A meta-analysis of host specificity in Neotropical hard ticks (Acari: Ixodidae)

S. Nava* and A.A. Guglielmone

Instituto Nacional de Tecnología Agropecuaria, Estación Experimental Agropecuaria Rafaela and Consejo Nacional de Investigaciones Científicas y Técnicas, CC 22, CP 2300 Rafaela, Santa Fe, Argentina

Abstract

Host specificity of Neotropical hard ticks (Acari: Ixodidae) was analyzed by using the number of hosts species for each tick species and the index of host specificity S_{TD}^* , which integrates phylogenetic and ecological information. The analyses were based on 4172 records of hard ticks collected from wild and domestic tetrapods. Most tick species included in this study were associated with three to 20 host species. No tick species has been associated either with a single species or with a single genus of host. It was found that the number of host species is sensitive to sampling effort, but not the S_{TD}^* . The most frequent values of S_{TD}^* were between 2.5 and 3.5, which shows that the host species more frequently used by Neotropical hard tick species belong to different families or different orders. Immature stages tend to use a broader taxonomic range of hosts than adults, and the interpretation of both measures of host specificity used in this study led to the conclusion that the impact of non-endemic hosts does not alter the patterns of host specificity in Neotropical hard ticks. The index S_{TD}* showed that a high proportion of tick species has phylogenetically unrelated species as principal hosts. The conclusion reached in this work indicates that strict host specificity is not common among Neotropical hard ticks and suggests that the influence of tick ecology and evolution of habitat specificity, tick generation time, phenology, time spent off the host and the type of life-cycle could be more important than hosts species.

Keywords: ticks, Ixodidae, host specificity, Neotropical region

(Accepted 3 July 2012; First published online 7 September 2012)

Introduction

One of the most important properties characterizing a parasite taxon is its host specificity. It is indicative of intrinsic biological characteristics of both host and parasite and an emergent property of their ecological and evolutionary relationship (Dick & Patterson, 2007). Host specificity can be defined as the extent to which a parasite taxon is restricted in the number of host species used at a given stage in the life cycle (Poulin, 2007). Thus, highly host-specific parasites have one host species, and specificity declines as the number of suitable

*Author for correspondence Fax: +54 03492440114 E-mail: snava@rafaela.inta.gov.ar host species increases (Poulin, 2007). The parasite-host association has been focused from two perspectives, the host centred view (the focus of coevolution is the host species) and the parasite-centred view (the resources for parasites are attributes of the host species, not the host species themselves) (Brooks & McLennan, 1993; Fenton & Pedersen, 2005; Brooks & Hoberg, 2007). The biological significance of these two visions lies in the frequency of host-switch events, which determines the host range and potential distribution of a parasite species. Host switching results in low host specificity because a parasite colonizes host lineages related or unrelated to its original host (Poulin, 2007). A parasite interacts with its host (physiology, immunity, ecology, phylogeny, size) but also with the environment. The external environment in which parasite and host interact can affect the strength and the evolution of the host-parasite interaction (Krasnov et al., 2008,

2010; Wolinska & King, 2009). Therefore, the degree of host specificity of a parasite and its variation in time and space not only are functions of intrinsic properties of host and parasite but also depends on external environmental conditions.

Hard ticks (Acari: Ixodidae) are haematophagous ectoparasites of amphibians, reptiles, birds and mammals, and they are divided into Prostriata (genus Ixodes) and Metastriata (genera Amblyomma, Anomalohimalaya, Bothriocroton, Compluriscutula⁺, Cornupalpatum⁺, Cosmiomma, Dermacentor, Haemaphysalis, Hyalomma, Margaropus, Nosomma, Rhipicentor, Rhipicephalus) (genera are listed according to Guglielmone et al., 2010). The life cycle of hard ticks encompasses parasitic and non-parasitic phases. In the former, larvae, nymphs and adults feed or mate on the host (members of Prostriata and a few species of Amblyomma and Bothriocroton have adults with ability to mate off host). However, hard ticks spend most of their life cycle off host, exposed to environmental conditions. In three-host cycle species, the non-parasitic phase includes important biological processes, such as moulting of engorged larvae and nymphs, oviposition of engorged females, incubation of eggs and host-seeking of all parasitic stages. Some ticks have a two-host or a one-host life cycle (i.e. species of the genera Rhipicephalus, Hyalomma, Dermacentor and Margaropus), where larvae or both nymphs and larvae moult on the host, respectively. Overall, it is evident that the life cycle of hard ticks is strongly influenced by both host and environmental factors.

Two contrasting hypotheses have been developed about host specificity in ticks. The first assumes that host specialisation was instrumental in the evolution of ticks and of their morphological characters (Hoogstraal, 1978; Hoogstraal & Aeschlimann, 1982; Hoogstraal & Kim, 1985) and is based on the idea of coevolution between ticks and terrestrial tetrapods. These authors state that phenotypic variations in mouthparts and coxae are the result of adaptation to a particular group of hosts, which lead to a high host specificity. Furthermore, Hoogstraal & Aeschlimann (1982) presented a classification system with six categories characterized by decreasing levels of host-specificity, namely: (i) strict-total, (ii) moderatetotal, (iii) strict-stage-stage, (iv) strict/moderate-stage-stage, (v) moderate-stage-stage and (vi) nonparticular. Klompen et al. (1996) raised an alternative hypothesis that stresses the importance of ecological specificity. According to this opinion, adaptation to a particular habitat is more relevant for tick evolution than adaptation to a particular host. Klompen et al. (1996) found a strong positive correlation between the total number of collections and the number of hosts, through an analysis using data from ixodid (Ixodes) and argasid (Carios) collections, and they concluded that "much of the distinction between strict-total and less-specific categories might be the difference between rarely and frequently collected species". Additionally, Klompen et al. (1996) criticized the lack of differentiation between the processes of adaptation and speciation in the Hoogstraal's studies. Klompen et al. (1996) stated that "although the presence of host adaptations may lead to host specificity, observed host specificity is not necessarily an indicator of host adaptation. Host specificity may also arise because of no opportunity to transfer to alternative hosts (which secondarily may result in cospeciation) or as a secondary effect of adaptation to off-host habitat". In the same way, Balashov (2004) rejected the idea of coevolution as a key factor in the evolution of ticks affirming that "phylogenetic parallelism between ticks and their hosts is absent or limited to short evolutionary periods". After the

