

Research Article

*Present address: Faculty of Medicine and Health Sciences, School of Veterinary Medicine and Science, University of Nottingham, UK

†Present address: Animal & Grassland Research and Innovation Centre, Teagasc, Grange, Dunsany, Co. Meath, Ireland

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Author for correspondence:

Anastasio Argüello, Email: tacho@ulpgc.es

Influence of treatment and refrigeration time on antimicrobial activity of goat and sheep colostrum

Maria D. Ruiz-Diaz^{1,*}, Anastasio Argüello^{1,†}, Daniel Padilla², Bernadette Earley³ and Noemi Castro¹

¹Animal Production and Biotechnology Group, Institute of Animal Health and Food Safety, Universidad de Las Palmas de Gran Canaria, Arucas, Las Palmas, Spain; ²Instituto Universitario de Sanidad Animal y Seguridad Alimentaria, Universidad de Las Palmas de Gran Canaria, Las Palmas, Spain and ³Animal & Grassland Research and Innovation Centre, Teagasc, Grange, Dunsany, Co. Meath, Ireland

Abstract

The aim of the studies presented in this research communication was to compare species of origin (goat and sheep) and the effect of treatment (pasteurization at 56, 63 and 72 °C, skimming and curding) and refrigeration time on colostrum antimicrobial activity (AnAc). Two experiments were performed. In experiment 1, twenty-four first milking colostrum samples were obtained (12 goats, 12 sheep) and an aliquot of each sample was subjected to 6 different treatments, control (untreated), pasteurization at 56, 63 and 72 °C, skimming and curding. Colostrum AnAc was tested directly against *E. coli* using disks in a Petri dish and Enrofloxacin (antibiotic) and saline serum as positive and negative control, respectively. Species had no effect ($P > 0.05$) on colostrum AnAc, and neither did pasteurization at different temperatures or skimming. However, curding showed the lowest colostrum AnAc ($P < 0.05$) in both species. In the second experiment, four treatments were assayed, control, pasteurization at 56 and 63 °C and skimming. An aliquot of twelve goat colostrum samples were refrigerated after treatments for 10 d at 4 °C. Colostrum AnAc was measured at 0, 2, 4, 6, 8, and 10 d. A reduction in colostrum AnAc was observed due to refrigeration time. The results suggest that if farmers use frozen colostrum for neonates, the process of curding colostrum or refrigeration at 4 °C longer than 4 d is not recommended.

The ingestion of colostrum by newborn ruminant is the only source of immunoglobulins (Ig) during the first month of life (Castro *et al.*, 2011). Colostrum is not only a source of Ig (Hernandez-Castellano *et al.*, 2014), it is also a rich source of multiple biomolecules, some of which confer the colostrum with antimicrobial activity (AnAc) (Bhanu *et al.*, 2015). On dairy farms, colostrum is usually milked and fed to neonates through bottles or buckets, so as to avoid the development of the mother-neonate bond, to prevent udder lesions, and to allow the colostrum to be treated in order to reduce presence of pathogens. Less common management practices are skimming or curding of the colostrum in order to enrich the Ig concentration (Castro *et al.*, 2007). The objective of the present study was to investigate the effect of species (goat and sheep), treatment (pasteurization at 56, 63 and 72 °C, skimming and curding) and refrigeration time on colostrum antimicrobial activity (AnAc).

Materials and methods

The first milking postpartum colostrum from 12 dairy Majorera nanny goats and 12 dairy Canarian ewes was collected in a milking parlour. Each colostrum sample was divided into aliquots (50 ml) and stored at –80 °C until subsequent analyses were performed. *Escherichia coli* were obtained from rectal faeces samples collected from Majorera goat kids. Three gram (g) of faeces were suspended in 5 ml of a saline solution. Then 400 µl of the solution were plated onto Petri dishes with blood agar medium (Oxoid, Hampshire, United Kingdom). Petri dishes were incubated at 37 °C for 24 h. After that, different colonies, which had grown up in this blood agar media, were plated into McConkey agar (37 °C, 24 h, Oxoid, Hampshire, United Kingdom). When these colonies had established, a Gram stain was performed to identify coliforms bacteria. Gram negative bacilli, which correspond with Enterobacteriaceae, were identified at 1000× magnification in an optic microscope. *Escherichia coli* was confirmed using an Analytical Profile Index (Biomerieux, USA).

