The association between invasive *Lantana camara* and seedlings/saplings of a plant community in Mudumalai Tiger Reserve, India

Gaius Wilson*,1, Ajay A. Desai†, Dalice A. Sim‡, Monica A. M. Gruber* and Philip J. Lester*

* School of Biological Sciences, Victoria University of Wellington, Kelburn Parade 6140, Wellington, New Zealand

† 84 BC Camp, Belgaum, Karnataka, 590 001, India

‡ School of Mathematics, Statistics and Operations Research, Victoria University of Wellington, Kelburn Parade 6140, Wellington, New Zealand (Received 17 February 2014; revised 1 July 2014; accepted 1 July 2014; first published online 8 August 2014)

Abstract: We examined changes in a community of seedlings/saplings 10–150 cm tall associated with the presence of a widely invasive plant, *Lantana camara* and environmental covariates along 67 randomly located transects, in Mudumalai, India. We compared plant species assemblage and grass cover in *L. camara*-invaded and uninvaded plots in three habitats. Multivariate analyses revealed a significant association of all environmental covariates with plant species assemblage. Pairwise tests indicated that *L. camara* was significantly associated with changes in plant species assemblage and grass cover within the moist and dry deciduous forest, but not in the thorn forest. The relationship between *L. camara* and that of elephant browse plants varied with species. A linear regression analysis indicated that *L. camara* invasion was the only significant predictor of grass occupancy. Our results indicate that in addition to other factors, *L. camara* was associated with altering plant species assemblage, some elephant browse plants and grass cover in the moist and dry deciduous forest. It appears that *L. camara* can have a major effect on diversity within this reserve, but whether this effect is by *L. camara* driving the change or being associated with other habitat change requires further experimental evidence.

Key Words: anthropogenic disturbance, elephants, grass cover, invasive weeds, Lantana camara, plant species assemblage

INTRODUCTION

Exotic plant species may modify native communities by altering soil properties such as nutrient cycling (Ehrenfeld *et al.* 2001), hydrology (Melgoza *et al.* 1990), be allelopathic (Gentle & Duggin 1997a), or compete with native species for light and nutrients (Braithwaite *et al.* 1989, Woods 1993). Native forage species used by herbivores as food may therefore receive only limited resources due to competition with exotic plants, thus causing native species to become locally extinct or to persist at very low densities (Bedunah 1992). Changes to the vegetation community through a decline of native forage species brought about by exotic weeds could have the potential to precipitate food-web-level, bottom-up meltdown (sensu Terborgh *et al.* 2001).

Exotic plants often require some form of disturbance for them to establish (Buckley *et al.* 2007, Duggin & Gentle 1998). In addition to the impact of exotic plant invasions on native plant communities, a number of studies have shown that anthropogenic disturbances can also alter plant communities (Godefroid & Koedam 2004). Biotic factors such as tree density, canopy cover, grass cover and abiotic factors such as fire, distance to roads and settlements are also responsible for changes to the vegetation community (Morrison *et al.* 1995, Oliveira-Filho *et al.* 1998).

Megaherbivores such as the Asian elephant (*Elephas maximus*) are adapted to live in diverse habitats and feed on a variety of plant species (Baskaran *et al.* 2010, Owen-Smith 1988). However, despite their ability to exploit a wide range of forage species, elephants may be influenced by the establishment and spread of exotic invasive plants especially if these exotic plants are not eaten by elephant and replace native forage species (Wilson *et al.* 2014). The establishment of exotic invasive plants often leads to displacement and decline of native forage species (Lym & Kirby 1987).

Mudumalai Tiger Reserve (hereafter Mudumalai) in southern India forms a part of the Nilgiri Biosphere Reserve which hosts the single largest Asian elephant

¹ Corresponding author. Email: gaiuswilson@hotmail.com



Figure 1. Mudumalai Tiger Reserve and its location in India showing the layout of 67 transects across the reserve within the three habitats. Habitat boundaries are delimited by dashed lines: moist deciduous, dry deciduous and thorn forest. Plantations are shown as black patches and settlements as grey patches. The road network within the reserve is shown.

population. In Mudumalai, one study estimates that browse forms 15% of elephant diet while grass forms nearly 85% of elephant diet (Baskaran *et al.* 2010). One of the physical impacts *Lantana camara* L. has is the reduction of grass cover. As *L. camara* spreads, grass cover declines (Kumar *et al.* 2012). This reduction in major elephant food source could lead to detrimental effects on elephants and their habitats (Prasad 2012). For large herbivores, whose populations are not regulated through natural predation, it is likely that the availability of food is the limiting resource (Owen-Smith 1988, Sinclair 1975). Thus food resources are vital to maintaining elephant health and abundance.

Lantana camara has invaded India's tropical dry forests and appears to be associated with a reduction in food species of native herbivores (Prasad 2012). Elsewhere, sites invaded by *L. camara* generally have lower plant species richness and diversity (Prasad 2010, Sharma & Raghubanshi 2007), and the weed is also thought to impede the growth of grass and native seedlings (Gooden *et al.* 2009, Kumar *et al.* 2012). For these reasons, many reserves manage habitat by investing resources in *L. camara* removal, especially by cutting and uprooting plants (Srivastava 2009). We tested the hypotheses that *L. camara*, along with other biotic and abiotic environmental covariates was significantly associated with (1) plant species assemblage, (2) three elephant browse plants present throughout the reserve in a plant community of seedlings/saplings 10–150 cm tall and (3) grass cover. Grass cover was examined because of the importance of grass in elephant diet (Baskaran *et al.* 2010, Wilson *et al.* 2014). We use the term 'association' here because this study was not a manipulative experiment.

METHODS

Study site and sampling design

The study was conducted in Mudumalai Tiger Reserve (Figure 1) southern India. Data were collected from 10×1 -m plots located every 100 m along 67 1-km long randomly located transects as described by Wilson *et al.* (2013). The 10×1 -m sampling plots located every 100 m were spaced in an attempt to ensure that the plots along each transect were independent given the size of the plots and the gap between each plot.

We first recorded *L. camara* presence or absence within each 10×1 -m plot along each transect to examine changes that were associated with differences in plant species assemblage and grass cover. We were also interested in whether the abundance of *L. camara* in a plot was associated with differences in three elephant browse plant species. To measure *L. camara* abundance, stem density of *L. camara* in each plot was recorded. To estimate *L. camara* invasion, the age of the stand, defined by average *L. camara* girth of all stems in a plot, was used because in field observations we noted that older stands had fewer *L. camara* plants (as few individuals dominate while others die out) as has been noted elsewhere (Swarbrick *et al.* 1998). By contrast, younger stands had more individual plants. The girth of all *L. camara* stems were measured at ground level within 10×1 -m plots and recorded in 1-cm categories. An estimate of the average girth for each plot was derived.

Plant species (shrubs and saplings, between 10 and 150 cm) were identified from herbarium specimens, field guides and knowledgeable field assistants and counted in plots measuring 10×1 -m located every 100 m along each transect in order to measure plant species assemblage at each plot (see Appendix 1 for a list of plant species). Biotic and abiotic environmental covariates that could potentially be associated with plant species assemblage were measured in each plot. Biotic covariates included tree density, canopy cover and grass cover. Tree density, canopy cover and percentage grass cover along each 1km transect was estimated every 100 m in 10×1 -m plots (Wilson et al. 2013). All grasses were grouped together without distinguishing the various species. The percentage of bare ground, other vegetation (trees, herbs, shrubs) and rocks, was also visually estimated at the same site. The percentage of grass occupancy (area of grass cover/area available to grass after deducting native vegetation, bare ground and rocks) was also calculated to provide a measure of the area in a plot that was actually occupied by grass or L. camara.

