# Repeated treatment with nitric oxide synthase inhibitor attenuates learned helplessness development in rats and increases hippocampal BDNF expression

Stanquini LA, Biojone C, Guimarães FS and Joca SR. Repeated treatment with nitric oxide synthase inhibitor attenuates learned helplessness development in rats and increases hippocampal BDNF expression.

**Background:** Nitric oxide synthase (NOS) inhibitors induce antidepressant-like effects in animal models sensitive to acute drug treatment such as the forced swimming test. However, it is not yet clear if repeated treatment with these drugs is required to induce antidepressant-like effects in preclinical models.

**Objective:** The aim of this study was to test the effect induced by acute or repeated (7 days) treatment with 7-nitroindazole (7-NI), a preferential inhibitor of neuronal NOS, in rats submitted to the learned helplessness (LH) model. In addition, we aimed at investigating if 7-NI treatment would increase brain-derived neurotrophic factor (BDNF) protein levels in the hippocampus, similarly to the effect of prototype antidepressants. **Methods:** Animals were submitted to a pre-test (PT) session with inescapable footshocks or habituation (no shocks) to the experimental shuttle box. Six days later they were exposed to a test with escapable footshocks. Independent groups received acute (a single injection after PT or before test) or repeated (once a day for 7 days) treatment with vehicle or 7-NI (30 mg/kg).

**Results:** Repeated, but not acute, treatment with 7-NI attenuated LH development. The effect was similar to repeated imipramine treatment. Moreover, in an independent experimental group, only repeated treatment with 7-NI and imipramine increased BDNF protein levels in the hippocampus.

**Conclusion:** The results suggest the nitrergic system could be a target for the treatment of depressive-like conditions. They also indicate that, similar to the positive control imipramine, the antidepressant-like effects of NOS inhibition could involve an increase in hippocampal BDNF levels.

# Laura Alves Stanquini<sup>1,2,\*</sup>, Caroline Biojone<sup>1,2,\*</sup>, Francisco Silveira Guimarães<sup>2,3</sup>, Sâmia Regiane Joca<sup>1,3</sup>

<sup>1</sup>Laboratory of Pharmacology, Department of Physics and Chemistry, School of Pharmaceutical Sciences of Ribeirão Preto (FCFRP), University of São Paulo, Ribeirão Preto, SP, Brazil; <sup>2</sup>Department of Pharmacology, School of Medicine of Ribeirão Preto (FMRP), University of São Paulo, Ribeirão Preto, SP, Brazil; and <sup>3</sup>Center for Interdisciplinary Research on Applied Neurosciences (NAPNA) from University of São Paulo, SP, Brazil

\*All authors contributed equally to this study.

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Dr. Sâmia R. L. Joca, School of Pharmaceutical Sciences of Ribeirão Preto (FCFRP), University of São Paulo, Avenida do café s/n, 14040-903, Ribeirão Preto, SP, Brazil. Tel: +55 16 36024705; Fax: +55 16 36024880; E-mail: samia@usp.br

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#### Significant outcomes

- In the present work the treatment with a preferential neuronal nitric oxide synthase (nNOS) inhibitor, 7-nitroindazole (7-NI), was tested in a behavioural model in which the effect of antidepressant drugs is usually observed after prolonged treatment.
- Similar to clinical antidepressants, only repeated, but not acute, treatment with 7-NI produced an antidepressant-like effect in the learned helplessness (LH) model.
- The antidepressant-like effects of 7-NI may involve increased brain-derived neurotrophic factor (BDNF) signalling in the hippocampus.

## Limitations

- To help us to elucidate the latency period of 7-NI effects, a time-course curve of BNDF levels should have been performed.
- BDNF levels were measured only in the hippocampus of non-stressed animals. Analysis of BDNF content in other structures, as well as in stressed animals, could add significant information.
- Analysis of expression and/or activity of nNOS was not performed.

### Introduction

Since the discovery of the antidepressants and their mechanism of action, it has been postulated that a monoaminergic imbalance in the brain was responsible for the development of depression, which would be restored by long-term antidepressant treatment (1–3). This so-called 'Monoaminergic Theory of Depression', however, does not explain why the therapeutic effects appear only after chronic treatment despite their acute effects on monoamine levels (4,5).

Amongst the several neurotransmitters that have been studied in an attempt to better understand the neurobiology of depression, nitric oxide (NO) has received considerable attention in the last decades. NO is an intercellular messenger which plays a critical role in several neural physiological processes (6-10). It is synthesised during the conversion of L-arginine to citrulline by the enzyme NOS. Of the three isoforms of NOS that have been identified nNOS), endothelial NOS (eNOS), and inducible NOS (iNOS) (11) - nNOS is the most expressed in the brain (12). nNOS is usually activated in response to  $Ca^{+2}$  influx, after the opening of *N*-methyl-Daspartate (NMDA) receptors by glutamate binding (13,14). NO diffuses rapidly and influences NOresponsive target cells in a widely extended area. modulating neuronal function in several ways (14,15). Besides its involvement in physiological processes, NO can also be harmful due to its high reactivity, thus triggering pathophysiological responses (7).

The observation that treatment with NMDA antagonists induces antidepressant-like effects in experimental protocols (16,17) raised the possibility that NO could also be involved in the neurobiology of depression. Since the first report by Jefferys and Funder (18), showing that systemic administration of NOS inhibitor reduces immobility time in the rat forced swimming test, several additional studies have confirmed an antidepressant-like effect in response to treatment with different NOS inhibitors in animals submitted to different models predictive of antidepressant activity (19–24). Besides, antidepressants drugs are able to reduce NOS activity in hippocampus (25) while the discontinuation with imipramine treatment induces opposite effects in rats (26). There

is also clinical evidence supporting the involvement of NO in the neurobiology of depression (27–29).

