

The benthos and fish of offshore sandbank habitats in the southern North Sea

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Meiofauna, macro-infauna and epifauna were sampled at two offshore sandbanks in the southern North Sea in 2006. The epifaunal and infaunal communities in the dynamic environment of the sandbank crests were species poor, with lesser weever, solenette, Crangon crangon, amphipods and deposit-feeding polychaetes the predominant benthic taxa. Abundant early life-history stages (24–39 mm) of the lesser weever Echiichthys vipera on the sandbank crests indicated that these habitats may be important nursery grounds for this species. Species diversity of infauna and epifauna was greater in the deeper waters parallel to the sandbanks. Contrasting patterns were evident for meiofaunal nematodes, where communities collected on the sandbank crests were more diverse than those recorded in the deeper off-bank sites. The fauna of sandbank crests is composed of a restricted range of the fauna typically associated with sandy habitats, particularly taxa adapted to live in this dynamic environment.

Keywords: nematodes, infauna, epifauna, demersal fish, Trachinidae, nursery ground, beam trawl, Swarte Bank, Broken Bank, Norfolk, North Sea

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INTRODUCTION

The North Norfolk Sandbanks and Saturn Reef are currently identified as a possible offshore Special Area of Conservation (SAC) under the Habitats Directive (EU, 1992). One of the major interest features in the area are ‘Sandbanks which are slightly covered by seawater all the time’, one of the habitats listed on Annex I of the Directive. For these purposes, sandbanks have been defined as ‘elevated, elongated, rounded or irregular topographic features, permanently submerged and predominantly surrounded by deeper water’, and that they are ‘slightly covered by sea water all the time’ means that the water depth above the sandbank is ‘seldom more than 20 m below chart datum’ (CEC, 2007).

The North Norfolk sandbanks site comprises ten linear sandbanks (Figure 1), in addition to several smaller, fragmented banks. Although the proposed SAC covers a total area of 4327 km², the area of the sandbanks themselves is smaller and individual banks range in size from approximately 16.5–135 km². The hydrodynamics and sediment movement associated with North Sea sandbanks have been reported in several studies (e.g. Howarth & Huthnance, 1984; Pan *et al.*, 2007), and these indicate that there is a local clockwise circulation of sand and water around the banks as part of the broad scale hydrodynamics of the southern North Sea (Collins *et al.*, 1995).

The characteristic fauna of such habitats may include polychaetes, crustaceans, anthozoans, bivalves, echinoderms and various fish species (including sand eels *Ammodytes* spp., dragonets *Callionymus* spp., sand gobies *Pomatoschistus* spp., lesser weever *Echiichthys vipera*, plaice *Pleuronectes*

platessa and dab *Limanda limanda*) (CEC, 2007). However, the seabed topography, shallow water depth and complex hydrodynamics make these environments difficult to sample, and so quantitative data on the fauna of offshore sandbanks are scarce, and there have been no recent published studies of the fauna of the North Norfolk sandbanks.

Sandbanks are important habitats that can help protect nearby coastlines (Pan *et al.*, 2007) and may also be exploited by the marine aggregate industry (e.g. Poiner & Kennedy, 1984; Moolaert & Hostens, 2007), but little is known about their ecological importance. Inshore sandbanks off the Dutch and Belgian coasts have been the subject of some investigations, including the sampling of meiofauna (Willems *et al.*, 1982b; Vanaverbeke *et al.*, 2000, 2002), macrofauna (Vanosmael *et al.*, 1982) and the suprabenthos (Dewicke *et al.*, 2003), but there is a lack of comparable information for the North Norfolk sandbanks which are of current conservation interest. Elsewhere in UK seas, Kaiser *et al.* (2004) undertook replicate 2 m and 4 m beam trawl sampling to examine the fish and larger epifauna associated with several sandbanks in Welsh coastal waters, which highlighted the low species diversity of such habitats. There are also some localized studies on estuarine and inshore sandbanks along the south coast of England (e.g. Holme, 1949; Withers & Thorp, 1978).

Effective conservation of such habitats in European seas requires specific conservation objectives. These need to be achieved by closely linked management measures that regulate the activities of those marine industries which may threaten the integrity of the habitat, or the viability of the associated populations (Pedersen *et al.*, 2009). Although the conservation objectives for the North Norfolk sandbanks have not yet been identified, those for sandbanks in German waters require the maintenance of characteristic morphological and hydrodynamic conditions, habitat structure and surface area,

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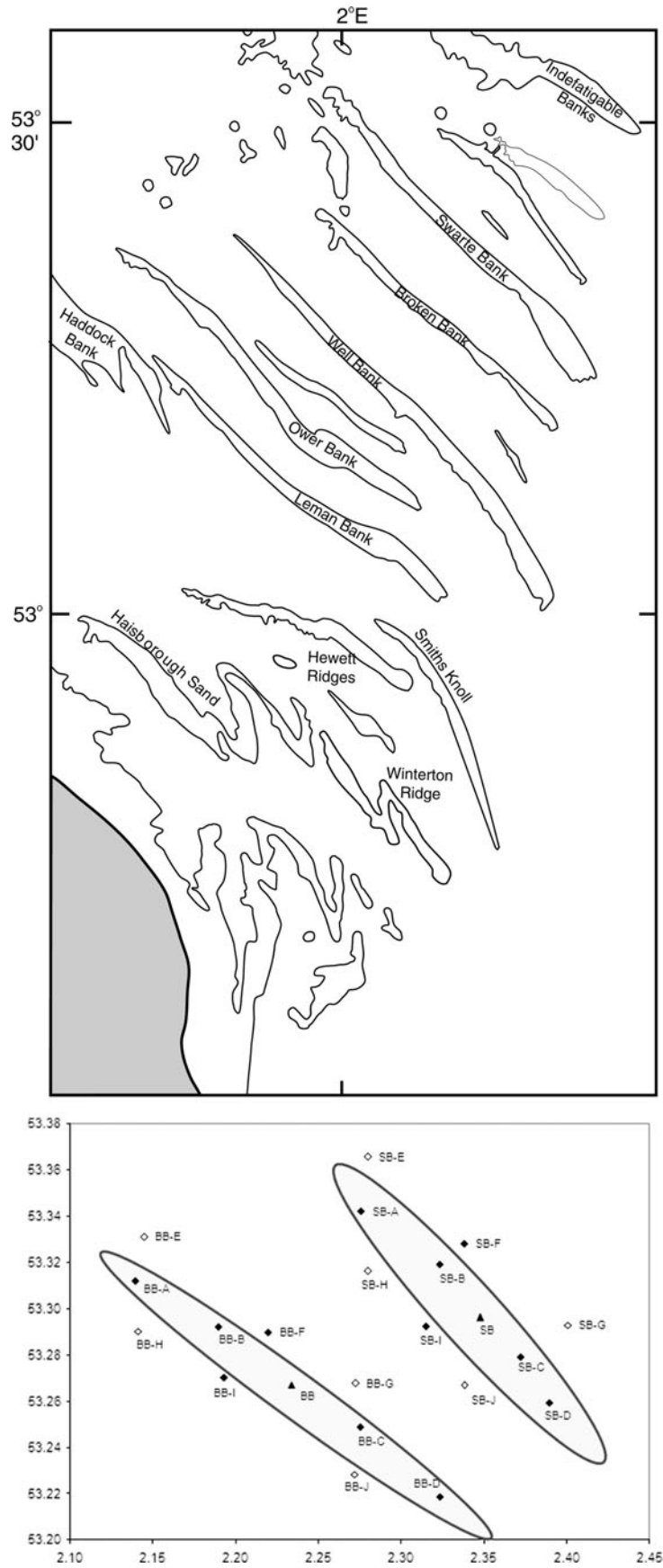


Fig. 1. Location of North Norfolk sandbanks (top) and sampling grid (bottom) indicating main sites (solid triangles: replicated 2 m beam trawl and infaunal sampling; solid diamonds: single 2 m beam trawl and infaunal sampling) and additional sites sampled with 2 m beam trawl and sediment sampling (open diamond).

Table 1. Physical parameters from the two sandbanks sampled, giving sediment composition for silt–clay (grain sizes <63 µm), sand (63 µm–1.9 mm), gravel content (≥2 mm), and water depth (m).

Physical parameter		Broken Bank		Swarte Bank	
		Sandbank	Off-bank	Sandbank	Off-bank
Silt–clay (%)	Mean	0.09	1.35	0.13	1.45
	Range	(0.06–0.16)	(1.06–1.91)	(0.08–0.18)	(0.48–3.63)
Sand (%)	Mean	99.9	97.43	99.86	96.83
	Range	(99.82–99.94)	(94.19–98.42)	(99.81–99.91)	(95.35–99)
Gravel (%)	Mean	0.01	1.22	0.02	1.73
	Range	(0–0.02)	(0.08–4.37)	(0–0.04)	(0.27–3.81)
Water depth (m)	Mean	18.9	35.9	17.4	35.4
	Range	(17.3–21.0)	(33.0–39.3)	(14.5–19.8)	(27.4–40.2)

and the characteristic benthic communities and benthic species (Pederson *et al.*, 2009). In order to achieve these management goals detailed descriptions of the conservation features, with an indication of their sensitivity to the pressures of offshore activities, are required. The aim of the present paper is therefore to provide the first detailed description of the fauna associated with the linear sandbanks off the North Norfolk coast, in preparation for the development of such conservation objectives. The communities observed are also compared with those described elsewhere in northern European seas.

MATERIALS AND METHODS

Description of study area

The North Norfolk sandbanks comprise a complex of linear sandbanks (Figure 1). The more inshore banks (Leman, Ower and Well Banks) have extensive shallower areas (water depth above the sandbank <10 m), while the more

offshore Broken Bank, Swarte Banks and Indefatigable Banks are slightly deeper (water depth above the sandbank <20 m). For practical reasons, field studies focused on the Swarte and Broken Banks, as other banks were either too shallow for surveying by research vessel and/or had offshore gas installations and pipelines in the area.

Biological sampling

The meiofauna, macro-infauna and epifauna of the Swarte and Broken Banks (Figure 1) were sampled with a range of benthic sampling gears during April 2006.

