

Association of selenium and copper with lipids in umbilical cord blood

E. M. Wells^{1*}, A. Navas-Acien^{2,3}, B. J. Apelberg⁴, J. B. Herbstman⁵, J. M. Jarrett⁶, Y. H. Lin⁷, C. P. Verdon⁶, C. Ward⁶, K. L. Caldwell⁶, J. R. Hibbeln⁷, R. U. Halden^{2,8}, F. R. Witter⁹ and L. R. Goldman¹⁰

¹School of Health Sciences, Purdue University, West Lafayette, IN, USA

²Department of Environmental Health Sciences, Johns Hopkins Bloomberg School of Public Health, Baltimore, MD, USA

³Department of Epidemiology, Johns Hopkins Bloomberg School of Public Health, Baltimore, MD, USA

⁴Office of Policy, Center for Tobacco Products, Food and Drug Administration, Rockville, MD, USA

⁵Columbia Center for Children's Environmental Health, Columbia University Mailman School of Public Health, New York, NY, USA

⁶National Center for Environmental Health, Centers for Disease Control and Prevention, Division of Laboratory Sciences, Atlanta, GA, USA

⁷National Institute of Alcohol Abuse and Alcoholism, National Institutes of Health, Rockville, MD, USA

⁸Center for Environmental Security, Biodesign Institute, Arizona State University, Tempe, AZ, USA

⁹Department of Gynecology and Obstetrics, Johns Hopkins University School of Medicine, Baltimore, MD, USA

¹⁰George Washington University School of Public Health and Health Services, Washington, DC, USA

Altered levels of selenium and copper have been linked with altered cardiovascular disease risk factors including changes in blood triglyceride and cholesterol levels. However, it is unclear whether this can be observed prenatally. This cross-sectional study includes 274 singleton births from 2004 to 2005 in Baltimore, Maryland. We measured umbilical cord serum selenium and copper using inductively coupled plasma mass spectrometry. We evaluated exposure levels vis-à-vis umbilical cord serum triglyceride and total cholesterol concentrations in multivariable regression models adjusted for gestational age, birth weight, maternal age, race, parity, smoking, prepregnancy body mass index, n-3 fatty acids and methyl mercury. The percent difference in triglycerides comparing those in the highest *v.* lowest quartile of selenium was 22.3% (95% confidence interval (CI): 7.1, 39.7). For copper this was 43.8% (95% CI: 25.9, 64.3). In multivariable models including both copper and selenium as covariates, copper, but not selenium, maintained a statistically significant association with increased triglycerides (percent difference: 40.7%, 95% CI: 22.1, 62.1). There was limited evidence of a relationship of increasing selenium with increasing total cholesterol. Our findings provide evidence that higher serum copper levels are associated with higher serum triglycerides in newborns, but should be confirmed in larger studies.

Received 14 November 2013; Revised 3 March 2014; Accepted 20 March 2014; First published online 22 April 2014

Key words: copper, infant, newborn, selenium, total cholesterol, triglyceride

Introduction

Triglycerides and total cholesterol are risk factors for future cardiovascular disease. Even among children and young adults, these risk factors have been associated with the development of atherosclerosis in both cross sectional^{1,2} and longitudinal^{3,4} studies. Improved understanding of determinants of elevated serum triglycerides and cholesterol could provide additional opportunities to prevent cardiovascular disease.

Selenium and copper are essential trace elements. Selenium, as part of selenoproteins, plays a role in protection against oxidative stress, thyroid function and immune function.⁵ Copper, incorporated into cuproenzymes, plays a role in heme and iron metabolism, immune function, and glucose and cholesterol metabolism.^{6,7} Although required for good health, either very low or very high concentrations may result in poor health outcomes, including cardiovascular disease.^{8–13}

Elevated selenium concentrations have been linked with elevated cholesterol and triglyceride concentrations.^{14–17} A few studies have found positive associations between copper and serum lipids;^{18,19} including a cross-sectional correlation of copper with triglyceride among hyperlipidemic Iraqi men.²⁰ Other studies, however, have found inverse relationships.^{21,22} Evidence in newborns is very limited: a study of selenium supplementation during pregnancy found higher cord blood triglyceride concentrations among those taking supplements,²³ and a cross-sectional study found umbilical cord serum copper concentrations were positively associated with triglycerides, but not with total or low-density lipoprotein (LDL) cholesterol.²⁴

In prior work, we showed a strong correlation between umbilical cord copper and selenium;²⁵ however, previous studies have only evaluated these elements in isolation instead of simultaneously. The goal of this study was to determine the joint association between umbilical cord serum selenium and copper concentrations with umbilical cord serum triglycerides and total cholesterol in a population of newborns from Baltimore, Maryland.

