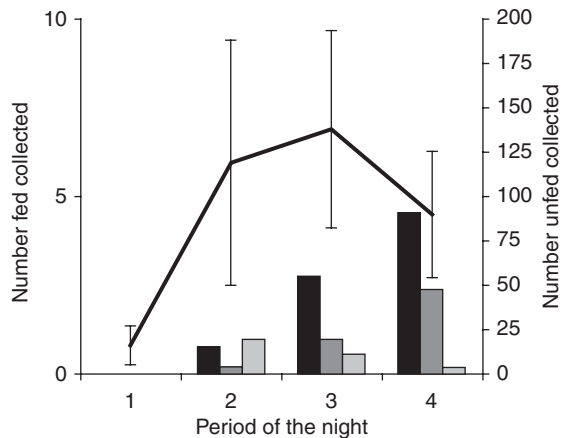


Fig. 2. Outdoor biting activity of M form *An. gambiae* determined using the Furvela tent trap, Okyereko, Ghana, June 2009 (■, part-fed; ■, engorged; □, gravid; —, unfed).

tent trap. Parous rates from both collections were very low throughout the experiment (table 2).

Mean numbers of unfed, part-fed, engorged and gravid females collected in the tent trap by night time from five collections in which collection bags were changed are shown in fig. 2. Most unfed and gravid females were collected in the middle two periods of the night, whereas numbers of part-fed and engorged insects, although small, increased during the night. The proportion of the collection that was parous decreased during the night (fig. 3) although the proportion of these that had sacs remained more or less constant, as did the proportion of first-feeding insects with mating plugs.

On one night of collection, the battery used in the light trap was not properly charged and, although numbers collected were sufficient for the purposes of dissection, they were a fraction of the number expected. Hence, the collection data from the light trap on this day was not included in the analysis.

Overall, approximately two thirds (0.62) of recently emerged mosquitoes had mating plugs in both traps (192 of 311 dissected from the light trap and 354 of 574 dissected from the tent trap). Similarly, there were no differences between overall proportions of the other age groups collected in the two traps. This again implies that both traps are sampling the same population of mosquitoes.

The wet season started on the 11th of June (148 mm being recorded in Winneba, the town 5 km from Okyereko). More *An. gambiae* than expected were caught in the light trap and less in the tent trap on this night. The sum of the numbers collected in both traps was, however, similar to the expected number, as was the proportion of newly emerged insects that had a mating plug (0.64). Thus, the rain had not

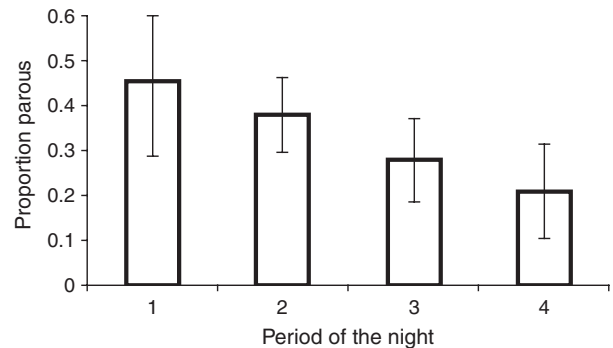


Fig. 3. Parous rates by collection period of the night of M form *An. gambiae* collected using the Furvela tent trap, Okyereko, Ghana, June 2009.

apparently affected mating behaviour nor overall host seeking activity.

After the 11th, a total of 251.2 mm of rain was recorded on 11 of the remaining 13 days of the study. On four of these days, two of them at night, more than 20 mm of rain fell. On those two nights, the proportion of the total collection was lower than usual in the tent trap (fig. 4). The number of mosquitoes in both traps was also less than expected on the 19th of June when 144.2 mm of rain fell from the evening until 02:40 am on the 20th, although the drop was greatest in the tent trap. The proportion of newly emerged insects with a mating plug was higher on this day than on all other days (0.9 having plugs compared to a mean of 0.63 on the other days). The correlation coefficient between the numbers collected in the light trap and tent trap for all nights (including those when more than 5 mm of rain fell) was 0.506. This improved to 0.761 when the nights when more than 5 mm of rain fell were excluded from the calculations. Variation in numbers collected in both traps was also greater in days subsequent to the rainstorm on the 19th than in the period leading up to it. After the rainstorm, numbers of *An. gambiae* in the tent trap declined, whilst numbers in the light trap continued to increase (fig. 5).

