Differential effects of reduced water potential on the germination of floodplain grassland species indicative of wet and dry habitats

Kristin Ludewig[1]*, Bianka Zelle[1], R. Lutz Eckstein[1], Eva Mosner[2], Annette Otte[1] and Tobias W. Donath[1]

[1]Institute of Landscape Ecology and Resource Management, Research Centre for Biosystems, Land Use and Nutrition (IFZ), Justus Liebig University Giessen, 35392 Giessen, Germany; [2]Bundesanstalt für Gewässerkunde (BfG), 56068 Koblenz, Germany

Abstract

Floodplain meadow ecosystems are characterized by high water level fluctuations and highly variable soil water potentials. Additionally, climate change scenarios indicate an increasing risk for summer drought along the northern Upper Rhine and the Middle Elbe River, Germany. While adult plants often persist even after strong changes in water availability, early life phases, such as seed germination and seedling establishment, might be more vulnerable. Therefore we tested whether reduced soil water potentials will affect the germination of meadow species and whether the response varies between (1) forbs indicative of wet and dry habitats and (2) seeds originating from sites along the rivers Elbe and Rhine. We exposed seeds of 20 floodplain meadow species with different moisture requirements from five plant families to a water potential gradient ranging from 0 to $-1.5\,\mathrm{MPa}$. While across species germination percentage and synchrony decreased, germination time increased at reduced water potentials. Germination of the species indicative of dry habitats decreased more strongly, was slower and less synchronous at reduced water potentials than that of species indicative of wet habitats. Seeds from sites along the rivers Elbe and Rhine did not differ in their germination characteristics. We propose that species of wet sites follow an all-or-nothing-strategy with fast and synchronous germination to maximize competitive advantages, betting on a high probability of moist conditions for establishment (optimists). In contrast, species from dry sites appear to follow a bet-hedging strategy with a moisture-sensing mechanism for unsuitable conditions (pessimists), resulting in a slower and less synchronous germination.

Introduction

Survival and persistence of plant populations strongly depend on reproduction, thus seed germination and seedling establishment are the most critical stages in the life of plant individuals (Grubb, 1977; Kitajima and Fenner, 2000). Without successful germination and establishment, populations are threatened by extinction. Germination processes are affected by the biotic environment, e.g. by competition from the established vegetation (Bakker and de Vries, 1992; Kitajima and Fenner, 2000) or by a litter layer (e.g. Jensen and Gutekunst, 2003; Loydi *et al.*, 2013), and by abiotic environmental conditions – the main abiotic drivers of germination processes being temperature and water availability (Baskin and Baskin, 2001; Fenner and Thompson, 2005). Most species require specific environmental conditions for the germination process and these factors are decisive for subsequent seedling establishment (Baskin and Baskin, 2001).

Floodplain grasslands are hydrologically highly dynamic ecosystems, characterized by the interplay of wet conditions during regularly or irregularly occurring floods and dry conditions over the summer (Hölzel and Otte, 2001). In directly inundated (functional) floodplains this gives rise to highly variable soil water potentials in time (i.e. within years) and space (i.e. along flooding gradients), resulting in a distinct zonation of plant communities (Leyer, 2005) which is also determined by seed and germination traits (Leyer and Pross, 2009). Due to the tightly intermingled vegetation zones, species density is high and floodplains harbour many rare and endangered species (Donath *et al.*, 2003;

*Correspondence
Email: kristin.ludewig@umwelt.uni-giessen.de

Toogood *et al.*, 2008). However, species-rich floodplain meadows have declined strongly over the past centuries due to habitat losses, mainly caused by land-use changes (e.g. Wesche *et al.*, 2012), river regulations (e.g. Tockner and Stanford, 2002) and river training (Brunotte *et al.*, 2009). Consequently, these meadows are of high conservation value and protected by the Habitats Directive (92/43/EEC, habitat type 6440: Alluvial meadows of river valleys of the *Cnidion dubii*) and subject to various restoration measures (Donath *et al.*, 2007; Schmiede *et al.*, 2012).

Climate change might pose an additional threat to the persistence of species-rich floodplain meadows. Regional climate change projections indicate higher temperatures and an increasing risk for summer droughts for the late 21st century due to less summer precipitation in western and northern Germany in relation to the reference period 1961–1990 (Jacob *et al.*, 2008; Görgen *et al.*, 2010). Accordingly, reduced river discharges during summers are projected for the large rivers Rhine (Görgen *et al.*, 2010) and Elbe (Conradt *et al.*, 2012). This, in turn, would lower the water table in the adjacent floodplain with negative effects on the soil water potential. In combination with increased transpiration at higher temperatures, these changes could induce severe drought-stress in plants of floodplain meadows (Jensen *et al.*, 2011). Additionally, increased temperatures could affect seed longevity, a prelude for building up viable seed banks (Ooi, 2012). As temperature and water availability are important drivers for the germination process, their changes will very likely affect this early stage in plant life (Walck *et al.*, 2011). Consequently, species abundance and population dynamics might be altered, leading to distribution shifts or extinctions (Thuiller *et al.*, 2005). To estimate future distribution and abundance of plant species, it is essential to gain knowledge on their specific requirements for seedling recruitment (e.g. Adler and HilleRisLambers, 2008; Walck *et al.*, 2011).

Recently, the impact of climate change on vegetation has received increasing attention. Different experimental approaches have already been conducted, focusing on CO_2 (e.g. Edwards *et al.*, 2001; Rasse *et al.*, 2005), temperature (e.g. Klanderud and Totland, 2005; Hudson *et al.*, 2011) or precipitation (e.g. Yahdjian and Sala, 2002; Beier *et al.*, 2012) and their effects on single species or plant communities. Some greenhouse and common garden experiments simulated drought (e.g. Jentsch *et al.*, 2011; Weisshuhn *et al.*, 2011) or flooding scenarios (e.g. van Eck *et al.*, 2006). While all of these approaches focused on the mature plant, little work has been done considering the early life stages. Here, one possible approach is to investigate germination at different water availabilities with seeds exposed to different water potentials (e.g. Fyfield and Gregory, 1989; Swagel *et al.*, 1997; Akhalkatsi and Lösch, 2001). To date, most studies in which water

potentials were manipulated focused on germination traits of single plant species, whereas comparative studies on a larger number of species are scarce (but see Evans and Etherington, 1990). Moreover, rare plant species and the influence of hydrological factors on their germination have only rarely been investigated (but see Geissler and Gzik, 2008) and, to our knowledge, only one study investigated whether germination differs between species indicative of different habitats (Evans and Etherington, 1990). However, analysing the responses of plant functional groups based on key life-history traits to climatic changes appears to be a promising approach (Ooi, 2012).

