Impact of an exotic parasitoid on *Plutella xylostella* (Lepidoptera: Plutellidae) population dynamics, damage and indigenous natural enemies in Kenya

B. Löhr¹*, R. Gathu¹, C. Kariuki², J. Obiero³ and G. Gichini¹

¹International Centre of Insect Physiology and Ecology, PO Box 30772, 00100 Nairobi, Kenya: ²Kenya Agricultural Research Institute, Katumani Research Centre, PO Box 340, 90700 Machakos, Kenya: ³Kenya Agricultural Research Institute, Muguga Research Centre, PO Box 30148, 00100 Nairobi, Kenya

Abstract

Diadegma semiclausum (Hellén) (Hymenoptera: lchneumonidae), an exotic diamondback moth parasitoid, was released in two pilot areas (Werugha in Coast Region and Tharuni in Central Province) in Kenya. Fifteen month before release, observations on the diamondback moth, Plutella xylostella (Linnaeus), and local natural enemy population dynamics and pest damage were initiated in both areas and continued for three years after release. The P. xylostella population was bimodal with higher records during dry seasons. At Werugha, the peak population of P. xylostella was 16.8 per plant (October 2001); at Tharuni it was 12.8 (February 2002). Populations at Werugha declined from three months after release and decreased from 5.4 per plant (before release) to 0.8 (year 3 after release). Concurrently, average damage (1.9 to 1.5) (on a 0–5 scale), proportion of attacked plants (72 to 31%) and proportion of plants in damage group >2 (plants with head damage) decreased (21.4 to 5.3%), while total parasitism increased from 14.4 (before) to 52.5% (year 3 after release, 90% due to D. semiclausum). At Tharuni, D. semiclausum was only recovered 3 months after release. Average populations of *P. xylostella* declined from 5.9 per plant (before release) to 2.4 (year 3 after release) and damage scores from 2.3 to 1.7. The proportion of plants in damage group >2 declined from 39.7 to 4.5% while overall parasitism increased from 4.2 to 40.6%(98.3% by D. semiclausum). Four species of indigenous parasitoids (Diadegma mollipla (Holmgren), Oomyzus sokolowskii (Kurdjumov), Apanteles sp. and Itoplectis sp., all primary parasitoids) were almost completely displaced by *D. semiclausum*. Possible reasons for the different parasitoid development between the two release areas and the displacement of the indigenous species are discussed.

Keywords: *Plutella xylostella,* biological control, *Diadegma semiclausum,* indigenous parasitoids, biocontrol impact, parasitoid displacement

*Fax: +254 20 8632001 E-mail: blohr@icipe.org

Introduction

Diadegma semiclausum (Hellén) (Hymenoptera: Ichneumonidae) is arguably the most successful parasitoid of the diamondback moth, Plutella xylostella (Linnaeus) (Lepidoptera: Plutellidae). It has been widely and successfully used over large areas of southeast and south Asia when pesticide resistance of P. xylostella had lead to the failure of crop protection programmes in crucifer production (Lim et al., 1996; Shelton et al., 1996; Uk & Harris, 1996; Verkerk & Wright, 1996). The first importation of the species was from England to New Zealand (Hardy, 1938) where it afforded partial control of the pest. From New Zealand, D. semiclausum was shipped to Sumatra in 1946 and released in the highland cabbage production areas. However, the releases did not result in noticeable improvement of the situation until broad-spectrum pesticides were replaced with Bt-based biopesticides that were harmless to the parasitoid (Sastrosiswojo & Sastrodiharjo, 1986). In the late 1980s and early 1990s, the parasitoid found its way to other countries of Southeast Asia (Ooi, 1992; Poelking, 1992; Talekar et al., 1992; Biever, 1996; Eusebio & Morallo-Rejesus, 1996) and Japan (Iga, 1997). In most situations, especially when the introductions were combined with a change to biocontrol-compatible pesticide use for the control of other cabbage pests, the parasitoid greatly contributed to reduced pesticide applications (Sastrosiswojo & Sastrodiharjo, 1986; Talekar et al., 1992; Biever, 1996).

In spite of this long-lasting effort to improve diamondback moth management through biological means, longterm studies of the impact of the parasitoid are scarce. In most biological control projects cited above, very little or no information was available about the existence and role of indigenous natural enemies or, more important in times of discussions about the importance of biodiversity, the fate of indigenous natural enemies after introduction and release of a supposedly superior competitor. Detailed studies on the parasitoids and diseases affecting P. xylostella were only conducted in countries where introduction biocontrol was never implemented, such as Canada (Harcourt et al., 1955); and the USA (Oatman & Platner, 1969; Hamilton, 1979; Ru & Workman, 1979). The only exception is South Africa, where Ullyett (1947) and Kfir (2003) have provided detailed information about occurrence and role of indigenous natural enemies. It was also in South Africa where the first introduction and release of D. semiclausum (as Angitia cerophaga Gravenhorst) in Africa took place in 1936 (Evans, 1939). The parasitoid was recovered in 1937 and assumed established by Greathead (1971). However, it was never again recovered and it is doubtful whether the parasitoid is established.

Crucifers are not as important and diverse vegetable crops in East Africa as they are in Southeast Asia. This may have been the major reason why the experiences in Asia took so long to be applied in Africa. Nevertheless, crucifers do play a major role in East Africa, particularly in Ethiopia (Ayalew *et al.*, 2004) and Kenya (Macharia *et al.*, 2005). In Kenya, kale is considered a valuable relish in many homesteads, providing necessary dietary vitamins and minerals in a maize-based diet. The crop is a source of cash for farmers in the rural and peri-urban areas, thus alleviating poverty and creating employment. Favourable weather conditions or availability of irrigation water allow for yearround production and in the highlands, farmers can grow at least three crops in a year. However, continuous production under poor crop management practice creates an environment for pest build-up including P. xylostella, which is the most economically important pest of crucifers in Kenya (Kibata, 1996; Oduor et al., 1996). Currently, synthetic pesticides are the predominant means of combating vegetable pests and diseases. Due to increasing pest pressure, farmers resorted to increased pesticide dosage and frequency of applications or application of cocktails, which has led to the development of widespread pesticide resistance (Kibata, 1996). In order to overcome this situation, the International Centre of Insect Physiology and Ecology, in collaboration with the Kenya Agricultural Research Institute and the Asian Vegetable Research and Development Centre, initiated a biological control programme based on importation and release of exotic parasitoids. In this paper, data are presented on temporal dynamics of P. xylostella and parasitoids for 15 months before and three years after the release of D. semiclausum in two pilot release areas in Kenya.

