

Statins and myocardial remodelling: cell and molecular pathways

Karen E. Porter* and Neil A. Turner

The advent of statins has revolutionised the treatment of patients with raised plasma cholesterol and increased cardiovascular risk. However, the beneficial effects of this class of drugs are far greater than would be expected from lowering of cholesterol alone, and they appear to offer cardiovascular protection at multiple levels, primarily as a result of their pleiotropic activity. Indeed, their favourable effects on the heart seem to be mediated in part through reduced prenylation and subsequent inhibition of small GTPases, particularly those of the Rho family. Such statin-mediated effects are manifested by reduced onset of heart failure and improvements in cardiac dysfunction and remodelling in heart failure patients. Experimental studies have shown that statins mediate their effects on the two major resident cell types of the heart—cardiomyocytes and cardiac fibroblasts—and thus facilitate improvement of adverse remodelling of ischaemic or non-ischaemic aetiology. This review examines evidence for the cellular effects of statins in the heart, and discusses the underlying molecular mechanisms at the level of the cardiomyocyte (hypertrophy, cell death and contractile function) and the cardiac fibroblast (differentiation, proliferation, migration and extracellular matrix synthesis). The prospects for future therapies and ongoing clinical trials are also summarised.

The earliest link between cholesterol deposition in arterial walls and death from coronary heart disease (CHD) was documented by Virchow over a century ago (Ref. 1), but it was not until the 1950s that prospective examination of the relationship between levels of plasma LDL (low-density lipoprotein)-C (cholesterol) and CHD was reported by the Framingham Study (Ref. 2). It is beyond doubt that the two are intrinsically linked, and there is a direct

correlation between the levels of plasma LDL-C and the risk of CHD (Ref. 3). The elucidation of the cholesterol synthetic pathway has facilitated the development of therapeutic strategies for lipid lowering and, as a result, hydroxymethylglutaryl coenzyme A (HMG-CoA) reductase inhibitors (statins) first came into clinical use during the 1980s. Statins inhibit the rate-limiting step in intracellular cholesterol synthesis, that of the conversion of HMG-CoA

Division of Cardiovascular and Neuronal Remodelling, Leeds Institute of Genetics, Health and Therapeutics (LIGHT) and Multidisciplinary Cardiovascular Research Centre (MCRC), University of Leeds, Leeds, UK

*Corresponding author: Karen E. Porter, Division of Cardiovascular and Neuronal Remodelling, Worsley Building, Clarendon Way, University of Leeds, Leeds LS2 9JT, UK. Email: medkep@leeds.ac.uk

to L-mevalonate, by binding to the active site of HMG-CoA reductase (Ref. 4). This effectively decreases cholesterol synthesis by the liver, leading to upregulated LDL receptor expression, with consequent increased clearance of cholesterol from the plasma.

The first statins used clinically were lovastatin and simvastatin, the latter being evaluated in a landmark trial – the Scandinavian Simvastatin Survival Study (4S) (Ref. 5). This study revealed an unequivocal reduction in all-cause mortality in 4444 hypercholesterolaemic patients with CHD who were followed up for 5 years, and provided the first definitive evidence of improved survival with statin therapy. Over the past two decades, statin therapy has become widely prescribed, and these drugs are probably the most thoroughly investigated primary therapeutic agents in clinical trials (Ref. 6).

Statins in clinical trials

In the 4S study, the ability of statins to reduce mortality in individuals with a history of CHD was considered to be attributable entirely to their cholesterol-lowering properties. These conclusions were supported by two further large studies, CARE (Ref. 7) and LIPID (Ref. 8), that evaluated the outcomes of cholesterol lowering in approximately 18 000 patients treated with pravastatin. These two studies also described a dramatic and consistent effect in patients with pre-existing CHD but without elevated cholesterol. A subsequent study of more than 20 000 patients in the UK provided significant new knowledge in this respect. The seminal findings of the Heart Protection Study (HPS) concluded that substantial benefit from statin therapy was apparent in individuals not only with existing CHD, but also with diabetes, cerebrovascular disease and peripheral arterial disease (Ref. 9). The study clearly showed that simvastatin therapy was equally beneficial to individuals with high and ‘normal’ levels of cholesterol and therefore raised the question of whether these benefits could be entirely attributable to effects on cholesterol alone.

A large number of subsequent clinical trials have concluded beyond doubt that statin therapy has proven effectiveness in decreasing CHD and reducing cardiovascular mortality. Subgroup analysis of some of these previous trials demonstrated that effects beyond lipid lowering were apparent. In the WOSCOPS

(Ref. 10) and CARE (Ref. 7) trials, risk of CHD was significantly reduced in statin-treated age- and sex-matched individuals with similar absolute levels of cholesterol. In 2005, the Cholesterol Treatment Trialists (CTT) Collaborators (Ref. 11) performed a meta-analysis of 14 randomised studies involving over 90 000 participants followed for 5 years. These collated data clearly demonstrated that statin therapy reduced the incidence of major coronary events, coronary revascularisation and stroke, and was related to absolute reduction in cholesterol, irrespective of initial levels. It is believed that half of all cases of myocardial infarction (MI) and stroke occur in apparently healthy individuals with plasma cholesterol levels below the threshold for lipid-lowering treatment (Ref. 12), suggesting that other contributing factors have a role. The JUPITER trial was a large randomised study conducted in 26 countries, recruiting almost 18 000 apparently healthy men and women with no history of cardiovascular disease. Although these individuals had ‘normal’ cholesterol levels, they exhibited increased levels of C-reactive protein (CRP), an inflammatory biomarker proposed to be predictive of future vascular events. Administration of rosuvastatin significantly reduced the rate of primary cardiovascular events in these patients (Ref. 12), the results being so clear-cut (major cardiovascular events in rosuvastatin group 142, versus 251 in placebo) that the trial was terminated after a median follow-up of 1.9 years (maximal 5 years).

The unequivocal outcomes of several clinical studies supported the widespread prescribing of statins to individuals at risk, such that some investigations have adopted more aggressive approaches to lipid lowering. Several studies, including PROVE-IT (Ref. 13), TNT (Ref. 14), IDEAL (Ref. 15), SEARCH (Ref. 16) and A to Z (Ref. 17), have reported that high-dose statin therapy can further reduce cardiovascular risk compared with standard regimens. These data from a total of almost 40 000 patients are collated in the most recent meta-analysis of 26 statin trials by the CTT Collaborators, comparing intensive versus regular statin treatment (Ref. 18).

Clinical evidence for statin pleiotropy

Pleiotropy refers to actions of a drug that extend beyond that for which it was specifically

developed. Such effects are generally unexpected and can be related to the drug's primary mode of action or may possibly be unrelated (Ref. 19). It has traditionally been assumed that the key benefits of statin therapy are attributable to the lowering of serum cholesterol levels, but the concept of pleiotropism first arose through retrospective analysis of some early clinical trials. In particular, in the 4S and TNT trials, a reduction in the incidence, morbidity and mortality of heart failure was reported (Refs 14, 20). In another clinical study, statin treatment for 14 weeks led to improved exercise endurance that corresponded to a significant increase in left ventricular (LV) ejection fraction and a decrease in plasma inflammatory cytokines: effects not observed in the placebo control group (Ref. 21).

