

# Diet and feeding strategy of the forkbeard *Phycis phycis* (Pisces: Phycidae) from the Portuguese continental coast

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*The diet and feeding behaviour of the forkbeard Phycis phycis was studied based on 246 stomachs collected between May 2011 and April 2012 from a commercial fleet operating off the central west coast of Portugal. A total of 44 prey items were identified in the stomachs which were merged into major groups to avoid problems with low expected frequencies. The following taxonomic categories were considered: non-decapod Crustacea, Caridea, Anomura, Munida spp., Processa spp., Brachyura, Pisces, Trisopterus luscus. In order to investigate possible diet differences between fish size classes, a cluster analysis was performed using the mean abundance of each prey group by forkbeard 5 cm length class, and three length groups (LG) were obtained: <22.5, 27.5–37.5 and >42.5 cm. Seasonally, Caridea was the main prey group during winter and autumn while Pisces was predominant during the rest of the year. Caridea was the most important prey group for LG1 and LG2 while in LG3 Pisces was the principal one. The forkbeard feeding behaviour may be characterized as presenting a shift pattern from a more generalist diet (small Crustacea, mainly Caridea) in the young adults to a more specialist strategy (teleosts) in the adults.*

**Keywords:** *Phycis phycis*, forkbeard, diet, feeding strategy

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## INTRODUCTION

Studies on diet and feeding strategy of fish species are fundamental to understand many aspects of their biology, ecology, physiology and behaviour (Gonçalves & Erzini, 1998). Such studies are even more crucial when involving upper trophic level species, which are especially important due to their recent global declines and the potential for associated ecosystem-level effects on species composition and diversity (Pauly *et al.*, 1998). In this context, stomach content analysis is the most widely used method for studying the diet of fish, allowing determination of the role of a species in the food chain (Hyslop, 1980) and therefore contributes to the study of intra- and inter-specific relationships in the ecosystem.

The forkbeard *Phycis phycis* (Linnaeus, 1766) is a gadiform benthopelagic fish with a wide distribution in the North-east Atlantic (from the Bay of Biscay to Morocco, south to Cape Verde and the Azores) and in the Mediterranean Sea (Svetovidov, 1986). The forkbeard lives on hard and sandy-muddy bottoms near rocks at depths up to 650 m (Svetovidov, 1986), where it looks for shelter in holes during the day and becomes an active predator during the night, feeding mainly on fish but also on decapods (Papaconstantinou & Caragitsou, 1989; Morato *et al.*, 1999). In the southern NE Atlantic, forkbeard is an important

commercial species, both in Portugal and Spain (Vieira *et al.*, 2014a, b, 2016a, b), with Portuguese landings reaching about 800 tons per year (INE, 2014). In Portuguese waters, this species is mainly caught by a longline fishery (trawl, trammel net and traps fisheries contribute with a small percentage of the landings) in coastal waters and offshore seamounts, in the mainland area and around Azores and Madeira archipelagos (Vieira *et al.*, 2014a, b). Despite its economic importance, little information is available on its biology (Silva, 1986; Abecasis *et al.*, 2009; Matic-Skoko *et al.*, 2011; Glavić *et al.*, 2014; Vieira *et al.*, 2014a, 2016b).

This study aims to present new data on diet composition and feeding strategy of the forkbeard in the Portuguese continental waters focusing on differences in the feeding habits according to fish length, season and sex.

## MATERIALS AND METHODS

### Sampling

Forkbeard specimens were obtained monthly between May 2011 and April 2012 from commercial vessels operating off mainland Portugal (mainly in the central west coast) (Figure 1), from depths between 55 and 310 m using mainly longline but also bottom trawl and trammel nets and landed at Peniche. Table 1 shows the number of individuals and the length range of the specimens caught by each gear and season (winter: January–March; spring: April–June;

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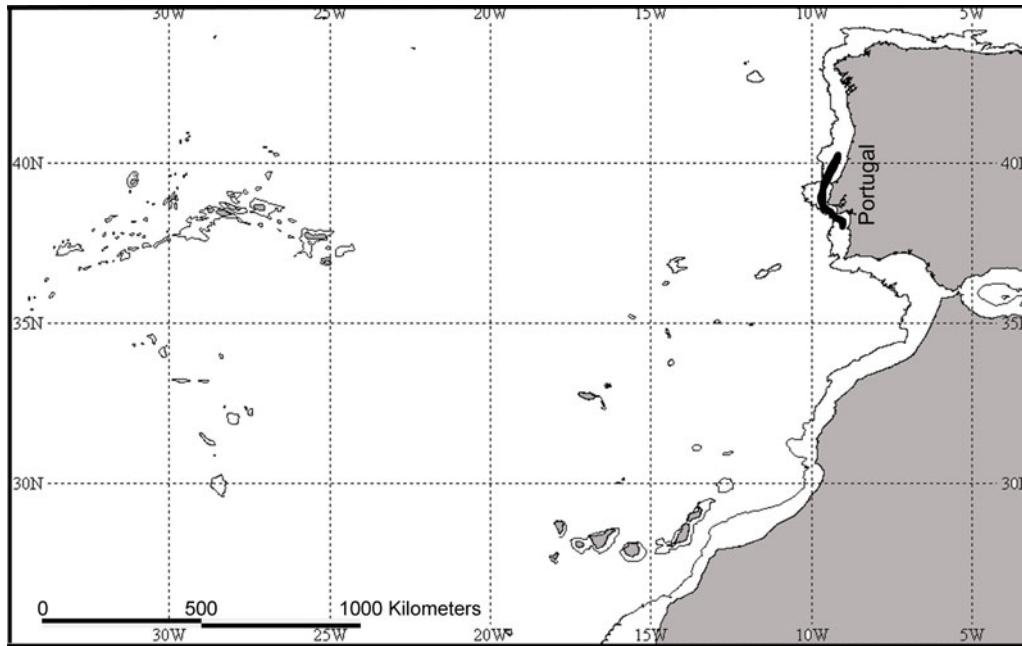


