

Rhythmic low-field magnetic stimulation may improve depression by increasing brain-derived neurotrophic factor

Le Xiao,¹ Christoph U. Correll,² Lei Feng,¹ Yu-Tao Xiang,³ Yuan Feng,¹ Chang-Qing Hu,¹
Rena Li,^{1,4} and Gang Wang^{1*}

¹ The National Clinical Research Center for Mental Disorders and Beijing Key Laboratory of Mental Disorders, Beijing An Ding Hospital, Capital Medical University, Beijing, China

² Department of Psychiatry, The Zucker Hillside Hospital, Northwell Health, Glen Oaks, New York, United States

³ Unit of Psychiatry, Faculty of Health Sciences, University of Macau, Macao SAR, China

⁴ Beijing Institute for Brain Disorders, Capital Medical University, Beijing, China

Background. Low-field magnetic stimulation (LFMS) has mood-elevating effect, and the increase of brain-derived neurotrophic factor (BDNF) is associated with antidepressant treatment. We evaluated the effects and association with BDNF of rhythmic LFMS in the treatment of major depressive disorder (MDD).

Methods. A total of 22 MDD patients were randomized to rhythmic alpha stimulation (RAS) or rhythmic delta stimulation (RDS), with 5 sessions per week, lasting for 6 weeks. Outcomes assessments included the 17-item Hamilton Depression Rating Scale (HAMD–17), the Hamilton Anxiety Rating Scale (HAMA), and the Clinical Global Impressions–Severity scale (CGI–S) at baseline and at weeks 1, 2, 3, 4, and 6. Serum BDNF level was measured at baseline and at weeks 2, 4, and 6.

Results. HAMD–17, HAMA, and CGI–S scores were significantly reduced with both RAS and RDS. RAS patients had numerically greater reductions in HAMD–17 scores than RDS patients (8.9 ± 7.4 vs. 6.2 ± 6.2 , effect size [ES] = 0.40), while RDS patients had greater improvement in HAMA scores (8.2 ± 8.0 vs. 5.3 ± 5.8 , ES = 0.42). RAS was associated with clinically relevant advantages in response (54.5% vs. 18.2%, number-needed-to-treat [NNT] = 3) and remission (36.4% vs. 9.1%, NNT = 4). BDNF increased significantly during the 6-week study period ($p < 0.05$), with greater increases in RAS at weeks 4 and 6 (ES = 0.66–0.76) and statistical superiority at week 2 ($p = 0.034$, ES = 1.23). Baseline BDNF in the 8 responders (24.8 ± 9.0 ng/ml) was lower than in the 14 nonresponders (31.1 ± 7.3 ng/ml, $p = 0.083$, ES = –0.79), and BDNF increased more in responders (8.9 ± 7.8 ng/ml) than in nonresponders (1.8 ± 3.5 ng/ml, $p = 0.044$). The change in BDNF at week 2 was the most strongly predicted response ($p = 0.016$).

Conclusions. Rhythmic LFMS was effective for MDD. BDNF may moderate/mediate the efficacy of LFMS.

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Key words: Major depressive disorder, rhythmic magnetic stimulation, brain-derived neurotrophic factor, moderator, mediator.

* Address for correspondence: Dr. Gang Wang, Mood Disorders Center, Beijing An Ding Hospital, No. 5 Ankang Lane, Deshengmenwai Avenue, Xicheng District, Beijing 100088, China.
(Email: gangwangdoc@gmail.com)

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Introduction

Low-field magnetic stimulation (LFMS) is a noninvasive neuromodulation technique. LFMS may stimulate the brain by emitting electromagnetic fields (similar to those produced by echo planar magnetic resonance imaging [MRI]) and could induce rapid mood elevation in bipolar depressive patients.^{1,2} The antidepressant-like effects of LFMS were also demonstrated by reducing immobility in the forced-swim test of the depressive animal model.³ Although previous studies demonstrated rapid

improvement in mood after a single LFMS intervention, the durability of these effects and the optimized stimulation parameters also need to be determined by further study.⁴

Magnetic stimulation is hypothesized to have antidepressant effects by resetting cortical oscillatory activity and reestablishing intrinsic cerebral rhythms.⁵ Based on the rationale and supportive findings of low-field synchronized transcranial magnetic stimulation in patients with major depressive disorder (MDD),⁶ deep-brain magnetic stimulation (DMS) has been developed with diverse rhythmic stimulations and was demonstrated to be effective for neuropsychiatric disorders, including depression.⁷ It has been reported that DMS as monotherapy or augmentation may improve the cognitive impairments in patients with Alzheimer's disease.⁸ On the molecular and cellular levels, DMS has been shown to greatly facilitate adult hippocampal neurogenesis and maturation, and to alleviate depression and stress-related responses in animal models.⁹ LFMS/DMS does not need to be orientated to specific brain regions prior to treatment due to its uniform, unidirectional, and pulsed magnetic field, which is hypothesized to have diffusive effects on neuronal regions.³

DMS seems to be a promising alternative treatment for MDD. However, there are still many uncertainties that require further investigation. For example, the role of the dorsal lateral prefrontal cortices (DLPFCs) in depression is asymmetric, with relative hypoactivity in the left DLPFC, along with relative hyperactivity in the right.¹⁰ High-frequency stimulation to the left DLPFC and low-frequency stimulation to the right DLPFC have both been proved effective in the treatment of MDD.^{11,12} A wide range of stimulation frequencies has been shown to modulate brain function, and the alpha band has been specifically focused.^{5,13,14} Alpha band power is involved in modulating connections among the dorsal anterior cingulate cortex, anterior insula, anterior prefrontal cortex, and thalamus.¹⁵ Our previous pilot study has proven that rhythmic alpha stimulation (RAS, 8–12 Hz) of DMS improved the depressive, anxiety, and sleep symptoms in treatment-resistant MDD patients, and as well increased the serum level of brain-derived neurotrophic factor (BDNF).¹⁶ Two recent randomized sham-controlled studies investigated the antidepressant efficacy of low-field synchronized alpha frequency stimulation for MDD and found that alpha stimulation greatly improved depression compared to sham.^{17,6} In Rohan's two studies of LFMS in the treatment of bipolar or unipolar depression,^{1,2} the 0.5-Hz paradigm, which is regarded as low-frequency delta stimulation, has been proved significantly effective compared to sham. However, there is still a lack of randomized and head-to-head studies that compare the antidepressant efficacy of LFMS by different rhythms.

