

On the food of the Antarctic sea anemone *Urticinopsis antarctica* Carlgren, 1927 (Actiniidae, Actiniaria, Anthozoa)

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The results of an investigation into coelenteron content of the Antarctic sea anemone Urticinopsis antarctica Carlgren, 1927 are presented. Remains of invertebrate animals and fishes were found in the gastrovascular cavity of anemones. Some of them were damaged by digestion and were considered as food items of U. antarctica. These items were molluscs Addamussium colbecki (Smith, 1902), Laevilacunaria pumilia Smith, 1879, Eatoniella caliginosa Smith, 1875 and one not strictly identified gastropod species from the family Rissoidae; a crinoid from the family Comatulida; sea-urchin Sterechinus neumayeri Meissner, 1900; ophiuroid Ophiurolepis brevirma Mortensen, 1936 and a fish Trematomus sp. In contrast to the prey mentioned above, three specimens of amphipods Conicostoma sp. were not destroyed by digestion. They may represent commensals, which live in the gastrovascular cavity of the anemone.

Keywords: Antarctica, *Urticinopsis antarctica*, prey capture, coelenteron content, diet, generalist

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INTRODUCTION

Sea anemones are well represented in marine benthic communities and are adapted to variable environmental conditions (Manuel, 1988). They are very common in shallow waters of the Antarctic, inhabit hard as well as soft substrata and often enter into symbiotic or commensal relationships with animals of other groups.

Life in diverse habitats requires differing strategies of obtaining food. As with other anthozoans, sea anemones have the following modes of nourishment: (1) capturing solid food, (2) absorbing dissolved organic matter (DOM) and (3) using assimilates of their symbiotic algae (Schlichter, 1978). All those kinds of nourishment can be applied together or separately.

There are different ways of prey collecting. When prey is suspended in the water column (usually zooplankton) it has to be intercepted by one or more tentacles and transferred to the mouth. Sessile prey, which can be dislodged by wave action or by foraging predators, is pushed into the tentacle crown. As concerns motile prey it simply blunders into the anemone's tentacles (Sebens, 1981). Each of these modes of prey capture requires corresponding morphological adaptations. For example, according to the observations realized by means of the Russian manned underwater vehicle 'Sever-2' during the 33rd cruise of research-vessel 'Odyssey', 1984 (Sirenko, 1993), a deep-water sea anemone *Actinostola callosa* extracted organic particles with numerous tentacles

disposed on the surface of a wide oral disc. The disc in this anemone can assume the form of a tube that allows selecting of food particles from water passing through it (Figure 1. 1–3). However, this species of anemone is able to capture big prey. According to German researchers, this anemone, an inhabitant of Norwegian fjords, was also discovered feeding on the coronate medusa *Periphylla periphylla*. Sea anemones were observed by means of a remotely operated vehicle (ROV); it turned out that *A. callosa* can completely swallow a medusa in half an hour. In laboratory experiments it took more than 40 min to engulf the medusa, the diameter of which exceeded the size anemone. The *Periphylla* population does not show any significant seasonality and its biomass is much larger than the biomass of the anemone, which might explain the high population density of *A. callosa* (Jarms & Tiemann, 2004).

In other cases anemones have long and numerous tentacles able to search for food by 'sweeping' the substrate, as it is typical for *Anemonia viridis* (Forsskål, 1775) (Chintiroglou & Koukouras, 1992).

The food of many sea anemones consists of almost any benthic organisms which can be caught and swallowed: crustaceans, worms, molluscs, fish, etc. The anemones are often able to eat objects of large size relative to their own bulk. Nevertheless in some cases laboratory experiments have demonstrated that *A. equina* indiscriminately ingests small prey, but size of prey is still restricted, because larger objects cannot be gulped (Davenport *et al.*, 2011). Many anemones are omnivorous animals. However, it does not mean that a species may not have a regular diet in which one or two items predominate (Stephenson, 1928). Moreover factors of environment (depths, different types of substrata, wave action) and time (seasons, daytime and night time) influence the accessibility of prey in communities and indirectly to the