Fig. 1. Map of the southern NE Atlantic with the location of the sampling area (shaded). Black line represents the 1000 m isobath.

summer: July–September; autumn: October–December). The depth interval at which specimens were caught is also shown.

In the laboratory, total length (TL, to the nearest 0.1 cm), gutted weight (GW, to the nearest 0.01 g), stomach weight (to the nearest 0.01 g) and sex were recorded from each fish. The stomachs were removed and frozen for later analysis.

The food items in the stomach contents, after being defrosted and dried in absorbent paper, were carefully separated, identified to the lowest taxonomic level possible, counted and weighed (to the nearest 0.0001 g). Whenever parts of uncompleted individuals were found, the number of individuals was assumed as the smallest possible number from which fragments could have originated. Food items that might have been used as bait in longline, mainly Atlantic chub mackerel *Scomber colias* Gmelin, 1789 and European sardine *Sardina pilchardus* (Walbaum, 1792), were excluded from the analysis if they were found whole or lightly digested.

## Feeding patterns

Dietary indices were used to quantify the prey relative importance (Hyslop, 1980; Cortés, 1997) by prey item and prey items aggregated by major taxonomic groups. The indices used were

(i) the frequency of occurrence (%O), (ii) the percentage by number (%N), (iii) the per cent by weight (%W), and (iv) the per cent index of relative importance (%IRI), described as:

$$(i) \%O = \frac{\text{number of stomachs with prey item } i}{\text{number of non empty stomachs}} \times 100$$

$$(ii) \%N = \frac{\text{number of prey item } i \text{ in all stomachs}}{\text{total number of food items in all stomachs}} \times 100$$

$$(iii) \%W = \frac{\text{total weight of prey item } i \text{ in all stomachs}}{\text{total weight of stomach contents}} \times 100$$

$$(iv) \%IRI = \left[ \frac{\%O \times (\%N + \%W)}{\sum (\%O \times (\%N + \%W))} \right] \times 100$$

The number of everted stomachs was estimated as well as the vacuity index (VI) considered as the percentage of empty stomachs in the sample (Ellis *et al.*, 1996). Full regurgitation was not taken into account and was included in the number of empty stomachs.

Table 1. Number of specimens of *Phycis phycis* caught by total length range, gear, season and depth.

		Spring	Summer	Autumn	Winter	Total
Longline	N	26	35	28	54	143
	TL (cm)	39–65	46–65	43–57	46–67	39–67
	Depth (m)					180–310
Trammel net	N	20	15	14	15	64
	TL (cm)	29–45	22–50	29–42	26–50	22–50
	Depth (m)					55–200
Trawl	N	8	24	3	4	39
	TL (cm)	19–28	27–43	18–34	15–26	15–43
	Depth (m)					55–150
	Total	54	74	45	73	246

Cumulative prey curves were performed for each group and season to determine if the number of specimens used was adequate to describe precisely the diet of the forkbeard. The order in which stomachs were analysed was randomized 10 times and the mean number of new prey species found consecutively in the stomachs plotted against the number of stomachs analysed (Ferry *et al.*, 1997). The presence of an

asymptotic relationship, which indicates that enough samples had been analysed, was investigated by the method developed by Bizzarro *et al.* (2007). The mean coefficient of variation of the four last points was additionally calculated to provide a standard measure of precision.

To evaluate the degree of feeding intensity of each individual, the stomach fullness index (SFI), described as the

**Table 2.** Diet composition of *Phycis phycis* by Length Group (LG) expressed as frequency of occurrence (%O), percentage by number (%N), percentage by weight (%W), and per cent index of relative importance (%IRI).