The epifauna was sampled using a steel 2 m beam trawl with chain mat (see Jennings *et al.* (1999) for a gear description). Tows were of 5 minutes duration with a warp:depth ratio of 3:1. Overall, 15 epifauna samples were collected for each bank, including five replicate samples (at one site) and four individual samples (four sites) along the crest of the bank, and single samples from three sites on either side of the bank. For logistic reasons, no trawling was undertaken on the steepest parts of the slopes. The larger epifauna and

Table 2. Dominant fish and epifauna (individuals per tow) captured by 2 m beam trawl on the sandbank crests (average similarity = 68.7%) and at off-bank sites (average similarity = 62.8%), showing the average abundance, average similarity, and the relative and cumulative contributions to the similarity. Data root-transformed.

Sandbank	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
<i>Echiichthys vipera</i>	6.16	21.05	7.83	30.67	30.67
<i>Buglossidium luteum</i>	3.72	12.42	5.46	18.09	48.76
<i>Crangon crangon</i>	3.49	10.77	2.20	15.69	64.44
<i>Arnoglossus laterna</i>	1.90	6.02	3.21	8.76	73.21
<i>Ophiura ophiura</i>	1.67	4.90	2.10	7.13	80.34
<i>Limanda limanda</i>	1.63	4.61	1.59	6.71	87.05
<i>Pomatoschistus</i> sp.	1.13	2.49	0.98	3.62	90.67
Off-bank	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
<i>Ophiura albida</i>	13.01	14.75	3.06	23.49	23.49
<i>Buglossidium luteum</i>	7.11	10.71	4.62	17.06	40.55
<i>Ophiura ophiura</i>	4.25	6.11	3.81	9.73	50.28
<i>Pomatoschistus</i> sp.	3.69	5.31	4.39	8.45	58.73
<i>Pagurus bernhardus</i>	2.86	4.00	2.21	6.37	65.11
<i>Liocarcinus holsatus</i>	2.22	2.76	2.88	4.40	69.51
<i>Crangon crangon</i>	1.90	2.67	1.69	4.26	73.76
<i>Crangon allmani</i>	1.85	2.60	3.32	4.15	77.91
<i>Arnoglossus laterna</i>	1.60	2.37	2.88	3.78	81.69
<i>Callionymus lyra</i>	1.69	2.17	1.71	3.45	85.14
Ammodytidae	2.52	1.93	1.22	3.08	88.22
<i>Asterias rubens</i>	1.36	1.34	1.01	2.13	90.35

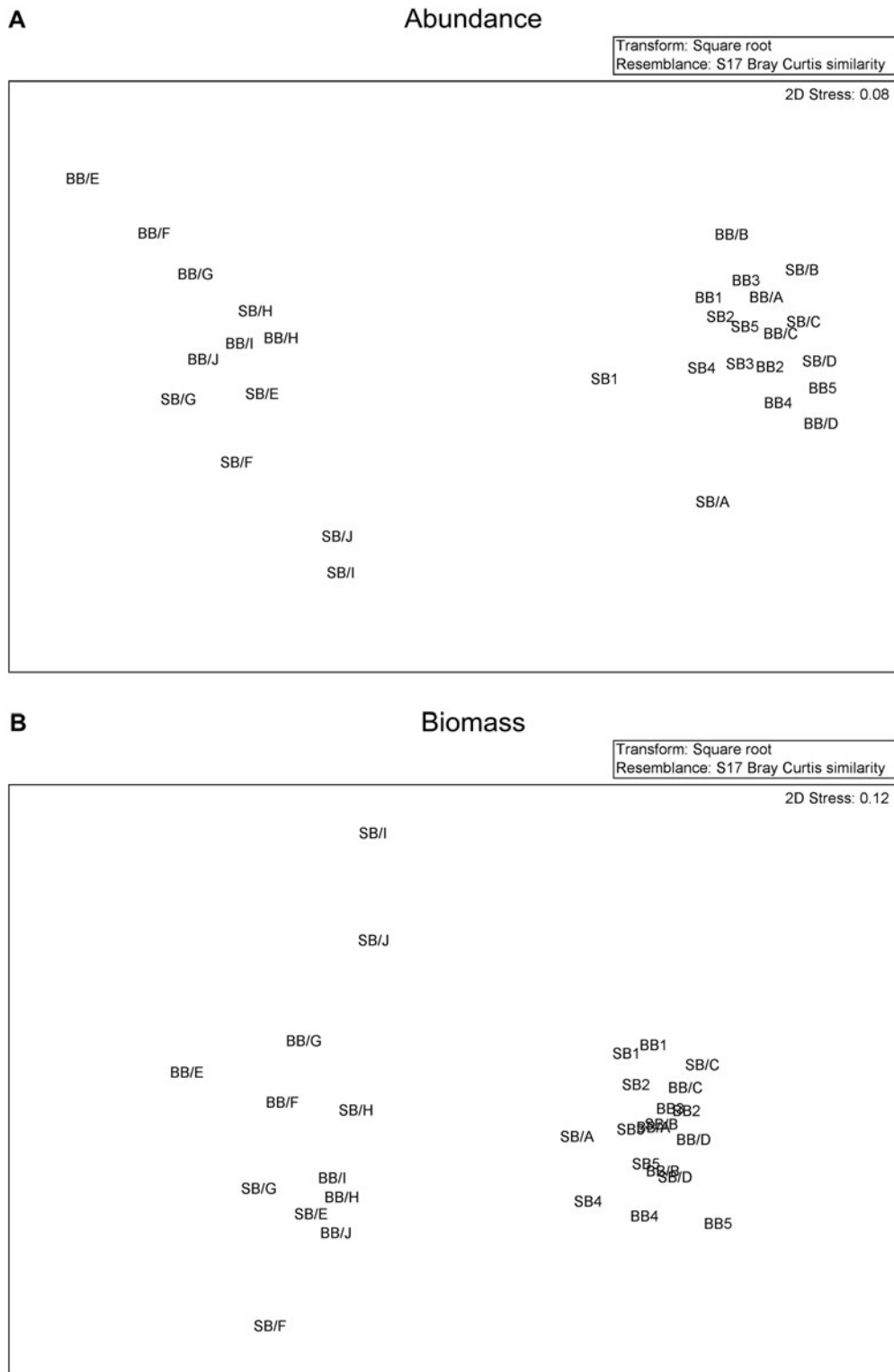


Fig. 2. Multidimensional scaling plots of fish and epifaunal catches in 2 m beam trawl from the Swarte Bank (SB) and Broken Bank (BB), with 1–5 replicate stations on the centre of the bank, A–D single stations along the bank, and E–J off-bank stations. Average dissimilarity between bank and off-bank samples by (A) abundance and (B) biomass data were 62.8% and 65.2%, respectively.

demersal fish were also sampled with 4 m beam trawl, with two tows (of 30 minutes duration) between and parallel to the sandbanks. Catches were fully sorted with invertebrates and demersal fish identified to the lowest taxonomic level, and the biomass and numbers of each species recorded. If a

species was very abundant, the total weight was recorded and a sub-sample of known weight was enumerated. Taxa that are not sampled effectively by 2 m beam trawl (e.g. mysids, nemertean and small polychaetes) were excluded from data analyses.

Table 3. Differences in the fish and epifauna of sandbank crests and off-bank sites (average dissimilarity = 62.8%), showing the average abundance, the average dissimilarity by species, and the relative and cumulative contributions to the dissimilarity. See Clarke & Gorley (2006) for further information.

Species	Average abundance					
	Sandbank	Off-bank	Av.Diss	Diss/SD	Contrib%	Cum.%
<i>Ophiura albida</i>	0.70	13.01	14.01	2.27	22.32	22.32
<i>Echiichthys vipera</i>	6.16	0.43	7.19	3.22	11.47	33.79
<i>Buglossidium luteum</i>	3.72	7.11	4.08	1.97	6.50	40.29
<i>Ophiura ophiura</i>	1.67	4.25	3.13	2.25	4.99	45.28
<i>Pomatoschistus</i> sp.	1.13	3.69	3.11	1.84	4.96	50.23
<i>Pagurus bernhardus</i>	0.57	2.86	3.02	1.45	4.81	55.04
<i>Echinocardium cordatum</i>	0.36	2.26	2.76	0.65	4.40	59.44
Ammodytidae	0.58	2.52	2.64	0.80	4.21	63.65
<i>Crangon crangon</i>	3.49	1.90	2.29	1.85	3.66	67.31
<i>Liocarcinus holsatus</i>	0.32	2.22	2.27	1.76	3.61	70.92
<i>Callionymus lyra</i>	0.22	1.69	1.85	1.69	2.94	73.86
<i>Spisula</i> sp.	0.00	1.37	1.60	1.38	2.55	76.41
<i>Asterias rubens</i>	0.28	1.36	1.51	1.22	2.41	78.81
<i>Crangon allmani</i>	1.01	1.85	1.45	1.33	2.30	81.12

Five replicate 0.1 m² Day grabs were collected at five stations along the crest of each sandbank and at single sites either side of the sandbanks. Additional, non-replicated Day grabs were collected at four other stations located on the deeper surrounding seabed of each sandbank. From each Day grab, two sub-samples were collected with a Perspex corer (3.6 cm diameter, 10 cm² surface area) to a depth of 5 cm; one for particle size analysis (PSA) and one for the study of meiofaunal nematodes. All faunal samples were fixed in 5% formaldehyde in 63 µm filtered seawater. Replicated sub-samples for PSA were combined by station and frozen at -20°C prior to analysis.

After thawing, the 22 sediment samples were wet sieved through a 500 µm sieve, and the fraction >500 µm was oven-dried at 90°C for 24 hours. This fraction was then dry sieved at 0.5 phi intervals, down to 1 phi (500 µm). The fraction <500 µm was freeze dried and analysed on a Coulter LS 130 Laser sizer.

A total of 35 macro-infauna samples were processed and analysed from each bank (i.e. five replicates from each of the five stations along the sandbank and the two off-bank sites). Samples were initially puddled over a 1 mm sieve to remove the excess sediment and the remaining fauna and sediment were fixed in 5% formaldehyde in 63 µm filtered

Table 4. Dominant fish and epifauna (biomass) captured by 2 m beam trawl on the sandbank crests (average similarity = 64.4%) and off-bank sites (average similarity = 53.0%). Data root-transformed. See Table 2 for further information on data presented.

