

Breast cancer gene discovery

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Many important advances have been made in the past decade in understanding breast cancer at the molecular level, and two important high-penetrance breast cancer genes – *BRCA1* and *BRCA2* – have been identified. However, germline mutations in these two genes are responsible for only a minority (~5%) of all breast carcinomas, and the genes responsible for the majority of breast cancer cases remain to be identified. There is evidence that there are additional high-to-moderate-penetrance breast cancer susceptibility genes but, given the high degree of molecular heterogeneity in breast carcinomas, it is likely that each of these genes is responsible for only a subset of cases. There are also many candidate low-penetrance breast cancer genes and many more are likely to be identified. In addition to germline, and somatic, sequence alterations, epigenetic changes in many genes are likely to play an important role in the pathobiology of breast cancer. Recently developed genomic technologies and the completion of the human genome sequence provide us with powerful tools to identify novel candidate breast cancer genes that could play an important role in breast tumourigenesis.

What is a breast cancer gene? In a strict definition, a breast cancer gene is a gene in which germline mutations or polymorphisms confer increased susceptibility to breast cancer. In a broader sense, breast cancer genes are genes that play a causative role in the pathogenesis of breast cancer even if they do not have any germline or somatic sequence alterations. This latter group includes genes that are aberrantly expressed in breast tumours due to epigenetic changes (such as methylation), and genes that may be abnormally activated/inactivated at the protein level. Some of these genes are specific for breast cancer, whereas others are involved in other cancer types as well.

Breast cancer is one of the most common cancers diagnosed in women worldwide and is a leading cause of cancer-related death (Ref. 1).

Therefore, the identification of breast cancer genes is a major scientific and social problem. The identification of such genes will not only enable the identification of individuals at high risk, but also aid in the design of more-effective therapies for breast cancer prevention and treatment. Pathological stages in breast tumourigenesis, along with an indication of underlying genetic alterations, are shown in Figure 1. Similar to the development of other cancer types, breast tumourigenesis is a multistep process; it starts with ductal hyperproliferation and progresses into in situ, then invasive, and finally metastatic carcinomas.

Breast cancer susceptibility is genetically determined, but the penetrance (i.e. the frequency of individuals with a specific genotype who manifest the trait in the phenotype) of known

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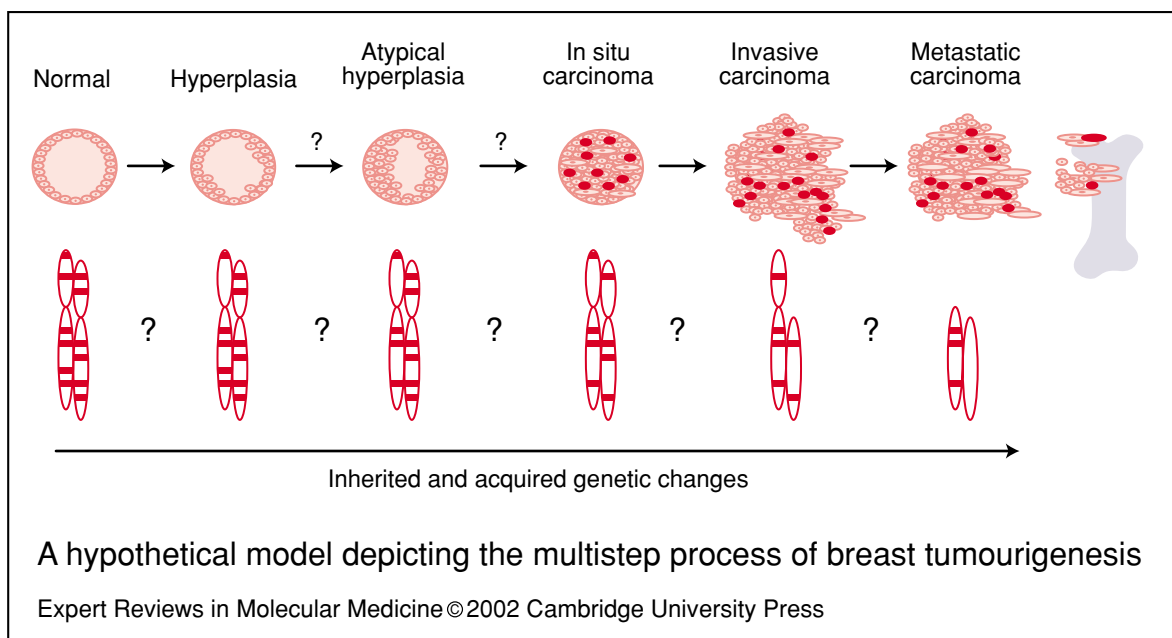


Figure 1. A hypothetical model depicting the multistep process of breast tumourigenesis. Clinical and pathological stages are indicated. The relationships among the earliest-stage tumours (hyperplasia, atypical hyperplasia, and in situ carcinoma) are not well understood and it is questionable if all these lesions represent obligate precursors of invasive carcinomas. Tumour progression is driven by inherited and acquired genetic changes, which are reflected in altered profiles of gene expression and of protein levels and modifications (fig001kpb).

genes that predispose to breast cancer varies widely (Ref. 2). A minority of breast cancer cases are attributable to inherited mutations in rare high-penetrance breast cancer susceptibility genes such as *BRCA1* and *BRCA2* (i.e. individuals with mutations in these genes frequently develop breast cancer), but the majority of cases are probably due to multiple low-penetrance genes (Refs 2, 3, 4). This assumption is based on multiple studies suggesting that most of the excess familial risk of breast cancer is genetic, yet known genes explain fewer than 25% of this excess risk, indicating that other breast cancer genes remain to be identified (Refs 5, 6, 7, 8). Analyses of genetic models to explain familial breast cancer risk that is not due to *BRCA1* and *BRCA2* suggest that the most likely scenario is a polygenic model with many susceptibility alleles, although the existence of an autosomal recessive mendelian allele cannot be excluded (Refs 9, 10). It is the interaction between these genes and environmental factors (e.g. geographical location, smoking, fruit/vegetable consumption) that determines the individual's overall breast cancer risk. In this sense, breast cancer, similar to diabetes, asthma or heart disease, is a complex genetic disease (Ref. 11).

