Helminth egg excretion with regard to age, gender and management practices on UK Thoroughbred studs

V. E. RELF¹*, E. R. MORGAN², J. E. HODGKINSON³ and J. B. MATTHEWS¹

¹ Moredun Research Institute, Pentlands Science Park, Midlothian EP26 0PZ, UK

² School of Biological Sciences, University of Bristol, Woodland Road, Bristol BS8 1UG, UK

³ Institute of Infection and Global Health, University of Liverpool, Liverpool L3 5RF, UK

(Received 6 July 2012; revised 30 August and 16 October 2012; accepted 16 October 2012; first published online 25 January 2013)

SUMMARY

Few studies have described the combined effect of age, gender, management and control programmes on helminth prevalence and egg shedding in grazing equines. Here, fecal samples collected from 1221 Thoroughbred horses, residing at 22 studs in the UK, were analysed. The distribution of strongyle eggs amongst individuals in relation to age, gender and management practices was investigated. Fecal worm egg counts (FWECs), described as the number of eggs per gramme (epg) of feces, were determined using a modification of the salt flotation method. The FWEC prevalence (mean%) of strongyles, *Parascaris equorum*, tapeworm spp. and *Strongyloides westeri* was 56, 9, 4 and 8%, respectively. Strongyle, *P. equorum*, tapeworm spp. and *Strongyloides westeri* was 56, 9, 4 and 8%, respectively. Strongyle, respectively. Within all age and gender categories, strongyle FWECs were highly over-dispersed (arithmetic mean = 95 epg, aggregation parameter k=0.111) amongst horses. Animal age, last anthelmintic type administered and management practices (for example, group rotation on grazing) most strongly influenced strongyle prevalence and level of egg shedding (P < 0.05). Overall, 11% of equines (range: 234–2565 epg) were responsible for excreting 80% of the strongyle eggs detected on FWEC analysis. The results confirm that the judicious application of targeted treatments has potential to control equine strongyle populations by protecting individual horses from high burdens, whilst promoting refugia for anthelmintic susceptible genotypes.

Key words: horse, helminths, prevalence, fecal egg shedding, age, gender, management practices, aggregation.

INTRODUCTION

Grazing equines are exposed to a number of helminth infections throughout their lives. Of these, cyathostomins (small strongyles) are considered the most problematic (Love et al. 1999). Since the introduction of macrocyclic lactone (ML) anthelmintics in the 1980s, cyathostomins have been reported as the most prevalent nematodes in equine populations, accounting for over 95% of the nematode infections detected (Lind et al. 1999; Kornas et al. 2010). Of the 3 modern anthelmintic classes (benzimidazoles (BZs), tetrahydropyrimidines (THPs) and MLs) licensed for use in equines, widespread cyathostomin resistance has been recorded against 2 (BZs and THPs), with ML resistance emerging (reviewed by Stratford et al. 2011; von Samson-Himmelstjerna, 2012). Reduced efficacy of the ML, ivermectin, against the small intestinal nematode Parascaris equorum is also an issue in equine populations containing large numbers of young horses (von Samson-Himmelstjerna, 2012). Thoroughbred (TB) breeding studs in the UK rely very heavily on anthelmintic

Parasitology (2013), **140**, 641–652. © Cambridge University Press 2013 doi:10.1017/S0031182012001941

treatments to control helminth infections: many premises administer frequent treatments to all animals, often using the same anthelmintic class (Relf *et al.* 2012). Such practices facilitate anthelmintic resistance (Kuzmina and Kharchenko, 2008). Should resistance to all 3 classes occur, this will have a major impact on equine health as no new products appear to be under development for horses and reversion to anthelmintic sensitivity does not readily occur in parasitic nematodes (Matthews, 2008).

The assessment of helminth distribution patterns in managed equine populations will yield useful information for developing improved control methods that are less reliant on chemical compounds. Previous studies indicate that, within populations, a relatively small proportion of individual horses are responsible for excreting the majority of strongyle eggs and that there is an element of consistency in the excretion patterns (Nielsen et al. 2006). It is proposed that by identifying animals regarded as 'high egg shedders', this will enable farms and studs to implement more targeted treatment approaches to helminth control. The acquisition of information on natural distribution patterns will help in establishing appropriate fecal worm egg count (FWEC) thresholds at which horses should be treated with

^{*} Corresponding author: Moredun Research Institute, Pentlands Science Park, Midlothian EP26 0PZ, UK. Tel: +44 131 445 5111. Fax: +44 131 445 6235. E-mail: valerie. relf@moredun.ac.uk

an adulticidal anthelmintic (commonly quoted as 200 eggs per gramme, epg; Uhlinger, 1993). Minimizing unnecessary anthelmintic treatments in animals excreting no or low numbers of eggs at certain times of year has been proposed to increase the proportion of parasites unexposed to anthelmintic (referred to as refugia), an important component in delaying anthelmintic resistance (van Wyk, 2001).

Several studies describe host factors influencing strongyle egg excretion in horses (Döpfer et al. 2004; Becher et al. 2010), as well as those environmental factors that affect development and survival of the free-living stages (reviewed by Nielsen et al. 2007); however, few describe the combined effect of age, gender and management on the prevalence of infection as determined by FWEC analysis (Kornas et al. 2010). To our knowledge, there are no published studies that have focused solely on breeding TBs, in which frequent anthelmintic administration is commonplace (Relf et al. 2012). On TB studs, the incidence of infection is likely to be elevated due to the high proportion of young animals, which have been demonstrated to have a higher odds ratio risk for strongyle egg shedding (Hinney et al. 2011). The aim here was to investigate the prevalence of patent helminth infections on a cohort of UK TB studs and to assess the level of strongylid and P. equorum egg shedding in relation to age, gender and management. The distribution of strongyle FWECs amongst individuals was also explored across age and gender categories. Helminth distribution is almost universally over-dispersed within host populations (Shaw and Dobson, 1995; Shaw et al. 1998) and, by characterizing the distribution of FWEC in terms of statistical theory, we aim to provide a basis for predicting the consequences of selective treatment strategies on pasture contamination rates.