	LG2				LG3			
	%O	%N	%W	%IRI	%O	%N	%W	%IRI
Echinodermata	0.0	0.0	0.0	0.0	1.0	0.4	0.0	0.0
Ophiuroidea	0.0	0.0	0.0	0.0	1.0	0.4	0.0	0.0
Mollusca	2.6	0.5	0.1	0.0	0.0	0.0	0.0	0.0
Cephalopoda	2.6	0.5	0.1	0.0	0.0	0.0	0.0	0.0
Non-decapod crustacea	23.1	7.5	1.6	3.0	11.8	6.9	0.1	1.1
Crustacea (N.I.)	17.9	5.6	1.5	2.8	2.9	1.3	0.0	0.1
Cirolanidae (N.I.)	5.1	1.9	0.1	0.2	7.8	5.2	0.0	1.0
Cirripedia	0.0	0.0	0.0	0.0	1.0	0.4	0.0	0.0
Penaeoidea	10.3	3.8	0.4	0.5	0.0	0.0	0.0	0.0
<i>Penaeus</i> spp.	7.7	2.3	0.1	0.4	0.0	0.0	0.0	0.0
<i>Solenocera</i> spp.	2.6	1.4	0.3	0.1	0.0	0.0	0.0	0.0
Caridea	82.1	51.2	6.4	54.0	12.7	15.5	0.2	2.6
Caridea (N.I.)	53.8	37.1	5.4	49.7	8.8	10.8	0.1	2.3
Pandalidae	10.3	3.3	0.1	0.7	2.0	3.0	0.1	0.2
<i>Processa</i> spp.	15.4	9.9	0.7	3.5	2.0	1.7	0.0	0.1
<i>Alpheus glaber</i>	2.6	0.9	0.2	0.1	0.0	0.0	0.0	0.0
Anomura	41.0	15.0	5.4	10.3	8.8	4.7	1.4	0.7
Anomura (N.I.)	0.0	0.0	0.0	0.0	1.0	0.4	0.0	0.0
<i>Galathea</i> spp.	15.4	4.2	0.3	1.5	2.0	1.3	0.1	0.1
<i>Munida</i> spp.	25.6	10.8	5.0	8.8	5.9	3.0	1.3	0.6
Brachyura	46.2	13.1	6.2	3.8	18.6	9.5	0.9	1.1
Brachyura (N.I.)	15.4	3.8	1.1	1.6	6.9	3.0	0.3	0.6
<i>Atelecyclus rotundatus</i>	7.7	2.3	1.7	0.7	4.9	3.0	0.4	0.4
Portunidae	0.0	0.0	0.0	0.0	0.0	0.4	0.0	0.0
<i>Bathynectes</i> spp.	2.6	1.4	1.0	0.1	2.0	0.9	0.0	0.0
<i>Liocarcinus</i> spp.	5.1	0.9	1.1	0.2	1.0	0.4	0.0	0.0
<i>Polybius henslowii</i>	2.6	0.5	0.2	0.0	0.0	0.0	0.0	0.0
<i>Pilumnus</i> spp.	0.0	0.0	0.0	0.0	1.0	0.4	0.0	0.0
<i>Pilumnus hirtellus</i>	10.3	3.8	0.9	1.0	1.0	0.4	0.0	0.0
<i>Monodaeus couchii</i>	2.6	0.5	0.2	0.0	0.0	0.0	0.0	0.0
<i>Goneplax rhomboides</i>	0.0	0.0	0.0	0.0	1.0	0.4	0.0	0.0
<i>Pisa</i> spp.	0.0	0.0	0.0	0.0	1.0	0.4	0.0	0.0
Pisces	46.2	8.9	79.9	28.3	97.1	62.9	97.5	94.5
Teleostei (N.I.)	17.9	3.3	10.7	5.5	34.3	18.1	5.5	19.9
<i>Conger conger</i>	5.1	1.4	0.2	0.2	1.0	0.4	0.1	0.0
<i>Sardina pilchardus</i>	0.0	0.0	0.0	0.0	6.9	5.6	7.2	2.2
<i>Phycis</i> spp.	0.0	0.0	0.0	0.0	2.0	0.9	6.1	0.3
<i>Merluccius merluccius</i>	0.0	0.0	0.0	0.0	1.0	0.4	1.0	0.0
<i>Micromesistius poutassou</i>	0.0	0.0	0.0	0.0	4.9	2.2	1.5	0.4
<i>Trisopterus luscus</i>	15.4	2.8	64.0	22.3	35.3	25.0	56.6	70.8
<i>Capros aper</i>	2.6	0.5	4.3	0.3	2.0	1.3	1.8	0.1
<i>Serranus</i> spp.	2.6	0.5	0.1	0.0	1.0	0.4	0.1	0.0
<i>Serranus cabrilla</i>	0.0	0.0	0.0	0.0	1.0	0.4	0.1	0.0
<i>Diplodus</i> spp.	0.0	0.0	0.0	0.0	1.0	0.4	0.5	0.0
<i>Mullus surmuletus</i>	0.0	0.0	0.0	0.0	1.0	0.4	2.7	0.1
Labridae	0.0	0.0	0.0	0.0	1.0	0.4	0.1	0.0
<i>Acantholabrus palloni</i>	0.0	0.0	0.0	0.0	1.0	0.4	1.8	0.1
<i>Symphodus melops</i>	0.0	0.0	0.0	0.0	1.0	0.4	0.1	0.0
Ammodytidae	0.0	0.0	0.0	0.0	2.0	2.6	1.7	0.2
<i>Pomatoschistus</i> spp.	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Scomber colias</i>	2.6	0.5	0.5	0.1	0.0	0.0	0.0	0.0
<i>Scomber scombrus</i>	0.0	0.0	0.0	0.0	1.0	3.4	10.7	0.3

percentage of the stomach content weight (SCW) in relation to the GW (Hyslop, 1980) was estimated.