*Address for correspondence: E. M. Wells, School of Health Sciences, Purdue University, 550 Stadium Mall Drive, HAMP 1269, West Lafayette, IN 47907, USA.
(Email: wells54@purdue.edu)

Methods

Study design

We obtained umbilical cord blood samples and medical record data from 300 singleton births between November 2004 and March 2005 at the Johns Hopkins Hospital in Baltimore, Maryland as part of the Baltimore Tracking Health Related to Environmental Exposures Study. This study was approved by the Johns Hopkins Medicine Institutional Review Board and was determined to be exempt from the Health Insurance Portability and Accountability Act. More details about the study were described previously.^{25,26}

There were 591 singleton births during the data collection period. Study staff collected 341 umbilical cord blood samples, of which only 300 had sufficient sample for laboratory analyses. Twenty-five births were excluded due to missing data for triglycerides and cholesterol ($n = 4$), missing data for selenium and copper ($n = 12$), missing data for n-3 fatty acids ($n = 9$). A subject with outlier values for total cholesterol, triglycerides and copper was also omitted, leaving a total of 274 births.

Data collection

Procedures for this study have been described previously.^{25,26} Briefly, trained hospital staff collected umbilical cord blood from the umbilical cord vein immediately following delivery. Cord blood was stored for <3 h at 4°C, then study staff separated samples into whole blood (for methyl mercury) and serum (for other analytes) and transferred samples to 2 mm polypropylene tubes prescreened for metals. Samples were stored at -80°C and were shipped on dry ice. Laboratory analysis of umbilical cord blood was conducted at the United States Centers for Disease Control and Prevention (selenium, copper, methyl mercury, triglycerides, total cholesterol and cotinine) and the United States National Institute on Alcohol Abuse and Alcoholism (n-3 fatty acids).

Selenium and copper were measured in umbilical cord serum with inductively coupled plasma dynamic reaction cell mass spectrometry (ICP-DRC-MS). Methyl mercury was measured in whole blood using high performance liquid chromatography coupled with ICP-DRC-MS. Limits of detection (LOD) were 5 µg/l (selenium), 4 µg/dl (copper) and 0.48 µg/l (methyl mercury). A total of 48 methyl mercury measurements below the LOD were replaced with LOD/√2 for analyses.²⁷

Triglycerides and total cholesterol were measured in umbilical cord serum using the same protocol as the CDC's National Health and Nutrition Examination Survey: an automated timed-endpoint method that marks breakdown products of the lipid and measures their quantity via wavelength absorbance of the marked product.

Cotinine was measured using liquid chromatography in conjunction with atmospheric pressure ionization mass spectrometry (LOD = 0.015 ng/ml); 26% of samples were below

the LOD. Cord serum cotinine and information from the medical record were used to determine smoking status, as described below. Fatty acids in umbilical cord serum were measured using fast gas chromatography in an automated system featuring a robotic apparatus.^{28,29} The ratio of n-3 highly unsaturated fatty acids (HUFAs)/total HUFAs was used in statistical analyses.

Gestational age, birth weight, maternal age, prepregnancy body mass index (BMI) and self-reported smoking were obtained from infant and maternal medical records; a random 10% sample was checked in duplicate for accuracy. Prepregnancy BMI was calculated from the most recent maternal height and weight before pregnancy from the maternal medical record. For mothers with missing height ($n = 3$), prepregnancy weight ($n = 7$), or both ($n = 1$), we imputed these data using multivariable regression models that controlled for placental weight, weight gain during pregnancy, race, education, height and prepregnancy weight.³⁰ We defined smoking as a report of smoking during pregnancy in the medical record or umbilical cord serum cotinine >10 ng/ml.

Data analyses

Statistical analyses were completed using Stata 11.2 (College Station, TX). Triglycerides, total cholesterol, mercury, and copper all had log-normal distributions. Therefore, these were described using geometric means and natural-log transformed for analyses, where appropriate. Linear regression, Student's *t*-tests, one-way ANOVAs and nonparametric smoothing (lowess) curves were used for bivariate analyses. Quartiles were used for exposure variables in regression analyses. Linear regression models on log-transformed triglyceride or total cholesterol concentrations were used to estimate the percent difference of lipid concentrations comparing quartiles 2–4 *v.* the lowest quartile of selenium or copper.