Given the heterogeneity of the therapeutic effects in patients with MDD, it will be important to identify clinically relevant moderators and mediators of treatment response. Most repetitive transcranial magnetic stimulation (rTMS) studies focused on BDNF as a potential moderator or mediator variable. BDNF is a critical prosurvival factor for the developing and adult central nervous system, acting through modulation of activity-induced neuronal plasticity,¹⁸ which has been repeatedly implicated in the etiology of depression. Moreover, the established antidepressant properties of BDNF led to elaboration of the neurotrophic hypothesis of depression.¹⁹ Finally, BDNF expression and signaling have been related to the efficacy of antidepressant treatments.^{20,21}

Based on the above, we aimed to evaluate: (1) the efficacy of DMS with two different rhythms in MDD, (2) the effect of DMS on BDNF, and (3) the relationship between baseline BDNF and BDNF change with antidepressant effects. Since serum BDNF levels in depressed patients are reduced compared to healthy controls and are normalized after antidepressant treatment,^{22,23} we hypothesized that the efficacy of DMS would be moderated by low baseline BDNF and mediated by treatment-related increases in BDNF.

Methods

Design

This was a randomized, double-blinded, active-controlled trial comparing rhythmic alpha stimulation (RAS) and rhythmic delta stimulation (RDS) that was conducted between May of 2010 and January of 2011 at the Beijing Anding Hospital. The study protocol was approved by the Human Research and Ethics Committee of Beijing Anding Hospital and was registered (clinicaltrials.gov, no. NCT02184221). All participants provided written informed consent.

Subjects

The inclusion criteria were as follows: (1) males and females; (2) aged 18–60 years; (3) right-handed; (4) out- or inpatient; (5) single or recurrent moderate or severe DSM-IV diagnosis of MDD; (6) 17-item Hamilton Rating Scale for Depression (HAM-D-17)^{24,25} total score ≥ 18 ; and (7) medication-free for ≥ 5 days before the trial. Patients who took psychotropic medications entered a washout phase that lasted for at least 7 days, during which psychotropic medications were tapered and stopped.

The exclusion criteria were as follows: (1) current diagnosis of DSM-IV axis I primary psychiatric illness other than MDD; (2) clinically significant medical diseases, including any cardiovascular, hepatic, renal,

respiratory, hematologic, endocrinological, or neurologic diseases; (3) organic mental disease; (4) clinically significant laboratory abnormality that was not stabilized or that was anticipated to require treatment during the study; (5) pregnant or lactating women, as well as females of childbearing potential without appropriate birth control measures; (6) treatment with electroconvulsive therapy within 12 months prior to screening; (7) significant risk of suicidal or self-harm behaviors; (8) cardiac pacemakers or implanted medical objects (other than dental work) within or near the head that could not be safely removed; and (9) treatment with fluoxetine during the current episode or had long-acting psychotropic medication/injection within the previous 6 months.

Interventions

Eligible patients were blindly randomized to 6 weeks of either RAS or RDS using a 1:1 ratio based on a computer-generated random number table. Treatment sessions were administered by two trained nurses for 20 minutes at a time, with 5 sessions per week, totaling 30 sessions over 6 weeks.

Only non-benzodiazepine short-acting hypnotics—including zolpidem, zopiclone, and zaleplon—were allowed for insomnia during the study period, and for a maximum of <7 days consecutively. Other medications for physical conditions with no effects on the central nervous system were allowed.

DMS device

The DMS device (model ADS-L-1, Beijing Aldans Biotechnology, Beijing, China; certified by the Beijing medical device quality supervision and inspection center at the State Food and Drug Administration inspection center and met the GB9706.1 standard; for investigational use only), included a pair of coils with a diameter of 360 mm and a distance of 300 mm between coils

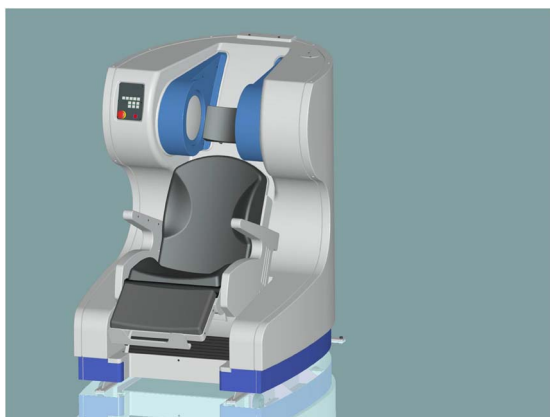
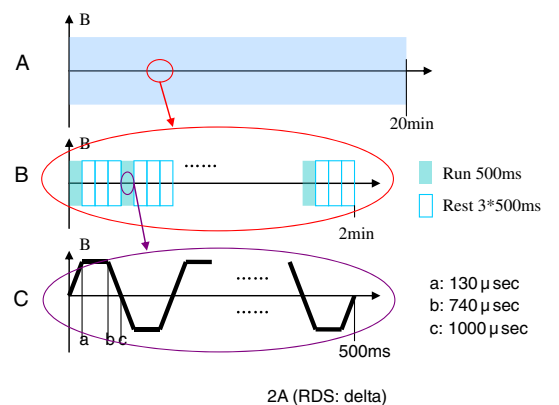


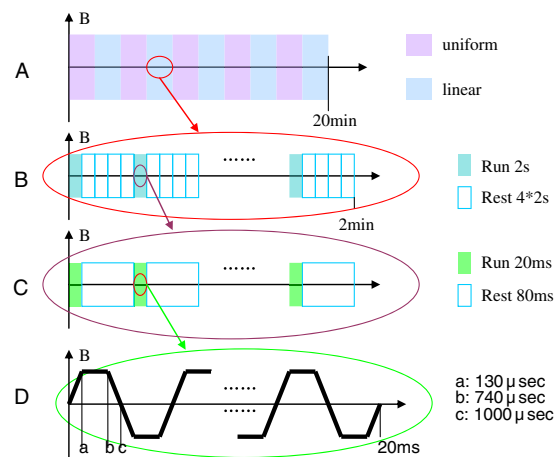
FIGURE 1. Deep-brain magnetic stimulation device.

