

# A possible biogenic origin for hydrogen peroxide on Mars: the Viking results reinterpreted

Joop M. Houtkooper<sup>1</sup> and Dirk Schulze-Makuch<sup>2</sup>

<sup>1</sup>Center for Psychobiology and Behavioral Medicine, Justus-Liebig-University of Giessen, Otto-Behaghel-Strasse 10F, D-35394 Giessen, Germany

e-mail: joophoutkooper@gmail.com

<sup>2</sup>School of Earth and Environmental Sciences, Washington State University, Pullman, WA 99164, USA

e-mail: dirksm@wsu.edu

**Abstract:** The adaptability of extremophiles on Earth raises the question of what strategies putative life might have used to adapt to the present conditions on Mars. Here, we hypothesize that organisms might utilize a water–hydrogen peroxide ( $\text{H}_2\text{O}$ – $\text{H}_2\text{O}_2$ ) mixture rather than water as an intracellular liquid. This adaptation would have the particular advantages in the Martian environment of providing a low freezing point, a source of oxygen and hygroscopicity. The findings by the Viking experiments are reinterpreted in light of this hypothesis. Our conclusion is that the hitherto mysterious oxidant in the Martian soil, which evolves oxygen when humidified, might be  $\text{H}_2\text{O}_2$  of biological origin. This interpretation has consequences for site selection for future missions to search for life on Mars.

Received 3 November 2006, accepted 29 March 2007

**Key words:** Mars, extremophile, hydrogen peroxide,  $\text{H}_2\text{O}_2$ – $\text{H}_2\text{O}$  hypothesis, Phoenix, oxidant, hyperhydration, Viking.

## A different kind of biochemistry?

The Viking Landers on Mars remain the only direct attempt to detect life on another world. All three experiments conducted by the Landers observed chemical changes that indicated the possible presence of life, although the expected signals were not as large as expected for a biological response and tapered off with time, casting doubt on a biological explanation. This led to a consensus view that Viking had detected reactive, oxidizing surface chemistry, but not biochemical metabolic processes (Klein 1999).

We propose here a reinterpretation of the Viking results, based on the assumption that microorganisms on Mars produce hydrogen peroxide to generate an  $\text{H}_2\text{O}$ – $\text{H}_2\text{O}_2$  intracellular solvent for biochemical processes selected by and adapted to the unique Martian environment.

Current environmental conditions near the surface of Mars are not incompatible with life. Various survival studies exposing terrestrial microbes to simulated near-surface conditions on Mars have revealed remarkably high survival rates below very shallow soil (Cockell *et al.* 2005; Diaz & Schulze-Makuch 2006). With regards to oxidant tolerance, some soil bacteria survive and grow to the stationary phase in 30 000 ppm  $\text{H}_2\text{O}_2$  (Mancinelli 1989). McDonald *et al.* (1998) tested the stability of organic macromolecules subjected to oxidation stress by 30%  $\text{H}_2\text{O}_2$  in water at three different temperatures relevant to Martian environmental conditions. Their data suggested that some organic macromolecules are

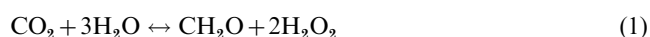
stable against oxidation on the Martian surface, at least in the polar regions, over the entire history of Mars. Mixtures of  $\text{H}_2\text{O}_2$  and  $\text{H}_2\text{O}$  would have remarkably useful properties for any organism in need of adapting to Martian environmental conditions.

Mixtures of  $\text{H}_2\text{O}_2$  and  $\text{H}_2\text{O}$  freeze at temperatures significantly below the freezing point of water. The lower eutectic point lies at  $-56.5^\circ\text{C}$  for a mixture with 61.2 wt%  $\text{H}_2\text{O}_2$  (Foley & Giguère 1951). Mixtures with a high  $\text{H}_2\text{O}_2$  concentration tend to supercool, sometimes resulting in the formation of glasses, down to liquid-air temperatures (Giguère & Secco 1954). Thus, putative Martian organisms could stay completely functional at temperatures far below the freezing point of water and even survive lower temperatures as the formation of ice crystals and piercing of cellular membranes would be prevented.  $\text{H}_2\text{O}_2$ – $\text{H}_2\text{O}$  mixtures are slightly acidic, with a pH of 4.5 for the 60 wt% mixture. Owing to the lower water vapour partial pressure in equilibrium with the liquid,  $\text{H}_2\text{O}_2$ – $\text{H}_2\text{O}$  mixtures tend to be hygroscopic compared with water, which would offer the opportunity for an organism to scavenge water molecules from the Martian atmosphere. These considerations point to at least the possibility that organisms might use  $\text{H}_2\text{O}_2$ , not only in storage as a convenient source of oxygen, but also as a major component of their intracellular fluid.