Sandbank	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
<i>Echiichthys vipera</i>	14.61	22.28	4.96	34.61	34.61
<i>Buglossidium luteum</i>	8.53	13.10	4.95	20.34	54.95
<i>Arnoglossus laterna</i>	5.98	8.50	3.16	13.20	68.14
<i>Limanda limanda</i>	5.51	5.04	0.94	7.82	75.97
<i>Crangon crangon</i>	3.30	4.69	2.58	7.28	83.24
<i>Ophiura ophiura</i>	3.33	4.18	2.18	6.49	89.74
<i>Pomatoschistus</i> sp.	0.98	0.96	0.97	1.49	91.22
Off-bank	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
<i>Buglossidium luteum</i>	16.18	11.64	4.44	21.98	21.98
<i>Ophiura ophiura</i>	7.86	5.63	3.64	10.62	32.61
<i>Ophiura albida</i>	8.67	5.02	2.56	9.47	42.08
<i>Callionymus lyra</i>	5.32	3.42	2.00	6.47	48.54
<i>Arnoglossus laterna</i>	4.91	3.33	2.79	6.28	54.82
<i>Echinocardium cordatum</i>	13.98	2.97	0.54	5.61	60.43
<i>Pomatoschistus</i> sp.	3.89	2.70	3.99	5.09	65.52
<i>Liocarcinus holsatus</i>	4.73	2.52	1.89	4.76	70.28
<i>Pagurus bernhardus</i>	4.19	2.47	2.02	4.66	74.94
<i>Asterias rubens</i>	4.81	1.97	0.96	3.72	78.67
Ammodytidae	3.88	1.69	1.19	3.19	81.86
<i>Sabellaria spinulosa</i>	5.43	1.33	0.43	2.52	84.38
<i>Solea solea</i>	4.71	1.17	0.41	2.21	86.58
<i>Crangon allmani</i>	1.72	1.15	3.07	2.17	88.75
<i>Limanda limanda</i>	3.29	0.97	0.63	1.84	90.59

Table 5. Differences in the fish and epifauna (biomass) of sandbanks and off-bank (average dissimilarity = 65.2%). See Table 3 for further information on the data presented.

Species	Average abundance					
	Sandbank	Off-bank	Av.Diss	Diss/SD	Contrib%	Cum.%
<i>Echiichthys vipera</i>	14.61	1.54	7.61	2.87	11.68	11.68
<i>Echinocardium cordatum</i>	2.22	13.98	7.44	0.76	11.42	23.10
<i>Ophiura albida</i>	0.39	8.67	4.64	2.02	7.12	30.23
<i>Buglossidium luteum</i>	8.53	16.18	4.32	1.92	6.62	36.85
<i>Sabellaria spinulosa</i>	0.00	5.43	3.40	0.70	5.22	42.07
<i>Limanda limanda</i>	5.51	3.29	2.81	1.19	4.31	46.37
<i>Ophiura ophiura</i>	3.33	7.86	2.66	2.00	4.09	50.46
<i>Callionymus lyra</i>	0.82	5.32	2.66	2.12	4.09	54.55
<i>Asterias rubens</i>	0.78	4.81	2.58	1.10	3.96	58.50
<i>Solea solea</i>	0.41	4.71	2.53	0.85	3.88	62.38
<i>Liocarcinus holsatus</i>	0.75	4.73	2.29	1.53	3.52	65.90
Ammodytidae	0.70	3.88	2.15	0.81	3.29	69.19
<i>Pagurus bernhardus</i>	1.25	4.19	2.00	1.60	3.06	72.25
<i>Pleuronectes platessa</i>	2.28	2.23	1.98	0.77	3.04	75.30
<i>Pomatoschistus</i> sp.	0.98	3.89	1.68	1.89	2.58	77.88
<i>Raja montagui</i>	0.00	2.26	1.45	0.30	2.22	80.10

seawater for subsequent analysis. Each individual was then identified to the lowest taxonomic level and the total biomass (blotted wet weight, g) and abundance (number of individuals) of each taxon in a sample was recorded. Animals were only used in the analysis if the head was present.

Four of the five meiofaunal replicates collected at each station were processed. After washing the samples onto a 63 µm sieve, meiofauna was extracted with Ludox™ 40 (Somerville & Warwick, 1996). The extraction was repeated three times before the extracted material was evaporated slowly in anhydrous glycerol and mounted on slides for taxonomic identification and counting.

Data analysis

All data analysis was conducted using the suite of statistical measures available in PRIMER v. 6 (Clarke & Gorley, 2006). Cluster analysis and multi-dimensional scaling of abundance and biomass data were conducted for epifaunal catches from beam trawl (square root transformed) and infaunal samples from Day grab samples (fourth root transformed) and using the Bray–Curtis similarity index. The similarity of percentages procedure (SIMPER) was used to identify species that

discriminated between catches on the bank and at off-bank sites.

The following diversity metrics were calculated for both the infaunal and epifaunal (excluding colonial species) samples: total number of species (S), total number of individuals (N), Margalef's species richness (d), Pielou's evenness (J'), Shannon's index (H') and the Simpson index ($1-\lambda'$). For further description of these metrics, see Magurran (1988) and Clarke & Gorley (2006).

RESULTS

Description of study area

The water depths over the crests of the banks ranged from 14.5–21 m, and the off-bank habitats sampled ranged from 27–40 m deep. The proportion of gravel and silt–clay were generally very low, with the sediments on the crests and off-bank sites ranging from 97–99.9% sand (Table 1), of which the greatest fractions were retained on the 125–250 µm sieves. Such subtle differences in sediment composition are an important factor in structuring the sandbank communities (see Discussion).

Fish and epifauna

The sandbank crests were typified by small catches of relatively few species (Table 2), with the main species sampled including *Crangon crangon*, *Ophiura ophiura* and five species of fish: lesser weever, solenette *Buglossidium luteum*, scaldfish *Arnoglossus laterna*, dab and sand gobies *Pomatoschistus* spp.). In contrast, sites in deeper water off the sandbanks were more speciose and the assemblages in these habitats were clearly distinct (Figure 2). Species that were relatively abundant between sandbanks included some species that occurred on the sandbank crests, as well as *Ophiura albida*, various crustaceans (*Pagurus bernhardus*, *Liocarcinus holsatus* and *Crangon allmanni*) and other fish species (e.g. dragonet and sandeels). The main differences

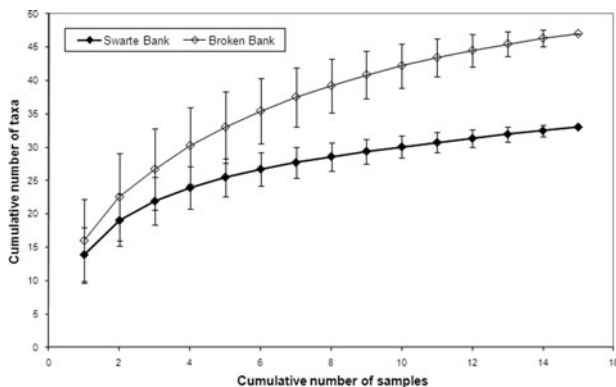
**Fig. 3.** Cumulative frequency of taxa caught in 2 m beam trawl sampling (mean ± SD) on and around the Swarte Bank and Broken Bank.

Table 6. Univariate indices (total number of species, S; total number of individuals, N; Margalef's species richness, d; Pielou's evenness, J'; Shannon's index, H'; and Simpson's index, 1-λ') for the epifauna of sandbank and off-bank habitats (mean ± standard deviation, range in parentheses).

Bank	Habitat	S	N	d	J'	H' (loge)	1-λ'
Broken Bank	Sandbank	11.78 ± 3.27 (8-18)	90.22 ± 30.72 (50-125)	2.42 ± 0.64 (1.46-3.52)	0.70 ± 0.08 (0.57-0.78)	1.69 ± 0.15 (1.53-1.94)	0.73 ± 0.06 (0.65-0.80)
	Off bank	22.50 ± 4.09 (17-29)	565.50 ± 388.43 (274-1276)	3.46 ± 0.40 (2.72-3.92)	0.47 ± 0.14 (0.25-0.61)	1.44 ± 0.36 (0.85-1.83)	0.57 ± 0.17 (0.31-0.75)
Swarte Bank	Sandbank	11.00 ± 1.73 (9-14)	92.33 ± 19.07 (59-121)	2.22 ± 0.35 (1.87-2.87)	0.70 ± 0.05 (0.64-0.78)	1.68 ± 0.14 (1.47-1.91)	0.74 ± 0.04 (0.66-0.79)
	Off bank	17.33 ± 3.14 (13-21)	230.50 ± 116.09 (83-351)	3.08 ± 0.46 (2.41-3.64)	0.68 ± 0.11 (0.56-0.84)	1.93 ± 0.27 (1.58-2.32)	0.78 ± 0.07 (0.69-0.88)

between bank and off-bank habitats were caused by high abundance of lesser weever and *C. crangon* on the crests of the banks, and larger numbers of species (especially *O. albida*, *O. ophiura*, *L. holsatus*, sand gobies and solenette) at off-bank sites (Table 3). In terms of biomass, these provided broadly similar results (Tables 4 & 5), although colonies of *Sabellaria spinulosa* were only recorded from off-bank habitats. Only one fish species (lumpsucker) that was recorded in the 4 m beam trawl was not observed in 2 m beam trawl catches. A taxonomic list of the fauna recorded from the sandbanks is provided in the Appendix.

Although the bank and off-bank assemblages were very similar on both the Swarte and Broken Banks, a larger number of species were observed on the Broken Bank (Figure 3; Table 6).

