Is asexual reproduction an evolutionary dead end in lichens?

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Abstract: Classical hypotheses in lichenology predict pairs of species in which sexual lineages are ancestral and long-lived evolutionarily and that these give rise to derived, evolutionarily transient asexual lineages. Extensive phylogenetic information generated over the last 20 years regarding relationships within and among various groups of lichens makes possible an investigation of polarity and lability in reproductive mode across diverse clades. To test the long-held hypothesis of asexual reproduction as an evolutionary dead end in lichens, existing phylogenetic data from 23 studies were utilized to reconstruct gains and losses of sexual and asexual reproduction in a model-based statistical framework. Summed across all studies, between 26–44 origins of asexual reproduction from asexual ancestors (forward transitions) and 14–25 origins of sexual reproduction from asexual reproduction was concentrated in a relatively low number of clades (e.g. *Dirina*). The greater number of forward compared to reverse transitions is consistent with dogma in both lichenology and evolutionary biology, but nonetheless this study documents numerous reverse transitions, suggesting that asexual lineages represent a source for evolutionary innovation.

Key words: ancestral state reconstruction, apothecia, isidia, propagule, reproductive mode, sexual, soredia

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Introduction

Across all domains of life, there arguably exists no single class of traits with more of an impact on the origin and fate of evolutionary lineages than those related to reproductive biology (Stebbins 1957; Grant 1958; Gomez-Mestre et al. 2012; Van der Niet et al. 2014). Reproductive traits determine the trajectory of a vast array of population-level processes and scale up to explain major macroevolutionary patterns (Fisher 1941; Maynard Smith 1978; Tripp & Manos 2008; Kerr et al. 2011). Among eukaryotes, fungi exhibit some of the greatest diversity in reproductive strategies, making them excellent model organisms for understanding the evolution and diversification of reproductive systems. Whereas most lineages of eukaryotes reproduce primarily through sexual means or asexual means with short-lived sexual cycles (but see Dacks & Roger 1999; Redecker 2002; Ramesh *et al.* 2005; Asplen *et al.* 2009), a sizeable proportion of the 1.5 million species of fungi on the planet (Hawksworth 2001) reproduce both sexually and asexually during time periods commonly referred to as teleomorphic and anamorphic phases.

Lichenized fungi (hereafter lichens) constitute one-fifth of all fungi on Earth and approximately half of all ascomycetes (Honegger 1997; Jaklitsch et al. 2016). An attractive hypothesis to explain their diversity and success worldwide, including in extreme or novel environments, is their plurality of reproductive strategies. Lichens are somewhat unusual among fungi in that teleomorphic and anamorphic stages are commonly persistent throughout the duration of an individual's lifetime (rather than ephemeral or in phases; see Table 1 in Bowler & Rundel 1975) and involve such divergent morphological forms that mode of reproduction has for the last half century or so been associated with

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| Genus* | Number of Species | Number of species showing sexual reproductive strategy | Number of species showing asexual reproductive strategy |
|-----------------------------|----------------------|--|---|
| Acarospora A. Massal. | 61 | 60 | 1 |
| Arthonia Ach. | 151 | 151 | 0 |
| Caloplaca Th. Fr. | 167 | 130 | 33 |
| Cladonia P. Browne | 172 | 87 | 18 |
| Heterodermia Trevis. | 30 | 14 | 16 |
| Hypogymnia (Nyl.) Nyl. | 35 | 18 | 17 |
| Lecidea Ach. | 135 | 132 | 3 |
| Lepraria Ach. | 38 | 0 | 38 |
| Parmotrema A. Massal. | 46 | 10 | 36 |
| Physcia (Schreb.) Michx. | 36 | 11 | 25 |
| Ramalina Ach. | 42 | 29 | 13 |
| Rhizocarpon DC. | 72 | 72 | 0 |
| Xanthoparmelia (Vain.) Hale | 85 | 49 | 34 |

 TABLE 1. An example of reproductive strategies among genera of lichens in North America. Data derive

 from unpublished research (E. Tripp & J. Lendemer, unpublished data), using version 17 of Esslinger's

 North American Lichen Checklist as a basis for taxon identification and enumeration.

*Taxonomic authorities follow Esslinger (2015).

identification of species (recently discussed in Brodo & Lendemer 2012). Diverse reproductive strategies in these organisms must in part be related to added complexity associated with being lichenized and may be driven by selection to maintain and enrich the symbiosis (see mechanisms discussed in Kroken & Taylor 2001a and Buschbom & Mueller 2005). Across lichen trees of life (e.g. Arnold et al. 2009; Miadlikowska et al. 2014), individual species to whole clades of species are obligate sexual (e.g. Toninia A. Massal.; Timdal 1991), obligate asexual (Lepraria Ach.; Lendemer 2013), almost entirely asexual with a few sexual species (e.g. Chrysothrix Mont.), almost entirely sexual with a few asexual species (e.g. Rinodina (Ach.) Gray; Sheard 2010) or more uncommonly, facultative asexual and/or sexual (e.g. Roccella galapagoensis Follmann; Tehler et al. 2009; Lobaria pulmonaria (L.) Hoffm.; Zoller et al. 1999). North America, as one example, is home to genera with considerable variation in ratios of sexual to asexual species: some lineages have no variation in mode of reproduction whereas others approximate to a 50:50 ratio (Table 1).

A given lineage of lichens such as *Porpidia* Körber (Buschbom & Barker 2006), *Letharia* (Th. Fr.) Zahlbr. (Kroken & Taylor 2001b; Altermann *et al.* 2014), or *Dirina* (Tehler *et al.* 2013) commonly contains close

relatives marked by either sexual or asexual reproduction. This suggests either a potentially strong correlation between mating system evolution and speciation in lichens, or widespread inaccuracy in taxonomic systems that have been in place for the last half century. We now know the latter to be untrue in numerous cases because studies have demonstrated strongly supported reciprocal monophyly of asexual versus sexual entities coupled with ecological or other forms of divergence typical of separately evolving lineages (e.g. Vulpicida pinastri (Scop.) J.-E. Mattsson & M. J. Lai vs. V. juniperinus (L.) J.-E. Mattsson & M. J. Lai: Saag et al. 2014; Porpidia degelii (H. Magn.) Lendemer vs. P. albocaerulescens (Wulfen) Hertel & Knoph: Lendemer & Harris 2014; various groups within the Lobaria meridionalis clade: Cornejo & Scheidegger 2015; in contrast see Buschbom & Mueller 2005 and Tehler et al. 2013). Such observations have long been of interest to lichenologists and underlie the classical notion of 'species pairs', in which a sexually reproducing species commonly has a closely related asexual species counterpart (Du Rietz 1924; Poelt 1970; Hale 1976), not dissimilar to species pairs consisting of self-compatible and self-incompatible plants (reviewed in Charlesworth 2006). The observation that the sexual species is ancestral evolutionarily,