Despite consistent evidence showing antidepressantlike effects of nNOS inhibitors in animals submitted to different preclinical models [for review (19) and (30)], it remains to be investigated if repeated treatment with NOS inhibitors is actually required to induce antidepressant-like effects. Given the fact that acute blockade of NMDA receptors, which are key modulators of NO synthesis in the CNS, is shown to promote rapid antidepressant-like effects (31-33) we investigated if acute treatment with nNOS inhibitor could induce similar effects. In addition, the effect of nNOS inhibition in the LH model has not been described before. In this context, the present work aimed at investigating the effects induced by acute and repeated treatment with a nNOS inhibitor in animals exposed to LH, a model that is irresponsive to acute (single injection) antidepressant treatment (34-37).

Repeated treatment with different antidepressants drugs increases the expression of BDNF, a neurotrophin critical for neuronal survival (38), in the hippocampus (39). Normalisation of BDNF levels in the hippocampus has been proposed as a crucial mechanism for antidepressant-induced behavioural effects (40,41). Evidence from *in vitro* and *in vivo* studies indicate BDNF levels can also be modulated by NO. For instance, administration of NO donors or NOS inhibitor decreases or increases, respectively, BDNF secretion in cultured cells (42). Besides, nNOS inhibition attenuated the decreased levels of BDNF in the hippocampus of animals exposed to chronic mild stress exposure (43).

Based on these pieces of evidence, the aim of present work was to investigate the effects induced by acute and repeated administration of 7-nitroindazole, a preferential nNOS inhibitor, in rats submitted to the LH model of depression. We also evaluated if the behavioural effect of this drug was temporarily associated with changes in hippocampal BDNF levels.

#### **Materials and methods**

#### Animals

Male Wistar rats weighing 200–220 g at the beginning of each experiment were individually housed in Plexiglas cages  $(30 \times 19 \times 13 \text{ cm})$  and kept in a temperature-controlled room  $(24 \pm 1^{\circ}\text{C})$ , under standard laboratory conditions with free access to food and water and a 12 h light/12 h dark cycle (lights on at 06:30 a.m.). The welfare of the animals was assessed daily. The cages and bedding were changed every 2 days as well as food and water replacement. All animal used came from a breeding facility from the University of São Paulo – Campus Ribeirão Preto, São Paulo, Brazil.

Animals were randomly assigned to the different experimental groups and procedures were performed in conformity with the Brazilian Council for the Control of Animals under Experiment (CONCEA), which are in compliance with international laws and politics. The local Ethical Committee approved the experimental protocol (number 08.1.233.53.5), and all efforts were made to minimise animal suffering.

## Drugs

Imipramine Hydrochloride (Sigma-Aldrich, Saint Louis, MO, USA) was administered i.p. at the dose of 15 mg/kg (according to (44)) and 7-nitroindazole (7-NI minimum 98%; Sigma-Aldrich, St Louis, MO, USA), a preferential nNOS inhibitor (45), was administered i.p. at the dose of 30 mg/kg (according to (21,46)). The drugs were dissolved in 40% dimethyl sulfoxide (DMSO; Gold Lab, Diadema, SP, Brazil), immediately before use, and administered at the volume of 2 ml/kg.

# Apparatus

The experiments were carried out in two automated shuttle boxes with two compartments of equal size  $(25 \times 22 \times 22 \text{ cm})$  separated by a wall with a central open door and equipped with a stainless-steel grid floor through which the scrambled shocks were delivered (Automatic Reflex Conditioner no. 7502 – UGO BASILE Biological Research Apparatus, Varese, Italy). One of the shuttle boxes was made of grey Plexiglas walls while the other was made of white Plexiglas walls and both had transparent covers. The behavioural tests took place in a sound attenuated, temperature-controlled  $(25 \pm 1^{\circ}C)$  room.

# **Behavioural procedure**

# LH

The procedures for the LH, a widely used behavioural test for the detection of antidepressant-like effects, were similar to those previously described by Joca et al. (44,47). The behavioural procedure consisted of two experimental sessions conducted in day 1 (pre-test or habituation) and day 7 (test) in different shuttle

boxes as described above. At the beginning of each experiment, naïve animals were randomly assigned to one of the following conditions: stressed group (pre-test with inescapable footshocks) or non-stressed group (habituation in the shuttle-box without footshocks). The animals in the pre-test group were placed in the shuttle box and submitted to 40 inescapable footshocks (1 mA, 10 s duration) given according to a variable time schedule with a mean interval of 60 s (range from 30 to  $90 \, \text{s}$ ). The rats in the habituation group were placed into the same apparatus for 30 min but no shock was given during this time. Six days later, animals of both groups were placed individually into the shuttle box and submitted to the test session (T). The test consisted of 30 escapable footshocks (0.8 mA, 10s duration, 30–90s interval) preceded by a tone (60 dB, 670 Hz) that started 5 s before each shock and lasted until the end of the footshock. Animals were free to cross to the opposite side of the chamber at any time during the test. They could avoid the shock (if they cross during sound presentation) or interrupt its presentation (if they cross during sound plus shock presentation). Absence of one of these behaviours was considered an escape failure, which was automatically registered during the test. The number of intertrial crossings were also automatically measured as an index of locomotion during the test (48).

# Hippocampal BDNF measurements

The animals were deeply anaesthetised (chloral hydrate 5%, 10 ml/kg) and sacrificed. Their hippocampi were dissected and homogenised in lysis buffer (NaCl 137 mM; Tris-HCl 20 mM pH 7.6; glycerol 10%) containing protease inhibitor cocktail (Sigma-Aldrich, Saint Louis, MO, USA, Cat# P2714). After centrifugation (20000 g, 15 min), the supernatant was collected and stored at -80°C until use. An aliquot of each sample was reserved and used to determine the total proteins levels using the Bradford method. Hippocampal BDNF was measured by ELISA (BDNF Emax® ImmunoAssay System kit; Promega, Madison, WI, USA, Cat# G7610) according to the manufacturer's instructions. Briefly, 96-well plate was pre-coated with a primary monoclonal antibody against BDNF overnight at 4°C. Following 1h blockade with BSA solution, supplied by the kit, we added the samples (2 h incubation at room temperature) and later the secondary polyclonal antibody (2h incubation at room temperature). After incubating for 2 h at room temperature with the tertiary Horseradish peroxidase (HRP)-conjugated, colorimetric detection of peroxidase activity was achieved by adding tetramethylbenzidine (TMB One) solution. The enzymatic reaction was stopped with hydrochloric acid (HCl)

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1 M and the colour intensity of each well was measured at 450 nm using a plate reader (VictorX3; Perkin Elmer, Boston, MA, USA). A standard curve was generated using values from the dilution series of a recombinant human BDNF standard (also supplied by kit) and was used to determine the BDNF concentration. Data was normalised by total protein levels in each sample.