In terms of demersal fish, lesser weever was more abundant on the crests of the sandbanks (mean catch per unit effort = 39.3 ind.tow⁻¹) than at off-bank sites (0.6 ind.tow⁻¹). The length-frequency of lesser weever included a cohort of recently recruited fish (24-39 mm total length, L_T) as well as larger fish (ranging from 50-155 mm L_T), although the smaller-sized fish were only present from samples collected

on the crests of the banks (Figure 4). Catch rates of scaldfish were broadly similar in both habitats (3.9 and 2.8 ind.tow⁻¹ on the sandbank and off-bank habitats), with fish ranging from 46-62 mm and 91-147 mm. The cohort of smallest fish was proportionally more abundant on the crest (23.9% of the total number of scaldfish caught on the sandbank) than at off-bank sites (14.7% of total individuals). Solenette were caught in greater numbers at off-bank sites than on the sandbanks themselves (53.7 and 14.6 ind.tow⁻¹, respectively), although the overall length distribution (30-117 mm) was comparable in both habitats. Although most solenette ranged from 60-105 mm L_T, there was also a cohort of fish 30-50 mm. Sand gobies were also more common on off-bank sites (15.3 ind.tow⁻¹) than on the crests of the sandbanks (1.9 ind.tow⁻¹), with a more restricted length-range observed on the sandbank (39-56 mm) than from off-bank sites (32-72 mm).

There were also subtle differences in the size distributions of some epifaunal invertebrates (Figure 5), although sample sizes were generally small. Samples of *Crangon allmanni* from the sandbank contained proportionally more smaller-sized individuals in contrast to off-bank sites, whereas the

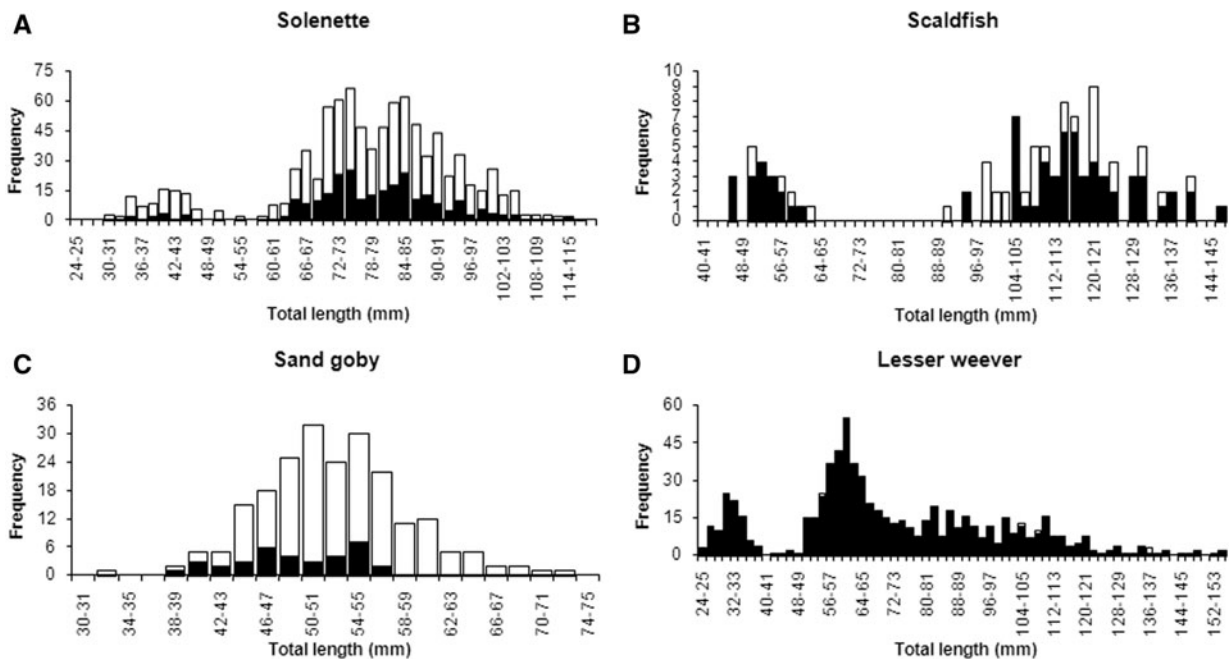


Fig. 4. Length-frequency distributions (by 2 mm length categories) for (A) solenette, (B) scaldfish, (C) sand goby and (D) lesser weever caught by 2 m beam trawl on the tops of sandbanks (black bars, 18 hauls) and from off-bank sites (white bars, 12 hauls).

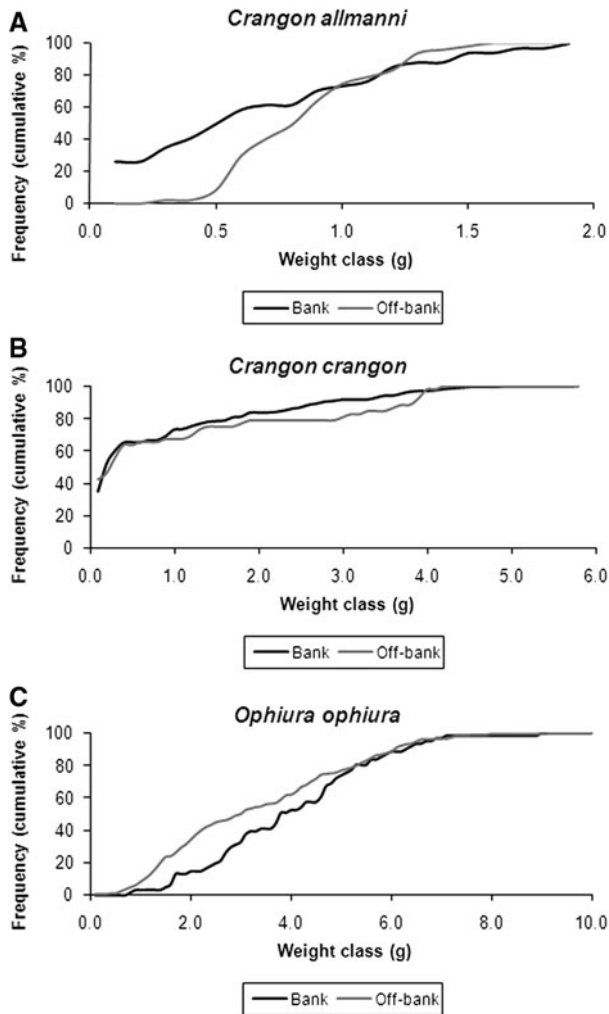


Fig. 5. Cumulative weight–frequency distributions for (A) *Crangon allmanni*, (B) *C. crangon*, and (C) *Ophiura ophiura* caught by 2 m beam trawl on the tops of sandbanks (black line, 18 hauls) and from off-bank sites (grey line, 12 hauls).

size distributions of *C. crangon* were similar in both habitats. The brittlestar *Ophiura ophiura* was more abundant at off-bank sites than on the crest, and proportionally more small individuals were found at off-bank sites.

Macrobenthic infauna

The sandbank crests were typified by low numbers of relatively few species (Table 7), with 10 species comprising more than 90% of the sampled fauna. With one exception (the brachyuran *Portunus latipes*), the dominant species were amphipods (*Bathyporeia elegans*, *B. guilliamsoniana* and *Urothoe brevicornis*), predatory polychaetes (*Nephtys cirrosa* and *Sthenelais limicola*) or deposit-feeding polychaetes (Spionidae and Magelonidae). In contrast, the assemblages at off-bank sites were clearly distinct (Figure 6; Table 8) and more speciose (particularly at the Broken Bank; see Table 9). Species that were relatively abundant at off-bank sites included those that occurred on the sandbank crests, as well as the deposit-feeding *Spiophanes bombyx*, two molluscs (*Euspira pulchellus* and *Tellina fabula*), sea potato (*Echinocardium cordatum*) and two further predatory polychaetes (*Nephtys hombergii* and *Goniada maculata*).

The main differences between bank and off-bank habitats were the reduced numbers of bivalves and echinoderms found on the crests of the banks, and an increased number of predatory species (e.g. *N. hombergii*, *Aglaophamus rubella*, *Glycera fallax*, *Anaitides* spp. and *Sigalion mathildae*) in off-bank habitats (Table 9). In terms of biomass, these provided broadly similar results to the abundance data. A list of species observed is included in the Appendix.

Meiofaunal nematodes

Two nematode feeding types dominated at all stations: non-selective deposit feeders and epigrowth feeders. Whilst the species diversity of nematode assemblages did not differ notably between both sandbanks, communities collected on the banks were more diverse than those recorded at the deeper off-bank sites. Heavily ornamented, small-sized species were abundant on the banks themselves, including *Xyala striata*, *Neochromadora trichophora*, *N. poecilosoma* and *Rhynchonema* sp. Off-bank locations were characterized by a high abundance of *Metadesmolaimus pandus*, *Microlaimus conothis*, *Sabatieria punctata* and *Paracanthonus platti*. Given the close association between nematodes and their sedimentary environments, further analyses of these data are given in Schratzberger *et al.* (in preparation).

DISCUSSION

Although offshore sandbanks are one of the habitats listed on Annex I of the EC Habitats Directive, there have been comparatively few studies examining the ecology of such habitats, especially in offshore areas (in this context, offshore refers to sites beyond 12 nautical miles from shore). Indeed, most previous studies on sandbanks in UK waters have been conducted in inshore areas (Holme, 1949; Withers & Thorp, 1978; Kaiser *et al.*, 2004), and the sandbank habitats studied previously in the North Sea have been either in coastal waters (Vanosmael *et al.*, 1982; Willems *et al.*, 1982a, b; Vanosmael & Heip, 1986; Vanaverbeke *et al.*, 2000, 2002; Dewicke *et al.*, 2003) or on the more extensive Dogger Bank (e.g. Kröncke & Knust, 1995; Kröncke & Wieking, 2003; Wieking & Kröncke, 2005), which is easier to sample.

Although there have been several general overviews of north-east Atlantic benthic communities (e.g. Jones, 1950; Glémarec, 1973), these have not differentiated the fauna of sandbanks from the more general sandy sediment communities on the inner continental shelf. The benthic community on the Dogger Bank has been described as a *Bathyporeia–Fabulina* (= *Tellina*) association (Wieking & Kröncke, 2005), and Tyler & Shackley (1980) considered sandbanks in the Bristol Channel to comprise a modified *Spisula* sub-community. Not only must those species that occur on sandbanks be able to adapt to local hydrodynamic conditions, but they are likely to also need a specialized trophic niche. Studies of the benthic macrofauna of the shallower parts of the Dogger Bank (18–32 m) have indicated that interface feeders (e.g. *Spiophanes bombyx*, *Magelona johnstoni*, *Tellina fabula* and *Amphiura brachiata*) and sand-licking amphipods (e.g. *Bathyporeia elegans*, *B. guilliamsoniana*, *B. tenuipes* and *Urothoe poseidonis*) are the predominant feeding guilds (Wieking & Kröncke, 2005), although other trophic guilds

Table 7. Infauna (individuals) captured by Day grab on the sandbank crests (average similarity = 59.61%) and at off-bank sites (average similarity = 47.52%). Data fourth root-transformed. See Table 2 for further information on the data presented.