Abiotic environmental covariates related to anthropogenic disturbances and included distance to roads and settlements, and time since last fire burn. Linear distances between each sampling plot and the closest road and settlement were measured from 1:50 000 topographic maps, using MapInfo Professional 7.8 (MapInfo Corporation, Troy, NY, USA). As the size and thus potential impact of roads and settlements varied throughout Mudumalai, we used three categorical factors for settlements: (1) if a plot fell more than 2 km from a minor settlement ($\leq 0.1 \text{ km}^2$); (2) if a plot fell within 2 km from a minor settlement; and (3) if a plot fell within 2 km of a major settlement (≥ 0.1 km²). Similarly, for roads: (1) if a plot fell more than 2 km from a forest road (grey lines, Figure 1); (2) if a plot fell within 2 km from a forest road (grey lines, Figure 1); and (3) if a plot fell within 2 km of a main/public road (grey double lines, Figure 1). Within Mudumalai, smaller forest roads that were used only by the forest department's tourist vehicles were assumed to have less impact than the main/public road and were presumed to have minimal impact on weed distribution. Data on anthropogenic fire during the 6 y prior to the study (2003-2008) in each plot were obtained using the same methods described by Wilson et al. (2013).

Statistical analyses

To assess differences in plant species assemblage in plots that were invaded and uninvaded by L. camara including environmental covariates, we used PERMANOVA+ using 9999 permutations implemented in PRIMER v 6.1.11 (Clarke & Gorley 2006). Only shrubs and saplings measuring between 10 and 150 cm in height were used. Lantana camara was excluded from the analysis of plant species assemblage, and was used only to define invaded/uninvaded groups. The data were logtransformed (log (x + 1)) prior to analyses. A Bray-Curtis index was used as a similarity measure for plant species assemblage (Clarke & Warwick 2001). We used non-metric Multi-Dimensional Scaling (nMDS), also implemented in PRIMER to determine whether plant species assemblage differed in the three habitats of the reserve. The nMDS was run over 1000 iterations using Kruskal stress formula 1 and a minimum stress of 0.01. To investigate the association of *L. camara* presence/absence with plant species assemblage further, we examined the output of PERMANOVA which includes pairwise tests within each habitat comparing plots with and without L. camara. The pairwise tests were conducted by including each habitat as a factor and L. camara presence/absence as a second factor. We were therefore able to define the association of L. camara within each habitat without analysing the data habitat-wise. As there were a large number of potential interactions between various factors, we a priori decided to examine only the interaction between habitat and L. camara presence/absence.

In order to examine how individual species contributed to the differences in plant species assemblage between L. camara-invaded and uninvaded plots, we used SIMPER subroutine (analysis of per cent similarity) (PRIMER v 6.1.11) based on a Bray-Curtis similarity measure, with a log-transformation of the data $(\log (x + 1))$. The top three elephant browse food plants (saplings between 10-150 cm tall) that contributed most to the dissimilarity from the SIMPER analysis were then used to examine the slope of the relationship with L. camara abundance using linear regression analysis conducted in SPSS Statistics, release version 20.0 (IBM SPSS Inc., Chicago, IL, USA). The effect size of L. camara on each of the species that contributed to the average dissimilarity among habitats and between invaded and uninvaded plots, were derived using 'adonis' function in the 'vegan' package in R (version 3.0.2; http://www.R-project.org).

Given the importance of grass in elephant diet (Baskaran *et al.* 2010, Wilson *et al.* 2014), we conducted an analysis on percentage grass cover to study the association of *L. camara* and other environmental covariates with percentage grass cover. A Bray–Curtis index was used as a similarity measure for percentage grass cover. PERMANOVA+ was used to run 9999

permutations to test for an association of *L. camara* presence/absence and environmental covariates (biotic and abiotic) with percentage grass cover. Biotic and abiotic factors used as environmental covariates were *L. camara* presence/absence, tree density, canopy cover, impact of roads, settlements and fire. PERMANOVA was used to conduct pairwise tests to compare plots with and without *L. camara* within each habitat to examine these differences. As above, only interactions between habitat and *L. camara* presence/absence were examined and not all factor interactions.

A linear regression analysis was then conducted on percentage grass occupancy (defined as given above), to test for an association of *L. camara* invasion (average girth per plot) along with other environmental covariates, which included impact of roads, settlements, canopy cover, fire, tree density, DDF × *L. camara* interaction and MDF × *L. camara* interaction term with the percentage grass occupancy using SPSS Statistics, release version 20.0 (IBM SPSS Inc., Chicago, IL, USA). The TF was used as the reference category for the dummy variable and hence its interaction term with *L. camara* was not included in the model. Percentage grass occupancy per plot which was the outcome variable was arcsine-square root transformed for normality.

RESULTS

Lantana camara has invaded large areas of Mudumalai. Overall, 59% of the sampling plots (n = 737) were invaded by *L. camara* throughout the reserve. The thorn forest (TF) (n = 165) had more *L. camara*-invaded sampling sites than the other habitats with 88% of sites invaded by *L. camara*. Of the sites sampled in the moist deciduous forest (MDF) (n = 132), 43% remained uninvaded while 52% in the dry deciduous forest (DDF) (n = 440) were uninvaded by *L. camara*. The density of *L. camara* varied throughout the reserve in different habitats from no *L. camara* to 39 stems per 10×1 -m plot with an interquartile range of 4 stems per 10×1 -m plot (25th percentile = 0 stems; 75th percentile = 4 stems). Of the 737 plots, only seven plots had no plants because they were occupied by bare earth or rock or by trees, while other plots had between 1 and 53 plant species. The total number of plant species identified within the plots throughout the reserve was 136 with *Catuneragum spinosa* (Rubiaceae) being the most common species (340 individuals). Plant species richness was highest in two plots along two different transects with a total of 53 plant species; one of these transects was found in the DDF while the other in the MDF.

Plant species assemblage

All environmental covariates, including the presence of L. camara in a plot were associated with the plant species assemblage (P < 0.001, Table 1). The largest component of variation was from habitat (14%), followed by L. camara presence/absence and roads (each 8%), settlements and the interaction of habitat and L. camara presence/absence (7%). The lowest component of variation was tree density (2%). Pairwise tests indicated that in both the MDF (t_{123} = 1.51, P = 0.006) and DDF (t_{430} = 4.39, P < 0.001), the presence of L. camara was significantly associated with differences in plant species assemblage, while there were no significant differences in the TF ($t_{157} = 1.13$, P = 0.238) whether L. camara was present or not. Twodimensional nMDS plot did not reveal clear differences in plant species assemblage in the three habitats, perhaps related to the poor fit of the data to a two-dimensional nMDS plot (Stress > 0.20, Figure 2).

Elephant browse plants

SIMPER analysis indicated that *C. spinosa* contributed 12.1% to the average similarity between habitats and

Table 1. Results of PERMANOVA (permutation analysis of variance) examining the association of *Lantana camara* presence/absence and environmental covariates with plant species assemblage in Mudumalai Tiger Reserve, India. All environmental covariates were statistically significant. PERMANOVA also gives a component of per cent variation for each predictor which is equivalent to the sum of the squared fixed effects divided by the degrees of freedom from standard ANOVA.

Environmental covariates	df	Pseudo-F	P(perm)	Estimate	Per cent variation
Habitat	2	20.7	< 0.001	265.2	13.6
Lantana camara (presence/absence)	1	14.7	< 0.001	93.2	8.1
Roads	2	7.8	< 0.001	87.4	7.8
Settlements	2	5.8	< 0.001	70.7	7.0
Habitat × Lantana camara (presence/absence)	2	3.0	< 0.001	59.5	6.5
Fire	1	18.8	< 0.001	54.9	6.2
Canopy cover	1	19.7	< 0.001	53.5	6.1
Grass cover	1	9.1	< 0.001	23.4	4.0
Tree density	1	3.8	< 0.001	8.1	2.4
Residuals	723			2110.9	38.4
Total	736				



Figure 2. A non-metric Multi-Dimensional Scaling (nMDS) plot generated in PRIMER v 6.1.11 showing the dissimilarities of the various species in the three habitats of Mudumalai Tiger Reserve. The two-dimensional nMDS plot shows that there is a high stress level (0.26) which is higher than the maximum stress level of 0.20. Given the high stress level and the poor representation of the data which is highly distorted we abandoned the nMDS. The three habitats are (1) moist deciduous forest represented by a star, (2) dry deciduous forest represented by a hollow circle and (3) thorn forest represented by a triangle.

between plots with and without *L. camara*, followed by *Phyllanthus emblica* (Phyllanthaceae) (7.3%), *Shorea roxburghii* (Dipterocarpaceae) (5.7%), *Cassia fistula* (5.1%) and *Grewia tiliifolia* (Malvaceae) (4.5%) (Table 2). All these plants except for *C. fistula* are elephant browse food plants and the most important species in differentiating those plots with and without *L. camara*. Of the browse species that were estimated to contribute most to elephant diet by Baskaran *et al.* (2010), bamboo spp. (Gramineae) and *Kydia calycina* (Malvaceae) contributed only 3% to the average similarity, and only bamboo spp. were found in all three habitats.