Parous mosquitoes classified as having sacs were more likely to have retained Stage V (i.e. unlaidd) eggs in their ovaries than mosquitoes classified as not having sacs (two-tailed probability from a Fisher's exact test  $P=0.005$ ) (table 3). This implies that such eggs are voided with time after oviposition as the sacs contract. Mean air temperatures in Apam decreased during the study (from a mean of 28.4°C on the 9th of June to 24.4°C on the 16th), as did the proportion of mosquitoes dissected without sacs ( $r^2$  between the proportion of parous insects with sacs and mean temperature = 0.299), giving rise to a lowered estimation of the mean oviposition cycle length over time (fig. 6).



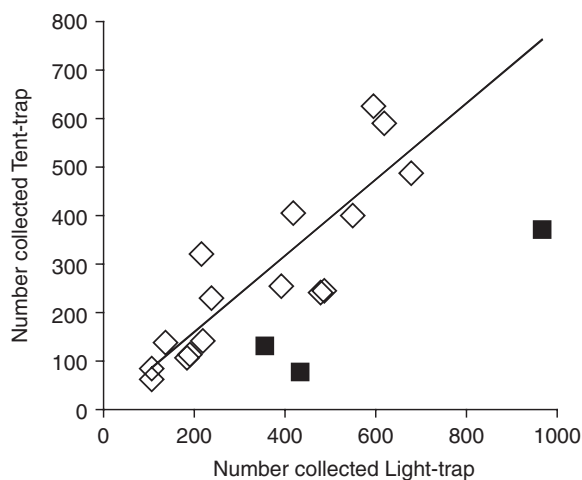


Fig. 4. Relationship between the numbers of M form *An. gambiae* collected in light trap and tent trap on nights with and without rain, Okyereko, Ghana, June 2009. (The regression equation for the trend line between the two traps is  $0.785x + 1.5078$ .) ( $\diamond$ , No-rain;  $\blacksquare$ , Rain).

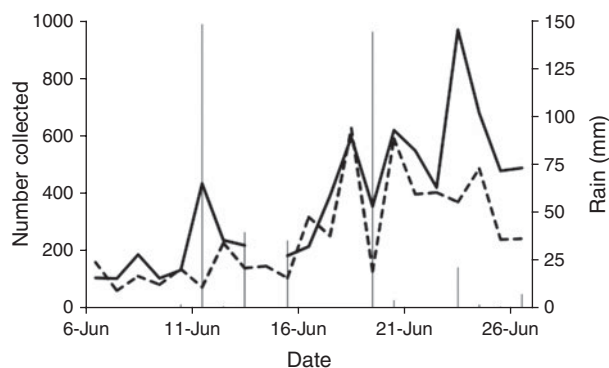


Fig. 5. Numbers of M form *An. gambiae* collected in CDC light traps indoors and Furlvela tent traps outdoors and daily rainfall, Okyereko, Ghana, June 2009 (—, Light trap; ---, Tent trap).

As expected from the low parous rates, infection in the mosquito was low. No oocysts were seen on the stomachs of parous insects dissected and only four (0.3%) of the 1296 mosquitoes analyzed in the ELISA tested positive for *P. falciparum* circumsporozoite protein. All of the positive mosquitoes came from the 753 tested from the light trap, each of the three days of collection yielding at least one positive mosquito, compared to none of the three days tested for the tent trap. Among the expected parous population from these days, overall sporozoite rates increased to 1.9%, whilst the rate from the light-trap samples increased to 5.2%.

## Discussion

Our results indicate that, in both East and West Africa, on calm nights, the Furlvela tent trap and CDC light trap sample similar fractions of the local *An. gambiae* populations. The tent trap has a number of advantages over the light trap, particularly for spatial studies. It collects host-seeking mosquitoes

Table 3. Number of parous M form *An. gambiae* with and without sacs and numbers with retained Stage V eggs dissected, Okyereko, Ghana, June 2009.