which then gives rise to asexual derivatives (self-incompatibility to self-compatibility in plants), has for many years been orthodox (Poelt 1970; Bowler & Rundel 1975; Tehler 1982; Takebayashi & Morrell 2001). This topic has been debated extensively (Tehler 1982; Buschbom & Mueller 2005), with the last 20 years or so yielding the phylogenetic methods as well as taxon sampling needed to advance a more quantitative understanding of the subject. In particular, a modern view has begun to emerge in which mostly asexual lineages may be viable for the long term evolutionarily, and may give rise to sexual lineages (Kroken & Taylor 2001b; Buschbom & Barker 2006; Cornejo et al. 2009).

The tremendous expansion of published lichen phylogenies over these two or more decades has yielded ample fodder for reexamining dogma pertaining to reproductive evolution in lichenology. More generally, probable links between reproductive modes and diversification dynamics in lichens (e.g., speciation, extinction, maintenance of existing and exploitation of new symbioses) indicate that trends in sexual versus asexual reproduction in lichens are of broad interest to evolutionary biologists. Without attempts at answering how and why diverse reproductive strategies evolve and are, or are not, maintained, as well as patterns in the frequency and directionality of character transitions, empirical and theoretical syntheses of reproductive biology across multiple domains of life will continue to be advanced without a lichenological perspective (see recent examples of syntheses from insects (Shuker & Simmons 2014), fish (Wootton & Smith 2014) and plants (Goodwillie et al. 2005; Karron et al. 2012; Pierre-Olivier 2012)). The present contribution seeks to make a first attempt at synthesizing knowledge of reproductive character evolution in lichens by explicit, model-based reconstruction of the frequency and polarity of evolutionary transitions between sexual and asexual reproduction. Here, asexuality as a dead end is defined as meeting two criteria: 1) that it is only a derived state and 2) that it is transient evolutionarily. I specifically aim to test whether dogma holds true and asexual reproduction is an evolutionary dead end in lichens, which has been argued in other lineages of life (Maynard Smith 1978; Takebayashi & Morrell 2001; Poulíčková *et al.* 2014), or conversely whether asexual lineages are a source for evolutionary innovation.

Materials and Methods

General strategy

The present investigation is based on ancestral state reconstructions of mode of reproduction. To accomplish this, only previously published phylogenies (i.e., tree files) with adequate taxon sampling and branch support were utilized (see below). Justification for using only published trees without further modification through new analyses of original datasets is that the enormous amount of work, knowledge, and expertise that accompanies any single phylogenetic study suggests that the best expert opinion regarding taxon sampling and phylogenetic interpretations is that conveyed in the original manuscript itself. Moreover, the present study is not focused on reassessing relationships presented in more than 20 prior works. As such, I refrain here from making any assertions or claims about what a given phylogeny 'should' look like, but rather make use of datasets that fit the criteria for analyses (see below).

Reproductive definitions

In the present study, sexually reproducing lichens are defined as species reproducing primarily through mycobiont spores that are products of meiosis (ascospores); these spores must then go on to encounter a suitable photobiont through various means (free-living, borrowed from a nearby thallus, etc.). Primary sexual structures in lichens are termed ascomata (specifically, apothecia (disc-like) or perithecia (flask-like)). Asexually reproducing lichens are defined here as species reproducing primarily through specialized propagules that are lichenized containing both the mycobiont and photobiont and are derived from mitosis. Primary asexual structures in lichens are termed soredia (ecorticate spherical bundles of hyphae and algae that usually form in the photobiont layer, ~20-100 µm diam.; Bowler & Rundle 1975), isidia (corticate finger-like outgrowths of the thallus containing hyphae and algae, variable in shape and size but typically larger than soredia) and phyllidia or schizidia and other outgrowths of the thallus that contain both symbionts, these often occurring along margins of lobes. A separate class of mitotic propagules occurs in lichens (conidia) but these disperse only the mycobiont (i.e. are non-lichenized) and are thought to function primarily in diploidization of sexually reproducing species but may also make contact with a suitable photobiont and establish new lichen thalli; as such, conidia are excluded from further consideration in this study.

Taxon sampling

It is widely appreciated that the single largest feature that has an impact on ancestral state reconstructions is taxon sampling (Salisbury & Kim 2001). In this study, I attempted to overcome this challenge as much as is possible, first by selecting lineages (clades, for the most part at the rank of genus and below) in which >50% of the total number of currently recognized species were included in phylogenetic sampling (Tier 1 analyses). The Lecanora varia (Hoffm.) Ach. group was included in this tier because, even though it was not possible to find published estimates of total clade size, it is likely that Pérez-Ortega et al.'s (2010) sampling of 33 species exceeds 50% of the total. However, because of the general paucity of studies that fit this criterion (this paucity is exacerbated by the lack of availability of data files in online repositories), a second threshold of taxon sampling was explored in which a minimum of 25% of species were included in the study (Tier 2 analyses). The present investigation represents a first attempt at understanding the generality of transitions in reproductive modes, and future studies with more complete taxon sampling are needed to fully explore the question at hand.