The association between *L. camara* presence/absence and *C. spinosa* in the MDF was significant (MDF: t_{130} = -2.38, P = 0.019), while there was no significant association between *L. camara* presence/absence and *C. spinosa* in the DDF (t_{438} = -0.38, P = 0.703) or TF (t_{163} = -1.09, P = 0.276) (Figure 3a). However, *L. camara* abundance was negatively associated with *C. spinosa* in the DDF only (t_{438} = -2.93, P = 0.004). *Lantana camara* presence/absence was significantly associated with *P. emblica* in the DDF ($t_{438} = -8.09$, P < 0.001), but not in the MDF ($t_{130} = 0.20$, P = 0.846) or TF ($t_{163} = 0.80$, P = 0.427) (Figure 3b). Shorea roxburghii was present only in the DDF, but was absent in the MDF and TF. Lantana camara presence/absence was significantly associated with *G. tiliifolia* in the DDF ($t_{438} = -2.48$, P = 0.014), but not in the MDF ($t_{130} = 1.35$, P = 0.180) or TF ($t_{163} =$ 1.34, P = 0.184). Lantana camara presence/absence was significantly associated with bamboo spp. in the MDF ($t_{130} =$ 2.56, P = 0.012), but not in the DDF ($t_{438} = 1.91$, P = 0.057) or TF ($t_{163} = 1.19$, P = 0.235) (Figure 3c).

Percentage grass cover and occupancy

The PERMANOVA analysis indicated that percentage grass cover differed significantly according to whether a plot was invaded or uninvaded by *L. camara* (P < 0.001, Table 3). In fact, the highest component contributing to the variation was *L. camara* presence/absence (16%), followed by the interaction term between habitat and

Table 2. Results of the SIMPER analysis showing the relative and cumulative contributions of various species that contributed to 90.05% of the average dissimilarity among habitats and between invaded and uninvaded plots in Mudumalai Tiger Reserve, India. The effect of *Lantana camara* on each of these species is shown as coefficients. Elephant browse food species are indicated with an asterisk.

	Average	Average bundance			%			
Species	Invaded	Uninvaded	Average dissimilarity	Contribution	Cumulative	Coefficient		
*Catunaregum spinosa	0.56	0.69	11.1	12.2	12.2	- 0.025		
*Phyllanthus emblica	0.09	0.46	6.66	7.32	19.5	-0.004		
* Shorea roxburghii	0.04	0.35	5.18	5.69	25.2	-0.005		
Cassia fistula	0.20	0.18	4.64	5.09	30.3	-0.005		
*Grewia tiliifolia	0.16	0.25	4.11	4.51	34.8	0.007		
*Kudia calucina	0.11	0.18	3.03	3.33	38.1	-0.001		
* Bambusa arundinacea	0.18	0.07	2.67	2.93	41	-0.001		
Glucosmis pentanhulla	0.14	0.13	2.64	2.95	43.9	0.004		
*Tectona arandis	0.11	0.10	2.01	2.9	46.8	0.001		
Anogeissus latifolia	0.02	0.10	2.0	2.60	49.4	0.001		
*Diospuros montana	0.02	0.20	2.1	2.04	51.8	0.000		
*Dalbaraja latifolia	0.10	0.10	2.19	2.41	54.2	0.004		
*Crawia hirauta	0.10	0.10	2.13	2.54	56.2	0.003		
Srewia nirsuta * Unlistense inorg	0.09	0.07	1.01	1.98	50.2	0.003		
* OL I: :	0.08	0.08	1.5/	1.75	57.9	- 0.004		
	0.04	0.10	1.46	1.61	59.5	0.000		
*Syzygium cumini	0.07	0.07	1.44	1.58	61.1	- 0.003		
*Desmodium pulchellum	0.05	0.09	1.42	1.56	62.6	-0.002		
*Ziziphus oenoplia	0.06	0.03	1.42	1.56	64.2	0.002		
Terminalia crenulata	0.00	0.10	1.3	1.43	65.6	0.002		
* Schleichera oleosa	0.06	0.04	1.24	1.36	67.1	0.005		
*Pterocarpus marsupium	0.02	0.08	1.18	1.3	68.3	0.000		
Dalbergia lanceolaria	0.05	0.05	1.11	1.22	69.51	-0.003		
<i>Opuntia</i> spp.	0.06	0.01	1.01	1.11	70.7	-0.001		
Zingiberaceae	0.11	0.00	0.98	1.07	71.7	-0.004		
Casearia esculenta	0.03	0.09	0.97	1.07	72.8	0.000		
Cordia wallichii	0.04	0.03	0.89	0.98	73.8	0.001		
Canthium dicoccum	0.05	0.03	0.76	0.84	74.6	-0.001		
Cinnamomum malabathrum	0.02	0.06	0.71	0.78	75.4	-0.001		
Coffea robusta	0.03	0.06	0.66	0.73	76.8	-0.001		
Olea glandulifera	0.01	0.07	0.66	0.72	77.6	0.001		
Cassia tora	0.05	0.01	0.64	0.7	78.3	-0.003		
Cipadessa baccifera	0.04	0.03	0.6	0.66	78.9	-0.002		
*Ziziphus xylopyrus	0.04	0.01	0.58	0.64	79.6	-0.001		
* Terminalia tomentosa	0.00	0.06	0.57	0.62	80.2	0.000		
Ougeinia oojeinensis	0.01	0.04	0.56	0.61	80.8	0.002		
Murraya paniculata	0.01	0.05	0.54	0.59	81.4	0.002		
*Albizia lebbeck	0.01	0.03	0.5	0.55	81.9	0.000		
Zizuphus rugosa	0.02	0.01	0.48	0.53	82.5	0.001		
*Solanum torvum	0.03	0.01	0.48	0.53	82.9	0.003		
* Acacia spp.	0.04	0.02	0.48	0.52	83.5	0.003		
Laaerstroemia microcarna	0.03	0.01	0.47	0.52	84	0.000		
Urena lohata	0.03	0.02	0.46	0.51	84 5	0.000		
Argeratum conuzoides	0.03	0.01	0.43	0.47	85	0.001		
Mautenus emarginata	0.03	0.01	0.43	0.47	85.5	0.001		
Flueagea leuconurus	0.03	0.01	0.13	0.47	85.9	-0.001		
* Solanum auriculatum	0.02	0.01	0.43	0.46	86.4	- 0.001		
Viburnum nunctatum	0.03	0.01	0.42	0.40	86.8	0.001		
* Acasia lausophlosa	0.01	0.04	0.4	0.44	80.8	0.000		
Darsaa maarantha	0.02	0.00	0.39	0.43	07.3 87.7	- 0.002		
r er seu mueranuna Tarminalia ababula	0.01	0.04	0.38	0.42	0/./	0.001		
renninana chebula Emitteran lum	0.01	0.03	0.38	0.42	00.1	0.000		
Eryunroxyuum monogynum	0.02	0.01	0.38	0.42	88.5	0.000		
Argyreia cuneata	0.00	0.02	0.36	0.39	88.9	0.000		
Tamilnadia uliginosa	0.01	0.03	0.34	0.37	89.3	0.000		
Actinodaphne malabarica	0.01	0.02	0.32	0.35	89.7	- 0.001		
Sterculia guttata	0.00	0.03	0.32	0.35	90	0.000		
Naringi crenulata	0.01	0.01	0.31	0.34	90.3	- 0.001		



Figure 3. Relative abundance of three plant taxa important for elephant food, in invaded and uninvaded plots in the three habitats. These plants were selected from the SIMPER analysis *Catuneragum spinosa* (a), *Phyllanthus emblica* (b) and bamboo spp. (c) in *Lantana camara* invaded and uninvaded plots in three habitats (moist deciduous forest (MDF); dry deciduous forest (DDF) and thorn forest (TF)) of Mudumalai Tiger Reserve, India. Numbers above the bars refer to the number of individual plants of each species in each habitat.