Family linkage studies are useful for the identification of high-penetrance, and perhaps even moderate-penetrance, breast cancer genes, whereas the identification of low-penetrance genes requires a combination of approaches including candidate-gene-based association studies in affected and non-affected populations, analysis of somatic mutations in tumours, and the use of novel, comprehensive and unbiased genomic techniques (Fig. 2) (Ref. 11). This article reviews the strategies and techniques that are available for the identification of additional breast cancer genes.

Linkage studies: high-penetrance genes

Families with multiple cases of cancer, frequently arising at an early age, have been useful for the identification of many cancer predisposition genes. Although inherited mutations in such genes are rare, these genes are frequently mutated in sporadic tumours as well. Breast cancer is particularly well suited for these types of studies, since one of the strongest risk factors for breast cancer is the occurrence of the disease in first-degree relatives, particularly if they are

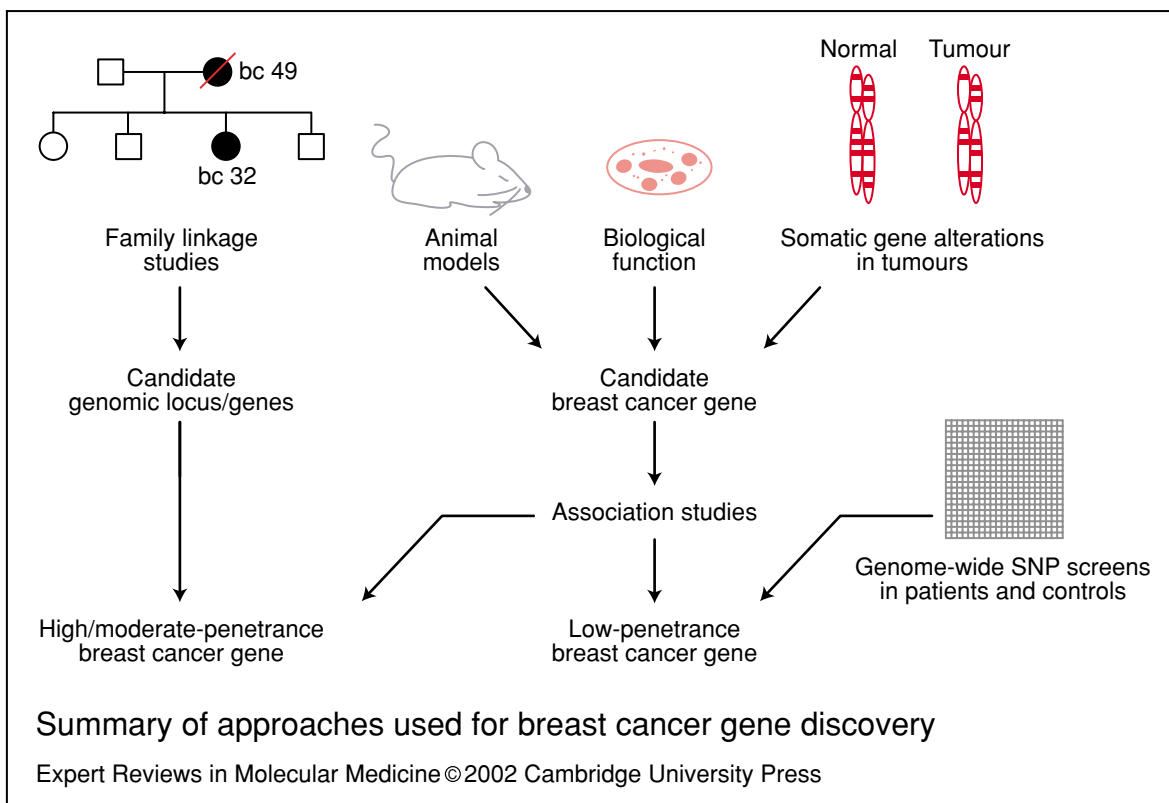


Figure 2. Summary of approaches used for breast cancer gene discovery. Family linkage studies are useful only for the identification of high-to-moderate-penetrance breast cancer genes. Association studies using unbiased genome-wide screens, or candidate genes selected on the basis of biological function, somatic gene alterations (genetic, epigenetic and gene expression changes) or animal models, usually identify low-penetrance breast cancer genes. bc 49, breast cancer arising at age 49 years; SNP, single-nucleotide polymorphism (**fig002kpb**).

diagnosed at an early age. The identification of a susceptibility locus on chromosome 17 was, therefore, of great importance, not only because it proved that early-onset breast cancer is inherited but also because it demonstrated that a high-penetrance breast cancer susceptibility gene exists (Ref. 12). The hunt for *BRCA1*, and subsequently *BRCA2*, was a 'brute-force' linkage mapping approach in affected families and was one of the most competitive races in the history of gene cloning (Refs 13, 14); *BRCA1* was found on chromosome 17q21 and *BRCA2* on 13q12.3. However, in contrast to most cancer predisposition genes identified through family studies, neither of these two genes is mutated in sporadic breast carcinomas (Refs 15, 16).