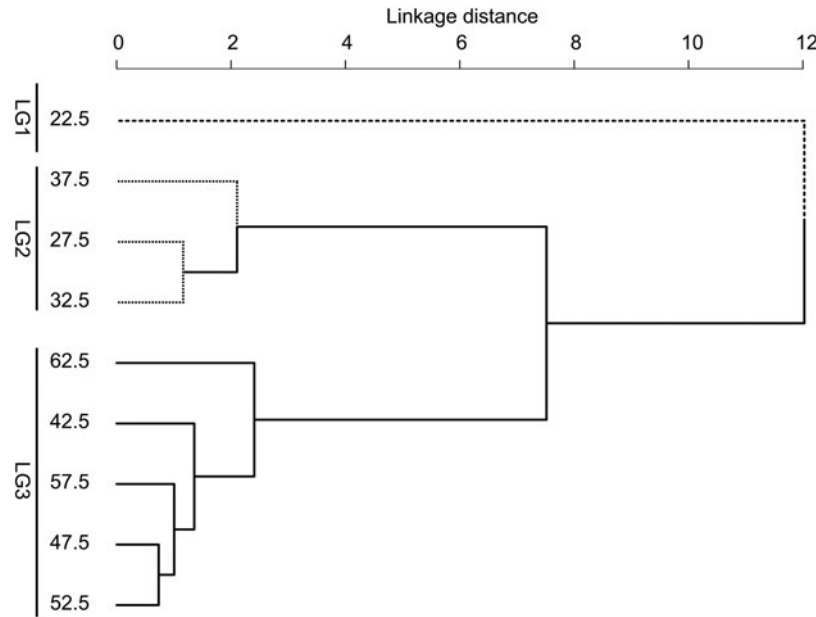
For the subsequent analyses, prey items were merged into major groups to avoid problems with low expected frequencies and the most common species/genera were included as separated items in these analyses. The higher taxonomic groups and the selected species/genera were as follows: non-decapod Crustacea, NDC; Caridea, CAR; Anomura, ANO;

*Munida* spp., MUN; *Processa* spp., PRO; Brachyura, BRA; Pisces, PIS; *Trisopterus luscus*, TRI. Echinodermata and Mollusca appeared with extremely low frequency (<1%) and therefore were excluded from the analysis.

To investigate possible diet differences between fish size classes, a cluster analysis (Ward's method, Manhattan distance) was performed using the mean abundance of each prey group by forkbeard 5 cm length class and, as a result,

**Table 3.** Diet composition of *Phycis phycis* by season expressed as frequency of occurrence (%O), percentage by number (%N), percentage by weight (%W), and per cent index of relative importance (%IRI).