Further support for the notion that additional mechanisms of action exist with statin therapy is that the risk of MI in statin-treated patients is lower than that of patients receiving alternative cholesterol-lowering strategies (Refs 22, 23, 24). For example, ezetimibe, a drug that lowers cholesterol by impeding intestinal absorption, does not offer equivalent cardioprotective effects to statins despite comparable reductions in serum cholesterol (Refs 25, 26). In the JUPITER trial, the significance of inflammatory mediators as a marker of future cardiovascular events was highlighted and the measured reduction in CRP following statin therapy was unrelated to a reduction in cholesterol levels. Moreover, the patients enrolled in that study had no history of cardiovascular disease and had cholesterol levels in the normal range (Ref. 12). In summary, accumulating evidence supports the existence of statin pleiotropy in the cardiovascular system. To what extent these effects justify their clinical benefits in the heart is less clear-cut but will now be reviewed in the light of current data.

Statins and the heart

The beneficial impact of statin therapy on cholesterol reduction is beyond doubt, and recognition of its far-reaching effects beyond lipid lowering is now widely documented. These properties include restoration of aberrant endothelial function, reducing oxidative stress, stabilisation of atherosclerotic plaques, inhibition of platelet aggregation, inhibition of vascular smooth muscle proliferation and migration, and reducing vascular inflammation (reviewed in Ref. 23).

Although most knowledge has been acquired through studies in vascular tissue and cells, data from clinical studies have provided evidence that statins also exert direct effects on the myocardium. In the Western world, congestive heart failure is a key cause of morbidity and mortality, and there are now several lines of evidence supporting the use of statins in heart failure patients. Indeed, it was retrospective analysis of the 4S study which showed that patients on long-term simvastatin therapy had reduced incidence of heart failure (Ref. 20). Statins reduce LV mass in hypertensive patients (Ref. 27) and lower the incidence of atrial fibrillation in patients with coronary artery disease (Ref. 28). Most large statin trials have subsequently documented favourable outcomes in heart failure patients (reviewed in Ref. 29), perhaps unsurprisingly, owing to the underlying commonality of atherosclerotic disease in ischaemic heart failure (Ref. 14). However, statins are also beneficial in patients with heart failure of non-ischaemic aetiology (Refs 21, 30, 31). Such observations support a mechanism that is independent of cholesterol and are reportedly additional to effects achieved with standard heart failure therapies such as ACE inhibitors and β -blockers (Ref. 32).

The consensus view of clinical studies with statins in patients with heart failure is the exhibition of several clinical and biological effects with a diversity of reported mechanisms: anti-inflammatory, antioxidant, improvements in endothelial function, increased angiogenesis, reduced cardiac hypertrophy and remodelling, reduced neurohormonal activation, improved autonomic regulation and anti-arrhythmic effects (reviewed in Ref. 32). However, whereas small prospective studies and post-hoc analyses of randomised trials support the notion that statins are effective in patients with heart failure, the recent randomised controlled GISSI-HF study specifically addressed their use in patients with heart failure. In this prospective trial of more than 4600 patients, it was reported that the standard 10 mg administration of rosuvastatin had no beneficial effect, irrespective of heart failure aetiology (Ref. 33). It is perhaps pertinent to note that the broad population of patients recruited to this study had established, symptomatic heart failure and were receiving traditional heart failure therapies. Therefore, it is conceivable that addition of statin therapy is less

effective in reducing cardiovascular events when heart failure is established, as opposed to prevention of heart failure in a population with CAD. This suggestion is endorsed by studies in which different statin regimens led to a consistent reduction in the development of heart failure in CAD patients (Refs 17, 20, 34, 35). Some studies propose that statin therapy may be beneficial early after acute ischaemic events that are associated with LV dysfunction and failure (Ref. 36). In further support of this proposition, statins do not appear to improve outcomes in patients with advanced heart failure (Ref. 37). The use of statins as an additional preventive strategy for patients at risk of atrial fibrillation does however appear to be beneficial (reviewed in Ref. 38).

Outside the clinical scenario, the weight of evidence from animal studies clearly supports the concept that statins confer beneficial effects on adverse myocardial remodelling. These include improvements in LV remodelling, hypertrophy and fibrosis, with resultant enhanced cardiac function (Refs 39, 40, 41, 42, 43). Regardless of the initiating stimulus (aortic banding, MI or a variety of transgenic models), the benefits of statin administration were clear. These studies were conducted in normocholesterolaemic animals with no induction of atherosclerosis; therefore, the documented effects were independent of correcting hyperlipidaemia. Rather, the mechanism of the effects of statins appears to be due to inhibition of small GTPases, which are important modulators of myocardial function.

Evidence for small GTPases in mediating myocardial remodelling

The multiplicity of reported pleiotropic effects of statins appears to be attributable to their ability to inhibit synthesis of intracellular mevalonate: a key upstream precursor of not only cholesterol, but also important isoprenoid intermediates, farnesyl pyrophosphate (FPP) and geranylgeranyl pyrophosphate (GGPP) (Ref. 44) (Fig. 1). These intermediates have key roles in post-translational modification of a range of proteins, including small GTPases (Ras, Rho, Rac), by serving as lipid attachments through a process known as prenylation. In this respect, isoprenoid moieties permit attachment, subcellular localisation and trafficking of membrane-bound proteins coupled to

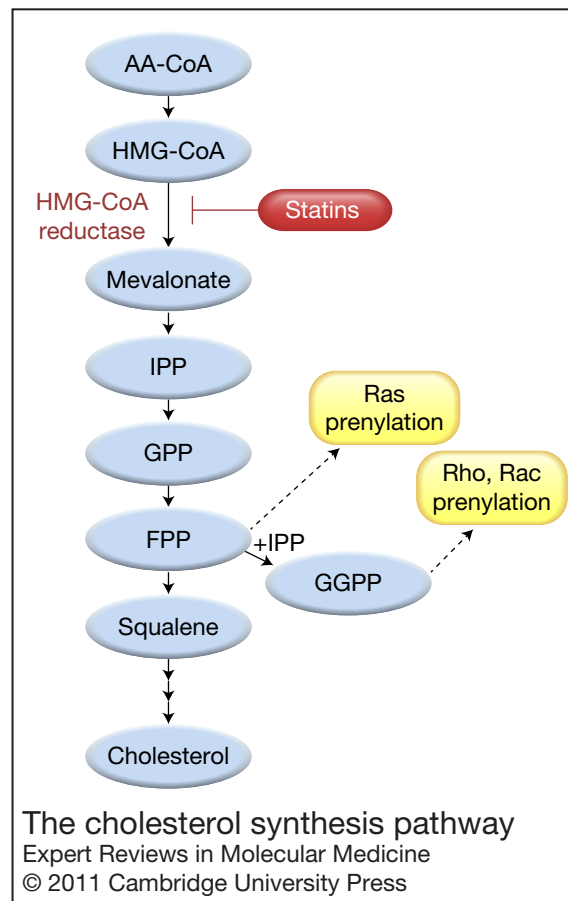


Figure 1. The cholesterol synthesis pathway. Statins inhibit HMG-CoA reductase, the rate-limiting enzyme in cholesterol synthesis. Resultant inhibition of FPP and GGPP synthesis inhibits prenylation of small GTPases of the Ras and Rho (Rho/Rac/cdc42) families, respectively, resulting in pleiotropic effects that are independent of effects on cholesterol synthesis. Abbreviations: AA-CoA, acetoacetyl coenzyme A; HMG-CoA, hydroxymethylglutaryl coenzyme A; IPP, isopentenyl pyrophosphate; GPP, geranyl pyrophosphate; FPP, farnesyl pyrophosphate; GGPP geranylgeranyl pyrophosphate.

intracellular signalling pathways, thus facilitating changes in cellular function (Ref. 45). The key role of prenylated proteins in the control of cellular function significantly affects cellular homeostatic activity in which most processes are regulated, either directly or indirectly, by Rho or Ras GTPases (Ref. 46). In addition, Rac1 GTPase contributes to myocardial pathologies both in experimental studies (Refs 47, 48) and in humans (Ref. 49). Hence it is likely, through modulation of such mechanisms,

that statins confer beneficial pleiotropic effects. Consistent with this proposal, many of the inhibitory effects of statins *in vitro* are reversible by isoprenoid (FPP/GGPP) supplementation, but not by cholesterol itself, or its precursor squalene (Fig. 1).