MATERIALS AND METHODS

Equines and establishments

From February 2010 to August 2011 inclusive, fecal samples were examined from 1237 TBs (range; 9-258, equines per stud) resident at 22 UK studs with available grazing area of 40-1200 acres (median 172.5 acres). Studs were selected on a first come/first served basis from questionnaire respondents (see Relf et al. 2012 for further details), but had to contain a minimum of 20 permanently resident equines. Of the 22 studs, 1 was in Scotland, while all others were located in England; 2 in the north, 4 in the Midlands and 15 in the south (8 within the county of Suffolk). Sixteen animals were excluded from data analysis as their age was not provided at the time of sampling. The remaining equines (N=1221) were categorised by age (<1, 1, 2–4, 5–14 and >14 years). Here, the term 'yearling' refers to equines born during the year prior to sample analysis (1 year old), 'foal' refers to

equines born in the same year as sampling (<1 yearold, including weanlings). Personnel at the studs were asked to provide the following information: animal age, gender, anthelmintic type administered prior to sampling and date of treatment. Fecal samples were obtained a minimum of 6 weeks post-IVM, pyrantel (PYR) or fenbendazole (FBZ) treatment, or 12 weeks post-moxidectin (MOX) treatment. Details of management practices were also available from a survey conducted in 2009-2010 (Relf et al. 2012). Stud managers or owners were asked to complete a questionnaire composed of 47 questions divided into 5 sections (general information, grazing practice, worm control practices, worm control in foals and knowledge/use of FWEC analysis). Here, results received from the 22 studs that participated in the FWEC analysis were considered in relation to FWECs to determine whether practices influenced strongyle prevalence and level of egg shedding.

Fecal worm egg counts

Participants were asked to collect freshly voided fecal samples from identified individuals and place each sample in a sealable plastic bag, expelling all excess air to minimize larval development during transit. All samples were analysed within 4 days of collection and were stored at 4 °C prior to analysis. FWECs were determined using a modification of a salt-flotation method (Jackson, 1974), with a detection limit of 1 epg. Briefly, 10 g of fecal material were thoroughly mixed in 100 ml of tap water. A 10 ml aliquot of the mixture was dispensed over a 1 mm sieve, washed through using 5 ml of tap water and centrifuged at 203 g for 2 min. After removal of the supernatant, the fecal pellet was re-suspended in saturated sodium chloride solution (NaCl), specific gravity 1.204 (Ministry of Agriculture, Fisheries and Food, 1986), and centrifuged at 203 g for a further 2 min. Eggs within the positive meniscus of the NaCl solution were transferred to a cuvette and examined under a compound microscope at 40X magnification. Initially, individual samples were analysed in triplicate (N=87 received from 3 studs); however, due to time constraints in handling the large volume of fecal samples, subsequent samples were analysed in duplicate and an average epg value determined. Eggs were differentiated as 'strongyles', P. equorum, tapeworm spp. and Strongyloides westeri following published guidelines (Thienpont et al. 1986).

Data analysis

Descriptive statistics were performed using Microsoft Excel (2007). Percentages were rounded to the nearest integer. Associations between management practices and prevalence and level of egg shedding, as determined by FWEC (≥ 200 epg), of strongyle and *P. equorum* infections were tested using

Table 1.	Variables	s included	in the initial	l logistic	regression	model:	MLs,	macrocyclic	lactones;
FBZ, fea	nbendazolo	e; PYR, p	yrantel						

Variable	Responses
Year	2010, 2011
Gender	Male, female, unknown
Age category	<1, 1, 2-4, 5-14, >14 years
Last anthelmintic class administered	MLs, FBZ, PYR, none
Season sample collected	Spring, Summer, Autumn, Winter
Number of equines at stud*	1 0, , , ,
Number of stallions at stud*	
Average number of visiting equines per annum*	
Permanent and visiting equines co-grazed	Yes, No, N/A
Equines de-wormed prior to integration	Yes, No, N/A
Visiting equines quarantined prior to co-grazing	Yes, No, N/A
Rotate grazing between groups of equines	Yes, No and subsequently \leq Monthly, \geq Annually, Never
Pastures rested from grazing	Yes, No
Pastures grazed by cattle and/or sheep	Yes, No
Harrow/crop pastures	\leq Monthly, \geq Annually
Spring (March-May) access to grazing	Yes, No and subsequently 0, 1–12, 13–24 h
Autumn (Sept.–Nov.) access to grazing	Yes, No and subsequently 1–12, 13–24 h
Winter (DecFeb.) access to grazing	Yes, No and subsequently 0, 1–12, 13–24 h
Remove feces from pasture	Yes, No
Method of fecal removal	Manual, Mechanical
Evidence of worm-related illness	Yes, No
Veterinary advice sought	Yes, No
Decision to worm	Set regime, At sign of disease, Following veterinary advice
Move equines to 'clean' pasture post-treatment	Yes, No
No of treatments administered per annum*	
MLs administered in 12 months prior	Yes, No
FBZ administered in 12 months prior	Yes, No
PYR administered in 12 months prior	Yes, No
Method of determining dosage required	Weigh each horse, Estimate weigh, One tube/packet

* Indicates a scale variable.

logistic regression (Minitab Version 15, Minitab Inc., State College, PA, USA). Regression models were initially populated with all potential explanatory factors and the least significant removed in turn until the most parsimonious model, with only significant factors, remained. The factors included in initial models are shown in Table 1. *P* values ≤ 0.05 indicate factors with a significant influence on strongylid or P. equorum egg shedding in the final model. For strongyles, in which variation in individual FWEC is most likely to serve as a criterion for treatment, FWEC distribution between individual horses was investigated using maximum likelihood. Counts were assumed to follow a negative binomial distribution (Shaw et al. 1998), and the likelihood, Pr, of observing a given FWEC, s, assuming a specified distribution mean, m, and aggregation parameter, k, was estimated using equation 1:

$$Pr\{o_i = s\} = \frac{\Gamma(k+s)}{\Gamma(k)s!} \left(1 + \frac{m}{k}\right)^k \left(\frac{m}{k+m}\right)^s \tag{1}$$

where Γ is the gamma distribution (Hilborn and Mangel, 1997).

Raw FWECs were divided by 25 and rounded to the nearest integer to avoid calculation overflow arising from factorials of high egg counts. The overall likelihood of the observed dataset given the assumed underlying distribution was expressed as the negative log of the likelihood, -LL:

$$-LL = \prod_{i=1}^{n} Pr(-\ln\{o_i = s\})$$
(2)

This was minimized by iteration using the Solver software add-in to Excel (Microsoft Corp, USA), to find the values of k and m that best explained the observed FWECs. The starting value of k before iterative fitting was the corrected moment estimate:

$$k_{CME} = \frac{m^2 - \frac{v}{n}}{v - m} \tag{3}$$

where m is the mean, v is the variance, and n is the number of counts.