Relevant confounders were identified based on previous work,^{14,18,21,25} as well as exploratory data analysis within this data set. Methyl mercury and n-3 fatty acids were included as potential confounders for effects of selenium as they share an exposure source (fish or seafood consumption), there are some indications that they affect cardiovascular health, and they may also affect each other.^{31,32}

Quadratic terms were used for gestational age and maternal prepregnancy BMI based on nonparametric smoothing curves of these variables with the outcomes. Triglyceride concentrations can be dependent on fasting. While fasting duration before delivery was not available, women are not allowed to eat during delivery and our results remained unchanged after further adjustment for duration of labor.

We modeled metal exposures in relation to blood lipids using the following sets of potential confounding variables: (1) gestational age; (2) further adjusted for maternal age, maternal race, parity, maternal smoking during pregnancy, prepregnancy BMI, n-3 HUFAs/total HUFAs and methyl mercury; and (3) further adjusted for either selenium or copper.

Results

The study population is described in Table 1. Mothers were, on average, 25.8 (95% confidence interval (CI): 25.0, 26.6) years old and 20.4% were under 20. There were 70.8% African Americans, 25.1% Caucasians and 7.7% Asian Americans. Before pregnancy, 24.1% of mothers were overweight and 24.1% were obese.

Geometric mean umbilical cord serum triglyceride was 34.5 mg/dl (95% CI: 32.9, 36.3) with a range of 13.3–110.4 mg/dl. Geometric mean total cholesterol was 64.6 mg/dl (95% CI: 62.5–66.7) with a range of 30.1–145.6. In bivariate analyses, higher cord blood serum triglyceride concentrations were associated with higher gestational age, prepregnancy BMI, n-3 HUFA/total HUFAs, primiparity, selenium, and copper. Higher total cholesterol concentrations were associated with lower gestational age, birth weight and n-3 HUFA/total HUFA ratio. Results were similar for the relationship between n-3 HUFA/total HUFA and selenium or cholesterol when the fatty acid ratio was included in multivariable models (data not shown).

After adjustment for gestational age, birth weight, maternal age and race, primiparity, prepregnancy BMI, smoking, n-3 HUFAs/total HUFAs and methyl mercury, umbilical cord triglyceride concentrations increased with increasing cord blood selenium or copper quartiles (Table 2, model 2). However, in models which also included both selenium and copper, only copper was associated with significantly higher triglycerides (Table 2, model 3).

There was a significant trend in adjusted models for selenium and total cholesterol (Table 3, model 2); however none of

the associations for selenium and total cholesterol by quartile of selenium were significant and the trend was not significant after adjustment for copper. There was also a borderline significant test for trend for the association of copper with total cholesterol in an adjusted model (Table 3, model 2), but not after adjustment for selenium (Table 3, model 3).

Discussion

This study shows a positive association of selenium and copper with triglycerides in umbilical cord serum. However, in models including both selenium and copper, only the association between copper and triglycerides remained statistically significant. For total cholesterol, we see a significant or borderline significant ($P < 0.10$) test for trend in adjusted models for selenium and copper, respectively; but not after models are adjusted for both selenium and copper. These results add valuable information regarding these compounds at an early stage of development, and suggest that simultaneous examination of multiple micronutrients may be beneficial in fully understanding the relationships between micronutrients and cardiovascular risk factors.

Selenium and copper are both essential nutrients and for optimal health their concentrations need to remain within a certain range. While the beneficial ranges for selenium and copper in adults are fairly well understood, relatively little information on optimal concentrations of these micronutrients in umbilical cord blood is available. Selenium concentrations

