

# The impact of parasites during range expansion of an invasive gecko

Louise K. Barnett<sup>1\*</sup>, Ben L. Phillips<sup>2</sup>, Allen C. G. Heath<sup>3</sup>, Andrew Coates<sup>2</sup>  
and Conrad J. Hoskin<sup>1</sup>

## Research Article

\*Present address: Global Ecology, College of Science and Engineering, Flinders University, Adelaide, South Australia 5001, Australia.

**Cite this article:** Barnett LK, Phillips BL, Heath AC G, Coates A, Hoskin CJ (2018). The impact of parasites during range expansion of an invasive gecko. *Parasitology* **145**, 1400–1409. <https://doi.org/10.1017/S003118201800015X>

Received: 3 September 2017  
Revised: 14 January 2018  
Accepted: 15 January 2018  
First published online: 14 February 2018

### Key words:

*Geckobia*; *Hemidactylus frenatus*; host–parasite dynamics; invasive species; mite; parasite release; parasite spillback; pentastome; *Waddycephalus*

Author for correspondence: Louise K. Barnett,  
E-mail: [louisekarnett@gmail.com](mailto:louisekarnett@gmail.com)

<sup>1</sup>College of Science and Engineering, James Cook University, Townsville, Queensland 4811, Australia; <sup>2</sup>School of Biosciences, University of Melbourne, Parkville, Victoria 3010, Australia and <sup>3</sup>AgResearch Ltd, Hopkirk Research Institute, Palmerston North 4442, New Zealand

### Abstract

Host–parasite dynamics can play a fundamental role in both the establishment success of invasive species and their impact on native wildlife. The net impact of parasites depends on their capacity to switch effectively between native and invasive hosts. Here we explore host-switching, spatial patterns and simple fitness measures in a slow-expanding invasion: the invasion of Asian house geckos (*Hemidactylus frenatus*) from urban areas into bushland in Northeast Australia. In bushland close to urban edges, *H. frenatus* co-occurs with, and at many sites now greatly out-numbers, native geckos. We measured prevalence and intensity of *Geckobia* mites (introduced with *H. frenatus*), and *Waddycephalus* (a native pentastome). We recorded a new invasive mite species, and several new host associations for native mites and geckos, but we found no evidence of mite transmission between native and invasive geckos. In contrast, native *Waddycephalus* nymphs were commonly present in *H. frenatus*, demonstrating this parasite's capacity to utilize *H. frenatus* as a novel host. Prevalence of mites on *H. frenatus* decreased with distance from the urban edge, suggesting parasite release towards the invasion front; however, we found no evidence that mites affect *H. frenatus* body condition or lifespan. *Waddycephalus* was present at low prevalence in bushland sites and, although its presence did not affect host body condition, our data suggest that it may reduce host survival. The high relative density of *H. frenatus* at our sites, and their capacity to harbour *Waddycephalus*, suggests that there may be impacts on native geckos and snakes through parasite spillback.

### Introduction

Invasive species pose an increasing threat to global biodiversity (Floerl *et al.* 2009). While invasives have numerous direct effects – through predation, competition or hybridization with native species (Menge, 1972; O'Dowd *et al.* 2003; Wanless *et al.* 2007) – they may also have powerful indirect effects, by spreading parasites and diseases (Dunn and Hatcher, 2015). Each introduced species brings an average of three parasite species with them from their native range (Torchin *et al.* 2003), and these parasites can have serious impacts on native species (Andreou *et al.* 2012). The impact of parasites during biological invasions is challenging to unravel, involving interactions between native and introduced hosts and their respective parasite communities (Dunn *et al.* 2012; Telfer and Bown, 2012; Perkins *et al.* 2017).

Introduced hosts may affect parasite prevalence in native hosts in three ways (Telfer and Bown, 2012). First, introduced parasites may switch to native hosts, causing a 'spillover' of parasites (Andreou *et al.* 2012; Miller *et al.* 2017). Naïve native species are often highly susceptible to introduced parasites, while invasive species, which have co-evolved with these parasites, may not be obviously affected (Andreou *et al.* 2012). Thus, spillover may be a primary mechanism of impact on native hosts (Hudson and Greenman, 1998). Second, invasive hosts acquire native parasites in the new range (Torchin and Mitchell, 2004). The presence of this new (often highly abundant) host, may cause a dramatic increase in native parasite abundance, which may affect native hosts through parasite 'spillback' (Kelly *et al.* 2009; Hartigan *et al.* 2011; Kelehear *et al.* 2013). Third, the opposite may happen if the introduced host acquires native parasites but the native parasite is not competent in this new host. In this case, an introduced host potentially reduces parasite prevalence by diluting parasite infection in native hosts (Telfer and Bown, 2012). A first step in resolving these possibilities is, of course, to assess parasite transmission from introduced to native hosts.

Conversely, if invasive species are susceptible to native parasites, acquisition of these parasites may impact the fitness of the invasive host (Krakau *et al.* 2006; Telfer and Bown, 2012). In such cases, native parasites might affect the establishment, impact, or range expansion of an invasive species (Case and Taper, 2000; Dunn, 2009; Perkins, 2012). Indeed, negative impacts of native parasites on invasive species have been recorded and likely affect the competitive ability of the invader (Krakau *et al.* 2006; Dunn, 2009; Gendron *et al.* 2012). Again, a first step in resolving these possibilities is to assess parasite transmission, this time from native to introduced hosts.

While invasive species do bring novel parasites and pathogens with them, they generally leave many of their natural parasites behind when they establish in their new range

(MacLeod *et al.* 2010). This ‘parasite release’ – driven by founder events or a lack of intermediate hosts in the new range – may facilitate establishment and range expansion of invasive species (Torchin *et al.* 2003; Prenter *et al.* 2004; Dunn and Hatcher, 2015). As well as this loss of parasites at the establishment, parasites often lag behind during subsequent range expansion, and populations on the leading edge of the invasion may be exposed to a lower prevalence of parasites than longer established populations (Phillips *et al.* 2010). This may give invasion front populations an advantage and facilitate the growth of dense populations in newly colonized areas (Phillips *et al.* 2010).

Clearly, host–parasite dynamics can play a crucial role in both the success of an invasion and the impact the invader has on recipient communities. The possible outcomes are manifold and complex, but the list of possible outcomes can be rapidly shortened with observational data on host-switching and impacts on host fitness. Issues of parasite loss during ongoing invasion can also be assessed by examining changes in prevalence through space. Here we address these questions for an invasive gecko spreading into the gecko-rich woodlands of northern Australia.

The Asian house gecko, *Hemidactylus frenatus*, is a human-commensal species that has been accidentally introduced to many tropical and subtropical areas globally (Lever, 2003; Hoskin, 2011). These geckos often establish large populations where they are introduced, and in some areas have caused declines and local extinctions of native species (Cole *et al.* 2005; Dame and Petren, 2006). Despite documented impacts in some areas, *H. frenatus* is typically considered a benign invader due to the belief that it is restricted to built areas (Vanderduys and Kutt, 2013). In northern Australia, however, *H. frenatus* are spreading from urban areas into bushland (Hoskin, 2011; Barnett *et al.* 2017). This range expansion is bringing *H. frenatus* into contact with a number of ecologically similar native species (Hoskin, 2011; Barnett *et al.* 2017). Are these native species acquiring the parasites of *H. frenatus*? Are *H. frenatus* acquiring native parasites? And, if so, does such host switching affect host fitness?