(Figure 1). The magnetic field generator can be discharged through two coils to produce a time-varying pulsed magnetic field. A linear gradient magnetic field is generated when the reverse current flows through the coils; conversely, a uniform magnetic field is generated when current flows in the same direction. The two coils was symmetrically positioned on both sides of the patient's head to provide a global magnetic field distributed broadly across the whole brain region. The waveform was a 1000-Hz train of alternating trapezoidal gradient pulses, and the maximum magnetic field intensity acting on the brain regions was 20 Gs. The time-varying rate was 150 Gs/ms. Notably, the strength of DMS fields are 100 to 1,000 times weaker than regular rTMS fields, penetrate throughout the whole brain, and are delivered at 1 kHz.

The delta stimulating paradigm (Program 1) is shown in Figure 2A. In this paradigm, the magnetic flux density was a linear gradient, divided into 10 2-minute outputs (Figure 2A-A). Each 2-minute output consisted of a 500 ms long RUN followed by a 1500 ms long REST and repeats (Figure 2A-B). Each RUN consisted of 500 alternating trapezoids, each lasting 1000 μ sec



2A (RDS: delta)



2B (RAS: alpha)

FIGURE 2. Schematic illustration of the DMS paradigm.

(Figure 2A-C). The alpha-stimulating paradigm (Program 2) is shown in Figure 2B. Different from Program 1, the magnetic flux density of Program 2 was a uniform alternating linear gradient. In this program, each 20-minute treatment consisted of 10 2-minute long microcycles (Figure 2B-A). Each microcycle started with a 2-second long RUN, followed by an 8-second long REST and repeats (Figure 2B-B). Each RUN generated 20 1-ms alternating trapezoid pulses at 8–12 Hz (Figure 2B-C). Figure 2B-D shows the microstructure of the 20-ms output pulses.

Assessments

Efficacy outcomes were assessed with the HAMD–17, the Hamilton Rating Scale for Anxiety (HAMA),²⁶ and the Clinical Global Impression–Severity (CGI–S)²⁷ at baseline and at weeks 1, 2, 3, 4, and 6. Three certified raters were blinded to study assignment. Safety outcomes in this study were assessed by adverse event reporting, clinical laboratory measurements, and physical examination. Treatment-emergent adverse events, vital signs, and body weight were monitored at each visit. Laboratory tests—including clinical chemistry, urinalysis and hematology panels, electrocardiogram (EKG), and electroencephalogram (EEG)—were recorded at baseline and at the 6-week endpoint or discontinuation visit.

BDNF measurement

At baseline and weeks 2, 4, and 6, venous blood (5 ml) was collected in anticoagulant-free tubes between 8:00 and 10:00 AM at fasting state of at least 10 hours, and were allowed to clot at room temperature for 60 min followed by centrifugation at 2000 rpm for 10 min at 4°C. Serum samples were stored at –80°C for tests. Serum BDNF was measured using an enzyme-linked immunosorbent assay (ELISA) kit according to the manufacturer's instructions. Absorbencies were measured by a Varioskan Flash Multimode Microplate Reader (Thermo Labsystems, Helsinki, Finland). All assays were conducted at the same time.

Outcome measures

The primary outcome was treatment response, defined as a reduction in HAMD–17 score of $\geq 50\%$ ²⁸ from baseline to week 6. Remission was defined as a HAMD–17 total score ≤ 7 at endpoint. Additional efficacy outcomes included change in HAMD–17, HAMA, and CGI–S scores. Further outcomes included BDNF levels and safety, as well as tolerability outcomes.

Statistical analysis

The analyses were conducted in the modified intent-to-treat sample (patients with a baseline and at least one

follow-up assessment). The Shapiro–Wilk test was used to check the normality of data. The data were analyzed using SPSS (v. 20.0, SPSS Inc., Chicago, Illinois, USA). Demographic and clinical characteristics at baseline were compared between the two groups using an independent sample *t* test, the Mann–Whitney *U* test, and the chi-square test, as appropriate.

The change from baseline over the entire 6-week study period within and across groups was analyzed for all outcomes by repeated measures analysis of variance (ANOVA). When Mauchly's sphericity test was significant, we adjusted the degrees of freedom and *F* ratios using the Greenhouse–Geisser adjustment.²⁹ The change from baseline was modeled as an effect of treatment group (treatment), treatment progression (visit), and the treatment \times visit interaction as fixed effects. In addition, last observation carried forward (LOCF) ANOVA analyses were conducted across groups for change from baseline to each individual study visit. Since the two treatment groups differed significantly on illness duration, the baseline-to-endpoint analyses across groups were also repeated covarying for illness duration using logistic regression analyses for response and remission.

Furthermore, pooling results from both treatment conditions together, the *t* test was used to compare the difference between responders and nonresponders regarding baseline BDNF and mean baseline-to-endpoint change of BDNF. Spearman's correlation coefficient was calculated between the change of HAMD–17 total score and BDNF level from baseline to week 6. Finally, in order to examine significant correlates of response and remission across both treatment conditions, multivariate logistic regression was carried out with response and remission as the dependent variables separately, and with treatment, age, sex, number of depressive episodes, illness duration, duration of the current episode, baseline HAMD–17 and HAMA scores, baseline BDNF, and change in change as independent variables.

All statistical tests were two-sided, with $\alpha = 0.05$. Since a type II error is likely in small samples with insufficient power to differentiate effects between two active treatments, and to provide estimates of the magnitude of observed effects in this pilot study, we also calculated effect sizes with their 95% confidence intervals ($CI_{95\%}$) for all group contrasts, even if between-group results were not statistically significant. Specifically, for categorical outcomes, we calculated numbers-needed-to-treat (*NNT*) by dividing 1 by the risk difference. In general, *NNT*s of 10 or less are considered clinically relevant.³⁰ For continuous outcomes, we calculated Cohen's *d* by dividing the difference in means by the pooled standard deviation. According to Cohen,³¹ 0.2 represents a small effect size, 0.5 a medium effect size, and 0.8 a large effect size.