Materials and methods

Site description

Two areas in the highland cabbage growing zone of Kenya were assigned by the Kenya Plant Health Inspectorate Service for pilot releases of *D. semiclausum*: Wundanyi Division in Coast Province and Limuru Division in Central Province. Data presented here were collected at the release sites in both areas, Werugha Location (03°26′16″ S; 38°20′24″ E) of Wundanyi Division in Taita Taveta District, Coast Province; and Tharuni Location (01°08′12″ S; 36°37′51″ E) of Limuru Division, Kiambu District, Central Province of Kenya. A detailed description of the pilot release areas is provided in Momanyi *et al.* (2006). The studies were started from April and June 2001 (Werugha and Tharuni, respectively) and carried through for 15 months before and three years after release.

Data collection

Sampling started in April 2001 at Werugha and, until July 2003, fortnightly samplings were conducted, which were changed to once every four weeks from August 2003 until the end of observations in July 2005. At Tharuni, fortnightly sampling was conducted from July 2001 to September 2003 followed by once every four weeks until September 2005. On 26 July 2002, 25 pairs of *D. semiclausum* were released in each of five fields at Werugha and the same number was released on 20 September 2002 at Tharuni.

The aim was to evaluate 15 farmer-managed farms in both areas at each sampling date. This was possible without problems at Werugha, where farmers used bucket irrigation and moved production from the terraces into valley bottoms in the dry season. At Tharuni, a place without access to irrigation water, the number of cabbage fields declined so much during the height of the long dry season, that in some occasions only six fields could be sampled. Fields were selected at random with the help of the local extension officer in each area. A field was eligible for sampling from two weeks after transplanting onwards, and the same field was visited until it was harvested. When a field had been harvested, a recently transplanted field in the immediate vicinity was chosen as replacement. Crop type and age, field management, pesticide applications and general conditions

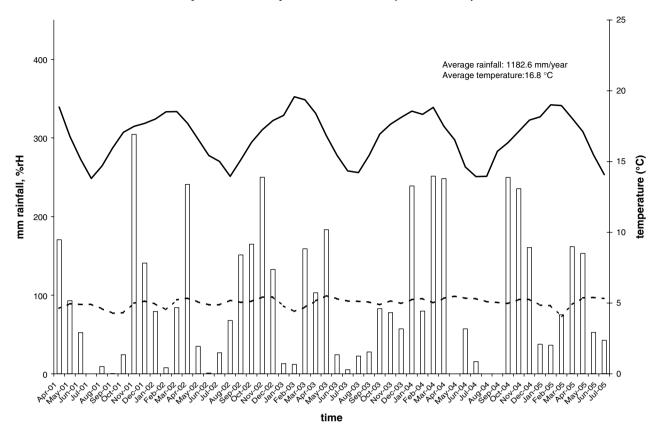


Fig. 1. Average monthly rainfall, relative humidity and temperature at Werugha, Wundanyi Division, Coast Province of Kenya. Averages were calculated for the three full years (2002–2004) of operations. (\Box , rainfall (mm/month); - - , % relative humidity; --, temperature (°C))

of the field were recorded. Ten plants were selected at random in each field and thoroughly checked. The number of small larvae, large larvae, pupae and adults of P. xylostella was counted and recorded separately. Other pests found on individual plants were also recorded. The damage caused by diamondback moth was estimated using a damage score of 0-5 (see Momanyi et al., 2006). Up to a maximum of five P. xylostella larvae (third instar or older) or pupae were collected from each plant and put in individual vials for further investigations in the laboratory. The first five encountered in these two stages were collected, which largely excludes bias, as the plants were systematically searched from the outer to the inner leaves. Larvae were retained singly on a fresh cabbage leaf in labelled 30 ml plastic vials at ambient temperatures of $21\pm2^{\circ}C$ and checked daily until emergence of adult moths or parasitoids. Emerged parasitoids were identified, sexed and counted. Parasitism was calculated as the number of parasitized larvae/pupae divided by the total number collected. A data logger (Hobo Pro Series, Onset Computer Corp. Pocasset, Massachusetts, USA) was used to record temperatures and relative humidity (hourly records), while rainfall records were obtained from the Kenya Meteorological Services.

Data analysis

The impact of the parasitoid on *P. xylostella* numbers was evaluated using three different methods: firstly, overall

average *P. xylostella* population of the year before release was compared to each year after release; secondly, in recognition of the influence of rainfall on *P. xylostella* populations, the before and after release years were also compared using the five months of highest and lowest population density to determine effects on the *P. xylostella* peaks and on the base population during unfavourable conditions. For graphic presentation, the data of population dynamics and parasitism data collected within the same month during the first 27 months of observation were pooled.

The changes in P. xylostella damage were compared using several approaches: firstly, whole year average damage score data were compared. In addition, the proportion of attacked plants and, finally, the percentage plants with a damage score >2 (where the market value is reduced) was calculated and compared for the years before and after release. Changes in parasitism were analysed comparing the before release overall parasitism rate to each year after release. One-way analysis of variance (ANOVA) was used to compare all these parameters; individual sampling dates were treated as repeated measures. Before calculations, the datasets were checked for normal distribution, which was fulfilled for all count data, hence those were not transformed. Where proportional data were used, raw data were arcsine transformed. However, all data presented in the tables are nontransformed values. Whenever ANOVA resulted in a significant F-value, means were separated using Tukey's honest significant difference (HSD) test at 5% level of

B. Löhr et al.

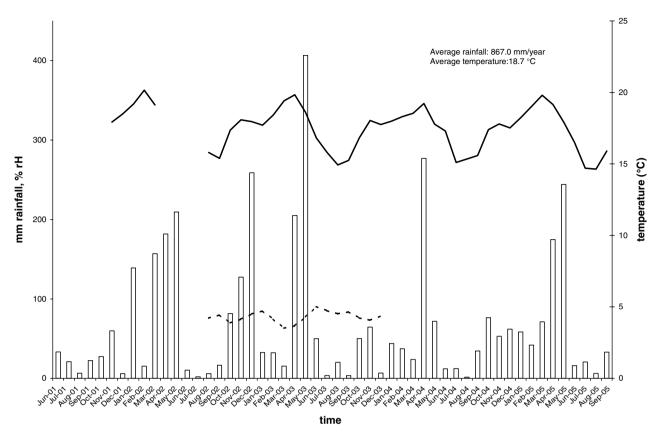


Fig. 2. Average monthly rainfall, relative humidity and temperature at Tharuni, Limuru Division, Central Province of Kenya. Averages were calculated for the three full years (2002–2004) of operations. (\Box , rainfall (mm/month); - - , % relative humidity; --, temperature (°C))

significance. Statistical software SAS version 9.1 (SAS Institute Inc., 2002) was used for all the analyses. For both release areas, linear regressions (PROC REG, SAS Institute, 2003) were calculated between *P. xylostella* population, parasitism by *D. semiclausum* and total parasitism and the progressing sampling dates starting from the date of release. A regression was also calculated between the total parasitism and parasitism by the introduced species. Original field data were used for all regression calculations, and the slopes of the regression lines were compared to determine differences (at *P* < 0.05).