It is now well established that small GTPases have important roles in regulating myocardial remodelling. Studies performed over a decade ago first demonstrated the importance of small GTPases, particularly Ras and Rho family proteins, in signalling from G-protein-coupled receptors to development of myocardial hypertrophy (reviewed in Refs 50, 51). Interestingly, Ras, Rho and Rac have all been implicated in promoting cardiac hypertrophy (Ref. 52). For example, typical hypertrophic agents such as phenylephrine and endothelin-1 (ET-1) have been shown to activate Rac1 through specific intracellular signalling pathways in cardiac myocytes (Ref. 53). In another study, cardiac-specific deletion of Rac1 in a murine model reduced angiotensin II (Ang II)-mediated hypertrophy (Ref. 54); conversely, cardiac-specific Rac1 overexpression led to atrial fibrillation (Ref. 47). Interestingly, increased Rac1 activity has been associated with cardiac hypertrophy in humans (Ref. 55).

Transgenic mice with cardiac-specific expression of RhoA, whilst showing no obvious hypertrophy, had clear evidence of heart failure accompanied by oedema, ventricular and atrial dilation, and increased fibrosis (Ref. 56). Rho kinase (ROCK) is a well-characterised downstream effector of RhoA, and studies in which ROCK was inhibited have indicated a role in diverse cardiovascular pathologies. Pharmacological inhibition of ROCK implicates it in the development of cardiac hypertrophy in rats (Refs 57, 58) and mice (Ref. 59). In murine studies, long-term inhibition of ROCK suppressed LV remodelling after MI (Ref. 60) and protected from ischaemia-reperfusion injury (Ref. 61). In transgenic models, ROCK1 haploinsufficient mice showed decreased cardiac fibrosis after MI (Ref. 62), and targeted deletion of ROCK significantly reduced fibrosis after three weeks of pressure overload (Ref. 63).

The *in vivo* function of Ras was first studied in transgenic mouse models (Ref. 64) in which animals displayed characteristic hypertrophy with features in common with human disease (Ref. 65). Indeed, a correlation between Ras

expression and the severity of hypertrophy has been reported in humans (Ref. 66).

Given the established roles of small GTPases in the aetiology of heart failure and other cardiac pathologies, inhibition of their activity by statins almost certainly underlies many of the pleiotropic effects of these drugs.

Effects of statins on resident myocardial cells

Evidence collected from animal models indicates that statins can ameliorate adverse myocardial remodelling largely through effects on Ras, Rho and Rac, but their precise site of action and specific cellular effects are difficult to elucidate in this setting. It is established that activation of small GTPase signalling in the heart is detrimental to cardiac function, but what is less clear is how blocking these pathways leads to beneficial effects at the cellular level. In the mammalian heart, cardiac function is dynamically regulated by the two key resident cell types, cardiomyocytes and cardiac fibroblasts (CFs). The normal human heart is composed of 30% cardiomyocytes and 70% nonmyocytes, the majority of which are CFs (Ref. 67). CFs are primarily responsible for extracellular matrix (ECM) homeostasis and regulate the structure of the heart, thereby providing coordinated signals between cellular and noncellular components (reviewed in Ref. 68). Cardiomyocytes, although fewer in number, occupy the bulk volume and provide mechanical force, transmission of which is a key function of the ECM. It is acknowledged that statins can also modulate other nonmyocyte cells resident in the heart, including endothelial cells, smooth muscle cells, neuronal cells and mast cells. However, these have been recently reviewed elsewhere (Ref. 69) and will not be discussed further here.

Historically, cardiac myocytes have been the key focus of clinical and experimental studies *in vivo* and *in vitro*. Although CFs have received considerably less attention, their role in regulating and maintaining the structural integrity of the heart through controlled proliferation and ECM turnover is pivotal to cardiac function (Refs 68, 70, 71, 72). CFs respond to a variety of stimuli (mechanical, electrical and chemical) to coordinate cell–cell and cell–matrix interactions in the maintenance of cardiac homeostasis (Refs 73, 74). CFs

regulate ECM turnover by balancing the synthesis of ECM components (collagens, laminins, glycosaminoglycans, matricellular proteins) with their breakdown, through the production of an array of specific proteases, including members of the matrix metalloproteinase (MMP) family (Ref. 68). Although electrically unexcitable, CFs are coupled to myocytes and can influence myocyte electrophysiology (Ref. 75).

The heart undergoes adaptive remodelling in response to pathophysiological stresses, including hypertension-associated pressure overload and ischaemia-reperfusion injury after MI: changes that in the longer term can progress to pathological remodelling and heart failure. At the level of the cardiomyocyte, these adaptive changes include loss of contractile function (through changes in contractile protein expression, excitation–contraction coupling and desensitisation to β -adrenergic signalling), increased cell hypertrophy, and augmented apoptotic and necrotic cell death (Ref. 76). Concurrently, CFs undergo phenotypic transformation to become myofibroblasts, display an increased propensity for cell proliferation and migration, and modulate ECM turnover through elevated MMP expression and enhanced collagen deposition (fibrosis) (Ref. 68). Therefore, complex alterations in both myocyte and fibroblast biology occur during adaptive and pathological myocardial remodelling.

Given this heterogeneity, it is clear that multiple cellular and molecular mechanisms underlie the benefits of statins on myocardial remodelling. Investigation of the modulatory effects of statins on individual myocardial cell types in vitro has therefore shed more light on their cellular and molecular mechanisms, as reviewed below.

Effects of statins on cardiac myocytes

In vivo and in vitro studies have provided evidence that statins can reduce the detrimental effects of myocardial injury on cardiomyocyte function at multiple levels, including hypertrophy, cell death and contractile function (Fig. 2a). The use of in vitro cell cultures in particular has enabled delineation of intracellular mechanisms of action of statins on these aspects of myocyte function. It should be noted, however, that the vast majority of these in vitro studies have utilised cultures of neonatal cardiomyocytes as model systems rather than

mature differentiated adult cardiomyocytes, so an element of caution is warranted in translating these findings to the adult in vivo system, particularly that of man.

Hypertrophy

Statins are widely accepted to have beneficial effects by reducing cardiac hypertrophy in experimental animal models (Ref. 77). In vitro studies on the effects of statins on markers of neonatal cardiomyocyte hypertrophy have also shown that the stimulatory effects of hypertrophic agonists (e.g. Ang II, ET-1, NE) are opposed by lipophilic statins, but not by the hydrophilic pravastatin which appears to have no effect in vitro (Refs 78, 79). The underlying mechanisms by which inhibition of HMG-CoA reductase modulates the hypertrophic response are mostly cholesterol independent, involving inhibition of GTPase prenylation (summarised in Fig. 3), particularly those of the Rho and Rac family that are integral to the hypertrophic mechanism (Ref. 52). By preventing geranylgeranylation and membrane association of RhoA, statins prevent downstream signalling to Rho substrates including ROCK, which is an important regulator of the actin cytoskeleton and cell size (Refs 79, 80, 81). The small GTPase Rac1 is crucial for activation of NADPH oxidase, a mediator of Ang II and ROS-mediated myocyte hypertrophy (Ref. 82). By reducing Rac1 geranylgeranylation, statins attenuate NADPH oxidase activity and subsequent Ang-II-induced myocyte hypertrophy (Refs 48, 80, 83). The ability of statins to inhibit farnesylation of the small GTPase p21Ras, and thus downstream ERK signalling, might also contribute to statin-mediated reductions in myocyte hypertrophy (Ref. 78). More recent studies have focused on the potential effects of statins on additional prenylated proteins, including large G-proteins that may modulate cellular responses to hypertrophic stimuli. For example, rosuvastatin was shown to inhibit α 1-adrenergic-receptor-induced cardiomyocyte hypertrophy by suppressing expression and membrane association of the large G-protein $G_{1\beta}$, resulting in reduced PKC–ERK signalling (Ref. 84). This is significant because it has been shown that coupling of the α 1-receptor to $G_{1\beta}$ can aggravate cardiac remodelling in the failing human heart (Ref. 85).