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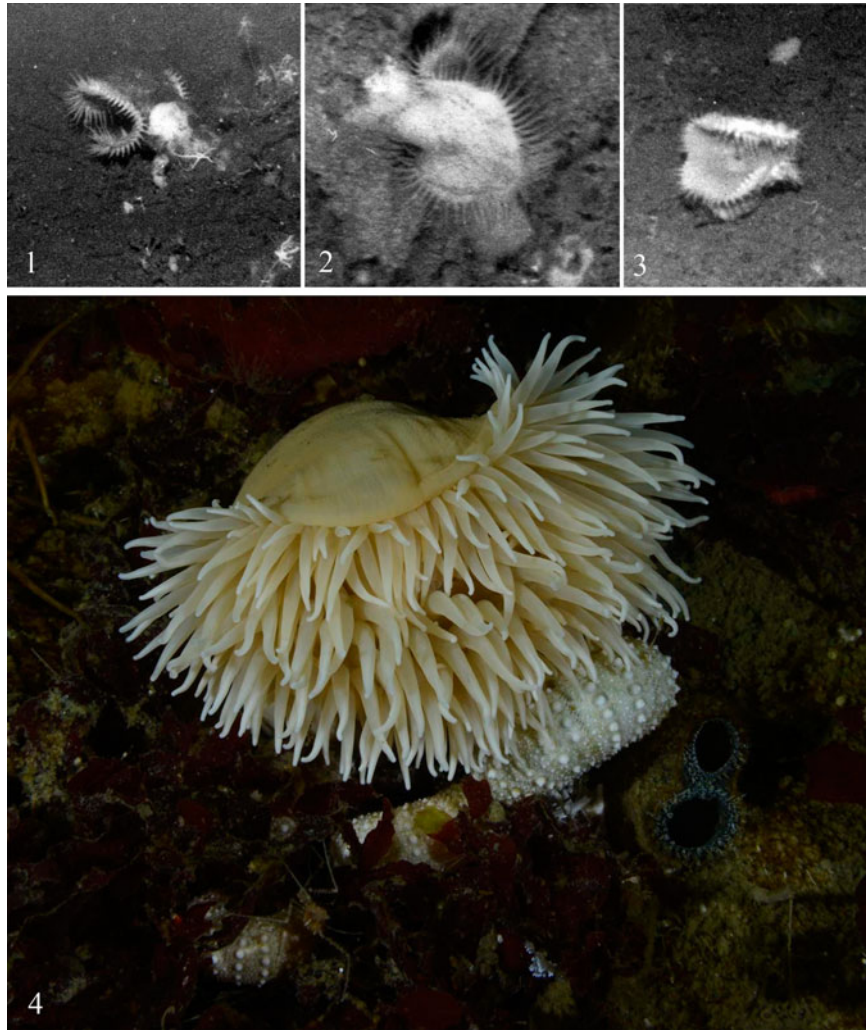


Fig. 1. Difference in form of the oral disc and tentacle distribution in deep-water sea anemone *Actinostola callosa* and Antarctic coastal inhabitant *Urticinopsis antarctica*. 1–3. *A. callosa* with tube-like rolled up oral disc (Photo by B. I. Sirenko). 4. *U. antarctica* with long and numerous tentacles with tests of three sea-urchins eaten by it. (Photo by O. V. Savinkin.)

diet of sea anemones (see Chintiroglou & Koukouras, 1992; Kruger & Griffiths, 1996; Davenport *et al.*, 2011; Quesada *et al.*, 2014).

The goal of this study was to examine the composition of the diet of *Urticinopsis antarctica*.

MATERIALS AND METHODS

Urticinopsis antarctica is one of the most common and plentiful sea anemones of the Antarctic coast, therefore it is well represented in the collection of Zoological Institute (the list of samples is presented below). Our study and data published before show that the species is widespread in Antarctic and Subantarctic waters (Figure 2). It is recorded from McMurdo Bay, South Shetland Islands, Prudz Bay, Cosmonauts Sea, Haswell Archipelago (Davis Sea) and the Weddell Sea. The predatory feeding habits of *Urticinopsis antarctica* could be confirmed by observations of research-divers and analysis of the coelenteron contents.