The parasitoid guild composition was compared before and after release using the Shannon diversity index (Mugurran, 1988). The diversity index obtained was subjected to a t-test to compare differences before and after release following the procedure described by Batten (1976).

Results

Rainfall, temperature and relative humidity in the release areas

Werugha received relatively regular rainfall (1182 mm annually during the study period, fig. 1) and complementary irrigation is easily available; while at Tharuni, rainfall was more erratic (fig. 2), the total amount was considerably lower (867 mm) and complementary irrigation is not available. The average monthly temperature was 2°C higher at Tharuni while relative humidity seemed to be considerably lower. Unfortunately, the data logger was faulty and most of the data records for relative humidity were corrupted. With these conditions, the two areas represent two extremes for cabbage production in Kenya.

Diamondback moth population and damage before release

Werugha

Diamondback moth dynamics was bimodal with peaks during the dry seasons (fig. 3). Higher counts were recorded during the months of September to November and again January to March, reflecting the rainfall pattern (fig. 1). The highest mean number of *P. xylostella* per plant in any of the collections was reached during the pre-release year on 16 October 2001 (16.8 per plant, not shown because of pooling of data), in spite of bi-weekly insecticide applications by most farmers. The build-up of the second peak started in December 2001, but heavy rains in late December and early January disrupted the population build-up (fig. 3). From May 2002, the population rose again until the release was made in August. The after-release population curve indicates a considerable reduction in numbers of P. xylostella starting already from three months after release (fig. 3). The population curve showed a steadily declining trend for the three after-release years with the bimodal population dynamics still clearly visible but greatly decreased amplitude (fig. 3).

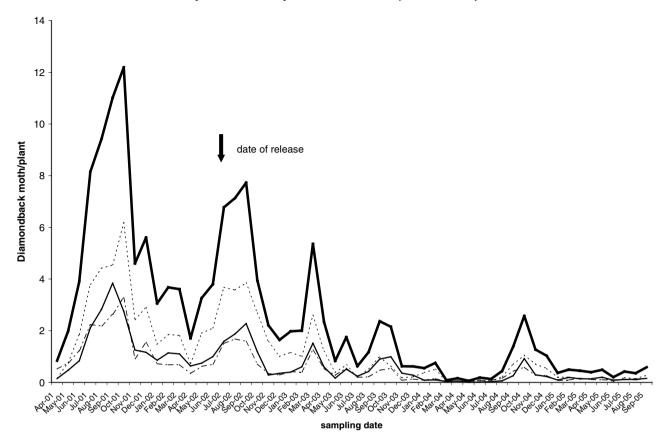


Fig. 3. Dynamics and composition of diamondback moth population before and after release of an exotic parasitoid, Werugha, Wundanyi Division, Coast Province of Kenya. Data were collected from ten plants in each of 15 farmer-managed fields. Sampling dates within one month were pooled for ease of presentation. (- - -, small larvae; ---, big larvae; ---, total)

Table 1. Diamondback moth (DBM) population and damage before and after introduction of the exotic parasitoid, *Diadegma semiclausum*, Werugha, Coast Province of Kenya.

Sampling Period	Observations	DBM per plant	Damage score	% plants attacked	% plants with damage score >2
Before release After release	486	$5.4 \pm 0.27a$	$1.9\pm0.03b$	72.3±1.31a	$21.4 \pm 1.21b$
Year 1 Year 2 Year 3	340 195 194	$\begin{array}{c} 3.6 \pm 0.24 b \\ 0.7 \pm 0.09 c \\ 0.8 \pm 0.09 c \end{array}$	$\begin{array}{c} 2.1 \pm 0.03 a \\ 1.8 \pm 0.04 b \\ 1.5 \pm 0.04 c \end{array}$	$\begin{array}{c} 63.6 \pm 1.71b \\ 24.8 \pm 1.91c \\ 30.9 \pm 1.79c \end{array}$	28.3±1.60a 11.4±1.60c 5.3±1.07d

Average population density of *P. xylostella* during the pre-release period was 5.4 per plant (table 1). This value declined significantly in the first year after release and even more sharply in the second year to 0.7 per plant and then remained stable. Similarly, the percentage of attacked plants declined in the first and second year after release and then remained stable at around 30%. The damage score increased from the year before release (1.9) to the first year after but then declined and attained a significantly lower value (1.5) in the third year after release (table 1). The proportion of plants in damage score >2 also increased in the first year after release but declined by a factor of four to 5.3% at the end of the observations.

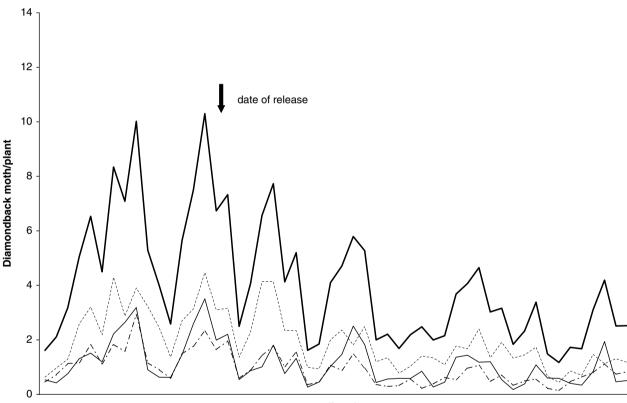
When the periods of highest population levels were compared over the four years, a factor eight reduction was recorded (table 2), while the reduction during the five months of low population levels was only by a factor of four. The age structure of the population did not change (data not shown); small larvae always constituted the largest part of the population.

Tharuni

The highest population of *P. xylostella* was recorded on 8 February 2002 (12.8 per plant, not shown); and peak periods were also in the dry seasons from June to October and February to March. Peak rainfall (fig. 2) coincided very

Table 2. Diamondback moth (DBM) population during the five months of lowest and highest infestation before and after introduction of the exotic parasitoid, *Diadegma semiclausum*, Werugha, Coast Province of Kenya.

Sampling period	5 months of lowest population		5 months of highest population		
	Observations	DBM per plant	Observations	DBM per plant	
Before release	75	1.5±0.21a	74	11.7±0.90a	
After release Year 1 Year 2 Year 3	75 75 74	$\begin{array}{c} 1.1 \pm 0.14 a \\ 0.3 \pm 0.07 b \\ 0.4 \pm 0.05 b \end{array}$	73 75 75	$7.7 \pm 0.73b$ $1.4 \pm 0.20c$ $1.4 \pm 0.22c$	



sampling date

Fig. 4. Dynamics and composition of diamondback moth population before and after release of an exotic parasitoid, Tharuni, Limuru Division, Central Province of Kenya. Data were collected from ten plants in each of up to 15 farmer-managed fields. Sampling dates within one month were pooled for ease of presentation. (- - -, small larvae; ---, big larvae; ---, total)

well with great reductions in the population (fig. 4). After release of the parasitoid, there was a declining trend of the population like at Werugha, but the final population remained higher (2.5 per plant).