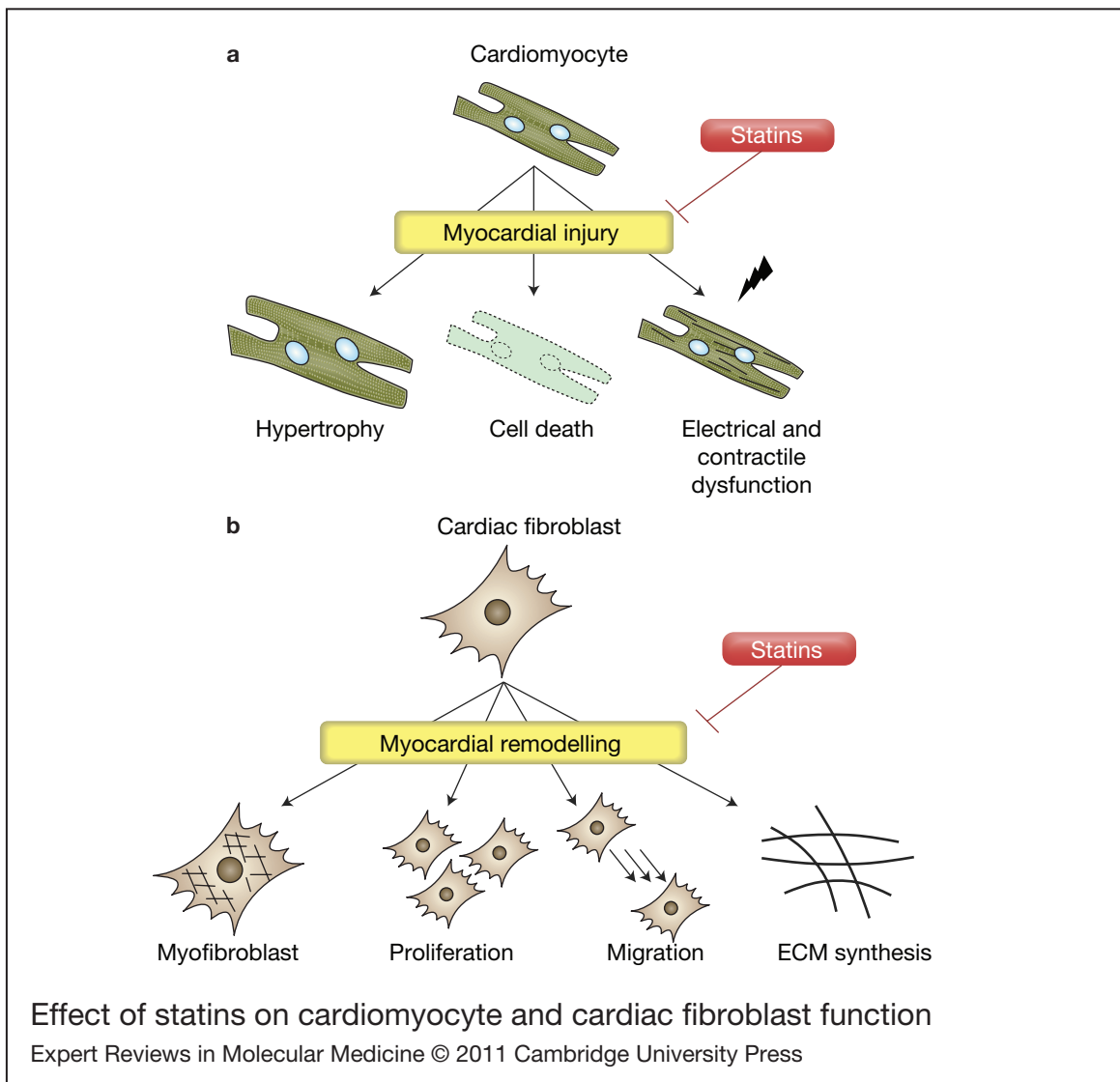


Figure 2. Effect of statins on cardiomyocyte and cardiac fibroblast function. Summary of some of the most well-established effects of statins on (a) cardiomyocyte and (b) cardiac fibroblast (CF) function. Statins can reduce cardiomyocyte hypertrophy and cell death, and improve electrical and contractile function following myocardial injury. Statins can also attenuate CF proliferation, migration, myofibroblast differentiation and extracellular matrix (ECM) synthesis.

Additional mechanisms of action of statins on hypertrophic signalling pathways have also been reported, which appear to be initiated independently of effects on G-proteins, including direct modulation of the JAK–STAT (Refs 86, 87) and PI3K–AKT (Refs 88, 89) pathways.

Cell death

Therapies aimed at reducing cardiomyocyte cell death hold promise in reducing infarct size and improving clinical outcomes (Refs 90, 91).

Statins are reported to exert cardioprotective effects by reducing myocyte cell injury and death (Ref. 91). In vitro studies have investigated the underlying mechanisms at the level of the cardiomyocyte and current thinking is that statins activate the PI3K–AKT pathway (Refs 92, 93) and other components of the reperfusion injury salvage kinase (RISK) pathway (Ref. 94), leading to increased NO synthesis (Refs 92, 95) and reduced mitochondrial dysfunction (Ref. 95). Rac1 might

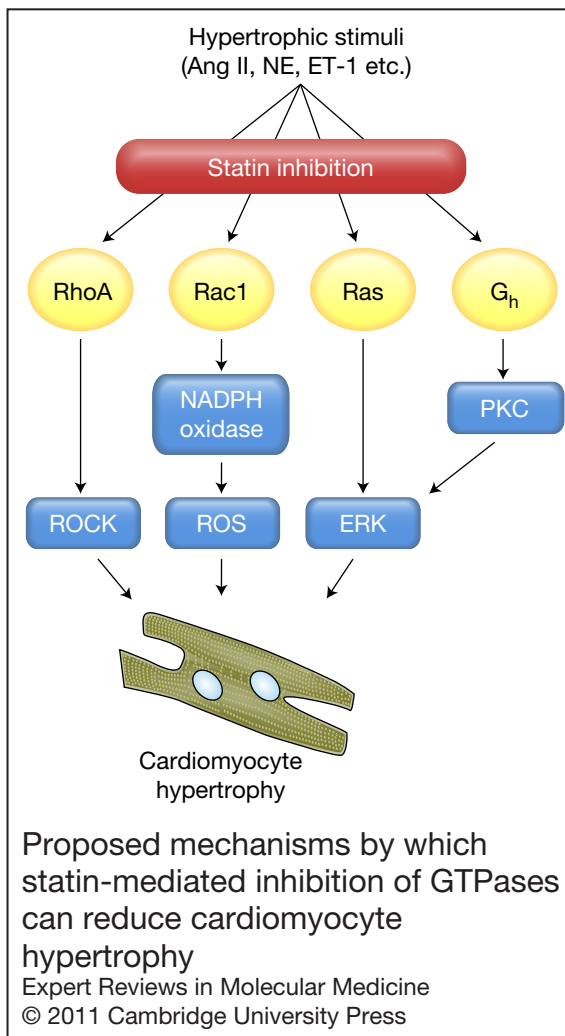


Figure 3. Proposed mechanisms by which statin-mediated inhibition of GTPases can reduce cardiomyocyte hypertrophy. Statins inhibit prenylation of small GTPases of the Ras, Rho and Rac families, resulting in reduced cardiomyocyte hypertrophy. Statins may also modulate large G-protein function (e.g. G_h) to similar effect.

also be a target for the antiapoptotic effects of statins, as cerivastatin reduced β -adrenergic-receptor-induced apoptosis and Rac1 and JNK activity in adult rat cardiomyocytes (Ref. 96).