	Spring				Summer				Autumn				Winter			
	%O	%N	%W	%IRI	%O	%N	%W	%IRI	%O	%N	%W	%IRI	%O	%N	%W	%IRI
Echinodermata	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	2.2	1.0	0.2	0.0	0.0	0.0	0.0	0.0
Ophiuroidea	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	2.2	1.0	0.2	0.0	0.0	0.0	0.0	0.0
Mollusca	2.3	0.7	0.0	0.1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Cephalopoda	2.3	0.7	0.0	0.1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Non-decapod crustacea	20.9	10.3	0.3	5.9	21.2	13.5	0.2	6.4	8.7	5.0	0.0	0.4	4.8	2.2	0.0	0.2
Crustacea (N.I.)	16.3	8.3	0.3	5.7	3.0	1.4	0.0	0.1	4.3	2.0	0.0	0.2	0.0	0.0	0.0	0.0
Cirrolanidae (N.I.)	2.3	1.4	0.0	0.1	18.2	12.2	0.1	6.3	4.3	3.0	0.0	0.2	4.8	2.2	0.0	0.2
Cirripedia	2.3	0.7	0.0	0.1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Penaeoidea	7.0	3.4	0.0	1.0	0.0	0.0	0.0	0.0	2.2	3.0	0.1	0.1	0.0	0.0	0.0	0.0
<i>Penaeus</i> spp.	7.0	3.4	0.0	1.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Solenocera</i> spp.	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	2.2	3.0	0.1	0.1	0.0	0.0	0.0	0.0
Caridea	27.9	16.6	0.2	9.1	21.2	24.3	1.0	8.9	43.5	86.0	0.9	38.5	47.6	48.3	0.5	22.9
Caridea (N.I.)	18.6	11.0	0.1	7.8	15.2	18.9	0.9	8.5	28.3	69.0	0.4	36.3	28.6	29.2	0.3	17.7
Pandalidae	2.3	0.7	0.0	0.1	3.0	1.4	0.0	0.1	8.7	11.0	0.4	1.8	4.8	3.4	0.0	0.3
<i>Processa</i> spp.	7.0	4.8	0.0	1.2	3.0	4.1	0.0	0.3	4.3	4.0	0.1	0.3	14.3	15.7	0.2	4.8
<i>Alpheus glaber</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	2.2	2.0	0.0	0.1	0.0	0.0	0.0	0.0
Anomura	34.9	19.3	1.3	14.8	3.0	2.7	0.3	0.3	10.9	8.0	2.6	1.1	23.8	9.0	2.6	3.1
Anomura (N.I.)	2.3	0.7	0.0	0.1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Galathea</i> spp.	9.3	3.4	0.0	1.7	0.0	0.0	0.0	0.0	4.3	4.0	0.0	0.3	14.3	6.7	0.5	2.2
<i>Munida</i> spp.	23.3	15.2	1.3	13.0	3.0	2.7	0.3	0.3	6.5	4.0	2.6	0.8	9.5	2.2	2.1	0.9
Brachyura	41.9	14.5	2.4	5.7	3.0	1.4	0.0	0.1	34.8	21.0	1.8	3.9	14.3	7.9	0.2	1.5
Brachyura (N.I.)	9.3	2.8	0.8	1.7	3.0	1.4	0.0	0.1	15.2	9.0	0.4	2.6	4.8	1.1	0.0	0.1
<i>Atelecyclus rotundatus</i>	9.3	4.8	0.9	2.0	0.0	0.0	0.0	0.0	8.7	5.0	0.8	0.9	0.0	0.0	0.0	0.0
Portunidae	2.3	0.7	0.0	0.1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Bathynectes</i> spp.	4.7	1.4	0.2	0.4	0.0	0.0	0.0	0.0	2.2	3.0	0.3	0.1	0.0	0.0	0.0	0.0
<i>Liocarcinus</i> spp.	7.0	2.1	0.3	0.9	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Polybius henslowii</i>	2.3	0.7	0.0	0.1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Pilumnus</i> spp.	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	2.2	1.0	0.1	0.0	0.0	0.0	0.0	0.0
<i>Pilumnus hirtellus</i>	4.7	1.4	0.1	0.4	0.0	0.0	0.0	0.0	2.2	1.0	0.0	0.0	9.5	6.7	0.2	1.4
<i>Monodaeus couchii</i>	2.3	0.7	0.0	0.1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Goneplax rhomboides</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	2.2	1.0	0.1	0.0	0.0	0.0	0.0	0.0
<i>Pisa</i> spp.	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	2.2	1.0	0.1	0.0	0.0	0.0	0.0	0.0
Pisces	83.7	35.2	95.8	63.4	84.8	58.1	98.5	84.3	80.4	46.0	94.4	55.9	81.0	32.6	96.7	72.3
Teleostei (N.I.)	25.6	9.7	4.9	14.5	18.2	8.1	5.6	7.0	41.3	22.0	8.4	23.2	28.6	7.9	3.7	6.9
<i>Conger conger</i>	4.7	1.4	0.4	0.4	3.0	2.7	0.0	0.2	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Sardina pilchardus</i>	4.7	1.4	1.0	0.5	9.1	5.4	7.1	3.2	2.2	1.0	2.1	0.1	4.8	6.7	22.6	2.9
<i>Phycis</i> spp.	0.0	0.0	0.0	0.0	6.1	2.7	22.2	4.3	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Merluccius merluccius</i>	0.0	0.0	0.0	0.0	3.0	1.4	3.7	0.4	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Micromesistius poutassou</i>	4.7	1.4	2.2	0.5	3.0	1.4	0.2	0.1	4.3	2.0	2.7	0.4	0.0	0.0	0.0	0.0
<i>Trisopterus luscus</i>	34.9	13.8	43.7	45.5	30.3	25.7	51.9	66.2	19.6	11.0	73.6	30.6	38.1	15.7	60.6	61.2
<i>Capros aper</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	6.5	4.0	7.0	1.3	0.0	0.0	0.0	0.0
<i>Serranus</i> spp.	2.3	0.7	0.3	0.1	3.0	1.4	0.0	0.1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Serranus cabrilla</i>	2.3	0.7	0.2	0.1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Diplodus</i> spp.	0.0	0.0	0.0	0.0	3.0	1.4	1.7	0.3	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Mullus surmuletus</i>	2.3	0.7	8.7	0.4	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Labridae	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	2.2	1.0	0.3	0.1	0.0	0.0	0.0	0.0
<i>Acantholabrus palloni</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	4.8	1.1	9.6	1.1
<i>Symphodus melops</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	2.2	1.0	0.3	0.1	0.0	0.0	0.0	0.0
Ammodytidae	0.0	0.0	0.0	0.0	6.1	8.1	6.1	2.4	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
<i>Pomatoschistus</i> spp.	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	2.2	4.0	0.1	0.2	0.0	0.0	0.0	0.0
<i>Scomber colias</i>	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	4.8	1.1	0.2	0.1
<i>Scomber scombrus</i>	2.3	5.5	34.4	1.4	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0



**Fig. 2.** Dendrogram resulting from the hierarchical cluster analysis of the mean abundance of each prey group by forkbeard (*Phycis phycis*) length class of 5 cm using Ward's method and Manhattan distance. Results from Similarity Profile Analysis are defined with different line types. LG1 (dashed line) <22.5 cm; LG2 (dot line) 27.5–37.5 cm; LG3 (solid line) >42.5 cm.

three length groups, henceforward referred to as LGs, were obtained. To test the number of statistical different clusters produced by the cluster analysis, a Similarity Profile Analysis (Simprof) was conducted.