Although there are some indications that somatic epigenetic inactivation of *BRCA1* and *BRCA2* occurs in some sporadic tumours, overwhelming evidence suggests the occurrence of differing tumourigenic pathways for inherited

and sporadic breast carcinomas (Refs 17, 18). Somewhat disappointingly, follow-up large-scale mutation screening studies determined that not only are *BRCA1* and *BRCA2* not mutated in sporadic tumours, but mutations in these two genes are responsible for only a fraction of inherited cases (Ref. 2). Since family history still remains a strong predictive factor in patients with no detectable mutations in these genes, additional high-to-moderate breast cancer susceptibility genes remain to be identified (Refs 5, 6, 7, 19, 20). Germline mutations in several other genes (Table 1), including *TP53* ('tumour protein p53'), *PTEN* ('phosphatase and tensin homologue'), *LKB1*, *CHK2* ('checkpoint homologue 2') and *MLH1/MSH2* ('MutL homologue 1/MutS homologue 2'), also predispose individuals to breast cancer, but these cases are very rare and are usually part of a multicancer and/or developmental abnormality syndrome (Ref. 21). Since the discovery of *BRCA1*

Table 1. Genes with germline mutations associated with breast cancer susceptibility (tab001kpb)

Gene name	Mode of discovery	Putative function	Refs
<i>BRCA1</i>	Linkage in families	DNA-damage checkpoint control, recombination, transcription	12, 13
<i>BRCA2</i>	Linkage in families	DNA-damage checkpoint control, recombination, transcription	14
<i>BRCAx-Chr13q21</i>	CGH plus linkage in families	?	24, 28, 29
<i>BRCAx-Chr8p</i>	Linkage in families	?	22
<i>TP53</i>	Candidate-gene testing in families	DNA-damage checkpoint control	92
<i>LKB1</i>	CGH plus linkage in families	Apoptosis/growth arrest?	30, 93, 94, 95
<i>PTEN</i>	RDA plus candidate-gene testing in families	Negative regulation of mitogenic/survival signals	51, 53
<i>MLH1/MSH2</i>	Linkage in families	Mismatch repair	96, 97
<i>ATM</i>	Linkage in families	DNA-damage checkpoint control	98
<i>CHK2</i>	Candidate-gene testing in families	DNA-damage checkpoint control	99
<i>HRAS1-VNTR</i>	Candidate-gene testing	Proto-oncogene: altered transcription/linkage disequilibrium/genomic instability	100
<i>CYP1A1</i>	Candidate-gene testing	Metabolism of oestrogen and polycyclic aromatic hydrocarbons	101
<i>CYP2D6</i>	Candidate-gene testing	Metabolism of steroid hormones	102, 103
<i>CYP17</i>	Candidate-gene testing	Metabolism of steroid hormones	104
<i>NAT1</i>	Candidate-gene testing	Metabolism of aromatic and heterocyclic amines	105, 106
<i>NAT2</i>	Candidate-gene testing	Metabolism of aromatic and heterocyclic amines	105, 107
<i>COMT</i>	Candidate-gene testing	Metabolism of catechol oestrogens	108, 109
<i>SOD2</i>	Candidate-gene testing	Metabolism of superoxide anions	110, 111
<i>SULT1A</i>	Candidate-gene testing	Metabolism of carcinogens and endogenous hormones	112, 113
<i>GSTM1</i>	Candidate-gene testing	Altered detoxification of carcinogens	114, 115
<i>GSTP1</i>	Candidate-gene testing	Altered detoxification of carcinogens	114
<i>GSTT1</i>	Candidate-gene testing	Altered detoxification of carcinogens	114
<i>XRCC1, 3, 5</i>	Candidate-gene testing	Base-excision repair	116

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Table 1. Genes with germline mutations associated with breast cancer susceptibility (tab001kpb) (continued)

Gene name	Mode of discovery	Putative function	Refs
<i>TNF-α</i>	Candidate-gene testing	Altered TNF- α levels/signalling	117, 118
<i>HSP70</i>	Candidate-gene testing	Altered oestrogen signalling?	117, 118
<i>ESR1 (ERα)</i>	Candidate-gene testing	Altered oestrogen signalling	119
<i>AR</i>	Candidate-gene testing	Altered androgen signalling	36
<i>PGR</i>	Candidate-gene testing	Altered progesterone signalling	37
<i>AIB1</i>	Candidate-gene testing	Altered oestrogen signalling	38
<i>HLA</i>	Candidate-gene testing	Immune surveillance	120

Abbreviations: AIB, amplified in breast cancer 1; AR, androgen receptor; ATM, ataxia telangiectasia mutated; BRCA 1, 2, x: breast cancer 1, 2, x; CGH, comparative genomic hybridisation; CHK2, checkpoint homologue; COMT: catechol-O-methyltransferase; CYP, cytochrome P 450; ER, estrogen (oestrogen) receptor; GST, glutathione transferase; HLA: histocompatibility antigen; HRAS1-VNTR, variable number of tandem repeats; HSP70, heat shock protein 70; MLH1, MutL homologue 1; MSH2, MutS homologue 2; NAT, N-acetyl transferase 2; PGR, progesterone receptor; PTEN, phosphatase and tensin homologue; RDA, representational difference analysis; SOD2, superoxide dismutase 2; SULT1A, sulphotransferase 1A; TP53, tumour protein p53; TNF- α , tumour necrosis factor α ; XRCC 1, 3, 5, X-ray repair complementing defective repair in Chinese hamster cells 1, 3, 5.

and *BRCA2*, two additional loci, on chromosomes 8p and 13q, have been implicated in breast cancer susceptibility, but so far no new breast cancer susceptibility gene (referred to here as *BRCAx*) has been identified (Refs 22, 23, 24).