Therefore, the objective of our study was to test how floodplain meadow species, preferring contrasting habitats with respect to soil moisture, respond (in terms of germination) to different water potentials. We exposed seeds of 20 floodplain meadow species to a water potential gradient ranging from no water limitation to the permanent wilting point. The overall aim was to understand possible impacts of climate change on the early life stage of floodplain meadow plants. We also included seeds of five of these species from populations at the River Elbe to test the effect of seed origin on germination characteristics. To include different aspects of the germination of species, we analysed: (1) germination percentage; (2) mean germination time; and (3) synchrony of germination. Germination percentage simply measures the recruitment success of a batch of seeds, and population fitness increases with increasing germination. In contrast, the success of recruitment does not necessarily monotonically increase with mean germination time or synchrony. Rather, the benefit of early or late germination (i.e. low or high germination time) and germination in one batch or scattered germination over time (i.e. high and low synchrony) may vary with environmental context.

Our research questions and hypotheses were: (1) Do reduced water potentials affect the germination patterns of all plant species? We hypothesize that all study species show reduced germination percentages at reduced water potentials. Further, we expect that germination will take longer and will be less synchronous at reduced water potentials. (2) Does the temperature regime affect germination? We expect germination time to decrease with increasing temperature. (3) Do seeds originating from populations along the Elbe differ in their response to reduced water potentials compared to those from populations along the Rhine? We expect germination of seeds from the River Elbe, where annual precipitation is lower, to be more successful at reduced water potentials than germination of seeds from the Rhine. (4) Does seed germination of floodplain plant species indicative of wet habitats differ to those floodplain plant species indicative of dry habitats?

We hypothesize that seed germination of plants indicative of wet habitats will decrease more strongly, to be slower and less synchronous at reduced water potentials than that of plants indicative of dry habitats.

Methods

Study species

We selected 20 species (four from each of five plant families) occurring in floodplain grasslands along the River Rhine (see Table 1). These species consist of typical and rare floodplain meadow plants, such as *Galium boreale*, *Pseudolysimachion longifolium* or *Peucedanum officinale* (according to Burkart, 2001) and more common and widespread grassland species, such as *Plantago media*, *Linaria vulgaris* or *Galium verum*. Moisture requirements of the study species were classified according to the Ellenberg indicator value for moisture (Ellenberg *et al.*, 1992). Moisture indicator values of the selected species varied from 3 (indicative of dry habitats) to 9 (indicative of wet habitats). We

sought to include pairs of genetically related species with different preferences for soil moisture to attain a phylogenetically balanced design. Plant nomenclature follows Wisskirchen and Haeupler (1998).

Seed collection

Seeds from 13 of the 20 species were hand-collected from autochthonous populations in floodplain meadows along the northern Upper Rhine, Germany (49°50′N 8°25′E–49°51′N 8°23′E). The seeds of six species (*Filipendula ulmaria*, *F. vulgaris*, *Linaria vulgaris*, *Plantago media*, *Sanguisorba minor*, *Veronica teucrium*) were obtained from a commercial supplier (Rieger & Hoffmann GmbH, Blaufelden-Raboldshausen, Germany) due to insufficient amounts of seeds from natural populations along the River Rhine. Seeds of *Galium palustre* originated from floodplain meadows at the Middle Elbe, Germany, (52°32′N 11°59′E–52°49′N 12°03′E) as seeds of this species could not be found along the River Rhine nor ordered from a commercial supplier. Hand-collected

Table 1. Study species with information on plant family, Ellenberg indicator value for moisture (EIV moist, ind = species with 'indifferent behaviour'), indicated habitat, seed viability (%) and germination capacity under outdoor conditions (%)

	EIV moist	Indicated habitat	Viability (%)[a]	Germination capacity (%)[b]
Apiaceae				
Pimpinella saxifraga	3	Dry	98.0 ± 1.2	78.0 ± 2.3
Peucedanum officinale[#]	4	Dry	96.7 ± 1.4	20.0 ± 5.3
Selinum carvifolia	7	Wet	99.0 ± 1.0	99.3 ± 0.7
Silaum silaus[#]	ind	–	100.0 ± 0.0	91.3 ± 4.4
Asteraceae				
Inula salicina	6	–	91.5 ± 1.4	84.0 ± 4.0
Inula britannica[#]	7	Wet	94.0 ± 1.2	76.0 ± 5.0
Centaurea jacea	ind	–	90.0 ± 1.2	77.3 ± 3.7
Serratula tinctoria	ind	–	89.5 ± 2.5	84.0 ± 1.2
Plantaginaceae				
Veronica teucrium	3	Dry	98.0 ± 2.0	72.7 ± 0.7
Linaria vulgaris	4	Dry	60.0 ± 12.0	60.7 ± 10.7
Plantago media	4	Dry	86.0 ± 2.0	79.3 ± 5.8
Pseudolysimachion longifolium[#]	8	Wet	88.0 ± 1.6	85.3 ± 2.7
Rosaceae				
Filipendula vulgaris	3	Dry	94.0 ± 2.0	72.7 ± 8.5
Sanguisorba minor	3	Dry	70.0 ± 2.0	51.3 ± 8.2
Sanguisorba officinalis[#]	7	Wet	92.6 ± 1.9	94.7 ± 2.4
Filipendula ulmaria	8	Wet	50.0 ± 2.0	61.3 ± 5.2
Rubiaceae				
Galium verum	4	Dry	89.2 ± 2.3	49.3 ± 1.8
Galium album	5	–	96.0 ± 1.0	78.7 ± 2.9
Galium boreale[#]	6	–	89.0 ± 2.5	24.7 ± 8.5
Galium palustre	9	Wet	56.0 ± 3.4	18.7 ± 0.7

[#] According to Burkart (2001) typical flood meadow species (river corridor plants).
[a] Tetrazolium chloride test with 25 seeds per replicate ($n = 2$).
[b] Tested in common garden, 50 seeds per replicate ($n = 3$).

seeds were sampled between August and October 2010 depending on species-specific seed maturation. Freshly matured seeds were collected from at least two populations of a minimum of 20 individuals. For comparing the germination characteristics of seeds from the Rhine with seeds from the Elbe, seeds of five species (*Centaurea jacea, Galium verum, Inula britannica, Pseudolysimachion longifolium, Silaum silaus*) were additionally collected in autumn 2010 at the Middle Elbe (52°32′N 11°59′E–52°49′N 12°03′E). These seeds were also collected from at least two populations of at least 20 individuals (with the exception of *S. silaus* from which only one population was available).

