Deployment of either a whole or dissected wild nuclear genome into the wheat gene pool meets the breeding challenges posed by the sustainable farming systems

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Abstract

Deploying whole and dissected nuclear genome of wild Triticeae species in the homoeologous wheat genetic background through inter-specific hybridization and introgression is a lower cost and effective option to prepare wheat germplasm with unexploited genes for disease resistance and enhanced grain yield and quality traits. The whole nuclear genomes of Dasypyrum villosum (Dv) and T. turgidum var durum have been combined, and an homoploid derivative of the original amphiploid displayed typical 'farro' spike morphology, tough rachis and the adaptive traits of Dv such as high resistance to diseases (caused by *Tilletia tritici*, Blumeria graminis f. sp. tritici, Puccinia triticina and P. graminis f. sp. tritici), heading earliness and fortified caryopses (high protein and micronutrient contents). The dissection of the Dv genome by either 'Triticum aestivum cv Chinese Spring (CS) × hexaploid amphiploid' or $(CS \times Dv) \times CS'$ hybridization and backcrossing provided wheat introgression breeding lines (IBLs) expressing one or more of the Dv adaptive traits. Molecular analyses revealed that either cryptic or Genomic In-situ Hybridization (GISH) detectable Dv chromatin introgression occurred in those IBLs. The IBLs, after 2 years of low-input field tests and genetic analyses in Italy and Hungary, showed simple inheritance, dominance and stability of the adaptive and disease resistance traits.

Keywords: Dasypyrum gene pool; gene transfer; introgression; wheat germplasm; wild plant species

Introduction

In order to meet the predicted global cereal grain demand for the next decades, efficient conventional

and molecular cereal breeding programmes and proper wheat genetic resources are necessary for selecting the suitable breeding lines (Baenziger *et al.*, 2008).

Many dominant genes for adaptation and trait enhancement have been lost during cereal crop domestication, but they have been retained in the genome of the wild components of the *Triticeae* gene pools (Dwivedi *et al.*, 2008). In natural habitat, wild *Triticeae* species

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such as *Dasypyrum villosum (Dv)*, whose genome was exposed to million of years of climatic and environmental changes, are now expressing increased heading earliness, density stands and plant biomass. Deploying whole and dissected *Dv* nuclear genome in the homoeologous wheat genetic background through interspecific hybridization and introgression could be a lower cost and effective option to help wheat breeders to merge and select the proper adapted gene pools to sustain the needed yearly grain yield increase. In this study, we show that combining the *Dv* genome with the *T. turgidum* var *durum* genomic background and deploying dissected *Dv* genome in *Triticum aestivum* provided wheat genetic resources with new trait enhancements.

Materials and methods

Combining the whole nuclear genomes of Dv and T. turgidum var. durum

A hulled and brittle rachis *Dv* ecotype (2n = 2x = 14; VV) collected near Bari (Puglia, Italy) was used as pollen parent in hybridization with the free-threshing and tough rachis *T. turgidum* var. *durum* cv 'Modoc' (2n = 4x = 28; AABB) (Jan *et al.*, 1986). The resulting hexaploid amphiploid (2n = 42; AABBVV), labelled $M \times V$ -b₁, was fertile and showed brittle rachis (Supplementary Fig. S1, available online only at http://journals.cambridge.org). A non-brittle rachis mutated amphiploid plant $(M \times V$ -nb₁; AABBVV; 2n = 6x = 42) was discovered in the $M \times V$ -b₁ plot grown in 1987 (De Pace *et al.*, 2003). In the following years, other genetically stable variants ('Mut 7-04', 'Mut 12-04' and 'Mut 16-04') were found among the $M \times V$ -nb₁ plants.

Dissection of the Dv genome by [T. aestivum cv 'Chinese Spring' (CS) \times Dv] \times CS hybridization and backcrossing

We selected six Dv-introgressed wheat breeding lines (IBLs), sharing a common CS (2n = 6x = 42; A'A'B'B' D'D') genetic background, from a population of 150 aneuploid lines developed at the University of Tuscia, Viterbo, Italy, from the backcross $(CS \times Dv) \times CS$ (Supplementary Fig. S2, available online only at http://journals.cambridge.org). The IBLs CS × V63 and $CS \times V32$ contained a disomic addition of 6V and a disomic substitution 6V(6B), respectively, and CS 1B-1V line contained a pair of 1BL-1VS chromosomes from a spontaneous exchange between chromosome 1B of wheat and 1V of Dv. The IBLs CS × V58, CS × V59 and $CS \times V60$ were morphologically different from CS, although they did not contain apparent GISH detectable V chromatin (Minelli et al., 2005; Caceres et al., 2008). The IBLs were evaluated for resistance to Blumeria graminis f. sp. tritici (Bgt), Puccinia triticina (Pt) and P. graminis f. sp. tritici (Pgt) isolates in Italy and Hungary, and for end-use grain quality traits.