To compare these parameters between equine age categories, likelihood ratio tests were used (Torgerson *et al.* 2005). Thus, a model using the arithmetic mean of each age class (<1 year, 1–2, 2–4, 5–14 and >14 years) and a separate fitted k for each age class was compared with the starting (null) model, in which a single k value was fitted, and the overall arithmetic mean count used. The difference in the negative log likelihoods of each model was multiplied by 2 and compared with the chi-square distribution, with degrees of freedom equal to the difference in the number of parameters between

V.E. Relf and others

	Age category (years)						
	<1	1	2–4	5-14	>14	All ages	
N	203	295	104	489	130	1221	
% Positive for strongyle	54	83	61	45	33	56	
% with $\geq 200 \text{ epg}$	6	27	19	6	4	11	
Max epg	1111.5	2430	1840.5	2565	1039.5	2565	
Mean epg	35	216	157	56	28	97	
Median epg	0.5	49	2	0	0	0.5	
N	203	268	100	420	118	1109	
% Positive for Parascaris equorum	38	4	3	1	0	9	
% with $\geq 200 \text{ epg}$	18	<1	0	0	0	3	
Max epg	3222	318	12.5	97.5	0	3222	
Mean epg	209	2	< 0.5	< 0.5	0	39	
Median epg	0	0	0	0	0	0	
N	194	153	65	199	61	672	
% Positive for tapeworm spp.	3	3	9	6	3	4	
% with $\geq 200 \text{ epg}$	0	0	0	0	0	0	
Max epg	1	6	7.5	5.5	1.5	7.5	
Mean epg	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	< 0.5	
Median epg	0	0	0	0	0	0	
N	194	153	65	199	61	672	
% Positive for Strongyloides westeri	16	13	0	1	2	8	
% with $\geq 200 \text{ epg}$	3	0	0	0	0	1	
Max epg	1552.5	6.5	0	1	1	1552.5	
Mean epg	23	< 0.5	0	< 0.5	< 0.5	7	
Median epg	0	0	0	0	0	0	

Table 2. The proportion (%) of horses positive for strongyle, *Parascaris equorum*, tapeworm spp. and *Strongyloides westeri* eggs (expressed as eggs per gramme of feces, epg), the proportion (%) observed to have counts ≥ 200 epg and, the maximum, mean and median epg values observed within each age category

models (Hilborn and Mangel, 1997). A value of P < 0.001 was taken to indicate a significantly better fit by the more complex model. Model complexity was then reduced in a step-wise fashion to identify the most parsimonious model. The fitted values of k that were closest to each other were replaced by a common value, and -LL minimized to achieve the best fit. The procedure was repeated for k and then for m until the minimum number of class-specific values was reached, which significantly improved the fit to the data in comparison with the null model. To detect gender differences in m and k, the final age-class model was further refined to include gender-specific m and k nested within each age-class. Following comparison of the fitted gender-specific model with the null age-class model using the likelihood ratio test, gender-specific parameters were fitted in turn to each age class, first for k across all classes and then for m, while retaining significant differences in k values. For this step-wise procedure, the critical P value was adjusted for multiple comparisons by dividing alpha = 0.05 by the number of comparisons made (Sokal and Rohlf, 1995). The final model therefore included age- and gender-specific values for k and m where these significantly improved the fit to the data, and common values where they did not. To avoid inaccuracies introduced by the rounding procedure, the final description of within-class parameters comprised arithmetic mean and a direct

maximum likelihood estimate of k using the method of Wessa (2008).

To evaluate the relationship between prevalence and mean FWEC, prevalence in each host class was predicted from the class mean using the term for the zero count of the negative binomial distribution (Hilborn and Mangel, 1997):

Predicted Prevalence =
$$\left(\frac{k}{k+m}\right)^k$$
 (4)

Prevalence predicted in this way was compared with that observed using Pearson correlation.

RESULTS

Fecal sample analysis: the distribution of strongyle and P. equorum *FWECs in populations*

Of the 1221 fecal samples analysed, 54% were from mares. The remainder were derived from yearlings (24%), foals (17%), stallions (1%), teasers (1%) and geldings (3%). Of the foals, 17% were 0–4 months old, 63% were 5–8 months old and the remainder were >8 months old at sampling. The mean values of strongyle, *P. equorum*, tapeworm spp. and *S. westeri* eggs were 97, 39, <0.5 and 7 epg (medians of 0.5, 0, 0 and 0 epg, respectively), with individual strongyle and *P. equorum* FWECs ranging from 0 to 2565 epg and 0 to 3222 epg, respectively (Table 2). Overall,



Fig. 1. Mean strongyle fecal worm egg count (eggs per gramme, epg) (A) and distribution of egg counts between individuals (B), with age. k= aggregation parameter of the negative binomial distribution, whereby high k indicates a less-aggregated distribution. MLE, maximum likelihood estimate using method of Wessa (2008); CME, corrected moment estimate (see Equation 3). Egg count and k are plotted separately for male and female 2–4 year olds, since these parameters were found to differ significantly with gender in this, and only this, age category. Mean epg in columns shaded differently are significantly different (see text).

11% of all TBs excreted 80% of the total strongyle eggs enumerated (FWECs ranged from 234 to 2565 epg). Of these animals, 7% were foals, 54% yearlings, 15% 2–4 year olds, 21% 5–14 year olds and 3% >14 year olds. In the case of *P. equorum*, 80% of the eggs enumerated were present in 2% of samples (FWECs ranged from 787.5 to 3222 epg), all of which were from 5–8 month old foals. All relevant raw data pertaining to the FWECs are attached as Supplementary material (Appendix I, online version only).

The distribution of strongyle eggs between individuals was highly over-dispersed, with an overall arithmetic mean of 95 epg and an inverse aggregation parameter, k, of 0·11. Strongyle mean FWEC and degree of aggregation between individuals varied significantly between age and gender categories (Fig. 1). The most parsimonious age-specific model retained 2 fitted values for m and 4 for k. This produced a significantly better fit to the data than the null model ($\chi^2 = 209.42$, 4 D.F., P < 0.001). Mean strongyle FWEC was significantly higher in horses aged 1-4 years than in other age classes (Fig. 1A), while there was no difference between classes above or below this range, or between horses aged 1 or 2-4 years, when gender was not taken into account. Foals (<1 year) and horses aged 5–14 years shared the same fitted k, while degree of aggregation in all other age classes differed (Fig. 1B). Fitting gender-specific k within these age classes further improved the model fit ($\chi^2 = 14.949$, 7 D.F., P = 0.037). Step-wise fitting of gender-specific k identified a significant departure from the age-specific null model only within the 2-4 years age class ($\chi^2 = 6.871$, 1 D.F., P=0.009). Fitting a separate k for every age and gender class did not improve on this model $(\chi^2 = 8.078, 6 \text{ D.F.}, P = 0.23)$. Mean FWEC also differed between genders within this age class, but not in others ($\chi^2 = 6.110, 10$ D.F., P = 0.013). In horses aged 2–4 years, *m* and *k* were higher in males (N=14,all geldings) than in females (N=89). Differences in fitted parameters between age and gender classes are summarized in Fig. 1 and all details of the model fit are presented as Supplementary material (Appendix II, online version only).