Sandbank	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
<i>Bathyporeia elegans</i>	1.46	11.19	6.65	18.78	18.78
<i>Nephtys cirrosa</i>	1.33	10.6	6.43	17.78	36.56
<i>Ophelia borealis</i>	1.35	10	4.79	16.77	53.33
<i>Urothoe brevicornis</i>	1.31	9.99	4.59	16.75	70.08
<i>Scoloplos armiger</i>	0.65	3.9	1.22	6.54	76.62
<i>Spio armata</i> (agg)	0.45	1.96	0.69	3.29	79.91
<i>Magelona johnstoni</i>	0.52	1.93	0.67	3.23	83.14
<i>Portumnus latipes</i>	0.42	1.76	0.69	2.96	86.1
<i>Bathyporeia guilliamsoniana</i>	0.37	1.29	0.52	2.16	88.25
<i>Sthenelais limicola</i>	0.36	1.16	0.53	1.94	90.19
Off-bank	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
<i>Magelona johnstoni</i>	1.3	4.94	11.27	10.4	10.4
<i>Nephtys cirrosa</i>	1.11	4.87	5.52	10.25	20.65
<i>Ophelia borealis</i>	1.2	4.33	3.86	9.11	29.76
<i>Euspira pulchellus</i>	0.88	4.11	27.16	8.64	38.4
<i>Spiophanes bombyx</i>	0.87	3.87	4.88	8.15	46.56
Ophiuridae	0.77	3.45	10.07	7.26	53.81
<i>Bathyporeia guilliamsoniana</i>	1.01	2.75	0.9	5.78	59.59
<i>Scoloplos armiger</i>	0.8	2.4	0.89	5.04	64.63
<i>Tellina fabula</i>	1.04	2.3	0.91	4.84	69.47
<i>Echinocardium cordatum</i>	0.8	2.27	0.91	4.78	74.25
<i>Chaetozone christiei</i>	0.74	2.13	0.91	4.47	78.72
<i>Nephtys hombergii</i>	0.61	1.69	0.9	3.55	82.27
<i>Urothoe poseidonis</i>	0.69	1.69	0.9	3.55	85.83
<i>Goniada maculata</i>	0.55	1.66	0.91	3.5	89.33

may be represented (e.g. the predatory *Euspira pulchellus*). Some of these species were also found on the Swarte and Broken Banks, although the brittlestar *Amphiura brachiata* and other ophiurids were not observed in the present study. The macro-infauna occurring on the offshore North Norfolk sandbanks is generally comparable to that of the Dogger Bank and sandbanks in Belgian coastal waters (e.g. Moulart & Hostens, 2007), and many of the infaunal species observed are widely distributed in the southern North Sea (Rees *et al.*, 2007). The epifaunal species observed were similar to other sandy habitats occurring in the southern North Sea (e.g. Ellis & Rogers, 1999) and other parts of the British Isles, albeit with a reduced species diversity (Ellis *et al.*, 2007).

In recent years there have been several broadscale surveys of the epibenthos of the North Sea (Jennings *et al.*, 1999; Zühlke *et al.*, 2001; Callaway *et al.*, 2002), however these surveys only provided few samples (from sites 21–27 m deep) from the present study area (Figure 7). The range of species collected by these surveys were broadly similar to that found in the present study, although they included some additional species records (see Appendix). These broad scale surveys, while providing a general overview of the fauna that are found on the North Norfolk sandbanks, are not able to provide faunal descriptions of a suitable quality for site-specific analysis.

Samples from the 2 m beam trawl generally yielded fewer mobile species on the tops of sandbanks (8–18 on the Broken Bank, and 9–14 on the Swarte Bank). In contrast, catches from off-bank sites were more speciose (13–21 and 17–29 off the Swarte and Broken Banks, respectively), and had a larger number of individuals. Although the off-bank

sites had the richest and most diverse fauna, evenness was slightly greater on the top of the banks. Lesser weever was more abundant on the top of the sandbanks than at off-bank sites, and the length–frequency of lesser weever caught on the top of the sandbanks included a cohort of small fish 24–39 mm L_T , which was not observed at off-bank sites. Given that lesser weever leave the plankton at 13–15 mm (Russell, 1976), these sandbanks may serve as an important nursery ground for this species. Sandbank habitats may be an important habitat for lesser weever, as they bury into sandy sediments, and are ambush predators feeding on crangonids and other hyperbenthic crustaceans (Ellis, unpublished data). Other fish species for which early life history stages were observed on the sandbanks included solenette (30–50 mm L_T) and scaldfish (46–62 mm L_T), and these species leave the plankton at approximately 10 mm and 16–30 mm, respectively (Russell, 1976). Further studies on the use of sandbanks by early life-history stages are required, in order to determine whether there is increased recruitment to such habitats, with fish descending to deeper water as they increase in size. This would mimic the distribution pattern frequently displayed by many other juvenile fish species in coastal waters (Heinke's law), but there is little evidence that this can operate further offshore and over such a small scale.

In terms of biodiversity, sandbanks are often species-poor (e.g. Wilson, 1982; Kaiser *et al.*, 2004), although there is a greater diversity on the habitats and substrates surrounding the banks. This was evident in both the epifaunal and macro-infaunal assemblages sampled during the present study, although a contrasting pattern was observed with nematode communities, which were species-rich on the tops of the banks. Although species diversity of larger fauna was lower

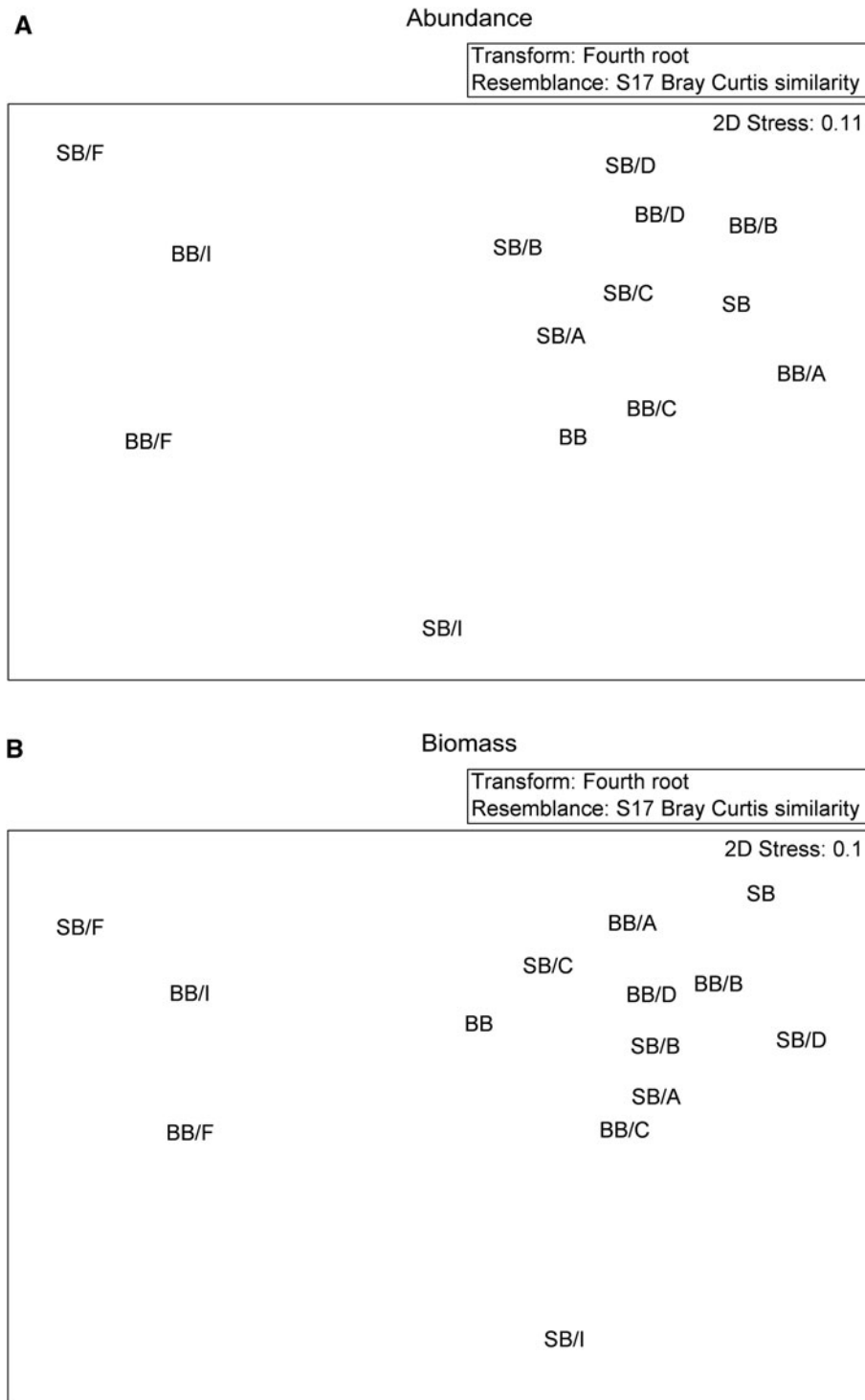


Fig. 6. Multidimensional scaling plots of infaunal catches by Day grab from seven stations at each of the Swarte and Broken Banks, including the main stations (SB and BB), other sites along the banks (suffixed A–D) and off-bank sites (suffixed F and I). The average dissimilarities between bank and off-bank samples were 60.89% by numbers (top) and 67.45% by biomass (bottom).

on the tops of sandbanks, certain epifaunal species (*C. crangon* and lesser weever) and macro-infaunal species (*Bathyporeia elegans* and *Urothoe brevicornis*) were more abundant on the banks than at off-bank sites. The increased diversity of macro-infauna and epifauna at off-bank sites may be attributable to an increased diversity of micro-habitats in such areas, both in terms of seafloor topography and sediment composition. For example, although the sediments at off-bank sites

still comprised primarily sand, there was an increased occurrence of both finer and coarser sediment types. It should be noted, however, that the distribution and relative abundance of meiofauna seemed to be influenced by subtle differences in sediment composition and this will be examined in more detail in a subsequent paper (Schratzberger *et al.*, in preparation). No targeted sampling was undertaken on the steeper slopes of the sandbanks, and future studies could usefully

Table 8. Differences in the macrobenthic infauna of sandbank crests and at off-bank sites (average dissimilarity = 60.89%). See Table 3 for further information on the data presented.