The area along the northern Upper Rhine where the seeds were sampled has a mean annual precipitation of 643 mm and a mean annual temperature of 10.6°C (1981–2010 Gernsheim, DWD). The area at the middle Elbe has a lower mean annual precipitation of 555 mm and a mean annual temperature of 9.0°C (1981–2010 Boizenburg, DWD).

Seed handling and germination tests

After collection, seeds were manually cleaned, air-dried and stored in darkness at room temperature (approximately 20°C) until the start of the experiment in December 2010. Viability of seeds was tested for each population (25 per replicate, $n = 2$) with 1% tetrazolium chloride solution.

To test the germination capacity under outdoor conditions, seeds were sown into trays (50 per replicate, $n = 3$) with sterile standard garden soil and placed in a common garden (50°32′12″N 8°41′35″E, 172 m above sea level) at the same time as the climate chamber experiment started. The sowing date in January 2011 ensured cold wet stratification. Seeds were watered regularly and seedlings were counted and removed at least every other week for 2 years.

Experimental design

We used a factorial experimental design to study the effects of *species* (20 species), *water potential* (0, −0.25, −0.5, −1.0, −1.5 MPa) and *temperature* (day/night: 15/5°C and 20/10°C) on seedling emergence. Each treatment combination (species × temperature × water potentials) was replicated five times, resulting in 1000 experimental units. Additionally, for the comparison of the germination patterns of seeds from along the River Rhine with those of seeds from along the River Elbe, the seeds of the above-mentioned five species from the Elbe were also treated in five replicates with the five water potentials (0, −0.25, −0.5, −1.0, −1.5 MPa) at both temperatures (15/5°C and 20/10°C), resulting in another 250 Petri dishes.

We used the osmotic agent mannitol (Euro OTC Pharma GmbH, Bönen, Germany) to establish defined water potentials. Mannitol concentrations of 0.1, 0.2, 0.4 and 0.6 mol l⁻¹ were prepared to generate water potentials of approximately −0.25, −0.5, −1.0 and −1.5 MPa, respectively (according to Swagel *et al.*, 1997). As a control (full water availability = water potential of 0 MPa) we used distilled water.

Fifty seeds of each species (25 seeds of *Peucedanum officinale,* due to its large seed size) were placed in sterile Petri dishes with one piece of filter paper moistened with 3 ml of D-mannitol solution or distilled water. In order to reduce evaporation, five Petri dishes were sealed together in a plastic bag.

As a stratification measure, the seeds were exposed to cold wet conditions for 5 weeks in climate chambers (Rumed type 3401, Rubarth Apparate GmbH, Laatzen, Germany) at 5°C to ensure breaking of potential seed dormancy. For incubation, we exposed the seeds to 12 h light and 12 h darkness and two diurnally fluctuating temperatures (15/5°C and 20/10°C) to simulate different spring temperature conditions.

Germination was defined as protrusion of the radicle. Germination was checked twice a week and seedlings were removed. After 4 weeks of incubation, germination decreased and Petri dishes were checked only once a week. While checking the Petri dishes for seedlings, the amount of mannitol solution in the Petri dishes was controlled. In order to keep osmotic potentials constant during the experiment, Petri dishes were carefully washed with 7.5 ml of mannitol solution or distilled water (control), emptied and re-filled with 3 ml of new mannitol solution or distilled water after 4 weeks of incubation. After 8 weeks of incubation the experiment ended, since almost no further germination was observed.

Germination parameters

As response variables, we calculated the germination percentage (%), mean germination time (days) and synchrony of germination (an index ranging from 0–1, without units) per replicate (according to Ranal and Santana, 2006; Ranal *et al.*, 2009). The germination percentage is the percentage of all germinated seeds from the initial number of seeds. Mean germination time and synchrony of germination were calculated based on seedling counts over time (Ranal *et al.*, 2009). Mean germination time is a measurement of the weighted average length of time required for germination (Ranal and Santana, 2006). The unit depends on the counting frequency and is days (d) in this study. The mean germination time $\bar{t}$ is:

$$\bar{t} = \frac{\sum_{i=1}^{k} n_i t_i}{\sum_{i=1}^{k} n_i}$$

where t_i is the time from the start of the experiment to the ith observation day and n_i is number of seeds germinated in the ith time, and k is the last time of germination (for details, see Ranal *et al.*, 2009).

The synchrony of germination indicates the germination variability over time, ranging from 0 to 1 (high values indicate highly synchronous germination). The synchrony of germination, Z, is:

$$Z = \frac{\sum Cn_{i,2}}{N} \qquad Cn_{i,2} = \frac{n_i(n_i - 1)}{2}$$

$$N = \frac{\sum n_i(n_i - 1)}{2}$$

where n_i is number of seeds germinated in the ith time.

Data handling and statistical analyses

For data analysis, plants with an Ellenberg moisture value of 3 or 4 were classified as indicators of dry habitats, and species with a moisture value of 7, 8 or 9 as indicators of wet habitats. Species with an intermediate Ellenberg moisture value of 5 or 6, or a so-called indifferent behaviour towards moisture (see Ellenberg *et al.*, 1992), were not included in the comparison between these two groups of species in our study.

For most species the osmotic threshold for germination was −1.0 or at least −1.5 MPa (see Fig. 1). To avoid zero inflation, the osmotic potentials of −1.0 and −1.5 MPa had to be omitted from the analyses. Moreover, we had to exclude the results of *Galium palustre*, due to the extremely low germination percentage of the seeds of this species in the climate chamber experiment (1.0 ± 0.5%).