In this study, we focussed on mites (*Geckobia* and *Neotrombicula*) and pentastomes of the genus *Waddycephalus* because they are visible externally on geckos in the field. At least seven species of *Geckobia* mites have been recorded on *H. frenatus* in other parts of its introduced range (Heath and Whitaker, 2015) but only *Geckobia bataviensis* is previously known to have been introduced with *H. frenatus* into Australia (Domrow, 1992; Hoskin, 2011). There are many native gecko mites in Australia, including *Geckobia* and Trombiculid mites of the genera *Ascoschoengastia*, *Neotrombicula* and *Trombicula* (Domrow and Lester, 1985). There are no records of *H. frenatus* hosting native Australian gecko mites, nor of native Australian geckos hosting introduced mites, but this has not been assessed systematically. *Geckobia* mites live their entire lives on geckos and are likely transmitted through direct contact between individuals (Bauer *et al.* 1990; Rivera *et al.* 2003), whereas the Trombiculid gecko mites have a free-living stage and geckos likely pick them up from their environment (Domrow and Lester, 1985). The impacts of haematophagous mites on the health and fitness of geckos are largely unknown (Hanley *et al.* 1995).

*Waddycephalus* are endoparasitic pentastomes that have been detected in many Australian snakes (definitive hosts) and lizards (one of the intermediate hosts) (Riley and Self, 1981; Riley *et al.* 1985; Barton, 2007; Paré, 2008; Kelehear *et al.* 2014b). Most species are described from Australia but the genus is also present in South-east Asia and Fiji (Riley and Self, 1981). *Waddycephalus* have a complex multiple host lifecycle: adults parasitize the lungs of snakes, and the two intermediate hosts are likely coprophagous insects (e.g. cockroaches) and insectivorous lizards, frogs and small mammals (Riley and Self, 1981; Paré, 2008;

Kelehear *et al.* 2014b). *Waddycephalus* nymphs encyst subcutaneously in geckos (Fig. 1A) and may excyst when the host is sick or stressed (Fig. 1B) (Paré, 2008). Infection with nymphal pentastomes can significantly affect the host, with migration and moulting of nymphs being associated with host morbidity (Paré, 2008). *Hemidactylus frenatus* have been recorded to host nymphs of one or more species of *Waddycephalus* in Australia (Barton, 2007; Coates *et al.* 2017). These are assumed to be native *Waddycephalus* given the diversity and prevalence of the genus in Australian reptiles, and given that *Waddycephalus* infection is only seen in bushland populations of *H. frenatus* and not in urban populations (Barton, 2007; Coates *et al.* 2017).

There has been one study of host–parasite dynamics in *H. frenatus* in Australia. Coates *et al.* (2017) assessed the prevalence of *Geckobia* mites (not identified to species), *Waddycephalus* (not identified to species) and *Raillietiella frenata*, an endoparasitic pentastome known to have been introduced with *H. frenatus* into Australia (Barton, 2007). They observed abrupt changes in parasite prevalence across the *H. frenatus* range edge (Coates *et al.* 2017). The prevalence of *Geckobia* mites declined past the urban edge, while *R. frenata* were completely absent outside of inner urban areas. In contrast, native *Waddycephalus* nymphs were found on *H. frenatus* in woodland environments and up to the urban edge (i.e. at the urban–woodland interface), but were absent in inner urban areas (Coates *et al.* 2017). The study concluded that during range expansion from urban to natural areas, *H. frenatus* may experience release from co-evolved parasites (*Geckobia* and *R. frenata*) but are exposed to novel native parasites (*Waddycephalus*).

In the current study, we document the abundance of both native and invasive gecko hosts in bushland habitat and investigate host specificity of mites and *Waddycephalus* where invasive and native geckos co-occur. We then investigate changes in parasite prevalence at the invasion front of *H. frenatus*, and assess factors affecting individual infection probability and intensity. Finally, we investigate the effects of these parasites on *H. frenatus* body condition and draw inferences on impacts on survival.

## Methods

### The parasites

We focused on mites and *Waddycephalus* nymphs in this study because they are visible externally on geckos in the field. Mites



**Fig. 1.** (A) *Waddycephalus* nymphs encysted subcutaneously are visible as protrusions on the mid-body of an adult *Hemidactylus frenatus* (an example is highlighted by the black arrow) and (B) a *Waddycephalus* nymph beginning to excyst after the gecko was captured. Photos: Matthew McIntosh

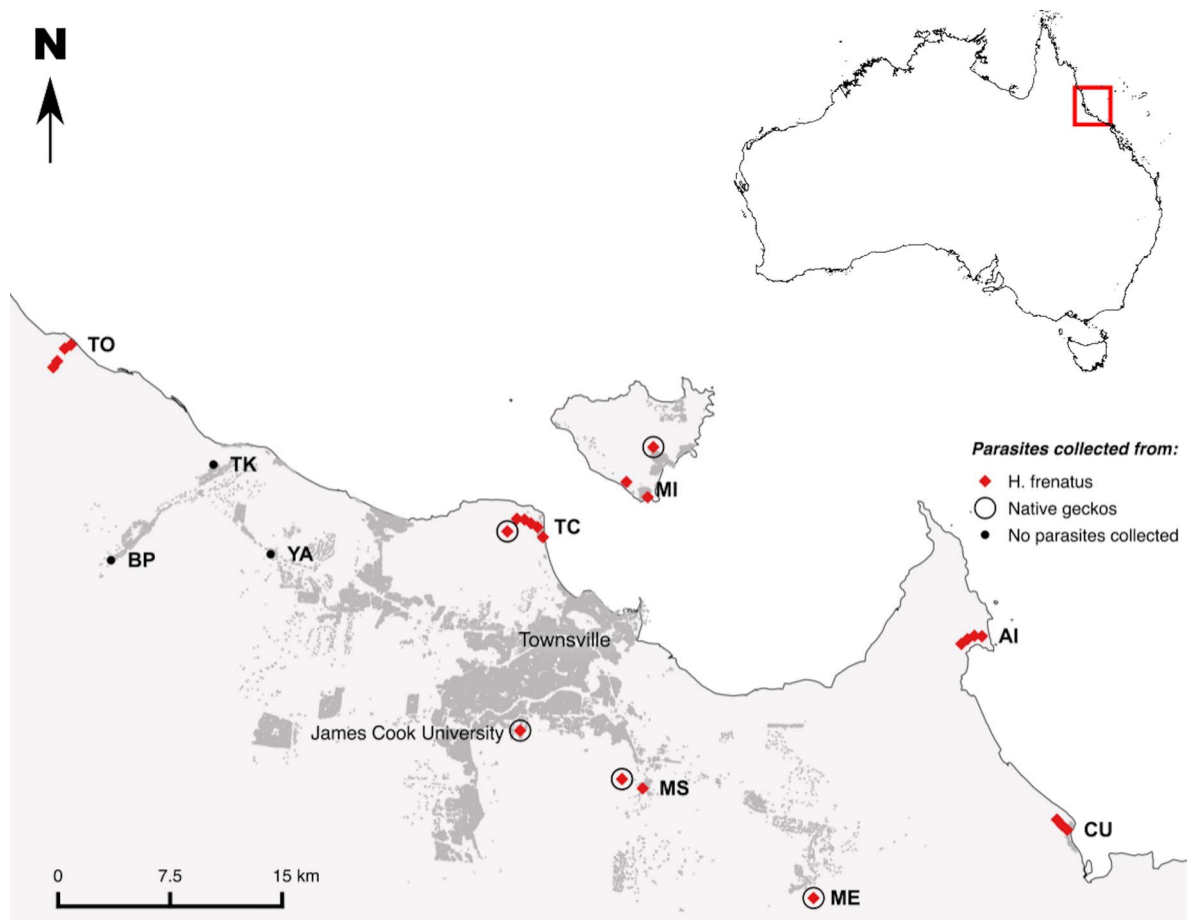
are visible as minute red or orange dots on the skin, often on the feet of geckos (Heath and Whitaker, 2015), while *Waddycephalus* nymphs are visible as small protrusions under the skin of geckos (e.g., Fig. 1A). We dissected three nymphs from one *H. frenatus* specimen to ensure that the protrusions under the skin were *Waddycephalus* nymphs. These nymphs were weighed to determine a representative weight to deduct in tests of host body condition.