Results

A total of 26 patients were screened, 22 of whom met the study criteria and were randomized to RAS ($n=11$) or RDS ($n=11$). Since all patients had at least one follow-up visit, all 22 randomized patients were included in the analyses. 15 patients had failed to respond to at least two antidepressant treatments. Patients were treated with sertraline, citalopram, escitalopram, paloxetine, venlafaxine, or mirtazapine before entering the study.

No significant differences were found between groups regarding demographic and clinical characteristics, including baseline HAMD-17, HAMA, and CGI-S scores; and BDNF level at baseline; except for a significantly longer duration of illness in the RAS vs. RDS group ($p=0.040$) (Table 1).

Efficacy outcomes

In the RAS group, 54.5% patients (6/11) responded at endpoint compared to only 2 in the RDS group (18.2%), but the difference did not reach statistical significance (adjusted $p=0.108$, $NNT=3$). There was also no significant difference in remission between the RAS (4/11, 36.4%) and RDS groups (1/11, 9.1%; adjusted $p=0.147$, $NNT=4$) (Table 2).

The improvement in HAMD-17 score was significant from baseline over the entire 6-week study period (Greenhouse–Geisser adjusted $F(1.5, 704.4)=24.94$, $p<0.001$). No significant difference was found between the RAS and RDS groups overall ($F(1, 182.01)=1.45$, $p=0.24$), and without significant time-by-treatment interaction (Greenhouse–Geisser adjusted $F(1.5, 21.93)=0.78$, $p=0.44$). The mean reduction in HAMD-17 score in the RAS group was numerically greater than that in the RDS group (8.9 ± 7.4 vs. 6.2 ± 6.2 , Cohen's $d=0.40$).

The improvement in HAMA score was also significant from baseline over the entire 6-week study period (Greenhouse–Geisser adjusted $F(2.0, 346.24)=13.67$, $p<0.001$). No significant difference was found between the RAS and RDS groups overall ($F(1, 112.76)=1.06$, $p=0.32$), and without significant time-by-treatment interaction (Greenhouse–Geisser adjusted $F(2.0, 17.16)=0.68$, $p=0.52$). However, different from the HAMD-17 results, RDS was associated with a numerically better improvement in HAMA change than RAS (8.2 ± 8.0 vs. 5.3 ± 5.8 , Cohen's $d=0.42$).

The improvement in CGI-S score was also significant from baseline over the entire 6-week study period

TABLE 1. Baseline demographic and clinical characteristics

| Characteristics | Total | | RAS ($n=11$) | | RDS ($n=11$) | | Statistics | |
|--|-------|------|----------------|------|----------------|------|-------------------|-------|
| | n | % | n | % | n | % | χ^2 | p |
| Male | 8 | 36.4 | 6 | 54.5 | 2 | 18.2 | — ^a | 0.183 |
| First episode | 6 | 27.3 | 2 | 18.2 | 4 | 36.4 | — ^a | 0.243 |
| Treatment resistance | 15 | 68.2 | 8 | 72.7 | 7 | 63.6 | — ^a | 0.319 |
| | Mean | SD | Mean | SD | Mean | SD | t | p |
| Age, years | 34.2 | 12.0 | 32.8 | 13.1 | 35.6 | 11.3 | -0.54 | 0.594 |
| Number of depressive episodes | 3.0 | 2.0 | 3.3 | 2.3 | 2.6 | 1.6 | 0.85 | 0.406 |
| Duration of illness, months | 60.2 | 76.6 | 72.5 | 81.2 | 48.0 | 73.5 | 2.07 ^b | 0.040 |
| Duration of current depressive episode, months | 8.5 | 13.5 | 10.6 | 17.5 | 6.4 | 8.1 | 0.06 ^b | 0.949 |
| HAMD-17 total | 22.1 | 3.1 | 21.6 | 3.3 | 22.9 | 2.8 | -0.98 | 0.339 |
| HAMA total | 16.7 | 5.2 | 15.3 | 3.4 | 18.1 | 6.3 | -1.30 | 0.208 |
| CGI-S total | 4.0 | 0.6 | 3.8 | 0.6 | 4.2 | 0.6 | 1.41 | 0.173 |
| BDNF, ng/ml | 28.8 | 8.3 | 27.1 | 8.7 | 30.6 | 7.9 | 0.98 | 0.338 |

a = Fisher's exact test; b = Mann–Whitney U test; BDNF = brain-derived neurotrophic factor; CGI-S = Clinical Global Impression–Severity; HAMA = Hamilton Rating Scale for Anxiety; HAMD-17 = 17-item Hamilton Rating Scale for Depression; RAS = rhythmic alpha stimulation; RDS = rhythmic delta stimulation; SD = standard deviation.

TABLE 2. Primary outcome measures of clinical efficacy from baseline to endpoint

| Outcome | RAS ($n=11$) | | RDS ($n=11$) | | Between-group comparison | | | |
|-----------|----------------|------|----------------|------|--------------------------|--------------|-------|--------------|
| | n | % | n | % | p | Adjusted p | NNT | $CI_{95\%}$ |
| Response | 6 | 54.5 | 2 | 18.2 | 0.183 | 0.108 | 3 | -0.86, 73.58 |
| Remission | 4 | 36.4 | 1 | 9.1 | 0.311 | 0.147 | 4 | -5.84, 60.39 |

$CI_{95\%}$ = 95% confidence interval; NNT = number needed to treat; RAS = rhythmic alpha stimulation; RDS = rhythmic delta stimulation.

