The lichen genus Topeliopsis, additions and corrections

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Abstract: Topeliopsis darlingtonii A. Frisch & Kalb and Topeliopsis elixii A. Frisch & Kalb from Australia and Topeliopsis meridensis Kalb & A. Frisch from Venezuela are described as new species. Thelotrema decorticans Müll. Arg. and Ocellularia subdenticulata Zahlbr. are transferred to Topeliopsis and are the correct names for T. corticola Kalb and T. vezdae Kalb, respectively. A revised key of the genus Topeliopsis is presented. Ascoconidia, produced in old ascospores of T. elixii, are described for the first time for the Thelotremataceae.

Key words: ascoconidia, Australia, *Graphidaceae*, lichenized ascomycetes, *Thelotremataceae*, *Topeliopsis*, Venezuela.

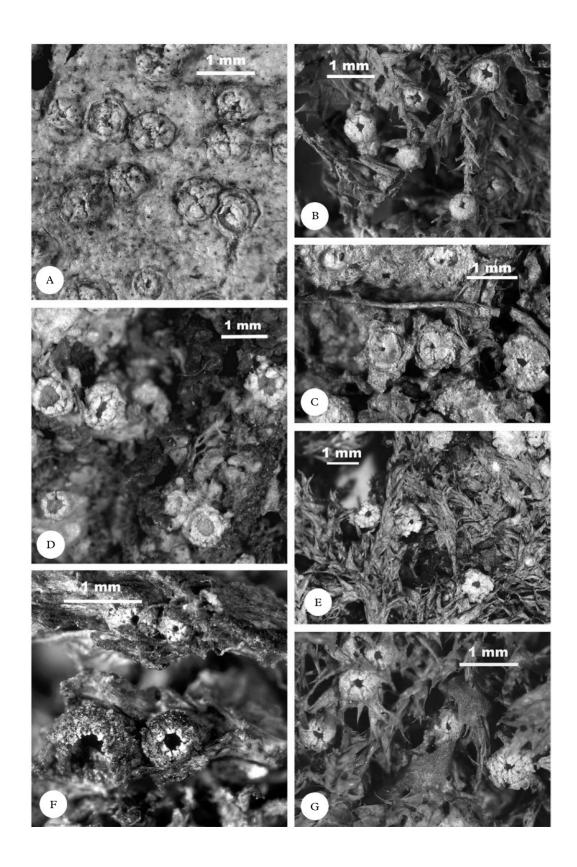
Introduction

The lichen genus Topeliopsis was described by Kantvilas & Vězda (2000) for three species of Thelotremataceae with adnate urceolate apothecia, a fused proper exciple with prominent periphysoids, and large hyaline, muriform, thin-walled ascospores turning reddish or purple in iodine. In this circumscription, Topeliopsis was noted by the authors to be rather heterogeneous and, in consequence, two of the species were removed from the genus (Kalb 2001). This author accepted only the generic type, Topeliopsis muscigena (Stiz.) Kalb (syn. T. muscicola Kantvilas & Vězda), but described three new species from Australia, namely Topeliopsis acutispora, T. corticola and T. vezdae. Also, the concept of Topeliopsis was emended to accommodate species with thick-walled ascospores, since it could be demonstrated that the ascospores Topeliopsis muscigena are not thin-walled as described in the protologue, but belong to the thick-walled type prevalent in the Thelotremataceae. This is apparent in the

slight thickening of the endospore, which in this species is always recognizable at the edges of the locules. Furthermore, the colour of the ascospore walls in iodine confirms the thick-walled type, as the amyloidity of the 'graphidean'-type ascospores of the Thelotremataceae is always located in the endospore. Current knowledge suggests that all species of Thelotremataceae with truly thin-walled ascospores lack any amyloid coloration (A. Frisch unpublished). Marked reduction of the endospore, as is observed for Topeliopsis muscigena and the two newly described species Topeliopsis elixii and T. meridensis, is a common phenomenon in densely muriform ascospores of the Thelotremataceae. This is largely an effect of the intensity of the cell division and the number of locules, and cannot be given taxonomic importance at genus level. Consequently, species with a markedly thickened endospore, all with transversely septate or submuriform ascospores, were accepted for the genus as they agree well with the type species in apothecium and thallus structure, as well as in ecology and distribution.