### Fieldwork

This study was conducted around the city of Townsville in North-east Australia, where *H. frenatus* have established large populations in dry sclerophyll woodland surrounding urban areas (Barnett *et al.* 2017). We used the same ten transects as Barnett *et al.* (2017), which each consisted of five survey sites, with site 1 on the urban edge and subsequent sites 500 m apart out to Site 5 at 2 km (in a straight line) from the urban edge (Fig. 2). Each site consisted of a surveyed area approximately 200 m long by 15 m wide. The habitat at Site 1 was urban housing and gardens, and the other four sites were in adjacent woodland. Each site was surveyed for *H. frenatus* and native geckos once a month between May 2013 and April 2014, giving a total of 12 surveys per site, except for one site (Yabulu, YA) where ten surveys were conducted. We, therefore, conducted a total of 590 site surveys over 12 months. Sites on the same transect were surveyed on the same night but the order of surveying these sites was randomized for each visit.

At each site, we conducted a 5-min auditory survey, where we counted each time we heard the distinctive ‘chuck chuck chuck...’ vocalization of *H. frenatus*, and a 15-min visual search, where we counted each *H. frenatus* or native gecko we found. During the 15-min visual survey, we walked slowly from a starting point, using head torches to locate geckos by their eye-shine. We also caught up to five *H. frenatus* on each survey to assess external parasites and to take measurements for body condition analyses. These geckos were placed inside a small snap-lock bag and weighed using a 10 g Pesola spring balance, and then measured for snout–vent length (SVL) using a small plastic ruler. Sex was determined by visually checking for testes bulges on males.

We began inspecting the captured *H. frenatus* for parasites during these surveys in September 2013 (the fifth month of surveys) and continued parasite screening through all surveys for the next eight months (a total of 400 surveys). Captured geckos ( $N = 617$ ) were examined visually for *Waddycephalus* nymphs and mites. These mites were not collected for identification to species and therefore in this section, we generalize all mites as ‘*Geckobia*’. The numbers of *Waddycephalus* nymphs and *Geckobia* mites on each gecko were counted. The 15-min survey time was paused while geckos were being measured and examined for parasites. After examination, geckos were released at the point of capture. Parasites were not quantified on native geckos because time and permit constraints meant that native geckos were sighted but not handled during these surveys. However, native geckos were often seen at close proximity and anecdotal observations of *Waddycephalus* nymphs and mites were made.



**Fig. 2.** Map of the Townsville region, showing transects (two-letter codes) where parasite prevalence and intensity data for *Hemidactylus frenatus* was collected. Transects were located at Mount Stuart (MS), Cungulla (CU), Mount Elliott (ME), the Australian Institute of Marine Science (AI), the Town Common (TC), Magnetic Island (MI), Yabulu (YA), Bluewater Park (BP), Toolakea (TK) and Toomulla (TO). *Hemidactylus frenatus* and native geckos occur at all ten transects. Mites were collected for identification at a sub-set of these transects, from either *H. frenatus* (red diamonds) or native geckos (open circles). Additional mites were collected from geckos at a site close to the Magnetic Island (MI) transect and from the James Cook University campus. Buildings are shaded in dark grey

### Targeted mite collection

To assess the potential for host-switching we collected mites from *H. frenatus* ( $N=95$ ) and native geckos ( $N=26$ ). Mite collection was targeted to areas where native geckos and *H. frenatus* co-occur. This included sites at seven of the permanent transects as well as some nearby areas where *H. frenatus* and native geckos coexist (Fig. 2). We also collected mites from geckos at the James Cook University campus (Fig. 2), an additional site where *H. frenatus* and native geckos co-occur on the urban–woodland interface. Mites were collected from the following native gecko species: *Gehyra dubia* ( $N=20$ ), *Amalosa rhombifer* ( $N=4$ ) and *Heteronotia binoei* ( $N=2$ ). All mites were identified morphologically to species by a specialist (AH).

### Statistical analyses

All analyses were conducted in R version 3.1.2 (R Core Team, 2017). All variables were normalized (converted to the same scale) with a mean of zero and a standard deviation of one, which aids in model fitting and the interpretation of results (Zuur *et al.* 2009). We used either linear mixed effects models (LMMs) or generalized linear mixed effects models (GLMMs), within the lme4 package (Bates *et al.* 2015), depending on the error structure of the response variable. To assess which factors affect parasite prevalence and the intensity of parasite infection, we analysed data for *Geckobia* mites and *Waddycephalus* nymphs in separate models (outlined below).

We chose parameters to include in analyses *a priori* based on current ecological knowledge of parasite–host relationships and specific questions we had about this study system (Crawley, 2007).

#### Which factors affect population-level parasite prevalence?

We used parasite prevalence data spanning 8 months, 10 transects and 617 individual geckos to build on the results of Coates *et al.* (2017), who assessed population-level parasite prevalence in the same region over one month, using seven transects and 231 individual geckos. Data for these studies were collected separately, and our approach allowed us to assess the impact of seasonal effects on parasite prevalence.

We calculated the first-principle component of *H. frenatus* seen in visual surveys and *H. frenatus* heard in auditory surveys (using the correlation matrix), and used this as an estimation of relative abundance. This was necessary because environmental conditions affect the detection probability of geckos differently for each survey technique (Barnett *et al.* in preparation), and conditions varied temporally and spatially throughout surveys. Across sites, we found relative abundance and distance to the urban edge were not highly correlated (Pearson's correlation coefficient =  $-0.31$ ), so we included both as independent variables in the following models.

We used GLMMs with binomial error structures and logit link functions (Bolker *et al.* 2009) to assess whether the distance from the urban edge, relative abundance, or season affected: (1) the number of infected/uninfected geckos with respect to *Geckobia* mites per survey, and (2) the number of infected/uninfected geckos with respect to *Waddycephalus* per survey. Seasonal changes were assessed by including a first-order Fourier function: Sine(day of the year) and Cosine(day of the year) as predictors in each model, where 'day of the year' was transformed to fall between 0 and  $2\pi$ . This approach allows a seasonal effect to be fitted as a sinusoidal curve across the course of the year (Stolwijk *et al.* 1999; Cox, 2006), which is suitable for this system because we expect temporal autocorrelation and annual cycles. In both models, we initially included the interaction between the distance from the urban edge and relative abundance but excluded this

parameter in the final model if the interaction was not significant (Crawley, 2007). The transect was included as a random effect in both models to account for broad variation in prevalence between transects.

#### Which factors affect infection probability and infection intensity on individual hosts?

GLMMs with binomial error structures were used to assess whether *H. frenatus* sex or body size (SVL) affected the likelihood of infection with (1) *Geckobia* mites or (2) *Waddycephalus* nymphs. In these analyses, we were less interested in the across the population and temporal effects, so we treated site and survey month as random effects such that our fixed (individual-level) effects were conditioned on the mean prevalence at each site.time.

To assess whether sex or body size (SVL) of individuals affected the intensity of infection with *Geckobia* mites or *Waddycephalus* nymphs we used zero-truncated GLMMs with negative binomial error distributions to account for overdispersion (Zuur *et al.* 2007; Bolker *et al.* 2012). Zero-truncated distributions were necessary because we only included infected individuals in these analyses: intensity of infection could not equal zero. Site and survey month were included as random effects in both models.