TABLE 3. Change in HAMD-17, HAMA, and CGI-S scores at each postbaseline visit

| Visit | Variable | RAS (n = 11) | | RDS (n = 11) | | Between-group comparison | | | |
|--------|----------|--------------|-----|--------------|-----|--------------------------|-------|------------|-------------------|
| | | Mean | SD | Mean | SD | F | p | Cohen's d* | CI _{95%} |
| Week 1 | HAMD-17 | 0.1 | 0.8 | 0.3 | 0.8 | 0.278 | 0.604 | -0.25 | -1.08, 0.60 |
| | HAMA | 1.6 | 5.2 | 1.6 | 6.3 | 0.001 | 0.971 | 0.00 | -0.84, 0.84 |
| | CGI-S | 0.1 | 0.8 | 0.4 | 0.7 | 0.714 | 0.408 | -0.40 | -1.23, 0.46 |
| Week 2 | HAMD-17 | 3.9 | 3.7 | 3.1 | 2.5 | 0.361 | 0.555 | 0.25 | -0.60, 1.08 |
| | HAMA | 3.4 | 5.2 | 3.6 | 5.9 | 0.013 | 0.910 | -0.04 | -0.87, 0.80 |
| | CGI-S | 0.5 | 1.1 | 0.6 | 0.5 | 0.059 | 0.811 | -0.12 | -0.95, 0.72 |
| Week 3 | HAMD-17 | 5.7 | 4.2 | 4.2 | 3.1 | 0.959 | 0.339 | 0.41 | -0.45, 1.23 |
| | HAMA | 4.3 | 5.5 | 5.3 | 7.2 | 0.133 | 0.719 | -0.16 | -0.99, 0.69 |
| | CGI-S | 0.6 | 1.1 | 0.7 | 0.8 | 0.049 | 0.828 | -0.10 | -0.94, 0.74 |
| Week 4 | HAMD-17 | 6.8 | 5.0 | 5.3 | 4.3 | 0.600 | 0.448 | 0.32 | -0.53, 1.15 |
| | HAMA | 4.7 | 5.4 | 6.3 | 8.0 | 0.279 | 0.603 | -0.23 | -1.06, 0.61 |
| | CGI-S | 0.6 | 1.0 | 1.2 | 1.2 | 1.828 | 0.191 | -0.54 | -1.37, 0.33 |
| Week 6 | HAMD-17 | 8.9 | 7.4 | 6.2 | 6.2 | 0.879 | 0.360 | 0.40 | -0.46, 1.22 |
| | HAMA | 5.3 | 5.8 | 8.2 | 8.0 | 0.948 | 0.342 | -0.42 | -1.24, 0.44 |
| | CGI-S | 0.6 | 1.3 | 1.5 | 1.1 | 3.106 | 0.093 | -0.75 | -1.58, 0.18 |

* Positive Cohen's *d* indicates superiority of RAS over RDS.
 CGI-S = Clinical Global Impression-Severity; CI_{95%} = 95% confidence interval; HAMA = Hamilton Rating Scale for Anxiety; HAMD-17 = 17-item Hamilton Rating Scale for Depression; RAS = rhythmic alpha stimulation; RDS = rhythmic delta stimulation; SD = standard deviation.

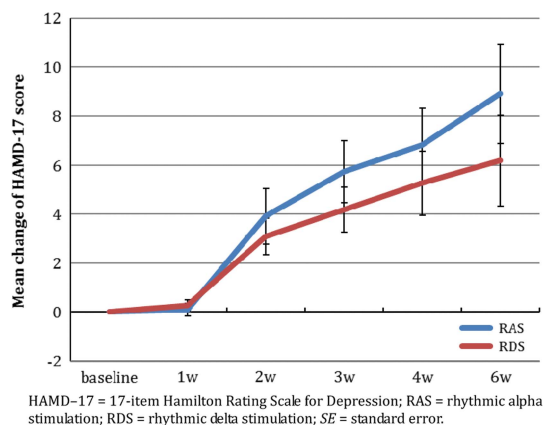


FIGURE 3a. Mean change of HAMD-17 total score over 6 weeks (mean ± SE).

(Greenhouse–Geisser adjusted $F(2.7, 6.57) = 10.65$, $p < 0.001$). Again, no significant difference was found between RAS and RDS overall ($F(1, 0.03) = 0.01$, $p = 0.92$), and without significant time-by-treatment interaction (Greenhouse–Geisser adjusted $F(2.7, 1.32) = 2.14$, $p = 0.11$). RDS had numerically greater improvement on the CGI-S than RAS (1.6 ± 1.2 vs. 0.6 ± 1.3 , Cohen's $d = 0.75$).

The HAMD-17, HAMA, and CGI-S score changes at each postbaseline visit are shown in Table 3, and the mean changes in HAMD-17 and HAMA scores over the 6-week period are shown in Figures 3a and 3b.

BDNF outcomes

The change in BDNF levels at each postbaseline visit over the 6-week period are shown in Table 4 and Figure 3c.

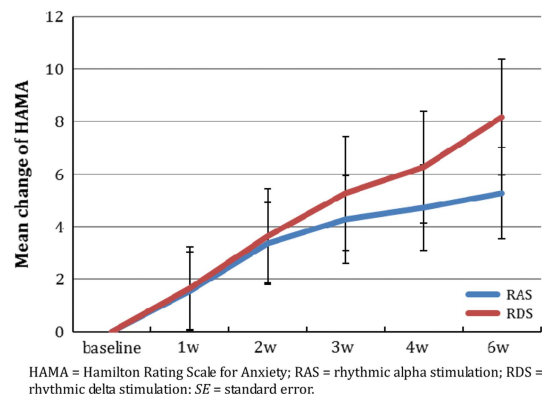


FIGURE 3b. Mean change of HAMA total score over 6 weeks (mean ± SE).

BDNF levels increased significantly after treatment with RAS, while it fluctuated with RDS (Figure 3c). The increase in BDNF was significant from baseline over the entire 6-week study period ($F(3, 99.01) = 4.09$, $p = 0.013$), but without significant time-by-treatment interaction ($F(3, 40.98) = 1.69$, $p = 0.184$) and without a significant group difference ($F(1, 0.90) = 0.003$, $p = 0.957$). The mean BDNF change was consistently numerically greater in the RAS than in the RDS group at each postbaseline study timepoint, including endpoint (8.0 ± 6.1 vs. 3.5 ± 7.5), but the difference between groups was only significant at week 2 ($p = 0.034$), and the effect sizes favoring RAS were moderate (Cohen's $d = 0.66$) to large (Cohen's $d = 1.23$).