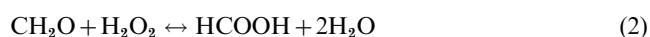
An intracellular  $\text{H}_2\text{O}_2$ – $\text{H}_2\text{O}$  mixture would not only provide a source of oxygen and be a favourable adaptation to low temperatures, but also convey hygroscopic abilities to the

putative Martian organisms. At little metabolic cost they would be able to scavenge the atmosphere for the little water vapour present. It has been suggested that some lichens on Earth may also survive on water vapour only (Rothschild 2007). On the other hand, this hygroscopic ability would mean vulnerability for exposure to liquid water. The organisms would be susceptible to death by hyperhydration. This could occur when the organisms are exposed to liquid water or even to a relatively warm atmosphere saturated with water vapour. At death, the cellular contents would be set free releasing O<sub>2</sub> as well as organic compounds. Furthermore, an exothermic reaction between the H<sub>2</sub>O<sub>2</sub> and the organics is likely, resulting in ultimate transformation into CO<sub>2</sub>, O<sub>2</sub> and water vapour (with some nitrogen and minor constituents).

The H<sub>2</sub>O<sub>2</sub> could be produced biochemically by the organisms themselves, through energy obtained from sunlight. A gross metabolic pathway could follow



The equation would proceed to the right using sunlight as an energy source and to the left in darkness or when work is exerted. However, when exchange with the environment is to be avoided, the sugars could be oxidized to formic acid,



However, this would quickly acidify the cell, in a similar manner to lactic acid in terrestrial animals. In an alternative reaction, H<sub>2</sub>O<sub>2</sub> could serve as a source of energy by simply decomposing into water and oxygen:



This reaction, though, is an energetically less efficient use of the H<sub>2</sub>O<sub>2</sub>, which has been produced at great metabolic cost.

Based on the freezing point of H<sub>2</sub>O–H<sub>2</sub>O<sub>2</sub> mixtures, the organisms would be well adapted if their intracellular fluid contains a substantial concentration of H<sub>2</sub>O<sub>2</sub>. This presumes that high amounts of intracellular H<sub>2</sub>O<sub>2</sub> would not require too high a cost in energy for the production of stabilizing compounds. Metabolic activity could theoretically occur at temperatures down to –56 °C (217 K) if a eutectic mixture is used. Temperatures even lower than the freezing point of the H<sub>2</sub>O<sub>2</sub>–H<sub>2</sub>O eutectic could also be tolerated because of likely supercooling of the liquid. An upper temperature limit depends on the stabilization mechanism, which must consume energy at a faster rate at high temperatures. As the cell contents are energy-rich, higher temperatures than those that usually occur on Mars may be withstood for only brief periods of time.

### The Viking life detection experiments

While the life detection experiments conducted by the Viking Landers were generally interpreted as a failure to detect life based on the biochemistry of microorganisms on Earth, doubts and inconsistencies about those results remain. In particular, the following doubts have been raised.

Table 1. Data from the PR experiment (Horowitz et al. 1976). The ‘Conditions’ column indicates whether the lamp was on or off, whether or not water vapour was injected and whether the soil sample was heat-sterilized (control is 175 °C for 3 h). The radioactivity of the Peak 2 column represents organic matter synthesized from the labelled gases.

Experiment	Conditions	Peak 2 (count min <sup>-1</sup> )
Chryse 1	Light, dry, active	96 ± 1.15
Chryse 2	Light, dry, control	15 ± 1.29
Chryse 3	Light, dry, active	27 ± 0.98
Chryse 4	Light, dry, active	35 ± 1.6
Utopia 1	Dark, dry, active	23 ± 1.7
Utopia 2	Light, wet, active	2.8 ± 0.92
Utopia 3	Dark, dry, active	7.5 ± 2.5

- (1) While no organic molecules were detected by gas chromatography–mass spectrometry (GC–MS), the requisite sensitivity may not have been achieved at the time.
- (2) Chemical explanations for the Viking Lander experiments (particularly the evolution of O<sub>2</sub> upon wetting) require a strong oxidizer at sufficiently high concentration, which has still not been identified.
- (3) There is no satisfactory explanation for the 30% rise in CO<sub>2</sub>, the near doubling of N<sub>2</sub> or the surprisingly large rise of O<sub>2</sub>, from 4 nmol to about 520 nmol, in the gas exchange (GEx) experiment (Oyama & Berdahl 1977).
- (4) No convincing mechanism had been proposed for the small but significant synthesis of organic material in the pyrolytic release (PR) experiment (Table 1). This amount could not come from the synthesis by ultraviolet radiation because an optical filter to screen out the ultraviolet wavelengths below 320 nm was included in the experiment.
- (5) The production of gas recorded from the labelled release (LR) nutrient when it was placed on Martian soil at both Lander sites was significant (Levin & Straat 1977). Decreases of released gas were observed at secondary injections. The reactant in the Mars soil was completely unreactive at the sterilizing temperature of 160 °C. In contrast, exposure to 18 °C for two Martian days did not inhibit the reaction.