In the first experiment, colostrum samples were thawed and heated at 37 °C in a water bath for 30 min. After that, each sample was divided into 6 tubes containing 5 ml each. Of these six samples, five were subsequently suspended in different treatments while one sample served as

control (no treatment). Treatments applied to samples were: pasteurization at 56 °C for 1 h, 63 °C for 30 min and 72 °C for 15 s according to Trujillo *et al.* (2007), skimming colostrum and curdling (to obtain whey) colostrum according to Castro *et al.* (2007). *Escherichia coli* suspension (McFarland No. 1 standard, approximately 3×10^8 CFU/ml) was used. Two hundred μ l of the *E. coli* suspension were plating into violet red bile agar medium Petri dishes (Oxoid, Hampshire, United Kingdom). Petri dishes were allowed to dry for 2 h. After that, 4 dried discs were located in each Petri dish: negative control (sterile saline serum 0.9%), positive control (Enrofloxacin, 250 μ g) and two colostrum samples (each sample was tested in duplicate in different dishes). Petri dishes were subsequently incubated at 37 °C for 24 h. Halos formed were measured using a digital scanner and the AnAc percentage of different samples was estimated. Positive control was defined as 100% of AnAc and negative control was defined as 0% of AnAc. Sample values were calculated as follow:

$$\text{AnAc \%} = (\text{Sample Diameter} - \text{Negative Control Diameter}) \times 100 / \text{Positive Control Diameter}$$

In the second experiment, twelve samples of goat colostrum (from the same animals involved in experiment 1) were used. Colostrum samples were thawed and heated at 37 °C in a water bath for 30 min. After that, each sample was divided into 4 tubes containing 30 ml each. Of these four tubes, three were subsequently distributed in different treatments while one tube was treated as control (no treatment). Treatments applied to samples were: pasteurization at 56 °C for 1 h, 63 °C for 30 min according to Trujillo *et al.* (2007) or skimmed. After treatments, each tube was aliquot into 6 tubes of 5 ml. All tubes were refrigerated at 4 °C, and the AnAc was assessed using the above equation on day 0, 2, 4, 6, 8 and 10 of refrigeration.

SAS 9.4 (SAS Institute Inc., Cary, NC, USA) program package was used for statistical analysis. Antimicrobial activity variable was transformed using a square root transformation due to absence of normal distribution. A factorial ANOVA was performed to evaluate the effect of species and treatments in experiment 1. For experiment 2, SAS PROC MIXED procedure for repeated measures was used to evaluate the fixed effect of colostrum treatments, refrigeration time and the interaction between both effects (treatment \times refrigeration time) on colostrum antimicrobial activity. Significant differences between means were identified using the Tukey–Kramer test. Data are presented untransformed in the tables.

Results and discussion

Species (goat vs. sheep) had no effect on colostrum AnAc ($P = 0.331$). Antimicrobial activity is the sum of the effects of different bioactive substances that have been reported with immune or antimicrobial activity such as antimicrobial peptides, oligosaccharides, glycoproteins, lactoferrin, lactoperoxidase, lactadherin and Ig (Cacho and Lawrence, 2017). Lactoferrin and Ig are amongst the most active biomolecules in the colostrum, but no differences between goats and sheep colostrum have been reported (Abd El-Gawad *et al.*, 1996; Hernandez-Castellano *et al.*, 2016). Lactoperoxidase expression has been reported to be higher in cow than in goat or sheep colostrum (Hernandez-Castellano *et al.*, 2016). To date, no major differences have been reported between goat and sheep colostrum

Table 1. Effect of pasteurization, skimming and curdling on antimicrobial activity (%) of goat and sheep colostrum

Species	Treatments				Statistical analysis					
	Control	56 °C	63 °C	72 °C	Skimmed	Curdling	SD	S	T	S \times T
Goat	14.77 ^a	12.28 ^a	14.41 ^a	9.71 ^a	16.22 ^a	6.35 ^b	0.82	0.331	0.001	0.466
Sheep	16.44 ^a	15.88 ^a	16.99 ^a	11.20 ^a	17.23 ^a	2.95 ^b	0.87			

S, species; T, treatment; SD, standard deviation.