The effects of the experimental predictor variables *species*, *water potential* and *temperature* on germination percentage, germination time and synchrony of germination were analysed by three-way analyses of variance (ANOVAs). To account for inherent effects of family identity, we first calculated one-way ANOVAs with

the factor *plant family* for every transformed (see below) response variable [germination percentage ($F_{4,565} = 39.6$, $P < 0.001$), germination time ($F_{4,565} = 80.3$, $P < 0.001$) and synchrony of germination ($F_{4,565} = 51.4$, $P < 0.001$)] and used the residuals for the three-way ANOVAs. As a measure of the relative contribution of each factor and interaction to the total variability in germination percentage, germination time and synchrony of germination, we used the ratio of the sum of squares of the factor or interaction of interest to the total sum of squares [i.e. variance contribution (vc) for all factors, their interactions and the error]. Following the three-way ANOVA, we conducted: (1) Tukey HSD-tests for the interaction of *species × water potential*, to analyse whether reducing the water potential to −0.25 and −0.5 MPa affected the seed germination percentages on the species level; and (2) contrast analyses to analyse if the species indicative of wet and dry habitats differ in their response to reduced water potential conditions. For the contrast analyses, the germination responses (germination percentage, germination time and synchrony of germination) of the seeds from species indicative of wet habitats were tested against the germination responses of the seeds from species indicative of dry habitats separately for the water potentials −0.25 and −0.5 MPa (but not for the non-stress control conditions).

For the analysis of the effects of seed origin (Rhine versus Elbe) on the germination parameters, we conducted ANOVAs with the factors *species*, *water potential*, *temperature* and *origin*. Data were transformed to approximate normal distribution and variance homogeneity [germination percentage: arcsin (square root/100); mean germination time and synchrony: log + 1]. All statistical tests were conducted using STATISTICA 10 (StatSoft Inc., Tulsa, Oklahoma, USA).

Results

General characteristics of seed material

Viability (%) of the seeds was generally high (Table 1). Most species had > 85% viable seeds, exceptions were *Filipendula ulmaria* (50 ± 2%), *Galium palustre* (56 ± 3%), *Linaria vulgaris* (60 ± 12%) and *Sanguisorba minor* (70 ± 2%). Under outdoor conditions 13 species had germination percentages of > 70%, whereas low germination percentages were exhibited by *Galium boreale* (24 ± 9%), *G. palustre* (19 ± 1%) and *Peucedanum officinale* (20 ± 5%) (Table 1).

Germination in response to water potential, temperature and origin

Generally, germination percentage was significantly reduced at lower water potentials and totally ceased at

Figure 1. Germination percentage at the tested water potentials (mean + SE); including all treatments and species.

a water potential of $-1.5\,$MPa (Fig. 1). The three-way ANOVA indicated that germination percentage varied significantly among *species*, this factor explained 41% of the total variation (vc; Table 2). Furthermore, germination percentage was significantly influenced by the *water potential* (vc = 30%; Table 2). At a water potential of $-0.5\,$MPa, the germination of most study species was significantly reduced. Only the germination of four species was not susceptible to a water potential of $-0.5\,$MPa: *Centaurea jacea, Inula britannica, Sanguisorba minor* and *S. officinalis* (P values > 0.05, Fig. 2). Mean germination time was likewise mostly influenced by the factors *species* (vc = 43%) and *water potential* (vc = 27%; Table 2). Additionally, germination time was significantly affected by the temperature regime at which the seeds germinated (vc = 12%; Table 2). On average, seeds needed 13 d for germination at the temperature regime 15/5°C and 17 d at the higher temperature regime 20/10°C (including all species except *Galium palustre*). Synchrony of germination mainly depended on the factor *species* (vc = 35%) whereas *water potential* explained only 11% of the total variation (Table 2).

The comparison of seed germination of five species originating from the Rhine and the Elbe indicated species-specific responses. *Species* identity explained the largest part of the total variation in germination percentages ($F_{4,240} = 299.8$, $P < 0.001$, vc = 48%). The main factor of interest, i.e. *origin*, had no effect on the germination percentages ($F_{1,240} = 0.04$, $P = 0.85$). Further, the response to the reduced water potential did not differ between origins as no interaction between *origin* and *water potential* was detected ($F_{2,240} = 0.71$, $P = 0.49$). *Species* also explained most of the variation in mean germination time ($F_{4,239} = 742.1$, $P < 0.001$, vc = 60%). Although *origin* had a significant effect on germination time, it only explained a small part of the variation ($F_{1,239} = 145.8$, $P < 0.001$, vc = 3%).

No *origin × water potential* interaction was found for germination time ($F_{2,239} = 1.1$, $P = 0.35$). Again, in the analysis of synchrony *species* explained most of the variation ($F_{4,239} = 198.4$, $P < 0.001$, vc = 51%), while *origin* had a significant, but small impact ($F_{1,239} = 31.9$, $P < 0.001$, vc = 2%) and no *origin × water potential* interaction was found ($F_{2,239} = 1.3$, $P = 0.27$).

Wet versus dry habitat species

Contrast analyses revealed that the seeds of species indicative of wet habitats had significantly higher germination percentages than species indicative of dry habitats at reduced water potentials (see Table 2 and Fig. 3a). On average, 77 ± 3% (mean ± SE, N = 50) of the seeds from species of wet habitats germinated at a water potential of $-0.25\,$MPa versus only 62 ± 3% (N = 80) of the seeds from species of dry habitats. At a water potential of $-0.5\,$MPa, 61 ± 4% (N = 50) of the seeds from species of wet habitats germinated versus only 44 ± 3% (N = 80) of the seeds from species of dry habitats. Besides, the seeds of the species from dry habitats needed approximately 2 d longer for germination at a water potential of $-0.5\,$MPa (see Table 2 and Fig. 3b) and germinated slightly less synchronously at a water potential of $-0.25\,$MPa (see Table 2 and Fig. 3c) than the seeds of the species from wet habitats.

Discussion

Germination responses

In accordance with our first hypothesis, the germination of all species was negatively affected by reduced water availability, and almost ceased at a water