analysis of a large data set of African ticks, Cumming (1998) reached a conclusion that can be interpreted as intermediate between the two aforementioned hypotheses. This author suggested that both host specificity and ecological specificity could be important and that it depends on each particular tick species. However, in a subsequent analysis on limiting factors for species ranges of African ticks, Cumming (2002) concluded that the distribution of ticks over a wide spatial scale is mainly determined by direct climatic effects, while hosts only generate heterogeneity in tick distribution at a smaller geographical scale.

In a broad sense, the host specificity of a parasite is quantified as the number of hosts species, or host range. The number of host species in itself is not informative because it implies that a parasite uses each host species equally. However, the utilization of a spectrum of host species by a given parasite is typically uneven. In fact, from an ecological perspective, some host species are used more intensively than others; and, from a phylogenetic perspective, some host species used by a parasite can be phylogenetically more closely related than others (Poulin et al., 2006; Poulin, 2007). Therefore, an attempt at evaluating the importance of host specificity must take into account all these aspects. In this work, a meta-analysis of host specificity in Neotropical hard ticks was performed. The number of hosts for each tick species and the index of host specificity (index S_{TD}*) proposed by Poulin & Mouillot (2005), which integrates phylogenetic and ecological information, were applied to the analysis. In addition, the two values were generated separately for adult and immature stages in order to investigate possible ontogenetic changes in host specificity, and the role of domestic animals in determining host specificity was also analyzed.

Materials and methods

The data set used in this study was obtained from a list of host records for all species of Neotropical ticks compiled from the literature (scientific papers, book sections and selected meeting proceedings) by one of us (AAG). The list is available under request. A record is defined as the finding of a tick species, on a determined host, and at a given locality, regardless of the number of host sampled and of ticks collected on a particular host. Findings on the same species of host and at the same locality, but on different dates, are each considered separate records. Only references that include tick stage, locality and scientific name of the host were considered, and tick species with less than 15 records were not included in the analysis. Records of immature stages made before the description of larva and nymph of the corresponding tick species became available were excluded. Exceptions were made for those references where the taxonomic determination method was explained (e.g. immature ticks identified to species after rearing the ticks to the adult stage in the laboratory or by using molecular tools). Records from humans, from hosts kept in captivity (laboratories, zoos, etc.) and from imported animals were not considered. When necessary, scientific names of mammal hosts have been updated following Wilson & Reeder (2005), Weksler et al. (2006) (for oryzomyne rodents), Voss & Jansa (2009) (for marsupials), Barquez et al. 2006 (for Felidae and Tayassuidae) and Francés & D'Elía (2006) and D'Elía et al. (2008) for some synonymized sigmodontine rodents. Scientific names of birds were taken from Clements (2007), and the nomenclature of amphibians and reptiles was the same chosen in Guglielmone & Nava

(2010). *Rhipicephalus (Boophilus) microplus* and the *Rhipicephalus sanguineus* species group were not considered in this work because they have a wide world distribution, and both taxa were recently introduced in the Neotropical region (approximately 400 years ago); therefore, an accurate analysis of their host-association should include records of other biogeographic regions. The list of host species recorded for each tick species is shown in the supplementary appendix 1.

The specificity index STD* proposed by Poulin & Mouillot (2005) was calculated for each hard tick species. This index measures the average taxonomic distinctness of all host species used by a parasite, weighted by the prevalence of the parasite in the different hosts (Poulin & Mouillot, 2005). Because of the lack of data on prevalence in most tick-host associations cited in the literature, we calculated instead, for each tick species, the proportion of records corresponding to each host species. It is obvious that the proportion of records is less informative than prevalence because it increases the risk of biases associated with differential sampling efforts. Another caveat to keep in mind is that presence of a representative of a given tick species on a given host does not mean this host is competent to sustain tick development and reproduction. The index STD* places hosts within the Linnean taxonomic hierarchy (phylum, class, order, family, genera, species). Taxonomic distinctness between two host species represents the mean number of steps up the taxonomic hierarchy that must be taken to reach a taxon common to both (for example: when two host species are congeners, one step species-togenus is necessary to reach a common node; if two host species belong to different genera but are included in the same family, two steps species-to-genus and genus-to-family are necessary) (Poulin & Mouillot, 2005). This value is weighted by the product of the parasite prevalence (in this work, proportion of records). The value of this index is inversely proportional to specificity. A high index value means that the host species more frequently used by a parasite are not closely related (Poulin & Mouillot, 2005). In absence of differences in the proportion values, the index STD* reaches its maximum value (5) when all host species belong to different classes, whereas it tends towards its minimum value (1) when all hosts belong to a same genus. Index STD* was calculated with a computer program developed by Poulin & Mouillot (2005) using Borland C++ Builder 6.0 (available online at http://www. otago.ac.nz/zoology/downloads/poulin/TaxoBiodiv2). The classifications used to place each host species within the correct classes, orders and families were Wilson & Reeder (2005) for class Mammalia, Clements (2007) for class Aves and Guglielmone & Nava (2010) for classes Reptilia and Amphibia.