Nevertheless, the consensus view was that the Viking life detection experiments detected reactive chemistry rather than biology, which was based on a convergence of (1) the evolution of O<sub>2</sub> upon wetting the soil, (2) the apparent absence of organic molecules in the soil and (3) the weakly positive result of the single control test in the PR experiment. With regard to the latter, the PR experiment was designed to detect carbon assimilation. The first test, Chryse 1, revealed a small but very significant incorporation of radioactive carbon in the pyrolyzable organic fraction of the sample (Table 1). Unfortunately, only one control experiment was performed after the first test. In this control test (Chryse 2), in which a sample was heated for 3 h at 175 °C, the reaction was absent (Klein *et al.* 1976) or diminished by 88% (Horowitz 1986). The test result from the control sample was 3.5σ above the baseline

expectation based on previously conducted laboratory tests on terrestrial soils (Horowitz *et al.* 1976). Although the incorporation of radioactive carbon means that a presumptive biological reaction has been found, such activity also occurring in a sterilized sample would falsify the biological hypothesis. However, the baseline was derived only from a limited number of tests ( $N=39$ , with one greater than 5 $\sigma$  outlier being rejected), so that not a normal but at least a Student *t*-distribution would be appropriate to assess the statistical likelihood. Moreover, it is questionable whether these baseline tests were representative for the Martian environment and whether, in fact, the whole sample in this single case was heated to 175 °C.

With regard to the control tests by the Viking experiments, we now know that some hyperthermophiles on Earth thrive at a temperature of at least 121 °C (Kashefi & Lovley 2003). The temperatures to sterilize the soil samples in the Viking control tests were 145, 160 and 175 °C, in the GEx, LR and PR experiments, respectively. There is no doubt that concentrated H<sub>2</sub>O<sub>2</sub>, by itself, would decompose quickly at such an elevated temperature. However, if H<sub>2</sub>O<sub>2</sub> is of biological nature, it must be assumed that it is situated in a particular biochemical environment created by biochemical processes. The closest Earth analogy would probably be the processes occurring in the peroxisomes of most eukaryotic cells (De Duve 1969). Radiation tolerance as a side effect of desiccation tolerance in bacteria is generally assumed based on pioneering work on *Deinococcus radiodurans* by Mattimore & Battista (1996). Although speculative, the possibility should be considered that H<sub>2</sub>O<sub>2</sub> as a major component of an intracellular fluid may require and has co-evolved with stabilization mechanisms, which have a side effect on temperature tolerance.

Temperature tolerance may have played a role in the evolution of O<sub>2</sub> in the GEx experiment. In the active tests, oxygen would be a metabolite, concomitant with the (auto-)oxidation of the organic cellular contents. Viable organisms may well have started to decompose at or above the freezing point of water and 100% moisture. The response in the two control tests of the GEx experiment was in one case diminished to about a third, in the other to about zero (Oyama & Berdahl 1977). The diminished positive result in the first control test could mean that H<sub>2</sub>O<sub>2</sub>-based organisms do not decompose completely at 145 °C, if, in fact, that temperature was imposed on the whole sample. Therefore, the results of the GEx experiment and the evolution of O<sub>2</sub> in particular are compatible with our hypothesis.

The failure to detect organic molecules by the GC-MS during the Viking mission to Mars was surprising, especially because some  $2.4 \times 10^8$  g of reduced carbon falls on Mars each year via asteroids, comets and other planetary material (Flynn 1996). The common assumption is that all of the organic material near the surface is oxidized by H<sub>2</sub>O<sub>2</sub> and other strong oxidizing compounds. In addition, a recent analysis indicates that the sensitivity of the Viking GC-MS was much less than originally thought owing to interference with minerals in the Martian soil and other factors (Navarro-González *et al.* 2006). Based on the reactivity of the surface

measured by the Viking GEx experiment, the amount of H<sub>2</sub>O<sub>2</sub> on the Martian surface was estimated to be between 1 ppm (Zent & McKay 1994) and 250 ppm (Mancinelli 1989). However, photochemical processes generate H<sub>2</sub>O<sub>2</sub> in the atmosphere at a much lower rate in the parts per billion range. Atmospheric H<sub>2</sub>O<sub>2</sub> abundances vary between 20 and 40 ppb by volume over the planet (Encrenaz *et al.* 2004), which appears to be a maximum concentration occurring during favourable weather conditions (Atreya & Gu 1994). Thus, there is a case to be made not only for the missing organics but also for the missing H<sub>2</sub>O<sub>2</sub>.