During the investigation of type material for a revision of the African *Thelotremataceae* (A. Frisch unpublished), we discovered two names belonging to *Topeliopsis* as recently circumscribed, namely *Thelotrema decorticans*

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Müll. Arg. and *Ocellularia subdenticulata* Zahlbr. These two species proved to be conspecific with *Topeliopsis corticola* Kalb and *T. vezdae* Kalb, respectively. In addition,

two new species were discovered recently in Australia and another in Venezuela. In this paper, we describe the new species and make the necessary new combinations.

Revised key to the species of the lichen genus Topeliopsis

1	Ascospores transversely septate
2(1)	Stictic acid present; ascospores 13–15-septate, $41–52 \times 9–10 \mu\text{m}$, with subacute ends
3(2)	Ascospores 25–35-septate, $90-130\times8-12~\mu m$, ends acute to subacute
4(1)	Ascospores 8/ascus, $50-60 \times 13-18 \mu m$
5(4)	Hymenium inspersed with minute spherical drops; excipulum brownish
6(5)	Hypostictic acid present; disc flat, white pruinose

The New Species

Topeliopsis darlingtonii A. Frisch & Kalb sp. nov.

Similis *Topeliopsidis subdenticulatae*, sed thallo acidum sticticum continente et apotheciis minus elevatis differt.

Typus: Australia, Queensland, Darlington Range, *c.* 15 km SSW of Canungra, in a cool temperate submontane rainforest, 28°13′32″S, 153°07′42″E, 980 m alt., 16 August 2002, *K. Kalb* 33979 (CANB—holotypus; hb. Kalb, hb. Frisch—isotypi).

(Figs 1A, 2 & 5A)

Thallus corticolous on soft bark, dirty grey to olive-grey, thin (c. 0.03–0.07 mm), with a smooth to slightly uneven, compact and glossy surface. Prothallus line thin, brownish.

Phenocortex (Büdel & Scheidegger 1996) 3–6 μm thick, without internal splitting, formed of strongly conglutinated periclinal hyphae. Photobiont layer 20–40 μm thick, without oxalate crystals. Medulla indistinct to absent.

Apothecia frequent, dispersed, moderately emergent, 0·5–0·8 mm diam., with a thick layered, incurved to rim-like, white to pale brown margin and a moderately wide, fissured pore. Hymenium pale brown, covered by a thin white, finely granular pruina. Phenocortex 3–6 μm thick. Photobiont layer 10–30 μm thick. Proper exciple cupular, hyaline, a strongly conglutinated prosoplectenchyma, 12–20 μm deep at

Fig. 1. Topeliopsis species. A, T. darlingtonii (holotype); B, T. acutispora (isotype, hb. Kalb); C, T. decorticans (holotype); D, T. elixii (holotype); E, T. muscigena (McOwan s.n. isotype PRE); F, T. meridensis (holotype); G, T. subdenticulata (Kalb 20481, hb. Kalb). Scales: A-G=1 mm.