Species	Average abundance					
	Sandbank	Off-bank	Av.Diss	Diss/SD	Contrib%	Cum.%
<i>Tellina fabula</i>	0.08	1.04	2.99	1.33	4.91	4.91
<i>Bathyporeia elegans</i>	1.46	0.57	2.88	1.43	4.73	9.64
<i>Urothoe brevicornis</i>	1.31	0.47	2.55	1.67	4.19	13.83
<i>Magelona johnstoni</i>	0.52	1.3	2.52	1.41	4.14	17.97
<i>Bathyporeia guilliamsoniana</i>	0.37	1.01	2.49	1.72	4.09	22.06
<i>Echinocardium cordatum</i>	0.21	0.8	2.09	1.54	3.43	25.49
<i>Chaetozone christiei</i>	0.13	0.74	2.07	1.57	3.4	28.89
<i>Urothoe poseidonis</i>	0	0.69	2.03	1.56	3.34	32.23
<i>Nephtys hombergii</i>	0	0.61	1.79	1.65	2.95	35.18
<i>Euspira pulchellus</i>	0.35	0.88	1.7	1.4	2.8	37.98
<i>Spiophanes bombyx</i>	0.35	0.87	1.7	1.37	2.79	40.77
<i>Goniada maculata</i>	0.07	0.55	1.67	1.41	2.74	43.52
<i>Tellimya ferruginosa</i>	0.07	0.55	1.62	1.02	2.65	46.17
<i>Scoloplos armiger</i>	0.65	0.8	1.51	1.3	2.48	48.65
<i>Ophelia borealis</i>	1.35	1.2	1.51	1.63	2.47	51.12
<i>Aglaophamus rubella</i>	0	0.45	1.45	0.93	2.38	53.5
Ophiuridae	0.37	0.77	1.4	1.23	2.3	55.8
<i>Spio armata</i> (agg)	0.45	0	1.4	1.17	2.3	58.1
<i>Sthenelais limicola</i>	0.36	0.58	1.34	1.16	2.21	60.31
<i>Portumnus latipes</i>	0.42	0	1.28	1.18	2.1	62.41
<i>Ophiura albida</i>	0.07	0.37	1.19	0.99	1.95	64.36
<i>Nephtys caeca</i>	0.28	0.17	0.95	0.89	1.57	65.93
<i>Ophelia neglecta</i>	0.23	0.17	0.94	0.81	1.54	67.47
<i>Periculodes longimanus</i>	0.2	0.17	0.82	0.8	1.34	68.82
<i>Nephtys cirrosa</i>	1.33	1.11	0.73	1.46	1.21	70.02
<i>Ophiura ophiura</i>	0	0.24	0.72	0.57	1.18	71.2
Nemertea	0.07	0.17	0.69	0.64	1.13	72.33
<i>Spio decorata</i>	0	0.24	0.67	0.57	1.1	73.42
<i>Nephtys assimilis</i>	0	0.2	0.6	0.57	0.99	74.41
<i>Magelona filiformis</i>	0	0.2	0.6	0.57	0.99	75.4
<i>Donax vittatus</i>	0	0.2	0.6	0.57	0.99	76.39
Turbellaria	0.07	0.17	0.6	0.64	0.99	77.37
<i>Hesionura elongata</i>	0	0.17	0.6	0.57	0.98	78.35
<i>Glycera fallax</i>	0	0.17	0.6	0.57	0.98	79.33
<i>Aricidea minuta</i>	0	0.17	0.6	0.57	0.98	80.31

examine the faunal communities on the lee and stoss slopes of such habitats.

Kaiser *et al.* (2004) recently undertook both 2 m and 4 m beam trawling on sandbanks in Welsh waters. This study highlighted that both species diversity and the number of individuals was lower on distinct sandbanks than on those sandbank-like habitats that were considered to be extensions of inshore sandy habitats, and our results would support this view. Many of the species that we observed on the

Swarte and Broken Banks were reported to be important components of the epifaunal communities of sandbanks in Welsh waters (Kaiser *et al.*, 2004), including *Urothoe* sp., *Pagurus bernhardus*, *Liocarcinus holsatus*, *Asterias rubens*, *E. vipera*, *Ammodytes tobianus* and *Pomatoschistus minutus*. Species recorded by Kaiser *et al.* (2004) as being important on sandbanks that were not recorded in the present study included greater sand eel *Hyperoplus lanceolatus*, sand sole *Pegusa lascaris* and the shrimp *Philocheras trispinosus*. The absence of

Table 9. Univariate indices for the macrobenthic infauna of sandbank and off-bank habitats (mean ± standard deviation, range in parentheses). See Table 6 for further information on indices used.

Bank	Habitat	S	N	d	J'	H' (loge)	1-λ'
Broken Bank	Sandbank	12.8 ± 3.63 (10–19)	18.56 ± 3.89 (13–23)	4.03 ± 1.01 (3.29–5.74)	0.71 ± 0.04 (0.67–0.76)	1.79 ± 0.13 (1.68–2.0)	0.82 ± 0.03 (0.79–0.85)
	Off-bank	25.5 ± 0.71 (25–26)	20.5 ± 9.19 (14–27)	8.38 ± 1.55 (7.28–9.47)	0.89 ± 0.02 (0.88–0.90)	2.89 ± 0.08 (2.83–2.94)	0.98 ± 0.03 (0.96–1.0)
Swarte Bank	Sandbank	13.6 ± 4.04 (8–18)	18.72 ± 3.86 (15–25)	4.29 ± 1.21 (2.46–5.26)	0.70 ± 0.04 (0.67–0.77)	1.80 ± 0.17 (1.61–1.94)	0.80 ± 0.03 (0.77–0.84)
	Off-bank	19.5 ± 2.12 (18–21)	38.8 ± 18.67 (26–52)	5.15 ± 0.13 (5.06–5.24)	0.63 ± 0.01 (0.62–0.64)	1.87 ± 0.10 (1.80–1.94)	0.75 ± 0.05 (0.72–0.78)

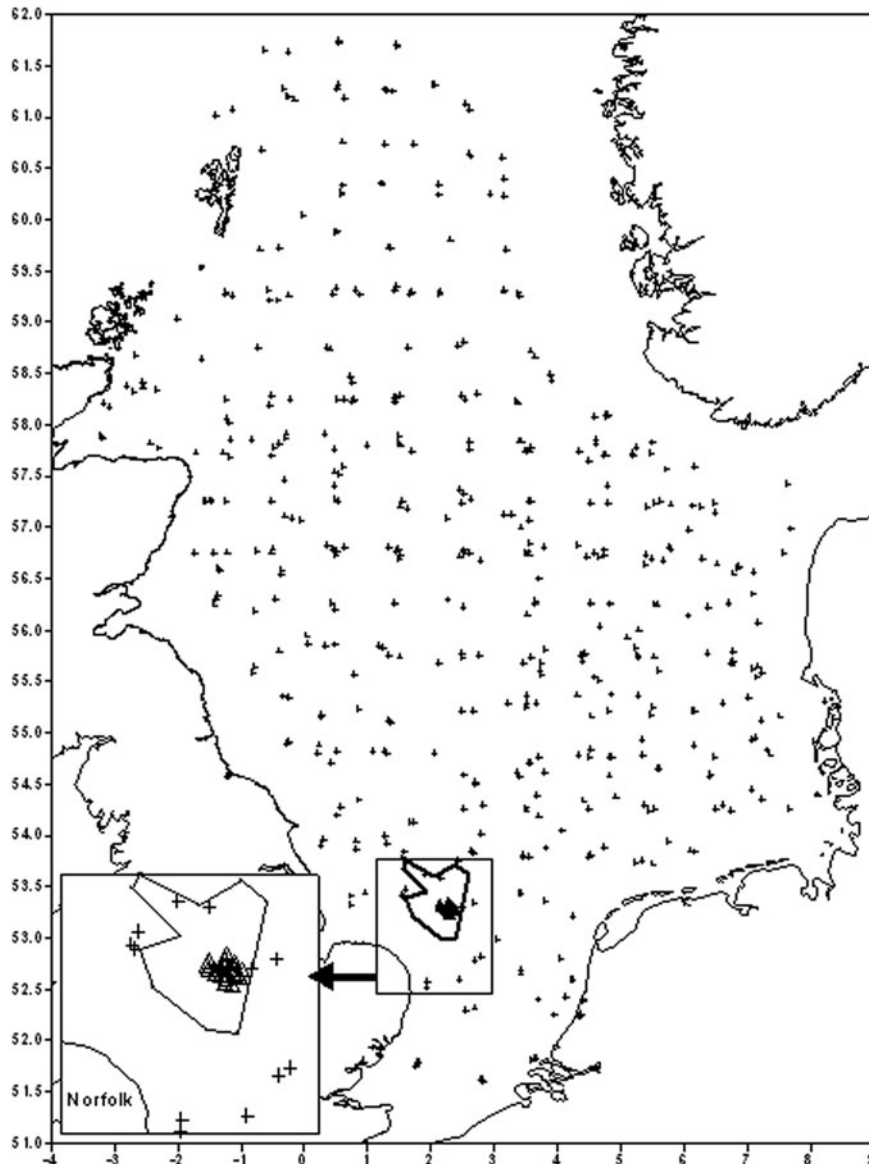


Fig. 7. Spatial coverage of 2 m beam trawl stations in the North Sea (+) indicating the low amount of sampling in the vicinity of the North Norfolk sandbanks and in the North Norfolk Sandbanks and Saturn Reef candidate SAC (inset), prior to the current sampling (Δ).

sand sole from our study is to be expected, as this species has a south-western distribution.