Despite these studies, the impact of statins on cardiomyocyte apoptosis is not as clear-cut as it is for myocyte hypertrophy, and several other studies have suggested that rather than reducing apoptosis, statins may actually augment myocyte death. For example, simvastatin promoted apoptosis of adult human cardiomyocytes, effects that were prevented

by co-supplementation with mevalonate or GGPP, indicative of a cholesterol-independent mechanism involving inhibition of protein geranylgeranylation (Ref. 97). Similarly, in neonatal rat cardiomyocytes, fluvastatin induced apoptosis that was reversed by mevalonate or GGPP and was mimicked by a protein prenylation inhibitor (perillic acid) or the ROCK inhibitor Y27632 (Ref. 98). It should be noted, however, that cardiomyocytes (especially those from adult hearts) are particularly difficult to maintain in culture, partly because the ECM transduces key survival signals in vivo that cannot be simulated in vitro (Ref. 99). One should therefore be tentative in applying knowledge from in vitro apoptosis experiments to the in vivo scenario.

Electrical and contractile function

Statins may improve cardiac contractile function by increasing expression of sarcoplasmic calcium regulatory proteins, including SERCA and RyR2 (Refs 84, 100). Statins can also modulate myocyte action potentials through indirect effects on Kv1.5 and Kv4.3 channel activity (Ref. 101), observations that might underlie the reported antiarrhythmic effects of statins both in experimental models and in humans (Ref. 102). A further beneficial effect of statins might be in opposing the potentially harmful positive inotropic effects of β -adrenergic stimulation. Atorvastatin can desensitise cardiomyocytes to the effects of β -adrenergic stimulation through a mechanism involving reduced isoprenylation of γ -subunits of heterotrimeric G-proteins and subsequent reductions in the cellular content of G α_s and hence cyclic AMP levels (Ref. 103). Similar effects have been observed in rat hearts in vivo (Ref. 104). Together, these data suggest that the ability of statins to improve cardiac function may be due, at least in part, to direct pleiotropic effects on cardiomyocyte calcium handling and excitation-contraction coupling.

Effects of statins on cardiac fibroblasts

The critical role of CFs in maintaining ECM homeostasis in the healthy heart is underscored by the need to provide structure, function and connectivity for all the myocardial cell types (Ref. 105). Through cell-cell interaction and secretion of growth factors, myocyte function is directly modulated by CFs (Ref. 106). During

the myocardial remodelling that follows MI and during heart failure progression, CFs undergo activation to a myofibroblast phenotype, expressing α -smooth muscle actin (α -SMA) and exhibiting proliferative, migratory and secretory properties (Refs 68, 71). Myofibroblasts can develop not only from pre-existing resident myocardial fibroblasts, but also from endothelial- and epithelial-to-mesenchymal transition, bone marrow stem cells and monocytes (Refs 107, 108). Myofibroblasts are not found in healthy myocardium, but are abundant in granulation tissue in infarct zones, where they have been observed to persist for months (Refs 109, 110), or years (Ref. 67), in mature scars. It is likely that this persistence facilitates fibrosis that directly influences pathological remodelling and compromises cardiac function that ultimately leads to heart failure.

As outlined in the preceding sections, experimental studies exploring the effects of statins on the heart have mainly focused on global myocardial remodelling in vivo or on isolated cardiomyocytes in vitro to provide evidence that targeting small G-proteins in the heart is beneficial. There are, however, considerably less data relating to the functional effects of statins at the level of CFs. The complexity of the in vivo scenario makes assessment of the direct effects of pharmacological agents on individual cell types difficult. Thus, studies on cultured CFs derived from a variety of sources (animal and human, adult and neonatal, atrial and ventricular) have been the predominant source of current knowledge. With the caveats that CF characterisation is paramount to cellular studies and the relevance of findings to the clinical scenario is interpreted with caution, experimental studies on the effects of statins have revealed important data with respect to myofibroblast differentiation, cell proliferation and migration, and ECM turnover (Fig. 2b).

Myofibroblast differentiation

Although in vivo evidence is lacking, it has been suggested that statins can reduce fibroblast–myofibroblast transformation. For example, simvastatin attenuated expression of α -SMA induced by transforming growth factor- β (TGF- β) in cultured canine atrial fibroblasts (Ref. 111) and similarly, pravastatin suppressed phenotypic transformation of rat CFs to myofibroblasts (Ref. 112). As ROCK has a key

role in differentiation of monocytes to cardiac myofibroblasts (Ref. 113), it is plausible that statins would interfere with this process in vivo; however, this has yet to be tested. Other evidence can be drawn from studies on fibroblasts from noncardiac sources. For example, inhibition of Rac (Ref. 114) or ROCK (Ref. 115) prevents human dermal fibroblasts adopting a myofibroblast phenotype, and statins can reduce myofibroblast differentiation (Ref. 116) and epithelial–mesenchymal transition (Ref. 117) by inhibiting RhoA signalling in fibroblasts of non-cardiac origin.

Proliferation and migration

Proliferation and migration of CFs are fundamental to the myocardial-remodelling process, and these functions have been well studied in cultured CFs. A range of statins consistently inhibit CF proliferation regardless of species or mitogenic stimulus. For example, DNA synthesis (a marker of proliferation) in rat (Refs 118, 119, 120, 121, 122), mouse (Ref. 123) and canine (Ref. 111) CFs was reduced by statin treatment. Few of these studies have investigated the underlying mechanism for these effects, although inhibition of ERK and AKT signalling pathways by statins has been proposed (Ref. 118). Our own studies using human CFs demonstrated that simvastatin reduced cell proliferation and cyclin A expression in a concentration-dependent manner through inhibition of RhoA geranylgeranylation (Ref. 124). Moreover, we further demonstrated in human CFs that simvastatin could inhibit TNF- α (tumour necrosis factor- α)-induced proliferation, migration and invasion (Refs 125, 126) by distinct mechanisms. Simvastatin reduced the ability of CFs to invade an ECM barrier by reducing secretion of the matrix metalloprotease MMP-9 through a novel post-transcriptional mechanism (Ref. 126). Additionally, simvastatin reduced CF migration (motility) per se by inducing cytoskeletal disruption through ROCK inhibition (Ref. 126). The known intracellular mechanisms by which statins can reduce CF proliferation and migration are summarised in Figure 4.