^{ab}Means within a row with different superscript differ ($P < 0.05$).

Table 2. Effect of refrigeration time in colostrum antimicrobial activity according to colostrum treatment

Treatment	Means						SD	Statistical analysis		
	Refrigeration days							T	D	T × D
	0	2	4	6	8	10				
Control	12.83 ^a	11.60 ^a	12.36 ^{ab}	5.92 ^b	6.83 ^b	3.12 ^c	5.09	0.599	0.001	0.001
56 °C	18.62 ^a	12.01 ^b	10.28 ^b	5.54 ^c	3.70 ^c	2.72 ^c	6.30			
63 °C	17.86 ^a	15.97 ^a	8.92 ^b	6.51 ^{bc}	3.78 ^c	1.76 ^c	6.66			
Skimmed	14.50 ^a	13.07 ^a	11.08 ^a	5.74 ^b	3.07 ^b	2.87 ^b	5.78			

T, treatment; D, day; sd, standard deviation.

^{abc}Means within a row with different superscript differ ($P < 0.05$).

for the concentration of biomolecules with immune function, which is in agreement with our results (Table 1).

Different pasteurization protocols tested in the present study did not affect the colostrum AnAc in either experiment (Tables 1 and 2). Colostrum pasteurization protocols applied in the present study are usually used for milk quality purposes (except 72 °C). Protocols using 56 and 63 °C have been described to avoid denaturation of Ig in colostrum (Trujillo *et al.*, 2007). Furthermore, Daniels *et al.* (2017) have reported 70% stability for lactoferrin in human milk using traditional sterilization methods. The low pasteurization method temperatures/times combination used in the present study could be the reason for not detecting statistical differences for AnAc between methods. Colostrum skimming or curding have been proposed by Castro *et al.* (2007) in order to increase the IgG concentration in colostrum. Our findings showed that skimmed colostrum AnAc was similar to pasteurization methods in both goat and sheep colostrum, however, the curding treatment showed lower colostrum AnAc. The curding method uses rennet in order to obtain the colostrum whey, Ig remains in the whey and the activity of this Ig was not affected according to Castro *et al.* (2007). Milk lactoferrin concentration is affected by curding. Ripolles *et al.* (2015) reported a reduction in antibacterial activity of bovine milk lactoferrin and its hydrolysates when different types of rennet were used, but lactoferrin is very resistant to heat treatments (Franco *et al.*, 2018). Lactoperoxidase is not affected by curding according to Nandini and Rastogi (2011). It was concluded from the results of these studies that lactoferrin is a suitable marker for investigating the reduction of colostrum AnAc after curding.

Colostrum AnAc evolution during the refrigeration (4 °C) time among different treatments (control, 56 °C, 63 °C and skimmed) is represented in Table 2. While treatment did not affect colostrum AnAc, refrigeration time reduced AnAc through the studied period ($P < 0.001$). The largest decrease for colostrum AnAc was observed after 4 d of refrigerated storage, and after ten days the degree of reduction was 75, 85, 90 and 80% in control, 56 °C, 63 °C and skimmed colostrum treatments, respectively. On dairy farms, in order to facilitate the management during the birth seasons, it is necessary to refrigerate the colostrum. The effects of refrigeration on IgG concentration have been studied in goat colostrum (Arguello *et al.*, 2003). In that study the reduction in IgG concentration in the colostrum during the refrigeration was 11% over 14 d, suggesting that Ig are not associated with the decline in AnAc during refrigeration. Lactoferrin stability has been reported during refrigeration in human milk during the first 4 d, but no references were found for longer times (Giribaldi

et al., 2013; Rollo *et al.*, 2014). Lactoperoxidase has been reported to be active in raw milk for 7 d, and due to low concentration of sodium thiocyanate and hydrogen peroxide in milk the activity will be greater at the beginning of the refrigeration period (Gaya *et al.*, 1991).

In conclusion, colostrum antimicrobial activity was similar in goat and sheep. Regarding treatments, curding the colostrum reduced the colostrum AnAc of the whey. Additionally, refrigeration at 4 °C for up to 4 d of previously frozen-thawed colostrum is not recommended.