## **Experimental design**

# LH

Independent groups of rats were submitted to pre-test or habituation (PT or H). In the repeated treatment group, animals received 7-NI (30 mg/kg), imipramine (15 mg/kg) or vehicle (1 ml/kg) for 7 days (the first injection was delivered immediately after PT or H, and the last one was delivered 1 h before test). In the acute treatment group, a single injection of 7-NI (30 mg/kg) or vehicle was administered either on the first day (immediately after pre-test or habituation) or on the seventh day (1 h before test) and the animals received vehicle injection on the other days.

## BDNF measurements

Independent groups of naïve animals received 7-NI (30 mg/kg), imipramine (15 mg/kg) or vehicle (1 ml/kg) once day for 7 days (repeated treatment). An additional experimental group received vehicle (1 ml/kg) injections for 6 days and a single injection with 7-NI (30 mg/kg), imipramine (15 mg/kg) or vehicle (1 ml/kg) on the last day (7th). One hour after the last injection, animals were killed and the hippocampus (both hemispheres) was collected.

# **Experimental conditions**

# Behavioural experiments

Animals were kept in groups of four animals/cage  $(41 \times 34 \times 16 \text{ cm})$  for at least 1 week before the beginning of the experiment. For behavioural experiments, animals were brought to the lab on day 1 and taken individually to experimental rooms, where they were exposed to PT or H. Immediately after completion of H or PT, animals received the administration of the drug or its vehicle. After that, they were housed individually in smaller cages  $(30 \times 19 \times 13 \text{ cm})$  and taken back to the animal house. Every day, at 07:00 am, the animals were brought to the lab experimental rooms, where they were weighted and injected with drug or vehicle, according to the groups they had been randomly assigned to and then taken back to the animal room.

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The animals assigned to BDNF analysis were maintained in the animal house for at least 1 week until the beginning of the treatment procedure, under the same conditions described above for animals submitted to behavioural experiments. On first day of treatment (day 1), animals were brought to the lab, where they were weighted and randomly assigned to the different treatment groups. After injection, they were housed individually in smaller cages  $(30 \times 19 \times 13 \text{ cm})$  and taken back to the animal house. Every day at 07:00 am, the animals were brought to the lab, where they were weighted and injected with drug or vehicle, and then taken back to the animal keeping room. On the treatment 7th day, the animals received the last injection and returned to their boxes where their remained undisturbed for 1 h. After that time, they were anaesthetised and sacrificed for hippocampus dissection. The last treatment injection and sacrifice were performed in random order to avoid circadian influences on BDNF analysis in individual treatment groups.

#### Sample size

The probable number of animals per group was calculated using the program G\*power 3.1.9.2. Based on that, the sample size required for LH experiments is about 13 animals per group. The total number of animals used was 175 male Wistar rats, and this number is close to the number predicted by G\*power program (204 animals). It is important to highlight that in some experimental groups the number of animals was smaller than expected due to the following exclusion criteria: animals that presented no crossing during test session, inflammation in the site of injection or other health issues.

# Statistical analysis

The total number of intertrial crossings and escape failures were calculated for each animal and analysed by Kruskal–Wallis test. BDNF levels were analysed by one-way analysis of variance (ANOVA) followed by Dunnett's Multiple Comparison test. Probability <0.05 was accepted as significant.

#### Results

Effect of imipramine or 7-NI repeated treatment on the number of escape failures in stressed and nonstressed animals (Fig. 1): (A) Kruskal–Wallis analysis indicated a significant effect of repeated pharmacological treatment with imipramine and 7-NI when compared with control (H=15, p=0.0006, n=12–15). The number of escape failures (mean ± SEM) for veh, imi, and 7-NI treatment, respectively,



*Fig. 1.* Repeated treatment with 7-NI counteracts the effect of stress in the learned helplessness model. Rats submitted to (A) pre-test (inescapable footshocks, stressed group) or (B) habituation (no shocks, non-stressed) were treated for 7 days with imipramine (imi), 7-NI or vehicle (vehic) and then submitted to learned helplessness test (escapable footshocks). Graphs represent the number of escape failures (median with interquartile ranges) measured during the test session. Kruskal-Wallis test indicated significant difference from corresponding vehicle-treated group (\*p < 0.05).



*Fig.* 2. Effect of imipramine or 7NI repeated treatment on the number of intertrial crossings during the test: Rats submitted to the pre-test (inescapable footshocks, stressed group) were treated for 7 days with imipramine (imi) or 7-NI and then submitted to learned helplessness test (escapable footshocks). Graphs represent the number of intertrial crossings (median with interquartile range) measured during the test session. Kruskal wallis test didn't indicate significant difference from vehicle-treated in the stressed group (p > 0.05).

were:  $18.67 \pm 2.35$ ,  $5.00 \pm 2.35$ , and  $5.25 \pm 1.89$ . Besides that, (B) in H group, Kruskal–Wallis analysis did not find any effect of repeated pharmacological treatment (H=1.101, p=0.5767, n=8-9). The number of escape failures (mean ± SEM) for veh, imi, and 7-NI treatment, respectively, were:  $0.375 \pm 0.1830$ ,  $0.2222 \pm 0.2222$ , and  $5.444 \pm 3.610$ .

Effect of imipramine or 7-NI repeated treatment on the number of intertrial crossings (Fig. 2): Kruskal– Wallis analysis did not find any effect on the number of intertrial crossings (H = 0.704, p = 0.7, n = 12-15). The number of intertrial crossing (mean ± SEM) for veh, imi, and 7-NI treatment, respectively, were:  $25.6 \pm 5.2$ ,  $32 \pm 7.9$ , and  $27.7 \pm 9.7$ .