Both of these new loci were identified through initial molecular studies performed on the tumours of the affected patients. In the case of chromosome 8p, multiple independent studies have demonstrated a high fraction of loss of heterozygosity (LOH) in this region in sporadic tumours (Refs 25, 26). Many of these studies were performed in early-stage tumours, such as in ductal carcinoma in situ [DCIS; before the tumour grows through the breast duct wall into the surrounding tissues (Fig. 1)], and are therefore presumably devoid of random chromosomal alterations. Thus, the probability that these LOH events target a legitimate breast tumour suppressor gene is high. Initial linkage studies performed with chromosome 8p markers in a small set of German breast cancer families established a positive LOD score (this being the logarithm of the total relative probability that a linkage relationship exists among selected loci) for this locus (Refs 22, 23). However, in a larger set of families, no evidence for linkage was

found (Ref. 27). Therefore, if a *BRCAx* gene exists on chromosome 8p, it is likely to be responsible for only a very small portion of inherited cases.

The candidate 13q21 *BRCAx* locus was identified on the basis of comparative genomic hybridisation (CGH) studies in tumours of patients with familial breast cancers (at least three cases per family, and without germline mutations in known high-penetrance breast cancer genes) (Ref. 24). The most frequently deleted locus in these tumours was chromosome 13q21, and this was then tested in linkage studies in the affected families where a positive LOD score was established. However, linkage analyses in two different sets of breast cancer families could not confirm positive linkage to 13q21 (Refs 28, 29). The discrepancy between these studies could be due either to the fact that the families were selected on the basis of different criteria or to the fact that the 13q21 *BRCAx* gene is responsible for only a minority of familial cases. Only the identification of the gene implicated will determine which of these possibilities is true. Nevertheless, a similar approach (CGH followed by targeted linkage) has been successfully used for the identification of the *LKB1* tumour suppressor gene (Ref. 30). Therefore,

it is likely to be useful for the identification of *BRCA* genes as well. The best way to perform these types of studies would be to analyse multiple early-stage tumours from the same patient using a high-resolution genome-wide genetic screen, such as the bacterial artificial chromosome (BAC) or cDNA-array CGH, followed by high-throughput candidate-gene testing of the implicated loci.

The fact that despite all these efforts no *BRCAx* gene has been identified in the past six years indicates that the search for *BRCAx* is a difficult task. There are multiple possible reasons why *BRCAx* is more difficult to find than either *BRCA1* or *BRCA2* was. First, *BRCA1* and *BRCA2* predispose to other relatively rare cancer types – ovarian (both genes) and male breast cancer (*BRCA2*) – in addition to early-onset breast cancer, making the identification of affected families easier. Second, and somewhat related to the first problem, breast cancer is a very common cancer in women; therefore, it is possible that, by selecting families solely based on the occurrence of breast cancer, sporadic cases (i.e. not due to inherited mutation in a high-penetrance gene) will be inadvertently included, hampering the power of the linkage studies. Third, the penetrance of *BRCAx* might be much lower than that of *BRCA1* or *BRCA2*, and observing the phenotype could require cooperation with multiple low-penetrance modifier genes. Finally, it is possible that there are multiple high-to-moderate-penetrance *BRCA* genes to be identified, each being responsible for only a small number of inherited cases.

Association studies: low-penetrance genes

Candidate genes based on function

Whereas linkage studies in families are useful for the identification of high-penetrance, and to a lesser extent even moderate-penetrance, genes, low-penetrance genes are impossible to identify using this approach (Ref. 11). Although low-penetrance genes confer only a small-to-moderate breast cancer risk, variants of these genes might occur at a high frequency in the population. Therefore, combinations of low-penetrance genes are likely to be responsible for a much larger fraction of breast cancer cases than are high-penetrance genes.

To date, most low-penetrance genes have been identified on the basis of candidate selection approaches followed by association studies in

affected and control populations (Table 1) (Ref. 3). The simplest and most efficient version of these association studies is a case–control approach that compares allele frequencies in breast cancer and in control, unaffected patients. In order to be confident that the observed association of allele frequencies with breast cancer is not due to the biased selection of patients, the affected and the unaffected populations have to be well matched (with respect to ethnicity, age, gender and so on). Genes to be tested have usually been selected if their biochemical function is thought to play a role in breast tumorigenesis (Table 1). Therefore, several of these low-penetrance breast cancer susceptibility genes are involved in DNA-damage signalling and repair, hormone and xenobiotic metabolism, anti-oxidant defence, and immune surveillance (see Refs in Table 1).

In addition, polymorphisms in high-penetrance genes have been investigated to determine if any of these have an effect on breast cancer risk. For example, several common polymorphisms in *TP53*, *ATM* ('ataxia telangiectasia mutated'), *BRCA1* and *BRCA2* have been extensively studied in several different populations (Refs 3, 31, 32, 33, 34, 35). Although the increased risk of breast cancer associated with the R841W *BRCA1* variant remains to be confirmed in larger populations, several *BRCA2* variants and the Arg72Pro polymorphism in *TP53* do appear to confer a slightly elevated risk of breast cancer (Refs 3, 33).