Diet is usually studied by analysis of gastrovascular cavity content. It often encounters some difficulties owing to different extent of digestion, egestion and poor preservation of prey

items. The possibility of accidental ingesting of shells and some non-food animals during sea anemone collecting also should be taken into account (Lampitt & Paterson, 1987). In spite of these difficulties this method is widely used, and was employed in our study too.

Twenty-three specimens of sea anemone *Urticinopsis antarctica* were dissected in the course of the study of its morphology and variability of taxonomically important characters (Figure 3. 6). These have been deposited in the collection of the Zoological Institute of RAS. These samples were collected by Soviet, Russian and International Antarctic expeditions (see Table 1).

All the specimens of *U. antarctica* were fixed in formaldehyde solution and preserved in 70% ethanol. Anemones were dissected by transverse cutting on the level of the pharynx and immediately above the base. Then they were also cut longitudinally along the endocoel of one of the directive pairs of mesenteries. This method of anatomical study gives a high possibility of more careful examination of the gastrovascular cavity. The specimens were dissected by use of scalpel or a blade in conformity with the size of the animals. The food items were counted and then identified to the lowest possible taxonomic level by specialists of ZIN RAS.

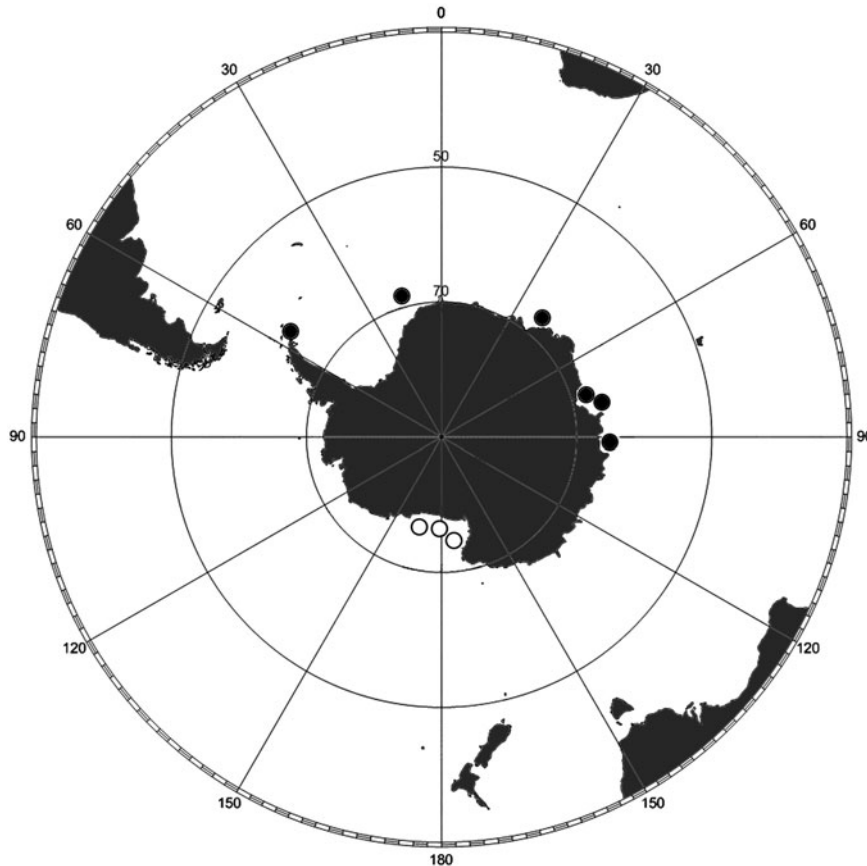


Fig. 2. Distribution of *Urticinopsis antarctica*. Black circles – our data, samples from collection of the Zoological Institute; empty circles – localities from literature data.

To study the quantitative diet composition we used the method detailed in Chintiroglou & Koukouras (1992). The following parameters were calculated:

$$\text{Vacuity coefficient } V = Ev \times 100/N$$

where Ev , the number of empty coelenterons, N , the total number of coelenterons examined.