A Pearson's correlation analysis was performed to assess the extent of co-variation between number of records of each tick species and number of host species and between number of records and S_{TD}^* values. The statistical significance of the differences in S_{TD}^* values between the samples which included domestic hosts and samples which excluded them was determined through a Student's *t*-test. The significance of the differences in number of host species between adult and immature stages was tested with a Mann-Whitney *U* test, while the significance of the differences in S_{TD}^* values between adult and immature stages was evaluated with a Student's *t*-test.

Results

A total of 4172 records of hard ticks (species of the genera Amblyomma, Dermacentor, Haemaphysalis, and Ixodes) collected from wild and domestic tetrapods were obtained. The data included 41 tick species for adult specimens (3007 records) and 22 for immature stages (1165 records) (table 1). Of the 4172 records, 1012 (24.2%) were from domestic mammals (cattle, goat, sheep, pig, horse, donkey, mule, dog, cat) and corresponded to adults of 22 tick species and immature stages of nine tick species. Number of host species, number of records and values of S_{TD}^* (calculated with and without domestic hosts) for each tick species are shown in table 1. In the cases where domestic hosts were excluded from the analysis, the difference in the values of S_{TD}^* for each species was not significant (see table 1) when the sample was analyzed as a whole (P=0.28, Student's *t*-test).

The distribution of number of host species was skewed for adults (fig. 1a), for immature ticks (fig. 1b) and also when immature and adults were grouped (fig. 1c). The figures clearly show that most tick species in this study parasitize between three and 20 different host species. No tick species has been associated either with a single species or with a single host genus. The mean number of hosts differed significantly (P=0.002, Mann-Whitney U test) between adults (mean= 12.56; median = 8; range = 3-49) and immatures (mean = 21.72; median = 15; range = 5–90). The frequency distribution of S_{TD}^* values are shown in fig. 2a for adults, in fig. 2b for immatures, and in fig. 2c for adults and immatures together. The most frequent values of S_{TD}^* were between 2.5 and 3.5. The average values of S_{TD}^* for adults and immatures were 2.78 (median =2.71; range=1.13-4.58) and 3.32 (median=3.31; range= 2.02–4.39), respectively (P=0.001, Student's t-test). A significant positive correlation between number of host species and number of records was found (r=0.64; $P \le 0.0001$) (fig. 3a). Conversely, there was no correlation of S_{TD}^* (r=0.19; P=0.12) with the number of records (fig. 3b). This absence of covariation indicates that S_{TD}^* , unlike the number of host species, is far less sensitive to sampling effort.

Discussion

This is the first attempt to describe general patterns of host specificity in Neotropical hard ticks through a quantitative approach. Although sometimes limited by incomplete data or lack of samples, in this study we generated a large database. We found a positive correlation between number of host species and number of records, which is in agreement with the findings reported by other authors for ticks (Klompen et al., 1996) and for other parasites (Poulin, 1992; Walther et al., 1995; Guégan & Kennedy, 1996; Walther & Morand, 1998). Therefore, the number of host species used by a particular tick species cannot be used as indicator of host specificity, because it can be an artefact caused by different sampling efforts. The index STD*, however, appears to be largely independent from sampling effort. Most tick species showed values of S_{TD}* ranging between 2.5 and 3.5, with an average close to 3 (table 1, fig. 2). These results show that, in general, an elevated proportion of tick species feeds on hosts that belong to different families or different orders. Naturally, some tick species are characterized by higher host specificity (STD* values between 1.13 and 2) (see table 1). However, none of them parasitize a single host species. When the analyses of number of hosts species and S_{TD}^* in adult and immature specimens were performed separately, both measures were significantly higher for immatures than for adults, which suggests that immature stages tend to use a broader taxonomic range of hosts than adults. Two hypotheses can be stated for

Table 1. Number of host species and values of the specificity index S _{TD} * proposed by Poulin & Mouillot (2005) (calculated with and without
domestic hosts) for the species of Neotropical hard ticks included in this study.