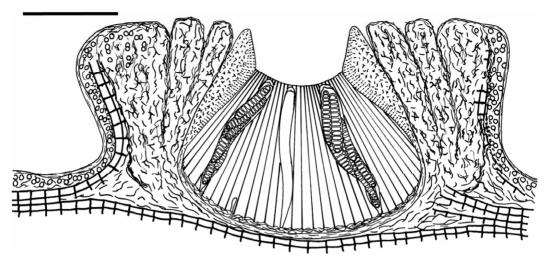


FIG. 2. Topeliopsis darlingtonii (holotype), section of thallus and apothecium. Scale=100 μm.

the base, split into up to three separate, 25–35 µm wide strands at the sides. *Periderm* layer present or absent, c. 25 µm wide, not conglutinated, formed of distinct rows of periderm cells. Periphysoids prominent, running down to the base of the proper exciple, perpendicular, up to 25 µm long and 2-2.5 μm wide, with 2-5 rectangular cells. Subhymenium 10–12 µm high. Hymenium 130-150 µm high, clear. Paraphyses simple, dense, $1.5-2 \mu m$ wide, not thickened at the apex. Epihymenium colourless, 7–10 µm high. Asci broadly clavate, c. $120-140 \times 15-$ 22 µm, with well-developed tholus when mature. Ascospores 8/ascus, hyaline, transversely 13–15-septate, $41–52 \times 9–10 \, \mu m$, with subacute ends, halonate in young stages, I+ pale purplish blue, spore development macrocephalic.

Pycnidia not seen.

Chemistry. Stictic acid (major), constictic acid (minor), α -acetylconstictic acid (minor) and consalazinic acid (minor) detected by TLC.

Etymology. The specific epithet refers to the type locality in the Darlington Range, Queensland.

Remarks. The layered margin of the apothecia of Topeliopsis darlingtonii is

reminiscent of some species of the *Thelotrema subtile*-group. However, the new species is distinguished by the fused proper exciple and paraphyses that are not thickened at the apex. The ascospores in the material investigated have only slightly thickened walls and are clearly halonate. It is possible that they are not fully mature in the material available. *Topeliopsis darlingtonii* is distinguished from the other *Topeliopsis* species with transversely septate ascospores (chiefly *T. subdenticulata*), by stictic acid in the thallus and by the less emergent apothecia with a layered margin.

Habitat and distribution. This new species is so far known only from the type locality where it grows abundantly on the smooth, weathered bark at the base of a deciduous tree, in a moist, cool temperate, submontane rainforest in Queensland/Australia.

Additional material examined. **Australia:** Queensland: Darlington Range, c. 15 km SSW of Canungra, in a cool temperate submontane rainforest, 28°13′32″S, 153°07′42″E, 980 m, 2002, *Kalb* 33980 (hb. Kalb).

Topeliopsis elixii A. Frisch & Kalb sp. nov.

Similis *Topeliopsidis muscigenae*, sed thallo acidum hyposticticum et acidum hypoconsticticum continente et apotheciis non distincte urceolatis, disco late aperto et pruinoso differt.

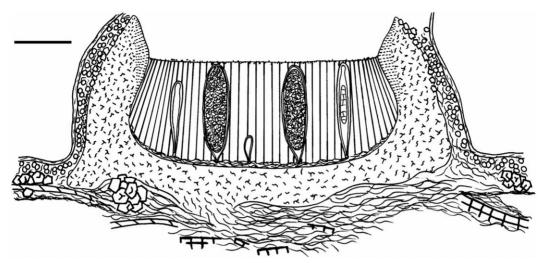


Fig. 3. Topeliopsis elixii (holotype), section of thallus and apothecium. Scale=100 μm.

Typus: Australia, Queensland, Darlington Range, c. 15 km SSW of Canungra, in a cool temperate submontane rainforest, 28°13′32″S, 153°07′42″E, 980 m alt., 16 August 2002, K. Kalb 33979 (CANB—holotypus; hb Kalb, hb Frisch—isotypi).

(Figs 1D, 3 & 5B)

Thallus muscicolous, olive-grey, thin (c. 0.03-0.12 mm), with a smooth to slightly verrucose, compact and glossy surface. Prothallus line thin, brownish. Phenocortex 10–15 μ m thick, without internal splitting, formed of strongly conglutinated periclinal hyphae. Photobiont layer 20–30 μ m thick, with a basal layer, up to 50 μ m thick, of oxalate crystals. Medulla indistinct to absent.