Table 1. Participant characteristics, THREE study, 2004–2005

Characteristic	Population		
	Q4 triglycerides ^a	Q4 Total cholesterol ^a	Entire cohort
<i>n</i> (%)	68 (24.8)	68 (24.8)	274 (100)
Maternal age (years); mean (95% CI)	26.0 (24.4, 27.6)	26.3 (24.7, 27.9)	25.8 (25.0, 26.6)
African American mothers; % (95% CI) ^b	64.7 (52.4, 75.3)	73.5 (61.5, 82.8)	70.8 (65.1, 75.9)
Maternal smoking; % (95% CI)	17.6 (10.1, 28.9)	16.2 (9.1, 27.2)	18.2 (14.0, 23.3)
Primiparity; % (95% CI)	58.8 (46.5, 70.1)	38.2 (27.2, 50.6)	42.3 (36.6, 48.3)
Maternal prepregnancy BMI (kg/m ²); mean (95% CI)	28.5 (27.1, 29.9)	26.8 (25.3, 28.3)	26.4 (25.6, 27.2)
Gestation (days); mean (95% CI)	273.4 (269.1, 277.6)	264.8 (260.0, 269.6)	271.8 (270.2, 273.5)
Birth weight (g); mean (95% CI)	3162 (3105, 3309)	3093 (2928, 3258)	3192 (3124, 3260)
n-3 HUFAs/total HUFAs; mean (95% CI)	18.9 (17.9, 19.9)	17.0 (16.2, 17.9)	18.1 (17.6, 18.5)
Methyl mercury (µg/l); geometric mean (95% CI)	0.95 (0.77, 1.17)	1.13 (0.92, 1.38)	1.05 (0.96, 1.15)
Selenium (µg/l); mean (95% CI)	72.7 (69.5, 75.9)	71.6 (68.4, 74.8)	69.8 (68.3, 71.2)
Copper (µg/dl); geometric mean (95% CI)	47.7 (43.5, 52.2)	37.6 (33.7, 42.0)	38.6 (36.7, 40.5)
Triglycerides (mg/dl); geometric mean (95% CI)	61.1 (57.8, 64.6)	39.2 (35.2, 43.7)	34.5 (32.9, 36.3)
Total cholesterol (mg/dl); geometric mean (95% CI)	70.1 (65.2, 75.3)	92.1 (88.5, 95.8)	64.6 (62.5, 66.7)

THREE, Tracking Health Related to Environmental Exposures; CI, confidence interval; BMI, body mass index; HUFA, highly unsaturated fatty acid; Q4, fourth quartile.

^aUmbilical cord serum triglycerides ≥ 46.0 mg/dl; umbilical cord serum total cholesterol ≥ 77.7 mg/dl.

^bPercent of Caucasians was Q4 triglycerides: 27.9 (95% CI: 18.4, 40.0); Q4 total cholesterol: 25.0 (95% CI: 16.0, 36.9); entire cohort: 21.5 (95% CI: 17.0, 26.8). Percent of Asians was Q4 triglycerides: 7.4 (95% CI: 3.0, 16.8); Q4 total cholesterol: 1.5 (95% CI: 0.2, 10.2); entire cohort: 7.7 (95% CI: 5.0, 11.5).

Table 2. Percent difference (95% confidence interval) in cord serum triglyceride by increasing quartile of selenium or copper, THREE study 2004–2005, n = 274

Model	Q1	Q2	Q3	Q4	Test for trend ^a
Selenium (µg/l)	42.0–62.0	63.0–69.0	70.0–78.0	79.0–114.0	
Model 1 ^b	1.00 (referent)	7.6 (–5.9, 23.1)	11.0 (–2.9, 27.0)	24.2 (8.1, 42.6)	0.008
Model 2 ^c	1.00 (referent)	11.8 (–1.1, 26.5)	13.1 (–0.1, 28.2)	22.3 (7.1, 39.7)	0.005
Model 3 ^{c,d}	1.00 (referent)	6.1 (–5.9, 19.6)	5.5 (–6.7, 19.2)	6.3 (–7.5, 22.2)	0.457
Copper (µg/dl)	12.1–28.4	28.5–38.0	38.1–51.6	51.7–109.0	
Model 1 ^b	1.00 (referent)	12.4 (–1.2, 28.0)	28.6 (12.7, 46.8)	50.6 (31.6, 72.2)	< 0.001
Model 2 ^c	1.00 (referent)	15.9 (2.6, 30.8)	30.3 (14.8, 48.0)	43.8 (25.9, 64.3)	< 0.001
Model 3 ^d	1.00 (referent)	15.0 (1.8, 30.0)	27.8 (11.7, 46.2)	40.7 (22.1, 62.1)	< 0.001

THREE, Tracking Health Related to Environmental Exposures; Q1, first quartile; Q2, second quartile; Q3, third quartile; Q4, fourth quartile; HUFA, highly unsaturated fatty acid.

^aBased on regression model using selenium or ln(copper) instead of quartiles.

^bAdjusted for gestational age (quadratic).

^cAdjusted for gestational age (quadratic), maternal age, maternal race, maternal smoking, primiparity, low birth weight, prepregnancy body mass index (quadratic), n-3 HUFAs/total HUFAs, and ln(methyl mercury).

^dAdjusted for variables in Model 2 as well as (3c) ln(copper) or (3d) selenium.