	With sac	Ret V	Without sac	Ret V
Tent-trap	136	12	269	15
Light-trap	80	15	183	10

Fisher's exact test (two-tail)  $P = 0.005$ .

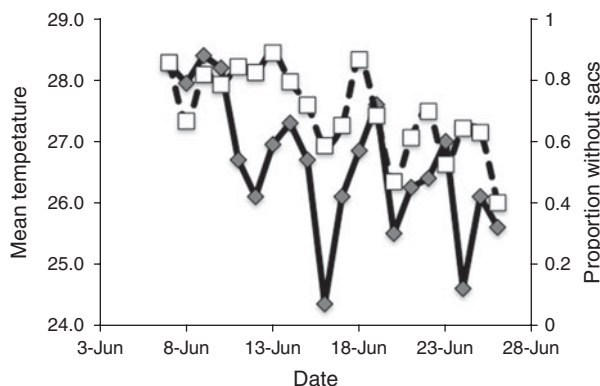


Fig. 6. Proportion of parous M form *An. gambiae* from both light and tent traps in Okyereko, Ghana, returning to feed without large ovariole sacs and mean daily temperature measured in Apam, 30 km to the west, June 2009 ( $\diamond$ —, mean temp).

that are attracted to kairomones rather than relying on light. Light traps collect mosquitoes in the act of departing from a house and can only be hung where there are houses. The kind of house in which a trap is hung influences the number of mosquitoes entering, hence the number caught. Tent traps, on the other hand, provide a uniform sample, and can, within reasonable limits, be located anywhere. They also use considerably less power than a light trap but do require the payment of a sleeper that increases their running costs.

The population of female *An. gambiae*, an archetypal 'invasive' species, went through a log-scale change in density during the three weeks of the study. Given this increase, it is not surprising that parous rates and malaria transmission were low and similar to the rates observed by Okoye *et al.* (2005) who found four of 2411 (0.2%) *An. gambiae* from Okyereko positive for sporozoites. Such low sporozoite rates have been described from other rice-growing areas of West Africa (Robert *et al.*, 1985, 1987; Ijumba & Lindsay, 2001; Diuk-Wasser *et al.*, 2007). Although in the study presented here we could not distinguish between catastrophic mortality in young insects vs. exodus from the study area, our data on parous rates and sporozoite rates provides insight into what might be happening in the population. Assuming age-specific mortality (Clements & Paterson, 1981), young insects should be the age group whose survival is at a premium, and even small mosquitoes that manage to take a blood meal should survive as well as any (Takken *et al.*, 1998a). Although sporozoite rates among the whole population were low, when only the expected parous population was taken into consideration, rates were typical for a population of African vector. Similarly, when first-feeding insects were removed from survival rate calculations, an estimate similar to other studies was obtained

(Gillies & Wilkes, 1965; Charlwood *et al.*, 1995, 1997; Takken *et al.*, 1998b). This implies that, in older age groups, survival was unexceptional, which itself implies that rather than dying, an excess of young insects were leaving the area. Dispersal of young *An. arabiensis* was also observed in Tanzania, a 'pulse' of insects being collected along a transect of traps into the village of Namawala following a mass emergence at the periphery (Takken *et al.*, 1998b). Greater dispersion of young *An. culicifaces*, compared to old ones, was also observed in Sri Lanka by Rawlings *et al.* (1981). Such a phenomenon may be occurring, but with overlapping generations, in Okyereko.

Dispersion of recently mated first-fed (pre-gravid) mosquitoes would explain the dearth of 'plug-blood' mosquitoes in the collection. Such a phenomenon does not necessarily require radically different behaviours among insects of different ages. Gravid mosquitoes might fly toward oviposition sites at a variable distance from the village by responding to, for example, oviposition-site odours or humidity gradients, whereas first-blood feeders might be expected to fly in less specific directions, responding to mating cues in the case of virgins with a first blood meal, or to host cues in the case of mated females that have taken just the first blood meal. This might lead to more of one group than the other leaving the study area, depending on the distribution of resources and the associated cues that stimulate directed movement. There is, however, no information available on the pattern of movement of gravid females to support or challenge this hypothesis.