Table 2. Results of the ANOVA for the climate chamber experiment. Effects of *species*, *water potential* and *temperature* on germination (%), mean germination time (d) and synchrony of germination; including the water potentials 0, -0.25 and $-0.5\,$MPa; df = degrees of freedom, F = variance ratio, P = error probability, vc (%) relative contribution of individual factors and their interactions to total variance

	df	Germination (%)			Mean germination time			Synchrony		
		F	P	vc (%)	F	P	vc (%)	F	P	vc (%)
Species (S)	18	76.9	<0.0001	40.9	106.0	<0.0001	42.9	38.7	<0.0001	35.0
Temperature (T)	1	<0.1	0.9527	0.0	532.6	<0.0001	12.0	34.4	<0.0001	1.7
Water potential (WP)	2	504.4	<0.0001	29.8	608.3	<0.0001	27.4	110.6	<0.0001	11.1
S × T	18	9.3	<0.0001	5.0	7.4	<0.0001	3.0	9.3	<0.0001	8.4
S × WP	36	8.8	<0.0001	9.4	3.8	<0.0001	3.1	6.6	<0.0001	12.0
T × WP	2	4.2	0.0153	0.3	0.6	0.5346	0.0	0.2	0.8226	0.0
S × T × WP	36	1.2	0.2517	1.2	1.8	0.0037	1.5	4.8	<0.0001	8.8
Error	456			13.5			10.3			22.9
Contrasts: wet versus dry species										
WP $-0.25\,$MPa		63.5	<0.0001		2.3	0.1316		9.4	0.0023	
WP $-0.5\,$MPa		71.6	<0.0001		5.1	0.0246		0.1	0.7199	

potential of −1.5 MPa. This emphasizes the importance of soil moisture as a key requisite for the germination process (Baskin and Baskin, 2001) and is in line with other studies in which germination was affected by low water potentials (Evans and Etherington, 1990; Swagel *et al.*, 1997; Akhalkatsi and Lösch, 2001; Springer, 2005; Daws *et al.*, 2008). Two of these studies included species that were able to germinate at reduced water potentials
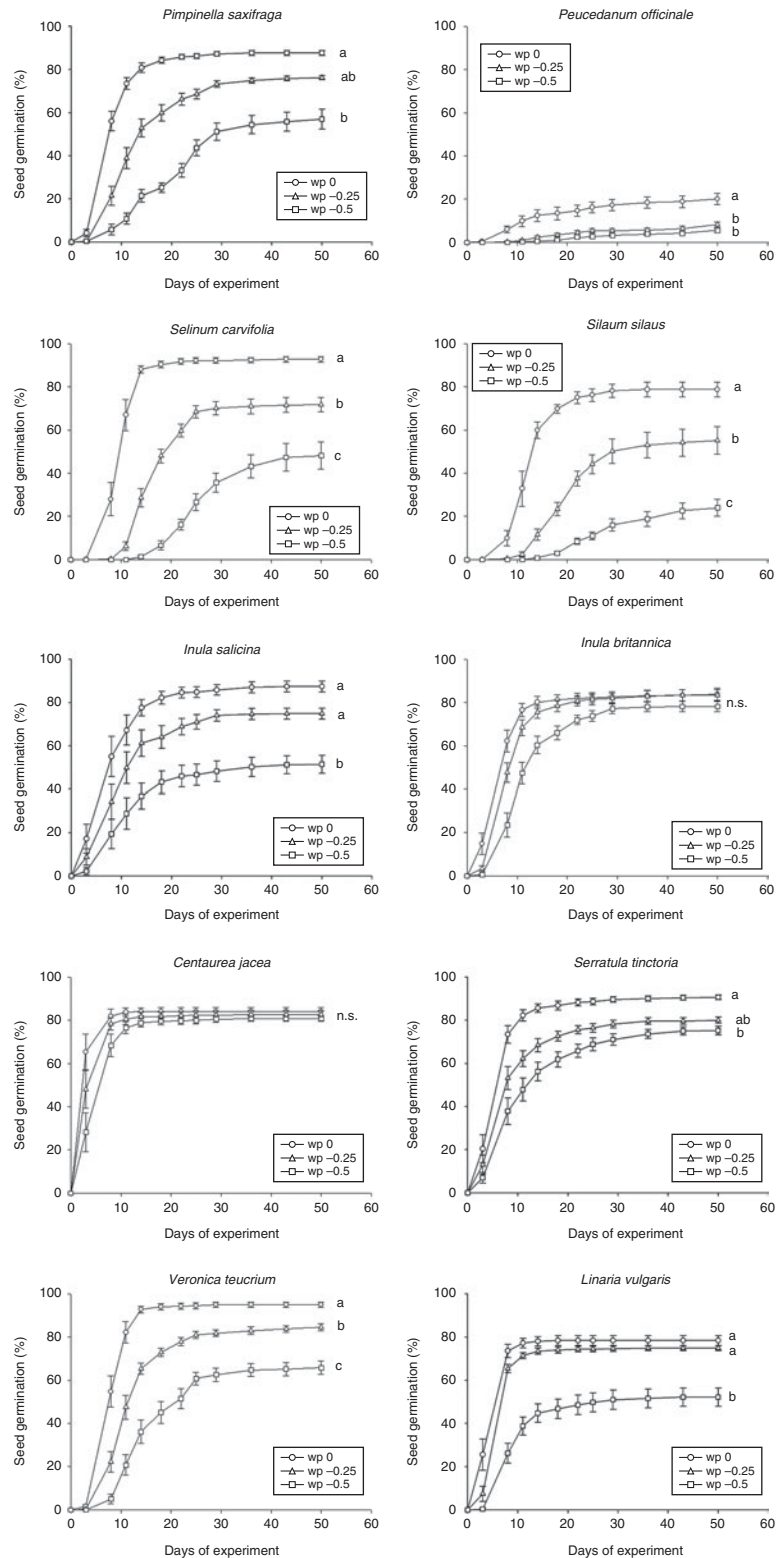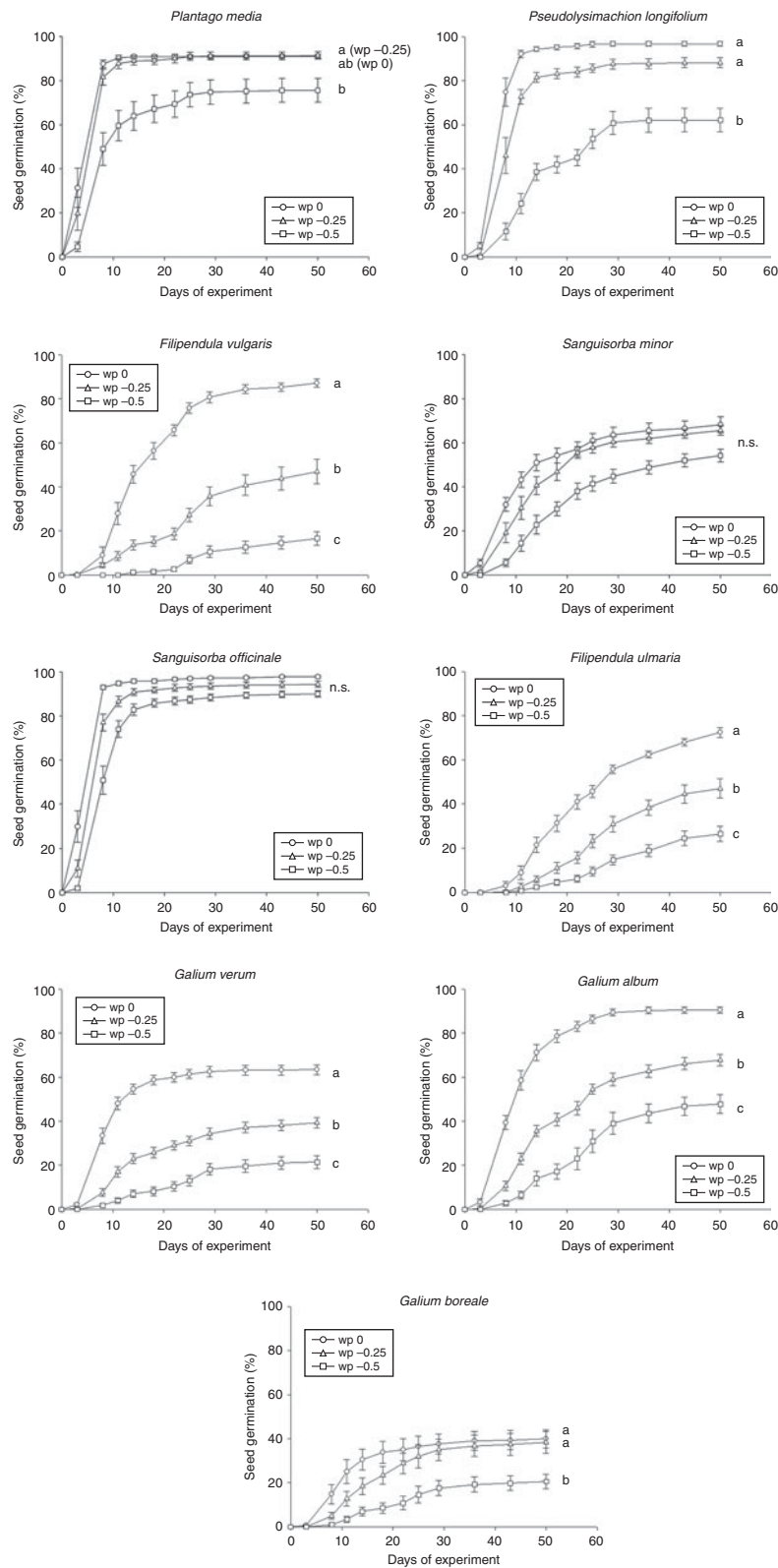

