# Organic weed control in white lupin (*Lupinus albus* L.)

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# Abstract

Legumes such as white lupin (*Lupinus albus* L.) provide a valuable nitrogen source in organic agriculture. With organic farming hectarage increasing and white lupin interest increasing in the southeastern USA due to newly released winter hardy cultivars, non-chemical weed control practices in lupin are needed. A two-year experiment was established at two locations in Alabama. Five weed control practices were evaluated: one pre-emergence (PRE)-applied herbicide (*S*-metolachlor), two mechanical (hand hoed) and two cultural (living mulch utilizing two black oat cultivars) weed control treatments. Fourteen weed species were encountered. *S*-metolachlor provided above 80% control of most weed species present in this experiment. The cultivation treatments and black oat companion crops also provided good weed control of many of the weeds encountered. Crop injury of all treatments was low on a 0 to 10 scale with 0 representing no injury: <2.0, <1.3 and <1.2 by *S*-metolachlor, the cultivation treatments and the black oat companion crops, respectively. Grain yield of cultivars ABL 1082, AU Alpha and AU Homer were 1540, 1130, 850 kg ha<sup>-1</sup>, respectively, when treated with the conventional treatment, *S*-metolachlor. Grain yield in the organic treatments was equivalent. The cultivation treatments and black oat companion crops, respectively. Grain yield of cultivars ABL 1082, AU Alpha and AU Homer were 1540, 1130, 850 kg ha<sup>-1</sup>, respectively, when treated with the conventional treatment, *S*-metolachlor. Grain yield in the organic treatments was equivalent. The cultivation treatments and black oat companion crops.

Key words: companion crop, cultivation, mechanical weed control

# Introduction

Organic production is an increasing sector in US agriculture. To be certified as organic, a farm must follow the United States Department of Agriculture National Organic Program (NOP) guidelines. Conventional agriculture is dependent on synthetic nitrogen (N) fertilizers and herbicides for maximizing crop performance<sup>1</sup>. The NOP prohibits the use of synthetic herbicides and fertilizers<sup>2</sup>; therefore, alternative strategies for sufficient nutrient availability as well as for weed, insect and disease control are implemented in organic production.

Worldwide, 450 lupin species have been identified but only four species are used agronomically<sup>3</sup>. White lupin (*Lupinus albus* L.) was first introduced into the southeastern USA in the 1930s and the production eclipsed 1 million ha in the early 1950s. Hectarage subsequently declined due to loss of government support, damage to seed nurseries because of mid-autumn freezes in two consecutive years and the increased availability of inorganic fertilizers<sup>4–6</sup>. White lupin is of increasing interest in the southeastern USA because newly released cultivars exhibit vernalization requirements similar to wheat (*Triticum aestivum* L.). Winter-type cultivars offer an economic opportunity as an alternative legume cover crop, grain crop or forage. White lupin used in a winter cover crop rotation increased lint yield in cotton (*Gossypium hirsutum* L.) as compared to traditional rotations<sup>6</sup>. Furthermore, white lupin is attractive as mid-winter forage for ruminants due to a forage quality similar to that of alfalfa (*Medicago sativa* L.)<sup>6</sup>.

*Lupinus* spp. are poor weed competitors during early establishment and canopy development is slow. Slow canopy development facilitates light penetration, weed seed germination and subsequent yield loss due to competition. Lupins reach maximum vegetative growth and competitiveness during flowering<sup>7</sup>. Weeds compete with lupin for water, nutrients and light; therefore, effective weed control, especially during early establishment, is necessary for the crop's success<sup>7,8</sup>.

*S*-metolachlor, a chloroacetamide, is one of three active ingredients currently registered for use in US lupin production<sup>9</sup>. This pre-emergence (PRE)-applied herbicide provided good control of annual bluegrass (*Poa annua* L.), shepherd's purse (*Capsella bursa-pastoris* L.) and common

chickweed (*Stellaria media* (L.) Vill.)<sup>10</sup>. *S*-metolachlor has been used for over a decade as a standard weed control program at Auburn University<sup>11,12</sup>.