RESULTS

Among 23 individuals of *Urticinopsis antarctica* only six contained eaten animals, giving a vacuity coefficient of ~74%. All identifiable animals found in the coelenterons are listed in Table 2. Four anemones had in their gastric cavity the remains of invertebrate animals and one enclosed the remains of a fish, rather damaged by digestion. These organisms were considered to be food items for the studied anemone species (Figure 3. 1–4). So we can regard as prey of *Urticinopsis antarctica* four mollusc species – *Laevilacunaria pumilia* Smith, 1879 and *Eatoniella caliginosa* Smith, 1875 (Gastropoda; 56 RAE, King George's Island, 4 m), *Addamussium colbecki* (Smith, 1902) (Bivalvia; 54 RAE, Prudz Bay, 5–6 m) and one not strictly identified representative of the family Rissoidae (Gastropoda; 13 SAE, Cosmonauts Sea, 28–30 m); a single specimen of crinoids (Crinoidea; 'Polarstern' 39 th cruise, Weddell Sea, 504–529 m); sea-urchin *Sterechinus neumayeri* Meissner, 1900

(Echinoidea, 56 RAE, King George Island, 4 m), needles of which were found in gastric cavity; a small ophiuroid *Ophiurolepis brevissima* Mortensen, 1936 (Ophiuroidea; 'Polarstern' 39th cruise, Weddell Sea, 504–529 m). In addition to this some fish bones (probably belonging to *Trematomus* sp., Nototheniidae; 54 RAE, Prudz Bay, 4–5 m) were also found. The fragments of unidentifiable algae were also discovered in one of the specimens.

All molluscs found in sea anemone guts were represented only by empty shells, which were nearly unharmed; only the edge of bivalve valves were slightly damaged (Figure 3. 1–4). Remains of the fish were represented by vertebrae. Findings of echinoderms were different. The test of sea-urchin was probably ejected since only needles were discovered in the actinopharynx. In contrast, crinoid skeleton (calyx and proximal parts of hands) and nearly intact ophiuroid were found in the gastric cavity (see Table 2).

In contrast to all listed animals, three specimens of scuds *Conicostoma* sp. (Amphipoda; 'Polarstern' 39th cruise, Weddell Sea, 504–529 m) were in very good condition (Figure 3. 5). They were not destroyed by digestion and therefore may represent commensal organisms, which live in close association with *Urticinopsis antarctica* (see discussion).

DISCUSSION

The analysis of *Urticinopsis antarctica* coelenteron content distinctly showed the ability to catch and eat different low