Many of the fish and benthic species observed on the sandbanks are widely distributed in other sandy habitats on the continental shelf, and the fauna of sandbank communities may simply be based on a specialized niche of the sand-associated fauna of the region. None of the taxa observed in the present study would seem to be obligate sandbank species and occur on other sandy habitats, as also reported in other regions (Kaiser *et al.*, 2004). However, certain taxa (e.g. *E. vipera*) may be locally abundant and potentially indicative of such habitats. The presence of ambush predators on the tops of sand ridges has also been observed in the north-west Atlantic, where species such as *Astroscopus guttatus* (Uranoscopidae) and *Trachinocephalus myops* (Synodontidae) are an important component of the sand ridge ichthyofauna (Vasslides & Able, 2008). The topographic features of sandbank habitats may create local hydrodynamic mechanisms that concentrate planktonic larvae (Ma *et al.*,

2006), which could explain both the importance of sandbanks habitats as settlement and nursery grounds, but also the abundance of those ambush predators that can retain their position in sites of potentially high velocity water movement by burying into the sediment.

Sandbanks are also topographically complex habitats, and further studies to examine the fine scale distribution and microhabitat use of certain species would clearly improve our understanding of the dynamics of the sandbank ecosystem. Indeed, certain species or life-history stages of fish may select specific microhabitats associated with sandy substrates (e.g. sand waves, ribbons, ripples and sand patches with emergent fauna), whether for shelter or trophic interactions (Auster *et al.*, 1995, 2003; Diaz *et al.*, 2003; Vasslides & Able, 2008).

Overall, 20 fish species were recorded during the survey, although more species are likely to occur in the area. The use of additional fish sampling techniques (e.g. longline and gillnet) is required to determine which larger piscivorous

fish forage around offshore sandbanks. Several large-bodied piscivorous fish species (e.g. cod, spurdog, tope, turbot and bass) are taken in the general area (see below) and further studies to examine whether sandbanks are important feeding grounds or topographic features for such species are required. Future sampling should also be undertaken at different tidal states, as the behaviour of both fish and invertebrates may vary depending on tidal state and currents (e.g. Medved & Marshall, 1983; Michalsen *et al.*, 1996; Jumars & Sato, 2008).

Commercial fishing by English vessels in the area of the North Norfolk sandbanks (ICES Rectangle 35F2) is relatively low (reported annual landings of 2–16 t for the period 2002–2007). The main species harvested from the general area include whelk *Buccinum undatum*, plaice, spurdog, edible crab and cod (Table 10), and although a variety of gears have been recorded (e.g. beam trawl, otter trawl, gillnet, pots and longline), the main commercial species in the area can be taken in static gears.

There is a lack of historical data on the fauna of these sandbanks, and so it is not possible to determine whether there have been any long-term temporal changes in the benthic communities, or how they have responded to human activities. Although early workers such as Davis (1925) collected samples in many parts of the southern North Sea, this study

had only a low number of samples from ICES Rectangle 35F2. As these samples were collected with a different gear and from east of the Swarte and Broken Banks (in waters of >27 m depth), the data are not comparable.

Macrobenthic infauna particularly polychaetes, act as an important food resource for commercially important flatfish such as plaice *Pleuronectes platessa* and sole *Solea solea* (e.g. Amezcua *et al.*, 2003). However, such fauna are themselves vulnerable to fishing, and trawling disturbance can affect the species composition, structure and production of infaunal communities (Jennings *et al.*, 2001). Larger benthic fauna (e.g. larger bodied bivalves, polychaetes and spatangoids) can suffer a higher mortality rate through crushing and capture than smaller bodied organisms. Analyses by Jennings *et al.* (2002) in the North Sea showed that production of small infauna or polychaetes was not significantly affected at trawling frequencies of 0.35–6.14 times yr⁻¹, although the biomass of larger infauna over similar trawling frequencies decreased by an order of magnitude, and production decreased 6-fold. Given the absence of historical data, it is not possible to identify whether the study site has been affected by human activities. In terms of the vulnerability of the sandbank fauna to fishing, most of the bivalves observed in the present study were small-bodied species (e.g. *Tellinomya*, *Abra* and *Tellina*), which may be less impacted by fishing disturbance, and few sessile filter-feeders (e.g. hydroids and bryozoans) were observed, possibly as a result of the turbidity and/or lack of appropriate substrate on which to settle. However, spatangoids (e.g. *Echinocardium cordatum*) and colonies of *Sabellaria spinulosa* were observed in the study area, the latter recorded exclusively from off-bank sites and such taxa are known to be susceptible to trawl disturbance (Bergman & van Santbrink, 2000). However, it should be recognized that many of the main commercial species in the area would be targeted with static gears, such as pots (e.g. whelk and edible crab) or longline (e.g. spurdog, cod and skates), and such gears are less damaging to the seafloor and will have a lower impact on the physical structure and integrity of sandbank habitats.

Table 10. Mean annual landings (tonnes) of commercial fish and shellfish from ICES Rectangle 35F2 by UK-registered fishing vessels (2002–2007).

Species	Mean (2002–2007)
Nephrops ⁽¹⁾	<i>Nephrops norvegicus</i> 5.855
Whelk	<i>Buccinum undatum</i> 3.244
Plaice	<i>Pleuronectes platessa</i> 1.921
Spurdog	<i>Squalus acanthias</i> 1.169
Edible crab	<i>Cancer pagurus</i> 1.023
Cod	<i>Gadus morhua</i> 0.885
Whiting	<i>Merlangius merlangus</i> 0.542
Skates and rays	Rajidae 0.517
Sole	<i>Solea solea</i> 0.497
Lemon sole	<i>Microstomus kitt</i> 0.312
Haddock	<i>Melanogrammus aeglefinus</i> 0.278
Dab	<i>Limanda limanda</i> 0.223
Tope	<i>Galeorhinus galeus</i> 0.182
Turbot	<i>Psetta maxima</i> 0.138
Red mullet	<i>Mullus surmuletus</i> 0.111
Flounder	<i>Platichthys flesus</i> 0.110
Gurnards	Triglidae 0.094
Brill	<i>Scophthalmus rhombus</i> 0.072
Other demersal fish ⁽²⁾	0.178
Crustaceans ⁽³⁾	0.096
Cephalopods ⁽⁴⁾	0.050
Other pelagic fish ⁽⁵⁾	0.031

⁽¹⁾Although there are reported landings of *Nephrops*, this species occurs on muddy substrates and are outside of the study area.

⁽²⁾Including halibut *Hippoglossus hippoglossus*, anglerfish *Lophius piscatorius*, witch *Glyptocephalus cynoglossus*, ling *Molva molva*, hake *Merluccius merluccius*, pout *Trisopterus* spp., bass *Dicentrarchus labrax* and John Dory *Zeus faber*.

⁽³⁾Including spider crab *Maja squinado*, and lobster *Homarus gammarus*.

⁽⁴⁾Including octopus, cuttlefish and squid.

⁽⁵⁾Including mackerel *Scomber scombrus*, scad *Trachurus trachurus* and grey mullet (Mugilidae).

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Appendix Taxonomic list of the marine fauna of the Broken and Swarte Banks (bank and off-bank habitats) taken during meiofaunal (M) and infaunal (G) sampling from 0.1 m² Day grab, 2 m beam trawl (T) and 4 m beam trawl (B). Species not observed in present study but recorded in prior 2 m beam trawl surveys (see Callaway *et al.*, 2002) are denoted T*.

Species	Gear
CNIDARIA	
Hydrozoa	
<i>Tubularia</i> sp.	T/G
<i>Coryne</i> sp.	G
<i>Dicoryne</i> sp.	T*
Bougainvilliidae	G
<i>Hydractinia echinata</i>	T
<i>Calycella syringa</i>	G
<i>Abietinaria abietina</i>	T*
<i>Hydrallmania falcata</i>	T
<i>Sertularia cupressina</i>	T
<i>Sertularia</i> spp.	G
<i>Nemertesia antennina</i>	T
<i>Nemertesia ramosa</i>	T*
<i>Campanularia volubilis</i>	T*
Campanulariidae	G
Hydroid indet.	B/T

Continued

Appendix Continued

Species	Gear
Anthozoa	
<i>Alcyonium digitatum</i>	T
Actiniaria indet.	B/T
PLATHELMINTHES	
<i>Turbellaria</i> indet.	G
NEMERTEA	
<i>Nemertea</i> indet.	G
NEMATODA	
Enoplida	
<i>Enoploides brunetti</i>	M
<i>Enoplolaimus</i> sp.	M
<i>Epacanthion</i> sp.	M
<i>Mesacanthion diplochma</i>	M
<i>Mesacanthion hirsutum</i>	M
<i>Chaetonema</i> sp.	M
<i>Anticoma acuminata</i>	M
<i>Anticoma eberthi</i>	M
<i>Dolicholaimus</i> sp.	M
<i>Thalassoalaimus</i> sp.	M
<i>Platycoma</i> sp.	M
<i>Halalaimus gracilis</i>	M
<i>Oncholaimellus</i> sp.	M
<i>Oncholaimus skawensis</i>	M
<i>Viscosia abyssorum</i>	M
<i>Viscosia cobbi</i>	M
<i>Viscosia elegans</i>	M
<i>Viscosia glabra</i>	M
<i>Belbolla</i> sp.	M
<i>Eurystomina</i> sp.	M
<i>Bathylaimus capacosus</i>	M
<i>Bathylaimus paralongisetosus</i>	M
<i>Bathylaimus tenuicaudatus</i>	M
<i>Gairleanema</i> sp.	M
<i>Tripylloides</i> sp.	M
<i>Rhabdodemia</i> sp.	M
Trefusiida	
<i>Rhabdocoma riemanni</i>	M
Chromadorida	
<i>Chromadorella duopapillata</i>	M
<i>Prochromadorella septempapillata</i>	M
<i>Prochromadorella</i> sp.	M
<i>Rhips</i> sp.	M
<i>Chromadorita tentabunda</i>	M
<i>Dichromadora cucullata</i>	M
<i>Neochromadora poecilosoma</i>	M
<i>Neochromadora trichophora</i>	M
<i>Spilophorella paradoxa</i>	M
<i>Sabatieria celtica</i>	M
<i>Sabatieria punctata</i>	M
<i>Vasostoma</i> sp.	M
<i>Comesa</i> sp.	M
<i>Trichethmolaimus hirsutus</i>	M
<i>Pomponema sedecima</i>	M
<i>Nannolaimoides effiliatus</i>	M
<i>Paracanthonchus longicaudatus</i>	M
<i>Paracanthonchus platti</i>	M
<i>Paracyatholaimus pentodon</i>	M
<i>Paracyatholaimoides multispiralis</i>	M
<i>Choniolaimus papillatus</i>	M
<i>Gammanema</i> sp.	M
<i>Synonchiella riemanni</i>	M
<i>Richtersia</i> sp.	M
<i>Desmodora schulzi</i>	M
<i>Desmodora tenuispiculum</i>	M