Mechanical and cultural weed control practices are important weed control alternatives in organic production. Hoeing is prohibitively expensive due to labor cost and hence is only used in high-value specialty crops or as supplement to other weed control practices. It is a successful weed control method for weed seedlings and annual/ biennial weeds<sup>13</sup>. Lentil (*Lens culinaris* Med.) yield was found to be higher in hand-hoed plots than in plots in which herbicides such as linuron and metribuzin were applied<sup>14</sup>.

Cover crops play a major role and are beneficial in any farming system such as conservation agriculture and organic farming. Some benefits include: reduction of soil erosion, reduction in pesticide use (herbicides, insecticides and fungicides), improved soil moisture, enhanced soil organic matter and breaking of pest cycles<sup>15</sup>. As a weed management tool, living cover crops are used to outcompete (smother) weeds, whereas desiccated cover crops are used as mulch, and both methods can release allelopathic chemicals<sup>13</sup>. Black oats (Avena strigosa Schreb.), a cool-season annual cereal, is a promising, relatively new cover crop in the southern USA and has been used successfully for many years as a cover crop for soybean [Glycine max (L.) Merr.] in Brazil<sup>15</sup>. Reasons for the success of this cover crop are its large biomass production and its exceptional allelopathic activity<sup>16</sup>. Even though black oat is successfully used as a weed management tool in soybean, cotton shows sensitivity to its allelopathic activity<sup>17</sup>.

The specific objectives were to determine the effect of organic weed control practices such as cultivation and companion crops on weed control, injury to lupin, lupin density, lupin reproductive maturity and yield.

# **Materials and Methods**

The experiment was conducted at two locations in October 2007 and 2008, respectively: the Alabama Agricultural Experiment Station E.V. Smith Research Center Field Crops Unit (FCU), near Shorter, AL (32.42N, 85.88W) on Compass loamy sand and at the E.V. Smith Research Center Plant Breeding Unit (PBU), near Tallassee, AL (32.49N, 85.89W) on Wickham sandy loam.

The experiment was a randomized complete block design (repeats = 4) with a 2 (year)  $\times$  2 (location)  $\times$  3 (lupin cultivar)  $\times$  5 (weed control) factorial treatment arrangement. Blocks were nested within year  $\times$  location  $\times$  cultivar combinations. The three lupin cultivars used were AU Homer (a high-alkaloid, indeterminate cover crop type), AU Alpha (a low-alkaloid, indeterminate forage type) and ABL 1082 (low-alkaloid, determinate grain-type experimental cultivar). The weed control factor had five levels: one pre-emergence (PRE)-applied herbicide (*S*-metolachlor), two mechanical (hand hoed) and two cultural (living mulch utilizing two black oat cultivars) weed control treatments. A non-treated control plot was also present.

Inoculated lupin were seeded in four rows with a John Deere<sup>®</sup> 1700 four row vacuum planter (Deere and Company, Moline, IL) with a row spacing of 90 cm at a depth of 1.25 cm in October 2007 and 2008. Smooth seed beds were prepared 2 weeks prior to planting in 2007. In 2008, raised beds were prepared with a KMC<sup>®</sup> four-row ripper/bedder (Kelley Manufacturing Company, Tifton, GA) due to concerns about waterlogging at both locations. The plot length was 6.0 or 7.5 m depending on location and year.

The conventional treatment *S*-metolachlor was applied at a rate of 1.12 kg a.i. ha<sup>-1</sup> (one day after planting) with a compressed CO<sub>2</sub> backpack sprayer delivering 140 liters ha<sup>-1</sup> at 147 kPa. The cultural control treatments, cv. SoilSaver and As 033 (a selection from PI 436103) black oat (*Avena strigosa* Schreb.) were hand sown 1 (2007) or 7 days (2008) after seeding of the lupin crop. The mechanical weed control treatments, between-row only cultivations and between and within row cultivation treatments were accomplished twice per year at 4 (2007) or 6 (2008) weeks after planting and 18 to 20 weeks, respectively.

Weed control ratings were recorded at both locations on a scale from 0% (no weed control) to 100% (complete weed control). The non-treated control was used to estimate the level of control in the treated plots. Two weed control ratings per treatment/plot were taken in each study year. The first rating was taken 6 weeks after planting and PRE application in both years. The second rating was taken 22 or 26 weeks after planting in 2007/2008 and 2008/2009, respectively.