Factors associated with treatment response

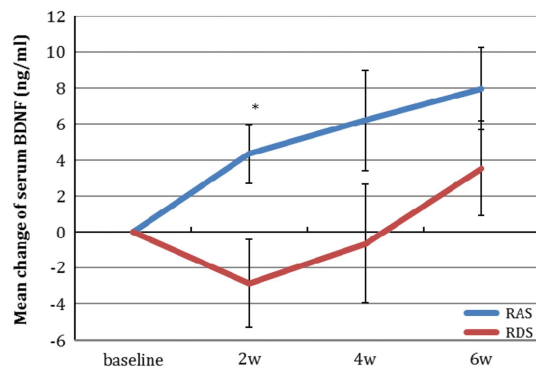
Baseline BDNF in the 8 responders was significantly lower than in the 14 nonresponders (24.8 ± 9.0 ng/ml vs.

TABLE 4. Change in BDNF levels from baseline to endpoint (ng/ml)

| Visit | RAS | | RDS | | Between-group comparison | | | |
|--------|------|-----|------|------|--------------------------|-------|--------------------|--------------------------|
| | Mean | SD | Mean | SD | F | p | Cohen's <i>d</i> * | <i>CI</i> _{95%} |
| Week 2 | 4.3 | 4.4 | -2.8 | 6.9 | 5.584 | 0.034 | 1.23 | 0.27, 2.09 |
| Week 4 | 6.2 | 7.4 | -0.6 | 10.2 | 2.152 | 0.166 | 0.76 | -0.13, 1.60 |
| Week 6 | 8.0 | 6.1 | 3.5 | 7.5 | 1.540 | 0.237 | 0.66 | -0.22, 1.49 |

* Positive Cohen's *d* indicates superiority of RAS over RDS.

BDNF = brain-derived neurotrophic factor; *CI*_{95%} = 95% confidence interval; RAS = rhythmic alpha stimulation; RDS = rhythmic delta stimulation; *SD* = standard deviation.



* $p < 0.05$.

BDNF = brain-derived neurotrophic factor; RAS = rhythmic alpha stimulation; RDS = rhythmic delta stimulation; *SE* = standard error.

FIGURE 3c. Mean change of BDNF level over 6 weeks (mean \pm SE).

31.1 ± 7.3 ng/ml, $df = 20$, $t = 1.822$, $p = 0.083$, Cohen's $d = -0.79$). The mean BDNF increase was significantly higher in responders than in nonresponders (8.9 ± 7.8 ng/ml vs. 1.8 ± 3.5 ng/ml, $df = 13$, $t = 2.307$, $p = 0.044$, Cohen's $d = 1.17$). However, Spearman's correlation analysis showed that the BDNF change was not significantly correlated with the change in HAMD-17 score ($r = 0.32$, $p = 0.244$): RAS ($r = 0.18$, $p = 0.699$) and RDS ($r = 0.38$, $p = 0.349$). Logistic regression showed that the BDNF change at week 2 was the most important variable in terms of predicting a response ($p = 0.016$).

Safety outcomes

DMS was well tolerated, and no seizures or headaches were reported throughout the study. Only one patient in the RAS group reported mild and transient fatigue during treatment (at week 1). No other spontaneously reported adverse effects occurred. Further, no abnormalities were found regarding vital signs, or laboratory, EEG, and EKG values.

Discussion

To the best of our knowledge, this is the first randomized controlled trial to investigate the effect of LFMS with two different rhythmic stimulations in the treatment of MDD.

We found that both alpha and delta stimulation significantly reduced depressive and anxiety symptoms. Although no statistically significant difference in efficacy was observed between the two groups, RAS tended to induce greater improvement in HAMD-17 scores than RDS ($ES = 0.40$), while RDS resulted in a potentially greater improvement in HAMA scores than RAS ($ES = 0.42$). The differences of antidepressant treatment response ($NNT = 3$) and remission of depression ($NNT = 4$), although not statistically significant, showed large and clinically relevant effect size advantages for RAS.

BDNF is expressed throughout the central nervous system and can cross the blood/brain barrier.³² Peripheral serum BDNF levels are highly correlated with cerebral levels in animal models.³³ Repetitive TMS enhances the expression of BDNF in both cortex and lymphocytes.³⁴ Consistent with this advantage for depression-related outcomes with RAS, BDNF increased with RAS but decreased slightly with RDS at week 2. Although group differences were only significant at week 2, favoring RAS, the effect sizes were moderate to large for greater BDNF increases in the RAS group ($ES = 0.66$ – 1.23). Some studies found that rTMS may increase serum BDNF,^{35,36} but others failed to replicate this finding.^{37,38} However, some studies suggested that high-frequency rTMS increases serum BDNF levels, whereas low-frequency stimulation reduces BDNF levels.^{34,39} Our results confirm prior findings that low-field magnetic stimulation could promote neural growth, with frequency-dependent characteristics.⁴⁰

The baseline level and change in serum BDNF levels after treatment were associated with the antidepressant efficacy of DMS, which supports the notion that BDNF may have potential use as a predictor of antidepressant efficacy.²³ Our findings indicate that DMS is more effective for depressed patients with comparatively lower BDNF levels, suggesting that low serum BDNF may be a moderator and predictive biomarker for the efficacy of DMS. The change in BDNF in responders was significantly higher than in nonresponders, and the increased BDNF at week 2 strongly predated the ultimate response at week 6. This relationship suggests that a BDNF increase is a relevant mediator of DMS activity that may be mechanistically and causally related to DMS efficacy, a finding that needs to be investigated further.