The non-parametric Kruskal–Wallis ANOVA and Mann–Whitney *U*-test were used to explore significant statistical differences in stomach fullness index by LG and season, and by sex, respectively. A three-factor crossed design permutational MANOVA (PERMANOVA) (Anderson, 2005) based on the Bray–Curtis distance measure was used to investigate statistical differences for the abundance of prey (diet) by LG, season, and sex. Since empty stomachs provide no valuable information for this analysis they were excluded. A simpler routine was performed to identify the most important species to discriminate among groups. The information on LG1 was also excluded since there were too few individuals in this group.

Feeding strategy by LG was analysed according to the graphical representation suggested by Cortés (1997) using the most representative prey items aggregated by the major taxonomic group.

All statistical analyses were executed in R environment (R Core Team, 2015) with packages clustsig (Whitaker & Christman 2015), vegan (Oksanen, 2011) and scatterplot3d (Ligges & Mächler, 2003).

## RESULTS

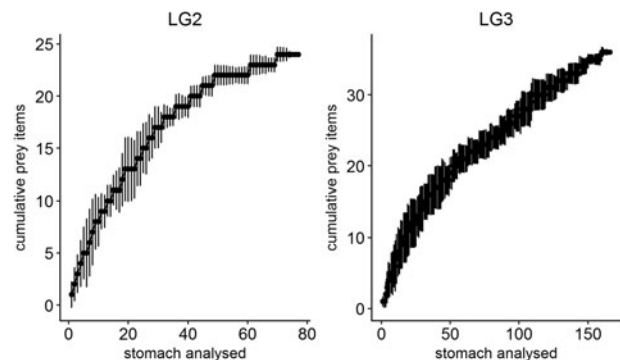
A total of 521 specimens were sampled but 275 had everted stomachs, representing 53% of the total. The remaining 246 individuals were used for the diet study (144 females and 102 males) and ranged between 15.5 and 67.1 cm TL. Specimens were captured mainly by longline at depths ranging between 180 and 310 m. This gear also captured the largest individuals while trawl captured the smallest ones (Table 1).

## Feeding patterns

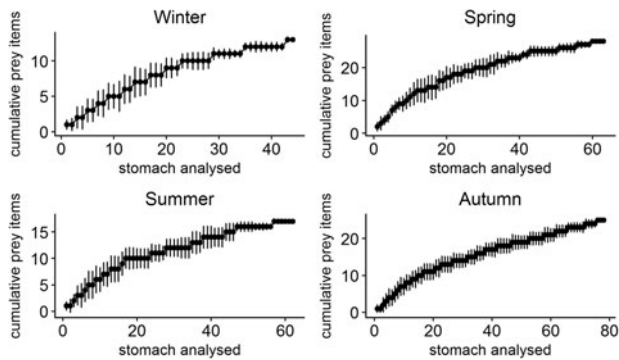
A total of 44 prey items were identified in the stomachs of the forkbeard (Tables 2 and 3). The species showed a high vacuity index with 42.1% of empty stomachs.

The cluster analysis indicated the presence of three groups which are statistically distinct (Figure 2): LG1, <22.5 cm ( $N = 4$ ); LG2, 27.5–37.5 cm ( $N = 77$ ); and LG3, >42.5 cm ( $N = 166$ ).

Table 2 shows the diet of forkbeard by length group. LG1 was represented by four individuals that fed mainly on Caridea and secondly on Pisces, especially *Pomatoschistus* spp. No indices were calculated for LG1 due to the low number of specimens involved. LG2 individuals presented a diet where Caridea, Anomura and Brachyura were the most consumed prey with the highest %N and %O. Finally, LG3 specimens fed mainly on Pisces, especially the pout *Trisopterus luscus* (Linnaeus, 1758), and secondly on Brachyura. Analysing the forkbeard diet by season (Table 3), Pisces were the most important feeding item in all seasons, seen by the



**Fig. 3.** Randomized cumulative prey curves for the forkbeard *Phycis phycis* samples by length group (LG). Mean values are plotted and error bars represent  $\pm$  SE.



**Fig. 4.** Randomized cumulative prey curves for the forkbeard *Phycis phycis* samples by season. Mean values are plotted and error bars represent  $\pm$  SE.

high values of IRI, while *T. luscus* was relevant in spring, summer and winter. Other important items were Caridae, showing high values of IRI in autumn and winter, non-decapod Crustacea in the summer and Brachyura in spring.

Cumulative prey curves were applied by LG (Figure 3) (except for LG1 that presented a low number of stomachs) and season (Figure 4) and the number of stomachs was considered adequate to describe the diet of the forkbeard. Table 4 shows the variability (CV) and the departure from zero slope of the four last points. The CV near 0 for both groups and seasons and the slope statistically equal to 0 ( $P > 0.1$ ) showed that the number of prey sampled in the last stomach reached stability.

SFI show significant differences between LGs ( $W = 5261.0$ ,  $P = 0.021$ ), but not among seasons ( $H = 128.2$ ,  $P = 0.773$ ) and sex ( $W = 8067.5$ ,  $P = 0.092$ ).