Whether the increase in numbers collected during the study was due to eclosion from several breeding sites or to the maturation of a single site is also unknown. Bearing in mind its limitations, the data indicates that the former took place. Certainly, the very productive field observed at the start of the study was no longer producing adult mosquitoes in any number by the end of the study. Not only had it dried up substantially but also the rice had grown from short to medium height. Changes observed in a number of population parameters indicate that at least two separate processes were occurring during the period of observation. The first is the drop in the estimated duration of the oviposition cycle (fig. 6) and the second is the rise in the proportion of first feeding insects (with follicles at Stage I) that were collected mated but without a mating plug (data not shown).

Temperature appears to determine the distribution of M and S forms of *An. gambiae* in Ghana (De Souza *et al.*, 2010), and it may have influenced the behaviour of the mosquitoes in the present study. Thus, the change in the observed proportion of parous mosquitoes collected with sacs could be associated with one or more of the following factors: a drop in temperatures as a result of the rain, slowing of the contraction of sacs, or that the population of mosquitoes was generally ovipositing closer to the collection site during the latter period of observation. Further studies determining the rates of sac contraction at different temperatures and mapping of breeding sites would help elucidate these points. We do not know why, among first-feeding mosquitoes collected in the first part of the study, a higher proportion had apparently mated earlier than those that were collected in the latter part of the study. Given the high densities of larvae observed in Okyereko, independent estimates of eclosion could be made using emergence-traps.

Heavy rainstorms tend to have a negative effect on mosquito populations, since they drown or flush out the larvae (Paaijmans *et al.*, 2007); but this, to a certain extent, depends on

the terrain and the level of the water table. In the present study, the two rainstorms, on the 11th and 19th of June, had different effects on the mosquito population. On the 11th of June, the behaviour of the *An. gambiae* was altered such that they were collected in greater numbers than expected in the light trap, whilst fewer than expected were collected in the tent trap. Overall, though, the rain did not affect the total number collected on this and subsequent days. Such an outcome in response to rain may enhance subsequent malaria transmission since the mosquitoes are more likely to take an infected bloodmeal inside than they are outside the house, where alternative hosts are to be found.

The rainstorm on the 19th had more profound effects. It severely reduced the numbers collected on that night and was associated with a greater subsequent variability in numbers in both traps. The water from the first rainstorm was largely absorbed within hours. Subsequent rain left numerous puddles in the village. The higher water table on the 19th June may have resulted in some flushing of late larvae in the rice fields, reducing numbers emerging and, hence, the numbers in the collections. The very high proportion of mated mosquitoes collected on that night was probably due to a deficit of virgin females in the collection rather than an exceptional amount of mating occurring.

A short but heavy rainstorm (71.5 mm of rain during three hours during the night) was shown to inhibit host-seeking behaviour, without affecting mortality, among *An. farauti* from Papua New Guinea (Charlwood *et al.*, 1988). In that case, however, mosquitoes survived and merely delayed coming to feed until the following day.

Despite the eventual presence of numerous puddles within the confines of the village, none were observed with anopheline larvae. These were sites typically preferred by the S form of *An. gambiae*. Less than 2% S form *An. gambiae* were collected during the study. These may have started to colonise the puddles, but numbers may have still been too low for them to be detected during the larval surveys.

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### References

- Charlwood, J.D., Birley, M.H., Dagar, H., Paru, R. & Holmes, P.R. (1985) Assessing survival rates of *Anopheles farauti* (Diptera, Culicidae) from Papua New Guinea. *Journal of Animal Ecology* **54**, 1003–1016.
- Charlwood, J.D., Graves, P.M. & Marshall, T.F. de C. (1988) Evidence for a 'memorised' home range in *Anopheles farauti* females from Papua New Guinea. *Medical and Veterinary Entomology* **2**, 101–108.
- Charlwood, J.D., Kihonda, J., Sama, S., Billingsley, P.F., Hadji, H., Verhave, J.P., Lyimo, E., Luttkhuizen, P.C. & Smith, T. (1995) The rise and fall of *Anopheles arabiensis* (Diptera, Culicidae) in a Tanzanian village. *Bulletin of Entomological Research* **85**, 37–44.