#### Do parasites affect body condition?

To investigate whether parasites affect body condition of *H. frenatus* we used LMMs with the natural log of gecko mass as the response variable. Here, we included *Waddycephalus* and *Geckobia* mites in the same models. First, we assessed whether the presence or absence of either parasite affected condition by including the following predictor variables: (1) the natural log of SVL, (2) the presence/absence of *Geckobia* mites and (3) the presence/absence of *Waddycephalus*. We then investigated whether the intensity of infection affected body condition. In this model, the predictor variables were: (1) the natural log of SVL, (2) the intensity of mite infection, and (3) the intensity of *Waddycephalus* infection. For individuals infected with *Waddycephalus*, gecko mass was first corrected by subtracting 0.005 g per nymph: the estimated mean weight of a nymph (calculated from weighing a subset of nymphs dissected out of a gecko). In both body condition analyses, we included site and survey month as random effects.

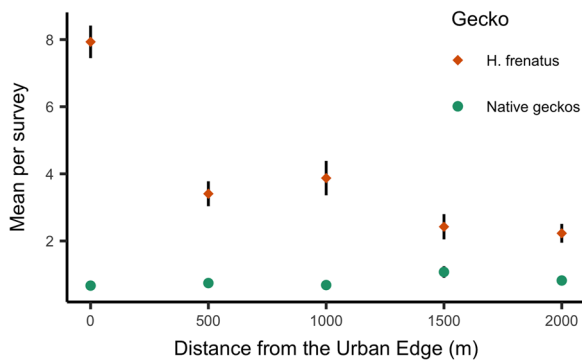
## Results

### Co-occurrence and relative abundance of hosts

*Hemidactylus frenatus* was detected at 46 of the 50 transect sites over the survey period. *Hemidactylus frenatus* was most common at the urban edge sites (site 1) but was present out to the furthest sites (site 5) at 2 km from the urban edge on nine out of ten transects. Native geckos were detected at 47 of the 50 sites, including 43 of the sites where *H. frenatus* was detected. The mean number of *H. frenatus* was higher than native geckos at all distances from the urban edge (all transects combined; Fig. 3), and, summed for all the woodland sites (Site 2–Site 5), *H. frenatus* was much more common than all six native gecko species combined (Fig. 4).

### Host specificity

Targeted mite collections for identification revealed that *H. frenatus* was infected exclusively with the introduced *Geckobia* mites, *G. bataviensis* and *G. keegani* (Table 1). This was the first record of *G. keegani* in Australia. Conversely, only native mite species were collected from three co-occurring native gecko species, *G. dubia*, *A. rhombifer* and *H. binoei* (Table 1). Both *G. dubia* and



**Fig. 3.** The mean number of *Hemidactylus frenatus* and native geckos seen per survey plotted for all transects combined.

*A. rhombifer* were new host records for *G. gymnodactyli*. This was also the first record of *A. rhombifer* hosting *Neotrombicula greenlyi*.

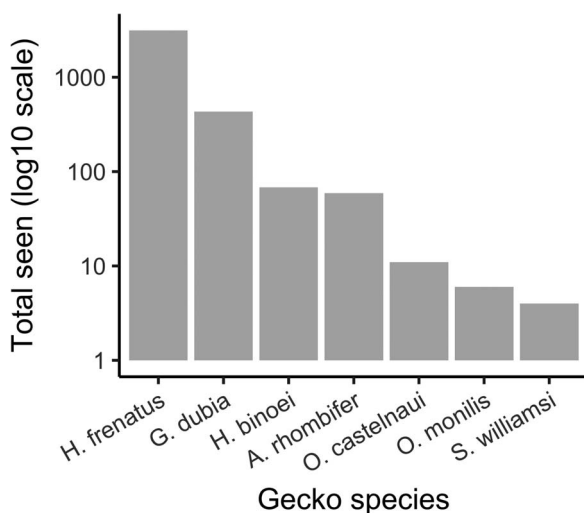
*Waddycephalus* nymphs were not identified to species so it was not possible to assess host specificity in detail. The nymph bulges were also observed multiple times on the native gecko *Gehyra dubia* during the transect surveys (authors, personal observation). Infection of this introduced host is taken to represent host switching, as the *Waddycephalus* found in *H. frenatus* are deemed to be one or more native species (Barton, 2007; Coates et al. 2017).

#### General patterns of parasite infection in *H. frenatus*

Mites were detected at high prevalence on *H. frenatus* on all transects (Table 2), and up to 2 km from the urban edge at five transects (TO, MI, MS, YA, TK). *Waddycephalus* nymphs were detected on *H. frenatus* on eight of the ten transects (Table 2), generally at low prevalence (Table 2). The maximum infection intensity for *Waddycephalus* was ten nymphs per gecko, observed in two individuals.

#### Which factors affect population-level parasite prevalence in *H. frenatus* populations?

Distance from the urban edge significantly affected the prevalence of *Geckobia* mites in *H. frenatus* populations, with lower proportions of infected individuals found farther from the urban edge



**Fig. 4.** The total number of each gecko species (on a log<sub>10</sub> scale) observed in visual surveys at woodland sites (>500 m from the urban edge) throughout the 12-month survey period.

( $P < 0.0001$ ,  $t = -4.53$ , Fig. 5A, Table 3). Relative abundance of *H. frenatus* did not affect the prevalence of mites directly ( $P = 0.59$ ,  $t = -0.54$ ), but there was a negative interaction between distance and abundance, with a steeper cline in mite prevalence in relatively larger host populations ( $P = 0.04$ ,  $t = -2.05$ ; Supplementary Fig. S1). In Supplementary Fig. S1, we show how the interaction between the distance from the urban edge and relative abundance affects *Geckobia* prevalence. Season also significantly affected the prevalence of *Geckobia* mites [Sine(day):  $P < 0.0001$ ,  $t = 3.88$ ; Cosine(day):  $P < 0.0001$ ,  $t = 6.95$ ; Supplementary Fig. S2A], with higher prevalence in the warmer summer months.

Prevalence of *Waddycephalus* nymphs was not affected by distance from the urban edge ( $P = 0.41$ ,  $t = 0.82$ ; Fig. 5B), or by relative abundance ( $P = 0.42$ ,  $t = 0.80$ ). The interaction between these two covariates was non-significant and we therefore dropped it from the model. This had no discernible effect on model fit (AIC without interaction = 181.62, AIC with interaction = 183.60,  $\Delta$ AIC = 1.98). Season significantly affected the prevalence of *Waddycephalus* [Sine(day):  $P = 0.03$ ,  $t = 2.11$ ; Cosine(day):  $P = 0.02$ ,  $t = 2.35$ ; Supplementary Fig. S2B, Table 3], with higher prevalence in the warmer summer months.

#### Which factors affect individual infection?

The likelihood of *Geckobia* mite infection (i.e. presence/absence of mites) was not significantly affected by either the sex ( $P = 0.94$ ,  $t = 0.08$ ) or SVL ( $P = 0.81$ ,  $t = 0.24$  Table 4; Fig. 6A) of geckos. Likelihood of infection with *Waddycephalus* sp. was similarly unaffected by the sex of individuals ( $P = 0.72$ ,  $t = 0.36$ ), but larger geckos were more likely to be infected with at least one *Waddycephalus* nymph ( $P < 0.01$ ,  $t = 2.93$ , Table 4; Fig. 6B).

When looking only at infected individuals, sex affected neither the intensity of mite ( $P = 0.79$ ,  $t = 0.27$ , Table 4), nor *Waddycephalus* infection ( $P = 0.32$ ,  $t = 1.00$ , Table 4). Gecko size positively affected the intensity of mite infection ( $P < 0.01$ ,  $t = 2.83$ , Table 4; Fig. 6C) but negatively affected the intensity of *Waddycephalus* infection, with smaller geckos having more *Waddycephalus* nymphs than larger geckos ( $P = 0.03$ ,  $t = -2.12$ , Table 4; Fig. 6D).