Table 3. Percent difference (95% confidence interval) in cord serum total cholesterol by increasing quartile of selenium or copper, THREE study 2004–2005, n = 274

Model	Q1	Q2	Q3	Q4	Test for trend ^a
Selenium (µg/l)	42.0–62.0	63.0–69.0	70.0–78.0	79.0–114.0	
Model 1 ^b	1.00 (referent)	2.7 (–5.8, 11.9)	3.3 (–5.3, 12.6)	6.4 (–2.7, 16.2)	0.038
Model 2 ^c	1.00 (referent)	3.4 (–5.2, 12.8)	2.5 (–6.2, 11.9)	7.9 (–1.7, 18.5)	0.035
Model 3 ^{c,d}	1.00 (referent)	2.3 (–6.3, 11.7)	1.0 (–7.7, 10.5)	4.9 (–5.2, 16.2)	0.132
Copper (µg/dl)	12.1–28.4	28.5–38.0	38.1–51.6	51.7–109.0	
Model 1 ^b	1.00 (referent)	1.4 (–7.1, 10.7)	7.2 (–1.9, 17.3)	3.0 (–6.0, 12.8)	0.222
Model 2 ^c	1.00 (referent)	2.1 (–6.6, 11.5)	10.6 (0.8, 21.3)	6.7 (–3.2, 17.6)	0.062
Model 3 ^d	1.00 (referent)	1.2 (–7.4, 10.6)	8.0 (–2.0, 19.2)	3.9 (–6.3, 15.2)	0.254

THREE, Tracking Health Related to Environmental Exposures; Q1, first quartile; Q2, second quartile; Q3, third quartile; Q4, fourth quartile; HUFA, highly unsaturated fatty acid.

^aBased on regression model using selenium or ln(copper) instead of quartiles.

^bAdjusted for gestational age (quadratic).

^cAdjusted for gestational age (quadratic), maternal age, maternal race, maternal smoking, primiparity, low birth weight, prepregnancy body mass index (quadratic), n-3 HUFAs/total HUFAs and ln(methyl mercury).

^dAdjusted for variables in model 2 as well as either (3c) ln(copper) or (3d) selenium.

are known to vary geographically, in large part due to differences in selenium content in soil and subsequently in the diet.³³ In the United States, the majority of the population is selenium-replete;¹⁴ therefore, slight increases of selenium may be sufficient to exceed the range of selenium's beneficial effect. To date, however, scientific evidence for this has been mixed.^{17,34}

We observed a positive relationship between selenium and triglycerides or total cholesterol, but this relationship did not persist after controlling for copper. Several cross-sectional studies conducted in adult populations from the United States or Western Europe report positive relationships between selenium and triglycerides,^{14,16,19,35} including one among newborns,²³

or selenium and cholesterol.^{14–17,19,35} However, other studies did not see a relationship of selenium with triglycerides³⁶ or found an inverse relationship of selenium with triglycerides and cholesterol.³⁷ It has also been suggested by a few researchers that the relationship between selenium and cardiovascular risk factors may depend on whether the selenium was derived from fish sources *v.* non-fish sources such as nuts.^{38–40} These studies suggested that non-fish sources were not related to increases in cardiovascular risk factors.

Longitudinal studies of selenium and lipids have recently been published. A short-term selenium supplementation trial among United Kingdom adults found selenium related to a reduction in total and non-high-density lipoprotein (HDL)

cholesterol; however, this seemed to be attenuated at higher levels of supplementation.⁴¹ Additionally, a Cochrane review of selenium supplementation trials, most of which did not include lipids as a primary outcome, did not find evidence of a relationship between selenium and cholesterol or triglycerides.^{36,42} Interestingly, two recent cohort studies among Finnish youth³⁴ and adult Italian males⁴³ both found positive associations of selenium with cholesterol on a cross-sectional, but not longitudinal basis.