Five additional criteria were applied to both Tier 1 and Tier 2 analyses in selecting datasets suitable for analyses: 1) the group was monophyletic (e.g. for lineages in which non-monophyly has recently been demonstrated, such as Hypotrachyna (Vain.) Hale (Divakar et al. 2010), only a monophyletic ingroup was analyzed, e.g., Remotrachyna Divakar & A. Crespo + Bulbothrix Hale); 2) the focal ingroup was composed of species with more or less clearly delimited modes of reproduction either sexual or asexual. Trees containing species commonly marked \overline{by} mixed modes of reproduction such as Lobaria pulmonaria (L.) Hoffm. were not included in this study but warrant future investigation; 3) ingroup taxa displayed a diversity in reproductive mode. Well-sampled phylogenies such as those of Psora Hoffm. (Ekman & Blaalid 2011) and Polyblastia A. Massal. (Savíc et al. 2008) were not included because reproductive mode is fixed among taxa; 4) phylogenies contained several (but not necessarily all) branches that were strongly supported by bootstrap analysis or posterior probabilities; 5) a tree file containing a phylogeny derived from parsimony, maximum likelihood (ML), or Bayesian analyses was available in Dryad, TreeBase, or via email contact with the authors of the study.

Outgroup selection

Like taxon sampling, it is widely understood that outgroup selection can and does have a major impact on ancestral state reconstructions and has thus been the subject of extensive investigation (Salisbury & Kim 2001; Brady *et al.* 2011). However, rigorous and detailed exploration of this topic across the diversity of lineages included here, which would best be addressed through a combination of empirical and simulation studies, is beyond the scope of this investigation. I selected two or more of the closest relatives to the ingroup based on the published phylogeny. In rare cases, tree files were unrooted; in these instances, rooting on the dataset was imposed using the phylogenetic results presented in the literature.

Character matrices and tree files

One of two major challenges facing this study was scoring characters. Several (to my knowledge, there has never been a published estimate) asexually reproducing species are known to pass through a sexual phase at some point during an individual's lifetime, such as the widespread Pyxine sorediata (Ach.) Mont. in eastern North America. In contrast, most sexual species are not known to pass through an asexual phase, but see Hestmark (1991) who reported contemporaneous phases present in numerous species of Umbilicaria Hoffm. Moreover, we lack data regarding how commonly polymorphisms occur across multiple populations of a given species, across multiple species in a given clade, and how reproductive structures develop over the course of an individual lichen's entire life cycle; elegant examples of documentation of this can be found in Denison (2003) and Sanders (2014). As such, currently available data on intrathalline polymorphisms are not amenable to phylogenetic reconstruction of this feature across most lineages of lichens. This limitation directed a discrete rather than continuous character approach. I thus coded species according to their primary mode of reproduction, defined as the mode of reproduction most commonly encountered in the field, herbarium, and/or literature. Extensive effort was made to reference species protologues and/or images of original material (available on JSTOR Plants) when possible to score reproductive mode as sexual (0) or asexual (1).

The second major challenge facing this study was the availability of datasets that are deposited in public repositories. Many academic publications still do not require this from authors and, as such, approximately half of all studies identified as suitable for this investigation lacked available data online. In several instances, I was able to retrieve tree files from generous authors. Nonetheless, this deficiency limited the scope of the present study. Tree files used for reconstructions are described in Table 2.

Ancestral state reconstructions

Reconstructions were conducted on tree files available in public repositories. These datasets provided a range of possible tree files, including a single type of file derived from only one analysis to multiple types of tree files derived from analyses using different optimality criteria. Only a fraction of datasets included posterior distributions of highly likely trees (i.e. tree sets) derived from Bayesian analyses. Reconstructing ancestral states of traits on posterior distributions of trees yields the ability to explore uncertainties in their topologies (Tripp & Manos 2008). However, the limited availability of this file type prevented widespread exploration of topological uncertainty. As such, and to enable easier comparison of patterns among all datasets, reconstructions were

 TABLE 2. Inferred numbers and rates of transitions in mode of reproduction (sexual, asexual) for 23 clades of lichens included in this study. Ancestral state reconstructions were implemented via maximum likelihood methods utilizing a two-rate (asymmetric) model of evolution. The number of estimated taxa in a given clade was derived from the reference included in parenthesis. Log likelihoods of optimized reconstruction under the asymmetric model are provided.

 See Materials and Methods for explanation of terms.

| | Number of taxa sampled | Forward | Reverse | | | Root |
|-------------------------------------|--|---------------|---------------|------|-------|----------|
| Clade | (reference) | Rate (0 to 1) | Rate (1 to 0) | Bias | –Ln L | State |
| TIER 1 | | | | | | |
| Biatora | 42 of 42 (Printzen 2014) | 6.36 | 37.53 | 0.17 | 20.62 | Sexual |
| Caloplaca saxicola group | 8 of 8 sensu Gaya et al. (2009) | 0.03 | 0.30 | 0.10 | 11.98 | Sexual |
| | (Gaya <i>et al.</i> 2011) | | | | | |
| <i>Cladonia furcata</i> group | 6 of 6(Pino-Bodas <i>et al.</i> 2015) | 4.94 | 16.59 | 0.30 | 16.16 | Sexual |
| Dirina | 24 of 24 (Tehler et al. 2013) | 9.79 | 2.3 | 4.82 | 22.16 | Sexual |
| Flavoparmelia | 21 of 38 (Del-Prado et al. 2013) | 23.19 | 22.54 | 1.03 | 18.88 | Sexual |
| Fulgensia s.s. | 8 of 10 sensu Poelt (1965) (Gava et al. 2008) | 0.02 | 0.05 | 0.40 | 5.66 | Sexual |
| Lecania + relatives | 30 of 50 (Naesborg <i>et al.</i> 2007) | 0.90 | 13.56 | 0.07 | 7.48 | Sexual |
| Lecanora varia group | 33 of <i>c</i> . 50 | 1.62 | 13.08 | 0.12 | 14.41 | Sexual |
| 0 1 | (Pérez-Ortega et al. 2010) | | | | | |
| Letharia | 6 of 6 (Kroken & Taylor 2001 <i>b</i>) | 8.42 | 7.95 | 1.06 | 8.06 | Sexual |
| Lobariella | 17 of 26 (Moncada et al. 2013) | 9.13 | 14.11 | 0.65 | 12.69 | Asexual |
| Melanohalea | 18 of 22 (Leavitt et al. 2013) | 7.06 | 25.13 | 0.28 | 21.34 | Sexual |
| Miriquidica incl. | 12 of 23 (Singh et al. 2015) | 1.49 | 12.72 | 0.12 | 4.66 | Sexual |
| Protoparmelia | (¹ ⁰) | | | | | |
| Montanelia | 5 of 5(Leavitt et al. 2015) | 25.67 | 11.54 | 2.22 | 7.98 | Asexual |
| Parmotrema perforatum group | 4 of 6 (Lendemer <i>et al.</i> 2015) | 0.24 | 0.13 | 1.85 | 7.05 | Asexual? |
| Protoparmelia | 14 of 25 (Singh et al. 2015) | 4.17 | 2.94 | 1.42 | 13.42 | Sexual |
| Punctelia rudecta group | 16 of 30 sensu Crespo et al. | 135.12 | 15.62 | 8.65 | 17.61 | Asexual |
| 0 1 | (2004) (Alors et al. 2016) | | | | | |
| Xanthomendoza | 11 of 21 (Leavitt et al. 2013) | 9.06 | 9.07 | 1.00 | 11.68 | Asexual |
| TIER 2 | | | | | | |
| Austroparmelina | 6 of 7 (Crespo et al. 2010) | 36.74 | 36.67 | 1.00 | 5.54 | ? |
| Bulbothrix + Remotrachyna | 18 of 49 (Crespo et al. 2010) | 29.57 | 28.94 | 1.02 | 14.67 | ? |
| Fuscopannaria incl. Moelleropsis | 12 of 30 (Ekman et al. 2014) | 29.90 | 85.34 | 0.35 | 7.96 | Sexual? |
| Hypogymnia | 25 of 100 | 45.27 | 81.18 | 0.56 | 32.56 | Sexual? |
| | (Miadlikowska <i>et al.</i> 2011) | | | | | |
| Pannaria | 17 of 51 (Ekman et al. 2014) | 12.24 | 58.78 | 0.21 | 9.46 | Sexual |
| Parmelina + Myelochroa | 9 of 33 (Crespo <i>et al.</i> 2010) | 9.68 | 11.89 | 0.81 | 8.84 | Sexual? |
| | · ······ (-···························· | , | 07 | | | |