#### Do parasites affect body condition?

Body condition of geckos was not affected by infection with *Geckobia* ( $P = 0.15$ ,  $t = -1.44$ ) or *Waddycephalus* ( $P = 0.94$ ,  $t = -0.07$ ). There was also no significant effect of the intensity of *Geckobia* ( $P = 0.71$ ,  $t = -0.37$ ) or *Waddycephalus* infections on body condition ( $P = 0.29$ ,  $t = 1.07$ ).

## Discussion

#### Host specificity and potential impact

We found large populations of *H. frenatus* out to the furthest sites at 2 km from the urban edge on some transects. Their relative abundance was, on average, over three times higher than co-occurring native gecko species (Figs 3 and 4). This suggests that *H. frenatus* are achieving higher density in natural environments than are native geckos. The presence of comparatively large *H. frenatus* populations in natural environments gives the potential for both parasite spillover and spillback, as well as dilution, to be occurring in this system.

We found no evidence for spillover of mites – the invasive gecko had introduced mites and the native geckos had native mites (Table 1). This was surprising, given the apparently ample opportunity for spillover. *Hemidactylus frenatus* co-occurs with native geckos at many of these sites, and is regularly found side-by-side with natives. Additionally, this invasive species has

**Table 1.** *Geckobia* and *Neotrombicula* mites collected on *Hemidactylus frenatus* and native geckos. *Geckobia* sp. in the final two columns refers to mites that could not be identified to species due to preservation condition

Gecko species	N	<i>G. bataviensis</i>	<i>G. keegani</i>	<i>G. bataviensis</i> & <i>G. keegani</i>	<i>G. gymnodactyli</i>	<i>Neotrombicula greenlyi</i>	<i>Geckobia</i> sp.	
							Adults	Larvae
<i>Hemidactylus frenatus</i>	95	71	13	8	–	–	1	2
<i>Gehyra dubia</i>	20	–	–	–	17	–	3	–
<i>Amalosia rhombifer</i>	4	–	–	–	2	2	–	–
<i>Heteronotia binoei</i>	2	–	–	–	1	–	1	–

been present at some of these sites for more than 20 years (Barnett *et al.* 2017). Prevalence of mites on *H. frenatus* was high, and infected *H. frenatus* were present at most sites (Table 2). The lack of evidence of host switching was unexpected because elsewhere in their introduced range, there are records of *H. frenatus* hosting *Geckobia* mites that are not present in their native range (Heath and Whitaker, 2015). It was particularly unexpected given the relative abundance of *H. frenatus* – spillover from invasive to native species is predicted to be more common in areas where invasive hosts are higher density than native species (Kelehear *et al.* 2014b).

There are three potential reasons for the absence of mite transmission between invasive and native geckos in this system. First, in natural environments, individual *Geckobia* mites may rarely switch hosts. These (Pterygosomatid) mites spend their entire lives on geckos, and may only be transmitted during close and prolonged contact, such as mating or fighting (Bauer *et al.* 1990; Rivera *et al.* 2003). Indeed, Hanley *et al.* (1995) found no evidence of mite transfer between geckos (*Lepidodactylus* spp.) even after keeping them confined together for 48 h (Hanley *et al.* 1995). While mites that have co-evolved with *H. frenatus* (i.e. *G. keegani* and *G. bataviensis*) have previously been recorded infecting other gecko species, including two members of the *Gehyra* genus (*G. oceanica* and *G. mutilata*), competency on these hosts has not been assessed (Heath and Whitaker, 2015). Second, although *H. frenatus* and native geckos co-occur in many of the areas where we collected mites, they may generally occupy different microhabitats or actively avoid encounters with each other. Elsewhere in their invasive range *H. frenatus* exclude native *Nactus* geckos from retreat sites, resulting in fewer encounters between the two species (Cole *et al.* 2005). Third, it is possible

that our sample of mites from native geckos ( $N=26$ ) was too small to detect transmission to native species. However, if that is the case, transmission frequency must be very low given the ample opportunity for transfer outlined above. Laboratory studies would further our understanding of host specificity of the introduced and native mites in this system.

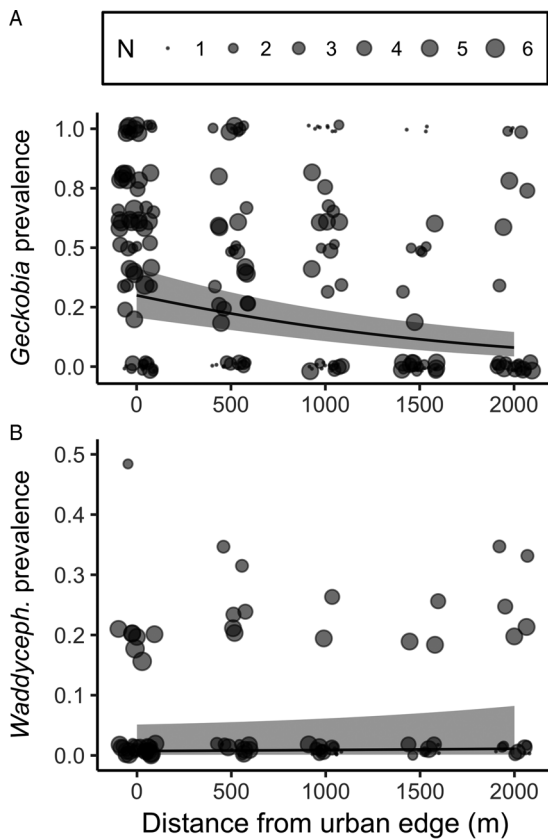
We found *H. frenatus* infected with *Waddycephalus* nymphs on eight of the ten transects (Table 2), and we recorded *Waddycephalus* infections at all distances from the urban edge (Fig. 5B). Prevalence was generally low; a result echoed in the scarcity of *Waddycephalus* records from museum specimens (Barton, 2007), but prevalence also appears low in native geckos in this system (authors pers. obs.). Despite this low prevalence, the high abundance of *H. frenatus* may increase the prevalence of *Waddycephalus* in the final hosts (native snakes) and in turn other intermediate hosts (coprophagous insects and native geckos). We witnessed native snakes (*Boiga irregularis* and *Morelia spilota*) preying on *H. frenatus* on multiple occasions during the study (authors, pers. obs.). It seems likely, therefore, that *H. frenatus* provide the same trophic link as native geckos, enabling *Waddycephalus* to complete its life cycle and potentially increasing its prevalence in native hosts through parasite spillover.

#### Do parasites affect the range expansion of *H. frenatus*?

Prevalence of mites in *H. frenatus* populations decreased with distance from the urban edge, which is consistent with Coates *et al.* (2017). There was also a significant negative interaction between relative abundance and distance, with larger *H. frenatus* populations having steeper declines in mite prevalence with distance from the urban edge (Supplementary Fig. S1). This pattern is

**Table 2.** Prevalence of parasites and mean intensity of infection in *Hemidactylus frenatus* assessed during transect surveys in 2013–2014

Transect	LAT	LONG	<i>H. frenatus</i> inspected	<i>Geckobia</i> mites			<i>Waddycephalus</i> nymphs		
				Prevalence	Mean Intensity	Sites detected	Prevalence	Mean Intensity	Sites detected
Town Common (TC)	–19.20	146.77	108	0.37	13.18	All sites	0.07	3.13	All sites
AIMS (AI)	–19.27	147.05	88	0.30	10.92	1,2,3,4	0.06	1.20	1,3,4,5
Mount Stuart (MS)	–19.36	146.84	78	0.51	10.60	All sites	0.06	1.80	1,2,3,5
Cungulla (CU)	–19.39	147.11	69	0.48	12.36	1,2,3,4	0.03	2.00	2,3
Magnetic Island (MI)	–19.17	146.84	102	0.43	10.23	All sites	0.02	3.00	2,3
Toomulla (TO)	–19.08	146.47	75	0.47	10.66	1,2,3,5	0.09	4.14	2,3,5
Mount Elliot (ME)	–19.43	146.95	33	0.42	7.07	1	0.03	1.00	1
Yabulu (YA)	–19.13	146.36	28	0.43	9.75	1,4,5	0.14	3.25	1,3,4,5
Bluewater Park (BP)	–19.23	146.48	22	0.41	6.67	1,3	0.00	0.00	–
Toolakea (TK)	–19.16	146.56	10	0.60	10.33	1,5	0.00	0.00	–