Table 3. Results of PERMANOVA (permutational analysis of variance) examining the association of *Lantana camara* presence/absence and environmental covariates on percentage grass cover in Mudumalai Tiger Reserve, India. The component of per cent variation (equivalent to the sum of squared fixed effects divided by the degrees of freedom from standard ANOVA) for each predictor is given. All environmental covariates except for tree density were significant predictors of percentage grass cover.

Environmental covariates	df	Pseudo-F	P(perm)	Estimate	Per cent variation
Lantana camara (presence/absence)	1	43.3	< 0.001	70.0	15.6
Habitat × Lantana camara (presence/absence)	2	6.8	0.001	43.2	12.3
Roads	2	5.9	0.002	15.6	7.4
Habitat	2	4.6	0.008	12.0	6.5
Settlements	2	3.9	0.015	10.7	6.1
Canopy cover	1	6.6	0.008	3.9	3.7
Fire	1	5.9	0.012	3.7	3.6
Tree density	1	3.0	0.086	1.5	2.3
Residuals	724			523.0	42.7
Total	736				

Table 4. Results of the linear regression of environmental covariates including *Lantana camara* invasion (average girth per plot) predicting percentage grass occupancy in Mudumalai Tiger Reserve, India. *Lantana camara* invasion was the only significant predictor of percentage grass occupancy. The interaction terms, moist deciduous forest (MDF) and dry deciduous forest (DDF) with *L. camara* are included in the model. The interaction term thorn forest (TF) and *L. camara* was set to zero because of redundancies in the model.

		Coefficien			
Environmental covariates	Unstand	lardized	Standardized		Р
	В	SE	Beta	t	
(Constant)	77.8	4.5		17.3	< 0.001
Lantana camara	-1.8	0.5	-0.2	-3.7	< 0.001
Canopy cover	-0.1	0.0	-0.1	-1.8	0.065
Fire	-0.2	0.8	0.0	-0.2	0.841
Tree density	- 5.9	22.5	0.0	-0.3	0.795
Roads	-0.2	1.9	0.0	-0.1	0.926
Settlements	-2.3	2.1	-0.1	-1.1	0.265
$MDF \times Lantana \ camara$	-0.8	0.6	-0.1	-1.3	0.184
DDF × Lantana camara	- 0.5	0.9	0.0	-0.6	0.559

L. camara presence/absence (12%), and roads (7%). Tree density was not a significant predictor of percentage grass cover (P = 0.086). However, all other environmental covariates were significantly associated with percentage grass cover (P < 0.015, Table 3). Pairwise tests indicated that percentage grass cover significantly differed in the MDF (t_{120} = 3.51, P = 0.003) and DDF (t_{424} = 1.97, P = 0.034) depending on whether *L. camara* was present or absent. However, the presence of *L. camara* made no difference to the percentage grass cover in the TF (t_{153} = 0.80, P = 0.441). Thus it is difficult to generalize on the common effects of *L. camara* across the different habitats.

The linear regression of the *L. camara* invasion (average girth per plot) and environmental covariates on percentage grass occupancy across habitats was statistically significant, although explained only a small amount of variation ($F_{8,736} = 6.7, R^2 = 0.07, P < 0.001$). *Lantana camara* invasion was the only significant predictor of the percentage grass occupancy (P < 0.001, Table 4), possibly indicating competition for the same space. There was a significant negative correlation between percentage

grass occupancy and *L. camara* in all three habitats, indicating that as *L. camara* invasion increased, grass occupancy declined.

DISCUSSION

Our results indicate a significant association between *L. camara* and a change in plant species assemblage, some elephant browse plants, percentage grass cover and occupancy in the moist and dry deciduous habitats of Mudumalai, but not in the thorn forest.

Plant species assemblage

While the three habitats in Mudumalai are clearly different in terms of their plant species assemblage, PERMANOVA pairwise tests of the interaction between habitat and plots with and without *L. camara* indicated that *L. camara* presence/absence made a significant

difference only to the moist deciduous forest (MDF) and dry deciduous forest (DDF) of Mudumalai and not the thorn forest. In the MDF, 43% of the sampled sites had L. camara present while in the DDF, 48% of the sampled sites were invaded by L. camara. The MDF has the highest shrub and sapling density and diversity compared with the DDF and thorn forest (TF) in Mudumalai (Kumar 2011). It is likely that *L. camara* is capable of changing the diversity and density of shrubs and saplings and hence we see an association of L. camara in the MDF because of the higher diversity and density of shrubs and saplings. The MDF is a closed-canopy forest and closed canopy is known to hamper L. camara growth (Duggin & Gentle 1998, Fensham et al. 1994). However, L. camara was recognized as a problem taking over the understorey and spreading rapidly in the Benne and Mudumalai blocks of the MDF and affecting the growth rate of teak in its early stages as early as 1924 in Mudumalai when timber extraction was carried out (Ranganathan 1941). The timber extractions may have opened up the canopy and facilitated L. camara invasion suggesting that L. camara may be the 'passenger' here, but further studies are required to confirm its role here. Nevertheless, L. camara has contributed significantly to a change in the plant species assemblage in the MDF.

Similarly, there was an association between *L. camara* presence and plant species assemblages in the DDF, where timber extraction continued until a ban on logging in the 1980s (Srivastava 2009). Anthropogenic disturbances such as logging may have opened up the canopy which has increased the amount of light penetrating into the forest floor. Opening up of the forest canopy and allowing more light, however, is an advantage to exotic invasive species such as *L. camara* that are known to germinate with an increase in light availability (Gentle & Duggin 1997b, Totland *et al.* 2005). Anthropogenic disturbances have also been known to facilitate exotic plant invasions (Buckley *et al.* 2007, Duggin & Gentle 1998) and may have facilitated *L. camara* invasion here.

In addition to logging, fire has also been regarded as having a major impact on native sapling regeneration in the DDF (Sivaganesan & Sathyanarayana 1995). Fires have been shown to facilitate the spread of L. camara elsewhere (Hiremath & Sundaram 2005). Fires suppress native saplings and facilitate germination and spread of L. camara (Berry et al. 2011, Raizada & Raghubanshi 2010) in the DDF. It is likely that the association of L. camara with plant species assemblage is seen in the DDF because of the impact of logging and fire in the DDF. Grasses can be fuel loads that influence fire frequency and intensity (Scholes & Archer 1997). In the MDF, however, fire has been suggested to have much less impact on native species regeneration because grasses in the MDF retain their moistness even in the dry season, which reduces fire frequency and intensity (Sivaganesan 1991). In the TF, a lack of litter accumulation and cattle grazing

results in reduced fire frequency and intensity (Daniel *et al.* 1995, Sivaganesan 1991). However, when interpreting the response of native species distribution and abundance to infestations of exotic plants, caution must be exercised because infrequent plants may just be rare because of their nature of being rare, or may have been displaced by weed invasions (Butler & Cogan 2004).

In addition to L. camara, the results of our study also show that biotic and abiotic environmental covariates such as tree density, canopy cover, grass cover, impact of roads, settlements and fire are also significantly associated with plant species assemblage. Elsewhere, the association of environmental covariates with plant species assemblage have also been documented indicating the role that biotic and abiotic factors have in the floristic assemblage. For example, Angold (1997) investigated the effect of a road on adjacent heathland vegetation in the UK, and found that there was an increase in the abundance of grasses in the vegetation near the road. In Australia, fire frequency was estimated to account for 60% of the floristic variation (Morrison et al. 1995). In a central Brazilian deciduous dry forest, plant species abundance and distribution was significantly correlated with canopy gaps (Oliveira-Filho et al. 1998). Thus, other environmental covariates are also responsible for changes in the plant community.