PERMANOVA revealed significant differences for the forkbeard diet among season and LG but no differences between sexes were found (Table 5). Discrimination among seasons and between LG was mainly due to Pisces, Caridea and *Trisopterus luscus*.

The three-dimensional graphical representation of the feeding strategy (Figure 5) reinforces the different food preferences of the forkbeard by group size. In LG2, Caridea present some significance both in occurrence and number and a particular fish species, the pout *Trisopterus luscus*, appears with an increasing importance in terms of biomass. The LG3 was the least generalist group, with Pisces and *Trisopterus luscus* being the main prey items.

## DISCUSSION

Diet and feeding strategy is quite difficult to assess since a large number of samples is needed to correctly evaluate the

**Table 4.** Results of the cumulative curves applied to the different length groups and seasons (b, last four points slope; t, t-test value; P, probability; CV, mean coefficient of variation and standard deviation of the four last points).

	b	t	P	CV
LG2	0.07 $\pm$ 0.03	2.65	0.117	0.003 $\pm$ 0.007
LG3	0.12 $\pm$ 0.05	2.77	0.110	0.005 $\pm$ 0.007
Spring	0.22 $\pm$ 0.08	2.70	0.114	0.005 $\pm$ 0.009
Summer	0.12 $\pm$ 0.04	2.67	0.116	0.006 $\pm$ 0.012
Autumn	0.12 $\pm$ 0.04	2.67	0.116	0.004 $\pm$ 0.008
Winter	0.20 $\pm$ 0.09	2.90	0.101	0.023 $\pm$ 0.009

**Table 5.** Statistical results for *Phycis phycis* diet comparison among sex, length groups and seasons with permutational multivariate analysis of variance.

	Fmodel	R <sup>2</sup>	Pr(>F)	df
Sex	1.0576	0.0067	0.3267	1
TL	<b>15.2701</b>	<b>0.0972</b>	<b>0.0010</b>	3
Season	<b>1.9356</b>	<b>0.0370</b>	<b>0.0410</b>	1
Residuals		0.8591		135
Total		1.0000		140

Formula = Diet  $\sim$  Sex + LG + Season, permutations = 1000; Pr(>F) – statistic P value; df, degrees of freedom. Significant values are given in bold.

species diet. In the present study, information was gathered using all fishing gears that capture the forkbeard. Longline, which operates at greater depths than bottom trawl and trammel nets, was the most important gear for capturing forkbeard. This can explain the high percentage of everted stomachs (similar to the one found by Morato *et al.*, 1999) most probably caused by decompression shock. Also, the high percentage of empty stomachs found (42.1%), higher than those reported for the forkbeard from Azorean waters (Morato *et al.*, 1999) and for *Phycis blennoides* (Brünnich, 1768) in the western Mediterranean (Morte *et al.*, 2002), might be a result of regurgitation caused by stress, common in fishes caught by hook (Morato *et al.*, 1999).

The longline may also affect the quality of stomach contents as the hauling can take several hours and the contents may become completely digested and non-identifiable (Morato *et al.*, 1999). This fishing gear is also size selective and does not capture the smaller specimens (Vieira *et al.*, 2014a), which explains the lack of small individuals in the study.

Cumulative prey curves were applied by LG and season with exception of LG1, due to the small number of specimens caught. However, we maintained LG1 data in the comparative analysis due to the lack of feeding information that exists for forkbeard smaller than 24 cm. In fact, Morato *et al.* (1999) sampled individuals larger than 24 cm and Papaconstantinou & Caragitsou (1989) sampled specimens larger than 15 cm but they did not mention their number.

The forkbeard is a demersal active predator that can be generally described as having a diet consisting predominantly of teleost fishes and decapod crustaceans, as Morato *et al.* (1999) and Papaconstantinou & Caragitsou (1989) have already reported. However, differences can be found considering the season and fish length.

According to the results of the present study, there was a significant difference in the forkbeard diet between seasons. In some species, high frequency occurrence index values are associated with a peak of abundance of the prey item in the environment as a result of a recent reproduction event of that prey item (Morte *et al.*, 2002), possible migratory movements among prey and its predator, temporal changes in water column productivity (Cartes *et al.*, 2008), and physico-chemical variables and biological features of the species (Fanelli & Cartes, 2008). During the whole year, Pisces is by far the most important food item but differences can be noted on other important items. In the autumn and winter, Caridea group represent an important part of the diet, eventually because of a decrease in the abundance of *Trisopterus*