The biological explanation for the lack of detected organics by the GC-MS could be that the oxidizing inventory of the H<sub>2</sub>O<sub>2</sub>-H<sub>2</sub>O solvent well exceeded the reducing power of the organic compounds of the organisms. Upon heating, therefore, the putative organisms might have auto-oxidized catastrophically, leaving the gases as detected by the GEx experiment plus very little solid residue without or with only very little organic content. The negative result of the GC-MS (Biemann *et al.* 1977) is therefore not a very reliable estimate of an upper bound on the biomass in the soil. A reasonable alternative explanation of the missing organics using a purely chemical explanation was advanced by Benner *et al.* (2000), who suggested that any organics on the surface of Mars would undergo a diagenesis to metastable compounds of carboxylic acids derivatives and would not be easily detected by the GC-MS. Explanations to the five questions presented above in terms of the H<sub>2</sub>O<sub>2</sub>-H<sub>2</sub>O hypothesis and traditional chemical explanations are provided in Table 2.

The fact that O<sub>2</sub> evolved from soil samples upon humidification deserves particular scrutiny in order to evaluate whether this is in accordance with a biological rather than a chemical origin. The release of O<sub>2</sub> was a surprise to the mission scientists, who then became convinced of a chemical origin as terrestrial life is not known for originating O<sub>2</sub> upon wetting. Our interpretation, however, is that under a dry or slightly humid atmosphere the release of oxygen would be a consequence of a metabolic pathway of putative H<sub>2</sub>O<sub>2</sub>-H<sub>2</sub>O-based Martian organisms (reaction (3)), while under wet conditions it would indicate the decomposition of organisms by hyperhydration after being exposed to excess water. A similar inference can be drawn from the PR experiment. Wetting all but inhibited any organic synthesis reaction in the Utopia 2 sample and the following Utopia 3 sample (Table 1), while organic synthesis reactions did occur under dry conditions (Chryse 1, and possibly to some minor degree also for Chryse 3 and 4, and Utopia 1). The mission scientists had difficulty explaining this phenomenon. Horowitz *et al.* (1977) called it 'startling', while Klein (1978) felt that an explanation for this phenomenon 'remains obscure'.

With regards to a chemical explanation for these findings, the past 30 years have seen numerous proposals of what the putative oxidants might be without reaching a definitive solution. Therefore, we feel that after 30 years of chemical indecision it is appropriate to propose a biological hypothesis.

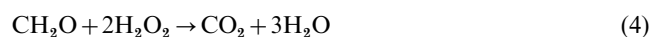
Not all results can be satisfactorily explained with our hypothesis. For example, the differences in amplitude of the