Elephant browse plants

Plant species are likely to respond to L. camara invasion differently, depending on different stages of its invasion (Gooden et al. 2009). While some native species are excluded more easily than others from invaded communities, the resistance of native species to invasion varies (Standish et al. 2001). For example, C. spinosa that forms only 0.15% of elephant diet in Mudumalai (Baskaran et al. 2010) was significantly associated with the presence of L. camara only in the MDF, but not in the DDF and TF, while the slope of the relationship between L. camara abundance and C. spinosa was negative only in the DDF and not in the MDF or TF. Further, bamboo spp. did not appear to be associated with L. *camara* presence in the DDF and TF but was significantly associated with L. camara presence in the MDF. In fact, the percentage of bamboo spp. saplings available was greater where there was more L. camara in all three habitats, and no bamboo spp. saplings were found in the TF where L. camara was absent. While this result does not indicate that this species requires L. camara to grow, it does appear to indicate that L. camara is affecting species composition by suppressing some species and facilitating the expansion of others such as bamboo spp. (A. A. Desai, pers. obs.). Such changes in the vegetation composition may have a cascading impact on the ecosystem and

would potentially impact all biodiversity. Further, we hypothesized that greater bamboo spp. sapling numbers occur within *L. camara* areas possibly because herbivores are unable to access these saplings. Other studies have shown that native plant species can benefit from invasive plant species by growing inside stands of the invasive species thereby experiencing lower levels of herbivory (Atwater et al. 2011). This association would allow these saplings to grow but herbivores may be feeding more on certain species where there is less L. camara thereby depleting their food resources in areas without *L. camara*. Although bamboo spp. are often suggested as being important elephant food plants, one estimate indicates that they made up only approximately 4.4% of elephant diet in Mudumalai (Baskaran et al. 2010). Therefore, our results suggest that L. camara presence and abundance, habitat and environmental covariates are associated with the abundance of some elephant food plants, but this association varies depending on the species and in which habitat these species are found.

Percentage grass cover and occupancy

The presence of *L. camara* was observed to have a significant negative association with grass cover in the MDF and DDF. The DDF was reported to have the maximum grass species richness, followed by the TF (Kumar 2011). In addition, the annual net primary productivity of grass was estimated to be highest (720 g m⁻²) in the DDF, 352 g m⁻² in the TF and 110 g m⁻² in the MDF (Baskaran *et al.* 2010). The association of *L. camara* may not be seen in the TF due to the lower grass biomass in this habitat when compared with the MDF and DDF.

In addition, there are other factors that could potentially contribute to the absence of any association of *L. camara* in the TF. For example, cattle grazing has been regarded as one of the causes of the depletion of grass in the thorn forest, and the TF has been considered as sub-optimal habitat for elephant due to low productivity of grass (Daniel *et al.* 1995) allowing *L. camara* to invade these sites (Silori & Mishra 2001) yet not have a significant association with grass cover in the TF.

From our observations within the reserve, the most visible association of *L. camara* on elephant habitat appeared to be the loss of grass cover. Our analysis indicated a significant negative association between percentage grass occupancy and *L. camara*. This result possibly indicates competition for the same space, nutrients or water. A previous study in Mudumalai indicated that in the dry deciduous forest, 85% of elephant diet was grass, while 78% and 53% of elephant diet consisted of grass in the MDF and TF respectively

(Baskaran *et al.* 2010). The reduction in grass cover could lead to food limitation for elephant and other herbivores that depend on grass in the reserve. Reduced grass cover could lead to a reduced carrying capacity of herbivores in the reserve. Any adverse impact on herbivores that are dependent on grass would in turn impact large carnivores such as tigers which are dependent on them (Prasad 2010).

Overall the replacement of grass by L. camara could have serious conservation implications for both herbivores and their predators. Unpalatable weeds such as L. camara may render some areas unsuitable to elephant through reduced forage, limiting food to fewer patches. Such changes in carrying capacity and distribution of food resources of the reserve could also result in elephants being forced to move out in search of better forage. This movement would likely occur with high elephant densities, if food becomes more limiting. Managers in particular need to recognize that reduced carrying capacity through loss of grazing areas can force elephants to move out of the reserve and come into increased conflict with the surrounding human settlement (Ishwaran 1993). It is important that managers take this into account and address this situation. For example, seeds of grass species such as Axonopus sp. that compete well with L. camara could be sowed to help increase forage for grazers (Kumar et al. 2012).

Conclusions

The results of our study indicate that *L. camara* appears capable of altering plant species assemblage, some elephant browse plants and percentage grass cover in the MDF and DDF in addition to other factors. It appears that L. camara invasion is not associated with plant species assemblage, elephant browse plants and grass cover in the TF despite the thorn forest having the highest number of invaded sites. These results suggest that L. *camara* may not be responsible for any changes brought about to the plant community within the TF. This lack of association also suggests that managers may instead focus on L. camara management in the MDF and DDF of Mudumalai where L. camara does have a significant association with the plant community. Nevertheless, as in many invaded systems, there is still uncertainty as to whether L. camara is the 'driver' of community changes or is just a 'passenger' that appears to be less affected by disturbance or environmental stressors and may just be an opportunistic invader (MacDougall & Turkington 2005). Further studies are required to empirically test whether L. camara is the 'driver' of plant community changes or just a 'passenger' that is a consequence of a disturbed habitat.

ACKNOWLEDGEMENTS

The Tamil Nadu Forest Department provided research permits (Ref. No. WL5/57210/2008) to conduct this study and we are particularly thankful to Dr Rajiv K. Srivastava, Field Director, Mudumalai Tiger Reserve and Dr N. Kalaivanan, Veterinary Assistant Surgeon, for facilitating this study. Thanks to the field trackers for their assistance with data collection and to Mr N. Mohanraj for providing maps of the study area. We are also grateful to Dr Stephen Hartley, Dr R. Nagarajan and two anonymous referees for their comments which have improved the manuscript. This project was funded by Rufford Small Grants, UK, United States Fish and Wildlife Services (96200–9-G171, Grant No. ASE-0435) and Mohammed Bin Zayed Species Conservation Fund (Project number: 1025959).

LITERATURE CITED

- ANGOLD, P. G. 1997. The impact of a road upon adjacent heathland vegetation: effects on plant species composition. *Journal of Applied Ecology* 34:409–417.
- ATWATER, D. Z., BAUER, C. M. & CALLAWAY, R. M. 2011. Indirect positive effects ameliorate strong negative effects of *Euphorbia esula* on a native plant. *Plant Ecology* 212:1655–1662.
- BASKARAN, N., BALASUBRAMANIAN, M., SWAMINATHAN, S. & DESAI, A. A. 2010. Feeding ecology of the Asian elephant *Elephas maximus* Linnaeus in the Nilgiri Biosphere Reserve, southern India. *Journal of the Bombay Natural History Society* 107:3–13.
- BEDUNAH, D. J. 1992. The complex ecology of weeds, grazing and wildlife. Western Wildlands 18:6–11.
- BERRY, Z. C., WEVILL, K. & CURRAN, T. J. 2011. The invasive weed Lantana camara increases fire risk in dry rainforest by altering fuel beds. Weed Research 51:525–533.
- BRAITHWAITE, R. W., LONSDALE, W. M. & ESTBERGS, J. A. 1989. Alien vegetation and native biota in tropical Australia: the impact of *Mimosa pigra*. *Biological Conservation* 48:189–210.
- BUCKLEY, Y. M., BOLKER, B. M. & REES, M. 2007. Disturbance, invasion and re-invasion: managing the weed-shaped hole in disturbed ecosystems. *Ecology Letters* 10:809–817.
- BUTLER, J. L. & COGAN, D. R. 2004. Leafy spurge effects on patterns of plant species richness. *Rangeland Ecology & Management* 57:305– 311.
- CLARKE, K. R. & GORLEY, R. N. 2006. PRIMER v6: user manual and tutorial. PRIMER-E, Plymouth. 190 pp.
- CLARKE, K. R. & WARWICK, R. M. 2001. Change in marine communities: an approach to statistical analysis and interpretation. PRIMER-E, Plymouth. 176 pp.
- DANIEL, J. C., DESAI, A. A., SIVAGANESAN, N., DATYE, H. S. & KUMAR, S. R. 1995. Ecology of the Asian elephant. Bombay Natural History Society, Bombay. 90 pp.
- DUGGIN, J. A. & GENTLE, C. B. 1998. Experimental evidence on the importance of disturbance intensity for invasion of *Lantana camara*