**Fig. 5.** (A) The prevalence of *Geckobia* mite infection in *Hemidactylus frenatus* per survey with distance from the urban edge. (B) The prevalence of *Waddycephalus* sp. infection with distance from the urban edge. Size of the circles indicates sample size.

driven by both higher prevalence at the urban edge and lower prevalence deeper into bushland. This is an intriguing pattern that requires further investigation. Higher density bushland populations of *H. frenatus* are generally also those that have been in the bushland the longest (Barnett *et al.* 2017). Therefore, a potential explanation is that parasite release occurs when *H. frenatus* establishes in bushland at low density and then as populations increase rapidly to high density, mite increase lags behind. Another explanation could be differences in behaviour between urban and woodland geckos. If, for example, woodland geckos move

more and are less territorial than urban geckos, they may be less likely to acquire mites (especially if this difference is magnified at high densities).

The prevalence of *Geckobia* mites and *Waddycephalus* nymphs was also affected by season, with higher prevalence in the warmer summer months (i.e. at the start and end of the year; Supplementary Fig. S2). This could be explained by a greater proportion of juvenile geckos being found towards the middle of the year (Supplementary Fig. S2), as larger geckos are more likely to be infected with *Waddycephalus*. However, for *Geckobia* mites, there was no relationship between individual-level prevalence and gecko size (discussed below). Therefore, seasonal patterns in prevalence may be driven by increased contact between geckos during the peak breeding season (i.e. summer in north Queensland; Barnett *et al.* 2017), which may lead to higher mite transmission rates. Future work should address the effects of season on these parasites more thoroughly by assessing prevalence over the entire year, as our data spanned only 8 months.

In terms of individual-level determinants of infection, we found that larger (and hence generally older) *H. frenatus* had higher intensity mite infections, suggesting that geckos steadily acquire mites through life. There was, however, no detectible impact of mite presence, or intensity of mite infection, on host body condition. Haematophagous mites can cause ulcerative dermatitis and inhibit skin sloughing in other lizards (Goldberg and Bursey, 1991; Goldberg and Holshuh, 1992; Walter and Shaw, 2002), but to date there is no evidence of gecko mites affecting the condition in wild populations (Hanley *et al.* 1995). Here, we found significantly more mites on larger geckos. If mites affected the lifespan of geckos, one would expect to see fewer highly infected large (i.e. older) individuals. Together with the lack of effect on body condition, our results suggest that the impact of mites on host fitness may be negligible. It is, therefore, unlikely that release from mites increases the rate of range expansion in this system, despite an apparently lower prevalence of mites on the invasion front.

The prevalence of *Waddycephalus* was not affected by distance from the urban edge or relative abundance of *H. frenatus*. Predictions regarding the prevalence of *Waddycephalus* in *H. frenatus* are complicated by the fact that *Waddycephalus* have a complex lifecycle, and so are limited by the abundance of their final hosts (native snakes) and their first intermediate hosts (likely coprophagous insects) (Ali and Riley, 1983; Kelehear *et al.* 2014a). The *Waddycephalus* that infect *H. frenatus* are most likely a native species (or multiple native species) and *H. frenatus* are infected at

**Table 3.** Factors that affect prevalence of (A) *Geckobia* mites, and (B) *Waddycephalus* nymphs in *Hemidactylus frenatus* populations

Model	Parameter	Estimate	Standard Error	t	P
(A) Mite prevalence	Distance from urban edge***	-0.56	0.12	-4.53	<0.0001
	Relative abundance	-0.05	0.10	0.54	0.59
	Sine (day)***	0.82	0.22	3.80	<0.0001
	Cosine (day)***	1.03	0.15	6.90	<0.0001
	Distance.abundance*	0.20	0.10	2.05	0.04
(B) Waddycephalus prevalence	Distance from urban edge	0.15	0.18	0.82	0.41
	Relative abundance	0.15	0.18	0.80	0.42
	Sine (day)*	1.60	0.76	2.11	0.03
	Cosine (day)*	1.52	0.65	2.35	0.02

\* $P < 0.05$ .

\*\*\* $P < 0.0001$ .

**Table 4.** Factors that affect the presence and infection intensity of *Geckobia* mites [models (A) and (B)] and *Waddycephalus* nymphs [models (C) and (D)] in individual *Hemidactylus frenatus*

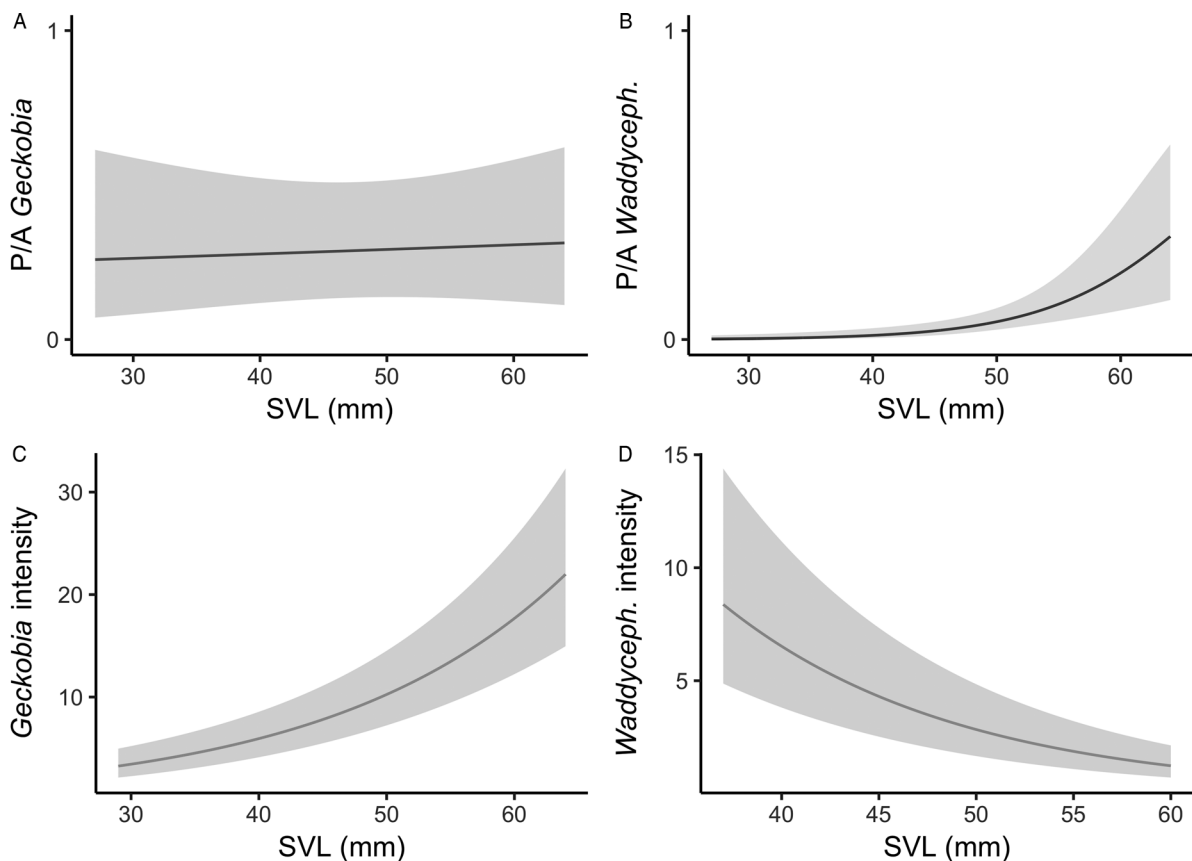
Parasite	Model	Parameter	Estimate	Std. error	<i>t</i>	<i>P</i>
<i>Geckobia</i> mites	(A) Presence/absence	SVL	0.05	0.19	0.24	0.81
		Sex (male)	0.02	0.27	0.08	0.94
	(B) Infection intensity (zero-truncated)	SVL**	0.28	0.10	2.83	<0.01
		Sex (male)	0.04	0.16	0.27	0.79
<i>Waddycephalus</i>	(C) Presence/absence	SVL**	0.99	0.34	2.93	<0.01
		Sex (male)	0.19	0.54	0.36	0.72
	(D) Infection intensity (zero-truncated)	SVL*	-0.56	0.26	-2.12	0.03
		Sex (male)	0.48	0.48	1.00	0.32