Aggregation parameter k tended to track the mean (Fig. 1), indicating a tendency for FWECs to be more evenly spread between individuals at high means, and more clumped between individuals at low means. However, k was highest in yearlings even though mean FWEC was higher in 2-4 year old males (Fig. 1). The corrected moment estimate of k provided a reasonably good approximation to the maximum likelihood estimation in most cases, although it underestimated k in yearlings (Fig. 1B). As might be expected, groups with a higher mean FWEC tended to have a higher prevalence of strongylid egg shedding (Fig. 2). Prevalence was quite predictable from the negative binomial distribution (Fig. 3). Prediction was improved by using a separate k parameter for each age-gender class, estimated from the regression of k on mean FWEC (Fig. 3A). This resulted in a similar correlation coefficient between predicted and observed prevalence to when a common k value was used (Fig. 3B), and spread the predicted prevalence over a more realistic range (Fig. 3C). The most precise prediction of prevalence was obtained when differences in aggregation between groups were taken into account through separate age-gender specific maximum likelihood estimates (Fig. 3D). Differences in aggregation modified the effects of mean FWEC on the proportion of horses shedding high numbers of eggs (Fig. 4).

Prevalence of strongylid and P. equorum infections and factors associated with prevalence

Although 44% of samples analysed were negative for strongyle eggs, this parasite group was by far the most



Fig. 2. Relationship between mean strongyle fecal worm egg count (eggs per gramme, epg) and prevalence in groups of horses in consistent age-gender groupings on different studs (A), and across the whole dataset (B). In (A), groupings were foals (<1 year old), yearlings (1 year old), geldings, mares and stallions/teasers on 22 UK studs (Pearson r=0.81, N=68, P<0.001). In (B), groups were <1 year old, 1 year old, 2–4 year old males, 2–4 year old females, 5–14 year olds and horses >14 years old (Pearson r=0.95, N=6, P=0.004). The horizontal and vertical axes are interchangeable since the mean can be used to predict prevalence and *vice versa*.

prevalent detected and strongylid eggs were identified in samples submitted from all studs. P. equorum, tapeworm spp. or S. westeri eggs were detected in samples received from 11/21, 9/20 and 8/20 of studs, respectively. Samples with P. equorum or S. westeri \geq 200 epg accounted for only 3% and 1% of the cohort, respectively (Table 2). Age had a significant effect on strongyle prevalence, with the highest level recorded in 1 year olds (83%, Table 2): horses in this age category were, on average, 3 times more likely to be positive for strongyle eggs than those <1 year old (Table 3). Horses in the \geq 5 year old category had the lowest prevalence of infection (33-45%, Table 2) and were 0.40-0.69 times more likely to have lower strongyle FWECs than those that were in the ≤ 1 year old category (Table 3). The prevalence of positive strongyle FWECs was also associated with management practices and year (Table 3). For example, horses positive for strongyle eggs were, on average, most likely to have been sampled during 2011, administered PYR at last treatment and resided at studs that did not perform rotational group

grazing. Rotational group grazing was the most significant factor: studs implementing rotation on a monthly (or more frequent) basis were nearly 18 times less likely to have animals positive for strongyle eggs at screening than those studs which did not implement rotation (Table 3).

Parascaris equorum eggs were predominantly observed in samples from horses <1 year old, whilst all samples from horses >14 years old were negative for these eggs (Table 2). Of the *P. equorum* FWECpositive foals (N=78), 82% were 5–8 months old, while the remainder were 0–4 or >8 months old (1 and 17%, respectively). In addition to age, equines positive for *P. equorum* by FWEC analysis were most likely have been sampled during 2011, sampled in summer or autumn at studs where pastures were 'rested' from grazing, and where feces were removed from pasture (Table 4).

Level of strongylid and P. equorum nematode egg shedding and factors associated with levels of shedding

Strongyle FWECs ≥ 200 epg were measured in samples received from 16/22 studs; however, these accounted for only 11% of the total samples screened (Table 2). Equines with ≥ 200 strongyle epg were most likely to have been administered FBZ at their last treatment (OR=30.61), were from studs that grazed >13 h per day from December to February (compared to those grazing for shorter periods of time) and from studs that moved horses to 'clean' pasture after treatment. Samples analysed during December to February had a higher OR, being >200 strongyle epg compared to those analysed in spring, (March to May) (Table 3). The highest OR (1.01, Table 4) associated with a *P. equorum* FWEC >200 epg was the number of horses present at the stud.

DISCUSSION

There is a critical need to deploy parasite control strategies that are less dependent on anthelmintic use and more reliant on management-based control on equine breeding enterprises. A prerequisite to developing such strategies is an improved understanding of the distributions of the relevant helminth species in field settings. The information generated here informs on the prevalence of the target pathogens and their distributions in horses of different age categories on UK TB studs. As observed many times previously (Mfitilodze and Hutchinson, 1989, 1990; Epe et al. 1993; Bucknell et al. 1995; Beelitz et al. 1996; Höglund et al. 1997; Epe et al. 2004; Kornas et al. 2010; Hinney et al. 2011), strongyles were the most prevalent helminth type detected. The prevalence of P. equorum was similar to previous studies (Epe et al. 1993, 2004; Kornas et al. 2010; Hinney et al. 2011). Prevalence and egg-shedding levels of this nematode



Fig. 3. Prevalence predicted from the group mean strongyle fecal worm egg count, using the negative binomial term for the zero class (see Equation 4). Over-dispersion parameter k was estimated from regression against the mean (A; Pearson r=0.636, P=0.175, N=6 in all cases), or by maximum likelihood, for each gender class (<1 year old, 1 year old, 2–4 year old males, 2–4 year old females, 5–14 year olds and horses >14 years old). The value of k used to predict prevalence was taken to be the common maximum likelihood estimate across the entire dataset (B; r=0.91, P=0.011), the age-gender class specific estimate from regression against the mean (C; r=0.90, P=0.016), or separate maximum-likelihood fits within each age-gender class (D; r=0.96, P=0.002). Trend lines represent the fitted linear regression (A) and the 1:1 relationship between predicted and observed prevalence (B-D).

species were greatest in 5-8 month old foals. This, coupled with reports of ML resistance in P. equorum (Reinemeyer, 2009), suggests that effective control of this potentially pathogenic parasite in young horses is essential. The finding of S. westeri eggs in foals was expected, given that they acquire infections via transmammary transmission as well as from the environment (Lyons et al. 1973). Environmental transmission during housing and creep feeding may account for the higher prevalence observed here (16%), than reported previously, where there was no indication of foals being creep fed (Lyons et al. 1993; Lyons and Tolliver, 2004). Lyons and Tolliver (2004) noted a decline in S. westeri prevalence, presumably due to the impact of ML treatments on transmammary transmission (Ludwig et al. 1983). The low prevalence of tapeworm here is similar to previous coprology-based studies (Epe et al. 1993; 2004; Lyons and Tolliver, 2004). However, our data must be interpreted with caution since the FWEC methodology used has not been thoroughly validated for this species and egg-counting methods modified for the detection of tapeworm eggs are typically performed on a larger volume of feces (Proudman and Edwards, 1992).