L. in dry rainforest open forest ecotones in north-eastern NSW, Australia. *Forest Ecology and Management* 109:279–292.

- EHRENFELD, J. G., KOURTEV, P. & HUANG, W. 2001. Changes in soil functions following invasions of exotic understory plants in deciduous forests. *Ecological Applications* 11:1287–1300.
- FENSHAM, R. J., FAIRFAX, R. J. & CANNELL, R. J. 1994. The invasion of *Lantana camara* L. in Forty Mile Scrub National Park, north Queensland. *Australian Journal of Ecology* 19:297–305.
- GENTLE, C. B. & DUGGIN, J. A. 1997a. Allelopathy as a competitive strategy in persistent thickets of *Lantana camara* L. in three Australian forest communities. *Plant Ecology* 132:85–95.
- GENTLE, C. B. & DUGGIN, J. A. 1997b. *Lantana camara* L. invasions in dry rainforest open forest ecotones: the role of disturbances associated with fire and cattle grazing. *Australian Journal of Ecology* 22:298–306.
- GODEFROID, S. & KOEDAM, N. 2004. The impact of forest paths upon adjacent vegetation: effects of the path surfacing material on the species composition and soil compaction. *Biological Conservation* 119:405–419.
- GOODEN, B., FRENCH, K., TURNER, P. J. & DOWNEY, P. O. 2009. Impact threshold for an alien plant invader, *Lantana camara* L., on native plant communities. *Biological Conservation* 142:2631–2641.
- HIREMATH, A. J. & SUNDARAM, B. 2005. The fire–Lantana cycle hypothesis in Indian forests. *Conservation and Society* 3:26–42.
- ISHWARAN, N. 1993. Ecology of the Asian elephant in lowland dry zone habitats of the Mahaweli River Basin, Sri Lanka. *Journal of Tropical Ecology* 9:169–182.
- KUMAR, M. A. 2011. Population, foraging and activity pattern of gaur (Bos gaurus H. Smith, 1827) in Mudumalai Tiger Reserve, southern India. Ph.D. thesis, Bharathidasan University, Tiruchirappally. 382 pp.
- KUMAR, M. A., NAGARAJAN, R., ILAYARAJA, R., SWAMINATHAN, S. & DESAI, A. A. 2012. Impact of plant weeds on grass availability in Gaur (*Bos gaurus* H. Smith, 1827) foraging areas of Mudumalai Tiger Reserve, southern India. *Indian Forester* 138:1131–1140.
- LYM, R. G. & KIRBY, D. R. 1987. Cattle foraging behavior in leafy spurge (*Euphorbia esula*)-infested rangeland. *Weed Technology* 1:314–318.
- MACDOUGALL, A. S. & TURKINGTON, R. 2005. Are invasive species the drivers or passengers of change in degraded ecosystems? *Ecology* 86:42–55.
- MELGOZA, G., NOWAK, R. S. & TAUSCH, R. J. 1990. Soil water exploitation after fire: competition between *Bromus tectorum* (cheatgrass) and two native species. *Oecologia* 83:7–13.
- MORRISON, D. A., GARY, G. J., PENGELLY, S. M., ROSS, D. G., MULLINS, B. J., THOMAS, C. R. & ANDERSON, T. S. 1995. Effects of fire frequency on plant species composition of sandstone communities in the Sydney region: inter-fire interval and time-since-fire. *Australian Journal of Ecology* 20:239–247.
- OLIVEIRA-FILHO, A. T., CURI, N., VILELA, E. A. & CARVALHO, D. A. 1998. Effects of canopy gaps, topography, and soils on the distribution of woody species in a Central Brazilian deciduous dry forest. *Biotropica* 30:362–375.
- OWEN-SMITH, R. N. 1988. *Megaherbivores, the influence of very large body size on ecology*. Cambridge University Press, Cambridge. 369 pp.
- PRASAD, A. E. 2010. Effects of an exotic plant invasion on native understory plants in a tropical dry forest. *Conservation Biology* 24:747–757.

- PRASAD, A. E. 2012. Landscape-scale relationships between the exotic invasive shrub *Lantana camara* and native plants in a tropical deciduous forest in southern India. *Journal of Tropical Ecology* 28:55– 64.
- RAIZADA, P. & RAGHUBANSHI, A. S. 2010. Seed germination behaviour of *Lantana camara* in response to smoke. *Tropical Ecology* 51:347–352.
- RANGANATHAN, C. R. 1941. Working plan for the Nilgiris Forest Division. Government Press, Madras. 117 pp.
- SCHOLES, R. J. & ARCHER, S. R. 1997. Tree-grass interactions in savannas. Annual Review of Ecology and Systematics 28:517–544.
- SHARMA, G. P. & RAGHUBANSHI, A. S. 2007. Effect of Lantana camara L. cover on local depletion of tree population in the Vindhyan tropical dry deciduous forest of India. Applied Ecology and Environmental Research 5:109–121.
- SILORI, C. S. & MISHRA, B. K. 2001. Assessment of livestock grazing pressure in and around the elephant corridors in Mudumalai Wildlife Sanctuary, South India. *Biodiversity and Conservation* 10:2181– 2195.
- SINCLAIR, A. R. E. 1975. The resource limitation of trophic levels in tropical grassland ecosystems. *Journal of Animal Ecology* 44:497–520.
- SIVAGANESAN, N. 1991. Ecology and conservation of Asian elephants (Elephas maximus) with special reference to the habitat utilization in Mudumalai Wildlife Sanctuary, Tamil Nadu. South India. Ph.D. thesis, Bharathidasan University, Tiruchirappally. 140 pp.
- SIVAGANESAN, N. & SATHYANARAYANA, M. C. 1995. Tree mortality caused by elephants in Mudumalai Wildlife Sanctuary, South India. Pp. 314–330 in Daniel, J. C. & Datye, H. S. (eds.). A week with elephants. Bombay Natural History Society and Oxford University Press, Bombay.
- SRIVASTAVA, R. K. 2009. *Mudumalai Tiger Reserve management plan*. Tamil Nadu Forest Department, Ootacamund. 408 pp.
- STANDISH, R. J., ROBERTSON, A. W. & WILLIAMS, P. A. 2001. The impact of an invasive weed *Tradescantia fluminensis* on native forest regeneration. *Journal of Applied Ecology* 38:1253–1263.
- SWARBRICK, J. T., WILLSON, B. B. & HANNAN-JONES, M. A. 1998. Lantana camara L. Pp. 119–140 in Panetta, F. D., Groves, R. H. & Shepherd, R. C. H. (eds.). The biology of Australian weeds (2). R.G. & F.J. Richardson, Melbourne.
- TERBORGH, J., LOPEZ, L., PERCY NUÑEZ, V., RAO, M., SHAHABUDDIN, G., ORIHUELA, G., RIVEROS, M., ASCANIO, R., ADLER, G. H., LAMBERT, T. D. & BALBAS, L. 2001. Ecological meltdown in predator-free forest fragments. *Science* 294:1923–1926.
- TOTLAND, Ø., NYEKO, P., BJERKNES, A.-L., HEGLAND, S. J. & NIELSEN, A. 2005. Does forest gap size affect population size, plant size, reproductive success and pollinator visitation in *Lantana camara*, a tropical invasive shrub? *Forest Ecology and Management* 215:329– 338.
- WILSON, G., DESAI, A. A., SIM, D. A. & LINKLATER, W. L. 2013. The influence of the invasive weed *Lantana camara* on elephant habitat use in Mudumalai Tiger Reserve, southern India. *Journal of Tropical Ecology* 29:199–207.
- WILSON, G., GRUBER, M. A. M. & LESTER, P. J. 2014. Foraging relationships between elephants and *Lantana camara* invasion in Mudumalai Tiger Reserve, India. *Biotropica* 46:194–201.

- WOODS, K. D. 1993. Effects of invasion by *Lonicera tatarica* L. on herbs and tree seedlings in four New England Forests. *American Midland Naturalist* 130:62–74.
 - **Appendix 1.** The list of plant species (shrubs and saplings measuring between 10 and 150-cm in height) identified in the plots along the 67 1-km in length transects in Mudumalai Tiger Reserve, India. Number refers to the total number of individual stems of the species counted throughout the reserve.