Tick species	No. of hosts species	No. of records	S_{TD}^*	S _{TD} * excluding domestic hosts
ADULTS				
Amblyomma albopictum	6	23	1.80	NA
Amblyomma argentinae	8	61	3.81	NA
Amblyomma aureolatum	20	152	3.04	3.31
Amblyomma auricularium	24	60	2.68	2.69
Amblyomma cajennense*	44	219	3.77	3.80
Amblyomma calcaratum	8	44	2.60	NA
Amblyomma coelebs	8	45	2.61	2.27
Amblyomma dissimile	49	231	3.48	3.41
Amblyomma dubitatum	5	76	2.99	2.97
Amblyomma fuscum	6	16	4.58	NA
Amblyomma geayi	6	37	3.40	NA
Amblyomma humerale	3	98	1.7	NA
Amblyomma incisum	3	92 70	1.82	NA 1.02
Amblyomma longirostre	14	70	1.99	1.93
Amblyomma naponense	8	33 150	2.47	NA 2 F7
Amblyomma neumanni	12 3		3.58	3.57
Amblyomma nodosum	3 19	31	2.31	NA 2.72
Amblyomma oblongoguttatum		43	3.59	3.72
Amblyomma ovale	33 6	152 40	3.40 2.76	3.29 2.75
Amblyomma pacae		40 19		
Amblyomma parkeri	3	19	1.67	NA
Amblyomma parvitarsum	3 33	330	2.20	2
Amblyomma parvum		19	3.64 1.41	3.65 NA
Amblyomma pecarium	3	35		
Amblyomma pseudoconcolor	12 8	35	2.54	2.43 2.74
Amblyomma pseudoparvum		38 17	2.88	
Amblyomma quadricavum	3 38	17 123	2.53	NA 2 E1
Amblyomma rotundatum	38 4	30	3.53 1.13	3.51 NA
Amblyomma sabanerae Amblyomma scalpturatum	4	63	1.13	NA
	15	117	2.50	2.69
Amblyomma tigrinum	15	45	3.65	3.62
Amblyomma triste Amblyomma varium	4	43 62	1.73	NA
Dermacentor nitens	11	117	2.31	**
Haemaphysalis juxtakochi	15	57	2.87	2.42
Haemaphysalis leporispalustris	4	29	1.84	NA
Ixodes auritulus	23	25	3.55	NA
Ixodes boliviensis	10	19	3.50	3.29
Ixodes loricatus	10	82	1.84	NA
Ixodes luciae	8	60	1.85	NA
Ixodes pararicinus	4	28	3.28	2
IMMATURE				
Amblyomma aureolatum	20	75	2.79	2.57
Amblyomma cajennense*	47	178	4.06	4.03
Amblyomma coelebs	12	16	4.39	4.44
Amblyomma dissimile	24	50	3.87	3.83
Amblyomma dubitatum	12	41	2.56	2.54
Amblyomma longirostre	90	140	3.30	NA
Amblyomma neumanni	12	77	3.61	3.57
Amblyomma nodosum	19	19	3.42	NA
Amblyomma ovale	18	30	3.87	3.74
Amblyomma parvum	11	22	3.04	3.12
Amblyomma rotundatum	13	24	3.34	NA
Amblyomma tigrinum	45	161	3.86	NA
Amblyomma triste	15	31	2.94	NA
Dermacentor nitens	5	59	2.73	***
Haemaphysalis juxtakochi	15	25	4.21	NA
Haemaphysalis leporispalustris	10	23	3.67	NA
Ixodes auritulus	42	53	3.26	NA
Ixodes longiscutatus	8	19	2.31	NA
Ixodes loricatus	15	35	3.02	NA
Ixodes luciae	20	42	3.05	NA
Ixodes pararicinus	13	17	3.99	NA
Ixodes sigelos	12	28	2.02	NA

NA, not applicable. * Recent studies suggested that *Amblyomma cajennense* is a species complex (Labruna *et al.*, 2011; Mastropaolo *et al.*, 2011). ** Because 89% of the records of adults of *D. nitens* were made on domestic animals, the analysis excluding domestic animals is unjustified. *** Because 87% of the records of immature stages of *D. nitens* were made on domestic animals, the analysis excluding domestic animals is unjustified.

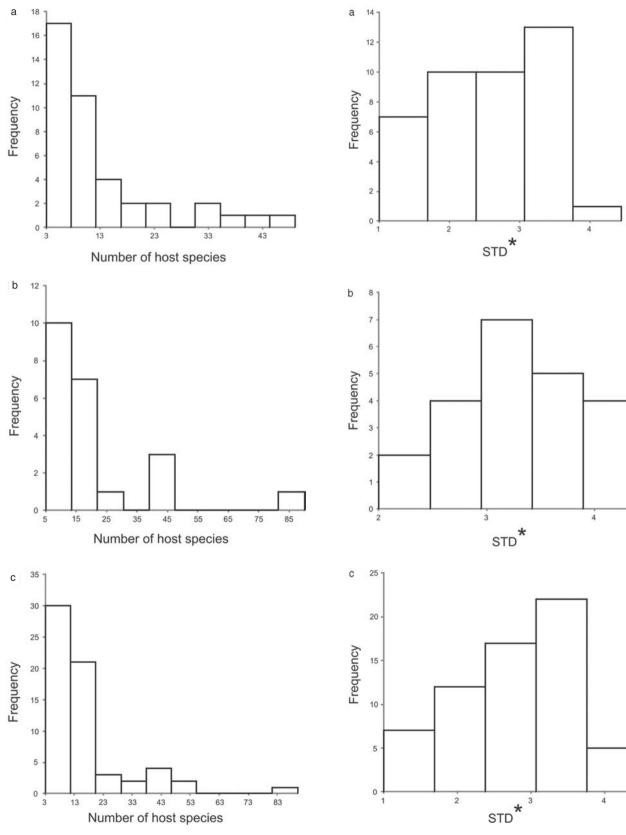


Fig. 1. Frequency distribution of number of host species among species of Neotropical hard ticks: (a) adults, (b) immature and (c) adults and immature grouped together.

Fig. 2. Frequency distribution of S_{TD}^* values among species of Neotropical hard ticks: (a) adults, (b) immature and (c) adults and immature grouped together.

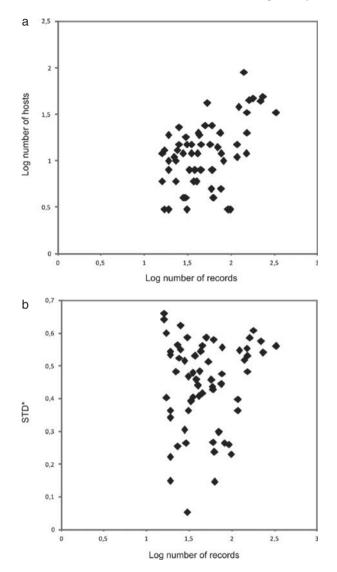


Fig. 3. Relationship between the number of records of each tick species included in this study and the number of (a) host species and (b) S_{TD}^* value per tick species (Pearson's correlation analysis).

these results. One of them is related to a physiological involvement, where larvae and nymphs may have greater adaptive plasticity than adults to feed on different host species. The other hypothesis is based on the number of host species available. If adult stages generally feed on hosts with larger body size (a fairly well-established trend), the number of host species available for those instars should be lower, given that there are relatively more small-bodied than largebodied host species in most environments.