The relatively low proportion of strongyle FWECpositive animals, compared to studies on non-TBs

(Beelitz et al. 1996; Osterman Lind et al. 1999; Hinney et al. 2011; Kyvsgaard et al. 2011), may reflect the extremely high frequency of ML administration on these studs (Relf et al. 2012). A cut-off for anthelmintic administration in horses of 100-300 helminth epg has been suggested, with 200 epg often chosen (Uhlinger, 1993). In the current dataset, a cut-off value of 200 strongyle epg would indicate that only 11% of the population sampled would have required treatment at the time of sampling. It is worthy of note here that, in all cases, the stud personnel involved were keen to administer anthelmintic treatments to all horses at the time that they submitted feces for FWEC analysis: indicating the substantial change in behaviour patterns that will be required for more sustainable helminth control on UK TB studs. In relation to the generally accepted adage that indicates that 20% of horses excrete 80% of the egg contamination (Stratford et al. 2011), only 11% of horses sampled here were responsible for excreting 80% of strongyle eggs based on the FWECs performed. Our findings also support the application of specific treatments for P. equorum in foals, since those responsible for excreting 80% of eggs were all <1 year old. Although age was identified as a contributing factor in determining strongyle and ascarid prevalence and level of egg shedding,



Fig. 4. Change in distribution of strongyle fecal worm egg count (eggs per gramme, epg) with age and gender. k is the aggregation (over-dispersion) parameter of the negative binomial distribution. P is the proportion of fecal worm egg counts above the arbitrary treatment threshold of 200 epg, indicated by the dotted vertical lines. Arrows indicate the mean strongyle fecal worm egg count, and dots the frequencies predicted by the negative binomial distribution for the first 4 count intervals.

FWECs can differ considerably among adult horses (Boersema et al. 1996; Eysker et al. 2008), indicating that some are susceptible to high levels of infection, and hence disease, throughout their lives (Jasko and Roth, 1984). In agreement with this, our results clearly demonstrate that strongyle FWECs were highly over-dispersed between horses within all age and gender categories, as observed previously (Nielsen et al. 2006; Vidyashankar et al. 2007; Hinney et al. 2011). Mean FWEC and degree of aggregation differed between genders in the 2-4 year old category with males displaying, on average, higher and less aggregated FWECs than females, possibly due to an effect of sex steroid expression on immunocompetence (Hamilton and Zuk, 1982; Klein, 2004; Bouman et al. 2005; Lamason et al. 2006; Monteiro et al. 2007). These findings must be interpreted with caution, however, as the low sample size of 2–4 year old males (N=14) meant that a small number of high individual counts were responsible for the high mean FWEC. An additional confounding factor may be differences between pastures on which the horses grazed, as male and female horses >1 year old were generally grazed separately.

Mean FWEC and k were highest in horses aged 1–4 years and lower in foals and in older horses, in

agreement with previous studies (Larsen et al. 2002; Fritzen et al. 2010; Kornas et al. 2010). Despite the lower degree of overdispersion within the 1-4 year class, the judicious application of a targeted treatment approach still has potential to protect younger horses from high burdens while leaving refugia for anthelmintic-susceptible genotypes (van Wyk, 2001): even at higher FWEC means, aggregation was marked, and k low relative to that for FWECs in grazing ruminants (Morgan et al. 2005). This provides considerable scope for the successful application of a targeted treatment plan based on FWECs in horses of all age categories. By characterizing the statistical distribution of FWECs in different age classes, we provide a basis for exploring the consequences and efficient application of targeted treatment strategies through simulation (Morgan et al. 2005; Eysker et al. 2008). Although in practice k is unlikely to be known for a given group of horses, our data indicate its likely range. Furthermore, the apparent correlation between mean FWEC and k provides the opportunity to apply an indicative k to horse populations based on observed mean FWEC. The value of k will affect the proportion of horses with high FWECs at a given group mean, the accuracy of observed FWEC in a subsample of horses as an estimate of underlying group mean epg and the proportion of the parasite population removed as a result of threshold-based anthelmintic treatment (Morgan and Wall, 2009). The data presented here will therefore assist development of strategic recommendations for implementation of targeted treatment approaches in TBs, and operational decision support tools for its practical application. Although other statistical methods could be used to investigate variation in FWEC between horses, and characterize distribution patterns, the explicit use of the negative binomial distribution is in keeping with valuable previous work in other taxa (Shaw and Dobson, 1995; Shaw et al. 1998) and will permit our conclusions to be generalized in support of a range of objectives in the wider equine population. We show that relationships between prevalence and mean FWEC are predictable, and can be more precisely characterized when differences in k between age categories are taken into account. The use of a highly sensitive FWEC method, with a detection limit as low as 1 epg, provided an opportunity to characterize parasite distribution and prevalence-mean relationships in some detail. Inference from data generated using other, less-sensitive FWEC methods would have to take into account measurement error, such that observed FWEC is a function of Poisson error around the true value (Torgerson et al. 2012).

In terms of management practices that affected prevalence or levels of nematode egg shedding, studs implementing rotational group grazing (especially between age categories) were more likely to have a lower prevalence and lower strongyle FWECs, Table 3. Significant predictors of the prevalence and level of strongyle egg shedding using logistic regression

(The dependent variable in each case was the presence of eggs in feces. Model fit was assessed using the Hosmer and Lemeshow test (X^2). ML, macrocyclic lactone; PYR, pyrantel;
FBZ, fenbendazole)

Infection category	χ^2 , d.f.	P, fit	Most significant variables	Variable category	Logit coefficient (s.e.)	OR	Р
Prevalence	7.570,8	0.477	Year		1.192 (0.251)	3.29	0.000
	,		Age category	<1 year		1	
			0 0 2	1 year	1.744 (0.249)	3.29	0.000
				5–14 years	-0.370(0.200)	0.69	0.064
				>14 years	-0.914(0.270)	0.40	0.001
			Last anthelmintic class administered	ML		1	
				PYR	1.751 (0.315)	5.76	0.000
			Number of stallions at stud		-0.268(0.089)	0.76	0.003
			Rotate grazing between groups of equines	≤ Monthly		1	
				Never	2.881(0.663)	17.84	0.000
			Harrow/crop pastures	≤ Monthly		1	
				≥ Annually	1.099(0.246)	3.00	0.000
			Spring (March–May) access to grazing	0 h		1	
			~F8 (1–12 h	-3.538(1.043)	0.03	0.001
				13–24 h	-3.838(0.982)	0.02	0.000
			Autumn (Sept – Nov.) access to grazing	1–12 h		1	
				13–24 h	0.938(0.263)	2.55	0.000
			Evidence of worm-related illness	No	0,000 (0,200)	1	0 000
				Yes	0.835(0.385)	2.30	0.030
			Decision to worm	Set regime	()	1	
				At sign of disease	-2.427(0.704)	0.09	0.001
				Following veterinary advice	-0.970(0.374)	0.38	0.009
≥ 200 epg	7.537.8	0.480	Age category	<1 vear		1	
2 10 10	· · · · , ·			5–14 years	-1.063(0.403)	0.35	0.008
				>14 years	-1.164(0.629)	0.31	0.064
			Last anthelmintic class administered	ML		1	
				FBZ	3.421 (0.751)	30.61	0.000
			Season sample collected	Spring (March–May)		1	
			1.	Winter (Dec.–Feb.)	1.238(0.298)	3.45	0.000
			Number of stallions at stud		-0.527(0.113)	0.59	0.000
			Equines de-wormed prior to integration	No		1	
				Yes	-2.014(0.446)	0.13	0.000
				N/A	-1.644 (0.484)	0.19	0.001
			Rotate grazing between groups of equines	No		1	0 001
				Yes	-2.336(0.730)	0.10	0.001
			Winter (DecFeb.) access to grazing	0 hours		1	
				13-24 hours	2.029(0.500)	7.61	0.000
			Move equines to 'clean' pasture post-treatment	No		1	0
				Yes	1.629(0.382)	5.10	0.000
						2.13	0 0 0 0