Plant species	Number
Acacia leucophloea (Roxb.) Willd.	13
Acacia spp.	22
Achyranthes aspera L.	3
Actinodaphne malabarica Balkr.	12
Albizia lebbeck (L.) Benth.	16
Allophylus cobbe (L.) Raeusch.	5
Anisochilus scaber Benth.	7
Anogeissus latifolia (DC.) Wallich ex Guill. & Perr.	62
Antidesma menasu (Tul.) Miq. ex MuellArg.	7
Ardisia solanacea Roxb.	8
Argeratum conyzoides L.	9
Argyreia cuneata (Willd.) Ker Gawler	10
Azadirachta indica Adr. Juss.	1
Bambusa arundinacea (Retz.) Willd.	94
Bauhinia racemosa Lam.	9
Bidens pilosa L.	6
Bischofia javanica Blume	7
Bridelia crenulata Roxb.	8
Bridelia retusa (L.) Spreng.	9
Butea monosperma (Lam.) Taubert	4
Callicarpa tomentosa (L.) Murr.	3
Canthium dicoccum (Gaertner) Teijsm. & Binnend.	30
Canthium parviflora Roxb.	1
Capparis zeylanica L.	1
Careya arborea Roxb.	7
Carissa carandas L.	2
Casearia esculenta Roxb.	42
Cassia fistula L.	132
Cassia sophera L.	4
Cassia tora L.	17
Catunaregum spinosa (Thunb.) Tirveng.	340
Celosia argentea L.	4
Celtis tetrandra Roxb.	1
Chloroxylon swietenia DC.	5
Cinnamomum malabathrum (Lam.) J.Presl	23
Cinnamomum spp.	3
Cipadessa baccifera (Roth) Miq.	25
Coffea robusta L.	23
Cordia wallichii Don.	33
Dalbergia lanceolaria L.f.	42
Dalbergia latifolia Roxb.	69
Desmodium pulchellum (L.) Benth.	43
Dichrostachys cinerea (L.) Wight & Arn.	8
Diospyros montana Roxb.	83
Elaeodendron glaucum (Rottb.) Pers.	1
Eriolaena nookeriana Wight & Arn.	5
Eriolaena quinquelocularis Wright	3
Eryunrina inaica Lam.	2
Eryunroxyium monogynum Koxb.	11
Eupnordia nirta L.	7

Appendix 1. Continued.

Plant species	Number
Ficus religiosa L.	2
Flacourtia indica (Burm.f.) Merr.	3
Flueggea leucopyrus Willd.	13
Gardenia gummifera L.f.	2
Garuga pinnata Roxb.	6
Givotia rottleriformis Griffith	1
Glochidion zeylanicum (Gaertn.) A.Juss.	5
Glycosmis pentaphylla (Retz.) DC.	67
Gmelina arborea Roxb.	8
Grewia hirsuta Vahl	58
Grewia orbiculata Wall.	2
Grewia tiliifolia Vahl	156
Haldina cordifolia (Roxb.) Ridsd.	3
Hardwickia binata Roxb.	1
Helicteres isora L.	45
Ipomoea hederifolia L.	6
Kydia calycina Roxb.	88
Lagerstroemia microcarpa Wight	21
Lagerstroemia parviflora Roxb.	5
Lannea coromandelica (Houtt.) Merr.	4
Lawsonia inermis L.	1
Litsea floribunda Gamble	3
Litsea mysorensis Gamble	10
Madhuca indica J. Gmelin	2
Maesa indica (Roxb.) A.DC.	4
Mallotus philippensis (Lam.) Muell. Arg.	6
Mangifera indica L.	4
Maytenus emarginata (Willd.) Ding Hou	13
Melia dubia Cav.	4
Mitragyna parviflora (Roxb.) Korth.	2
Murraya paniculata (L.) Jack	19
Naringi crenulata (Roxb.) Nicolson	12
Nothapodytes foetida (J. Graham) Mabb.	2
Ocimum tenuiflorum L.	1
Olea dioica Roxb.	45
Olea glandulifera Desf.	19
Opuntia spp.	30
Ougeinia oojeinensis (Roxb.) H. Ohashi	17
Pavetta indica L.	5
Persea macrantha (Nees) Kosterm.	16
Phoenix loureirii Kunth	7

Appendix 1. Continued.