Modifiers of high-penetrance genes

Another potential way to identify low-penetrance breast cancer susceptibility genes is to search for genes that modify the penetrance of high-penetrance genes, although many investigators believe that modifier genes probably account for only a tiny fraction of the total variance in these families. According to some investigators, the power of this approach might be that because there are many breast cancer cases in the affected cohort, even genes with slight effects on penetrance can be revealed. However, there are potential problems as well, including the possibility that modifiers of high-penetrance genes might not have an effect in the general population, and difficulties associated with selecting enough unaffected controls from the mutation carriers. In the case of *BRCA1* and *BRCA2*, several studies have been performed to identify genes that influence the age of onset and the occurrence of associated ovarian cancer

(Ref. 20). Among others, genes involved in oestrogen and androgen signalling (oestrogen, androgen and progesterone receptors, and an oestrogen receptor co-activator, AIB1), as well as detoxifying enzymes [glutathione S-transferase T1 (GSTT1)], might affect cancer penetrance in mutation carriers (Refs 36, 37, 38). In addition, targeted genotyping approaches indicate the presence of a modifier locus at 5q33-4, but the gene responsible has not been identified (Ref. 2). An interesting finding of an unselected case-control association study was that one of the alleles of *BRCA2* (N372H) had a different distribution among females and males, with females showing an excess of heterozygotes but no homozygotes, whereas the reverse was found in males (Ref. 34). This might indicate that there is a sex-specific selection for certain alleles either during the generation of the germ cells or in utero during embryonic development, owing to the fact that *BRCA2* function could be important for these processes. This result also indicates that breast cancer patients should be analysed for associated phenotypes unrelated to the cancer, such as infertility, since these might aid the discovery of new modifier alleles.

In addition to polymorphisms in the coding region of the candidate breast cancer susceptibility genes, recent results indicate that sequence variations in the non-coding regions such as promoters and introns could also have an effect on breast cancer risk. The molecular effects of these alterations are not always clear, but one possibility is that they influence the mRNA/protein levels of the genes. The importance of this type of effect is highlighted by a recent analysis of the effect of a sequence change in the *APC* ('adenomatous polyposis coli') gene in colon cancer (Ref. 39). An unidentified sequence alteration outside of the coding exons led to a twofold decrease in *APC* protein levels that was sufficient to initiate colonic polyp formation and increase colon cancer risk. Similarly, a polymorphism in the promoter region in the gene encoding insulin-like growth factor I (IGF-I) appears to be associated with increased breast cancer risk (Ref. 40).

SNP-ing the genome

Although the candidate-gene testing approach has been successful in identifying several putative new low-penetrance breast cancer genes, and will continue to be used in the future, it is inherently

limited to characterised genes (which currently constitute the minority of genes encoded by the human genome). Unbiased comprehensive studies are needed to uncover additional, uncharacterised breast cancer genes using polymorphic markers. So far, these studies have been technically challenging to complete in the general population. However, recent advances in many different fields will probably make these studies possible. The completion of the human genome project and subsequent re-sequencing of parts of the genome in different individuals has led to the discovery of a large number of single-nucleotide polymorphisms (SNPs) (Ref. 41). The identification of these SNPs, in combination with recently developed high-throughput SNP-genotyping methods, will enable us to perform genome-wide association studies in large populations (Refs 42, 43, 44). The recent finding that the human genome is probably composed of blocks (tens to hundreds of kilobases) of haplotypes might make the execution of genome-wide association studies easier (Ref. 45), since instead of individual SNPs of unknown relevance one can look at the frequency of haplotypes (containing multiple SNPs) in the affected and control populations. Although no study of this type has been performed in breast cancer, it is likely that such studies will be conducted in the future once the haplotype map of the human genome is defined. In addition to the technical challenges involved, there are several other difficulties that might make these types of studies impossible to perform or interpret. For example, the extent of linkage disequilibrium in surrounding common gene variants in various human populations is unknown. Similarly, the number of SNPs required to perform genome-wide studies is still subject to debate. Several recent reviews discuss the current status of SNP genotyping and future challenges (Refs 46, 47, 48, 49, 50).

Somatic genetic alterations in tumours Homozygous deletions and mutations

In addition to analysing germline mutations, studying somatic genetic alterations in breast tumours might also lead to the identification of candidate breast cancer susceptibility genes. The identification of the *PTEN* and *DPC4/Smad4* ('deleted in pancreatic carcinoma 4'/SMA and MAD related protein 4') tumour suppressor genes through mapping homozygous deletions in breast

and pancreatic tumours, respectively, proves the validity of this approach (Refs 51, 52). Similarly, the locus responsible for Peutz–Jeghers syndrome (the *LKB1* gene), which is an autosomal dominant condition associated with multiple gastrointestinal hamartomatous polyps and melanocytic spots of the lips, buccal mucosa and digits, was initially localised by CGH studies in tumours of affected patients (Ref. 30).

There are many different techniques that could be used for the identification of genes deleted or amplified in tumours. The *PTEN* tumour suppressor gene was discovered based on representational difference analysis (RDA) of a breast tumour followed by mapping of the deleted fragment and candidate-gene testing (Ref. 51). In the case of *DPC4*, detailed LOH mapping in multiple tumours led to the definition of a locus that corresponded to a region of consensus LOH and a homozygous deletion (Ref. 52). Although *PTEN* is almost never mutated in sporadic breast carcinomas, germline mutations within the gene are responsible for two inherited cancer predisposition syndromes: Cowden’s disease and Bannayan–Riley–Ruvalcaba syndrome (Refs 53, 54, 55).