649

Table 4. Significant predictors of the prevalence and level of Parascaris equorum egg shedding using logistic regression

(The dependent variable in each case was the presence of eggs in feces. Model fit was assessed using the Hosmer and Lemeshow test (X^2) . ML, macrocyclic lactone; FBZ, fenbendazole)

Infection category	χ ² , D.F.	P, fit	Most significant variables	Variable category	Logit coefficient (s.e.)	OR	Р
Prevalence	8.158,8	0.418	Year		1.729 (0.678)	5.64	0.011
	,		Age category	<1 year		1	
			0 0 2	1 year	-1.924(0.488)	0.15	0.000
				2–4 years	-2.222(0.757)	0.11	0.003
				5–14 years	-2.928(0.581)	0.05	0.000
			Last anthelmintic class	ML		1	
			administered	FBZ	-1.386(0.812)	0.25	0.088
				None	-2.361(0.587)	0.09	0.000
			Season sample collected	Spring (March–May)		1	
				Summer (June–Aug.)	1.626 (0.519)	5.08	0.002
				Autumn (Sept –Nov.)	1.991 (0.700)	7.32	0.004
			Number of equines at stud	(~···)	0.007(0.004)	1.01	0.052
			Pastures rested from grazing	No	, , ,	1	
			0 0	Yes	1.355(0.660)	3.88	0.040
			Spring (March–May) access	0 h	· · · · ·	1	
			to grazing	1–12 h	-4.016(1.642)	0.02	0.014
			0 0	13–24 h	-4.319(1.633)	0.01	0.008
			Remove feces from pasture	No		1	
				Yes	1.691 (0.758)	5.42	0.026
			Move equines to 'clean'	No		1	
			pasture post-treatment	Yes	-2.832(1.188)	0.06	0.017
≥ 200 epg	0.300, 2	0.861	Age category	<1 Year		1	
				1 Year	-2·329 (1·119)	0.10	0.037
			Number of equines at stud		0.014 (0.004)	1.01	0.002
			Number of stallions at stud		-0.242(0.138)	0.78	0.079

confirming the value of rotation to reduce transmission via pasture, as previously described in sheep (Leathwick et al. 2008). Not surprisingly, time spent grazing also influenced the parasitological parameters, presumably due to longer grazing periods increasing exposure to third stage larvae. Regular removal of roughs, areas where larvae can remain protected from desiccation (Herd and Willardson, 1985), by harrowing or cropping, was associated with reduced strongyle prevalence, similar to observations on German stud farms (Fritzen et al. 2010). FWECs of ≥ 200 epg were more likely to be detected on studs where horses were moved to 'clean' pasture after anthelmintic treatment suggesting that the perception of 'clean' may be inappropriate, as highlighted previously (Larsen et al. 2002; Hinney et al. 2011). In addition, anthelmintic inefficacy will also affect this, with animals still excreting eggs after treatment (and not detected as doing so) transferring infection to clean pasture. Animal movement to clean pasture immediately after anthelmintic treatment is no longer advised because of potential detrimental effects on levels of refugia (van Wyk, 2001): further knowledge on the impact of refugia needs to be translated to UK veterinary surgeons and stud managers. Where PYR and FBZ were administered as the last

treatment prior to FWEC analysis, strongyle prevalence and FWECs were likely to be higher, respectively. The less persistent activity of PYR and FBZ (Eysker et al. 1992; Uhlinger, 1992) along with the higher possibility of resistance in cyathostomins to these active ingredients (Kaplan, 2002, 2004; Traversa et al. 2007), are likely reasons for these observations. The increased likelihood of higher levels of strongyle egg-shedding in animals sampled during winter (December to February), in comparison to those sampled in spring (March to May) may reflect changes in grazing management patterns, keeping in mind that those horses that grazed for >13 h per day at this time of year were more likely to have higher levels of strongyle egg shedding. This, linked to the potential effects of climate change, i.e. milder autumns/winters in the UK, may lead to an increase in development and survival of nematode larvae in winter (van Dijk et al. 2010) resulting in alterations in egg-shedding patterns and hence the epidemiology of these infections. The fact that fecal samples obtained in summer, in this instance, were not associated with higher levels of shedding may be linked with a higher anthelmintic treatment frequency, and hence strongyle egg suppressive effect, at this time of year.

Few studies have attempted to investigate the prevalence, distribution and level of strongyle and P. equorum egg shedding in relation to host and management factors (Kornas et al. 2010). The findings here indicate that a number of specific host and management factors play an important role in determining the prevalence, distribution and intensity of nematode infections in breeding TBs, all of which need to be taken into account in designing sustainable control programmes for the future.

ACKNOWLEDGEMENTS

The authors would like to thank the studs and veterinary surgeons that participated in this study.

FINANCIAL SUPPORT

This work was supported by funding from The Horse Race Betting Levy Board, HBLB (grant number HBLB vet/prj/ 755).

REFERENCES

Becher, A. M., Mahling, M., Nielsen, M. K. and Pfister, K. (2010). Selective anthelmintic therapy of horses in the Federal states of Bavaria (Germany) and Salzburg (Austria): An investigation into strongyle egg shedding consistency. *Veterinary Parasitology* **171**, 116–122.

Beelitz, P., Göbel, E. and Gothe, R. (1996). Spectrum of species and incidence of endoparasites in foals and their mother mares from breeding farms with and without anthelmintic prophylaxis in upper Bavaria. *Tierärztliche Praxis* **24**, 48–54.

Boersema, J. H., Eysker, M., Maas, J. and van der Aar, W. M. (1996). Comparison of the reappearance of strongyle eggs on foals, yearlings and adult horses after treatment with ivermectin or pyrantel. *Veterinary Ouarterly* **18**, 7–9.