conducted on only a single tree including a most likely tree derived from ML₂ a maximum clade credibility tree derived from Bayesian analysis, or a consensus summary tree derived from parsimony analysis (Table 2). In cases where more than one tree file was available, reconstructions were conducted on the most favoured phylogeny presented in the published study. For likelihood-based studies (ML₂ Bayesian), ancestral state reconstructions made use of estimated branch lengths derived from the original study.

The only manipulation made to tree files prior to analysis was the removal of taxa not included in the ingroup or retained as outgroups for rooting trees. For example, for the *Fulgensia* A. Massal. & De Not. analysis all taxa in Gaya *et al.* (2008) were deleted except sampled *Fulgensia* plus the two nearest outgroups (*Letrouitia domingensis* (Pers.) Hafellner & Bellm. and *Letrouitia parabola* (Nyl.) R. Sant & Hafellner); from this matrix Fulgensia australis (Arnold) Poelt. was further pruned because the phylogenetic results in Gava et al. (2008) demonstrated that this taxon was not resolved within the otherwise monophyletic Fulgensia s. str. clade. Other than taxon pruning, no matrices or tree files were manipulated prior to analyses except in two instances where this was necessary, thus enabling a direct comparison of character evolution derived from this study with previously presented phylogenetic data. Firstly, no published phylogeny of Punctelia rudecta (Ach.) Krog exists but both a matrix and a tree file were available on TreeBase. However, because the tree file lacked branch support, the original matrix was used to conduct a parsimony bootstrap search in PAUP* using default settings and 100 replicates. Secondly, no online data were available for Tehler et al.'s (2013) phylogenetic study of Dirina Fr., but A. Tehler kindly provided the DNA matrix used in that publication; this was used to conduct an ML search and ML bootstrap search (100 replicates) in Garli (default settings). Finally, names of taxa/terminals were not altered prior to analysis except in instances where terminals contained no information indicative of species identification. In these cases, I added the specific epithet plus an underscore to the beginning of the name of the OTU. Information on data matrices, tree file names, and public repositories is provided in Appendix 1.

Rates of transitions

Using marginal reconstructions and comparative methods based on Brownian motion (cf. Felsenstein 2012) implemented in Mesquite v3.04, reproductive modes were estimated across nodes in target phylogenies using a simple Q matrix for a single character with two discrete states. This matrix contains four terms that describe stasis in either of the states (q_{00}, q_{11}) plus two instantaneous transition rates between the states: one forward and one reverse (q_{01}, q_{10}) ; the present study focused only on rates associated with transitions, not those associated with stasis. I used an asymmetric Markov continuous model of character evolution that permitted different rates for q₀₁ (forward rate of change) versus q10 (reverse rate of change). The forward rate is calculated as the overall rate × square root of the bias whereas the reverse rate is calculated as the overall rate/ square root of the bias. The overall bias in transition rates was calculated as the rate of gain of asexual reproduction (q_{01}) / the rate of gain of sexual reproduction (q_{10}) . Thus, values >1 indicate higher rates of gain of asexual reproduction from sexual ancestors whereas values <1 indicate higher rates of gain of sexual reproduction from asexual ancestors. A two-rate model was used instead of a one-rate model (where forward and reverse transition rates are equal) because the former vielded reconstructions with higher likelihoods in all cases, except in two instances in which the log likelihood estimated under the two different models was the same. The root state was optimized under a model that assumes root transition rates consistent with the overall model (i.e. the 'Root State Frequencies Same as Equilibrium' option in Mesquite).

Numbers of transitions

The overall number of state transitions (forward and reverse) was estimated as follows. The proportional likelihoods of states 0 and 1 for a given node were calculated and statistical support for each reconstructed node was based on the difference in log likelihood between the two character states: a difference >2 was taken to indicate support for the reconstruction. As a qualitative assessment, a 50% proportional likelihood cut-off was used to estimate whether a given node was most likely sexually or asexually reproducing. However, tallies of numbers of transitions were considered significant only if two conditions were met: 1) proportional likelihoods themselves were significant in that the difference between the two states was >2 log likelihood

units, and 2) relevant branches were supported because they were $\geq 70\%$ likelihood bootstrap (LB), $\geq 70\%$ parsimony bootstrap (PB), and/or $\geq 95\%$ posterior probability (PP)). Ancestral states were depicted in the figures only for nodes relevant to transitions (q_{01} or q_{10}) rather than on every node in a phylogeny (instances of q_{00} or q_{11}). Root state was inferred as the state reconstructed with the higher proportional likelihood but was not reported as significant unless the difference was >2 log likelihood units. The resulting files upon which ancestral state reconstructions are based have been deposited in Dryad (doi:10.5061/dryad.29h6k).