Effect of 7-NI acute treatment on the number of escape failures in stressed and non-stressed animals (Fig. 3): Kruskal–Wallis analysis did not find any effect of acute pharmacological treatment, after PT and before T, with 7-NI in (A) stressed group when compared with control (H=0.1944, p=0.9074,

n=9–15). The number of escape failures (mean  $\pm$  SEM) for veh, 7-NI after PT, and 7-NI before T, respectively, were: 15.87  $\pm$  2.713, 15,88  $\pm$  4.311, and 18.22  $\pm$  2.100. The same occurred on (B) habituated group with Kruskal–Wallis analyses (H=0.1864, p=0.3937, n=8–20). In this case, the number of escape failures (mean  $\pm$  SEM) for veh, 7-NI after PT, and 7-NI before T, respectively, were: 10.45  $\pm$  2.155, 9.375  $\pm$  3.946, and 13.75  $\pm$  3.048.

Effect of imipramine or 7-NI repeated or acute treatment on BDNF content in hippocampus (Fig. 4): (A) Imipramine and 7-NI (in the same dose and drug regimen that induced antidepressant-like effect in LH) increased hippocampal BDNF, as indicated by one-way ANOVA (F(2,18) = 7.25), p < 0.05) followed by Dunnett's multiple comparison test (veh vs. imi: p < 0.05, veh vs. 7-NI: p < 0.05, n=8). (B) The single injection with Imipramine and 7-NI was ineffective in increasing hippocampal indicated by one-way BDNF, as ANOVA (F(2,20) = 0.7907, p > 0.05, n = 8) followed by Dunnett's multiple comparison test (veh vs. imi: p > 0.05, veh vs. 7-NI: p > 0.05).

#### Discussion

The present work demonstrated that: (1) repeated, but not acute, treatment with 7-NI induced an antidepressant-like effect in the LH paradigm; (2) BDNF levels were increased in the hippocampus of animals repeatedly treated with 7-NI. This data agrees with several studies that pointed to the involvement of NO in the development of stressrelated disorders (18,21,22,46,49–51). However, none of the previously published studies investigated if acute inhibition of nNOS either during the test assessment or immediately after stress presentation would be sufficient to induce antidepressant-like effect. Therefore, this is the first report that nNOS inhibition fails to induce rapid antidepressant-like



*Fig. 3.* Acute treatment with the preferential nNOS inhibitor 7-NI is not sufficient to counteract the effect of stress in the learned helplessness model. Rats submitted to (A) the pre-test (inescapable footshocks, stressed group) or (B) habituation (no shocks, non-stressed group) received a single injection of 7-NI either immediately after the pre-test or 1h before the test, and then were submitted to learned helplessness test (escapable footshocks). The graph represents the number of escape failures (median with interquartile range) measured during the test session. Kruskal wallis test didn't indicate significant difference from vehicle and the treatments in the stressed and habituated groups (p > 0.05).



*Fig. 4.* 7-NI and imipramine repeated treatment increases BDNF protein in hippocampus. BDNF from total hippocampus of naïve animals treated with 7-NI or imipramine (A) during 7 days or (B) with a single injection 1h before the sacrifice was quantified by ELISA method. Data represent mean $\pm$ sem normalized by control (vehicle group). \*p < 0.05 versus vehicle group.

effects in an animal model that is insensitive to acute effects of conventional antidepressant treatments (35).

The clinical effect of antidepressant drugs has been associated with repeated treatment, as the mood improvement in depressed patients can only be observed only after 4–5 weeks of continuous treatment (5). Based on this observation, animal models that are sensitive to repeated but not acute antidepressant treatment are believed to have a better predictive validity to the antidepressant effect (52). Thus, these models could provide additional and valuable information about the time course for the effects of new experimental drugs.

The LH does not respond to acute (single injection) treatment, with the antidepressant-like effect being observed after at least 3–4 days of continuous treatment (53–56). Moreover, LH is not sensitive either to anxiolytic drugs (diazepam, lorazepam) or to stimulants (amphetamine, caffeine), what confers a good predictive validity to the model (34,57). This model also presents good face validity,

as the behavioural deficit in controlling the presentation of aversive stimuli in animals previously exposed to uncontrollable stress (LH) is usually accompanied by endocrine, neurochemical and neuroanatomical changes that are also observed in depression (58,59). Helplessness has been found in depressed patients and, as a consequence, it has been the main focus of important preclinical and clinical depression research (58,59). Therefore, in the present study the LH model was used to investigate if helplessness behaviour induced by pre-exposure to stress could be counteracted by treatment with a NOS inhibitor. We also tried to identify the critical period when the NOS inhibition would be required.

Our findings demonstrated that repeated treatment (7 days) with a preferential nNOS inhibitor, 7-NI, counteracted the development of LH, thus inducing an effect similar to that induced by the prototype antidepressant imipramine. This effect does not seem to be associated with unspecific locomotor changes as no difference in the number of intertrial crossings was

found. Interestingly, it was observed that the acute 7-NI administration (either immediately after the pre-test stress or 1 h before the test) was not sufficient to induce antidepressant-like effect. These results suggest that a prolonged inhibition of NOS is necessary to achieve the antidepressant-like effect in the LH model. It is not clear why repeated treatment, rather than the acute, is needed. Some studies have suggested that stress could imbalance the normal nitrergic neurotransmission by increasing the nNOS expression (46,60). We speculate pre-test stress could induce a sustained increase in NO production that would be counteracted only by repeated administration of NOS inhibitors. Additional investigation is necessary to test this possibility and its triggering mechanisms, as well as to verify the time course of this process.