Regulation of ECM

CFs are key regulators of ECM metabolism, responding to altered levels of a range of growth factors and cytokines to balance

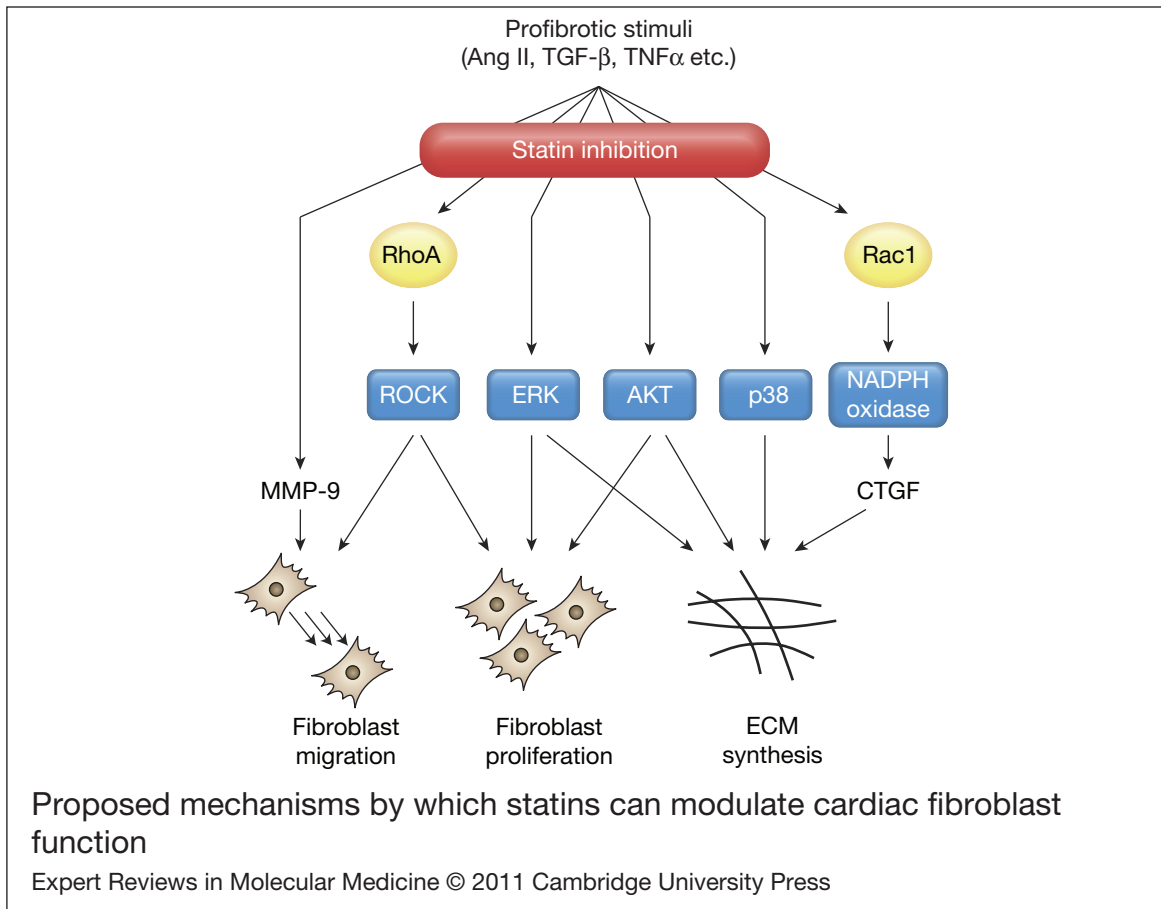


Figure 4. Proposed mechanisms by which statins can modulate cardiac fibroblast function. Statins reduce CF migration, proliferation and ECM synthesis by inhibition of RhoA and Rac1 small GTPases. Statins can also inhibit ERK, AKT and p38 MAP kinase signalling pathways, albeit through undefined mechanisms, leading to reduced fibroblast proliferation and collagen synthesis. Additionally, MMP-9 secretion by CFs can be inhibited by statins, thus impeding cell migration through the ECM.

synthesis and degradation of ECM components (Ref. 127). Statins have been consistently found to attenuate collagen synthesis by CFs. For example, procollagen expression was mildly inhibited by pravastatin in Ang-II-treated mouse CFs, an effect that was augmented by co-treatment with pioglitazone, an insulin-sensitising agent (Ref. 123). The underlying mechanism for this appeared to be the result of reduced Ang-II-induced activation of p38 MAPK and ERK. In neonatal rat CFs, Ang-II- or TGF-β-induced expression of procollagen mRNA and collagen deposition were inhibited by atorvastatin in a concentration-dependent manner (Ref. 119). The same study also reported similar effects on human CFs. In addition to directly attenuating collagen synthesis, statins might also exert

antifibrotic effects by reducing the secretion of profibrotic growth factors, such as TGF-β or connective tissue growth factor (CTGF). Ang II induces expression of CTGF by activation of Rac1 and NADPH oxidase, and this can be reduced by simvastatin in rat CFs through inhibition of Rac1 (Refs 119, 128). Atorvastatin can also reduce TGF-β-induced collagen synthesis by inhibiting the PI3K–AKT–Smad3 pathway and reducing expression of endoglin, a membrane glycoprotein that acts as a co-receptor for TGF-β (Ref. 129). Thus, there are multiple mechanisms by which statins can reduce ECM synthesis at the level of the CF (summarised in Fig. 4).

Although CFs derived from several species, including humans, have been shown to express several members of the MMP family (Ref. 68),

modulation of this group of proteases by statins is less well studied. Ang II upregulated MMP-3 and MMP-9 expression in mouse CFs and although MMP activity was inhibited by pravastatin, this was only observed when in combination with pioglitazone (Ref. 123). In our own laboratory, we showed that simvastatin inhibited TNF- α -induced MMP-9 secretion from human CFs and this led to reduced invasive capacity (Ref. 125). Studies in other cell types (e.g. vascular smooth muscle and inflammatory cells) have identified MMPs as being potential targets of the pleiotropic effects of statin inhibition (Refs 130, 131), so it is likely that similar mechanisms exist in CFs.

Anti-inflammatory effects of statins

In statin-treated heart failure patients, reduced plasma levels of proinflammatory cytokines are observed (Refs 132, 133). Moreover, both experimental and clinical studies suggest that statins can reduce local expression of proinflammatory cytokines in the myocardium (Refs 134, 135). Because CFs are an important source of myocardial cytokines, in our own studies we investigated whether statins could modulate proinflammatory cytokine expression in human CFs (Ref. 136). TNF- α treatment stimulated expression of interleukins IL-1 α , IL-1 β and IL-6, but none was modulated by statin therapy, indicating that CFs are probably not the cellular targets for the anti-inflammatory effects of statins on the heart.

However, we found that exposure of human endothelial cells to inflammatory mediators increased the expression of adhesion molecules and augmented leukocyte adhesions in a flow model (Ref. 137), both of which were abrogated by statin treatment. It appears therefore that the anti-inflammatory effects of statins in the inflammatory milieu of atherosclerosis, CHD and MI are due predominantly to effects on endothelial and inflammatory cells (reviewed in Refs 69, 138).

In summary, a large body of evidence supports the concept of both cardiomyocytes and CFs as distinct targets for the beneficial modulatory effects of statins on myocardial remodelling. The ability of statins to regulate cellular function at different levels underscores their utility as versatile therapeutic agents through their pleiotropic effects in addition to their primary role as cholesterol-lowering drugs.

Future directions and outstanding research questions

It is beyond question that the widespread prescribing of statins has resulted in significant reductions in the morbidity and mortality of cardiovascular disease. Moreover, the consensus opinion is that these drugs provide beneficial effects that are greater than would be expected from lipid lowering alone.