Results

A total of 23 datasets were utilized in the analyses: 17 comprised Tier 1 and six comprised Tier 2. Overall, transition rates were not predictive of numbers of inferred gains and losses of asexuality (Table 2, Fig. 1). Most lineages displayed higher rates of gain of sexual reproduction from asexual ancestors (i.e. reverse transitions with biases <1), despite the fact that in these lineages there were generally greater numbers of inferred gains of asexuality than losses (Fig. 1).

Across all 23 datasets, ancestral state reconstructions indicated dynamic histories of mode of reproduction in lichens (Figs 1-5; Supplementary Material Figures 1-19, available online). Gains of asexual reproduction from sexual ancestor was the more common pattern of transition but this transition was not irreversible; in several lineages including Hypogymnia (Nyl.) Nyl., Melanohalea O. Blanco, Montanelia Divakar, Parmotrema A. Massal., Protoparmelia M. Choisy, Punctelia Krog, and Xanthomendoza S. Y. Kondr. & Kärnefelt, statistically supported gains of sexual reproduction from asexual ancestors were documented (Table 2). The overall total number of gains of asexual reproduction from sexual ancestors was 26 (supported), plus additional unsupported gains (Fig. 1). The total number of gains of sexual reproduction from asexual ancestors was 14 (supported), plus additional unsupported gains (Fig. 1).

Discussion

Reproductive traits are among the most important and thus frequently studied aspects of the ecology and life history of



FIG. 1. Summary of numbers of inferred transitions from reconstructions of ancestral state of mode of reproduction in lichens. For each lineage, forward transitions, or gains of asexuality from sexual ancestors, are shown on the right (A) and reverse transitions, or gains of sexuality from asexual ancestors, are shown on the left (S). Black columns indicate significant transitions whereas grey columns indicate potentially additional but non-significant transitions. Data from 20 of 23 clades are presented; *Austroparmelia, Bulbothrix* and *Parmelina* were omitted because directionality of transition cannot be inferred from ancestral state reconstruction. Lineages are labelled by focal genus. From left to right, these are: *Biatora* (Bi), *Caloplaca* (Ca), *Cladonia* (Cl), *Dirina* (Di), *Flavoparmelia* (Fl), *Fulgensia* (Fs), *Lecania* (Le), *Lecanora* (Ln), *Lobariella* (Lo), *Letharia* (Lt), *Melanohalea* (Me), *Miriquidica* (Mi), *Montanelia* (Mo), *Punctelia* (Pc), *Pannaria* (Pn), *Parmotrema* (Pr), *Protoparmelia* (Pt), and *Xanthomendoza* (Xa).

organisms (Tehler 1982; Case et al. 2008; Asplen et al. 2009; Gomez-Mestre et al. 2012; Poulíčková et al. 2014; Oliveira et al. 2015). In particular, the question of whether asexuality (including 'selfing' in vascular plants) represents a terminal versus ancestral state and a stable versus transient state evolutionarily is both longstanding and pervasive across the tree of life (Bowler & Rundel 1975; Judson & Normark 1996; Goodwillie 1997; Normark et al. 2003; Igic et al. 2006; Gioti et al. 2013; Castagnone-Sereno Danchin & 2014: Hespeels et al. 2014). In this study, I amassed published datasets to explicitly reconstruct the frequency and polarity of evolutionary transitions between sexual and asexual reproduction in lichens. The resulting analyses indicate that gains of asexuality from sexual ancestors, consistent with dogma in lichenology, are indeed common and widespread across divergent groups of lichens. Summarizing the 23 clades studied here, the total number of forward transitions ranged from 26 (supported only) to 44 (including non-supported transitions). However, the analyses also yielded numerous reverse transitions: from 14 (supported) to 25 (including non-supported transitions) (Table 2). There are a few instances of



FIG. 2. Ancestral state reconstructions of reproductive mode using the *Biatora* clade (Tier 1 analysis). Proportional likelihoods of reconstructed state shown in circles on nodes, white = sexual; black = asexual. Nodes pertinent to transitions are labelled as follows: above horizontal lines are relative proportional likelihoods for sexual ("S") or asexual ("A") states, and an asterisk indicates reconstruction was significant for that state; below horizontal lines are support values (PP = Posterior Probability, LB = Likelihood Bootstrap, PB = Parsimony Bootstrap, NS = non-significant (i.e. <95% PP or <70% LB or PB)). Numbers of inferred transitions and directionality of transitions given to the right of taxon labels. NS indicates that the reconstruction was non-significant (i.e. that it did not meet both criteria for significance as described under Materials and Methods).</p>

non-supported transitions in which the direction of change is unknown. Thus, the numerous 'reverse transitions' from asexual ancestors to sexual derivatives documented here indicate ample diversity in reproductive mode evolution in lichens.



(Fig. continued on following page)



Asexuality is not a dead end in lichens

The results presented here reinforce a theme that has begun to emerge in lichenology in recent years, with several studies documenting the evolutionary potential of asexual lineages (e.g. Cornejo et al. 2009; Lendemer 2013). The work of Buschbom & Barker (2006) was one of the first and most important demonstrations that sexual lineages can arise from asexual ancestors. Through model-based reconstructions of reproductive mode in Porpidia s. lat. and other phylogenetic methods, these authors found evidence for the presence of traditional species pairs, statistical support for the nonmonophyly of different asexual lineages, extremely high rates of gain of sexual reproduction from asexual ancestors, and high (although non-significant) conditional probabilities of asexuality serving as the ancestral state for four of six key nodes.