Table 2. *Explanations for some remaining questions after Viking.*

Question	Chemical explanation	H <sub>2</sub> O <sub>2</sub> –H <sub>2</sub> O hypothesis
Lack of identified organic molecules	The organics have been oxidized to non-volatile salts of benzenecarboxylic acids, and perhaps oxalic and acetic acid (Benner <i>et al.</i> 2000).	Upon death of the organisms, the organics are spontaneously oxidized by the previously intracellularly bound H <sub>2</sub> O <sub>2</sub> with no or very little organic residue. Non-biology bound organic molecules are oxidized chemically (Benner <i>et al.</i> 2000) and/or consumed by organisms. The release of 50–700 ppm of CO <sub>2</sub> by the Viking GC–MS may indicate that oxidation of organic material took place (Navarro-González <i>et al.</i> 2006).
Lack of identified oxidant	There is some yet unidentified mechanism producing H <sub>2</sub> O <sub>2</sub> or other oxidants. The oxidant might be present in the form of a compound that has no analogue on Earth. Suggested inorganic oxidants include metal oxides such as Fe- and Ti-oxides (Quinn & Zent 1999) and superoxide ions (Yen <i>et al.</i> 2000).	The H <sub>2</sub> O <sub>2</sub> in the H <sub>2</sub> O <sub>2</sub> –H <sub>2</sub> O mixture is part of the biochemistry of the putative Martian organisms. It would explain the oxidizing potential observed in the Viking results. However, some soil chemistry reactions certainly play a role in the response to the Viking Lander experiments as well.
Release and partial resorption of O <sub>2</sub> , CO <sub>2</sub> and N <sub>2</sub> in the GEx experiment	Evolution of O <sub>2</sub> upon humidification was suggested to involve one or more reactive species such as ozonides, superoxides and peroxides (Oyama & Berdahl 1977). CO <sub>2</sub> production in the wet mode can be interpreted to be related to the oxidation of nutrient organic compounds (Oyama <i>et al.</i> 1977), and N <sub>2</sub> release can be interpreted to be related to an initial N <sub>2</sub> desorption from soil by water vapour and subsequent resorption in liquid water (Oyama <i>et al.</i> 1977).	The release of O <sub>2</sub> (and possibly CO <sub>2</sub> to a lesser degree) can be interpreted as the result of an energy-producing metabolism. Upon humidification it could also point to the decomposition of dying Martian biota, as could the increase of N <sub>2</sub> . The decrease of N <sub>2</sub> can be understood as biological fixation if it exceeded the amount due to physical sorption, a possibility also entertained by Oyama <i>et al.</i> (1977).
Synthesis of organic material in PR experiment	No consistent explanation has been provided, but attempts to explain the observations include instrument malfunction, incorporation of <sup>14</sup> C into carbon suboxide polymer preformed on the Martian surface and reduction of <sup>14</sup> C by H <sub>2</sub> O <sub>2</sub> in the surface material (Horowitz <i>et al.</i> 1977).	Some of the putative organisms were able to metabolize and synthesize organic compounds before they died being overwhelmed by water.
Responses in the LR experiment	Laboratory tests on Earth using inorganic oxidants and clay minerals simulated many of the key findings, but not the decrease of responses after storage at elevated temperatures (Klein 1999).	Limited metabolism (Levin & Straat 1977, 1981) before the organisms died due to hyperhydration, osmotic pressure and/or heat shock.

response of the PR experiment remain a puzzle. However, this might be understood by assuming that the Martian surface is not covered by a homogeneous population of organisms. The apparent difference in heat resistance observed in some of the Viking experiments may also be due to a heterogeneous microbial population. In addition, some chemical reactions certainly play a role in the response to the Viking Lander experiments. From the perspective of the H<sub>2</sub>O<sub>2</sub>–H<sub>2</sub>O hypothesis on Martian life, the Viking experiments were both too warm and too wet. In particular, the combination of high temperatures (relative to average Martian conditions) and saturation with water vapour represent extremely un-Martian conditions and both the GEx and the LR experiments employed these conditions. The results of these two experiments might best be explained by a gradual failure of heterotrophic metabolism (which the experiments were designed to detect) to cope with adverse conditions. The putative Martian organisms were overwhelmed by too much water vapour, a condition against which they had no defence, so that they failed because of a fatal rise in osmotic pressure.

If we assume H<sub>2</sub>O<sub>2</sub>–H<sub>2</sub>O-based life as a working hypothesis, we may roughly estimate the biomass fraction of Martian soil. Like Mancinelli (1989), we derive an estimate of H<sub>2</sub>O<sub>2</sub> concentrations from the amounts of evolved gases in the GEx experiment. Here we assume the evolved O<sub>2</sub> to originate from the decomposition of biogenic H<sub>2</sub>O<sub>2</sub>, but we also make the assumption that the evolved CO<sub>2</sub> originated in the auto-oxidation of the microbes. The idea that the CO<sub>2</sub> is released by desorption is neglected, because in the LR experiment released CO<sub>2</sub> was absorbed upon further wetting of the samples (Levin & Straat 1976). Also we neglect in our simplified calculations the evolved N<sub>2</sub>. We assume a model composition of the microbes consisting of organic macromolecules (simplified as CH<sub>2</sub>O) and equal weights of H<sub>2</sub>O and H<sub>2</sub>O<sub>2</sub>. The decomposition reactions playing a role are



and



From the GEx experiment, the maximum amounts of evolved gases, from a nominal 1.3 g sample, were 9800 nmol CO<sub>2</sub> and 790 nmol O<sub>2</sub>. From these figures, the molecular ratio of reactions (4) and (5) follows at about 12.4 to 1. This leads to the overall equation



The molecular ratio of CH<sub>2</sub>O to H<sub>2</sub>O<sub>2</sub>, 12.4 to 26.8, leads to 17% of the microbe weight being CH<sub>2</sub>O and 41.5% of the weight being H<sub>2</sub>O<sub>2</sub> and H<sub>2</sub>O, respectively. The evolved CO<sub>2</sub> in the GEx was 9800 nmol, from 1.3 g of soil (Oyama & Berdahl 1977), therefore 7538 nmol g<sup>-1</sup>. This CO<sub>2</sub> is assumed to originate from 7538 nmol g<sup>-1</sup> CH<sub>2</sub>O or 226 μg g<sup>-1</sup> of soil. As CH<sub>2</sub>O constitutes 17% of the biomass, the total biomass is 1330 μg g<sup>-1</sup> or 1330 ppm.