Bouman, A., Heineman, M. J. and Faas, M. M. (2005). Sex hormones and the immune response in humans. *Human Reproduction Update* 11, 411–423. doi: 10.1093/humupd/dmi008.

Bucknell, D. G., Gasser, R. B. and Beveridge, I. (1995). The prevalence and epidemiology of gastrointestinal parasites of horses in Victoria, Australia. *International Journal for Parasitology* 25, 711–724.

Döpfer, D., Kerssens, C. M., Meijer, Y. G. M., Boersema, J. H. and Eysker, M. (2004). Shedding consistency of strongyle-type eggs in Dutch boarding horses. *Veterinary Parasitology* **124**, 249–258.

Epe, C., Coati, N. and Schnieder, T. (2004). Results of parasitological examinations of faecal samples from horses, ruminants, pigs, dogs, cats, hedgehogs and rabbits between 1998 and 2002. *Deutsche Tierärztlicher Wochenschrift* **111**, 243–247.

Epe, C., Ising-Volmer, S. and Stoye, M. (1993). Parasitological fecal studies of equids, dogs, cats and hedgehogs during the years 1984–1991. *Deutsche Tierärztlicher Wochenschrift*, **100**, 426–428.

Eysker, M., Bakker, J., van den Berg, M., van Doorn, D. C. K. and Ploeger, H. W. (2008). The use of age-clustered pooled faecal samples for monitoring worm control in horses. *Veterinary Parasitology* **151**, 249–255.

Eysker, M., Boersema, J. H. and Kooyman, F. N. J. (1992). The effect of ivermectin treatment against inhibited early third stage, late third stage and fourth stage larvae and adult stages of the cyathostomes in Shetland ponies and spontaneous expulsion of these helminths. *Veterinary Parasitology* **42**, 295–302.

Fritzen, B., Rohn, K., Schnieder, T. and Von Samson-Himmelstjerna, G. (2010). Endoparasite control management on horse farms-lessons from worm prevalence and questionnaire data. *Equine Veterinary Journal* 42, 79–83. doi: 10.2746/042516409x471485.

Hamilton, W.D. and Zuk, M. (1982). Heritable true fitness and bright birds: a role for parasites. *Science* 218, 384–387.

Herd, R.P. and Willardson, K.L. (1985). Seasonal distribution of infective strongyle larvae on horse pastures. *Equine Veterinary Journal* **17**, 235–237. doi: 10.1111/j.2042-3306.1985.tb02481.x.

Hilborn, R. and Mangel, M. (1997). The Ecological Detective: Confronting Models with Data, Princeton University Press, Princeton, NJ, USA. Hinney, B., Wirtherle, N., Kyule, M., Miethe, N., Zessin, K.-H. and Clausen, P.-H. (2011). Prevalence of helminths in horses in the state of Brandenburg, Germany. *Parasitology Research* **108**, 1083–1091. doi: 10.1007/s00436-011-2362-z.

Höglund, J., Ljungström, B.L., Nilsson, O., Lundquist, H., Osterman, E. and Uggla, A. (1997). Occurrence of *Gasterophilus intestinalis* and some parasitic nematodes of horses in Sweden. Acta *Veterinaria Scandinavica* 38, 157–165.

Jackson, F. (1974). New technique for obtaining nematode ova from sheep faeces. *Laboratory Practice* 23, 65–66.

Jasko, D. J. and Roth, L. (1984). Granulomatous colitis associated with small strongyle larvae in a horse. *Journal of the American Veterinary Medical Association* **185**, 553–554.

Kaplan, R. M. (2002). Anthelmintic resistance in nematodes of horses. *Veterinary Research* 33, 491–507.

Kaplan, R.M. (2004). Drug resistance in nematodes of veterinary importance: a status report. *Trends in Parasitology* **20**, 477–481.

Klein, S. L. (2004). Hormonal and immunological mechanisms mediating sex differences in parasite infection. *Parasite Immunology* **26**, 247–264. doi: 10.1111/j.0141-9838.2004.00710.x.

Kornas, S., Cabaret, J., Skalska, M. and Nowosad, B. (2010). Horse infection with intestinal helminths in relation to age, sex, access to grass and farm system. *Veterinary Parasitology* **174**, 285–291.

Kuzmina, T. A. and Kharchenko, V. O. (2008). Anthelmintic resistance in cyathostomins of brood horses in Ukraine and influence of anthelmintic treatments on strongylid community structure. *Veterinary Parasitology* **154**, 277–288.

Kyvsgaard, N. C., Lindbom, J., Andreasen, L. L., Luna-Olivares, L. A., Nielsen, M. K. and Monrad, J. (2011). Prevalence of strongyles and efficacy of fenbendazole and ivermectin in working horses in El Sauce, Nicaragua. *Veterinary Parasitology* **181**, 248–254.

Lamason, R., Zhao, P., Rawat, R., Davis, A., Hall, J., Chae, J., Agarwal, R., Cohen, P., Rosen, A., Hoffman, E. and Nagaraju, K. (2006). Sexual dimorphism in immune response genes as a function of puberty. *BMC Immunology* **7**, 2.

Larsen, M., Lendal, S., Chriel, M., Olsen, S. and Bjorn, H. (2002). Risk factors for high endoparasitic burden and the efficiency of a single anthelmintic treatment of Danish horses. *Acta Veterinaria Scandinavica* **43**, 99–106.

Leathwick, D. M., Miller, C. M., Atkinson, D. S., Haack, N. A., Waghorn, T. S. and Oliver, A. M. (2008). Managing anthelmintic resistance: Untreated adult ewes as a source of unselected parasites, and their role in reducing parasite populations. *New Zealand Veterinary Journal* 56, 184–195. doi: 10.1080/00480169.2008.36832.

Lind, E. O., Höglund, J., Ljungström, B. L., Nilsson, O. and Uggla, A. (1999). A field survey on the distribution of strongyle infections of horses in Sweden and factors affecting faecal egg counts. *Equine Veterinary Journal* **31**, 68–72. doi: 10.1111/j.2042-3306.1999.tb03793.x.

Love, S., Murphy, D. and Mellor, D. (1999). Pathogenicity of cyathostome infection. *Veterinary Parasitology* **85**, 113–122.

Ludwig, K.G., Craig, T.M., Bowen, J.M., Ansari, M.M. and Ley, W.B. (1983). Efficacy of ivermectin in controlling *Strongyloides westeri* infections in foals. *American Journal of Veterinary Research* 44, 314-316.

Lyons, E. T., Drudge, J. H. and Tolliver, S. C. (1973). On the life cycle of *Strongyloides westeri* in the equine. *Journal of Parasitology* **59**, 780–787.

Lyons, E. T. and Tolliver, S. C. (2004). Prevalence of parasite eggs (*Strongyloides westeri*, *Parascaris equorum* and strongyles) and oocysts (*Emeria leuckarti*) in the feces of Thoroughbred foals on 14 farms in central Kentucky in 2003. *Parasitology Research* 92, 400–404. doi: 10.1007/s00436-003-1068-2.