Learning and memory mechanisms should be considered when evaluating drug effects on LH paradigm. Escape failures in the test (the core representation of LH) have been interpreted as animal learning during PT that shock and its response are not contingent, thus leading to impairments in forming relevant contingencies in the future test session (61). Alternatively, it is seen as impaired associative learning during the test, due to increased fear (and freezing) and/or attentional deficits (62.63). Nevertheless, evidence from the literature has consistently shown that LH animals, either congenitally (64) or stress-induced (65,66), present attentional, learning and memory impairments, along with increased fearful associations and decreased fear extinction. These effects are similar to deficits of attention and working memory observed in patients with major depression (67–69), what gives further support to LH face validity as an animal model of depression. Accordingly, chronic antidepressant treatment attenuates cognitive impairments in LH animals (65,70) and in depressed humans (67-69).

Learning and memory mechanisms, particularly memory consolidation can be modulated by NO (71–73). In that scenario, it is important to consider that 7-NI effects could have resulted from disruption of consolidation of PT experience, impairments in retrieval of the aversive situation (PT) and/or impairments in the associative learning during test. Regarding aversive memories, evidence from contextual conditioning paradigm indicates that 7-NI administration, at the same dose used in the present study (30 mg/kg), immediately after the conditioning session with footshocks (consolidation period), did not interfere with the expression of learned fear later on (74). This is in agreement with our results observed with single injection of 7-NI after PT, which resulted in no effect in the test session. 7-NI treatment also did not impair the retrieval of already existing memory trace (74), again supporting our evidence that a single injection before the test was ineffective in the LH paradigm. In addition, in the same work (74), authors reported 7-NI disrupted context-shock associative learning, which could potentially decrease learning to escape in LH paradigm, thus leading to increased escape failures, which were not observed herein. Based on that, it is unlike that 7-NI effects observed in the present work resulted from impairments in learning and memory mechanisms. Alternatively, as the attenuation of fear and anxiety during the test facilitates associative learning and improve performance in LH (75), it is more likely that 7-NI effects in attenuating fear and anxiety (76,77) could have facilitated the attenuation of escape failures. Accordingly, L-NAME administration into the dorsal raphe nucleus reduced freezing induced by previous inescapable shock exposure and decreased escape failures during the test (78).

Emotional and cognitive effects of stress can result from impairments in cellular plasticity, including reduced neurogenesis, dendrite atrophy, loss of synaptic connection and neuronal cell death (79–82). This is supported by evidence that chronic, but not acute, treatment with monoaminergic antidepressants counteracts stress-induced effects in neural plasticity (38,83,84). In fact, prevention of stress-induced impairments in neural plasticity seems to be crucial to the antidepressant effect (85,86). Similar to conventional antidepressants, NOS inhibition increases cell proliferation (87,88) and prevents hippocampal neurogenesis impairment induced by chronic mild stress (46).

In line with that, previous evidence indicated NO/cGMP pathway modulates BDNF levels (89), a neurotrophin widely associated with cell proliferation, survival and differentiation (38). In hippocampal cell culture NO donors (SNP, NOR3) decreased BDNF release whereas the inhibition of NO production increased its levels (42). Also, rats systemically treated with L-NAME (a non-selective NOS inhibitor) for 7 days presented increased levels of BDNF mRNA in the dentate gyrus (90). However, in that study, behavioural effects of L-NAME were not investigated. Accordingly, in the present work, 7 days treatment with 7-NI increased hippocampal levels of BDNF, whereas acute injection was ineffective. Therefore, our data present evidence for a temporal association between the antidepressant-like effect of NOS inhibition and increased hippocampal BDNF levels. However, as BDNF levels were not investigated in stressed animals, it does not allow was to suggest a causal relationship between the increased levels of BDNF observed in non-stressed animals and the antidepressant-like effect induced by 7-NI.

Previous evidence suggests nNOS inhibitors increase serotonin availability in the hippocampus (91) and serotonin mediates their antidepressant-like

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effect in the Forced Swimming Test (FST) (92). On the other hand, different classes of antidepressants can decrease hippocampal NOS activity *in vivo* and *in vitro* (25), indicating there is a complex interaction between the nitrergic and serotonergic systems. Altogether, it is possible to suggest that the behavioural and molecular effects of 7-NI in the LH model could involve the modulation of serotonergic neurotransmission. However, this hypothesis warrants further investigation.

In conclusion, our findings, showing that inhibition of NO production induces antidepressant-like effects, corroborate the hypothesis that an increased production of NO might be related to the development of behavioural consequences of stress, such as LH. In addition, the antidepressant-like effect of nNOS inhibition could be mediated by increased BDNF signalling in the hippocampus.

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## **Conflicts of Interest**

The authors declare no conflict of interest. S.J. is an Associate Editor of Acta Neuropsychiatrica. However, S.J. did not handle the manuscript nor was involved in any decisions related to the present work.

## Supplementary material

To view supplementary material for this article, please visit https://doi.org/10.1017/neu.2017.28

#### References

- SCHILDKRAUT JJ, KETY SS. Biogenic amines and emotion. Science. 1967;156:21–37.
- HENINGER GR, DELGADO PL, CHARNEY DS. The revised monoamine theory of depression: a modulatory role for monoamines, based on new findings from monoamine depletion experiments in humans. Pharmacopsychiatry. 1996;29:2–11.
- 3. CASTREN E. Is mood chemistry? Nat Rev Neurosci 2005; 6:241–246.
- BLIER P. The pharmacology of putative early-onset antidepressant strategies. Eur Neuropsychopharmacol. 2003; 13:57–66.
- Kiss JP. Theory of active antidepressants: a nonsynaptic approach to the treatment of depression. Neurochem Int. 2008;52:34–39.