The amount of biogenic H<sub>2</sub>O<sub>2</sub> in our simplified model is 552 ppm, compared with a value of 25–250 ppm of adsorbed or chemically bound H<sub>2</sub>O<sub>2</sub> as calculated by Mancinelli (1989).

In future experiments on Mars, the detection of decomposing microbes might be attempted by measuring the heat produced from the exothermic decomposition reactions. The overall reaction (6) produces an enthalpy of 8454 kJ for 12.4 mol CH<sub>2</sub>O. As the soil contains 7538 nmol g<sup>-1</sup> of CH<sub>2</sub>O, the enthalpy per gram of soil is 5.1 J g<sup>-1</sup> based on the calculations above. These estimates are maximum values because we used the maximum gas evolution recorded by the GEx experiment and further assumed that all of the evolved CO<sub>2</sub> resulted from biological decomposition. Another uncertainty is that in other locations on Mars any biomass in the soil will vary owing to nutrient availability, local temperatures and availability of water vapour in the atmosphere.

Adding water to the postulated H<sub>2</sub>O<sub>2</sub>–H<sub>2</sub>O-based life as done in the Viking life detection experiments would have ambiguous results. At the test cell temperatures of about 10 °C, organisms might survive 50% humidity for some time, whereas 100% humidity at that temperature seems to be fatal within a few days at most. The hygroscopicity of the H<sub>2</sub>O<sub>2</sub>–H<sub>2</sub>O mixture and the probable lack of a mechanism to exclude too much water are the likely causes of the sensitivity for water, even as vapour. The argument that Viking detected reactive chemistry rather than biology based on the fact that there are no known Earth organisms that can be shown to reproduce all Viking results is wanting, as is the argument that Viking discovered biology because there is no known mineral or reactive Earth-analogue chemistry that produces all Viking results. Any explanation of the Viking results must be intrinsically linked to the Martian environment with its differing geochemistry and organisms, if they exist. Recent Mars missions including the Rovers Spirit and Opportunity were not capable of detecting oxidants in the Martian soil.

### Terrestrial analogues

H<sub>2</sub>O<sub>2</sub>–H<sub>2</sub>O solutions are mostly known as disinfectants and sterilizing agents on Earth. Thus, the compatibility of H<sub>2</sub>O<sub>2</sub> with biological processes might seem questionable. However,

some microbial organisms produce hydrogen peroxide (e.g. certain *Streptococcus* and *Lactobacillus sp.* (Eschenbach *et al.* 1989; Ryan & Kleinberg 1995)), while other microbes utilize H<sub>2</sub>O<sub>2</sub> (e.g. *Neisseria sicca*, *Haemophilus segnis*, *H. parainfluenzae*, *Actinomyces viscosus* and *Staphylococcus epidermidis* (Ryan & Kleinberg 1995)). The microbe *Acetobacter peroxidans* even uses H<sub>2</sub>O<sub>2</sub> in its metabolism (overall reaction H<sub>2</sub>O<sub>2</sub>(aq) + H<sub>2</sub>(aq) ↔ 2H<sub>2</sub>O (Tanenbaum 1956)). However, the high reactivity of H<sub>2</sub>O<sub>2</sub> poses a problem to most microorganisms, which control it by the use of stabilizing compounds. Colloidal silicate and pyrophosphate are often used in commercial products, compounds such as phenacetin, an aromatic amine (N-ethoxy-acetanilide), may be more applicable to organisms. Most microbes that come into contact with H<sub>2</sub>O<sub>2</sub> protect themselves with scavenging enzymes such as catalase, glutathione peroxidase and peroxiredoxin. H<sub>2</sub>O<sub>2</sub> is commonly used as a defence mechanism by microbes, antibodies, immune cells and even certain insects. The Bombardier beetle, *Brachinus crepitans*, for example, has in its posterior a chitinous chamber in which a mix of fluids can be injected, one of which is a 25% solution of H<sub>2</sub>O<sub>2</sub> (Eisner 2003). This is combined with hydroquinone and a catalyst to produce a steam explosion in the chamber, which can be directed at a pursuing predator. The uses of H<sub>2</sub>O<sub>2</sub> in biology are surprisingly diverse. Mammalian cells are known to produce H<sub>2</sub>O<sub>2</sub> to mediate diverse physiological responses such as cell proliferation, differentiation and migration (Sundaresan *et al.* 1995; Rhee *et al.* 2000), and biological redox reactions catalysed by H<sub>2</sub>O<sub>2</sub> typically involve the oxidation of cysteine residues on proteins (Rhee 2006). Thus, high concentrations of H<sub>2</sub>O<sub>2</sub> can be produced and utilized biochemically even in terrestrial organisms. There does not appear to exist a basic reason why H<sub>2</sub>O<sub>2</sub> could not be used by living systems. On Earth, utilizing H<sub>2</sub>O<sub>2</sub> in the intracellular fluid has little advantage with regard to temperature and availability of oxygen and water, thus the majority of Earth organisms never developed extensive adaptation mechanisms. On Mars, on the other hand, directional selection may have favoured organisms that developed on an early warm and wet Mars to adapt to the progressive cooling and desiccation. Minor sources of inorganic H<sub>2</sub>O<sub>2</sub> produced in the Martian environment (e.g. Atreya *et al.* 2006) would increase the likelihood of an evolutionary trajectory using hydrogen peroxide for biochemical purposes. While for organisms on Earth it was advantageous to include large amount of salts into their intracellular fluids (e.g. Schulze-Makuch & Irwin 2004), H<sub>2</sub>O<sub>2</sub> may have been more suitable for organisms to optimally adapt to the very dry and cold environmental conditions on Mars.