We report a significant positive relationship between copper and triglycerides, but did not observe a relationship between copper and total cholesterol among newborns. This is consistent with a study conducted among Spanish newborns²⁴ and a cross-sectional study among hyperlipidemic Iraqi men,²⁰ although there was no correlation between copper and triglycerides among controls in the Iraqi study. Several other studies in children or adults have found positive relationships between copper and several forms of cholesterol,¹⁹ HDL cholesterol,¹⁵ or an unfavorable lipid profile (LDL/HDL >2.2).⁴⁴ However, other studies among adults report inverse associations between copper and cholesterol²¹ or no relationship between copper and triglycerides^{15,19,44,45} or cholesterol.^{45,46}

A few studies in adult populations have evaluated both selenium and copper with respect to lipid concentrations.^{15,19,22} In Lebanese adults, both selenium and copper were correlated with total cholesterol but only selenium was correlated with triglycerides.¹⁹ Portuguese men with hyperlipidemia had higher selenium concentrations but lower copper concentrations compared to controls, with no differences among women.²² In adults from the United Kingdom, HDL cholesterol was positively related to copper and total cholesterol positively related to selenium concentrations in multivariable analyses.¹⁵ The comparison with our findings, however, is uncertain because those studies did not adjust for the other metal, two of them did not adjust for other key variables^{19,22} and they were conducted in adult instead of newborn populations. Average concentrations of selenium and copper differ substantially between adults and children and the relationship of these micronutrients with lipids could be life-stage dependent.

Some results from our regression models (Tables 2–3) have a statistically significant test for overall trend of triglycerides or cholesterol levels across quartiles of selenium and/or copper, while comparisons of the second, third and/or fourth quartile *v.* the lowest quartile levels were not statistically significant, as confidence intervals included zero. This suggests that the overall magnitude of changes we observed were perhaps too small and too variable to be observed for the numbers of individuals in each quartile (68), but large enough to be detected by a trend test that incorporates the entire range of exposures and therefore has more statistical power.

One limitation of this study is that we did not have access to some variables which may have been useful in analyses. Specifically, maternal blood samples were not collected; and laboratory analysis of zinc, chromium, HDL cholesterol or LDL cholesterol were not available. In particular, several

studies have reported different results for HDL cholesterol in comparison with total cholesterol or LDL cholesterol for both selenium^{17,19,36,39} and copper¹⁵ as we do not have HDL and LDL measurements we are unable to directly compare our results to this work.

Promotion of healthy serum lipid concentrations, a major risk factor for cardiovascular disease, is an important public health goal. Our findings indicate that at birth, higher triglyceride concentrations in umbilical cord blood are associated with elevated copper concentrations and maybe also with elevated selenium. Our work adds important early life evidence to a growing body of evidence on the relationship between trace metals and lipid concentrations.

Acknowledgments

The authors thank Jochen Heidler for help with data collection; Ruth Quinn for study management; as well as Norman Salem, Jr. and John Bernert for their assistance with laboratory analyses. The U.S. EPA has not officially endorsed this work and the views presented herein are those of the author and may not reflect those of the agency. Likewise, the findings and conclusions in this report are those of the authors and do not necessarily represent official positions of the Centers for Disease Control and Prevention, the Food and Drug Administration or the National Institutes of Health.

Financial Support

This study received funding from the Maryland Cigarette Restitution Program Research Grant, the United States National Institute for Environmental Health Sciences (R.U.H., grant #:1R01ES015445), and the United States STAR Fellowship Program (E.M.W.).

Conflicts of Interest

None.

Ethical Standards

The authors assert that all procedures contributing to this work comply with the ethical standards of the United States Department of Health and Human Services (HHS 45CFR 46) and with the Helsinki Declaration of 1975, as revised in 2008, and has been approved by the institutional committees at the Johns Hopkins Hospital, in Baltimore, Maryland, United States.

References

1. Berenson GS, Srinivasan SR, Bao W, *et al.* Association between multiple cardiovascular risk factors and atherosclerosis in children and young adults. The Bogalusa Heart Study. *N Engl J Med.* 1998; 338, 1650–1656.
2. Owen CG, Rudnicka AR, Nightingale CM, *et al.* Retinal arteriolar tortuosity and cardiovascular risk factors in a multi-ethnic