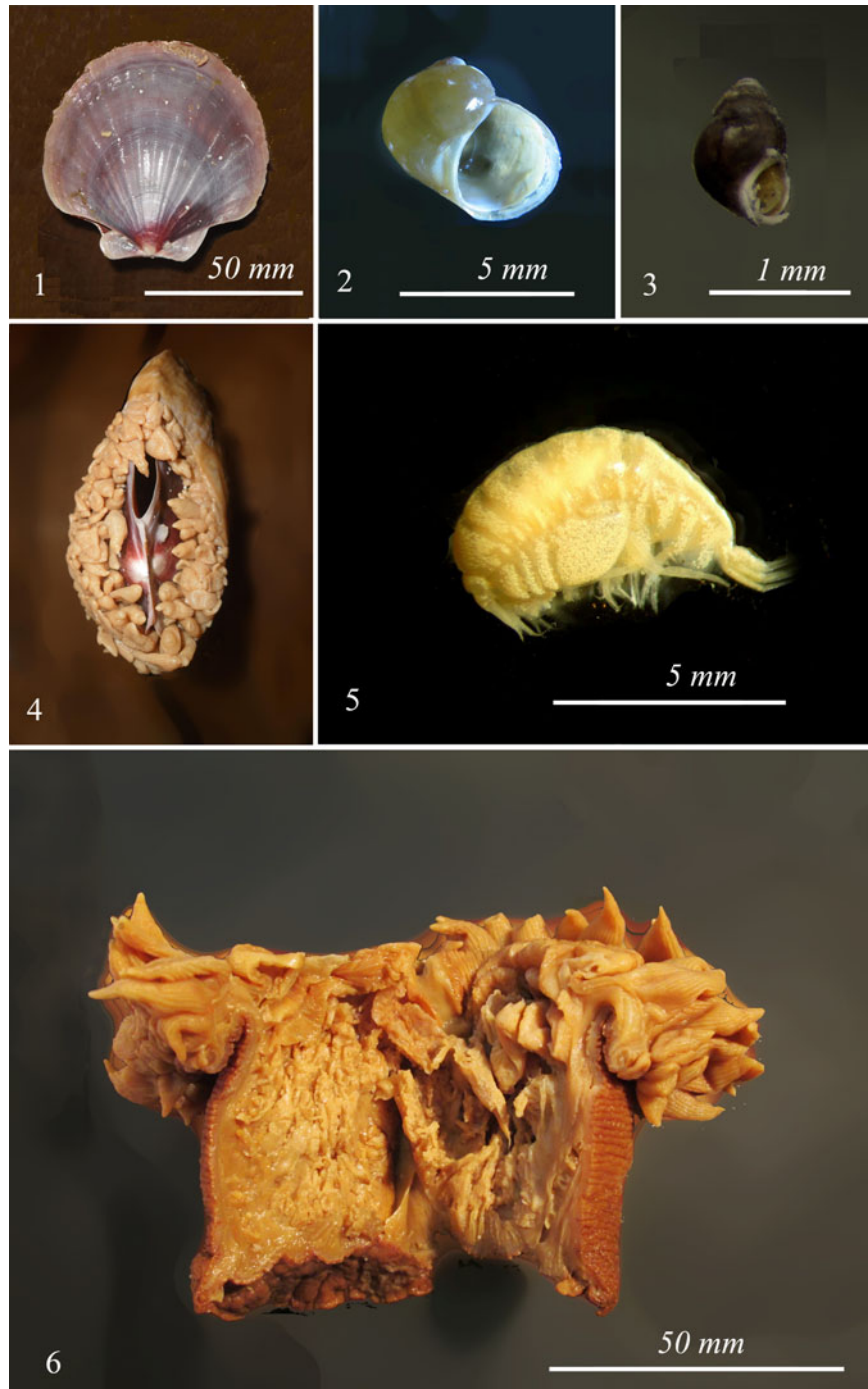


Fig. 3. Dissected specimen of the *Urticinopsis antarctica* and the most intact food items from the gastric cavity of sea anemones. 1. *Addamussium colbecki* (Smith, 1902) (Bivalvia). 2. *Laevilacunaria pumilia* Smith, 1879 (Gastropoda). 3. *Eatoniella caliginosa* Smith, 1875 (Gastropoda). 4. The anemone with its prey in pharynx – *A. colbecki* (Smith, 1902). 5. Symbiotic amphipod *Conicostoma* sp. 6. Dissected sea anemone.

mobile invertebrate animals and also some fish, *Trematomus* sp., which live on the seafloor and are primarily benthic feeders (Brueggeman, 1998). As with other motile animals, fish occasionally blunder into an anemone's tentacles and are captured. Sessile organisms are probably dislodged by wave action or by foraging predators (see Introduction). Big body size (up to 120 mm height in preserved condition) and presence of numerous long tentacles also indicate the ability to capture quite large benthic animals.

We don't see that the actinian species has a distinct food preference. So we consider this anemone to be a generalist, according to the classification of Kruger & Griffiths (1996). However, a small number of studied specimens and poor gastric cavity content are not enough for reliable judgement. However, our observations and data from literature show that *Urticinopsis antarctica* slightly prefers to feed on Echinodermata. They are sea stars *Perknaster fuscus antarcticus* (Koehler 1906), *Odontaster validus* Koehler 1906, *Diplasterias*

Table 1. Samples of *Urticinopsis antarctica* collected by Soviet, Russian and International Antarctic expeditions.