Figure 2. (*Continued*).

Figure 2. Cumulative germination percentage over time of the study species (except *Galium palustre*), averaging the data of the two temperature regimes (mean ± SE). Species order as in Table 1. Different letters indicate differences in the final germination percentage between water potential treatments (0, −0.25 and −0.5 MPa).

Figure 3. Germination percentage, germination time and synchrony, averaging the data of the two temperature regimes (mean + SE) of the species indicative of dry (*Pimpinella saxifraga, Peucedanum officinale, Veronica teucrium, Linaria vulgaris, Plantago media, Filipendula vulgaris, Sanguisorba minor, Galium verum*) and wet (*Selinum carvifolia, Inula britannica, Pseudolysimachion longifolium, Sanguisorba officinalis, Filipendula ulmaria*) habitats, including the water potentials -0.25 and -0.5 MPa (excluding *Galium palustre*). Significance levels: * = $P < 0.05$, *** = $P < 0.001$.

(for details see Evans and Etherington, 1990; Springer, 2005). Also in our study it is surprising that germination of four species (*Centaurea jaceae, Inula britannica, Sanguisorba minor* and *S. officinalis*) was unaffected by a water potential as low as -0.5 MPa. A further seven species germinated to an equal percentage at water potential -0.25 MPa compared to the control conditions (*Galium boreale, Inula salicina, Linaria vulgaris, Pimpinella saxifraga, Plantago media, Pseudolysimachion longifolium, Serratula tinctoria*). Germination time and synchrony of germination were also influenced by the water potential in our study. This finding corresponds to other studies in which a delayed onset of germination

was reported at reduced water potentials (Evans and Etherington, 1990; Daws *et al.*, 2008).

While the three response variables were mainly influenced by species identity (which fits with the results of Burmeier *et al.*, 2010) followed by water potential (see results of the three-way ANOVA, Table 2), it has to be acknowledged that all response variables were affected by plant family. Concerning germination time, an accumulation of species with the ability of very fast germination (<24 h) within certain plant families was recently reported in a review on this topic (Parsons, 2012). It seems that fast germination may be especially abundant in high-stress

germination time between the two groups of species is rather small (the seeds of species from wet habitats germinated approximately 2 d earlier than the seeds from species indicative of dry habitats) at the water potential -0.5 MPa. It remains unclear if this finding is ecologically relevant under field conditions. Nevertheless, especially in the productive wet meadows, early germination could be advantageous for establishment. This corresponds to a study on the germination of 91 species in response to a temperature gradient, where species of productive grassland germinated rapidly (Olff *et al.*, 1994). Therefore, we assume that the species of wet sites follow an all-or-nothing-strategy, with fast and synchronous germination to maximize competitive advantages and betting on a high probability of moist conditions for establishment ('optimists'). In contrast, species of dry sites follow a bet-hedging strategy with a moisture-sensing mechanism for the right conditions, betting on a high probability for unsuitable conditions ('pessimists' *sensu* Jones, 1992, who coined this terminology for strategy types of photosynthetic reactions to drought stress), resulting in a slower and less synchronous germination.

Conclusions

The germination of almost all studied species was decreased by lower water potentials, which strengthens the results of former studies (Evans and Etherington, 1990; Swagel *et al.*, 1997; Akhalkatsi and Lösch, 2001; Springer, 2005; Daws *et al.*, 2008) and demonstrates that floodplain meadow species will be negatively influenced in their earliest life stage (i.e. seed germination) by decreasing water availabilities during future climate change. However, the species indicative of wet and of dry habitats of floodplain meadows might be affected differently. Our experimental data suggest that seeds of species indicative of dry habitats show sensitivity to the moisture status of their immediate environment. Their 'pessimistic' response (germinating only when they sense sufficient moist conditions) probably enables them to track the time windows with high probability for successful germination and establishment. Seeds of species indicative for wet habitats do not possess such a mechanism since the conditions in their typical habitat are usually sufficiently moist. Their 'optimistic' response to this environmental factor probably makes them comparably vulnerable to climate change. Shifts of these species further down the elevation gradient in floodplains (where conditions are still moist enough for successful establishment) might be the consequence. Further, faster seed germination under low temperatures could also lead to a shift to earlier germination, when the soil is still moist from precipitation during winter and spring. Then other

factors, such as the occurrence or timing of spring floods or frosts may gain importance. More research is needed concerning the timing of germination under changing climatic conditions.