Approximately 24% of the records were from domestic mammals, but the variation in S_{TD}^* was statistically insignificant when domestic hosts were excluded from the analysis (see table 1). The capacity of some domestic mammals to totally or partially sustain the life cycle of endemic tick species is well known, as in the cases, for instance, of *Amblyomma tigrinum*, *Amblyomma aureolatum*, *Amblyomma parvum*, *Amblyomma neumanni* and *Amblyomma triste* (Guglielmone

et al., 2000; Pinter *et al.*, 2004; Nava *et al.*, 2008, 2009, 2011). Although this fact highlights the ability of some tick species for rapid adaptation to recently introduced host species, the impact of non-endemic hosts on the ecology of native ticks does not alter the patterns of host specificity.

The application of index S_{TD}* to quantify host specificity in Neotropical hard ticks shows that a high proportion of ticks feed on phylogenetically distant host species. Similarities must be sought in other host characteristics, such as morphology, physiology or habitat usage, which appear to be more relevant. It has been established for other parasite taxa (e.g. helminthes of fishes, fleas of small mammals) that ecological similarity among host species is more important than host phylogeny (Poulin, 2005; Krasnov et al., 2007) and that external environmental factors may play a key role in the evolution of host-parasite associations (Krasnov et al., 2010). These conclusions are in agreement with the concept of ecological fitting, a process where organisms form a novel association with other species or use novel resources by ecological readjustment unrelated to previous evolutionary history, as a result of the suites of traits that they carry at the time they encounter the novel condition (Janzen, 1985; Agosta & Klemens, 2008). Ecological fitting is applicable to parasites when these can exploit a specific type of resource that is distributed across different host species (Brooks et al., 2006a,b). In these cases, the main requirement for the parasite is the resource itself, not the way that resource is packaged (the host species) (Brooks et al., 2006b). Support for ecological fitting in systems that involve parasites and hosts emerges from ecological and macroevolutionary evidence (Kethley & Johnston, 1975; Hoberg & Brooks, 2008; Kelly et al., 2009; Agosta et al., 2010). Following this concept, the association of a tick species with its hosts may be primarily determined by the environment occupied by an assemblage of suitable hosts, regardless of host phylogenetic relatedness. Host associations of larvae and nymphs of Ixodes loricatus and Ixodes luciae clearly illustrate ecological fitting. Small rodents (Cricetidae: Sigmodontinae) and marsupials (Didelphidae) are the principal hosts for immature stages of these two ticks (Guglielmone & Nava, 2011). Sigmodontine rodents and didelphids occupy the same ecological niche, but they are not phylogenetically related and have different evolutionary histories. The diversification of marsupials of the family Didelphidae in South America probably occurred between the Oligocene and Miocene (Steiner et al., 2005), while sigmodontines appeared on the continent only in the Pliocene (Pardiñas et al., 2010). This example, which could be applied to other tick species in this study, highlights the key role played by host ecological similarities in shaping hostparasite relationships in ticks. Also, our conclusions reject the hypothesis that coevolution was relevant in determining present tick-host associations, stressing the significance of host switching events during tick evolutionary history.

The widespread use of laboratory animals in experimental studies on life cycles or reproductive barriers is another issue that reinforces the idea of ticks as parasites with low host specificity. Hosts, such as rabbits (*Oryctolagus cuniculus*), guinea pigs (*Cavia porcellus*), chickens (*Gallus gallus*), rats (*Rattus norvegicus*), mice (*Mus musculus*), dogs (*Canis familiaris*), domestic pigs (*Sus scrofa*), horses (*Equus caballus*), among others, were successfully utilized to feed a wide spectrum of ticks under laboratory conditions. Some examples of this type of study in Neotropical hard ticks include the life cycle of *Amblyomma auricularium*, *A. parvum*, *A. neumanni*, *Amblyomma*

brasiliense, Amblyomma pseudoconcolor, Haemaphysalis leporispalustris and I. loricatus (Guglielmone et al., 1991; Aguirre et al., 1999; Labruna et al., 2000, 2002, 2003; Schumaker et al., 2000; Chacon et al., 2004; Pinter et al., 2004; Sanches et al., 2008; Szabó et al., 2009; Faccini et al., 2010; Olegário et al., 2011). Cross-mating trials to elucidate reproductive barriers between Amblyomma pseudoparvum and A. parvum, and among the members of the Amblyomma cajennense species complex, were carried out using rabbits as hosts (Guglielmone & Mangold, 1993; Labruna et al., 2011; Mastropaolo et al., 2011). These examples show the physiological plasticity exhibited by ticks to feed successfully when they are exposed to a novel host not related to the natural hosts.

The results of this investigation provide evidence that supports the hypothesis of Klompen *et al.* (1996) that ticks are habitat- rather than host-specialists. When one considers that hard ticks spend most of their life cycle off host, this conclusion is not unexpected. However, and regardless of habitat specificity, other variables, such as size, parasite generation time, phenology, time spent off host and the type of life-cycle (three-, two- or one-host), should be considered for future studies, as they are factors which can influence tick host associations.