Despite the extensive use and high safety profile of statins, they have been found to exhibit unfavourable effects in some patients. Lowering of plasma cholesterol levels is primarily attributable to inhibition of HMG-CoA reductase in the liver, whereas the cholesterol-independent effects can feasibly affect every cell in the body. Essential roles for many derivatives of mevalonate in normal physiology suggest that global inhibition of this pathway (by HMG-CoA reductase) might explain some of the reported side effects of statins. As discussed in this review, clinical and experimental research has shown that many of the pleiotropic effects of statins are likely to be attributable to inhibition of small GTPase prenylation, predominantly inhibition of the Rho family GTPases. On the contrary, the reported detrimental effects of statins appear to be associated with other intermediates in the mevalonate pathway (see Fig. 1). For example, statins inhibit the synthesis of coenzyme Q10 (ubiquinone), whose benzoate ring structure requires a polyisoprene side chain that is synthesised from FPP (Ref. 139). Q10 is present in all cells and essentially required for both mitochondrial function and its antioxidant properties. Statin-induced Q10 inhibition has been documented in animal and human studies, and has been proposed to underlie statin-induced myopathy (Ref. 140). By inhibiting mevalonate synthesis, statins also inhibit synthesis of isopentenyl pyrophosphate (see Fig. 1), which is required for activation of selenocysteine transfer RNA (Ref. 141). Inhibition of this step leads to deficiency or dysfunction of selenoproteins required for muscle metabolism and hence has been associated with statin-induced muscle myopathies.

Given that the unwanted side effects of statins arise from inhibition of the mevalonate pathway, and that the beneficial pleiotropic effects of statins are perceived to be due predominantly to inhibition of Rho-family small GTPases, it is logical to investigate whether inhibition of Rho

signalling itself offers clinical benefits to myocardial remodelling without the side effects of statins. Despite growing interest in ROCK as a therapeutic target, there are surprisingly few clinical trials that have used ROCK inhibitors (Refs 142, 143). Currently, the isoquinoline derivative fasudil (HA-1077) is the only ROCK inhibitor available for clinical use, although this agent has limited specificity for ROCK over other kinases, especially PKA (Ref. 144). Human trials have assessed the efficacy of fasudil in stroke, angina, vascular resistance, hypertension, atherosclerosis and aortic stiffness (Ref. 142). In addition, fasudil is currently undergoing clinical trials for treating atherosclerosis and hypercholesterolaemia (ClinicalTrials.gov identifier: NCT00120718). The potential of orally active ROCK inhibitors for treating MI, LV hypertrophy and heart failure has not yet been examined in humans (Ref. 143). The established safety record of fasudil suggests that ROCK inhibition per se is a promising target for therapies aimed at reducing cardiac dysfunction. Other more selective inhibitors of ROCK (e.g. Y27632, GSK269962A, SB772077B, SR-715, SR-899 and SLx-2119) have not yet been assessed in man (Ref. 143). Therefore, although new concepts are currently being explored to mimic some of the pleiotropic effects of statins, the major rationale for prescribing these drugs is that of cholesterol lowering. It remains to be proven whether there are any additional clinical advantages in developing new therapies to specifically target small GTPases.

In this review we have discussed the clinical and experimental benefits of statins on the heart and its key constituent cells, cardiomyocytes and CFs. The fact that statins exhibit properties that are distinct and nonoverlapping with current medical therapies for heart failure leads to the proposition of added benefit for patients. Understanding the mechanisms of the pleiotropic effects of statins on the heart remains an important goal if we are to expose targets for more focused therapies to reduce adverse myocardial remodelling in man. Although some such trials are in progress, there are many more avenues to be explored in the future.

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References

- 1 Steinberg, D. and Gotto, A.M., Jr. (1999) Preventing coronary artery disease by lowering cholesterol levels: fifty years from bench to bedside. *Journal of the American Medical Association* 282, 2043-2050
- 2 Kannel, W.B. and McGee, D.L. (1979) Diabetes and cardiovascular risk factors: the Framingham study. *Circulation* 59, 8-13
- 3 Grundy, S.M. et al. (2004) Implications of recent clinical trials for the National Cholesterol Education Program Adult Treatment Panel III Guidelines. *Journal of the American College of Cardiology* 44, 720-732
- 4 Istvan, E.S. and Deisenhofer, J. (2001) Structural mechanism for statin inhibition of HMG-CoA reductase. *Science* 292, 1160-1164
- 5 Scandinavian Simvastatin Survival Study Group (1994) Randomised trial of cholesterol lowering in 4444 patients with coronary heart disease: the Scandinavian Simvastatin Survival Study (4S). *Lancet* 344, 1383-1389
- 6 Brugts, J.J. et al. (2009) The benefits of statins in people without established cardiovascular disease but with cardiovascular risk factors: meta-analysis of randomised controlled trials. *British Medical Journal* 338, b2376
- 7 Sacks, F.M. et al. (1996) The effect of pravastatin on coronary events after myocardial infarction in patients with average cholesterol levels. Cholesterol and Recurrent Events Trial investigators. *New England Journal of Medicine* 335, 1001-1009
- 8 The Long-Term Intervention with Pravastatin in Ischaemic Disease (LIPID) Study Group (1998) Prevention of cardiovascular events and death with pravastatin in patients with coronary heart disease and a broad range of initial cholesterol levels. *New England Journal of Medicine* 339, 1349-1357
- 9 Heart Protection Study Collaborative Group (2002) MRC/BHF Heart Protection Study of cholesterol lowering with simvastatin in 20,536 high-risk individuals: a randomised placebo-controlled trial. *Lancet* 360, 7-22
- 10 Shepherd, J. et al. (1995) Prevention of coronary heart disease with pravastatin in men with hypercholesterolemia. West of Scotland Coronary Prevention Study Group. *New England Journal of Medicine* 333, 1301-1307
- 11 Baigent, C. et al. (2005) Efficacy and safety of cholesterol-lowering treatment: prospective