- Charlwood, J.D., Billingsley, P.F., Takken, W., Lyimo, E.O.K., Smith, T. & Meuwissen, J.H.E.T. (1997) Survival and infection probabilities of anthropophilic Anophelines from an area of high prevalence of *Plasmodium falciparum* in humans. *Bulletin of Entomological Research* **87**, 445–453.
- Charlwood, J.D., Pinto, J., Sousa, C.A., Ferreira, C., Petrarca, V. & do Rosario, V.E. (2003a) A mate or a meal? – Pre-gravid behaviour of female *Anopheles gambiae* from the islands of São Tomé and Príncipe, West Africa. *Malaria Journal* **2**, 7.
- Charlwood, J.D., Pinto, J., Sousa, C.A., Ferreira, C., Gil, V. & de Rosario, V. (2003b) Mating does not affect the biting behaviour of *Anopheles gambiae* from the islands of São Tomé and Príncipe, West Africa. *Annals of Tropical Medicine and Parasitology* **97**, 751–756.
- Clements, A.N. & Paterson, G.D. (1981) The analysis of mortality and survival rates in wild populations of mosquitoes. *Journal of Applied Ecology* **18**, 373–399.
- Collins, F.H., Petrarca, V., Mpfu, S., Brandling-Bennett, A.D., Were, J.B., Rasmussen, M.O. & Finnerty, V. (1988) Comparison of DNA probe and cytogenetic methods for identifying field collected *Anopheles gambiae* complex mosquitoes. *American Journal of Tropical Medicine & Hygiene* **39**, 545–550.
- Costantini, C., Gibson, G., Brady, J., Merzagora, L. & Coluzzi, M. (1993) A new odour-baited trap to collect host-seeking mosquitoes. *Parassitologia* **35**, 5–9.
- De Souza, D., Kelly-Hope, L., Lawson, B., Wilson, M. & Boakye, D. (2010) Environmental factors associated with the distribution of *Anopheles gambiae* s.s. in Ghana; an important vector of lymphatic filariasis and malaria. *PLoS ONE* **5**(3), e9927.
- Diabate, A., Dao, A., Yaro, A.S., Adamou, A., Gonzalez, R., Manoukis, N.C., Traoré, S.F., Gwadz, R.W. & Lehmann, T. (2009) Spatial swarm segregation and reproductive isolation between the molecular forms of *Anopheles gambiae*. *Proceedings of Biological Sciences* **7**, 4215–4222.
- Diuk-Wasser, M., Touré, M.B., Dolo, G., Bagayoko, M., Sogoba, N., Sissoko, I., Traoré, S.F. & Taylor, C.E. (2007) Effect of rice cultivation patterns on malaria vector abundance in rice-growing villages in Mali. *American Journal of Tropical Medicine and Hygiene* **76**, 869–874.
- Donnelly, M.J., Licht, M.C. & Lehmann, T. (2001) Evidence for recent population expansion in the evolutionary history of the malaria vectors *Anopheles arabiensis* and *Anopheles gambiae*. *Molecular Biology and Evolution* **18**, 1353–1364.
- Dzodzomenyo, M. & Simonsen, P.E. (1999) Bancroftian filariasis in an irrigation project community in southern Ghana. *Tropical Medicine and International Health* **4**, 13–18.
- Fanello, C., Santolamazza, F. & della Torre, A. (2002) Simultaneous identification of species and molecular forms of the *Anopheles gambiae* complex by PCR-RFLP. *Medical & Veterinary Entomology* **16**, 461–464.
- Govella, N.J., Chaki, P.P., Geissbuehler, Y., Kannady, K., Okumu, F., Charlwood, J.D., Anderson, R.A. & Killeen, G.F. (2009) A new tent trap for sampling exophagic and endophagic members of the *An. gambiae* complex. *Malaria Journal* **8**, 157.
- Gillies, M.T. & Wilkes, T.J. (1965) A study of age composition of population of *Anopheles gambiae* Giles and *Anopheles funestus* Giles in North-eastern Tanzania. *Bulletin of Entomological Research* **56**, 129–135.