Overall, average P. *xylostella* population declined year after year and ended at 2.4 per plant in the third year after release. A similar development was recorded for the damage (table 3). There was no clear change in the proportion of plants attacked; however, the plants in the damage score group >2 declined significantly in the first two years to stabilize between 9.2 and 4.5% (table 3). As at Werugha, the

highest reduction (by a factor of three) of the population was registered during the period of population peaks while the reduction during the low population period was only by 50% (table 4). The age distribution of the population did not alter much and small larvae were always the most frequent.

Development of parasitism

Four species of indigenous parasitoids were collected at both sites: *Diadegma mollipla* (Holmgren) (Hymenoptera:

Sampling period	Observations	DBM per plant	Damage score	% plants attacked	% plants with damage score > 2
Before release After release	401	$5.9\pm0.25a$	$2.3 \pm 0.03a$	$88.9\pm0.85a$	39.7±1.54a
Year 1 Year 2 Year 3	338 149 146	$\begin{array}{c} 4.3 \pm 0.25 b \\ 3.0 \pm 0.21 c \\ 2.4 \pm 0.12 c \end{array}$	$2.0 \pm 0.02b$ $1.8 \pm 0.03c$ $1.7 \pm 0.03d$	86.6±0.95ab 83.3±1.57b 87.8±1.45a	$15.7 \pm 1.17b$ $9.2 \pm 1.27c$ $4.5 \pm 0.77c$

Table 3. Diamondback moth (DBM) population and damage before and after introduction of the exotic parasitoid, *Diadegma semiclausum*, Tharuni, Central Province of Kenya.

Table 4. Diamondback moth (DBM) population during the five months of lowest and highest infestation before and after introduction of the exotic parasitoid, *Diadegma semiclausum*. Tharuni, Central Province of Kenya.

Sampling Period	5 months of lowest population		5 months of highest population		
	Observations DBM per plant		Observations	DBM per plant	
Before release	52	$2.9 \pm 0.35a$	54	10.2±1.14a	
After release Year 1 Year 2 Year 3	45 36 33	$\begin{array}{c} 1.7 \pm 0.15 b \\ 2.1 \pm 0.26 a b \\ 1.5 \pm 0.15 b \end{array}$	47 42 34	$8.0 \pm 1.23a$ $4.2 \pm 0.62b$ $3.5 \pm 0.12b$	

Means with different letters within a column are significantly different (P < 0.05), Tukey's honest significant difference test.

Ichneumonidae), *Oomyzus sokolowskii* (Kurdjumov) (Hymenoptera: Eulophidae), *Apanteles* sp. (Hymenoptera: Braconidae, undescribed species) and *Itoplectis* sp. (Hymenoptera Ichneumonidae), all primary parasitoids. A single specimen of a suspected hyperparasitoid (Hymenoptera: Ichneumonidae) was collected at Werugha, but the cocoon was discarded before a dissection had been conducted to confirm the status of the parasitoid.

Werugha

Pre-release parasitism was low from April 2001 until February 2002 when the total parasitism surpassed 20% for the first time (fig. 5). The highest level reached was 38.3% at the beginning of April (not shown) and from the end of May the rate dropped sharply again. Oomyzus sokolowskii was the parasitoid responsible for this rise and was the only parasitoid with strongly expressed seasonality (fig. 5). Diadegma mollipla was present in most collections, even though at a very low level. The first collection after release (August 2002) yielded D. semiclausum and even though numbers remained low until mid February 2003, the species was present in all collections. From the end of February, D. semiclausum parasitism surged and peaks of 80% parasitism were reached in March 2004 and again in June 2005. From June 2003, the indigenous parasitoids disappeared almost completely, and only D. mollipla parasitism recovered slightly starting from June 2004 (fig. 5).

Even though the number of specimens collected for laboratory rearing to establish levels of parasitism cannot be considered strictly as a parameter (and was thus not subjected to statistical analysis), the decline of almost 50% in the first year and of 89.8 and 88.6% (year 2 and 3, respectively) indicates the huge reduction in the field population of the pest (table 5). The major parasitoid species recorded before release of the exotic species were *O. sokolowskii* and *D. mollipla*. The introduced parasitoid surpassed the overall parasitism rate of all indigenous species combined even in the first year after its release, and the final rate of parasitism was around 50% (table 5).

Tharuni

Parasitism was almost non-existent until March 2002 (fig. 6). From then, low rates of D. mollipla were recorded. After release, there was a slight surge in parasitism by the former and O. sokolowskii in January 2003, which faded again in April. The first recovery of D. semiclausum was in January 2003, when eight specimen were recovered. Parasitism remained low until September 2003 but increased to 54% in November. After very erratic rises and declines, the introduced parasitoid finally became firmly established from September 2004 onwards and reached the highest rate of parasitism (75.0%) at the end of the observations in September 2005 (fig. 6). Diamondback moth larvae collected for laboratory observation of parasitism declined by 33% and 65% (first and second year after release, respectively) and only marginally thereafter (table 6). Parasitism by the indigenous parasitoids increased slightly in the first year after release, then these species almost vanished. Diadegma semiclausm parasitism increased very slowly in the first year and reached 39.9% in the third year after release, comprising almost 100% of overall parasitism (table 6).

Comparison of the two release areas

Development of *P. xylostella* populations and parasitism after release were compared between the two pilot

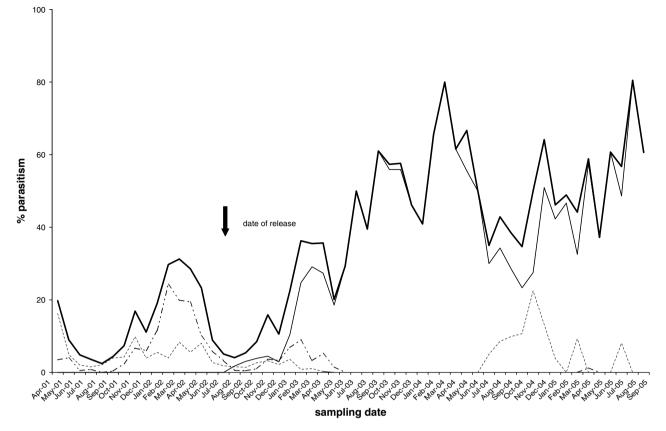


Fig. 5. Variation in field parasitism and parasitoid guild composition of diamondback moth before and after release of an exotic parasitoid, Werugha, Taita Hills, Coast Province of Kenya. Data were collected from ten plants in each of 15 farmer-managed fields. Sampling dates within one month were pooled for ease of presentation. (- - -, *Diadegma mollipla; ---, Oomyzus sokolowskii; --, Diadegma semiclausum; --,* Total parasitism)

Table 5. Field parasitism of diamondback moth before and after introduction of the exotic
parasitoid, Diadegma semiclausum, Werugha, Coast Province of Kenya.