Crop injury ratings were taken on a scale from 0 (no injury/alive) to 10 (complete injury/dead). In 2007/ 2008, crop injury ratings were taken 3 weeks after planting and PRE application and 15 weeks after planting. In 2008/ 2009, injury ratings were taken 4 weeks after planting and PRE application and 18 weeks after planting.

Stand counts were taken in the two center rows of each four-row plot along a 3-m pole. In 2007/2008, the first, second and third counts were taken 6, 11 and 16 weeks after planting, respectively. In 2008/2009, two counts were taken 4 and 8 weeks after planting. Due to frequent rains in winter 2008/2009, plots were inaccessible for a third stand count.

Maturity ratings on a scale from 0 (not in bloom) to 100% (full bloom) were taken to determine whether any treatments delayed maturity. The rating was taken at the end of March at both locations in both years.

To determine the plot yield as influenced by weed management practice, the two center rows of each plot were harvested with a plot combine. Plot samples were dried to constant weight, weighed and a sample taken to determine test weight and mean seed mass on a plot basis.

Mixed model procedures as implemented in SAS<sup>®</sup> PROC GLIMMIX (SAS Institute Inc., Cary, NC) were used to analyze weed control, crop injury, plant density, maturity and yield. Cultivar, weed control treatment and their interaction were treated as fixed effects. Location,

year and block (cultivar), their interaction with each other and fixed effects were considered to be random effects. All response variables, including weed control, were analyzed using the normal distribution function based on plots of standardized residuals. Occasionally, the GROUP option was employed to create variance groups for groups of treatments. Statistical significance was declared at Dunnett's P < 0.1.

# Results

#### Weed control

During the experiment 14 weed species were encountered. None of the interactions and treatment main effects were significant for Geranium carolinianum L., Trifolium incarnatum L. and Medicago lupulina L. Cultivar by treatment interaction was non-significant, but treatment main effects were significant for all other weed species mentioned in Table 1. At the first rating 6 weeks after planting in 2007 and 2008, lesser swinecress [Coronopus didymus (L.) Sm] was controlled by S-metolachlor to 86%. Both organic living mulch black oat treatments provided much lower control ( $\leq 28\%$ ). S-metolachlor provided 94% control of henbit (Lamium amplexicaule L.). With less than 25% control by both cultivation treatments and less than 62% control by both black oat cultivars, the organic treatments were inferior to S-metolachlor. The continuous germination of henbit<sup>18</sup> is likely the reason for poor control by organic treatments. Cutleaf-evening primrose (Oenothera laciniata Hill) was controlled to 99% by S-metolachlor and to 90% by both black oat cultivars. Both cultivation treatments provided much lower control ( $\leq 25\%$ ). S-metolachlor controlled annual bluegrass (Poa annua L.) to 91%. With the exception of between-within row cultivation (73%), all of the remaining organic treatments provided lower control of this weed ( $\leq 21\%$ ). Black oat cultivars SoilSaver (88%) and As 033 (82%) were as successful as S-metolachlor (82%) in controlling wild radish (Raphanus raphanistrum L.). With less than 35% control both cultivation treatments were inferior to S-metolachlor. Heartwing sorrel (Rumex hastatulus Baldw.) was controlled to 90% by S-metolachlor. With less than 60% control both black oat cultivars provided lower control than S-metolachlor. S-metolachlor provided 88% control of corn spurry (Spergula arvensis L.). The cultivation and companion crop treatments provided lower control than S-metolachlor. SoilSaver (90%) and As 033 (88%) were as successful as the conventional treatment (86%) in controlling winter vetch (Vicia villosa Roth).