By using two measures of host specificity, we describe a general pattern based on a restricted data set pertaining to specific region of the world (Neotropical region). We do not pretend that our conclusions will be applicable everywhere else. Local adaptations or the size of the geographical range of a parasite species may cause intraspecific variations in host specificity across its distribution (Kaltz & Shykoff, 1998; Jackson & Tinsley, 2005; Krasnov *et al.*, 2005, 2011; Morgan *et al.*, 2005; Gandon *et al.*, 2008), and these variables should also be included in future analyses. Similarly, it would be interesting to evaluate the levels of fitness achieved by a particular tick species after feeding on different hosts species, through the measurement of biological parameters. Nevertheless, the evidence presented in this work shows that strict host specificity is not common among hard ticks.

Acknowledgements

We are very grateful to Lorenza Beati, Robert Poulin and Marcelo B. Labruna for their valuable comments on an earlier version of the manuscript. We thank the financial support of Instituto Nacional de Teconlogía Agropecuaria, Asociación Cooperadora of EEA-INTA Rafaela and Consejo Nacional de Investigaciones Científicas y Técnicas, Argentina.

Supplementary material

The online appendix can be viewed at http://journals. cambridge.org/ber.

References

- Agosta, S.J. & Klemens, J.A. (2008) Ecological fitting by phenotypically flexible genotypes: implications for species associations, community assembly and evolution. *Ecology Letters* 11, 1123–1134.
- Agosta, S.J., Janz, N. & Brooks, D.R. (2010) How specialists can be generalists: resolving the "parasite paradox" and implications for emerging disease. *Zoologia*. 27, 151–162.
- Aguirre, D.H., Viñabal, A.E. & Guglielmone, A.A. (1999) The life cycle of *Amblyomma neumanni* Ribaga, 1902 (Acari: Ixodidae)

in the laboratory. *Experimental and Applied Acarology* **23**, 159–164.

- Balashov, Y.S. (2004) The main trends in the evolution of ticks (Ixodida). Entomological Review 84, 814–824.
- Barquez, R.M., Díaz, M.M. & Ojeda, R.A. (2006) Mamíferos de Argentina: Sistemática y Ddistribución. Tucumán, Argentina, Sociedad Argentina para el Estudio de los Mamíferos (SAREM).
- Brooks, D.R. & Hoberg, E.P. (2007) How will global climate change affect parasite-host assemblages? *Trends in Parasitology* 23, 571–574.
- Brooks, D.R. & McLennan, D.A. (1993) Parascript: Parasites and the Language of Evolution. Washington DC, USA, Smithsonian Institution Press.
- Brooks, D.R., Leon-Regagnon, V., McLennan, D.A. & Zelmer, D. (2006a) Ecological fitting as a determinant of the community structure of platyhelminth parasites of anurans. *Ecology* 87, 76–85.
- Brooks, D.R., McLennan, D.A., Leon-Regagnon, V. & Hoberg, E. (2006b) Phylogeny, ecological fitting and lung flukes: helping solve the problem of emerging infectious diseases. *Revista Mexicana de Biodiversidad* 77, 225–233.
- Chacon, S.C., Faccini, J.L.H. & Bittencourt, V.R.E.P. (2004) Successful infestation by *Amblyomma pseudoconcolor* and *A. cooperi* (Acari: Ixodidae) on horses. *Annals of New York Academic Sciences* 1026, 232–234.
- Clements, J.F. (2007) *The Clements Checklist of Birds of the World*. 6th edn. Ithaca, NY, USA, Cornell University Press.
- Cumming, G.S. (1998) Host preference in African ticks (Acari: Ixodida): a quantitative data set. Bulletin of Entomological Research 88, 379–406.
- Cumming, G.S. (2002) Comparing climate and vegetation as limiting factors for species ranges of African ticks. *Ecology* 83, 255–268.
- D'Elía, G., Pardiñas, U.F.J., Jayat, P. & Salazar-Bravo, J. (2008) Systematics of *Necromys* (Rodentia, Cricetidae, Sigmodontinae): species limits and groups, with comments on historical biogeography. *Journal of Mammalogy* 89, 778–790.
- Dick, C.W. & Patterson, B.D. (2007) Against all odds: explaining high host specificity in dispersal-prone parasites. *Inter*national Journal for Parasitology 37, 871–876.
- Faccini, J.L.H., Cardoso, A.C.B., Onofrio, V.C., Labruna, M.B. & Barros-Battesti, D.M. (2010) The life cycle of Amblyomma auricularium (Acari: Ixodidae) using rabbits (Oryctolagus cuniculus) as experimental host. Experimental and Applied Acarology 50, 71–77.
- Fenton, A. & Pedersen, A.B. (2005) Community epidemiology framework for classifying disease threats. *Emerging Infectious Diseases* 11, 1815–1821.
- Francés, J. & D'Elia, G. (2006) Oligoryzomys delticola es sinónimo de O. nigripes (Rodentia, Cricetidae, Sigmodontinae). Mastozoología Neotropical 13, 123–131.
- Gandon, S., Buckling, A., Decaestecker, E. & Day, T. (2008) Host-parasite coevolution and patterns of adaptation across time and space. *Journal of Evolutionary Biology* 21, 1861–1866.
- Guégan, J.F. & Kennedy, C.R. (1996) Parasite richness/sampling effort/host range: the fancy three-piece jigsaw puzzle. *Parasitology Today* 12, 367–369.
- Guglielmone, A.A. & Mangold, A.J. (1993) Cross mating between Amblyomma parvum Aragao, 1908 and Amblyomma pseudoparvum Guglielmone, Mangold and Keirans, 1990. Folia Parasitologica 40, 144–145.