Studies have shown that LFMS significantly affects cerebral metabolism and neuronal activity⁴¹ as well as having antidepressant-like effects.^{1,3,42} Our results support the notion that low-intensity magnetic fields constitute an effective treatment for MDD, whether or not the stimulation is delivered at high or low frequency. While the antidepressant effects of RAS may be due to an elevation of BDNF levels, the mechanism of RDS remains incompletely understood. MDD involves dysfunction in a number of cortical regions, such as the dorsolateral

prefrontal cortex and anterior cingulate cortex, as well as deep gray matter structures, such as the nuclei of the thalamus and hypothalamus.^{5,43} Furthermore, MDD is increasingly understood as a disorder of connectivity in the brain networks linking these regions.⁴⁴ It is assumed that DMS may alter neural activity and induce network effects in the brain.⁹ The process of repetitive magnetic stimulation is hypothesized to reset brain oscillatory activity and connectivity, and to increase neuroplasticity, which together lead to antidepressant effects.⁵ Our results add to the growing body of literature suggesting that LFMS is effective in the treatment of MDD. Further research should investigate different parameters and durations of stimulation to determine the optimal DMS strategy. Furthermore, future studies with significantly larger samples should investigate whether different patient and illness variables predict greater treatment improvement with RAS or RDS.

There are several limitations of our study. First, the sample size was relatively small, which limits the power for group comparisons, rendering most results of this pilot study exploratory. However, we calculated effect sizes showing that, although not reaching statistical significance, most of the treatment group differences for antidepressant activity and increase in BDNF favored RAS versus RDS with effect sizes that were clinically meaningful. Second, we only compared two active treatments, and there was a lack of a sham control. However, at least, raters were blind to treatment assignment. Third, despite randomization, illness duration was significantly longer in the RAS versus the RDS group, but we adjusted all group comparisons for this variable. Fourth, peripherally measured BDNF levels are an imperfect assessment of centrally active BDNF, and previous antidepressants and withdrawal could affect baseline BDNF. Finally, this was a 6-week study, indicating that long-term effects need to be examined in future studies. Nevertheless, to our knowledge, this is the first randomized controlled trial of DMS comparing two stimulating paradigms providing preliminary evidence for the utility of DMS in treating MDD and providing relevant effect size information for the design of adequately powered studies.

Conclusion

We found that rhythmic DMS is a noninvasive and effective treatment for MDD without significant risk and a low rate of adverse events. Therefore, DMS may constitute a valuable neuromodulation treatment option for MDD. The antidepressant efficacy of rhythmic alpha frequency appeared to be larger than that of delta stimulation and was correlated with BDNF elevation. Furthermore, a low baseline serum BDNF level may be a predictive biomarker for better efficacy of rhythmic

magnetic stimulation, and the change in BDNF at week 2 may mediate efficacy. The results from our study need to be replicated in larger samples, ideally including a sham control group, and the long-term effects of rhythmic magnetic stimulation need to be assessed further.

Disclosures

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REFERENCES:

1. Rohan M, Parow A, Stoll AL, Demopoulos C, *et al.* Low-field magnetic stimulation in bipolar depression using an MRI-based stimulator. *Am J Psychiatry*. 2004; **161**(1): 93–98.
2. Rohan ML, Yamamoto RT, Ravichandran CT, *et al.* Rapid mood-elevating effects of low field magnetic stimulation in depression. *Biol Psychiatry*. 2014; **76**(3): 186–193.
3. Carlezon WA, Rohan ML, Mague SD, *et al.* Antidepressant-like effects of cranial stimulation within a low-energy magnetic field in rats. *Biol Psychiatry*. 2005; **57**(6): 571–576.
4. Shafi M, Stern AP, Pascual-Leone A. Adding low field magnetic stimulation to noninvasive electro-magnetic neuromodulatory therapies. *Biol Psychiatry*. 2014; **76**(3): 170–171.
5. Leuchter AF, Cook IA, Jin Y, Phillips B. The relationship between brain oscillatory activity and therapeutic effectiveness of transcranial magnetic stimulation in the treatment of major depressive disorder. *Front Hum Neurosci*. 2013; **7**: 37.
6. Leuchter AF, Cook IA, Feifel D, *et al.* Efficacy and safety of low-field synchronized transcranial magnetic stimulation (sTMS) for treatment of major depression. *Brain Stimul*. 2015; **8**(4): 787–794.