The utilization of H<sub>2</sub>O<sub>2</sub> is not without some drawbacks. H<sub>2</sub>O<sub>2</sub> decomposes spontaneously, thus an organism needs some stabilization mechanism. The situation is even more demanding for photoautotrophic organisms exposed to sunlight, which on Mars includes a considerable flux of ultraviolet with wavelengths down to about 200 nm. H<sub>2</sub>O<sub>2</sub> will decompose under ultraviolet radiation and has to be protected by pigments in the cellular membrane or by an active stabilization mechanism. This does not necessarily require

chlorophyll, but could involve bacteriorhodopsin embedded in the cell membrane such as in halophilic organisms, or involve some inorganic compound such as cycloocta sulphur for efficient ultraviolet protection (Schulze-Makuch *et al.* 2004). However, to date, no suitable ultraviolet protection compound has been identified to exist on Mars, perhaps indicating that any such organisms, if they exist, would have to pursue an endolithic lifestyle comparable to the microbes in the Antarctic Dry Valleys (Friedmann 1982). Notably, these problems are of lesser magnitude at lower temperatures requiring fewer resources.

### Consequences for future Mars missions

In contrast to water-based organisms, the putative Martian autotrophs would need to avoid liquid water. However, in the generally arid environment, it is beneficial if the water vapour partial pressure is above about 50% for a significant fraction of time. Also, a generally low ambient temperature is beneficial in view of the stabilization of the cellular contents. If organisms on Mars exist that use the proposed biochemistry, they would likely be active in colder areas on Mars with high water vapour concentrations as would be expected along the polar ice fringes. As the Martian tropical areas are warmer and drier than optimum, they were not the best location for the detection of life based on a H<sub>2</sub>O<sub>2</sub>–H<sub>2</sub>O intracellular solvent.

Our hypothesis of Martian organisms that would utilize a H<sub>2</sub>O<sub>2</sub>–H<sub>2</sub>O mixture as an intracellular liquid is of great consequence for future missions searching for extant life on Mars. Rather than exploring in the equatorial belt, where temperatures might allow liquid water to exist for only brief periods of time, life may well exist in temperate or sub-arctic regions, where temperatures are colder and the atmosphere contains more water vapour. These concerns would also have to be addressed in future sample return missions to Mars and are relevant to the Mars Phoenix, ExoMars and Mars Science Laboratory missions.