- Guglielmone, A.A. & Nava, S. (2010) Hosts of Amblyomma dissimile Koch, 1844 and Amblyomma rotundatum Koch, 1844 (Acari: Ixodidae). Zootaxa 2541, 27–49.
- Guglielmone, A.A. & Nava, S. (2011) Rodents of the subfamily Sigmodontinae (Myomorpha: Cricetidae) as hosts for South American hard ticks (Acari: Ixodidae) with hypothesis on life history. Zootaxa 2904, 45–65.
- Guglielmone, A.A., Mangold, A.J. & Garcia, M.D. (1991) The life cycle of *Amblyomma parvum* Aragao, 1908 (Acari: Ixodidae) under laboratory conditions. *Experimental and Applied Acarology* 13, 129–136.
- Guglielmone, A.A., Mangold, A.J., Luciani, C.E. & Viñabal, A.E. (2000) Amblyomma tigrinum (Acari: Ixodidae) in relation to phytogeography of central-northern Argentina with note on hosts and seasonal distribution. Experimental and Applied Acarology 24, 983–989.
- Guglielmone, A.A., Robbins, R.G., Apanaskevich, D.A., Petney, T.A., Estrada-Peña, A., Horak, I.G., Shao, R. & Barker, S.C. (2010) The Argasidae, Ixodidae and Nuttalliellidae (Acari: Ixodida) of the world: a list of valid names. *Zootaxa* 2528, 1–28.
- Hoberg, E.P. & Brooks, D.R. (2008) A macroevolutionary mosaic: episodic host-switching, geographical colonization and diversification in complex host-parasite systems. *Journal of Biogeography* 35, 1533–1550.
- Hoogstraal, H. (1978) Biology of ticks. pp. 3–14 in *Tick borne diseases and their vectors: Proceedings of the International Conference of Edinburgh*, 1978, University of Edinburgh, September 1976, Edinburgh, UK.
- Hoogstraal, H. & Aeschlimann, A. (1982) Tick-host specificity. Bulletin de la Société Entomologique Suisse 55, 5–32.
- Hoogstraal, H. & Kim, K.C. (1985) Tick and mammal coevolution, with emphasis on *Haemaphysalis*. pp. 505–568 *in* Kim, K.C. (*Ed.*) *Coevolution of Parasitic Arthropods and Mammals*. London, UK, London University Press.
- Jackson, J.A. & Tinsley, R.C. (2005) Geographic and withinpopulation structure in variable resistance to parasite species and strains in a vertebrate host. *International Journal for Parasitology* 35, 29–37.

Janzen, D.H. (1985) On ecological fitting. Oikos 45, 308-310.

- Kaltz, O. & Shykoff, J.A. (1998) Local adaptation in host-parasite systems. *Heredity* 81, 361–370.
- Kelly, D.W., Paterson, R.A., Townsend, C.R., Poulin, R. & Tompkins, D.M. (2009) Parasite spillback: a neglected concept in invasión ecology? *Ecology* **90**, 2047–2056.
- Kethley, J.B. & Johnston, D.E. (1975) Resource tracking patterns in bird and mammal ectoparasites. *Miscellaneous Publications* of the Entomological Society of America 9, 231–236.
- Klompen, J.S.H., Black, W.C., Keirans, J.E. & Oliver, J.H. (1996) Evolution of ticks. Annual Review of Entomology 41, 141–161.
- Krasnov, B.R., Poulin, R., Shenbrot, G.I., Mouillot, D. & Khokhlova, I.S. (2005) Host specificity and geographic range in haematophagous ectoparasites. *Oikos* 108, 449–456.
- Krasnov, B.R., Korine, C., Burdelova, N.V., Khokhlova, I.S. & Pinshow, B. (2007) Between-host phylogenetic distance and feeding efficiency in hematophagous ectoparasites: rodent fleas and a bat host. *Parasitology Research* **101**, 365–371.
- Krasnov, B.R., Korallo-Vinarskaya, N.P., Vinarski, M.V., Shenbrot, G.I., Mouillot, D. & Poulin, R. (2008) Searching for general patterns in parasite ecology: host identity versus environmental influence on gamasid mite assemblages in small mammals. *Parasitology* 135, 229–242.
- Krasnov, B.R., Mouillot, D., Shenbrot, G.I., Khokhlova, I.S., Vinarski, M.V., Korallo-Vinarskaya, N.P. & Poulin, R.

(2010) Similarity in ectoparasites faunas of Palaeartic rodents as a function of host phylogenetic, geographic or environmental distances: which matters the most? *International Journal for Parasitology* **40**, 807–817.