Candidate-gene approaches are also applied in searches for genes somatically inactivated in tumours, and almost all newly identified candidate tumour suppressor genes tested for mutations in breast carcinomas show varying results. Examples for these genes are those encoding ‘Patched’ (*PTCH*), mitogen-activated protein kinase kinase 4 (*MKK4*) and E-cadherin (*CDH1*) (Refs 56, 57, 58, 59, 60) (Table 2). *PTCH* is a receptor for ‘Sonic hedgehog’ (*SHH*) and is mutated in a hereditary cancer predisposition known as Gorlin syndrome. In one study, analysis of *PTCH* in seven breast tumours identified two missense mutations of unknown relevance, whereas a larger study analysing 45 breast tumours found no evidence of genetic alterations (Refs 56, 57). Thus, alterations in *PTCH*/*SHH* signalling are unlikely to play an important role in breast tumorigenesis. The *MKK4* gene was implicated as a tumour suppressor gene since it was found to be homozygously deleted in two human tumour cell lines, derived from pancreatic and lung carcinomas (Ref. 58). E-cadherin is a cell adhesion protein with growth/invasion suppressor function and shows germline mutations in hereditary diffuse gastric carcinoma patients (Ref. 61). In addition, the *CDH1* gene

encoding E-cadherin is located on chromosome 16q, a region demonstrating frequent LOH in sporadic breast carcinomas (Ref. 62). Analysis of 42 medullary and infiltrating ductal breast carcinoma samples revealed no alterations in *CDH1*, but four out of seven lobular invasive carcinomas harboured a truncating mutation (Ref. 59). Since this initial study, many other groups have independently confirmed the presence of frequent *CDH1* mutations in lobular breast carcinomas, indicating that mutations in this gene play an important role in the development of lobular carcinomas. Despite the fact that *CDH1* mutations have been identified very infrequently in ductal carcinomas (Ref. 63), the expression of *CDH1* is frequently abolished by promoter methylation, and LOH at 16q is also frequently observed in these tumour types (Ref. 64). This might indicate either that elimination of *CDH1* protein function is also important in ductal carcinomas or that LOH at 16q targets an as-yet-unidentified tumour suppressor gene.

High-throughput genotyping and mutation screens

Recent technical advances make the search for somatic genetic alterations easier to perform and allow large-scale ‘brute-force’ approaches. The above-described genome-wide SNP arrays can be used for the localisation of regions deleted or amplified in the tumours. Similarly, a genome-wide BAC or cDNA-array CGH can facilitate the identification of new breast cancer genes (Refs 65, 66, 67). The simultaneous comprehensive genotype and gene expression analysis of tumours – using CGH or SNP arrays for genotyping, and cDNA/oligo arrays or serial analysis of gene expression (SAGE) for the analysis of gene expression patterns – promises to be especially powerful for the uncovering of new candidate genes.

A massive ‘brute-force’ project for new cancer gene discovery was recently initiated by the Wellcome Trust Fund at the Sanger Institute (Cambridge, UK) (see <http://www.sanger.ac.uk/CGP/> for details). One of the goals of this project is to conduct a genome-wide homozygous deletion screen in over 1200 human cancer cell lines using polymorphic CA/GT repeats followed by STS screens (‘STS’ stands for sequence-tagged site; short stretches of unique DNA sequence that can be specifically detected by a PCR assay), with the aim of identifying new tumour suppressor

Table 2. Genes with somatic genetic and epigenetic alterations in breast cancer (tab002kpb)

Gene	Alteration	Putative function	Refs
<i>MKK4</i>	Mutation	Signal transduction	58, 72
<i>PTCH</i>	Mutation	Hedgehog signalling	56, 57
<i>CDH1</i>	Mutation/methylation	Cell adhesion	60, 64
<i>17q-erbB2</i>	Amplification	Receptor tyrosine kinase	73, 121
<i>8q-c-myc</i>	Amplification	Transcription factor	73, 122
<i>11q-CCND1</i>	Amplification	Cell cycle/transcription	73, 123
<i>11q-FGF3</i>	Amplification	Growth factor	73
<i>11q-FGF4</i>	Amplification	Growth factor	73
<i>20q-AIB1</i>	Amplification	Transcription co-activator	124
<i>20q-ZNF217</i>	Amplification	Transcription factor	125
<i>CDKN2A</i>	Methylation	Cell cycle	126, 127
<i>CCND2</i>	Methylation	Cell cycle	128
<i>SFN</i>	Methylation	Cell cycle	77, 129
<i>RARβ2</i>	Methylation	Transcription factor	130
<i>HIN-1</i>	Methylation	Secreted growth inhibitor	76
<i>BRCA1</i>	Methylation	Recombination/transcription	131
<i>GSTP1</i>	Methylation	Metabolism	132
<i>FABP3</i>	Methylation	Secreted growth inhibitor	133
<i>HOXA5</i>	Methylation	Transcription factor	134
<i>ARHI</i>	Imprinting/LOH	Ras/Rap signalling	135

Abbreviations: AIB, amplified in breast cancer 1; ARHI, Ras homologue gene family member I; BRCA1, breast cancer 1; CCND1, cyclin D1; CCND2, cyclin D2; CDH1, E-cadherin; CDKN2A, cyclin-dependent kinase inhibitor 2A (p16); erbB2, erythroblastic leukemia viral oncogene homologue 2; FABP3, fatty-acid-binding protein 3; FGF 3, 4: fibroblast growth factor 3, 4; GST, glutathione transferase; HIN-1, 'high in normal' 1; HOXA5, homeo box A5; MKK4, mitogen-activated protein kinase kinase 4; myc, v-myc myelocytomatosis viral oncogene homologue; PTCH, patched; RARβ2, retinoic acid receptor β2; SFN, stratifin (14-3-3σ); ZNF217, zinc finger protein 217.

genes. In addition to the deletion mapping in cell lines, the investigators are also planning to sequence systematically all the coding exons and flanking splice junctions of every transcribed or predicted human gene in multiple tumour types including 16 primary breast carcinomas and matched normal genomic DNA (for details,

see <http://www.sanger.ac.uk/CGP/>). The completion of this project could lead to the identification of many new breast cancer genes.