\**P* < 0.05.\*\**P* < 0.01.

low prevalence at most of our woodland sites. In contrast, the parasite has been found to be completely absent in adjacent inner urban sites, probably due to smaller snake populations (Coates *et al.* 2017).

Due to their presence in natural areas but absence in inner urban environments, *Waddycephalus* could potentially affect range expansion of *H. frenatus* in natural habitats if infection impacts body condition or survival. We found no effect of presence or intensity of *Waddycephalus* nymphs on body condition of *H. frenatus*. In infected individuals, however, smaller geckos had a greater intensity of infection than larger geckos (Table 4). One

explanation for this pattern is that high-intensity *Waddycephalus* infection increases mortality with time, so adult geckos with high-intensity infection are missing from the population. While data on the effects of nymphal pentastomes on *H. frenatus* are lacking, they can kill reptile hosts when they migrate or moult, particularly when they burrow from the stomach to the body wall (Paré, 2008), or affect the likelihood of predation through changes to host behaviour (Lefèvre *et al.* 2009). Another explanation is that some nymphs are lost through time, either because they excyst or because adult geckos can somehow shed them. Future studies should further explore the impact of *Waddycephalus* nymphs on geckos.



**Fig. 6.** The predicted relationship ( $\pm$  standard error) between the snout-vent length of *Hemidactylus frenatus* and the presence/absence of (A) *Geckobia* mites and (B) *Waddycephalus* nymphs; and intensity of infection with (C) *Geckobia* mites and (D) *Waddycephalus* nymphs, from our generalized linear mixed effects models. In these models we incorporated Site and Survey Number as random effects such that the fixed (individual-level) effects shown here are conditioned on the mean prevalence/intensity at each site.time.



### Concluding remark

This study examined the potential impacts of an invasive gecko on native species through parasite transmission and the potential effect of parasites on range expansion of the invasive gecko. We found no evidence of invasive mites infecting native geckos in this system, or *vice versa*, despite apparently ample opportunity in terms of time and fine-scale co-occurrence. *Hemidactylus frenatus* are, however, susceptible to infection by native *Waddycephalus* nymphs, and high-intensity infections may reduce the survival of individual *H. frenatus*. We explored the complex interactions between parasites and range expansion of this invasive gecko and found that range expansion into natural environments means both release from co-evolved mites and exposure to novel *Waddycephalus* nymphs. The relatively high density of *H. frenatus* makes parasite spillback (of *Waddycephalus*) to native host species a concern. Future work should investigate parasite spillback by assessing whether the prevalence of *Waddycephalus* nymphs in native gecko and snake populations is higher where they co-occur with *H. frenatus*.

**Supplementary material.** The supplementary material for this article can be found at <https://doi.org/10.1017/S003118201800015X>

**Financial Support.** This research received no specific grant from any funding agency, commercial or not-for-profit sectors.