At the second rating 22 weeks and 26 weeks after planting in 2007 and 2008, respectively, henbit, a winter annual weed species, was not present since its growing season ended earlier. Shepherd's purse [*Capsella bursa-pastoris* (L.) Medik.] was better controlled by the cultivation (>93%) and black oat companion crops (>90%) than by S-metolachlor. Control of lesser swinecress by

S-metolachlor was reduced to 72%. Between-within row cultivation (94%) provided better control than S-metolachlor. Providing less than 51% control, the black oat companion crops were inferior to S-metolachlor. Cutleafevening primrose control by S-metolachlor (36%) was inferior at the second rating as compared to the first rating. With more than 70% control both cultivation treatments and both black oat companions provided better control than S-metolachlor. Germination of this weed species continued after the first rating. Annual bluegrass control by S-metolachlor was 93%. Between-row cultivation (60%) provided lower control than S-metolachlor. S-metolachlor provided 86% wild radish control. Between-within row cultivation (97%) provided superior control. Heartwing sorrel control by S-metolachlor was 77%. Both cultivation treatments provided better control (>90%) than S-metolachlor. Corn spurry control by S-metolachlor was 81%. Both cultivation treatments performed as successfully as the conventional treatment. SoilSaver and As 033 provided 58% and 66% control, respectively. S-metolachlor provided 79% winter vetch control.

Cultivar by treatment interaction was significant for annual ryegrass (*Lolium multiflorum* Lam.) (Table 2). Annual ryegrass was present only at the second rating. Mean control of annual ryegrass in cultivar ABL 1082 was 98%. Black oat cultivar As 033 provided excellent control (91%), but this was lower than control by *S*-metolachlor. AU Alpha weed control treatments provided equivalent weed control. Annual ryegrass was controlled 90% by *S*-metolachlor. With the exception of between-row cultivation (64%), none of the organic treatments provided lower control than *S*-metolachlor in AU Homer.

Similarly, cultivar by treatment interaction also was significant for yellow nutsedge (*Cyperus esculentus* L.). At the first rating, yellow nutsedge was controlled to 90% by *S*-metolachlor in ABL 1082. Between-row and between-within row cultivation both provided 12% control. Similar results were observed in cultivars AU Alpha and AU Homer. *S*-metolachlor provided 96% and 99% control of yellow nutsedge in AU Alpha and AU Homer, respectively. Both cultivation treatments (<37% control) were significantly inferior in both cultivars.

At the second rating, S-metolachlor provided  $\ge 90\%$  control of yellow nutsedge in all three cultivars. Betweenrow cultivation in ABL 1082 was the only treatment that provided lower control of yellow nutsedge than S-metolachlor. In AU Alpha the organic treatments controlled this weed to more than 77%. S-metolachlor and black oat companions each provided 99% control of yellow nutsedge at this rating.

# Lupin injury

Treatment by cultivar interaction was non-significant at P = 0.10. Treatment main effects were significant. At the first rating, both cultivation and black oat treatments reduced crop injury compared to the *S*-metolachlor

	S-metolachlor		Cultivation between rows		Cultivation between and within rows		Black oat 'SoilSaver'		Black oat 'As 033'	
Weed species	% Control	SE	% Control	Dunnett's <i>P</i> -value	% Control	Dunnett's <i>P</i> -value	% Control	Dunnett's <i>P</i> -value	% Control	Dunnett's <i>P</i> -value
Rating 1										
Coronopus didymus (L.) Sm.	86	10.7	71	0.2146	83	0.9859	26	< 0.0001	28	0.0002
Lamium amplexicaule L.	94	11.4	46	< 0.0001	51	< 0.0001	62	< 0.0001	56	< 0.0001
Oenothera laciniata Hill	99	8.7	25	< 0.0001	21	< 0.0001	90	0.6814	90	0.5781
Poa annua L.	91	9.4	67	0.0521	73	0.4028	59	0.0490	42	0.0019
Raphanus raphanistrum L.	82	12.6	34	0.0094	17	< 0.0001	88	0.9252	82	1.0000
Rumex hastatulus Baldw.	90	9.7	81	0.0310	87	0.8187	59	0.0007	53	< 0.0001
Spergula arvensis L.	88	8.5	43	< 0.0001	49	< 0.0001	59	0.0003	56	0.0001
Vicia villosa Roth	86	10.3	25	< 0.0001	21	< 0.0001	90	0.9781	88	0.9958
Rating 2										
Capsella bursa-pastoris (L.) Medik.	82	7.2	93	0.0098	94	0.0066	90	0.1011	93	0.0131
Coronopus didymus (L.) Sm.	72	11.1	74	0.9949	94	0.0548	51	0.0691	36	0.0273
Oenothera laciniata Hill	36	5.8	71	< 0.0001	85	< 0.0001	74	< 0.0001	75	< 0.0001
Poa annua L.	93	3.9	60	< 0.0001	84	0.1954	86	0.3631	86	0.3631
Raphanus raphanistrum L.	83	6.1	90	0.7898	97	0.0474	85	0.9950	81	0.9944
Rumex hastatulus Baldw.	77	6.1	94	0.0054	98	0.0007	77	1.0000	74	0.9806
Spergula arvensis L.	81	13.1	81	1.0000	81	1.0000	58	< 0.0001	66	0.0329
Vicia villosa Roth	79	5.8	91	0.1299	91	0.1014	75	0.9229	76	0.9881