- Kiss JP. Role of nitric oxide in the regulation of monoaminergic neurotransmission. Brain Res Bull 2000;52:459–466.
- GUIX FX, URIBESALGO I, COMA M, MUNOZ FJ. The physiology and pathophysiology of nitric oxide in the brain. Prog Neurobiol. 2005;76:126–152.
- MONCADA S, BOLANOS JP. Nitric oxide, cell bioenergetics and neurodegeneration. J Neurochem. 2006;97:1676–1689.
- JOCA SR, GUIMARAES FS, DEL-BEL E. Inhibition of nitric oxide synthase increases synaptophysin mRNA expression in the hippocampal formation of rats. Neurosci Lett. 2007; 421:72–76.
- GARTHWAITE J. Concepts of neural nitric oxide-mediated transmission. Eur J Neurosci 2008;27:2783–2802.
- 11. PAAKKARI I, LINDSBERG P. Nitric oxide in the central nervous system. Ann Med. 1995;27:369–377.
- MORENO-LOPEZ B, NOVAL JA, GONZALEZ-BONET LG, ESTRADA C. Morphological bases for a role of nitric oxide in adult neurogenesis. Brain Res. 2000;869:244–250.
- GARTHWAITE J, CHARLES SL, CHESS-WILLIAMS R. Endothelium-derived relaxing factor release on activation of NMDA receptors suggests role as intercellular messenger in the brain. Nature. 1988;336:385–388.
- PRAST H, PHILIPPU A. Nitric oxide as modulator of neuronal function. Prog Neurobiol. 2001;64:51–68.
- 15. BRENMAN JE, BREDT DS. Synaptic signaling by nitric oxide. Curr Opin Neurobiol 1997;7:374–378.
- TRULLAS R, SKOLNICK P. Functional antagonists at the NMDA receptor complex exhibit antidepressant actions. Eur J Pharmacol 1990;185:1–10.
- PETRIE RX, REID IC, STEWART CA. The N-methyl-D-aspartate receptor, synaptic plasticity, and depressive disorder. A critical review. Pharmacol Ther. 2000;87:11–25.
- JEFFERYS D, FUNDER J. Nitric oxide modulates retention of immobility in the forced swimming test in rats. Eur J Pharmacol 1996;295:131–135.
- JOCA SR, MOREIRA FA, WEGENER G. Atypical Neurotransmitters and the neurobiology of depression. CNS Neurol Disord Drug Targets 2015;14:1001–1011.
- ULAK G, MUTLU O, TANYERI P, KOMSUOGLU FI, AKAR FY, ERDEN BF. Involvement of serotonin receptor subtypes in the antidepressant-like effect of TRIM in the rat forced swimming test. Pharmacol Biochem Behav 2010;95:308–314.
- YILDIZ F, ERDEN BF, ULAK G, UTKAN T, GACAR N. Antidepressant-like effect of 7-nitroindazole in the forced swimming test in rats. Psychopharmacology (Berl) 2000; 149:41–44.
- JOCA SR, GUIMARAES FS. Inhibition of neuronal nitric oxide synthase in the rat hippocampus induces antidepressant-like effects. Psychopharmacology (Berl) 2006;185:298–305.
- MONTEZUMA K, BIOJONE C, LISBOA SF, CUNHA FQ, GUIMARAES FS, JOCA SR. Inhibition of iNOS induces antidepressant-like effects in mice: pharmacological and genetic evidence. Neuropharmacology. 2012;62:485–491.
- WEGENER G, HARVEY BH, BONEFELD B et al. Increased stressevoked nitric oxide signalling in the Flinders sensitive line (FSL) rat: a genetic animal model of depression. Int J Neuropsychopharmacol 2010;13:461–473.
- 25. WEGENER G, VOLKE V, HARVEY BH, ROSENBERG R. Local, but not systemic, administration of serotonergic antidepressants decreases hippocampal nitric oxide synthase activity. Brain Res. 2003;959:128–134.

- HARVEY BH, RETIEF R, KORFF A, WEGENER G. Increased hippocampal nitric oxide synthase activity and stress responsiveness after imipramine discontinuation: role of 5HT 2A/C-receptors. Metab Brain Dis 2006;21:211–220.
- 27. SUZUKI E, YAGI G, NAKAKI T, KANBA S, ASAI M. Elevated plasma nitrate levels in depressive states. J Affect Disord 2001;63:221–224.
- OLIVEIRA RM, GUIMARAES FS, DEAKIN JF. Expression of neuronal nitric oxide synthase in the hippocampal formation in affective disorders. Braz J Med Biol Res 2008;41:333–341.
- NAYLOR GJ, SMITH AH, CONNELLY P. A controlled trial of methylene blue in severe depressive illness. Biol Psychiatry. 1987;22:657–659.
- WEGENER G, VOLKE V. Nitric oxide synthase inhibitors as antidepressants. Pharmaceuticals (Basel) 2010;3:273–299.
- BERMAN RM, CAPPIELLO A, ANAND A et al. Antidepressant effects of ketamine in depressed patients. Biol Psychiatry. 2000;47:351–354.
- DUMAN RS, LI N, LIU RJ, DURIC V, AGHAJANIAN G. Signaling pathways underlying the rapid antidepressant actions of ketamine. Neuropharmacology. 2012;62:35–41.
- ABDALLAH CG, SANACORA G, DUMAN RS, KRYSTAL JH. Ketamine and rapid-acting antidepressants: a window into a new neurobiology for mood disorder therapeutics. Annu Rev Med 2015;66:509–523.
- MAIER SF. Learned helplessness and animal models of depression. Prog Neuropsychopharmacol Biol Psychiatry 1984;8:435–446.
- SHERMAN AD, SACQUITNE JL, PETTY F. Specificity of the learned helplessness model of depression. Pharmacol Biochem Behav 1982;16:449–454.
- WILLNER P. Validation criteria for animal models of human mental disorders: learned helplessness as a paradigm case. Prog Neuropsychopharmacol Biol Psychiatry 1986;10: 677–690.
- 37 WILLNER P. Animal models of depression: an overview. Pharmacol Ther 1990;**45**:425–455.
- CASTREN E, RANTAMAKI T. The role of BDNF and its receptors in depression and antidepressant drug action: reactivation of developmental plasticity. Dev Neurobiol. 2010;70:289–297.
- BJORKHOLM C, MONTEGGIA LM. BDNF a key transducer of antidepressant effects. Neuropharmacology. 2016;102:72–79.
- DUMAN RS, HENINGER GR, NESTLER EJ. A molecular and cellular theory of depression. Arch Gen Psychiatry 1997;54: 597–606.
- DUMAN RS, MONTEGGIA LM. A neurotrophic model for stress-related mood disorders. Biol Psychiatry. 2006;59: 1116–1127.
- 42. CANOSSA M, GIORDANO E, CAPPELLO S, GUARNIERI C, FERRI S. Nitric oxide down-regulates brain-derived neurotrophic factor secretion in cultured hippocampal neurons. Proc Natl Acad Sci USA 2002;99:3282–3287.
- 43. YAZIR Y, UTKAN T, ARICIOGLU F. Inhibition of neuronal nitric oxide synthase and soluble guanylate cyclase prevents depression-like behaviour in rats exposed to chronic unpredictable mild stress. Basic Clin Pharmacol Toxicol 2012;111:154–160.
- JOCA SR, PADOVAN CM, GUIMARAES FS. Activation of postsynaptic 5-HT(1A) receptors in the dorsal hippocampus prevents learned helplessness development. Brain Res. 2003;978:177–184.