Expeditions	Stations	Regions	Depth, m	No ^a
11 SAE	Molodezhnaya	Cosmonauts Sea	12	11,381
13 SAE	Molodezhnaya	Cosmonauts Sea	26–28	11,386
13 SAE	Molodezhnaya	Cosmonauts Sea	28–30	11,382
13 SAE	Mirny	Davis Sea	10–30	11,390
16 SAE	Progress	Prydz Bay, Sodruzhestvo Sea	3–20	11,388
16 SAE	Progress	Prydz Bay, Sodruzhestvo Sea	12–25	11,383; 11,389
Ice-breaker 'Polarstern' 39th cruise	–	Weddell Sea 71°30.30' S 12°27.80' W	504–529	11,387
54 RAE	Progress	Prydz Bay, Sodruzhestvo Sea	5–6	11,391; 113,91b
54 RAE	Progress	Prydz Bay, Sodruzhestvo Sea	4–5	11,378; 11,380; 11,384; 11,385a; 11,385b; 11,385c
54 RAE	Progress	Prydz Bay, Sodruzhestvo Sea	12–13	11,652
56 RAE	Bellinshauzen	King George Island, 62°12.372'S; 58°56.818'W	4	11,379; 11,392a; 113,92b; 113,93a; 11,393b
59 RAE	Progress	Prydz Bay, Sodruzhestvo Sea	30–50	11,653

^aNumbers of specimens are noted following Incoming catalogue of Department of Porifera and Coelenterata, Zoological Institute, Russian Academy of Sciences.

Table 2. Content of gastrovascular cavity of Antarctic sea anemone *Urticinopsis antarctica*.

Group of organisms	Condition (degree of digestion)	Identifiable taxa		No. (specimens)
		Algae	Not identifiable	
Plants	Fragments			
Animals				
Molluscs	Only shells	Bivalvia	<i>Addamussium colbecki</i> (Smith, 1902)	1
		Gastropoda	Rissoidea (species is unidentified)	1
			<i>Laevilacunaria pumilia</i> Smith, 1879	1
			<i>Eatoniella caliginosa</i> Smith, 1875	1
Sea lilies	Skeleton of calyx and proximal part of hands	Crinoidea	Comatulida (species is unidentified)	1
Sea-urchins	Needles	Echinoidea	<i>Sterechinus neumayeri</i> Meissner, 1900	– [needles of one or many exemplars]
Brittle stars	Nearly intact	Ophiuroidea	<i>Ophiurolepis brevirima</i> Mortensen, 1936	1
Sea cucumber	Intact	Holothuroidea	<i>Heterocucumis steineni</i> (Ludwig, 1898)	1
Side-swimmers	Intact, not damaged	Amphipoda	<i>Conicostoma</i> sp.	3
Fishes	Vertebrae	Nototheniidae	<i>Trematomus</i> sp.	1

brucei (Koehler 1908) and sea-urchin *Sterechinus neumayeri* Meissner, 1900, which compose about 77% of its diet (Dayton *et al.*, 1970). Another representative of this invertebrate group which also could be part of the diet of *U. antarctica* is the holothurian *Heterocucumis steineni* (Ludwig, 1898); unfortunately this animal was identified only in a photo (Figure 4).

Moreover, the predatory feeding habits of *Urticinopsis antarctica* were confirmed by observations of research-divers B.I. Sirenko and A.F. Pushkin, the participants of Soviet Antarctic expeditions (ZIN RAS). According to the divers' personal observation of Sirenko, a number of sea-urchin naked tests or 'shells' frequently surround large specimens of *Urticinopsis antarctica* at the bottom of Prydz Bay (Sodruzhestvo Sea, see Figure 1. 4). Even more exhaustive experiments were carried out by Pushkin in the natural habitat (Haswell Archipelago, Davis Sea). During his dives he regularly put a quite large sea-urchin (probably *Sterechinus neumayeri*) at the oral disc of *Urticinopsis*. It

turned out that the anemone can digest a sea-urchin and throw out its 'shell' in a few days.

Study of the stinging capsules set of *U. antarctica* showed that the capsules typical for scyphozoan medusas (Scyphozoa) sometimes occur in endodermal epithelium of the pharynx (our observations). These capsules have proper orientation there and seem to function as cleptocnidae, known in other invertebrates (for example, nudibranch molluscs), which are able to eat cnidarian polyps and medusas and to use their stinging capsules. The latest observation concurs with those of American researchers, who believed that medusas constitute 21% of *Urticinopsis antarctica* diet (Dayton *et al.*, 1970).