**Table 1.** Weed control as affected by treatment 6 (2007 and 2008), 22 (2007) or 26 (2008) weeks after planting. Dunnett's test was performed to compare cultivation treatments and companion black oat treatments to a conventionally grown crop treated with S-metolachlor. Each species was analyzed separately, hence the different standard errors.

**Table 2.** Control of annual ryegrass (*Lolium multiflorum* Lam.) and yellow nutsedge (*Cyperus esculentus* L.) in *L. albus* L. cultivars as affected by treatment 6 (2007 and 2008), 22 (2007) or 26 (2008) weeks after planting. Dunnett's test was performed to compare cultivation treatments and companion black oat treatments to a conventionally grown crop treated with *S*-metolachlor. The group option was employed to account for heterogeneous variances among treatments, hence the varying standard errors.

	Lolium	n multiflo	rum Lam.	Cyperus esculentus L.						
		Rating	2	Rating 1			Rating 2			
Cultivar treatment	% control	SE	Dunnett's <i>P</i> -value	% control	SE	Dunnett's <i>P</i> -value	% control	SE	Dunnett's <i>P</i> -value	
ABL 1082										
S-metolachlor	98	5.6		90	9.5		94	13.7		
Cultivation between rows	97	8.1	1.0000	12	9.5	< 0.0001	35	13.7	0.0158	
Cultivation between and within rows	99	5.5	0.9944	12	9.5	< 0.0001	45	13.7	0.0519	
Black oat 'SoilSaver'	99	5.8	0.9996	91	9.5	1.0000	99	13.7	0.9973	
Black oat 'As 033'	91	5.1	0.0765	89	9.5	1.0000	99	13.7	0.9973	
AU Alpha										
S-metolachlor	94	5.6		96	11.0		92	6.6		
Cultivation between rows	96	8.5	0.9957	37	11.0	0.0002	77	6.6	0.2783	
Cultivation between and within rows	95	5.6	0.9973	25	11.0	< 0.0001	94	6.6	0.9969	
Black oat 'SoilSaver'	94	5.9	1.0000	98	11.0	0.9998	98	6.6	0.8980	
Black oat 'As 033'	92	5.1	0.8737	97	11.0	1.0000	99	6.6	0.8398	
AU Homer										
S-metolachlor	90	5.8		99	11.0		99	6.6		
Cultivation between rows	64	8.9	0.0090	24	11.0	< 0.0001	99	6.6	1.0000	
Cultivation between and within rows	87	5.7	0.9286	23	11.0	< 0.0001	99	6.6	1.0000	
Black oat 'SoilSaver'	85	6.1	0.6616	99	11.0	1.0000	99	6.6	1.0000	
Black oat 'As 033'	94	5.3	0.5368	96	11.7	0.9995	99	7.7	1.0000	

treatment (0.97). Crop injury increased over time. At the second rating only between-row cultivation (1.04) and black oat cultivar As 033 (0.87) caused lower injury than the *S*-metolachlor treatment (1.61). Overall, crop injury was negligible and well below reported thresholds.