Apothecia frequent, dispersed, strongly emergent or with a slightly constricted base adnate to the thallus, 0·6-1·0 mm diam., with an incurved to rim-like margin and a narrow to wide, fissured pore. Thallus margin exfoliating and typically eroded at the apex, the crenate to deeply fissured, white to brownish white proper exciple exposed. Hymenium pale brown, finally flat, covered by a white, coarsely granular pruina. Phenocortex 10–15 μm thick. Photobiont layer up to 40 μm thick, with a basal layer of oxalate crystals. Periderm layer 10–15 μm wide, hyaline to pale brownish, of similar structure to the proper exciple, indistinct. Proper

exciple cupular, hyaline, with a strongly conglutinated prosoplectenchyma which often becomes almost paraplectenchymatous in the central portions, up to 120 µm deep at the base and 40-60 µm wide laterally. Periphysoids prominent, running down to the base of the proper exciple, perpendicular to typically slightly down-turned, up to 20 µm long and 2.5 μm wide, with up to three rectangular cells. Hymenium 170-220 µm high, clear. Paraphyses simple, dense and straight, 1.5–2 um wide, not thickened at the apex. Epihymenium colourless to very pale brown, 8-15 µm high, inspersed by fine brownish granules and a few small oxalate crystals. Asci broadly clavate, c. 145- $190 \times 25-48 \,\mu\text{m}$, without a well-developed tholus when mature. Ascospores 1/ascus, hyaline, densely muriform, halonate in young stages, $105-170 \times 22-45 \,\mu\text{m}$, with rounded ends, I+ purplish blue, spore development microcephalic.

Pycnidia not seen.

Chemistry. Hypostictic acid (major), hypoconstictic acid (minor) detected with TLC.

Etymology. The specific epithet refers to our friend and colleague, Prof. John A. Elix,

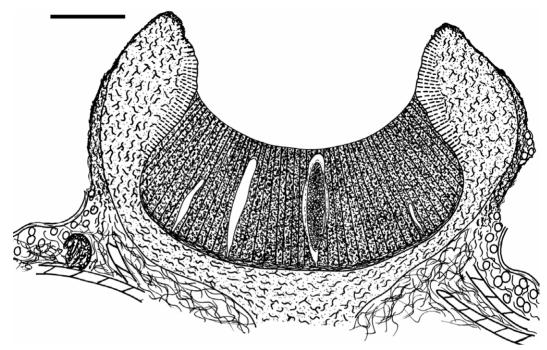


Fig. 4. Topeliopsis meridensis (holotype), section through thallus and apothecium. Scale = 100 μm.

Canberra, for his untiring support in chemistry as well as in the field.

Remarks. Topeliopsis elixii differs from T. muscigena, the only other species with a single densely muriform ascospore per ascus, by the presence of hypostictic and hypoconstictic acids in the thallus, and a flat, wide open disc which is covered by a distinct pruina. The apothecia are not typically urceolate.

Habitat and distribution. This new species is known only from the type locality where it grows abundantly over mosses at the base of a deciduous tree, in a moist, cool temperate, submontane rainforest in Queensland/ Australia together with *T. darlingtonii*.

Topeliopsis meridensis Kalb & A. Frisch sp. nov.

Similis *Topeliopsidis muscigenae*, sed hymenio dense insperso et excipulo brunneolo differt.

Typus: Venezuela, Merida, Distr. Rangel, between Laguna Mucubaji and Laguna Negra, c. 15 km SE of Apartaderos, over mosses in a paramo, 08°45′S, 70°45′E, 3500 m alt. 15 August 1989, K. & A. Kalb & López-Figueiras 34793 (hb. Kalb—holotypus).