### References

- Atreya, S.K. & Gu, Z. (1994). *J. Geophys. Res.* **99**, 13 133–13 145.
- Atreya, S.K., Wong, A.-S., Renno, N.O., Farrell, W.M., Delory, G.T., Sentman, D.D., Cummer, S.A., Marshall, J.R., Rafkin, S.C.R. & Catling, D.C. (2006). *Astrobiology* **6**, 429–450.
- Benner, S.A., Devine, K.G., Matveeva, L.N. & Powell, D.H. (2000). *Proc. Natl Acad. Sci. USA* **97**, 2425–2430.
- Biemann, K. *et al.* (1977). *J. Geophys. Res.* **82**, 4641–4658.
- Cockell, C.S., Schuerger, A.C., Billi, D., Friedmann, E.I. & Panitz, C. (2005). *Astrobiology* **5**, 127–140.
- Diaz, B. & Schulze-Makuch, D. (2006). *Astrobiology* **6**, 332–347.
- De Duve, C. (1969). *Ann. N. Y. Acad. Sci.* **168**, 369–381.
- Eisner, T. (2003). *For Love of Insects*. Harvard University Press, Cambridge, MA.
- Encrenaz, Th., Bezaud, B., Greathouse, T.K., Richter, M.J., Lacy, J.H., Atreya, S.K., Wong, A.S., Lebonnois, S., Lefevre, F. & Forget, F. (2004). *Icarus* **170**, 424–429.
- Eschenbach, D.A., Davick, P.R., Williams, B.L., Klebanoff, S.J., Young-Smith, K., Critchlow, C.M. & Holmes, K.K. (1989). *J. Clin. Microbiol.* **27**, 251–256.
- Flynn, G.J. (1996). *Earth Moon Planets* **72**, 469–474.
- Foley, W.T. & Giguère, P.A. (1951). *Can. J. Chem.* **29**, 123–132.
- Friedmann, E.I. (1982). *Science* **215**, 1045–1053.
- Giguère, P.A. & Secco, E.A. (1954). *Can. J. Chem.* **32**, 550–556.
- Horowitz, N.H. (1986). *To Utopia and Back: The Search for Life in the Solar System*. W.H. Freeman & Co., New York.
- Horowitz, N.H., Hobby, G.L. & Hubbard, J.S. (1976). *Science* **194**, 1321–1322.
- Horowitz, N.H., Hobby, G.L. & Hubbard, J.S. (1977). *J. Geophys. Res.* **82**, 4659–4662.
- Kashefi, K. & Lovley, D.R. (2003). *Science* **301**, 934.
- Klein, H.P. (1978). *Icarus* **34**, 666–674.
- Klein, H.P. (1999). *Origins Life Evol. Biosphere* **29**, 625–631.
- Klein, H.P. *et al.* (1976). *Science* **194**, 99–105.
- Levin, G.V. & Straat, P.A. (1976). *Science* **194**, 1322–1329.
- Levin, G.V. & Straat, P.A. (1977). *J. Geophys. Res.* **82**, 4663–4667.
- Levin, G.V. & Straat, P.A. (1981). *Icarus* **45**, 494–516.
- Mancinelli, R.L. (1989). *Adv. Space Res.* **9**, 191–195.
- Mattimore, V. & Battista, J.R. (1996). *J. Bacteriol.* **178**, 633–637.
- McDonald, G.D., de Vanssay, E. & Buckley, J.R. (1998). *Icarus* **132**, 170–175.
- Navarro-González, R. *et al.* (2006). *Proc. Natl Acad. Sci. USA* **103**, 16 089–16 094.
- Oyama, V.I. & Berdahl, B.J. (1977). *J. Geophys. Res.* **82**, 4669–4676.
- Oyama, V.I., Berdahl, B.J. & Carle, G.C. (1977). *Nature* **265**, 110–114.
- Quinn, R.C. & Zent, A.P. (1999). *Origins Life Evol. Biosphere* **29**, 59–72.
- Rhee, S.G., Bae, Y.S., Lee, S.R. & Kwon, J. (2000). *Signal Transduction Knowledge Environ.* **pe1**, 5353.
- Rhee, S.G. (2006). *Science* **312**, 1882–1883.
- Rothschild, L.J. (2007). *Planetary Systems and the Origins of Life*, eds Pudritz, R.E., Higgs, P. & Stone, J. Cambridge University Press, Cambridge.
- Ryan, C.S. & Kleinberg, I. (1995). *Arch. Oral. Biol.* **40**, 753–763.
- Schulze-Makuch, D., Grinspoon, D.H., Abbas, O., Irwin, L.N. & Bullock, M. (2004). *Astrobiology* **4**, 11–18.
- Schulze-Makuch, D. & Irwin, L.N. (2004). *Life in the Universe: Expectations and Constraints*. Springer, Berlin.
- Sundaresan, M., Yu, Z.X., Ferrans, Y.J., Irani, K. & Finkel, T. (1995). *Science* **270**, 296–299.
- Tanenbaum, S.W. (1956). *Biochim. Biophys. Acta* **21**, 335–342.
- Yen, A.S., Kim, S.S., Hecht, M.H., Frant, M.S. & Murray, B. (2000). *Science* **289**, 1909–1912.
- Zent, A.P. & McKay, C.P. (1994). *Icarus* **108**, 146–157.