- Holmes, P.R. & Birley, M.H. (1987) An improved method for survival rate analysis from time series of haematophagous Dipteran populations. *Journal of Animal Ecology* **56**, 427–440.
- Ijumba, J.N. & Lindsay, S.W. (2001) Impact of irrigation on malaria in Africa: Paddies paradox. *Medical and Veterinary Entomology* **15**, 1–11.
- Klowden, M.J. (2001) Sexual receptivity in *Anopheles gambiae* mosquitoes: absence of control by male accessory gland substances. *Journal of Insect Physiology* **47**, 661–666.
- Mutero, C.M. & Birley, M.H. (1989) The effect of pre-gravid development on the estimation of mosquito survival rates. *Journal of Applied Entomology* **107**, 96–101.
- Noriega, F.G. (2004) Nutritional regulation of JH synthesis: a mechanism to control reproductive maturation in mosquitoes? *Insect Biochemistry and Molecular Biology* **34**, 687–693.
- Okoye, P.N., Wilson, M.D., Boakye, D.A. & Brown, C.A. (2005) Impact of the Okyereko irrigation project in Ghana on the risk of human malaria infection by *Anopheles* species (Diptera: Culicidae). *African Entomology* **13**, 249–253.
- Paaijmans, K.P., Wandango, M.O., Githeko, A.K. & Takken, W. (2007) Unexpected high losses of *Anopheles gambiae* larvae due to rainfall. *PLoS One* **2**: e1146.
- Rawlings, P., Curtis, C.F., Wickramasinghe, M.B. & Lines, J. (1981) The influence of age and season on dispersal and recapture of *Anopheles culicifacies* in Sri Lanka. *Ecological Entomology* **6**, 307–319.
- Robert, V., Gazin, P., Boudin, C., Molez, J.F., Ouedraogo, V. & Carnevale, P. (1985) La transmission du paludisme en zone de savane arboree et en zone rizicole des environs de Bobo Dioulasso (Burkina Faso). *Annales de Societe Belge Medicines Tropicales* **65**(Supplement 2), 201–214.
- Robert, V., Gazin, P. & Carnevale, P. (1987) Malaria transmission in three sites surrounding the area of Bobo-Dioulasso (Burkina Faso): The savanna, a rice field and the City. *Bulletin of the Society of Vector Ecology* **12**, 541–543.
- Takken, W., Klowden, M.J. & Chambers, G.M. (1998a) Effect of body size on host seeking and blood meal utilization in *Anopheles gambiae* sensu stricto (Diptera: Culicidae): The disadvantage of being small. *Journal of Medical Entomology* **35**, 639–645.
- Takken, W., Charlwood, J.D., Billingsley, P.F. & Gort, G. (1998b) Dispersal and survival of *Anopheles funestus* and *A. gambiae* s.l. (Diptera, Culicidae) during the rainy season in southeast Tanzania. *Bulletin of Entomological Research* **88**, 561–566.
- Wirtz, R., Zavala, F., Charoenvit, Y., Campbell, G.H., Burkot, T.R., Schneider, I., Esser, K.M., Beaudoin, R.L. & Andre, R.G. (1987) Comparative testing of *Plasmodium falciparum* sporozoite monoclonal antibodies for ELISA development. *Bulletin WHO* **65**, 39–45.
- Yawson, A.E., McCall, P.J., Wilson, M.D. & Donnelly, M.J. (2004) Species abundance and insecticide resistance of *Anopheles gambiae* in selected areas of Ghana and Burkina Faso. *Medical and Veterinary Entomology* **18**, 372–377.
- Yawson, A.E., Weetman, D., Wilson, M.D. & Donnelly, M.J. (2007) Ecological Zones Rather Than Molecular Forms Predict Genetic Differentiation in the Malaria Vector *Anopheles gambiae* s.s. in Ghana. *Genetics* **175**, 751–761.