In contrast to molluscs, echinoderms and fish, three specimens of side-swimmers *Conicostoma* sp. were not destroyed by digestion. It is known that representatives of this and related genera are associated with sea anemones and sponges (Barnard & Karaman, 1991). Thus the observed



Fig. 4. Holothurian *Heterocucumis steineni* half-swallowed by *Urticinopsis antarctica*. (Photo by O. V. Savinkin.)

specimens may represent not prey items, but commensals of *Urticinopsis antarctica*.

Hence, all available data show that the large and common in the Antarctic anemone *Urticinopsis antarctica* is probably able to feed on any big benthic and even some nektonic animals. Moreover, according to American researchers, cannibalism is also possible in this species (Dayton *et al.*, 1970, Figure 4).

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REFERENCES

Barnard J.L. and Karaman G.S. (1991) The families and genera of marine gammaridean Amphipoda (except marine gammaroids). Part 2. *Records of the Australian Museum, Supplement* 13, 419–866.

Bruggeman P. (1998) Underwater field guide to Ross Island & McMurdo sound, Antarctica. Chordata: fish. <http://www.peterbruggeman.com/nsf/fguide/chordata--fish.pdf>

Chintiroglou Ch. and Koukouras A. (1992) The feeding habits of three Mediterranean sea anemone species, *Anemonia viridis* (Forsk.), *Actinia equina* (Linnaeus) and *Cereus pedunculatus* (Pennant). *Helgolander Meeresunters* 46, 53–68.

Davenport J., Moloney T.V. and Kelly J. (2011) Common sea anemones *Actinia equina* are predominantly sessile intertidal scavengers. *Marine Ecology Progress Series* 430, 147–155.

Dayton P.K., Robilliard G.A. and Paine R.T. (1970) Benthic faunal zonation as a result of anchor ice at McMurdo Sound, Antarctica. In Holgate M.W. (eds) *Antarctic ecology*. Volume 1. New York, NY: Academic Press.

Jarms G. and Tiemann H. (2004) *Actinostola callosa* (Verrill, 1882) (Actinostolidae, Anthozoa), a medusivorous sea anemone and its mass occurrence in the Lurefjord, Norway. *Helgoland Marine Research* 58, 15–17.

Kruger L.M. and Griffiths Ch.L. (1996) Sources of nutrition in intertidal sea anemones from the south-western Cape, South Africa. *Southern African Journal of Zoology* 31, 110–119.

Lampitt R.S. and Paterson G.L.J. (1987) The feeding behaviour of an abyssal sea anemone from *in situ* time lapse photographs and trawl samples. *Oceanologica Acta* 10, 455–461.

Manuel R.L. (1988) *British Anthozoa (Coelenterata: Octocorallia and Hexacorallia)*. Synopses of the British Fauna; new ser., no. 18 (rev). London: Academic Press.

Quesada A.J., Acuña F.H. and Cortés J. (2014) Diet of the sea anemone *Anthopleura nigrescens*: composition and variation between daytime and nighttime high tides. *Zoological Studies* 53, 26.

Schlichter D. (1978) On the ability of *Anemonia sulcata* (Coelenterata: Anthozoa) to absorb charged and neutral amino acids simultaneously. *Marine Biology* 45, 97–104.

Sebens K.P. (1981) The allometry of feeding, energetics, and body size in three sea anemone species. *Biological Bulletin* 161, 152–171.

Sirenko B.I. (1993) Distribution of benthos in some areas of the continental slope of Kurile Islands. In Sirenko B.I. and Vassilenko S.V. (eds) *The fauna of the continental slope of the Kurile Islands. Based on collections of 33 voyage r/v "Odyssey"*. 46(54). Saint Petersburg: Zoological Institute Russian Academy of Sciences, pp. 5–44. [In Russian].

and

Stephenson T.A. (1928) *The British sea anemones*. Volume 1. London: The Ray Society.

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