Sampling	Specimen collected	Contribution of species to overall parasitism					
period		Diadegma mollipla	Oomyzus sokolowskii	Diadegma semiclausum	Total		
Before release	6285	5.4 ± 0.51 a	9.0±0.80a		$14.4\pm0.9c$		
After release Year 1 Year 2 Year 3	3292 667 740	$2.2 \pm 0.54 bc$ $1.4 \pm 0.93 c$ $5.0 \pm 0.97 ab$	$4.3 \pm 0.69b$ 0c $0.1 \pm 0.05c$	$17.4 \pm 1.59b$ $50.5 \pm 3.37a$ $47.4 \pm 2.74a$	$23.9 \pm 1.7b$ $51.9 \pm 3.3a$ $52.5 \pm 2.6a$		

areas using regression analysis. The population decline at Werugha was considerably, and significantly, faster than at Tharuni (F 1,98, 24.91, P < 0.0001) (fig. 7). While the regression line for Werugha cuts the zero population mark at sampling 43, the position of the regression line for Tharuni indicates that the population decline may not have reached a final level yet.

The increase in parasitism rates of *D. semiclausum* after the first field recovery was similar at both sites (F 1,97, 0.20, P = 0.645) even though the first recovery at Tharuni was only

three months after release (fig. 8). No differences were found between the release areas in total parasitism development after release and within release areas between total parasitism and the contribution of *D. semiclausum* (not shown).

Parasitoid species diversity and evenness at Werugha were similar before and one year after release (table 7). In the second and third year after release, both indices declined sharply with the displacement of almost all indigenous parasitoid species. At Tharuni, diversity and evenness increased in the first after-release year and then declined

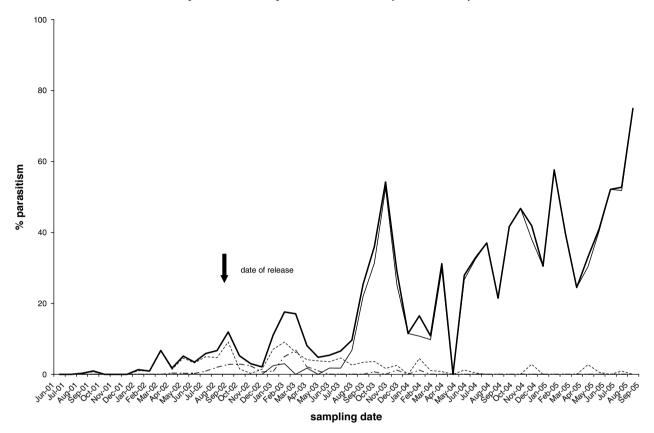


Fig. 6. Variation in field parasitism and parasitoid guild composition of diamondback moth before and after release of an exotic parasitoid, Tharuni, Limuru Division, Central Province of Kenya. Data were collected from ten plants in each of up to 15 farmer-managed fields. Sampling dates within one month were pooled for ease of presentation. (- - -, *Diadegma mollipla*; ---, *Oomyzus sokolowskii*; --, *Diadegma semiclausum*; --, Total parasitism)

Table 6. Field parasitism of diamondback moth before and after introduction of the exotic
parasitoid, Diadegma semiclausum, Tharuni, Central Province of Kenya.

Sampling period	No of specimens	Contribution of species to overall parasitism				
	collected	Diadegma Mollipla	Oomyzus sokolowskii	Diadegma semiclausum	Total	
Before release	7449	$3.3\pm0.42ab$	$0.9\pm0.31ab$		$4.2 \pm 0.56d$	
After release Year 1 Year 2 Year 3	4968 1722 1366	$4.8 \pm 0.54a$ $1.3 \pm 0.31bc$ $0.7 \pm 0.29c$	$\begin{array}{c} 2.3 \pm 0.40 a \\ 0.4 \pm 0.17 b \\ 0 b \end{array}$	$3.1 \pm 0.51c$ 24.9 $\pm 2.03b$ 39.9 $\pm 2.05a$	$\begin{array}{c} 10.2 \pm 0.84 c \\ 26.6 \pm 2.07 b \\ 40.6 \pm 2.07 a \end{array}$	

to below before-release levels in years 2 and 3 (table 7). Indigenous parasitoids also disappeared almost completely.

Discussion

In spite of the great number of introductions of *D. semiclausum* against *P. xylostella* in Southeast Asia, long-term studies of parasitoid impact on *P. xyostella* populations and indigenous parasitoids are not available. Poelking (1992) gave a detailed account of the early impact of *D. semiclausum*

releases in the Philippines with >95% parasitism towards the end of the first after-release cabbage season. In Taiwan, Talekar *et al.* (1992) reported establishment after a single release in the highlands. Establishment at Werugha was similarly fast as reported in the Philippines and was documented by Momanyi *et al.* (2006) in field mortality studies of larvae and pupae. However, establishment of the parasitoid at Tharuni was only noted three months after introduction and impact on the *P. xylostella* population took even longer to show. Momanyi *et al.* (2006) adduce climatic

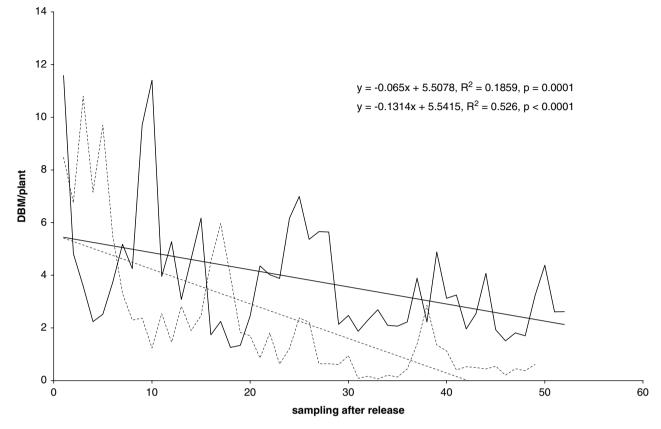


Fig. 7. Diamondback moth population development after release of an exotic parasitoid in two pilot release areas, Werugha, Wundanyi Division, Coast Province and Tharuni, Limuru Division, Central Province of Kenya. (—, Tharuni; - - -, Werugha; ——, linear regression Tharuni; - - -, linear regression Werugha)

differences, in particular considerably lower rainfall and resulting dusty conditions, to explain the delay in parasitoid establishment. We believe that the erratic rainfall with an evapotranspiration deficit of probably nine out of twelve months every year and the resulting discontinuity of cabbage production at Tharuni must also have contributed to the slow pace of *D. semiclausum* establishment. The difficulties of the exotic species are not surprising as marginal conditions for parasitoids at Tharuni were already indicated by the extremely low parasitism rate by indigenous species before introduction. Natural expansion of D. semiclausum from later releases in neighbouring areas (approximately 10 km distance) may have contributed to the fast increase in parasitism when expanding populations could provide much higher and permanent parasitoid numbers for invasion of the marginal areas than the initial release.