Clearly, asexual reproduction in lichens does not represent an evolutionary dead end based on Criterion 1 laid out in the Introduction, that it is a derived state only; instead, asexuality also serves as an ancestral state. Criterion 2, that whole lineages are known to be entirely asexual (Lepraria (57 spp.): Saag et al. 2009; Lendemer 2013) or nearly entirely asexual (Chrysothrix (c. 20 spp.): Nelsen et al. 2009; Leprocaulon s. str. Lamy (c. 12 spp.): Lendemer & Hodkinson 2013; J. Lendemer, pers. comm.) indicates that asexual lineages obviously undergo speciation and thus cannot be described as transient but are rather stable evolutionarily. However, the degree to which asexual versus sexual lineages persist remains an open question that will be among the most difficult to answer in the coming years. One potential means of exploring this question would be through more complete taxon

sampling and analyses that account for time, for example, in a divergence time framework that allows for the calculation of dates of gain of asexuality as well as an estimation of longeveity of asexuality (the latter could then be compared to longeveity of sexuality). Such an approach would benefit from datasets in which authors have explicitly delimited phylogenetic species through gene tree-species tree reconciliation approaches, such as those made possible through coalescent based inference (e.g. Leavitt *et al.* 2015; Singh *et al.* 2015).

Whether reverse transitions are truly reflective of a complete regaining of sexuality rather than representing shifts from predominantly asexual to predominantly sexual states merits discussion. Regarding evolutionary trait reversals in other lineages, the genetic architecture underlying the trait of interest might not always be fully 'lost', so that the term 'gain' could be misleading (reviewed and discussed extensively in Collin & Miglietta 2008). Instead, sexual reproduction could be suppressed (functional and under selection, but down-regulated) via one or more regulatory mechanisms for some evolutionary time period. If accurate, regains of sexual reproduction from asexual ancestors might instead reflect 'latent homology' (Osborn 1902; Carroll 2008) of sexuality, in which case the terms gain and loss should be interpreted with caution. Lichens that commonly display both asexual and sexual modes of reproduction contemporaneously (e.g. Leptogium dactylinum Tuck.; E. Tripp, pers. obs. (Lendemer, Tripp, et al. 30272, NY)) or at different life stages (e.g. Lasallia pustulata (L.) Mérat; Hestmark 1992) lend evidence to support this hypothesis (see Hestmark et al. 2011 for further discussion). In lichens, I am not aware of any study that has described the

FIG. 3. Ancestral state reconstructions of reproductive mode using the *Dirina* clade (Tier 1 analysis). The genera *Dirina* and *Roccella* are presented within this clade. Proportional likelihoods of reconstructed state shown in circles on nodes: white = sexual; black = asexual; grey = ambiguous reconstruction owing to missing data. Nodes pertinent to transitions are labelled as follows: above horizontal lines are relative proportional likelihoods for sexual ("S") or asexual ("A") states, and an asterisk indicates reconstruction was significant for that state; below horizontal lines are support values (PP = Posterior Probability, LB = Likelihood Bootstrap, PB = Parsimony Bootstrap, NS = non-significant (i.e. <95% PP or <70% LB or PB)). Numbers of inferred transitions and directionality of transitions given to the right of taxon labels. Key: ap = apotheciate; sor = sorediate; cort = corticolous; sax = saxicolous.</p>



FIG. 4. Ancestral state reconstructions of reproductive mode using the *Hypogymnia* clade (Tier 2 analysis). Proportional likelihoods of reconstructed state shown in circles on nodes: white = sexual; black = asexual; grey = ambiguous reconstruction owing to missing data. Nodes pertinent to transitions are labelled as follows: above horizontal lines are relative proportional likelihoods for sexual ("S") or asexual ("A") states, and an asterisk indicates reconstruction was significant for that state; below horizontal lines are support values (PP = Posterior Probability, LB = Likelihood Bootstrap, PB = Parsimony Bootstrap, NS = non-significant (i.e. <95% PP or <70% LB or PB)). Numbers of inferred transitions and directionality of transitions given to the right of taxon labels. NS indicates that the reconstruction was non-significant (i.e. it did not meet both criteria for significance as described in Materials and Methods).

genetic architecture underlying sexual versus asexual reproduction, but I predict that a complete loss of sexual reproduction would

be an exception rather than the rule (the genus *Lepraria* representing one possible exception).



FIG. 5. Ancestral state reconstructions of reproductive mode using the *Punctelia* clade (Tier 1 analysis). Proportional likelihoods of reconstructed state shown in circles on nodes, white = sexual; black = asexual; grey = ambiguous reconstruction owing to missing data. Nodes pertinent to transitions are labelled as follows: above horizontal lines are relative proportional likelihoods for sexual ("S") or asexual ("A") states, and an asterisk indicates reconstruction was significant for that state; below horizontal lines are support values (PP = Posterior Probability, LB = Likelihood Bootstrap, PB = Parsimony Bootstrap, NS = non-significant (i.e. <95% PP or <70% LB or PB)). Numbers of inferred transitions and directionality of transitions given to the right of taxon labels.

Selfing as a dead end in other lineages?

To date, the majority of research that has explored micro- and macroevolutionary consequences of reduced outcrossing has done so from a vascular plant perspective of 'selfing', most commonly sporophytic selfing (and thus not selfing in the gametophytic or truly asexual That selfing evolves from selfsense). incompatible ancestors is axiomatic in plant reproductive biology and has been intensively investigated ever since Fisher's (1941) earliest theoretical models describing reduced fitness but transmission advantage of selfers over outcrossers. An exhaustive review of selfing as a dead end in plants was undertaken by Takebayashi & Morrell (2001), who concluded that the evidence to date was largely consistent with the dead-end hypothesis but recognized that directionality in plant mating system evolution may be more dynamic than previously appreciated. Since that review, evidence from plants as well as other organisms has cast doubt on the robustness of the dead end dogma. In an exhaustive study involving 571 taxa of Asteraceae, Ferrer & Good-Avila (2007) used phylogenetic methods to demonstrate both forward and reverse transitions between selfincompatibility and self-compatibility, suggesting that selfing is not a dead end. In nematodes, unique genomic rearrangements are associated with mitotic, highly successful clades of asexual parasites (Castagnone-Sereno & Danchin 2014), similarly suggesting that selfing is not a dead end. In an extensive review of estimates of the ages of asexual lineages from across the tree of life, Neiman et al. (2009) demonstrated that asexual lineages are not always short-lived, as has been commonly held.