Dissection of the Dv genome by hybridization of T. aestivum cv CS and $M \times V$ -nb₁ hexaploid amphiploid

An F_2 -like breeding population with broad genetic diversity was obtained from selfing the F_1 plants obtained after crossing the hexaploid amphiploid $M \times V$ -nb₁ to CS (Supplementary Fig. S3, available online only at http://journals.cambridge.org). After two generations of selfing, three F_4 lines, named '8-1', '41-3' and 'Mut 3-04', were tested for two consecutive growing seasons

Table 1. Mean values for heading time, plant height, number of spikelets/spike, 1000 kernel weight and protein content in $M \times V$ -nb₁ hexaploid amphiploid and derived variants by spontaneous mutation in comparison with 'farro' species used as controls^a

Entry	Heading time (days from 1st April)	Plant height (cm)	Number of spikelets/spike	1000 Kernel weight (g)	Protein content (% dry weight)
Hexaploid amphiploid					
M × V-nb₁ ΄	114	101	14.7	27.1	21.1
Derived variants from s	pontaneous mutation in t	the hexaploid amp	hiploid		
Mut 7-04	116	105	14.6	23.5	21.5
Mut 12-04	124	104	16.3	42.6	16.5
Mut 16-04	124	106	15.9	42.6	16.9
Control 'farro' species					
T. monococcum	144	109	17.7	20.0	20.0
T. dicoccoides	132	125	15.5	35.5	22.0
T. spelta	135	124	17.0	39.8	18.8

^a Data were taken from plots at S. Angelo Lodigiano (Lodi, Italy) in 2007 and 2008.

								srabender farinog	graph	Chopi	n alveogi	raph	Br	ead test	
Line	Year	TW (kg/hl)	PC (%dm)	SSV (ml)	SSSV	Gluten index	Stab. (min)	Degree of softness (BU)	Water abs (%)	P (mm)	L (mm)	ЬЛ	$(\times 10^{-4}J)$	Vol (cm ³)	Height (mm)
8-1	2008 2009	74.9 79.6	12.6 13.8	91.0 88.5	7.2 6.4	99 84	18.2 17.4	23 25	57.8 63.0	104 80	128 130	0.81	400 286	710 735	100 103
41-03	2008	76.4 79.8	12.3	87.5	7.2 6.1	- 80 80	14.6 17.8	25 21	57.7 62.6	92 72	119	0.77	334 234 234	685 770	101
Mut 3-04	2008 2008	77.0 80.0	12.6	91.0 88.5	7.2	99 76	13.1	33 12	58.1 62.9	93 81	136	0.68	375 266	720 790	105
Control CS	2008	64.7	12.2	48.0	3.9	32	3.9	93	56.4	5 5	61	0.89	06	560	84
Bologna PR22R58	2009	80.4 76.4	13.9	86.5 87.5	6.2 7.2	97 97	18.5 19.0	22 21	58.5 54.5	78	140 115	0.57 0.49	396 219	720 695	105 98
Best check	2009		1	2	!	5					-	5	1	785	109
Stab., stability,	; BU, Bra	thender U	Inits; abs,	absorptic	m; P, pre	ssion, mea	asures the	e tenacity of the	: dough; L, lei	ngth, mea	sures the	e extens	ibility of the	dough; M	/, work,

Table 2. Average values for test weight (TW), protein content (PC), SDS sedimentation volume (SSV), specific SDS sedimentation volume (SSSV = SSV/PC), gluten index, rheological flour dough properties (Brabender farinograph and Chopin alveograph) and bread quality, determined for three inbred breeding lines obtained from hybridization of *T. aestivum* cv CS and M × V-nb₁ hexaploid amphiploid^a

measures the strength of the dough. ^a The values are mean of plots raised at two locations (S. Angelo Lodigiano and Viterbo) in Italy for 2 years (2008 and 2009).

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(2007/2008 and 2008/2009), in the field at two sites: S. Angelo Lodigiano, SAL, near Lodi in northern Italy, and Tolentino, TOL, near Macerata in central Italy. The plots were managed using low-input criteria. The hexaploid wheat cultivars 'Bologna' and 'PR22R58' were used as checks. Heading time (days from 1st April), yield components and rheological properties of flour dough were evaluated at both sites.