## Lupin density

The interactions and treatment main effects were nonsignificant (P = 0.10). Plant density of the cultivars decreased over time from rating 1 to 3. ABL 1082 plant density was 9.8, 9.1 and 8.8 plants m<sup>-2</sup> for ratings 1, 2 and 3, respectively. Density of AU Alpha was 10.6, 9.5 and 7.6 plants m<sup>-2</sup>. AU Homer was 10.6, 9.6 and 9.3 plants m<sup>-2</sup>. The slight decrease in plant density may be related to plant death over time due to insects, deer browsing and disease.

#### Lupin maturity

The interactions and treatment main effects were nonsignificant (P = 0.10). At the March rating it was observed that ABL 1082 is slightly later maturing because 82% of the plants were in full bloom as compared to 89% for AU Alpha and AU Homer. This is consistent with the breeding history of these cultivars (van Santen, 2006, unpublished results).

#### Lupin yield

The treatment by cultivar interaction was significant for test weight and yield, but not for seed mass. Treatment main effects were significant for test weight, seed mass and yield. Plants of ABL 1082 treated with S-metolachlor had a test weight of 79.6 kg 100 liter<sup>-1</sup>. Only black oat As 033 (78.6 kg 100 liter<sup>-1</sup>) resulted in a lower test weight (Table 3). Neither cultivation nor black oat cultivar treatments altered the mean seed mass of ABL 1082 as compared to S-metolachlor (210 mg seed<sup>-1</sup>). ABL 1082 plants treated with the conventional treatment had a grain yield of 1540 kg ha<sup>-1</sup>. With the exception of between-row cultivation (1904 kg ha<sup>-1</sup>) none of the organic treatments significantly increased the yield of ABL 1082.

The test weight of AU Alpha treated with *S*-metolachlor was 77.5 kg 100 liter<sup>-1</sup> (Table 3). Both black oat cultivars reduced the test weight ( $\leq$ 76.3 kg 100 liter<sup>-1</sup>) compared to *S*-metolachlor. None of the organic treatments either increased or decreased mean seed mass in comparison to *S*-metolachlor (227 mg seed<sup>-1</sup>). AU Alpha treated with *S*-metolachlor yielded 1126 kg ha<sup>-1</sup>. Neither cultivation

**Table 3.** Test weight (kg  $100 \text{ liter}^{-1}$ ), seed mass (mg seed<sup>-1</sup>) and yield (kg ha<sup>-1</sup>) of L. albus L. as affected by treatment.

Treatment	Test wei	ght	Seed 1	nass	Grain yield		
Name	Class	Mean (kg 100 liter <sup>-1</sup> )	Dunnett's <i>P</i> -value	Mean (mg seed <sup>-1</sup> )	Dunnett's <i>P</i> -value	Mean (kg ha <sup>-1</sup> )	Dunnett's <i>P</i> -value
ABL 1082							
S-metolachlor	Conventional	79.6		210		1543	
Cultivation between rows	Organic	80.1	0.800	217	0.445	1904	0.013
Cultivation between and within rows	Organic	80.2	0.610	214	0.886	1678	0.690
Black oat 'SoilSaver'	Organic	79.5	0.996	202	0.482	1424	0.779
Black oat 'As 033'	Organic	78.6	0.085	213	0.959	1290	0.138
Non-treated	Control	80.1	0.715	215	0.783	1705	0.524
AU Alpha							
S-metolachlor	Conventional	77.5		227		1126	
Cultivation between rows	Organic	77.7	0.985	231	0.895	1214	0.919
Cultivation between and within rows	Organic	77.5	1.000	233	0.722	1151	1.000
Black oat 'SoilSaver'	Organic	76.2	0.145	230	0.954	939	0.390
Black oat 'As 033'	Organic	76.3	0.068	233	0.744	909	0.296
Non-treated	Control	77.6	0.993	233	0.674	1330	0.305
AU Homer							
S-metolachlor	Conventional	76.4		222		849	
Cultivation between rows	Organic	76.5	0.997	231	0.278	1020	0.473
Cultivation between and within rows	Organic	76.3	0.999	236	0.046	1018	0.482
Black oat 'SoilSaver'	Organic	76.3	0.999	235	0.070	934	0.931
Black oat 'As 033'	Organic	76.9	0.672	232	0.246	1067	0.247
Non-treated	Control	76.3	1.000	236	0.034	887	0.998
SE		0.48		11.6		232.4	