Studies of the competitive displacement of indigenous parasitoids of *P. xylostella* after the introduction of exotic species are not available in spite of the introductions made in many parts of the world. In our case, the displacement of the indigenous species by *D. semiclausum* may have been caused by the superior host searching capability and better association with cruciferous host plants, at least as far as its congeneric indigenous species is concerned (Rossbach *et al.*, 2005). The aforementioned authors actually consider *D. mollipla* a generalist parasitoid as it was originally described as *Limneria mollipla* (Holmgren) from potato tuber

moth, Phthorimaea operculella (Zeller) (Lepidoptera: Gelechiidae). It is reported to be indigenous to southern and eastern Africa but the original host is unknown (Broodryk, 1971; Gupta, 1974; Azidah et al., 2000). Unfortunately, similar information about the other indigenous parasitoids is not available. Nevertheless, we assume that all indigenous parasitoids must have alternative hosts in addition to *P. xylostella* and have just made use of a largely unexploited resource, P. xylostella larvae, in the absence of other more specialized parasitoids. After the introduction of D. semiclausum, they were displaced from the system. Similar cases have been reported from other host-parasitoid systems. Bennett (1993) reported the displacement of Pseudhomalopoda prima Girault (Hymenoptera: Encyrtidae), a local opportunist parasitoid of the introduced scale Chrysomphalus anoidum (Linnaeus) (Homoptera: Diaspididae), after the introduction of Aphytis holoxanthus DeBach (Hymenoptera: Aphelinidae), an exotic specialist parasitoid, to California. In West Africa, Anagyrus niombae Boussienguet (Hymenoptera: Encyrtidae) (Boussienguet, 1988) and other species of Anagyrus adapted to the introduced cassava mealybug but were displaced after the introduction of Apoanagyrus lopezi (DeSantis) (Hymenoptera: Encyrtidae), a host-specific cassava mealybug parasitoid from South America (Bassiangama et al., 1989; Boussienguet et al., 1991). Nevertheless, the speed of the displacement in the present case was surprising; it was almost complete when parasitism by D. semiclausum was still

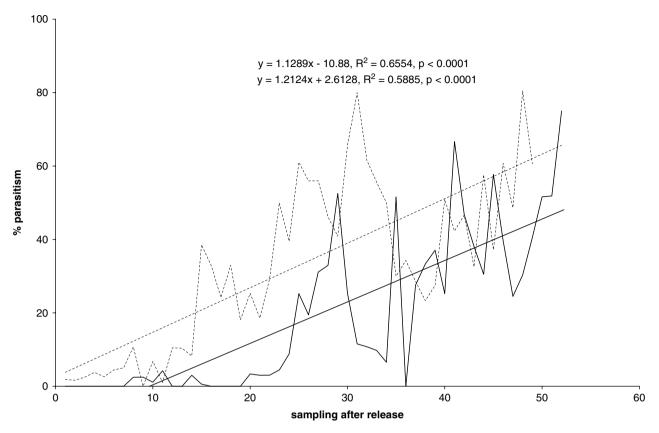


Fig. 8. Development of parasitism of diamondback moth after release of an exotic parasitoid in two pilot release areas (Werugha, Taita Hills, Coast Province and Tharuni, Limuru Division, Central Province of Kenya). (—, Tharuni; - - -, Werugha; ——, linear regression Tharuni; - - -, linear regression Werugha)

moderate and many unparasitized diamondback moth larvae were available.

A close look at the dramatic increase in parasitoid guild diversity and evenness at Tharuni in the year immediately after introduction suggests that this was rather an effect of the much better rainfall (five months with around 100 mm or considerably more, fig. 2) allowing better crop growth and hence parasitoid activity than a direct effect of the introduced parasitoid. In comparison, in the period before release from June 2001 to February 2002, there were seven consecutive months with a rainfall deficit.

The almost complete absence of hyperparasitoids is surprising and must have contributed to the fast establishment and impact of D. semiclausum. During the course of the work presented here, 3943 parasitized P. xylostella larvae and pupae were collected and there was only one doubtful case of hyperparasitism at Werugha by an ichneumonid. This is in stark contrast to the situation in South Africa, where Kfir (1997) and Ullyett (1947) identified a number of hyperparasitoids, some of which were shared between D. mollipla and Cotesia plutellae (Kurdjumov) (Hymenoptera: Braconidae). Poelking (1992) stated briefly that D. semiclausum is attacked by the same hyperparasitoids as C. plutellae. We have observed hyperparasitoids (yet unidentified Pteromalidae and Eulophidae) of Apanteles sp. attack cocoons of C. plutellae in the lowlands; however, so far none of these has attacked D. semiclausum in more than 12 months of observations (B. Löhr et al., unpublished data).

The present data indicate that the effect of the release of *D. semiclausum* on *P. xylostella* populations and damage, both under favourable (Werugha) and unfavourable (Tharuni) conditions, are substantially larger than in the assumptions made for the prediction of the economic impact of this biological control project for Kenya by Macharia *et al.* (2005). In that paper, a cost benefit ratio of 1:23.7 for cabbage production in Kenya was predicted with the assumptions of a 50% reduction in pesticide use and a reduction of the damage by 30%. For the damage we can now consider a 50% reduction from our data. When the corresponding loss abatement figure is inserted in the sensitivity analyses conducted by Macharia *et al.* (2005), the cost benefit value increases to 1:34.4 and the internal rate of return to 100.1%.

The reduction of pesticide overuse was one of the major aims of the Diamondback moth biocontrol project and activities to document this are still ongoing. However, at the pilot sites this was not the major thrust of the work. Assessing pesticide reduction is therefore difficult, but a rough estimate can be obtained using intervention thresholds for control operations for which between 0.3 and 1.0 larvae per plant have been suggested (Simonet & Morisak, 1982; Cartwright *et al.*, 1987; Amend & Mangalli, 1992). If an intervention threshold of one large *P. xylostella* larva per cabbage is assumed valid for Kenyan conditions, where quality requirements are lower than in developed countries, a pesticide application would have been required before parasitoid release after 76% of the sampling occasions at

Table 7. Shannon index for species diversity and evenness of the diamondback moth parasitoid guild before and after the introduction of an exotic parasitoid species, Werugha, Coast Province and Tharuni, Central Province of Kenya.