The use of somatic genetic changes to uncover new breast cancer genes is limited by the fact that, owing to genomic instability, tumours can have very high rates of somatic genetic events. Many

of these somatic changes might be random and not related to the tumourigenic process. The use of early-stage tumours, possibly multiple tumours from the same individual, can overcome this problem. The first recognised obligate precursor of invasive tumours is DCIS, as described above, but unfortunately even DCIS tumours have already accumulated multiple genetic alterations and some are aneuploid (Refs 68, 69). Earlier-stage tumours, such as usual or atypical ductal hyperplasia, are not easy to diagnose and their malignant potential and relationship to invasive carcinomas is unclear at this point.

Another complicating problem with searching for breast cancer genes based on LOH studies in tumours is the fact that certain chromosomal regions show frequent alterations that are not due to the genes located in the region but instead to fragile chromosome structure. The most common fragile sites in the human genome are 3p, 6q, 7q, 16q and 17q (Ref. 70). Even homozygous deletions, which are relatively rare and thought to be the clear indication of a tumour suppressor gene, can be misleading, since two recent studies reported polymorphic homozygous deletions without any specific gene targets (Refs 71, 72).

Despite all the potential problems associated with the analysis of somatic genetic alterations, one clear advantage of this approach is that it allows the identification of genes involved in tumour progression, including those required for invasion, angiogenesis and metastasis. In summary, although analyses of somatic genetic alterations have led to the identification of several breast cancer genes, it has thus far proven to be a fairly inefficient method. However, the use of new high-throughput sequencing approaches, like the one currently applied by the Cancer Genome Project at the Sanger Institute, will probably yield many new putative breast cancer genes in the near future.

Amplifications and epigenetic changes

Certain genes and chromosomal regions are amplified in breast tumours (Table 2). Increased gene copy number changes can be detected by CGH or as homogeneously staining (HS) regions (chromosomal regions that show unusual homogenous cytogenetic staining) or double minutes (DMs; very small accessory chromosomes) and they usually lead to the overexpression of genes included in the region of

amplification (Ref. 73). Contrary to deletions, where homozygous inactivation of a gene is a definitive proof that the gene is the target of the deletion, in the case of amplifications it is frequently difficult to determine which gene is the critical target of this genetic event. This is due to multiple reasons: (1) amplifications are frequently large and include many genes; (2) the amplified gene might not always be overexpressed in the tumour; and (3) there might be a cluster of genes targeted by the event. Because of these facts, the above-described combined genotype/gene expression pattern analysis of breast tumours could be particularly useful for the identification of new genes targeted by amplifications. The best-characterised amplicons in breast cancer are on chromosome 8q, 11q, 17q and 20q (Ref. 73). For all of these amplicons, there are many candidate genes identified, such as *c-myc* (8q), *CCDN1* (11q; encoding cyclin D1), *FGF3* and *FGF4* (11q; encoding fibroblast growth factor 3 and 4), *HER2/erbB2* (17q; encoding erythroblastic leukemia viral oncogene homologue 2), *AIB1* (20q) and *ZNF217* (20q; encoding zinc finger protein 217).

In addition to genetic changes, epigenetic alterations including methylation or imprinting could also lead to the somatic inactivation of candidate tumour suppressor genes. Examples of genes showing epigenetic changes are listed in Table 2. Genes methylated in breast cancer could also be identified using comprehensive gene expression profiling (if the gene is consistently downregulated in tumours) or by arrays or subtractive hybridisation approaches specifically designed for the detection of methylated genes (Refs 74, 75). Examples of genes identified using the former approach – in this case, SAGE analysis of normal and cancerous mammary epithelial cells – are *HIN-1* ('high in normal 1') and *14-3-3 σ* (encoding stratifin) (Refs 76, 77).

As demonstrated by the effect of *AIB1* on *BRCA1* penetrance, polymorphisms in amplified or epigenetically altered genes might influence breast cancer susceptibility, but not many such studies have been performed to date. In the case of amplified genes, sequence variants of the promoter region that alter expression, or in the coding region that alter activity, could modify breast cancer risk. Similarly, polymorphisms of CpG sites in the promoter of methylated genes could confer increased resistance to breast cancer. In addition, genes targeted by these events are excellent therapeutic or diagnostic targets, since

amplifications or overexpression usually result in gain-of-function phenotypes that are necessary for the tumour cells, whereas genes specifically methylated in cancers could be used for breast cancer diagnosis.

Animal models of breast cancer

The nearly completed mouse and rat genome sequences and the almost complete synteny between these rodent genomes and the human genome makes possible the use of animal models of breast cancer for the identification of new candidate cancer susceptibility genes. This is especially true in the mouse where genetic crosses can be performed to investigate complex genetic diseases. In the mouse, there are already several mammary tumour models that to some extent reflect the human disease (Ref. 78). The most promising approach for the generation of these models is the somatic random inactivation of tumour suppressor genes or the activation of oncogenes only in the desired cell type or tissue. An example of this is the generation of mammary-gland-specific knock-outs of *BRCA1* and *BRCA2* using cre recombinase under the control of the whey acidic protein (WAP) promoter (Refs 79, 80). These mouse models then can be used to search for modifiers of the penetrance of the phenotype.

However, a major problem with the use of mice as models of human breast cancer is that mouse strains are very inbred, and are therefore unlikely to reflect the highly variable human populations. In addition, the differing reproductive and environmental factors and lifespans of rodents and humans are problems that severely limit the usefulness of this approach for the identification of genes relevant to human disease. This is demonstrated by the use of mice that carry a mutated multiple intestinal neoplasia (*Min*) gene, which is the mouse homologue of the human *APC* gene, resulting in the development of intestinal tumours. The *Min* mouse is a fairly faithful model of human colon cancer and has therefore been used to identify *APC* modifier genes. However, although one of the putative modifiers identified ['modifier of *Min*' (MOM1), a secretory phospholipase *Pla2g2a*] strongly inhibited intestinal polyp formation in mice, it does not seem to play a role in human colorectal tumours (Refs 81, 82, 83).