- 45. BUSH MA, POLLACK GM. Pharmacokinetics and pharmacodynamics of 7-nitroindazole, a selective nitric oxide synthase inhibitor, in the rat hippocampus. Pharm Res 2001;18:1607–1612.
- ZHOU QG, HU Y, HUA Y et al. Neuronal nitric oxide synthase contributes to chronic stress-induced depression by suppressing hippocampal neurogenesis. J Neurochem. 2007;103: 1843–1854.
- 47. JOCA SR, ZANELATI T, GUIMARAES FS. Post-stress facilitation of serotonergic, but not noradrenergic, neurotransmission in the dorsal hippocampus prevents learned helplessness development in rats. Brain Res. 2006;1087:67–74.
- GEOFFROY M. Psychomotor stimulants versus antidepressants in the learned helplessness model of depression. Drug Development Research 1993;29:48–55.
- GHASEMI M, SADEGHIPOUR H, MOSLEH A, SADEGHIPOUR HR, MANI AR, DEHPOUR AR. Nitric oxide involvement in the antidepressant-like effects of acute lithium administration in the mouse forced swimming test. Eur Neuropsychopharmacol. 2008;18:323–332.
- HARKIN AJ, BRUCE KH, CRAFT B, PAUL IA. Nitric oxide synthase inhibitors have antidepressant-like properties in mice. 1. Acute treatments are active in the forced swim test. Eur J Pharmacol 1999;**372**:207–213.
- MUTLU O, ULAK G, LAUGERAY A, BELZUNG C. Effects of neuronal and inducible NOS inhibitor 1-[2-(trifluoromethyl) phenyl] imidazole (TRIM) in unpredictable chronic mild stress procedure in mice. Pharmacol Biochem Behav 2009;92:82–87.
- WILLNER P, MITCHELL PJ. The validity of animal models of predisposition to depression. Behav Pharmacol. 2002; 13:169–188.
- 53. PORSOLT RD, LE PICHON M, JALFRE M. Depression: a new animal model sensitive to antidepressant treatments. Nature. 1977;**266**:730–732.
- CRYAN JF, MARKOU A, LUCKI I. Assessing antidepressant activity in rodents: recent developments and future needs. Trends Pharmacol Sci 2002;23:238–245.
- 55. SAARELAINEN T, HENDOLIN P, LUCAS G et al. Activation of the TrkB neurotrophin receptor is induced by antidepressant drugs and is required for antidepressant-induced behavioral effects. J Neurosci. 2003;23:349–357.
- 56. RANTAMAKI T, HENDOLIN P, KANKAANPAA A et al. Pharmacologically diverse antidepressants rapidly activate brain-derived neurotrophic factor receptor TrkB and induce phospholipase-C gamma signaling pathways in mouse brain. Neuropsychopharmacology. 2007;**32**:2152–2162.
- MARTIN P, PUECH AJ. Antagonism by benzodiazepines of the effects of serotonin-, but not norepinephrine-, uptake blockers in the learned helplessness paradigm in rats. Biol Psychiatry. 1996;39:882–890.
- PRYCE CR, AZZINNARI D, SPINELLI S, SEIFRITZ E, TEGETHOFF M, MEINLSCHMIDT G. Helplessness: a systematic translational review of theory and evidence for its relevance to understanding and treating depression. Pharmacol Ther. 2011; 132:242–267.
- 59 SELIGMAN MEP. Helplessness: on depression, development, and death. San Francisco: WH Freeman and Company, 1975.
- DE OLIVEIRA RM, APARECIDA DEL BEL E, MAMEDE-ROSA ML, PADOVAN CM, DEAKIN JF, GUIMARAES FS. Expression of neuronal nitric oxide synthase mRNA in stress-related brain areas after restraint in rats. Neurosci Lett. 2000; 289:123–126.