- population study of 10-year-old children; the Child Heart and Health Study in England (CHASE). *Arterioscler Thromb Vasc Biol.* 2011; 31, 1933–1938.
3. Li S, Chen W, Srinivasan SR, et al. Childhood cardiovascular risk factors and carotid vascular changes in adulthood: the Bogalusa Heart Study. *JAMA.* 2003; 290, 2271–2276.
 4. Raitakari OT, Juonala M, Kahonen M, et al. Cardiovascular risk factors in childhood and carotid artery intima-media thickness in adulthood: the Cardiovascular Risk in Young Finns Study. *JAMA.* 2003; 290, 2277–2283.
 5. Barceloux DG. Selenium. *J Toxicol Clin Toxicol.* 1999; 37, 145–172.
 6. Barceloux DG. Copper. *J Toxicol Clin Toxicol.* 1999; 37, 217–230.
 7. Stern BR, Solioz M, Krewski D, et al. Copper and human health: biochemistry, genetics, and strategies for modeling dose-response relationships. *J Toxicol Environ Health B Crit Rev.* 2007; 10, 157–222.
 8. Flores-Mateo G, Navas-Acien A, Pastor-Barriuso R, Guallar E. Selenium and coronary heart disease: a meta-analysis. *Am J Clin Nutr.* 2006; 84, 762–773.
 9. Navas-Acien A, Bleys J, Guallar E. Selenium intake and cardiovascular risk: what is new? *Curr Opin Lipidol.* 2008; 19, 43–49.
 10. Rayman MP, Bode P, Redman CW. Low selenium status is associated with the occurrence of the pregnancy disease preeclampsia in women from the United Kingdom. *Am J Obstet Gynecol.* 2003; 189, 1343–1349.
 11. Reunanen A, Knekt P, Marniemi J, et al. Serum calcium, magnesium, copper and zinc and risk of cardiovascular death. *Eur J Clin Nutr.* 1996; 50, 431–437.
 12. Salonen JT, Salonen R, Korpela H, Suntioinen S, Tuomilehto J. Serum copper and the risk of acute myocardial infarction: a prospective population study in men in eastern Finland. *Am J Epidemiol.* 1991; 134, 268–276.
 13. Stranges S, Marshall JR, Trevisan M, et al. Effects of selenium supplementation on cardiovascular disease incidence and mortality: secondary analyses in a randomized clinical trial. *Am J Epidemiol.* 2006; 163, 694–699.
 14. Bleys J, Navas-Acien A, Stranges S, et al. Serum selenium and serum lipids in US adults. *Am J Clin Nutr.* 2008; 88, 416–423.
 15. Ghayour-Mobarhan M, Taylor A, New SA, Lamb DJ, Ferns GA. Determinants of serum copper, zinc and selenium in healthy subjects. *Ann Clin Biochem.* 2005; 42(Pt 5), 364–375.
 16. Laclaustra M, Stranges S, Navas-Acien A, Ordovas JM, Guallar E. Serum selenium and serum lipids in US adults: National Health and Nutrition Examination Survey (NHANES) 2003–2004. *Atherosclerosis.* 2010; 210, 643–648.
 17. Stranges S, Laclaustra M, Ji C, et al. Higher selenium status is associated with adverse blood lipid profile in British adults. *J Nutr.* 2010; 140, 81–87.
 18. Ghayour-Mobarhan M, Taylor A, Kazemi-Bajestani SM, et al. Serum zinc and copper status in dyslipidaemic patients with and without established coronary artery disease. *Clin Lab.* 2008; 54, 321–329.
 19. Obeid O, Elfakhani M, Hlais S, et al. Plasma copper, zinc, and selenium levels and correlates with metabolic syndrome components of Lebanese adults. *Biol Trace Elem Res.* 2008; 123, 58–65.
 20. Al-Sabaawy OM. The relationship between serum lipid profile and selected trace elements for adult men in mosul city. *Oman Med J.* 2012; 27, 300–303.
 21. Bo S, Durazzo M, Gambino R, et al. Associations of dietary and serum copper with inflammation, oxidative stress, and metabolic variables in adults. *J Nutr.* 2008; 138, 305–310.
 22. Pavao ML, Figueiredo T, Santos V, et al. Whole blood glutathione peroxidase and erythrocyte superoxide dismutase activities, serum trace elements (Se, Cu, Zn) and cardiovascular risk factors in subjects from the city of Ponta Delgada, Island of San Miguel, The Azores Archipelago, Portugal. *Biomarkers.* 2006; 11, 460–471.
 23. Boskabadi H, Maamouri G, Rezagholizade Omran F, et al. Effect of prenatal selenium supplementation on cord blood selenium and lipid profile. *Pediatr Neonatol.* 2012; 53, 334–339.
 24. Bastida S, Vaquero MP, Veldhuizen M, Sanchez-Muniz FJ. Selected trace elements and minerals in cord blood: association with lipids and lipoproteins at birth. *Acta Paediatr.* 2000; 89, 1201–1206.
 25. Wells EM, Jarrett JM, Lin YH, et al. Body burdens of mercury, lead, selenium and copper among Baltimore newborns. *Environ Res.* 2011; 111, 411–417.
 26. Herbstman JB, Sjodin A, Apelberg BJ, et al. Determinants of prenatal exposure to polychlorinated biphenyls (PCBs) and polybrominated diphenyl ethers (PBDEs) in an urban population. *Environ Health Perspect.* 2007; 115, 1794–1800.
 27. Hornung RW, Reed RD. Estimation of average concentration in the presence of nondetectable values. *Appl Occup Environ Hyg.* 1990; 5, 46–51.
 28. Lin YH, Salem N Jr., Wells EM, et al. Automated high-throughput fatty acid analysis of umbilical cord serum and application to an epidemiological study. *Lipids.* 2012; 47, 527–539.
 29. Masood MA, Salem N Jr. High-throughput analysis of plasma fatty acid methyl esters employing robotic transesterification and fast gas chromatography. *Lipids.* 2008; 43, 171–180.
 30. Wells EM, Navas-Acien A, Herbstman JB, et al. Low-level lead exposure and elevations in blood pressure during pregnancy. *Environ Health Perspect.* 2011; 119, 664–669.
 31. Park K, Mozaffarian D. Omega-3 fatty acids, mercury, and selenium in fish and the risk of cardiovascular diseases. *Curr Atheroscler Rep.* 2010; 12, 414–422.
 32. Shearer GC, Savinova OV, Harris WS. Fish oil – how does it reduce plasma triglycerides? *Biochimica et Biophysica Acta.* 2012; 1821, 843–851.
 33. Navarro-Alarcon M, Cabrera-Vique C. Selenium in food and the human body: a review. *Sci Total Environ.* 2008; 400, 115–141.
 34. Stranges S, Tabak AG, Guallar E, et al. Selenium status and blood lipids: the cardiovascular risk in Young Finns study. *J Intern Med.* 2011; 270, 469–477.
 35. Yang KC, Lee LT, Lee YS, et al. Serum selenium concentration is associated with metabolic factors in the elderly: a cross-sectional study. *Nutr Metab.* 2010; 7, 38.
 36. Suadicani P, Hein HO, Gyntelberg F. Serum selenium concentration and risk of ischaemic heart disease in a prospective cohort study of 3000 males. *Atherosclerosis.* 1992; 96, 33–42.
 37. Karita K, Yamanouchi Y, Takano T, et al. Associations of blood selenium and serum lipid levels in Japanese premenopausal and postmenopausal women. *Menopause.* 2008; 15, 119–124.