### References

- Ali JH and Riley J (1983) Experimental life-cycle studies of *Raillietiella gehyrae* Bovien, 1927 and *Raillietiella frenatus* Ali, Riley and Self, 1981: pentastomid parasites of geckos utilizing insects as intermediate hosts. *Parasitology* **86**, 147–160.
- Andreou D, Arkush KD, Guégan J-F and Gozlan RE (2012) Introduced pathogens and native freshwater biodiversity: a case study of *Sphaerothecum destruens*. *PLoS ONE* **7**, e36998.
- Barnett LK, Phillips BL and Hoskin CJ (2017). Going feral: time and propagule pressure determine range expansion of Asian house geckos into natural environments. *Austral Ecology* (in press) **42**, 165–175
- Barton DP (2007) Pentastomid parasites of the introduced Asian house gecko, *Hemidactylus frenatus* (Gekkonidae), in Australia. *Comparative Parasitology* **74**, 254–259.
- Bates D, Mächler M, Bolker B and Walker S (2015) Fitting linear mixed-effects models using lme4. *Journal of Statistical Software* **67**, 1–48.
- Bauer AM, Russell AP and Dollahan NR (1990) Skin folds in the gekkonid lizard genus *Rhacodactylus*: a natural test of the damage limitation hypothesis of mite pocket function. *Canadian Journal of Zoology* **68**, 1196–1201.
- Bolker BM, Brooks ME, Clark CJ, Geange SW, Poulsen JR, Stevens MHH and White J-SS (2009) Generalized linear mixed models: a practical guide for ecology and evolution. *Trends in Ecology & Evolution* **24**, 127–135.
- Bolker B, Brooks M, Gardner B, Lennert C and Minami M (2012). Owls example: a zero-inflated, generalized linear mixed model for count data. 1–35. Available at <https://groups.nceas.ucsb.edu/non-linear-modeling/projects/owls/WRITUEUP/owls.pdf>
- Case TJ and Taper ML (2000) Competition, environmental gradients, gene flow and the coevolution of species' borders. *The American Naturalist* **155**, 583–605.
- Coates A, Barnett LK, Hoskin C and Phillips BL (2017) Living on the edge: parasite prevalence changes dramatically across a range edge in an invasive gecko. *The American Naturalist* **189**, 178–183.
- Cole NC, Jones CG and Harris S (2005) The need for enemy-free space: the impact of an invasive gecko on island endemics. *Biological Conservation* **125**, 467–474.
- Cox NJ (2006) Speaking Stata: in praise of trigonometric predictors. *Stata Journal* **6**, 561–579.
- Crawley MJ (2007). *The R Book*, 1st edn. Imperial College London at Silkwood Park, UK: John Wiley & Sons Ltd.
- Dame EA and Petren K (2006) Behavioural mechanisms of invasion and displacement in Pacific island geckos (Hemidactylus). *Animal Behaviour* **71**, 1165–1173.
- Domrow R (1992) Acari Astigmata (excluding feather mites) parasitic on Australian vertebrates: an annotated checklist, keys and bibliography. *Invertebrate Systematics* **6**, 1459.
- Domrow R and Lester LN (1985) Chiggers of Australia (Acari: Trombiculidae): an annotated checklist, keys and bibliography. *Australian Journal of Zoology* **114**, 1–111.
- Dunn AM (2009) *Parasites and Biological Invasions*, 1st edn. London, UK: Elsevier Ltd. doi:10.1016/S0065-308X(08)00607-6.
- Dunn AM and Hatcher MJ (2015) Parasites and biological invasions: parallels, interactions, and control. *Trends in Parasitology* **31**, 189–199.
- Dunn AM, Torchin ME, Hatcher MJ, Kotanen PM, Blumenthal DM, Byers JE, Coon CAC, Frankel VM, Holt RD, Hufbauer RA, Kanarek AR, Schierenbeck KA, Wolfe LM and Perkins SE (2012) Indirect effects of parasites in invasions. *Functional Ecology* **26**, 1262–1274.
- Floerl O, Inglis GJ, Dey K and Smith A (2009) The importance of transport hubs in stepping-stone invasions. *Journal of Applied Ecology* **46**, 37–45.
- Gendron AD, Marcogliese DJ and Thomas M (2012) Invasive species are less parasitized than native competitors, but for how long? The case of the round goby in the Great Lakes-St. Lawrence Basin. *Biological Invasions* **14**, 367–384.
- Goldberg SR and Bursey CR (1991) Integumental lesions caused by ectoparasites in a wild population of the side-blotched lizard (*Uta stansburiana*). *Journal of Wildlife Diseases* **27**, 68–73.
- Goldberg SR and Holshuh HJ (1992) Ectoparasite-induced lesions in mite pockets of the Yarrow's spiny lizard, *Sceloporus jarrovi* (Phrynosomatidae). *Journal of Wildlife Diseases* **28**, 537–541.
- Hanley KA, Fisher RN and Case TJ (1995) Lower mite infestations in an asexual gecko compared with its sexual ancestors. *Evolution* **49**, 418–426.
- Hartigan A, Fiala I, Dyková I, Jirků M, Okimoto B, Rose K, Phalen DN and Šlapeta J (2011) A suspected parasite spill-back of two novel *Myxidium* spp. (Myxosporae) causing disease in Australian endemic frogs found in the invasive Cane toad. *PLoS ONE* **6**, e18871.
- Heath ACG and Whitaker AH (2015) Mites (Acari: Pterygosomatidae, Macronyssidae) taken from lizards intercepted at the New Zealand border. *Systematic and Applied Acarology* **20**, 739.
- Hoskin CJ, (2011) The invasion and potential impact of the Asian House Gecko (*Hemidactylus frenatus*) in Australia. *Austral Ecology* **36**, 240–251.
- Hudson P and Greenman J (1998) Competition mediated by parasites: biological and theoretical progress. *Trends in Ecology and Evolution* **13**, 387–390.
- Kelehear C, Brown GP and Shine R (2013). Invasive parasites in multiple invasive hosts: the arrival of a new host revives a stalled prior parasite invasion. *Oikos* **122**, 1317–1324.
- Kelehear C, Spratt DM, O'Meally D and Shine R (2014a) Pentastomids of wild snakes in the Australian tropics. *International Journal for Parasitology: Parasites and Wildlife* **3**, 20–31.
- Kelehear C, Saltonstall K and Torchin ME (2014b) An introduced pentastomid parasite (*Raillietiella frenata*) infects native cane toads (*Rhinella marina*) in Panama. *Parasitology* **142**, 675–679.
- Kelly DW, Paterson RA, Townsend CR, Poulin R and Tompkins DM (2009) Parasite spillback: a neglected concept in invasion ecology. *Ecology* **90**, 2047–2056.
- Krakau M, Thielges DW and Reise K (2006) Native parasites adopt introduced bivalves of the North Sea. *Biological Invasions* **8**, 919–925.
- Lefèvre T, Lebarbenchon C, Gauthier-Clerc M, Missé D, Poulin R and Thomas F (2009) The ecological significance of manipulative parasites. *Trends in Ecology and Evolution* **24**, 41–48.
- Lever C (2003) *Naturalized Reptiles and Amphibians of the World*. New York: Oxford University Press.
- MacLeod CJ, Paterson AM, Tompkins DM and Duncan RP (2010) Parasites lost – do invaders miss the boat or drown on arrival? *Ecology Letters* **13**, 516–527.
- Menge BA (1972) Competition for food between two intertidal starfish species and its effect on body size and feeding. *Ecology* **53**, 635–644.
- Miller MA, Kinsella JM, Snow RW, Hayes MM, Falk BG, Reed RN, Mazzotti FJ, Guyer C and Romagosa CM (2017) Parasite spillover: indirect effects of invasive Burmese pythons. *Ecology and Evolution* **8**, 830–840.
- O'Dowd DJ, Green PT and Lake PS (2003) Invasional “meltdown” on an oceanic island. *Ecology Letters* **6**, 812–817.
- Paré JA (2008) An overview of pentastomiasis in reptiles and other vertebrates. *Journal of Exotic Pet Medicine* **17**, 285–294.

- Perkins TA** (2012) Evolutionarily labile species interactions and spatial spread of invasive species. *The American Naturalist* **179**, E37–E54.
- Perkins SE, White TA, Pascoe EL and Gillingham EL** (2017) Parasite community dynamics in an invasive vole – from focal introduction to wave front. *International Journal for Parasitology: Parasites and Wildlife* **6**, 412–419.
- Phillips BL, Kelehear C, Pizzatto L, Brown GP, Barton D and Shine R** (2010) Parasites and pathogens lag behind their host during periods of host range advance. *Ecology* **91**, 872–881.
- Prenter J, Macneil C, Dick JTA and Dunn AM** (2004) Roles of parasites in animal invasions. *Trends in Ecology and Evolution* **19**, 385–390.
- R Core Team** (2017). *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing.
- Riley J, and Self J**, (1981) A redescription of *Waddycephalus teretiusculus* (Baird, 1862) Sambon, 1922 and a revision of the taxonomy of the genus *Waddycephalus* (Sambon, 1922), pentastomid parasites of Asian, Australian and Indonesian snakes, with descriptions of eight new species. *Systematic Parasitology* **3**, 243–257.
- Riley J, Spratt DM, and Presidente PJA**, (1985) Pentastomids (Arthropoda) parasitic in Australian reptiles and mammals. *Aust. J. Zool.* **33**, 39–54.
- Rivera CCM, Negrón AG and Bertrand M** (2003) *Hemidactylus mabouia* (Sauria: Gekkonidae), host of *Geckobia hemidactyli* (Actiniedida: Pterygosomatidae), throughout the Caribbean and South America. *Caribbean Journal of Science* **39**, 321–326.
- Stolwijk AM, Straatman H and Zielhuis GA** (1999) Studying seasonality by using sine and cosine functions in regression analysis. *Journal of Epidemiology and Community Health* **53**, 235–238.
- Telfer S and Bown K** (2012) The effects of invasion on parasite dynamics and communities. *Functional Ecology* **26**, 1288–1299.
- Torchin ME and Mitchell CE** (2004) Parasites, pathogens, and invasions by plants and animals. *Frontiers in Ecology and the Environment* **2**, 183–190.
- Torchin ME, Lafferty KD, Dobson AP, McKenzie VJ and Kuris AM** (2003) Introduced species and their missing parasites. *Nature* **421**, 628–630.
- Vanderduys EP and Kutt AS** (2013) Is the Asian house gecko, *Hemidactylus frenatus*, really a threat to Australia's biodiversity? *Australian Journal of Zoology* **60**, 361–367.
- Walter DE and Shaw M** (2002) First record of the mite *Hirstiella diolii* Baker (Prostigmata: Pterygosomatidae) from Australia, with a review of mites found on Australian lizards. *Australian Journal of Entomology* **41**, 30–34.
- Wanless RM, Angel A, Cuthbert RJ, Hilton GM and Ryan PG** (2007) Can predation by invasive mice drive seabird extinctions? *Biology Letters* **3**, 241–244.
- Zuur AF, Ieno EN and Smith GM** (2007) *Analyzing Ecological Data in R*. New York, USA: Springer.
- Zuur AF, Ieno EN, Walker N, Saveliev AA and Smith GM** (2009). *Mixed Effects Models and Extensions in Ecology with R*. New York, NY: Springer.