Lyons, E. T., Tolliver, S. C., Harold Drudge, J., Granstrom, D. E. and Collins, S. S. (1993). Natural infections of *Strongyloides westeri:* prevalence in horse foals on several farms in central Kentucky in 1992. *Veterinary Parasitology* **50**, 101–107.

Matthews, J.B. (2008). An update on cyathostomins: Anthelmintic resistance and worm control. *Equine Veterinary Education* **20**, 552–560. doi: 10.2746/095777308x363912.

Mfitilodze, M. W. and Hutchinson, G. W. (1989). Prevalence and intensity of non-strongyle intestinal parasites of horses in northern Queensland. *Australian Veterinary Journal* 66, 23–26. doi: 10.1111/j.1751-0813.1989.tb09708.x.

Mfitilodze, M.W. and Hutchinson, G.W. (1990). Prevalence and abundance of equine strongyles (Nematoda: Strongyloidea) in tropical Australia. *Journal of Parasitology* **76**, 487–494.

Ministry of Agriculture, Fisheries and Food (1986). Manual of Veterinary Parasitological Laboratory Techniques. In *Reference Book 418* pp. 36–39. HMSO, London, UK.

Monteiro, R. V., Dietz, J. M., Beck, B. B., Baker, A. J., Martins, A. and Jansen, A. M. (2007). Prevalence and intensity of intestinal helminths found in free-ranging golden lion tamarins (*Leontopithecus rosalia*, Primates, Callitrichidae) from Brazilian Atlantic forest. *Veterinary Parasitology* **145**, 77–85.

Morgan, E.R., Cavill, L., Curry, G.E., Wood, R.M. and Mitchell, E.S.E. (2005). Effects of aggregation and sample size on composite faecal egg counts in sheep. *Veterinary Parasitology* **131**, 79–87.

Morgan, E. R. and Wall, R. (2009). Climate change and parasitic disease: farmer mitigation? *Trends in Parasitology* 25, 308–313.

Nielsen, M. K., Haaning, N. and Olsen, S. N. (2006). Strongyle egg shedding consistency in horses on farms using selective therapy in Denmark. *Veterinary Parasitology* **135**, 333–335.

Nielsen, M. K., Kaplan, R. M., Thamsborg, S. M., Monrad, J. and Olsen, S. N. (2007). Climatic influences on development and survival of free-living stages of equine strongyles: Implications for worm control strategies and managing anthelmintic resistance. *The Veterinary Journal* 174, 23–32.

Osterman Lind, E., Hoglund, J., Ljungstrom, B., Nilsson, O. and Uggla, A. (1999). A field survey on the distribution of strongyle infections of horses in Sweden and factors affecting faecal egg counts. *Equine Veterinary Journal* **31**, 68–72.

Proudman, C. J. and Edwards, G. B. (1992). Validation of a centrifugation/flotation technique for the diagnosis of equine cestodiasis. *Veterinary Record* 131, 71–72. doi: 10.1136/vr.131.4.71.

Reinemeyer, C. (2009). Diagnosis and control of anthelmintic-resistant Parascaris equorum. Parasites and Vectors 2(Suppl. 2), S8.

Relf, V.E., Morgan, E.R., Hodgkinson, J.E. and Matthews, J.B. (2012). A questionnaire study on parasite control practices on UK breeding Thoroughbred studs. *Equine Veterinary Journal* **44**, 466–471. doi: 10.1111/j.2042-3306.2011.00493.x.

Shaw, D.J. and Dobson, A.P. (1995). Patterns of macroparasite abundance and aggregation in wildlife populations: a quantitative review. *Parasitology* **111**(Suppl. S1), S111-S133.

Shaw, D.J., Grenfell, B.T. and Dobson, A.P. (1998). Patterns of macroparasite aggregation in wildlife host populations. *Parasitology* **117**, 597-610.

Sokal, R. and Rohlf, F. (1995). *Biometry: The Principles and Practice of Statistics in Biological Research*. W. H. Freeman and Co., New York, USA.

Stratford, C. H., McGorum, B. C., Pickles, K. J. and Matthews, J. B. (2011). An update on cyathostomins: Anthelmintic resistance and diagnostic tools. *Equine Veterinary Journal* **43**, 133–139. doi: 10.1111/j.2042-3306.2011.00397.x.

Thienpont, D., Rochette, F. and Vanparijs, O. F. J. (1986). *Diagnosing Helminthiasis by Coprological Examination*, 2nd Edn. Janssen Research Foundation, Beerse, Belgium.

Torgerson, P. R., Paul, M. and Lewis, F. I. (2012). The contribution of simple random sampling to observed variations in faecal egg counts. *Veterinary Parasitology* **188**, 397–401.

Torgerson, P. R., Schnyder, M. and Hertzberg, H. (2005). Detection of anthelmintic resistance: a comparison of mathematical techniques. *Veterinary Parasitology* **128**, 291–298.

Traversa, D., Klei, T. R., Iorio, R., Paoletti, B., Lia, R. P., Otranto, D., Sparagano, O. A. E. and Giangaspero, A. (2007). Occurrence of anthelmintic resistant equine cyathostome populations in central and southern Italy. *Preventive Veterinary Medicine* 82, 314–320.

Uhlinger, C. (1992). Preliminary studies into factors affecting the variability of egg reappearance period and anthelmintic treatment intervals in the control of equine cyathostomes. In *The 6th International Equine Infectious Diseases Conference* pp. 157–161. R & W Publications Ltd, Newmarket, UK.

Uhlinger, C. (1993). Uses of faecal egg count data in equine practice. *Compendium on Continuing Education for the Practicing Veterinarian* **15**, 742–748.

van Dijk, J., Sargison, N.D., Kenyon, F. and Skuce, P.J. (2010). Climate change and infectious disease: helminthological challenges to farmed ruminants in temperate regions. *Animal* **4**, 377–392.

van Wyk, J. A. (2001). Refugia: overlooked as perhaps the most potent factor concerning the development of anthelmintic resistance. *Onderstepoort Journal of Veterinary Research* **68**, 55–67.

Vidyashankar, A. N., Kaplan, R. M. and Chan, S. (2007). Statistical approach to measure the efficacy of anthelmintic treatment on horse farms. *Parasitology* **134**, 2027–2039.

von Samson-Himmelstjerna, G. (2012). Anthelmintic resistance in equine parasites-detection, potential clinical relevance and implications for control. *Veterinary Parasitology* 185, 2–8.

Wessa, P. (2008). Maximum-likelihood Negative Binomial Distribution Fitting (v1.0.2) in Free Statistics Software (v1.1.23-r7). Office for Research Development and Education, URL http://www.wessa.net/ rwasp_fitdistrnegbin.wasp/