- Krasnov, B.R., Mouillot, D., Shenbrot, G.I., Khokhlova, I.S. & Poulin, R. (2011) Beta specificity: the turnover of host species in space and another way to measure host specificity. *International Journal for Parasitology* **41**, 33–41.
- Labruna, M.B., Cerqueira Leite, R., Faccini, J.R.L. & Ferreira, F. (2000) Life cycle of the tick *Haemaphysalis leporis-palustris* (Acari: Ixodidae) under laboratory conditions. *Experimental* and Applied Acarology 24, 683–694.
- Labruna, M.B., Souza, S.L.P., Menezes, A.C., Horta, M.C., Pinter, A. & Gennari, S.M. (2002) Life cycle and host specificity of *Amblyomma tigrinum* (Acari: Ixodidae) under laboratory conditions. *Experimental and Applied Acarology* 24, 115–125.
- Labruna, M.B., Fugisaki, E.Y.M., Pinter, A., Duarte, J.M.B. & Szabó, M.J.P. (2003) Life cycle and host specificity of *Amblyomma triste* (Acari: Ixodidae) under laboratory conditions. *Experimental and Applied Acarology* **30**, 305–316.
- Labruna, M.B., Soares, J.F., Martins, T.F., Soares, H.S. & Cabrera, R.R. (2011) Cross-mating experiments with geographically different populations of *Amblyomma cajennense* (Acari: Ixodidae). *Experimental and Applied Acarology* 54, 41–49.
- Mastropaolo, M., Nava, S., Guglielmone, A.A. & Mangold, A.J. (2011) Biological differences between two allopatric populations of *Amblyomma cajennense* (Acari: Ixodidae) in Argentina. *Experimental and Applied Acarology* 53, 371–375.
- Morgan, A.D., Gandon, S. & Buckling, A. (2005) The effect of migration on local adaptation in a coevolving host-parasite system. *Nature* 437, 253–256.
- Nava, S., Mangold, A.J. & Guglielmone, A.A. (2008) Aspects of the life cycle of *Amblyomma parvum* (Acari: Ixodidae) under natural conditions. *Veterinary Parasitology* 156, 270–276.
- Nava, S., Estrada-Peña, A., Mangold, A.J. & Guglielmone, A.A. (2009) Ecology of *Amblyomma neumanni* (Acari: Ixodidae). *Acta Tropica* 111, 226–236.
- Nava, S., Mangold, A.J., Mastropaolo, M., Venzal, J.M., Fracassi, N. & Guglielmone, A.A. (2011) Seasonal dynamics and hosts of *Amblyomma triste* (Acari: Ixodidae) in Argentina. *Veterinary Parasitology* 181, 301–308.
- Olegário, M.M., Gerardi, M., Tsuruta, S.A. & Szabó, M.P. (2011) The life cycle of the tick *Amblyomma parvum* Aragao, 1908 (Acari: Ixodidae) and suitability of domestic hosts under laboratory conditions. *Veterinary Parasitology* **179**, 203–208.
- Pardiñas, U.F.J., Teta, P. & D'Elia, G. (2010) Roedores sigmodontinos de la región pampeana: historia evolutiva, sistemática y taxonomía. pp. 9–36 in Polop, J.J. & Busch, M. (Eds) Biología y Ecología de Pequeños Roedores en la Región Pampeana Argentina: Enfoques y Perspectivas. Córdoba, Argentina, Editorial de la Universidad Nacional de Córdoba.
- Pinter, A., Dias, R.A., Gennari, S.M. & Labruna, M.B. (2004) Study of the seasonal dynamics, life cycle, and host specificity of *Amblyomma aureolatum* (Acari: Ixodidae). *Journal of Medical Entomology* **41**, 321–332.
- Poulin, R. (1992) Determinants of host-specificity in parasites of freshwater fishes. *International Journal for Parasitology* 22, 753–758.
- **Poulin, R.** (2005) Relative infection levels and taxonomic distances among the host species used by a parasite: insights into parasite specialization. *Parasitology* **130**, 109–115.

- Poulin, R. (2007) Evolutionary Ecology of Parasites. 2nd edn. Princenton, NJ, USA, Princeton University Press.
- Poulin, R. & Mouillot, D. (2005) Combining phylogenetic and ecological information into a new index of host specificity. *Journal of Parasitology* 91, 511–514.
- Poulin, R., Krasnov, B.R., & Morand, S. (2006) Patterns of host specificity in parasites exploiting small mammals. pp. 233–256 in Morand, S., Krasnov, B.R. & Poulin, R. (Eds) Micromammals & Macroparasites: From Evolutionary Ecology to Management. Tokyo, Japan, Springer-Verlag.
- Sanches, G.S., Bechara, G.H., Garcia, M.V., Labruna, M.B. & Szabó, M.P.J. (2008) Biological aspects of Amblyomma brasiliense (Acari: Ixodidae) under laboratory conditions. Experimental and Applied Acarology 44, 43–48.
- Schumaker, T.T.S., Labruna, M.B., Dos Santos, A.I., Soares Clerici, P.T. (2000) Life cycle of *Ixodes* (*Ixodes*) *loricatus* under laboratory conditions. *Journal of Medical Entomology* 37, 714–720.
- Steiner, C., Tilak, M., Douzery, E.J.P. & Catzeflis, F.M. (2005) New DNA data from a transthyretin nuclear intron suggest an Oligocene to Miocene diversification of living South America opossums (Marsupialia: Didelphidae). *Molecular Phylogenetics and Evolution* 35, 363–379.

- Szabó, M.P.J., Pereira, L.F., Castro, M.B., Garcia, M.V., Sanches, G.S. & Labruna, M.B. (2009) Biology and life cycle of Amblyomma incisum (Acari: Ixodidae). Experimental and Applied Acarology 48, 263–271.
- Voss, R.S. & Jansa, S.A. (2009) Phylogenetic relationships and classification of didelphid marsupials, an extant radiation of New World metatherian mammals. *Bulletin of the American Museum of Natural History* 322, 1–177.
- Walther, B.A. & Morand, S. (1998) Comparative performance of species richness estimation methods. *Parasitology* 116, 393–405.
- Walther, B.A., Cotgreave, P., Price, R.D., Gregory, R.D. & Clayton, D.H. (1995) Sampling effort and parasite species richness. *Parasitology Today* 11, 306–310.
- Weksler, M., Percequillo, A.R. & Voss, R.S. (2006) Ten new genera of oryzomyne rodents (Cricetidae: Sigmodontinae). *American Museum Novitates* 3537, 1–29.
- Wilson, D.E. & Reeder, D.M. (2005) Mammal Species of the World: A Taxonomic and Geographic Reference. 3rd edn. Baltimore, MD, USA, Johns Hopkins University Press.
- Wolinska, J. & King, K.C. (2009) Environment can alter selection in host-parasite interactions. *Trends in Parasitology* 25, 236–244.