Conflicts of interest

None.

References

Adler, P.B. and HilleRisLambers, J. (2008) The influence of climate and species composition on the population dynamics of ten prairie forbs. *Ecology* **89**, 3049–3060.

Akhalkatsi, M. and Lösch, R. (2001) Changes in water relations, solute leakage and growth characters during seed germination and seedling development in *Trigonella coerulea* (Fabaceae). *Journal of Applied Botany* **75**, 144–151.

Bakker, J.P. and de Vries, Y. (1992) Germination and early establishment of lower salt-marsh species in grazed and mown salt marsh. *Journal of Vegetation Science* **3**, 247–252.

Baskin, C.C. and Baskin, J.M. (2001) *Seeds – ecology, biogeography, and evolution of dormancy and germination.* San Diego, Academic Press.

Beier, C., Beierkuhnlein, C., Wohlgemuth, T., Penuelas, J., Emmett, B., Korner, C., de Boeck, H., Christensen, J.H., Leuzinger, S., Janssens, I.A. and Hansen, K. (2012) Precipitation manipulation experiments – challenges and recommendations for the future. *Ecology Letters* **15**, 899–911.

Brunotte, E., Dister, E., Günther-Diringer, D., Koenzen, U. and Mehl, D. (2009) *Flussauen in Deutschland – Erfassung und Bewertung des Auenzustandes.* Bundesamt für Naturschutz (BFN), Bonn – Bad Godesberg.

Burkart, M. (2001) River corridor plants (Stromtalpflanzen) in Central European lowland: a review of a poorly understood plant distribution pattern. *Global Ecology and Biogeography* **10**, 449–468.

Burmeier, S., Donath, T.W., Otte, A. and Eckstein, R.L. (2010) Rapid burial has differential effects on germination and emergence of small- and large-seeded herbaceous plant species. *Seed Science Research* **20**, 189–200.

Cochrane, A., **Daws, M.I. and Hay, F.R.** (2011) Seed-based approach for identifying flora at risk from climate warming. *Austral Ecology* **36**, 923–935.

Conradt, T., **Koch, H.**, **Hattermann, F.F. and Wechsung, F.** (2012) Spatially differentiated management-revised discharge scenarios for an integrated analysis of multi-realisation climate and land use scenarios for the Elbe River basin. *Regional Environmental Change* **12**, 633–648.

Daws, M.I., **Crabtree, L.M.**, **Dalling, J.W.**, **Mullins, C.E. and Burslem, D.F.R.P.** (2008) Germination responses to water potential in neotropical pioneers suggest large-seeded species take more risks. *Annals of Botany* **102**, 945–951.

Donath, T.W., **Hölzel, N. and Otte, A.** (2003) The impact of site conditions and seed dispersal on restoration success in alluvial meadows. *Applied Vegetation Science* **6**, 13–22.

Donath, T.W., **Bissels, S.**, **Hölzel, N. and Otte, A.** (2007) Large scale application of diaspore transfer with plant material in restoration practice – impact of seed and microsite limitation. *Biological Conservation* **138**, 224–234.

Edwards, G.R., **Clark, H. and Newton, P.C.D.** (2001) The effects of elevated CO_2 on seed production and seedling recruitment in a sheep-grazed pasture. *Oecologia* **127**, 383–394.

Ellenberg, H., **Weber, H.E.**, **Düll, R.**, **Wirth, V.**, **Werner, W. and Paulißen, D.** (1992) Zeigerwerte von Pflanzen in Mitteleuropa. *Scripta Geobotanica* **18**, 1–248.

Evans, C.E. and Etherington, J.R. (1990) The effect of soil water potential on seed germination of some British plants. *New Phytologist* **115**, 539–548.

Fenner, M. and Thompson, K. (2005) *The ecology of seeds*. Cambridge, Cambridge University Press.

Fyfield, T.P. and Gregory, P.J. (1989) Effects of temperature and water potential on germination, radicle elongation and emergence of mungbean. *Journal of Experimental Botany* **40**, 667–674.

Geissler, K. and Gzik, A. (2008) The impact of flooding and drought on seeds of *Cnidium dubium*, *Gratiola officinalis*, and *Juncus atratus*, three endangered perennial river corridor plants of Central European lowlands. *Aquatic Botany* **89**, 283–291.

Görgen, K., **Beersma, J.**, **Brahmer, G.**, **Buiteveld, H.**, **Carambia, M.**, **de Keizer, O.**, **Krahe, P.**, **Nilson, E.**, **Lammersen, R.**, **Perrin, C. and Volken, D.** (2010) *Assessment of climate change impacts on discharge in the Rhine Basin: Results of the RheinBlick2050 Project, Lelystad*. CHR-report, Commission for the Hydrology of the Rhine Basin (CHR).

Grubb, P.J. (1977) The maintenance of species-richness in plant communities: The importance of the regeneration niche. *Biological Reviews* **52**, 107–145.

Hölzel, N. and Otte, A. (2001) The impact of flooding regime on the soil seed bank of flood-meadows. *Journal of Vegetation Science* **12**, 209–218.

Hölzel, N. and Otte, A. (2004) Ecological significance of seed germination characteristics in flood-meadow species. *Flora* **199**, 12–24.

Hudson, J.M.G., **Henry, G.H.R. and Cornwell, W.K.** (2011) Taller and larger: shifts in Arctic tundra leaf traits after 16 years of experimental warming. *Global Change Biology* **17**, 1013–1021.

Jacob, D., **Göttel, H.**, **Kotlarski, S.**, **Lorenz, P. and Sieck, K.** (2008) *Klimaauswirkungen und Anpassung in Deutschland – Phase 1: Erstellung regionaler Klimaszenarien für Deutschland*. 08/11, UBA – Umwelt Bundesamt, Dessau-Roßlau.

Jensen, K. and Gutekunst, K. (2003) Effects of litter on establishment of grassland plant species: the role of seed size and successional status. *Basic and Applied Ecology* **4**, 579–587.