Site	Parasitoid species	Before release	Years after release		
			1	2	3
Werugha	Diadegma mollipla	267	52	1	58
0	Oomyzus sokolowskii	374	92	-	1
	Apanteles sp.	2	1	_	1
	Itoplectis sp.	74	45	3	1
	Diadegma semiclausum		443	353	300
	п	717	633	357	361
	Species	4	5	3	5
	Diversity index	0.958	0.934	0.068	0.497
	Evenness	0.691	0.580	0.062	0.309
			t=0.6	t = 18.9	t = 8.6
			P > 0.05	P < 0.001	P < 0.001
Tharuni	D. mollipla	138	224	58	21
	O. sokolowskii	31	93	3	0
	Apanteles sp.	1	12	1	0
	Itoplectis sp.	6	0	15	2 2
	Brachymeria sp.	3	3	0	2
	D. semiclausum	-	49	576	700
	п	179	381	654	725
	Species (S)	5	5	5	4
	Diversity index	0.716	1.067	0.447	0.169
	Evenness	0.445	0.596	0.323	0.122
			t = 4.35 P < 0.001	t = 11.48 P < 0.001	t = 6.16 P < 0.001

t-tests were conducted to compare differences with the previous year.

Tharuni and 57% at Werugha. Three years after release only 7.7% of the samplings at Tharuni and none at Werugha would have triggered pesticide application, a reduction of >90%. This figure is also realistic considering experience with farmers from various cabbagegrowing areas. When a conservative 70% reduction in pesticide use is added to 50% loss abatement for the economic impact analysis, the benefit increases to 1:38.1 and the internal rate of return to 104.5% (I. Macharia, personal communication). We therefore expect that this biological control project will rank as a very highly successful one when the final economic impact assessment is concluded.

Acknowledgements

Due to the long duration of the study and the spatial separation of the two pilot areas, a number of different persons and institutions were involved in this work: F. Nang'ayo and M. Kusewa of the Kenya Agricultural Research Institute were transferred and did not see this effort through until the end; N. Mwikya, F. Nyamu, R. Mukiti, G. Gachanja, C. Kanyi of the International Centre of Insect Physiology and Ecology and P. Onano and S. Juma of the District Agricultural Office, Taita Taveta District, are gratefully acknowledged for their huge input into these studies. Anthony Wanjoya and L. Nedorezov provided advice and guidance with statistical analysis. We thank all the farmers who allowed us to work in their fields, over 200 in total over the period of four years. Thanks are also extended to the German Federal Ministry of Economic Cooperation and Development (BMZ) who funded the Diamondback Moth Biocontrol Project through the German Agency of Technical Cooperation (GTZ).

References

- Amend, J. & Mangalli, T. (1992) Field release of the parasitoid Diadegma semiclausum (Hellen) (Hymenoptera: Ichneumonidae) against Plutella xylostella (L.) (Lepidoptera: Yponomeutidae) in seed Petchay in Northern Luzon. Philippine Journal of Plant Industry 57, 53–62.
- Ayalew, G., Löhr, B., Baumgärtner, J. & Ogol, C.K.P.O. (2004) Diamondback moth, *Plutella xylostella* (Lepidoptera: Plutellidae) and its parasitoids in Ethiopia. p. 140 in Kirk, A.A. & Bordat, D. (Eds) Proceedings of the International Symposium Improving Biocontrol of Plutella xylostella. Montpellier France, 21–24 October 2002.
- Azidah, A.A., Fitton, M.G. & Quicke, D.L.J. (2000) Identification of the *Diadegma* species (Hymenoptera: Ichneumonidae, Campopleginae) attacking the diamondback moth, *Plutella xylostella* (Lepidoptera: Plutellidae). *Bulletin of Entomological Research* **90**, 375–389.
- Bassiangama, A., le Rü, B., Iziquel, Y., Kiyindou, A. & Bimangou, A.S. (1989) L'entomocénose inféodée à la cochenille du manioc, *Phenacoccus manihoti* (Homoptera: Pseudococcidae), au Congo, cinq ans après l'introduction d'Epidinocarsis lopezi (Hymenoptera: Encyrtidae). Annales de la Sociétée Entomologique de France 25, 315–320.
- Batten, L.A. (1976) Bird communities of some Killarney woodlands. *Proceedings of the Royal Irish Academy* 76, 285–313.
- Bennett, F. (1993) Do introduced parasitoids displace native ones? Florida Entomologist 76, 54–63.

- Biever, K.D. (1996) Development and use of a biological control–IPM system for insect pests of crucifers. p. 257 in Sivapragasam, A., Kole, W.H., Hassan, A.K. & Lim, G.S. (Eds) The management of diamondback moth and other crucifer pests. Proceedings of the Third International Workshop, Kuala Lumpur, Malaysia, 29 October–1 November 1996.
- Boussienguet, J. (1988) Morphologie et biologie d'Anagyrus nyombae, n. sp., parasite de Phenacoccus manihoti, au Gabon (Hymenoptera, Encyrtidae; Homoptera, Pseudococcidae). Revue Francaise d'Entomologie 10, 277–283.
- Boussienguet, J., Neuenschwander, P. & Herren, H.R. (1991) Le complexe entomophage de la cochenille du manioc au Gabon. 4. Etablissement du parasitoide *Epidinocarsis lopezi*. *Entomophaga* 36, 455–469.
- Broodryk, S.W. (1971) The biology of *Diadegma stellenboschense* (Cameron) (Hymenoptera: Ichneumonidae), a parasitoid of potato tuber moth. *Journal of the Entomological Society of South Africa* 34, 413–423.
- Cartwright, B., Edelson, J.V. & Chambers, C. (1987) Composite action thresholds for the control of lepidopterous pests on fresh-market cabbage in the lower Rio Grande Valley of Texas. *Journal of Economic Entomology* 80, 175–181.
- Eusebio, J.E. & Morallo-Rejesus, B. (1996). Integrated pest management of diamondback moth: the Philippine highlands' experience. p. 253 in Sivapragasam, A., Kole, W.H., Hassan, A.K. & Lim, G.S. (Eds) The management of diamondback moth and other crucifer pests. Proceedings of the Third International Workshop, Kuala Lumpur, Malaysia, 29 October–1 November 1996.
- Evans, I.B.P. (1939) Solving the Union's pasture, crop and insect problems. Annual Report of the Division of Plant Industry. *Farming in South Africa* **117**, 22 pp.
- Greathead, D. (1971) A review of biological control in the Ethiopian region. Commonwealth Institute of Biological Control Technical Communication No. 5, Commonwealth Agricultural Bureaux, Slough, UK, 162 pp.
- Gupta, V.K. (1974) Studies on certain porizontine ichneumonids reared from economic hosts (parasitic Hymenoptera). Oriental Insects 3, 99–116.
- Hamilton, J.T. (1979) Seasonal abundance of *Pieris rapae* (L.), *Plutella xylostella* (L.) and their diseases and parasites. *General and Applied Entomology* **11**, 59–66.
- Harcourt, D.G. Backs, R.H. & Case, L.M. (1955) Abundance and relative importance of caterpillars attacking cabbage in Eastern Ontario. *Canadian Entomologist* 87, 400–406.
- Hardy, J.E. (1938) Plutella maculipennis Curt., its natural and biological control in England. Bulletin of Entomological Research 29, 343–373.
- Iga, M. (1997) Effect of release of the introduced ichneumonid parasitoid, *Diadegma semiclausum* (Hellen), on the diamondback moth, *Plutella xylostella* (L.) in an experimental cabbage field. *Japanese Journal of Applied Entomology and Zoology* 44, 195–199.
- Kfir, R. (1997) Parasitoids of diamondback moth, *Plutella xylostella* (L.) (Lepidoptera: Yponomeutidae), in South Africa: an annotated list. *Entomophaga* 42, 517–523.
- Kfir, R. (2003) Effect of parasitoid elimination on populations of diamondback moth on cabbage. p. 197 *in* Endersby, N.M. & Ridland, P.M. (*Eds*) *The management of diamondback moth and* other crucifer pests. Proceedings of the Fourth International Workshop, 26–29 November 2001, Melbourne, Victoria, Australia, Australian Entomological Society.
- Kibata, G.N. (1996) The diamondback moth: a problem pest of brassica crops in Kenya. p. 47 *in* Sivapragasam, A., Kole,