nor black oat cultivars influenced the test weight of AU Homer as compared to *S*-metolachlor (76.4 kg 100 liter<sup>-1</sup>) (Table 3). Plants treated with *S*-metolachlor resulted in a seed mass of 222 mg seed<sup>-1</sup>. Seed mass was higher in non-treated plants (236 mg seed<sup>-1</sup>), as compared to the seed mass of plants in which between–within row cultivation (236 mg seed<sup>-1</sup>) and black oat cultivar SoilSaver (235 mg seed<sup>-1</sup>) were used. None of the organic treatments reduced or increased the grain yield of AU Homer as compared to *S*-metolachlor (849 kg ha<sup>-1</sup>).

# **Discussion and Conclusion**

Our results show that between and within row cultivation successfully reduced most weed species that were present in our experiment to more than 80%, including Shepherd's purse, annual ryegrass, winter vetch and heartwing sorrel. The cultivation treatments, between row and between-within row (hoeing) yielded as high or higher than the non-treated control and higher than *S*-metolachlor. This coincides with observations made in lentil<sup>13</sup>, in which hand-hoed plots yielded higher than chemical weed control. Test weight and seed mass were not reduced or increased by cultivation treatments. Yield potential may be higher in these treatments since crop injury was lower compared to the *S*-metolachlor treatment.

The two black oat cultivars provided very good control of annual ryegrass, shepherd's purse and yellow nutsedge (>90%), but were not successful in the control of other weed species, especially corn spurry (Spergula arvensis L.). Black oat can be grown as a companion crop in lupin, growing in the same field at the same time. In conventional farming systems, the black oat companion may be terminated by a selective grass herbicide (i.e. aryloxyphenoxypropionates and/or cyclohexanediones) prior to lupin harvest. Black oat, especially SoilSaver, out-competes weeds by shading due to its large biomass production<sup>15</sup>. Black oat also produces allelopathic compounds that inhibit weed growth<sup>15</sup>. This is particularly important since lupins have a slowly developing canopy during their early establishment which inhibits their ability to shade weeds themselves. Neither of the black oat cultivars resulted in crop injury. This may indicate that white lupin, unlike cotton<sup>16</sup>, is not sensitive to the allelopathic activity of black oat. Neither of the black oat companions affected the yield of any lupin cultivar.

Stand count reductions appear more closely related to yield loss than other parameters, including crop injury<sup>19</sup>. Based on previous research it was found that crop injury greater than 3 is unacceptable in white lupin<sup>20</sup>.

We conclude that organic production of white lupin in the southeastern USA is possible. The cultivation Organic weed control in white lupin (Lupinus albus L.)

treatments and companion crops used in this experiment provide satisfactory to excellent weed control without causing unacceptable crop injury. Plants treated with these organic methods yielded as high or higher than the chemical control, which makes the production of organic seed feasible.

# References

- 1 Clark, M.S., Horwath, W.R., Shennan, C., Scow, K.M., Lantni W.T., and Ferris, H. 1999. Nitrogen, weeds and water as yield-limiting factors in conventional, low-input, and organic tomato systems. Agriculture, Ecosystem and Environment 73:257–270.
- 2 Cornell Cooperative Extension Publication. 2009. Integrated Crop and Pest Management Guidelines for Commercial Vegetable Production 2009. Available at Web site http:// www.nysaes.Cornell.edu/recommends/11frameset.html (verified June 30, 2010).
- 3 Dierauer, H., Böhler, D., Kranzler, A., and Zollitsch, W. 2004. Merkblatt Lupinen 2004. Ausgabe Österreich © BIO ERNTE AUSTRIA & FiBL.
- 4 Payne, W.A., Chen, C., and Ball, D.A. 2004. Alternative crops agronomic potential of alternative crops agronomic potential of narrow-leafed and white lupins in the Inland Pacific Northwest. Agronomy Journal 96:1501–1508.
- 5 van Santen, E. and Reeves, D.W. 2003. Tillage and rotation effects on lupin in double-cropping systems in the southeastern USA. In E. van Santen and G.D. Hill (eds). Wild and Cultivated Lupins from the Tropics to the Poles. Proceedings of the 10th International Lupin Conference, Laugarvatn, Iceland, 19–24 June 2002. International Lupin Association, Canterbury, New Zealand.
- 6 Noffsinger, S.L. and van Santen, E. 2005. Evaluation of *Lupinus albus* L. Germplasm for the Southeastern USA. Crop Science 45:1941–1950.
- 7 Putnam, D.H., Oplinger, E.S., Hardman, L.L., and Doll, J.D. 2007. Lupine, Alternative Field Crops Manual. University of Wisconsin-Extension, Cooperative Extension. University of Minnesota: Center for Alternative Plant and Animal Products and the Minnesota Extension Service. Available at Web site http://www.hort.purdue.edu/newcrop/afcm/lupine.html (verified June 30, 2010).
- 8 Poetsch, J. 2006. Pflanzenbauliche Untersuchungen zum ökologischen Anbau von Körnerleguminosen an sommertrockenen Standorten Südwestdeutschlands. Institut für Pflanzenbau und Grünland der Universität Hohenheim, Salzgitter.