One limitation of the above comparison is that asexual reproduction in lichens is not equivalent to (sporophytic) selfing in vascular plants. A more appropriate comparison would be that of homothallism and heterothallism in lichens to self-compatibility and self-incompatibility in vascular plants, but data available on homo/heterothallism in lichens are far more scarce (before Scherrer *et al.* 2005; Singh *et al.* 2012) than the basic knowledge of sexual versus asexual modes of reproduction. Nonetheless, taken together, evidence from lichens, plants, and other organisms suggests that re-examination of the dead-end hypothesis is needed across the board: primarily asexual and/or selfing lineages can and do undergo speciation, may be old evolutionarily, and can and do give rise to primarily sexual lineages.

Origins, maintenance, and success of asexual lichens

The origin, maintenance, and success of asexuality or selfing as a primary reproductive strategy call for efforts to understand the underlying genetic mechanisms and population-level processes. Regarding origins of asexuality, the loss of mitochondria from conidia, which has been documented in some Xanthoria (Fr.) Th. Fr., effectively renders these propagules (that presumably function in fertilization of sexual species) functionless, without the capacity to germinate (Honegger 1984). Such a mechanism may be related to the complete loss of sexual reproduction in these organisms. Secondly, origins of asexuality at a population level may be explained by an imbalance in mating type alleles among individuals wherein heterothallic species fail to form sexual structures because of a paucity or lack of compatible mating types (Zoller et al. 1999; Singh et al. 2012; see below for further discussion). Regardless of the mechanisms of the origins of asexuality, it must be remembered that sexual reproduction may in fact rarely be fully 'lost' (see above). Future studies at the genetic level are needed to clarify the molecular basis for sexual trait expression in lichens.

Maintenance of asexuality is likely explainable through some mechanism or set of mechanisms to acquire new genetic variation, perhaps to a degree greater than previously appreciated. First, both Kroken & Taylor (2001*a*) and Buschbom & Mueller (2005) cited recombination in asexual lineages of lichens, perhaps made possible through rare sexual reproduction, and similar evidence derives from non-lichenized fungi (Burt *et al.* 1996; Geiser *et al.* 1998). Second, new variation may additionally be acquired through mutation. Dal Grande *et al.* (2012) found that, among 2229 thalli spanning 62 populations of *Lobaria pulmonaria*, 15% of genetic diversity in the mycobiont might be explainable by somatic mutations; more generally, appreciation for a role for somatic mutations in generating new genetic diversity is growing (Frank 2010; Yong 2012). Third, fungi may acquire new genetic material through non-sexual processes such as parasexuality, or the fusion of vegetative cells from adjacent thalli (Clutterbuck 1996).

Success of asexual lineages has been addressed in a variety of previous works. In general, co-dispersal of both symbionts in vegetative propagules of asexual species is a highly efficient means of reproduction in lichens (Honegger 1984), and some asexually reproducing species have geographical ranges that are nearly twice as large as those of sexual species (Tripp et al. 2016). Furthermore, Bowler & Rundel (1975) speculated that species with asexual reproduction have, on the whole, broader ecological amplitudes, such as the capacity of sorediate taxa to colonize both hardwoods and conifers (vs. some sexual taxa that colonize only hardwoods). Such 'generalist' strategies may correlate to the evolutionary longevity of asexual lineages. Other authors have hypothesized that asexuality is an effective mode of reproduction in degraded landscapes such as urban areas or industrial sites (Howe & Lendemer 2010). For example, LeBlanc & De Sloover (1970) found increased production of soredia associated with atmospheric pollution. Nonetheless, we currently lack a large-scale metasynthesis of correlations among environmental conditions, mode of reproduction, and other lichen functional traits. Future studies are needed to help understand the repeated origins, maintenance, and successes of asexual lineages.

Discrepancy in transition rates versus transition numbers

This study reiterates caveats associated with inferring rates of evolution during ancestral state reconstructions of character states. In particular, taxon sampling, branch lengths, models of evolution, and variable rates of evolution are among the many features that have an impact on reconstructions (Salisbury & Kim 2001; Wiens et al. 2007; Cusimano & Renner 2014). In this study, I utilized an asymmetric two-rate model of evolution that was favoured over a symmetric one-rate model. However, the model implemented assumed constant rates of character evolution through time, no differences in rates of evolution across branches, and no differences in extinction rates among lineages. It is likely that the violation of one, two, or all three of these assumptions contributed to the discrepancy between transition rates and transition numbers documented here. For example, the Dirina and Biatora Fr. analyses yielded qualitatively similar patterns in transitions, in that both showed gains of asexuality from sexual ancestors and none in the reverse direction, but yielded very different transition rate biases (biases >1 in Dirina but <1 in Biatora). These two datasets additionally differed in their distribution of transitions: gains of asexuality were inferred only among extreme tips of the Biatora phylogeny, whereas gains of asexuality characterized deeper nodes of the Dirina phylogeny. Because of limitations of the method of ancestral state reconstruction implemented here, I have erred on the side of caution and emphasize inferred numbers of transitions over rates of transition in interpreting results. Future studies on lichen reproductive evolution should explore the impacts of different methods on ancestral state reconstructions, including methods that account for differential rates of character evolution and extinction, as well as those that account for relative time (e.g. rate smoothing approaches) or absolute time (e.g. divergence time analyses calibrated preferably by primary fossils).

Additional limitations

Interpretations from the present study rely on the robustness of the patterns documented here. Namely, it is well known that ancestral state reconstructions are complex analyses that depend heavily on several parameters in addition to those described above, not least of which is working with trees that reflect true evolutionary history (Duchêne & Lanfear 2015). Igic et al. (2006) demonstrated that spurious results can arise from analyses that use only character states of extant taxa to infer ancestral states. At present, inference of major macroevolutionary patterns among lichens is limited by current availability of thoroughly (to completely) sampled phylogenies of extant taxa that form monophyletic groups; only with focused efforts to build such datasets can we begin to better understand the extent to which incomplete taxon sampling alters our assessment of these patterns. Both taxon and molecular (genetic) sampling similarly affect branch lengths, which are also well known to influence ancestral state reconstructions (Pagel 1999; Litsios & Salamin 2012). The impact of basing reconstructions on raw phylogenetic distances that account only for instead of ultrametricized substitutions distances that additionally account for time has been debated, but recent studies have suggested that the two methods might yield qualitatively similar results more often than not (Cusimano & Renner 2014). Here, I used trees based on raw phylogenetic distances not transformed in any way to account for relative or absolute time; further study is needed to explore the impact of reconstructions conducted on ultrametricized trees, if any.