W.H., Hassan, A.K. & Lim, G.S. (Eds) The management of diamondback moth and other crucifer pests. Proceedings of the Third International Workshop, Kuala Lumpur, Malaysia, 29 October–1 November 1996.

- Lim, G.S., Sivapragasam, A. & Loke, W.H. (1996) Crucifer insect pest problems: trends, issues and management strategies. p. 3 in Sivapragasam, A., Kole, W.H., Hassan, A.K. & Lim, G.S. (Eds) The management of diamondback moth and other crucifer pests. Proceedings of the Third International Workshop, Kuala Lumpur, Malaysia, 29 October–1 November 1996.
- Macharia, I., Löhr, B. & DeGroote, H. (2005) Assessing the potential impact of biological control of the diamondback moth in cabbage production in Kenya. *Crop Protection* 24, 981–989.
- Momanyi, C., Löhr, B. & Gitonga, L. (2006) Biological impact of the exotic parasitoid, *Diadegma semiclausum* (Hellen), of diamondback moth, *Plutella xylostella* L., in Kenya. *Biological Control* 38, 254–263.
- Mugurran, A.E. (1988) *Ecological diversity and its measurement*. 179 p. Princeton, New Jersey, Princeton University Press.
- Oatman, E.R. & Platner, G.R. (1969) An ecological study of insect populations on cabbage in Southern California. *Hilgardia* 40, 1–40.
- Oduor, G.I., Löhr, B. & Seif, A.A. (1996) Seasonality of major cabbage pests and incidence of their natural enemies in Central Kenya. p. 37 in Sivapragasam, A., Kole, W.H., Hassan, A.K. & Lim, G.S. (Eds) The management of diamondback moth and other crucifer pests. Proceedings of the Third International Workshop, Kuala Lumpur, Malaysia, 29 October–1 November 1996.
- **Ooi, P.** (1992) Role of parasitoids in managing diamondback moth in the Cameron Highlands, Malaysia. p. 255 *in* Talekar, N.S. (*Ed.*) *Diamondback moth and other crucifer pests. Proceedings of the Second International Workshop*, Tainan, Taiwan, 10–14 December 1990.
- Poelking, A. (1992) Diamondback moth in the Philippines and its control with *Diadegma semiclausum*. p. 271 *in* Talekar, N.S. (*Ed.*) *Diamondback moth and other crucifer pests*. *Proceedings of the Second International Workshop*, Tainan, Taiwan, 10–14 December 1990.
- Rossbach, A., Löhr, B. & Vidal, S. (2005) Generalism versus specialism: response of *Diadegma mollipla* (Holmgren) and *Diadegma semiclausum* (Hellen) to the host shift of the diamondback moth (*Plutella xylostella* L.) to peas. *Journal of Insect Behaviour* 18, 491–503.
- Ru, N. & Workman, R.B. (1979) Seasonal abundances and parasites of imported cabbage worm, diamondback moth and cabbage webworm in Northeast Florida. *Florida Entomologist.* 62, 68–69.
- SAS Institute Inc. (2002) SAS/STAT Users Guide, Cary, North Carolina, USA.
- Sastrosiswojo, S. & Sastrodihardjo, S. (1986) Status of biological control of diamondback moth by introduction of parasitoid *Diadegma eucerophaga* in Indonesia. p. 185 in Talekar, N.S. & Griggs, T.D. (Eds) Diamondback moth management: Proceedings of the First International Workshop, Shanhua, Taiwan, 11–15 March 1985.
- Shelton, A.M., Perez, C.J., Tang, J.D. & Van den Berg, J.D. (1996) Prospects for novel approaches towards management of the diamondback moth. p. 17 in Sivapragasam, A., Kole, W.H., Hassan, A.K. & Lim, G.S. (Eds) The management of diamondback moth and other crucifer pests. Proceedings of the Third International Workshop, Kuala Lumpur, Malaysia, 29 October–1 November 1996.

- Simonet, D.E. & Morisak, D.J. (1982) Utilizing action thresholds in small-plot insecticide evaluations against cabbage-feeding lepidopterous larvae. *Journal of Economic Entomology* 75, 43–46.
- Talekar, N.S., Yang, J.C. & Lee, S.T. (1992) Introduction of Diadegma semiclausum to control diamondback moth in Taiwan. p. 263 in Talekar, N.S. (Ed.) Diamondback moth and other crucifer pests. Proceedings of the Second International Workshop, Tainan, Taiwan, 10–14 December 1990.
- Uk, S. & Harris, J.G. (1996) The historical failure of insecticide resistance management of the diamondback moth and the way forward. p. 222 in Sivapragasam, A., Kole, W.H., Hassan, A.K. & Lim, G.S. (Eds) The management of diamondback moth and other crucifer pests. Proceedings of the

Third International Workshop, Kuala Lumpur, Malaysia, 29 October–1 November 1996.

- Ullyett, G.C. (1947) Mortality factors in populations of Plutella maculipennis Curt. (Tineidae: Lep) and their relation to the problem of control. Entomological Memoirs, Department of Agriculture and Forestry, Union of South Africa 2, 77–202.
- Verkerk, R.H.J. & Wright, D.J. (1996) Multitrophic interactions and management of the diamondback moth: a review. Bulletin of Entomological Research 86, 205–216.

(Accepted 26 November 2006) © 2007 Cambridge University Press