References

- Abd El-Gawad IA, El-Sayed EM, Mahfouz MB and El-Salam AM (1996) Changes of lactoferrin concentration in colostrum and milk from different species. *Egyptian Journal of Dairy Science* **24**, 12.
- Arguello A, Castro N, Capote J, Gines R, Acosta F and Lopez JL (2003) Effects of refrigeration, freezing-thawing and pasteurization on IgG goat colostrum preservation. *Small Ruminant Research* **48**, 135–139.
- Bhanu LS, Amano M, Nishimura SI and Aparna HS (2015) Glycome characterization of immunoglobulin G from buffalo (*Bubalus bubalis*) colostrum. *Glycoconjugate Journal* **32**, 625–634.
- Cacho NT and Lawrence RM (2017) Innate immunity and breast milk. *Frontiers in Immunology* **8**, 584. <https://doi.org/10.3389/fimmu.2017.00584>.
- Castro N, Capote J, Morales L, Quesada E, Briggs H and Arguello A (2007) Short communication: addition of milk replacer to colostrum whey: effect on immunoglobulin G passive transfer in Majorera kids. *Journal of Dairy Science* **90**, 2347–2349.
- Castro N, Capote J, Bruckmaier RM and Arguello A (2011) Management effects on colostrumogenesis in small ruminants: a review. *Journal of Applied Animal Research* **39**, 85–93.
- Daniels B, Schmidt S, King T, Israel-Ballard K, Mansen KA and Coutsoudis A (2017) The effect of simulated flash-heat pasteurization on immune components of human milk. *Nutrients* **9**, E178. doi: 10.3390/nu9020178
- Franco I, Perez MD, Conesa C, Calvo M and Sanchez L (2018) Effect of technological treatments on bovine lactoferrin: an overview. *Food Research International* **106**, 173–182.
- Gaya P, Medina M and Nuñez M (1991) Effect of the lactoperoxidase system on *Listeria monocytogenes* behavior in raw milk at refrigeration temperatures. *Applied and Environmental Microbiology* **57**, 3355–3360.
- Giribaldi M, Ortoffi MF, Giuffrida MG, Gastaldi D, Peila C, Coscia A, Raia M, Arslanoglu S, Moro GE, Cavallarin L and Bertino E (2013) Effect of prolonged refrigeration on the protein and microbial profile of human milk. *International Dairy Journal* **31**, 121–126.
- Hernandez-Castellano LE, Almeida AM, Ventosa M, Coelho AV, Castro N and Arguello A (2014) The effect of colostrum intake on blood plasma proteome profile in newborn lambs: low abundance proteins. *BMC Veterinary Research* **10**, 85. doi: 10.1186/1746-6148-10-85

- Hernandez-Castellano LE, Almeida AM, Renaut J, Arguello A and Castro N** (2016) A proteomics study of colostrum and milk from the two major small ruminant dairy breeds from the Canary Islands: a bovine milk comparison perspective. *Journal of Dairy Research* **83**, 366–374.
- Nandini KE and Rastogi NK** (2011) Integrated downstream processing of lactoperoxidase from milk whey involving aqueous two-phase extraction and ultrasound-assisted ultrafiltration. *Applied Biochemistry and Biotechnology* **163**, 173–185.
- Ripolles D, Harouna S, Parron JA, Calvo M, Perez MD, Carraminana JJ and Sanchez L** (2015) Antibacterial activity of bovine milk lactoferrin and its hydrolysates prepared with pepsin, chymosin and microbial rennet against foodborne pathogen *Listeria monocytogenes*. *International Dairy Journal* **45**, 15–22.
- Rollo DE, Radmacher PG, Turcu RM, Myers SR and Adamkin DH** (2014) Stability of lactoferrin in stored human milk. *Journal of Perinatology* **34**, 284–286.
- Trujillo AJ, Castro N, Quevedo JM, Arguello A, Capote J and Guamis B** (2007) Effect of heat and high-pressure treatments on microbiological quality and immunoglobulin G stability of caprine colostrum. *Journal of Dairy Science* **90**, 833–839.