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- MAIER SF, JACKSON RL. Learned helplessness: all of us were right (and wrong): inescapable shock has multiple effects. In: Bower B, editor. Advances in Learning and Motivation. New York: Academic Press, 1979; p. 155–215.
- 62. JACKSON RL, ALEXANDER JH, MAIER SF. Learned helplessness, inactivity, and associative deficits: effects of inescapable shock on response choice escape learning. J Exp Psychol Anim Behav Process 1980;6:1–20.
- MINOR TR, JACKSON RL, MAIER SF. Effects of task-irrelevant cues and reinforcement delay on choice-escape learning following inescapable shock: evidence for a deficit in selective attention. J Exp Psychol Anim Behav Process 1984;10:543–556.
- 64. RICHTER SH, ZEUCH B, LANKISCH K, GASS P, DURSTEWITZ D, VOLLMAYR B. Where have I been? Where should I go? Spatial working memory on a radial arm maze in a rat model of depression. PLoS One. 2013;8:e62458.
- SONG L, CHE W, MIN-WEI W, MURAKAMI Y, MATSUMOTO K. Impairment of the spatial learning and memory induced by learned helplessness and chronic mild stress. Pharmacol Biochem Behav 2006;83:186–193.
- 66. YANG Y, WANG ZH, JIN S et al. Opposite monosynaptic scaling of BLP-vCA1 inputs governs hopefulness- and helplessness-modulated spatial learning and memory. Nat Commun. 2016;7:11935.
- CLARK L, CHAMBERLAIN SR, SAHAKIAN BJ. Neurocognitive mechanisms in depression: implications for treatment. Annu Rev Neurosci 2009;32:57–74.
- HAMMAR A, ARDAL G. Cognitive functioning in major depression-a summary. Front Hum Neurosci 2009;3:26.
- MARAZZITI D, CONSOLI G, PICCHETTI M, CARLINI M, FARAVELLI L. Cognitive impairment in major depression. Eur J Pharmacol 2010;626:83–86.
- SCHULZ D, MIRRIONE MM, HENN FA. Cognitive aspects of congenital learned helplessness and its reversal by the monoamine oxidase (MAO)-B inhibitor deprenyl. Neurobiol Learn Mem 2010;93:291–301.
- BOHME GA, BON C, STUTZMANN JM, DOBLE A, BLANCHARD JC. Possible involvement of nitric oxide in long-term potentiation. Eur J Pharmacol 1991;199:379–381.
- SCHUMAN EM, MADISON DV. A requirement for the intercellular messenger nitric oxide in long-term potentiation. Science. 1991;254:1503–1506.
- MIZUTANI A, SAITO H, ABE K. Involvement of nitric oxide in long-term potentiation in the dentate gyrus in vivo. Brain Res. 1993;605:309–311.
- 74. CHEN W, YAN M, WANG Y, WANG X, YUAN J, LI M. Effects of 7-nitroindazole, a selective neural nitric oxide synthase inhibitor, on context-shock associative learning in a twoprocess contextual fear conditioning paradigm. Neurobiol Learn Mem 2016;**134**(Pt B):287–293.
- MAIER SF, GRAHN RE, KALMAN BA, SUTTON LC, WIERTELAK EP, WATKINS LR. The role of the amygdala and dorsal raphe nucleus in mediating the behavioral consequences of inescapable shock. Behav Neurosci. 1993;107:377–388.
- GUIMARAES FS, DE AGUIAR JC, DEL BEL EA, BALLEJO G. Anxiolytic effect of nitric oxide synthase inhibitors microinjected into the dorsal central grey. Neuroreport. 1994;5:1929–1932.

- 77. SPOLIDORIO PC, ECHEVERRY MB, IYOMASA M, GUIMARAES FS, DEL BEL EA. Anxiolytic effects induced by inhibition of the nitric oxide-cGMP pathway in the rat dorsal hippocampus. Psychopharmacology (Berl) 2007;195:183–192.
- 78. GRAHN RE, WATKINS LR, MAIER SF. Impaired escape performance and enhanced conditioned fear in rats following exposure to an uncontrollable stressor are mediated by glutamate and nitric oxide in the dorsal raphe nucleus. Behav Brain Res 2000;**112**:33–41.
- LANFUMEY L, MONGEAU R, COHEN-SALMON C, HAMON M. Corticosteroid-serotonin interactions in the neurobiological mechanisms of stress-related disorders. Neurosci Biobehav Rev 2008;32:1174–1184.
- CASTREN E, VOIKAR V, RANTAMAKI T. Role of neurotrophic factors in depression. Curr Opin Pharmacol 2007;7:18–21.
- LUCASSEN PJ, MEERLO P, NAYLOR AS et al. Regulation of adult neurogenesis by stress, sleep disruption, exercise and inflammation: Implications for depression and antidepressant action. Eur Neuropsychopharmacol. 2010;20:1–17.
- McEwen BS. Glucocorticoids, depression, and mood disorders: structural remodeling in the brain. Metabolism. 2005;54(Suppl. 1):20–23.
- MALBERG JE, EISCH AJ, NESTLER EJ, DUMAN RS. Chronic antidepressant treatment increases neurogenesis in adult rat hippocampus. J Neurosci. 2000;20:9104–9110.
- NIBUYA M, MORINOBU S, DUMAN RS. Regulation of BDNF and trkB mRNA in rat brain by chronic electroconvulsive seizure and antidepressant drug treatments. J Neurosci. 1995;15:7539–7547.
- SANTARELLI L, SAXE M, GROSS C et al. Requirement of hippocampal neurogenesis for the behavioral effects of antidepressants. Science. 2003;301:805–809.
- MALBERG JE, DUMAN RS. Cell proliferation in adult hippocampus is decreased by inescapable stress: reversal by fluoxetine treatment. Neuropsychopharmacology. 2003;28:1562–1571.
- MORENO-LOPEZ B, ROMERO-GRIMALDI C, NOVAL JA, MURILLO-CARRETERO M, MATARREDONA ER, ESTRADA C. Nitric oxide is a physiological inhibitor of neurogenesis in the adult mouse subventricular zone and olfactory bulb. J Neurosci. 2004:24:85–95.
- PACKER MA, STASIV Y, BENRAISS A et al. Nitric oxide negatively regulates mammalian adult neurogenesis. Proc Natl Acad Sci USA 2003;100:9566–9571.
- BIOJONE C, CASAROTTO PC, JOCA SR, CASTREN E. Interplay between nitric oxide and brain-derived neurotrophic factor in neuronal plasticity. CNS Neurol Disord Drug Targets 2015;14:979–987.
- PINNOCK SB, HERBERT J. Brain-derived neurotropic factor and neurogenesis in the adult rat dentate gyrus: interactions with corticosterone. Eur J Neurosci 2008;27:2493–2500.
- 91. WEGENER G, VOLKE V, ROSENBERG R. Endogenous nitric oxide decreases hippocampal levels of serotonin and dopamine in vivo. Br J Pharmacol 2000;**130**:575–580.
- HARKIN A, CONNOR TJ, WALSH M, ST JOHN N, KELLY JP. Serotonergic mediation of the antidepressant-like effects of nitric oxide synthase inhibitors. Neuropharmacology. 2003;44:616–623.