Results and discussion

Combining the whole nuclear genomes of Dv and T. turgidum var durum

The hexaploid amphiploid $M \times V$ -nb₁ and the derived homoploid variants displayed 'farro' traits (tenacious glumes but tough rachis) and the typical adaptive traits of *Dv*, such as high resistance to diseases (caused by *Tilletia tritici*, *Bgt*, *Pt* and *Pgt*), fortified caryopses (+17 to +21% protein contents and >+25% of Fe and Zn content, compared with CS) and heading earliness. When compared with the conventional 'farro' (*T. monococcum*, *T. dicoccoides*, and *T. spelta*), new 'farro' types expressed earlier heading and shorter culms (Table 1), and also caryopses (Mut 12-04 and Mut 16-04) with larger size.

Dissection and deployment of the Dv genome by $(CS \times Dv) \times CS'$ hybridization and backcrossing

This type of hybridization favoured the haploidization of the V genome in the heterohaploid $A'B'D'V F_1$, the random assortment (dissection) of V chromosomes in 7A' + 7B' + 7D' + 1V gametes and the formation of wheat IBLs with the disomic addition of one of the V chromosome or V chromosome arm after backcrossing to CS. The IBLs $CS \times V32$ and $CS \times V63$ contained the chromosome 6V#4 (De Pace et al., 2011), conferring multiple disease resistance to virulent strains of Bgt, Pt and Pgt. The IBL CS_1BL-1VS expressed superior Sodium Dodecyl Sulfate (SDS) sedimentation and rheological properties of flour dough than CS. Other IBLs (CS \times V58, CS \times V59 and CS \times V60) had cryptic *Dv* chromatin introgression exhibiting a -7 to -9d earliness in heading time compared with CS in the field. Using specific primers for several DNA targets in the genome of these lines revealed Dv alleles, but not CS alleles, at two loci (Vrn-A1 and Vrn-B3) involved in the vernalization response pathway (Caceres et al., 2008).

 F_3 progenies derived from the hybridization of two disomic addition lines for chromosome 6V, CS × V63 (+6V#4) and CS + 6V#1 (produced by Sears, 1982, susceptible to *Bgt*, *Pt* and *Pgt*; Supplementary Fig. S4,

available online only at http://journals.cambridge.org) expressed simple inheritance, due to *Dv*-derived Mendelian dominant genes governing resistance response to each of the *Bgt*, *Pt* and *Pgt* pathogen. The high SDS sedimentation value of CS_1BL-1VS was traced to the effect of one high-molecular weight glutenin subunit (1v in Supplementary Fig. S5, available online only at http://journals.cambridge.org) encoded at the *Ghu-V1* locus in 1VS of CS_1BL-1VS (Vaccino *et al.*, 2010).

Dissection and deployment of the Dv genome by hybridization of T. aestivum cv CS and $M \times V$ -nb₁ hexaploid amphiploid

'New' chromosome assortments were achieved using the AABBVV hexaploid amphiploid as a bridge to combine the A and B genomes from durum wheat; the A', B' and D' genomes of bread wheat, and the V genome of Dv. Tetraploid AA'BB' lines, hexaploid AA'BB'D'D' lines, and aneuploid AA'BB' lines with the addition of one or more V and D' chromosomes were obtained (Supplementary Figs. S3 and S6, available online only at http://journals.cambridge.org). Promising outcomes of this breeding scheme were the high frequency of euploid segregants (Supplementary Fig. S3, available online only at http://journals.cambridge.org), the phenotypic uniformity observed in the progenies after only two generations of selfing that followed the $CS \times (M \times V-nb_1)$ triparental hybridization and the good agronomic values of those lines. The three inbred breeding lines, '8-1', '41-3' and 'Mut 3-04', had chromosome counting of 2n = 42 and were as good as the best check for yield components, grain quality and rheological flour dough properties (Table 2 and Supplementary Table S1, available online only at http://journals. cambridge.org).

In conclusion, it is suggested that a sustainable breeding response to mitigate the severe threat to the world's wheat supply is necessary (Hovmøller *et al.*, 2010). We evidenced that deploying whole and dissected Dvnuclear genome in the homoeologous wheat genetic background, it is possible to prepare wheat germplasm with unexploited genes for rust disease resistance and enhanced grain yield and quality traits.

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