Continued

Appendix Continued

Species	Gear
<i>Parallelocoilas</i> sp.	M
<i>Spirinia parasitifera</i>	M
<i>Chromaspirina parapontica</i>	M
<i>Sigmophoranema litorale</i>	M
<i>Pseudonchus</i> sp.	M
<i>Leptonemella aphanothecae</i>	M
Desmodoridae (indet.)	M
<i>Calomicrolaimus</i> sp.	M
<i>Microlaimus conothelis</i>	M
<i>Microlaimus monstrosus</i>	M
<i>Microlaimus ostracion</i>	M
<i>Microlaimus</i> sp. 1	M
<i>Nudora bipapillata</i>	M
<i>Alaimella</i> sp.	M
<i>Leptolaimodes</i> sp.	M
<i>Leptolaimus</i> sp.	M
<i>Stephanolaimus elegans</i>	M
<i>Stephanolaimus yayasreei</i>	M
<i>Camacolaimus</i> sp.	M
<i>Haliplectus</i> sp.	M
<i>Tarvaia angusta</i>	M
<i>Aegialolaimus elegans</i>	M
<i>Cyartonema elegans</i>	M
<i>Southernia</i> sp.	M
<i>Ceramonema yunfengi</i>	M
<i>Dasyemoides albaensis</i>	M
<i>Metadasyemoides</i> sp.	M
<i>Pterygonema</i> sp.	M
<i>Tricoma brevirostris</i>	M
Monhysterida	
<i>Daptonema hirsutum</i>	M
<i>Daptonema invagiferoum</i>	M
<i>Daptonema normandicum</i>	M
<i>Daptonema oxycera</i>	M
<i>Daptonema</i> sp. 1	M
<i>Theristus acer</i>	M
<i>Theristus denticulatus</i>	M
<i>Theristus interstitialis</i>	M
<i>Tricotheristus mirabilis</i>	M
<i>Metadesmolaimus pandus</i>	M
<i>Metadesmolaimus</i> sp.	M
<i>Daptonemna fallax</i>	M
<i>Theristus</i> sp.	M
<i>Paramonohystera</i> sp.	M
<i>Gonionchus cumbraensis</i>	M
<i>Gonionchus longicaudatus</i>	M
<i>Rhynchonema</i> sp.	M
<i>Xyala striata</i>	M
<i>Siphonolaimus</i> sp.	M
<i>Desmolaimus zeelandicus</i>	M
<i>Disconema</i> sp.	M
<i>Eleutherolaimus</i> sp.	M
<i>Eumorpholaimus</i> sp.	M
<i>Linhomoeus</i> sp.	M
<i>Metalinhomoeus filiformis</i>	M
<i>Terschellingia communis</i>	M
<i>Axonolaimus helgolandicus</i>	M
<i>Axonolaimus hexapilus</i>	M
<i>Axonolaimus orcombensis</i>	M
<i>Ascolaimus</i> sp.	M
<i>Odontophora</i> sp.	M
<i>Odontophoroides</i> sp.	M
<i>Campylaimus</i> sp.	M
<i>Diplopeltula</i> sp.	M

Continued

Appendix Continued

Species	Gear
<i>Coninckia macrospirifera</i>	M
Secernentea	
<i>Rhabditis</i> sp.	M
ANNELIDA	
Polychaeta	
<i>Harmothoe impar</i>	G
<i>Lepidonotus squamatus</i>	G
<i>Sigalion mathildae</i>	G
<i>Sthenelais limicola</i>	G
<i>Hesionura elongata</i>	G
<i>Hypereteone foliosa</i>	G
<i>Anaitides lineata</i>	G
<i>Anaitides rosea</i>	G
<i>Glycera fallax</i>	G
<i>Glycera lapidum</i> (agg)	G
<i>Goniada maculata</i>	G
<i>Goniada maculata</i> (epitoke)	G
<i>Podarkeopsis capensis</i>	G
<i>Aglaophamus rubella</i>	G
<i>Nephtys assimilis</i>	G
<i>Nephtys caeca</i>	G
<i>Nephtys cirrosa</i>	G
<i>Nephtys hombergii</i>	G
<i>Nephtys longosetosa</i>	G
Nephtyidae (indet.)	T/G
<i>Scoloplos armiger</i>	G
<i>Aricidea minuta</i>	G
<i>Paraonis fulgens</i>	G
<i>Poecilochaetus serpens</i>	G
Spionidae (larva)	G
<i>Scolelepis bonnieri</i>	G
<i>Spio armata</i> (agg)	G
<i>Spio decorata</i>	G
<i>Spiophanes bombyx</i>	G
<i>Magelona filiformis</i>	G
<i>Magelona johnstoni</i>	G
<i>Chaetozone christiei</i>	G
<i>Ophelia bicornis</i>	G
<i>Ophelia borealis</i>	G
<i>Ophelia neglecta</i>	G
<i>Ophelia</i> sp.	T/G
<i>Ophelina acuminata</i>	G
<i>Sabellaria spinulosa</i>	B/T
CRUSTACEA	
Cirripedia	
<i>Balanus balanus</i>	T*
Amphipoda	
<i>Perioculodes longimanus</i>	G
<i>Pontocrates altamarinus</i>	G
<i>Urothoe brevicornis</i>	G
<i>Urothoe poseidonis</i>	G
<i>Orchomenella nana</i>	G
<i>Scopelocheirus hopei</i>	G
<i>Atylus falcatus</i>	G
<i>Atylus swammerdamei</i>	G
<i>Bathyporeia elegans</i>	G
<i>Bathyporeia guilliamsoniana</i>	G
<i>Dyopodos monacanthus</i>	G
<i>Iphinoe trispinosa</i>	G
Cumacea	
<i>Diastylis bradyi</i>	G
Isopoda	
<i>Idotea linearis</i>	T
Decapoda (Natantia)	

Continued

Appendix Continued

Species	Gear
<i>Processa</i> sp.	T
<i>Pandalina brevirostris</i>	T
<i>Pandalus montagui</i>	B/T
<i>Crangon allmani</i>	B/T/G
<i>Crangon crangon</i>	B/T
<i>Philocheras trispinosus</i>	G
Decapoda (Anomura)	
<i>Anapagurus laevis</i>	T
<i>Pagurus bernhardus</i>	B/T
<i>Pisidia longicornis</i>	T
Decapoda (Brachyura)	
<i>Ebalia cranchii</i>	T
<i>Macropodia rostrata</i>	T
<i>Corystes cassivelaunus</i>	T
<i>Liocarcinus depurator</i>	B/T
<i>Liocarcinus holsatus</i>	B/T
<i>Portumnus latipes</i>	T/G
MOLLUSCA	
Gastropoda	
<i>Euspira pulchella</i>	T/G
Opisthobranchia	
<i>Acanthodoris pilosa</i>	T*
Bivalvia	
<i>Tellinomya ferruginosa</i>	G
<i>Spisula solida</i>	B/G
<i>Spisula subtruncata</i>	T
<i>Phaxas pellucidus</i>	T
<i>Tellina fabula</i>	G
<i>Donax vittatus</i>	T/G
<i>Abra alba</i>	T/G
<i>Abra prismatica</i>	G
<i>Chamelea gallina</i>	T
<i>Dosinia lupinus</i>	T
<i>Mya truncata</i>	T
Cephalopoda	
<i>Sepiola atlantica</i>	T
<i>Loligo forbesi</i>	T*
BRYOZOA	
<i>Alcyonidium diaphanum</i>	B/T
<i>Alcyonidium</i> sp.	G
<i>Alcyonidium parasiticum</i>	T/G
<i>Vesicularia spinosa</i>	G
<i>Eucreatea loricata</i>	T
<i>Electra pilosa</i>	G
<i>Flustra foliacea</i>	B/T

Continued

Appendix Continued

Species	Gear
<i>Scrupocellaria reptans</i>	T*
<i>Scrupocellaria</i> sp.	T
<i>Celleporella hyalina</i>	G
ECHINODERMATA	
<i>Astropecten irregularis</i>	T*
<i>Asterias rubens</i>	B/T
<i>Crossaster papposus</i>	T*
<i>Ophiura affinis</i>	T
<i>Ophiura albida</i>	B/T/G
<i>Ophiura ophiura</i>	B/T/G
<i>Ophiocten affinis</i>	G
<i>Psammechinus miliaris</i>	T*
<i>Echinocardium cordatum</i>	B/T/G
ELASMOBRANCHII	
<i>Raja montagui</i>	B/T
TELEOSTEI	
<i>Enchelyopus cimbrius</i>	T
<i>Merlangius merlangus</i>	B/T
<i>Entelurus aequoreus</i>	T
<i>Syngnathus acus</i>	T
<i>Eutrigla gurnardus</i>	B/T
<i>Myoxocephalus scorpius</i>	B/T
<i>Agonus cataphractus</i>	B/T
<i>Echiichthys vipera</i>	B/T/G
<i>Cyclopterus lumpus</i>	B
<i>Ammodytes marinus</i>	T*
<i>Ammodytes tobianus</i>	T
<i>Gymnammodytes semisquamatus</i>	T/G
Ammodytidae	B
<i>Callionymus lyra</i>	B/T
<i>Callionymus reticulatus</i>	T
<i>Pomatoschistus</i> sp.	B/T
<i>Lepidorhombus whiffiagonis</i>	T*
<i>Arnoglossus laterna</i>	B/T
<i>Limanda limanda</i>	B/T
<i>Pleuronectes platessa</i>	B/T
<i>Buglossidium luteum</i>	B/T/G
<i>Solea solea</i>	B/T

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