(Figs 1F, 4 & 5C)

Thallus muscicolous, continuous, pale grey to pale brownish grey-olive, thin (c. 0.05-0.1 mm) to evanescent, with a smooth to slightly verrucose, compact and glossy surface. Prothallus line not seen. Phenocortex $8-12~\mu m$ thick, without internal splitting, formed of strongly conglutinated periclinal hyphae. Photobiont layer $20-80~\mu m$ thick. Medulla indistinct.

Apothecia few, dispersed, moderately to strongly emergent, 0·5–0·9 mm diam., broadly hemispherical to barrel-shaped, with an incurved rim and a c. 0·15–0·25mm wide, rugged to radially-fissured, white to brownish white pore. Thallus margin exfoliating, largely eroded, the brown to black proper exciple exposed. Hymenium deeply urceolate, dark, epruinose. Phenocortex 8–12 μm thick. Photobiont layer c. 25 μm thick. Periderm layer 20–25 μm wide, brown, of similar

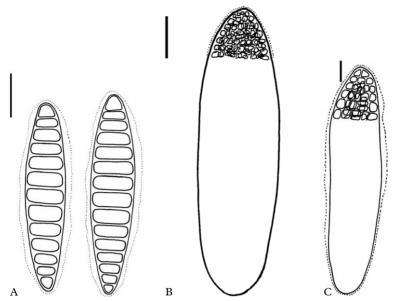


Fig. 5. Ascospores of *Topeliopsis* species. A, *T. darlingtonii* (holotype); B, *T. elixii* (holotype); C, *T. meridensis* (holotype). Scales: A & C=10 µm; B=20 µm.

structure to the proper exciple, indistinct. Proper exciple cupular, brown, composed of a strongly conglutinated prosoplectenchyma becoming almost paraplectenchymatous in the central portions, up to 60 µm deep at the base and 55-80 µm wide laterally. Periphysoids prominent, running down to the base of the proper exciple, perpendicular to slightly down-turned, up to 30 µm long and $2.5-3.0 \,\mu m$ wide, with up to four rectangular cells. Hymenium 170–190 µm high, strongly inspersed with minute spherical droplets. Paraphyses simple, dense and straight, 2-2.5 µm wide, not thickened at the apex. Epihymenium pale brown to pale greenish brown, 10-15 µm high. Asci broadly clavate, c. $140-155 \times 28-39 \,\mu\text{m}$, without a well-developed tholus when mature. Ascospores 1/ascus, hyaline, densely muriform, halonate in young stages, 105- $112 \times 22-28 \,\mu\text{m}$, with rounded ends, I – (see remarks!), spore development microcephalic.

Pycnidia not seen.

Chemistry. K - , C - , PD - ; material too scanty for TLC.

Etymology. The specific epithet refers to the type locality in the state of Merida in Venezuela.

Remarks. The inspersed hymenium and the brown pigmentation of the proper exciple of Topeliopsis meridensis are unique in the genus. These features separate the new species from the similar Topeliopsis muscigena. Topeliopsis elixii, the only other Topeliopsis species with a single muriform ascospore per ascus, differs by the finally flat, pruinose disc and by its chemistry, i.e. the presence of hypostictic and hypoconstictic acids. The type material of Topeliopsis meridensis is scanty and the ascospores appear to be not fully mature. This might explain the lack of amyloidity of the endospore, which is otherwise characteristic of the genus.

Habitat and distribution. This new species is known only from the type locality where it was found growing sparsely over mosses at the base of a deciduous tree, in a moist, light paramo-vegetation along a creek. This

ecology agrees well with that of the other species in the genus.

The New Combinations

Topeliopsis subdenticulata (Zahlbr.) A. Frisch & Kalb comb. nov.

Basionym: Ocellularia subdenticulata Zahlbr., in Skottsberg, C. (ed.), The Natural History of Juan Fernandez and the Easter Island, vol. 2, Botany: 329 (1924); type: Juan Fernandez Islands, Masafuera, "nördlicher Teil des hohen Kammes, auf bemoosten Dicksonia-Stämmen", C. & I. Skottsberg (W—holotype).