Jensen, K., **Reisdorff, C.**, **Pfeiffer, E.M.**, **v. Oheimb, G.**, **Schmidt, K.**, **Schmidt, S.**, **Schrautzer, J.**, **Meyer-Grünefeldt, M. and Härdtle, W.** (2011) Klimabedingte Änderungen in terrestrischen und semi-terrestrischen Ökosystemen. pp. 143–176 *in* Storch, H.; Claussen, M. (Eds) *Klimabericht für die Metropolregion*. Hamburg, Springer.

Jentsch, A., **Kreyling, J.**, **Elmer, M.**, **Gellesch, E.**, **Glaser, B.**, **Grant, K.**, **Hein, R.**, **Lara, M.**, **Mirzae, H.**, **Nadler, S.E.**, **Nagy, L.**, **Otieno, D.**, **Pritsch, K.**, **Rascher, U.**, **Schadler, M.**, **Schloter, M.**, **Singh, B.K.**, **Stadler, J.**, **Walter, J.**, **Wellstein, C.**, **Wollecke, J. and Beierkuhnlein, C.** (2011) Climate extremes initiate ecosystem-regulating functions while maintaining productivity. *Journal of Ecology* **99**, 689–702.

Jones, H.G. (1992) *Plants and microclimate: A quantitative approach to environmental plant physiology*. Cambridge, Cambridge University Press.

Jurado, E. and Westoby, M. (1992) Germination biology of selected central Australian plants. *Australian Journal of Ecology* **17**, 341–348.

Kitajima, K. and Fenner, M. (2000) Ecology of seedling regeneration. pp. 331–359 *in* Fenner, M. (Ed.) *Seeds – the ecology of regeneration in plant communities*. Wallingford, CABI Publishing.

Klanderud, K. and Totland, O. (2005) Simulated climate change altered dominance hierarchies and diversity of an alpine biodiversity hotspot. *Ecology* **86**, 2047–2054.

Leyer, I. (2005) Predicting plant species' responses to river regulation: the role of water level fluctuations. *Journal of Applied Ecology* **42**, 239–250.

Leyer, I. and Pross, S. (2009) Do seed and germination traits determine plant distribution patterns in riparian landscapes? *Basic and Applied Ecology* **10**, 113–121.

Loydi, A., **Eckstein, R.L.**, **Otte, A. and Donath, T.W.** (2013) Effects of litter on seedling establishment in natural and semi-natural grasslands: a meta-analysis. *Journal of Ecology* **101**, 454–464.

Olff, H., **Pegtel, D.**, **Vangroenendael, J. and Bakker, J.** (1994) Germination strategies during grassland succession. *Journal of Ecology* **82**, 69–77.

Ooi, M.K.J. (2012) Seed bank persistence and climate change. *Seed Science Research* **22** (suppl. S1), S53–S60.

Parsons, R.F. (2012) Incidence and ecology of very fast germination. *Seed Science Research* **22**, 161–167.

Qi, M.Q. and Redmann, R.E. (1993) Seed germination and seedling survival of C3 and C4 grasses under water stress. *Journal of Arid Environments* **24**, 277–285.

Ranal, M.A. and de Santana, D.G. (2006) How and why to measure the germination process? *Revista Brasileira de Botânica* **29**, 1–11.

Ranal, M.A., **de Santana, D.G.**, **Ferreira, W.R. and Mendes-Rodrigues, C.** (2009) Calculating germination measurements and organizing spreadsheets. *Revista Brasileira de Botânica* **32**, 849–855.

Rasse, D.P., **Peresta, G. and Drake, B.G.** (2005) Seventeen years of elevated CO_2 exposure in a Chesapeake Bay Wetland: sustained but contrasting responses of plant growth and CO_2 uptake. *Global Change Biology* **11**, 369–377.

Romo, J.T., **Grilz, P.L.**, **Bubar, C.J. and Young, J.A.** (1991) Influences of temperature and water stress on germination of plains rough fescue. *Journal of Range Management* **44**, 75–81.

Schmiede, R., **Otte, A. and Donath, T.W.** (2012) Enhancing plant biodiversity in species-poor grassland through plant material transfer – the impact of sward disturbance. *Applied Vegetation Science* **15**, 290–298.

Springer, T.L. (2005) Germination and early seedling growth of chaffy-seeded grasses at negative water potentials. *Crop Science* **45**, 2075–2080.

Swagel, E.N., **Bernhard, A.V.H. and Ellmore, G.S.** (1997) Substrate water potential constraints on germination of the strangler fig *Ficus aurea* (Moraceae). *American Journal of Botany* **84**, 716–722.

Thuiller, W., **Lavorel, S.**, **Araújo, M.B.**, **Sykes, M.T. and Prentice, I.C.** (2005) Climate change threats to plant diversity in Europe. *Proceedings of the National Academy of Sciences, USA* **102**, 8245–8250.

Tockner, K. and Stanford, J.A. (2002) Riverine flood plains: present state and future trends. *Environmental Conservation* **29**, 308–330.

Toogood, S., **Joyce, C. and Waite, S.** (2008) Response of floodplain grassland plant communities to altered water regimes. *Plant Ecology* **197**, 285–298.

van Eck, W., **Lenssen, J.**, **van de Steeg, H.**, **Blom, C. and de Kroon, H.** (2006) Seasonal dependent effects of flooding on plant species survival and zonation: a comparative study of 10 terrestrial grassland species. *Hydrobiologia* **565**, 59–69.

Walck, J.L., **Hidayati, S.N.**, **Dixon, K.W.**, **Thompson, K. and Poschlod, P.** (2011) Climate change and plant regeneration from seed. *Global Change Biology* **17**, 2145–2161.

Weisshuhn, K., **Auge, H. and Prati, D.** (2011) Geographic variation in the response to drought in nine grassland species. *Basic and Applied Ecology* **12**, 21–28.

Wesche, K., **Krause, B.**, **Culmsee, H. and Leuschner, C.** (2012) Fifty years of change in Central European grassland vegetation: Large losses in species richness and animal-pollinated plants. *Biological Conservation* **150**, 76–85.

Wisskirchen, R. and Haeupler, H. (1998) *Standardliste der Farn- und Blütenpflanzen Deutschlands*. Stuttgart, Ulmer.

Yahdjian, L. and Sala, O.E. (2002) A rainout shelter design for intercepting different amounts of rainfall. *Oecologia* **133**, 95–101.