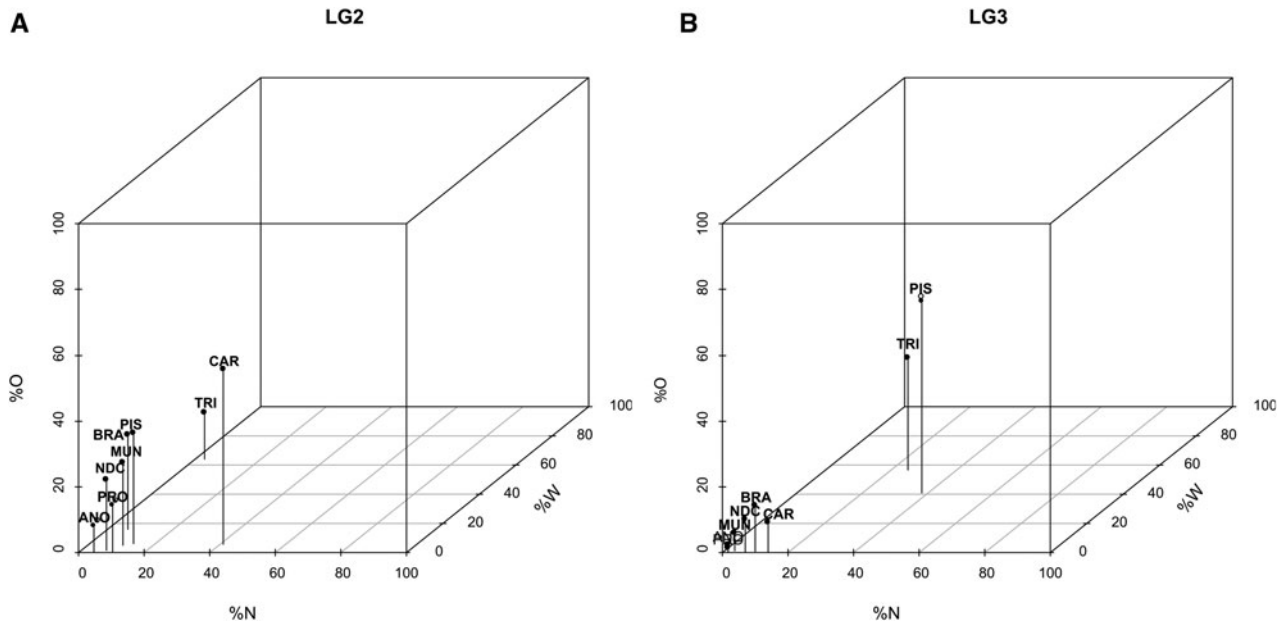


Fig. 5. Three-dimensional representation of feeding strategy of the forkbeard *Phycis phycis* by total length groups (LG). (A) LG2, (B) LG3. NDC, non-decapod Crustacea; CAR, Caridea; ANO, Anomura; MUN, *Munida* spp.; PRO, *Processa* spp.; BRA, Brachyura; PIS, Pisces; TRI, *Trisopterus luscus*.

*luscus*, especially in autumn. In spring and summer, there is an increase in consumption of *T. luscus*, which has a reproductive cycle during the whole year but with a peak in spring (Alonso-Fernández *et al.*, 2008). On the other hand, the fact that the forkbeard reproduces between September and January (Vieira *et al.*, 2016b) can also explain the decrease in feeding activity during these months of the year, as occurs in the other gadiformes such as the silver hake *Merluccius bilinearis* (Bowman, 1984).

Besides the diet variation between seasons, the forkbeard diet varied with fish size. The feeding strategy analysis confirmed the increasing consumption of Teleost and a decreasing consumption of the other prey groups, such as Caridea, with increase of fish length, which might be related to the large prey size preference as the predator grows (Morato *et al.*, 1999; Carrassón & Cartes, 2002). Furthermore, the depth where the individuals of different LG occur coincides with the depth of the most important prey in each group. Similarly to *Phycis chesteri* Goode & Bean, 1878 (Wenner, 1983), smaller forkbeard specimens were found in the shallower part of the depth distribution range. Larger forkbeard specimens were found at higher depths (Wenner, 1983) where they predate species such as conger eel *Conger conger* (Linnaeus, 1758), blue whiting *Micromesistius poutassou* (Risso, 1827) and European hake *Merluccius merluccius* (Linnaeus, 1758). Individuals belonging to LG2 eventually make the transition between LG1 and LG3, with a diet including items from the two adjacent length groups.

According to the results of the present study, no significant differences were found for the forkbeard diet between sexes. The same results were also obtained by Morato *et al.* (1999) for forkbeard from Azorean waters.

Although SFI did not show significant differences among sex, season and LGs, it is important to note that there was an increase in the mean SFI values as fish became larger. The absence of significant differences may be related to the great variability of the data particularly the small number of

individuals caught in LG1. The positive relationship between SFI and fish length was found in ambush piscivorous species like the *Cygnodraco mawsoni* Waite, 1916 (Pakhomov, 1988), however a standard behaviour for all piscivorous species is difficult to find. In fact, in cod *Gadus morhua* Linnaeus, 1758 both a decreasing and increasing trend of SFI with length could be found (Lilly, 1983) and in bluemouth *Helicolenus dactylopterus* (Delaroche, 1809), it was the middle length class specimens that presented the highest SFI values (Neves *et al.*, 2011).

The results of this study demonstrated the change in diet preference of the forkbeard during its life cycle, which is conditioned by their growth, the season and the depth. Forkbeard feeding behaviour may be characterized as presenting a shift pattern from a more generalist diet (small Crustacea, mainly Caridea) in the young adults to a more specialist strategy (teleosts) in the adults. This shift of feeding pattern has also been related to the different energy content and evacuation rate between crustacean and fish prey (Andersen, 1999).

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