In summary, although taxon sampling, extinction, variable rates of evolution, and many other factors are well-known complicators of phylogenetic and character evolution inferences (Salisbury & Kim 2001; O'Meara 2012), much knowledge can still be gained through ancestral state reconstructions and I predict that the overall dynamic histories of lichen reproductive evolution herein documented will hold through time.

What's next?

In lichens, as well as in many nonlichenized fungal lineages, genetic research on reproductive biology has focused on homothallism and heterothallism, roughly equivalent to self-compatibility versus incompatibility in flowering plants. Even though not as exhaustively researched for non-lichenized ascomycetes, it is well

understood that lichens have two extremely divergent mating type alleles found at a single locus (MAT-1, MAT-2) that determine reproductive compatibility (Coppin et al. 1997; Honegger et al. 2004; Rydholm et al. 2007). Heterothallic lichens reproduce sexually when two nuclei containing different alleles fuse, whereas homothallic lichens reproduce sexually when nuclei containing the same mating type allele fuse (note that some homothallic species harbour both alleles; Scherrer et al. 2005; Singh et al. 2012). Future research in this area will undoubtedly yield important insights into mating system evolution, especially studies that combine next generation sequencing technologies with careful laboratory work such as analyses of single sporelings (Honegger et al. 2004). What remains to be explored in any detail is the relationship between mode of reproduction (sexual, asexual) and breeding system (homothallic, heterothallic). It seems clear that sexual versus asexual reproduction in lichens is largely one with genetic underpinnings rather than environmental plasticity. Yet the genetic basis of reproductive structures in lichens has never been studied. This is the most obvious area of extension for further research in lichen reproductive biology and is, to my knowledge, essentially wide open.

Conclusions

The present study represents a first attempt to synthesize general patterns in macroevolutionary transitions in mode of reproduction across disparate groups of lichenized fungi. Ancestral state reconstructions on phylogenies from several different groups indicate longheld assumptions about directionality in reproductive evolution are supported by empirical data in some cases, but not in others. In fact, that asexual lineages are long-lived evolutionarily and can give rise to sexual lineages (i.e. are not evolutionary dead ends) has been minimally appreciated in the past. Ancestral state reconstruction remains a powerful approach to understanding major trends in character evolution across groups. However, methods are well known to be sensitive to a wide variety of parameters including those

pertaining to taxon sampling, character coding, model of character evolution, branch lengths, and rate heterogeneity among branches and through time (Ekman *et al.* 2008; Cusimano & Renner 2014; King & Lee 2015). A fully elucidated picture of lichen reproductive trait evolution awaits more densely sampled phylogenies and rigorous exploration of the impact of alternative reconstruction methods and models of evolution. For now, we can appreciate that asexuality in lichens may not be the evolutionary end point it was once viewed as.

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SUPPLEMENTARY MATERIAL

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| Clade | File name | Study number | Optimality criterion | Reference* |
|------------------------------------|---|---|----------------------|-----------------------------------|
| TIER 1 | | | | |
| Biatora | T68835.nex | S15023 (TreeBase) | ML | Printzen (2014) |
| Caloplaca saxicola group | 20723.nex | S10717 (TreeBase) | Parsimony | Gaya et al. (2011) |
| Cladonia furcata group | [none deposited, sent by authors] | [none] | Bayesian | Del-Prado et al. (2013) |
| Flavoparmelia | T63498.nex | S14222 (TreeBase) | Bayesian | Del-Prado et al. (2013) |
| Fulgensia s.s. | T49479.nex | S12085 (TreeBase) | Parsimony | Gaya et al. (2008) |
| Lecania + relatives | T972.nex | S1737 (TreeBase) | Bayesian | Naesborg et al. (2007) |
| Lecanora varia group | [none deposited, sent by authors] | [none] | Bayesian | Pérez-Ortega et al. (2010) |
| Letharia | T27653 | SN376-1131 (TreeBase) | Parsimony | Kroken & Taylor (2001a) |
| Lobariella | [none deposited, sent by authors] | [none] | ML | Moncada et al. (2013) |
| Melanohalea | T50986.nex | S12364 (TreeBase) | ML | Leavitt et al. (2013) |
| Miriquidica incl. Protoparmelia | [none deposited, sent by authors] | [none] | ML | Singh et al. (2015) |
| Montanelia | T76977.nex | S16237 (TreeBase) | ML | Leavitt et al. (2015) |
| Parmotrema perforatum group | RAxML_bestTree.Parmotrema_ 1_ITS.nex | http://dx.doi.org/10.5061/ dryad.6c605 | ML | Lendemer et al. (2015) |
| Protoparmelia | [none deposited, sent by authors] | [none] | ML | Singh et al. (2015) |
| Punctelia rudecta group | T90392.nex | S18070 (TreeBase) | Bayesian | Alors et al. (2016) |
| Xanthomendoza | T66987.nex | S14794 | Bayesian | Leavitt et al. (2013) |
| TIER 2 | | | | |
| Austroparmelina | combined_min3_max2_ex_2.mb. com.nex | http://dx.doi.org/10.5061/ dryad.400b6 (Dryad) | Bayesian | Crespo et al. (2010) |
| Bulbothrix + Remotrachyna | combined_min3_max2_ex_2.mb. | http://dx.doi.org/10.5061/ | Bayesian | Crespo et al. (2010) |
| Europe and and a in al | Com.nex | aryad.400b6 (Dryad) | Daviasian | Element d (2014) |
| Moelleropsis | 1 08203.nex | S14978 (TreeBase) | Bayesian | Ekman <i>et al.</i> (2014) |
| Hypogymnia | T73442.nex | S11110 (TreeBase) | ML | Miadlikowska <i>et al.</i> (2011) |
| Pannaria | T68563.nex | S14978 (TreeBase) | Bayesian | |
| Parmelina + Myelochroa | combined_min3_max2_ex_2.mb. con.nex | http://dx.doi.org/10.5061/ dryad.400b6 (Dryad) | Bayesian | Crespo et al. (2010) |

Appendix 1. Information on datasets used in this study including original file name as provided in TreeBase or DRYAD, study number provided in TreeBase or DRYAD, optimality criterion used to construct phylogeny and reference from which tree file was derived.

*see main reference list.

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