Phyllanthus emblica L.177Pongamia pinnata (L.) Pierre3Premna tomentosa Willd.1Pterocarpus marsupium Roxb.36Radermachera xylocarpa (Roxb.) Schumann8Schleichera oleosa (Lour.) Oken50Schrebera swietenoides Roxb.7Scolopia crenata Clos12Shorea roxburghii Don81Solanum auriculatum14Solanum torvum Sw.18Sterculia gutata Roxb.8Sterculia urens Roxb.3Stercospermum personatum (Hassk.) Chatterjee4Sterospermum tetragonum DC.5Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Terminalia crenulata Roth27Terminalia crenulata Roth27Terminalia crenulata Roth21Viburum punctatum BuchHam.ex D.Don14Viburum punctatum BuchHam.ex D.Don14Viburum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Ziziphus rugosa Lam.2Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30 </th <th>Plant species</th> <th>Number</th>	Plant species	Number
Pongamia pinnata (L.) Pierre3Premna tomentosa Willd.1Pterocarpus marsupium Roxb.36Radermachera xylocarpa (Roxb.) Schumann8Schleichera oleosa (Lour.) Oken50Schrebera swietenoides Roxb.7Scolopia crenata Clos12Shorea roxburghii Don81Solanum auriculatum14Solanum torvum Sw.18Sterculia guttata Roxb.8Sterculia guttata Roxb.3Stereoulia guttata Roxb.3Stereoulia urens Roxb.3Stereospermum personatum (Hassk.) Chatterjee4Stychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamihadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia chebula Retz.10Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Widyntum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Ziziphus mauritiana Lam.2Ziziphus mauritiana Lam.2Ziziphus nauritiana Lam.2Ziziphus nauritiana Lam.2Ziziphus vylopyrus (Retz.) Willd.25Ziziphus vylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) W	Phyllanthus emblica L.	177
Premna tomentosa Willd.1Pterocarpus marsupium Roxb.36Radermachera xylocarpa (Roxb.) Schumann8Schleichera oleosa (Lour.) Oken50Schrebera swietenoides Roxb.7Scolopia crenata Clos12Shorea roxburghii Don81Solanum auriculatum14Solanum torvum Sw.18Sterculia guttata Roxb.8Sterculia guttata Roxb.3Stercoulia guttata Roxb.3Stercoulia urens Roxb.3Stercospermum personatum (Hassk.) Chatterjee4Syzygium cumini (L.) Skeels50Tamaindus indica L.2Taminadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia chebula Retz.10Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Widynum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Ziziphus mauritiana Lam.2Ziziphus mauritiana Lam.2Ziziphus mauritiana Lam.2Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyru	Pongamia pinnata (L.) Pierre	3
Pterocarpus marsupium Roxb.36Radermachera xylocarpa (Roxb.) Schumann8Schleichera oleosa (Lour.) Oken50Schrebera swietenoides Roxb.7Scolopia crenata Clos12Shorea roxburghii Don81Solanum auriculatum14Solanum torvum Sw.18Sterculia guttata Roxb.8Sterculia guttata Roxb.3Stereospermum personatum (Hassk.) Chatterjee4Stereospermum personatum (Hassk.) Chatterjee4Stychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamaindus indica L.2Tamihadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia chebula Roth27Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Wiek altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Ziziphus galabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus mauritiana Lam.2Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30	Premna tomentosa Willd.	1
Radermachera xylocarpa (Roxb.) Schumann8Schleichera oleosa (Lour.) Oken50Schrebera swietenoides Roxb.7Scolopia crenata Clos12Shorea roxburghii Don81Solanum auriculatum14Solanum torvum Sw.18Sterculia guttata Roxb.8Sterculia guttata Roxb.3Stereospermum personatum (Hassk.) Chatterjee4Stereospermum personatum (Hassk.) Chatterjee4Stereospermum tetragonum DC.5Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamaindus indica L.2Tamihadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Wiendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Ziziphus mauritiana Lam.2Ziziphus mauritiana Lam.2Ziziphus mauritiana Lam.2Ziziphus nugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30	Pterocarpus marsupium Roxb.	36
Schleichera oleosa (Lour.) Oken50Schrebera swietenoides Roxb.7Scolopia crenata Clos12Shorea roxburghii Don81Solanum auriculatum14Solanum torvum Sw.18Sterculia guttata Roxb.8Sterculia guttata Roxb.3Stereospermum personatum (Hassk.) Chatterjee4Stereospermum personatum (Hassk.) Chatterjee4Stereospermum tetragonum DC.5Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth2Terminalia aneuclata Roth2Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Wiendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrighta tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus galabrata Heyne ex Roth1Ziziphus nauritiana Lam.2Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (L.) Miller42Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30	Radermachera xylocarpa (Roxb.) Schumann	8
Schrebera swietenoides Roxb.7Scolopia crenata Clos12Shorea roxburghii Don81Solanum auriculatum14Solanum torvum Sw.18Sterculia guttata Roxb.8Sterculia urens Roxb.3Stereospermum personatum (Hassk.) Chatterjee4Stereospermum tetragonum DC.5Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrighta tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus galabrata Heyne ex Roth1Ziziphus rugosa Lam.9Ziziphus rugosa Lam.9Ziziphus rugosa Lam.25Ziziphus rugosa Lam.18	Schleichera oleosa (Lour.) Oken	50
Scolopia crenata Clos12Shorea roxburghii Don81Solanum auriculatum14Solanum torvum Sw.18Sterculia guttata Roxb.8Sterculia guttata Roxb.3Stereospermum personatum (Hassk.) Chatterjee4Stereospermum tetragonum DC.5Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrighta tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus galabrat Heyne ex Roth1Ziziphus nauritiana Lam.2Ziziphus nauritiana Lam.2Ziziphus vylopyrus (Retz.) Willd.25Ziziphus vylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (L.) Miller30Ziziphus xylopyrus (L.) Willd.30	Schrebera swietenoides Roxb.	7
Shorea roxburghii Don81Solanum auriculatum14Solanum torvum Sw.18Sterculia guttata Roxb.8Sterculia guttata Roxb.3Stereospermum personatum (Hassk.) Chatterjee4Stereospermum tetragonum DC.5Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus galabrata Heyne ex Roth1Ziziphus nauritiana Lam.2Ziziphus vylopyrus (Retz.) Willd.25Ziziphus vylopyrus (Retz.) Willd.30Ziziphus vylopyrus (L.) Miller42Ziziphus vylopyrus (L.) Willd.30Ziziphus vylopyrus (L.) Willd.30	Scolopia crenata Clos	12
Solanum auriculatum14Solanum torvum Sw.18Sterculia guttata Roxb.8Sterculia urens Roxb.3Stereospermum personatum (Hassk.) Chatterjee4Stereospermum tetragonum DC.5Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Shorea roxburghii Don	81
Solanum torvum Sw.18Sterculia guttata Roxb.8Sterculia urens Roxb.3Stereospermum personatum (Hassk.) Chatterjee4Stereospermum tetragonum DC.5Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrighta tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus galabrat Heyne ex Roth1Ziziphus rugosa Lam.9Ziziphus rugosa Lam.9Ziziphus rugosa Lam.25Ziziphus rugosa Lam.18	Solanum auriculatum	14
Sterculia guttata Roxb.8Sterculia urens Roxb.3Stereospermum personatum (Hassk.) Chatterjee4Stereospermum tetragonum DC.5Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrighta tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus galabrata Heyne ex Roth1Ziziphus nauritiana Lam.2Ziziphus rugosa Lam.9Ziziphus rugosa Lam.25Ziziphus rugosa Lam.18	Solanum torvum Sw.	18
Sterculia urens Roxb.3Stercospermum personatum (Hassk.) Chatterjee4Stereospermum tetragonum DC.5Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia comentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrighta tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Sterculia guttata Roxb.	8
Stereospermum personatum (Hassk.) Chatterjee4Stereospermum tetragonum DC.5Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia comulata Roth22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrighta tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Sterculia urens Roxb.	3
Stereospermum tetragonum DC.5Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia comulata Roth22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Stereospermum personatum (Hassk.) Chatterjee	4
Strychnos potatorum L.f.4Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia comulata Roth2Terminalia comulata Roth22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Stereospermum tetragonum DC.	5
Syzygium cumini (L.) Skeels50Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia comulata Roth2Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Strychnos potatorum L.f.	4
Tamarindus indica L.2Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tactona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia crenulata Roth2Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Syzygium cumini (L.) Skeels	50
Tamilnadia uliginosa (Retz.) Tirveng. & Sastre12Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia comentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Tamarindus indica L.	2
Tectona grandis L.f.80Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia crenulata Roth2Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.25Ziziphus rugosa Lam.30Ziziphus rugosa Lam.18	Tamilnadia uliginosa (Retz.) Tirveng. & Sastre	12
Tephrosia purpurea (L.) Pers.6Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia paniculata Roth2Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus galabrata Heyne ex Roth1Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugasa Lam.18	Tectona grandis L.f.	80
Terminalia chebula Retz.10Terminalia crenulata Roth27Terminalia paniculata Roth2Terminalia paniculata Roth2Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.25Ziziphus rugosa Lam.18	Tephrosia purpurea (L.) Pers.	6
Terminalia crenulata Roth27Terminalia paniculata Roth2Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus rugosa Lam.9Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Terminalia chebula Retz.	10
Terminalia paniculata Roth2Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Terminalia crenulata Roth	27
Terminalia tomentosa Wight & Arn.22Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Terminalia paniculata Roth	2
Toona ciliata Roemer4Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus (L.) Miller18	Terminalia tomentosa Wight & Arn.	22
Urena lobata L. ssp. sinuata (L.) Borssum.11Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus rugosa Lam.9Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus xylopyrus Lam.18	Toona ciliata Roemer	4
Viburnum punctatum BuchHam.ex D.Don14Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus rugosa Lam.9Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Urena lobata L. ssp. sinuata (L.) Borssum.	11
Vitex altissima L.f.4Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus rugosa Lam.9Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Viburnum punctatum Buch.–Ham.ex D.Don	14
Wendlandia thyrsoidea (Roem. & Schult.) Steud.4Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus rugosa Lam.9Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Vitex altissima L.f.	4
Wrightia tinctoria (Roxb.) R.Br.7Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus rugosa Lam.9Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Wendlandia thyrsoidea (Roem. & Schult.) Steud.	4
Xanthium indicum J. Koenig2Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus rugosa Lam.9Ziziphus vylopyrus (Retz.) Willd.25Ziziphus xylopyrus (L.) Miller42Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Wrightia tinctoria (Roxb.) R.Br.	7
Zingiberaceae20Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus rugosa Lam.9Ziziphus vylopyrus (Retz.) Willd.25Ziziphus vylopyrus (L.) Miller42Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Xanthium indicum J. Koenig	2
Ziziphus glabrata Heyne ex Roth1Ziziphus mauritiana Lam.2Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.25Ziziphus oenoplia (L.) Miller42Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Zingiberaceae	20
Ziziphus mauritiana Lam.2Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.25Ziziphus oenoplia (L.) Miller42Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Ziziphus glabrata Heyne ex Roth	1
Ziziphus rugosa Lam.9Ziziphus xylopyrus (Retz.) Willd.25Ziziphus oenoplia (L.) Miller42Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Ziziphus mauritiana Lam.	2
Ziziphus xylopyrus (Retz.) Willd.25Ziziphus oenoplia (L.) Miller42Ziziphus xylopyrus (Retz.) Willd.30Ziziphus ruaosa Lam.18	Ziziphus rugosa Lam.	9
Ziziphus oenoplia (L.) Miller42Ziziphus xylopyrus (Retz.) Willd.30Ziziphus rugosa Lam.18	Ziziphus xylopyrus (Retz.) Willd.	25
Ziziphus xylopyrus (Retz.) Willd. 30 Ziziphus rugosa Lam. 18	Ziziphus oenoplia (L.) Miller	42
Ziziphus rugosa Lam. 18	Ziziphus xylopyrus (Retz.) Willd.	30
	Ziziphus rugosa Lam.	18