7. Kammer T, Spitzer M. Brain stimulation in psychiatry: methods and magnets, patients and parameters. *Curr Opin Psychiatry*. 2012; **25**(6): 535–t41.
8. Peng DT, Zhu R, Yuan XR, Zhang X. Clinical study of deep brain magnetic stimulation technique in the treatment of Alzheimer's disease [in Chinese]. *Chin J Geriatr*. 2012; **31**(11): 929–931.
9. Zhang Y, Mao RR, Chen ZF, *et al*. Deep-brain magnetic stimulation promotes adult hippocampal neurogenesis and alleviates stress-related behaviors in mouse models for neuropsychiatric disorders. *Mol Brain*. 2014; **7**: 11.
10. Garcia-Toro M, Montes JM, Talavera JA. Functional cerebral asymmetry in affective disorders: new facts contributed by transcranial magnetic stimulation. *J Affect Disord*. 2001; **66**(2–3): 103–109.
11. Burt T, Lisanby SH, Sackeim HA. Neuropsychiatric applications of transcranial magnetic stimulation: a meta analysis. *Int J Neuropsychopharmacol*. 2002; **5**(01): 73–103.
12. Gershon AA, Dannon PN, Grunhaus L. Transcranial magnetic stimulation in the treatment of depression. *Am J Psychiatry*. 2003; **160**(5): 835–845.
13. Thut G, Veniero D, Romei V, Miniussi C, Schyns P, Gross J. Rhythmic TMS causes local entrainment of natural oscillatory signatures. *Curr Biol*. 2011; **21**(14): 1176–1185.
14. Veniero D, Brignani D, Thut G, Miniussi C. Alpha-generation as basic response-signature to transcranial magnetic stimulation (TMS) targeting the human resting motor cortex: a TMS/EEG co-registration study. *Psychophysiology*. 2011; **48**(10): 1381–1389.
15. Sadaghiani S, Scheeringa R, Lehongre K, Morillon B, Giraud AL, Kleinschmidt A. Intrinsic connectivity networks, alpha oscillations, and tonic alertness: a simultaneous electroencephalography/functional magnetic resonance imaging study. *J Neurosci*. 2010; **30**(30): 10243–10250.
16. Xiao L, Feng Y, Feng L, Hu C, Zhang C, Wang G. Effects of deep-brain magnetic stimulation on brain derived neurotrophic factor in treatment-resistant depression [in Chinese]. *J Clin Psychiatry*. 2015; **25**(6): 361–364.
17. Jin Y, Phillips B. A pilot study of the use of EEG-based synchronized transcranial magnetic stimulation (sTMS) for treatment of major depression. *BMC Psychiatry*. 2014; **14**: 13.
18. Park H, Poo M. Neurotrophin regulation of neural circuit development and function. *Nat Rev Neurosci*. 2013; **14**(1): 7–23.
19. Groves JO. Is it time to reassess the BDNF hypothesis of depression? *Mol Psychiatry*. 2007; **12**(12): 1079–1088.
20. Adachi M, Barrot M, Autry AE, Theobald D, Monteggia LM. Selective loss of brain-derived neurotrophic factor in the dentate gyrus attenuates antidepressant efficacy. *Biol Psychiatry*. 2008; **63**(7): 642–649.
21. Duman RS, Monteggia LM. A neurotrophic model for stress-related mood disorders. *Biol Psychiatry*. 2006; **59**(12): 1116–1127.
22. Brunoni AR, Lopes M, Fregni F. A systematic review and meta-analysis of clinical studies on major depression and BDNF levels: implications for the role of neuroplasticity in depression. *Int J Neuropsychopharmacol*. 2008; **11**(8): 1169.
23. Sen S, Duman R, Sanacora G. Serum brain-derived neurotrophic factor, depression, and antidepressant medications: meta-analyses and implications. *Biol Psychiatry*. 2008; **64**(6): 527–532.
24. Hamilton M. A rating scale for depression. *J Neurol Neurosurg Psychiatry*. 1960; **23**: 56–62.
25. Xie GR, Shen QJ. Use of the Chinese version of the Hamilton Rating Scale for Depression in general population and patients with major depression. *Chin J Nerv Ment Dis*. 1984; **10**: 364.
26. Hamilton M. The assessment of anxiety states by rating. *Br J Psychiatry*. 1959; **32**(1): 50–55.
27. Guy W. Clinical global impressions. In: *ECDEU Assessment Manual for Psychopharmacology*. Rockville, MD: National Institute for Mental Health; 1976: 218–222.
28. Trivedi MH, Rush AJ, Wisniewski SR, *et al*. Evaluation of outcomes with citalopram for depression using measurement-based care in STAR*D: implications for clinical practice. *Am J Psychiatry*. 2006; **163**(1): 28–40.
29. Greenhouse SW, Geisser S. On methods in the analysis of profile data. *Psychometrika*. 1959; **24**(2): 95–112.
30. Citrome L. Number needed to treat: what it is and what it isn't, and why every clinician should know how to calculate it. *J Clin Psychiatry*. 2011; **72**(3): 412–413.
31. Cohen J. *Statistical Power Analysis for the Behavioral Sciences*, 2nd ed. Hillsdale, NJ: Erlbaum; 1988.
32. Pan W, Banks WA, Fasold MB, Bluth J, Kastin AJ. Transport of brain-derived neurotrophic factor across the blood-brain barrier. *Neuropharmacology*. 1998; **37**(12): 1553–1561.
33. Sartorius A, Hellweg R, Litzke J, *et al*. Correlations and discrepancies between serum and brain tissue levels of neurotrophins after electroconvulsive treatment in rats. *Pharmacopsychiatry*. 2009; **42**(6): 270–276.
34. Wang HY, Crupi D, Liu J, *et al*. Repetitive transcranial magnetic stimulation enhances BDNF–TrkB signaling in both brain and lymphocyte. *J Neurosci*. 2011; **31**(30): 11044–11054.
35. Yukimasa T, Yoshimura R, Tamagawa A, *et al*. High-frequency repetitive transcranial magnetic stimulation improves refractory depression by influencing catecholamine and brain-derived neurotrophic factors. *Pharmacopsychiatry*. 2006; **39**(2): 52–59.
36. Zanardini R, Gazzoli A, Ventriglia M, *et al*. Effect of repetitive transcranial magnetic stimulation on serum brain-derived neurotrophic factor in drug resistant depressed patients. *J Affect Disord*. 2006; **91**(1): 83–86.
37. Lang C, Schüler D. Biogenic nanoparticles: production, characterization, and application of bacterial magnetosomes. *J Phys Condens Matter*. 2006; **18**(38): S2815–S2828.
38. Gedde L, Beaudoin A, Lazowski L, du Toit R, Jokic R, Milev R. Effects of electroconvulsive therapy and repetitive transcranial magnetic stimulation on serum brain-derived neurotrophic factor levels in patients with depression. *Front Psychiatry*. 2012; **3**: 12.
39. Angelucci F, Oliviero A, Pilato F, *et al*. Transcranial magnetic stimulation and BDNF plasma levels in amyotrophic lateral sclerosis. *Neuroreport*. 2004; **15**(4): 717–720.
40. Rusovan A, Kanje M, Mild KH. The stimulatory effect of magnetic fields on regeneration of the rat sciatic nerve is frequency dependent. *Exp Neurol*. 1992; **117**(1): 81–84.
41. Volkow ND, Tomasi D, Wang GJ, *et al*. Effects of low-field magnetic stimulation on brain glucose metabolism. *NeuroImage*. 2010; **51**(2): 623–628.
42. Rokni-Yazdi H, Sotoudeh H, Akhondzadeh S, Sotoudeh E, Asadi H, Shakiba M. Antidepressant-like effect of magnetic resonance imaging-based stimulation in mice. *Prog Neuropsychopharmacol Biol Psychiatry*. 2007; **31**(2): 503–509.
43. Drevets WC, Price JL, Furey ML. Brain structural and functional abnormalities in mood disorders: implications for neurocircuitry models of depression. *Brain Struct Funct*. 2008; **213**(1–2): 93–118.
44. Leuchter AF, Cook IA, Hunter AM, Cai C, Horvath S. Resting-state quantitative electroencephalography reveals increased neurophysiologic connectivity in depression. *PLoS One*. 2012; **7**(2): e32508.