Although rat strains are less inbred than mice, and rat mammary carcinomas are more

similar to human breast cancers both in terms of histopathological progression and hormone responsiveness, many of the problems associated with the use of mice also hold true in rats (Ref. 84). In addition, rats are less amenable to genetic manipulation, and so most of the rat breast cancer models are based on chemical-induced carcinogenesis that is unlikely to reflect the human disease.

Thus, although rodent models of breast cancer have the potential to aid in the identification of new breast cancer genes, their usefulness remains to be proven. However, one definite advantage of animal models of cancer is that they can be used for the identification of new cancer preventive and therapeutic strategies that can be translated into human clinical trials.

Clinical implications and applications

What is the clinical importance of identifying breast cancer genes? In the case of inherited cancer susceptibility, identifying and testing the genes responsible is useful if the result of the test would change the clinical management of the patient (Ref. 85). However, this is true only if there are effective early diagnostic and intervention strategies available for the particular cancer type the patient is predisposed to. In the case of the known high-penetrance breast cancer susceptibility genes *BRCA1* and *BRCA2*, the value of genetic testing is still only presumed and not yet established (Ref. 86). Currently, the most effective preventive therapy of choice is bilateral mastectomy, but many women with familial risk of breast cancer and even proven mutation carriers do not want to undergo this treatment (Ref. 87). Therefore, new technologies for the early detection of, and new preventive therapies for, breast and ovarian cancer are needed.

A recently described proteomic pattern technology appears to be very promising since, in a preliminary study, it demonstrated 100% sensitivity and 95% specificity for the diagnosis of ovarian cancer (Ref. 88). The approach is based on surface-enhanced laser desorption/ionisation time-of-flight mass spectrometry (SELDI-TOF MS), which combines chromatography and mass spectrometry, together with bioinformatic analysis of serum from affected and unaffected patients, resulting in the definition of a 'normal' and 'cancer' pattern. Similar approaches are currently being developed for the diagnosis of breast and other cancer types.

Polymorphism screens for candidate low-penetrance breast cancer genes are currently performed only for research purposes. Genetic testing for multiple low-penetrance genes might eventually become reality, but only if accurate prediction is possible. The complexity of polygenic susceptibility and the possible combination of genotype with environmental and behavioural factors make the feasibility and usefulness of this type of test somewhat questionable (Ref. 89).

Ideally, understanding the genetic basis of breast cancer should lead to more-effective cancer prevention and treatment, since genetic defects are potential therapeutic targets. However, in many cases, genetic abnormalities are not tractable pharmaceutically. The analysis of tumour samples with associated clinical data using genomic technologies promises to be useful for the identification of new molecular targets that determine the clinical behaviour of the tumours. Many such studies on many different cancer types have been performed in the past few years, and many candidate targets have been identified. However, future studies are required to evaluate the clinical potential of these newly identified genes. The success of anti-oestrogen therapy (e.g. tamoxifen) for the prevention and treatment of oestrogen-receptor-positive breast tumours and the use of the recombinant humanised anti-Her2/ErbB2 monoclonal antibody trastuzumab (Herceptin®) for the treatment of Her2/ErbB2-positive tumours suggest that further molecular-based therapies will probably be identified and successfully used in the clinical management of breast cancer patients (Ref. 90).

Outstanding research questions and future prospects

Despite all of the efforts, the genetic basis of the majority of breast carcinomas is still poorly defined. However, it is clear that there is not a single breast cancer gene, an equivalent of *APC* in colorectal tumours, and that hereditary and sporadic breast tumours might have their own tumourigenic pathways. Therefore, the search must continue to find additional breast cancer genes using all the available traditional and new genomic techniques. The major challenge of identifying other breast cancer genes might be solved by using the new technical tools becoming available. However, the major obstacle has been, and continues to be, the translation of these

findings into clinical practice. As demonstrated by the success of Herceptin® and STI571 (Gleevec™), an inhibitor of the Bcr-Abl tyrosine kinase for the treatment of chronic myelogenous leukaemia (Ref. 91), molecularly targeted cancer treatment is much more effective, with fewer side effects, than the other currently used anti-cancer therapies. Many new putative breast cancer therapeutic and diagnostic targets are being investigated in the clinic and many more are likely to follow. The identification of additional breast cancer susceptibility genes is sure to aid the identification of individuals at risk and the design of targeted cancer-preventive therapies.

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Further reading, resources and contacts

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Meghan A. Arnold and Michael Goggins (2001) BRCA2 and predisposition to pancreatic and other cancers. *Exp. Rev. Mol. Med.* 14 May, <http://www.expertreviews.org/0100309Xh.htm>

The Wellcome Trust Fund Sanger Institute's website has details on the Cancer Genome Project:

<http://www.sanger.ac.uk/CGP/>

Comparative genomic hybridisation data on 38 human breast cancer cell lines are collated at the US National Human Genome Research Institute website:

<http://www.nhgri.nih.gov/DIR/CGB/CR2000>

Loss of heterozygosity studies in breast cancer are summarised at the University of Nottingham, UK, website:

<http://www.nottingham.ac.uk/~pdzmgh/loh>

Features associated with this article

Figures

Figure 1. A hypothetical model depicting the multistep process of breast tumourigenesis (fig001kpb).

Figure 2. Summary of approaches used for breast cancer gene discovery (fig002kpb).

Tables

Table 1. Genes with germline mutations associated with breast cancer susceptibility (tab001kpb).

Table 2. Genes with somatic genetic and epigenetic alterations in breast cancer (tab002kpb).

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