38. Cominetti C, de Bortoli MC, Garrido AB Jr., Cozzolino SM. Brazilian nut consumption improves selenium status and glutathione peroxidase activity and reduces atherogenic risk in obese women. *Nutr Res.* 2012; 32, 403–407.
39. Miyazaki Y, Koyama H, Nojiri M, Suzuki S. Relationship of dietary intake of fish and non-fish selenium to serum lipids in Japanese rural coastal community. *J Trace Elem Med Biol.* 2002; 16, 83–90.
40. Strunz CC, Oliveira TV, Vinagre JC, *et al.* Brazil nut ingestion increased plasma selenium but had minimal effects on lipids, apolipoproteins, and high-density lipoprotein function in human subjects. *Nutr Res.* 2008; 28, 151–155.
41. Rayman MP, Stranges S, Griffin BA, Pastor-Barriuso R, Guallar E. Effect of supplementation with high-selenium yeast on plasma lipids: a randomized trial. *Ann Intern Med.* 2011; 154, 656–665.
42. Rees K, Hartley L, Day C, *et al.* Selenium supplementation for the primary prevention of cardiovascular disease. *Cochrane Database Syst Rev.* 2013; 1, CD009671.
43. Stranges S, Galletti F, Farinero E, *et al.* Associations of selenium status with cardiometabolic risk factors: an 8-year follow-up analysis of the Olivetti Heart study. *Atherosclerosis.* 2011; 217, 274–278.
44. Elcarte Lopez T, Villa Elizaga I, Gost Garde JI, *et al.* Cardiovascular risk factors in relation to the serum concentrations of copper and zinc: epidemiological study on children and adolescents in the Spanish province of Navarra. *Acta Paediatr.* 1997; 86, 248–253.
45. Kim MH, Choi MK. Seven dietary minerals (Ca, P, Mg, Fe, Zn, Cu, and Mn) and their relationship with blood pressure and blood lipids in healthy adults with self-selected diet. *Biol Trace Elem Res.* 2013; 153, 69–75.
46. DiSilvestro RA, Joseph EL, Zhang W, Raimo AE, Kim YM. A randomized trial of copper supplementation effects on blood copper enzyme activities and parameters related to cardiovascular health. *Metabolism.* 2012; 61, 1242–1246.