Available at Web site http://opus.ub.uni-hohenheim.de/volltexte/2007/193/2,4-DB (verified June 30, 2010).

- 9 Crop Protection Referenc. 2007. Greenbook's Crop Protection Reference. 23rd ed. Vance Publishing Corporation, Lenexa, KS.
- 10 Mitich, L.W., Cassman, K.G., Larson, K.J., and Smith, N.L. 1987. Evaluation of preemergence herbicides for control of winter annual weeds in "Minnesota Ultra" lupins. Research Progress Report, p. 222–223.
- 11 Noffsinger, S.L. 1998. Physiology and management of wintertype white lupin (*Lupinus albus* L.). PhD Dissertation, Auburn University, Auburn, AL.
- 12 Noffsinger, S.L., Huyghe, C., and van Santen, E. 2000. Analysis of grain-yield components and inflorescence levels in winter-type white lupin. Agronomy Journal 92: 1195–1202.
- 13 Anderson, W.P. 1996. Weed Science: Principles and Applications, 3rd ed. West Publishing Company, St. Paul, MN.
- 14 Sandhu, P.S., Dhingra, K.K., Bhandari, S.C. and Gupta, R.P. 1991. Effect of hand-hoeing and application of herbicides on nodulation, nodule activity and grain yield of *Lens culinaris* Med. Plant and Soil 135:293–296.
- 15 Bowman, G., Shirley, C. and Cramer, C. 1998. Managing Cover Crops Profitably, 2nd ed. Sustainable Agriculture Network Handbook Series Book. 3. Sustainable Agriculture Network, National Agricultural Library, Beltsville, MD.
- 16 Price, A.J., Stoll, M.E., Bergtold, J.S., Arriaga, F.J., Balkcom, K.S., Kornecki, T.S., and Raper, R.L. 2008. Effect of cover crop extracts on cotton and radish radicle elongation. Communications in Biometry and Crop Science 3:60–66. Available at Web site http://agrobiol.sggw.waw.pl/~cbcs/ pobierz.php?plik=CBCS\_3\_1\_6 (verified June 30, 2010).
- 17 CTAHR—College of Tropical Agriculture and Human Resources. University of Hawai'i. Sustainable Agriculture in Hawai'i. 2002. Green Manures: non-legumes. Black oat (Avena strigosa). Available at Web site http://www.ctahr. hawaii.edu/sustainag/Database.asp (verified June 30, 2010).
- 18 Murphy, T.R. Preemergence Control of Winter Weeds. University of Georgia. Available at Web site http:// commodities.caes.uga.edu/turfgrass/georgiaturf/WeedMngt/ weedcontrol/turfwin.PDF (verified June 30, 2010).
- 19 Taylor-Lovell, S., Wax, L.M., and Nelson, R. 2001. Phytotoxic response and yield of soybean (*Glycine max*) varieties treated with sulfentrazone or flumioxazin. Weed Technology 15:95–102.
- 20 Knott, C.M. 1996. Tolerance of autumn-sown determinate lupins (*Lupinus albus*) to herbicides. Test of agrochemicals and cultivars 17. Annals of Applied Biology 128 (Supplement): 52–53.