Topeliopsis vezdae Kalb, Mycotaxon 79: 323 (2001); type: Australia, Queensland, Styx River State Forest, c. 68 km E of Armidale, cool temperate rainforest along a rivulet, 30°34′S, 152°13′E, 800 m alt., 11 viii 1988, K. Kalb & J. Williams 19199 (CANB—holotype).

Ocellularia patagonica Imshaug ined.; Chile, Magallanes, Isla Desolacion, Hebe scrub along S shore of Caleta San José, Bahia Tuesday, 52°51′S, 74°28′W, 04 x 1969, H. Imshaug & K. Ohlsson 44750 (H).

(Fig. 1G)

Topeliopsis decorticans (Müll. Arg.) A. Frisch & Kalb comb. nov.

Basionym: *Thelotrema decorticans* Müll. Arg., *Bull. Herb. Boissier* 1: 54 (1893); type: Australia, Victoria, Black Spur, 1892, *F. R. M. Wilson* 514 (G!—holotype).

Topeliopsis corticola Kalb, Mycotaxon 79: 322 (2001); type: Australia, Blue Mountains National Park, Mount Wilson, Chimney Cottage, in a moist cool temperate rainforest, 33°29′S, 152°25′E, 1000 m alt., 10 viii 1988, K. Kalb & J. Williams 21227 (CANB—holotype).

(Fig. 1C)

Ascoconidia

Pycnidia have not been reported from any *Topeliopsis* species so far. However, ascoconidia are produced in old ascospores of *Topeliopsis elixii* (and of *T. muscigena*; G. Kantvilas, pers. comm.), a type of conidiogenesis that has already been described for several lichenized ascomycetes (Hafellner & Bellemère 1983; Santesson 1952). Ascospore development and conidiogenesis could not be studied in all details because of the lack of adequate material, and only some

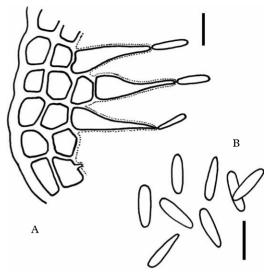


FIG. 6. Ascoconidia in *Topeliopsis elixii* (holotype). A, transverse section of old ascospore showing 2–3 layers of ascospore cells and ascoconidia; B, ascoconidia. Scales: A & $B=5~\mu m$.

major observations are recorded here. The microcephalic ascospores of Topelipsis elixii are produced singly from the beginning and are already muriform in the earliest stages observed. A large cavity is formed in some of the older ascospores by the separation and disintegration of the central cells. This cavity is bordered by 2-3 layers of ascospore cells. Ascoconidia are formed from the innermost circle of cells that borders the central cavity, into which they are finally released. The ascoconidia are produced acrogenously on bottle-shaped phialides, c. 5–13 \times 1·5–2 μ m, which appear first as small papillae and later grow out to their normal size. Using light microscopy, it could not be demonstrated without doubt whether the phialides are separated from the ascospore cells by a transverse septum or not. However, the former seems to be the case. Therefore, the conidiophores are of Vobis' types I or II (Vobis 1980). The ascoconidia are 4.5– $6 \times 1.2 - 1.5$ µm in size and rather irregularly bacilliform to narrow elliptical or claviform (Fig. 6A & B).

The conidiogenesis of *Topeliopsis elixii* differs from that described in Hafellner &

Bellemère (1983) for *Brigantiaea leucoxantha* mainly in the formation of distinct conidiogenous cells, whereas the ascoconidia of *B. leucoxantha* are produced on small papillae which are formed pleurogenously on the isolated central ascospore cells. Furthermore, the production of conidia in *B. leucoxantha* was described as occurring on all central ascospore cells, whereas in *Topeliopsis elixii* it is restricted to the cell layer bordering the central cavity.

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