- meta-analysis of data from 90,056 participants in 14 randomised trials of statins. *Lancet* 366, 1267-1278
- 12 Ridker, P.M. et al. (2008) Rosuvastatin to prevent vascular events in men and women with elevated C-reactive protein. *New England Journal of Medicine* 359, 2195-2207
 - 13 Cannon, C.P. et al. (2004) Intensive versus moderate lipid lowering with statins after acute coronary syndromes. *New England Journal of Medicine* 350, 1495-1504
 - 14 LaRosa, J.C. et al. (2005) Intensive lipid lowering with atorvastatin in patients with stable coronary disease. *New England Journal of Medicine* 352, 1425-1435
 - 15 Pedersen, T.R. et al. (2005) High-dose atorvastatin vs usual-dose simvastatin for secondary prevention after myocardial infarction: the IDEAL study: a randomized controlled trial. *Journal of the American Medical Association* 294, 2437-2445
 - 16 Armitage, J. et al. (2010) Intensive lowering of LDL cholesterol with 80 mg versus 20 mg simvastatin daily in 12,064 survivors of myocardial infarction: a double-blind randomised trial. *Lancet* 376, 1658-1669
 - 17 de Lemos, J.A. et al. (2004) Early intensive vs a delayed conservative simvastatin strategy in patients with acute coronary syndromes: phase Z of the A to Z trial. *Journal of the American Medical Association* 292, 1307-1316
 - 18 Baigent, C. et al. (2010) Efficacy and safety of more intensive lowering of LDL cholesterol: a meta-analysis of data from 170,000 participants in 26 randomised trials. *Lancet* 376, 1670-1681
 - 19 Davignon, J. (2004) Beneficial cardiovascular pleiotropic effects of statins. *Circulation* 109, III39-III43
 - 20 Kjekshus, J. et al. (1997) The effects of simvastatin on the incidence of heart failure in patients with coronary heart disease. *Journal of Cardiac Failure* 3, 249-254
 - 21 Node, K. et al. (2003) Short-term statin therapy improves cardiac function and symptoms in patients with idiopathic dilated cardiomyopathy. *Circulation* 108, 839-843
 - 22 Corsini, A. et al. (1999) New insights into the pharmacodynamic and pharmacokinetic properties of statins. *Pharmacology and Therapeutics* 84, 413-428
 - 23 Liao, J.K. and Laufs, U. (2005) Pleiotropic effects of statins. *Annual Review of Pharmacology and Toxicology* 45, 89-118
 - 24 Pekkanen, J. et al. (1990) Ten-year mortality from cardiovascular disease in relation to cholesterol level among men with and without preexisting cardiovascular disease. *New England Journal of Medicine* 322, 1700-1707
 - 25 Liu, P.Y. et al. (2009) Evidence for statin pleiotropy in humans: differential effects of statins and ezetimibe on rho-associated coiled-coil containing protein kinase activity, endothelial function, and inflammation. *Circulation* 119, 131-138
 - 26 Ostad, M.A. et al. (2009) Flow-mediated dilation in patients with coronary artery disease is enhanced by high dose atorvastatin compared to combined low dose atorvastatin and ezetimibe: results of the CEZAR study. *Atherosclerosis* 205, 227-232
 - 27 Su, S.F. et al. (2000) Effects of pravastatin on left ventricular mass in patients with hyperlipidemia and essential hypertension. *American Journal of Cardiology* 86, 514-518
 - 28 Young-Xu, Y. et al. (2003) Statins reduce the incidence of atrial fibrillation in patients with coronary artery disease. *Journal of the American College of Cardiology* 41 (Suppl. A), 301A
 - 29 Tsouli, S.G. et al. (2008) Should a statin be prescribed to every patient with heart failure? *Heart Failure Reviews* 13, 211-225
 - 30 Foody, J.M. et al. (2006) Statins and mortality among elderly patients hospitalized with heart failure. *Circulation* 113, 1086-1092
 - 31 Horwich, T.B., MacLellan, W.R. and Fonarow, G.C. (2004) Statin therapy is associated with improved survival in ischemic and non-ischemic heart failure. *Journal of the American College of Cardiology* 43, 642-648
 - 32 Ramasubbu, K. et al. (2008) Experimental and clinical basis for the use of statins in patients with ischemic and nonischemic cardiomyopathy. *Journal of the American College of Cardiology* 51, 415-426
 - 33 Tavazzi, L. et al. (2008) Effect of rosuvastatin in patients with chronic heart failure (the GISSI-HF trial): a randomised, double-blind, placebo-controlled trial. *Lancet* 372, 1231-1239
 - 34 Khush, K.K. et al. (2007) Effect of high-dose atorvastatin on hospitalizations for heart failure: subgroup analysis of the treating to new targets (TNT) study. *Circulation* 115, 576-583
 - 35 Scirica, B.M. et al. (2006) Intensive statin therapy and the risk of hospitalization for heart failure after an acute coronary syndrome in the PROVE IT-TIMI 22 study. *Journal of the American College of Cardiology* 47, 2326-2331
 - 36 Kumar, A. and Cannon, C.P. (2008) The role of statins in the prevention of heart failure after acute

- coronary syndrome. *Heart Failure Clinics* 4, 129-139
- 37 Laufs, U., Custodis, F. and Bohm, M. (2008) Who does not need a statin: too late in end-stage renal disease or heart failure? *Heart* 94, 1138-1140
- 38 Adam, O. et al. (2008) Prevention of atrial fibrillation with 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors. *Circulation* 118, 1285-1293
- 39 Bauersachs, J. et al. (2001) Improvement of left ventricular remodeling and function by hydroxymethylglutaryl coenzyme A reductase inhibition with cerivastatin in rats with heart failure after myocardial infarction. *Circulation* 104, 982-985
- 40 Hasegawa, H. et al. (2003) 3-Hydroxy-3-methylglutaryl coenzyme A reductase inhibitors prevent the development of cardiac hypertrophy and heart failure in rats. *Journal of Molecular and Cellular Cardiology* 35, 953-960
- 41 Hayashidani, S. et al. (2002) Fluvastatin, a 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitor, attenuates left ventricular remodeling and failure after experimental myocardial infarction. *Circulation* 105, 868-873
- 42 Luo, J.D. et al. (1999) Simvastatin inhibits cardiac hypertrophy and angiotensin-converting enzyme activity in rats with aortic stenosis. *Clinical and Experimental Pharmacology and Physiology* 26, 903-908
- 43 Patel, R. et al. (2001) Simvastatin induces regression of cardiac hypertrophy and fibrosis and improves cardiac function in a transgenic rabbit model of human hypertrophic cardiomyopathy. *Circulation* 104, 317-324
- 44 Liao, J.K. (2002) Isoprenoids as mediators of the biological effects of statins. *Journal of Clinical Investigation* 110, 285-288
- 45 Van Aelst, L. and Souza-Schorey, C. (1997) Rho GTPases and signaling networks. *Genes and Development* 11, 2295-2322
- 46 Lezoualc'h, F. et al. (2008) Small GTP-binding proteins and their regulators in cardiac hypertrophy. *Journal of Molecular and Cellular Cardiology* 44, 623-632
- 47 Adam, O. et al. (2007) Role of Rac1 GTPase activation in atrial fibrillation. *Journal of the American College of Cardiology* 50, 359-367
- 48 Custodis, F. et al. (2006) Association of RhoGDI α with Rac1 GTPase mediates free radical production during myocardial hypertrophy. *Cardiovascular Research* 71, 342-351
- 49 Maack, C. et al. (2003) Oxygen free radical release in human failing myocardium is associated with increased activity of rac1-GTPase and represents a target for statin treatment. *Circulation* 108, 1567-1574
- 50 Clerk, A. and Sugden, P.H. (2000) Small guanine nucleotide-binding proteins and myocardial hypertrophy. *Circulation Research* 86, 1019-1023
- 51 Hunter, J.J. and Chien, K.R. (1999) Signaling pathways for cardiac hypertrophy and failure. *New England Journal of Medicine* 341, 1276-1283
- 52 Brown, J.H., Del Re, D.P. and Sussman, M.A. (2006) The Rac and Rho hall of fame: a decade of hypertrophic signaling hits. *Circulation Research* 98, 730-742
- 53 Clerk, A. et al. (2001) Regulation of mitogen-activated protein kinases in cardiac myocytes through the small G protein Rac1. *Molecular and Cellular Biology* 21, 1173-1184
- 54 Satoh, M. et al. (2006) Requirement of Rac1 in the development of cardiac hypertrophy. *Proceedings of the National Academy of Sciences of the United States of America* 103, 7432-7437
- 55 Lu, H. et al. (2006) Integrin-linked kinase expression is elevated in human cardiac hypertrophy and induces hypertrophy in transgenic mice. *Circulation* 114, 2271-2279
- 56 Sah, V.P. et al. (1999) Cardiac-specific overexpression of RhoA results in sinus and atrioventricular nodal dysfunction and contractile failure. *Journal of Clinical Investigation* 103, 1627-1634
- 57 Higashi, M. et al. (2003) Long-term inhibition of Rho-kinase suppresses angiotensin II-induced cardiovascular hypertrophy in rats in vivo: effect on endothelial NAD(P)H oxidase system. *Circulation Research* 93, 767-775
- 58 Satoh, S. et al. (2003) Chronic inhibition of Rho kinase blunts the process of left ventricular hypertrophy leading to cardiac contractile dysfunction in hypertension-induced heart failure. *Journal of Molecular and Cellular Cardiology* 35, 59-70
- 59 Wang, Y.X. et al. (2005) Inhibition of Rho-kinase by fasudil attenuated angiotensin II-induced cardiac hypertrophy in apolipoprotein E deficient mice. *European Journal of Pharmacology* 512, 215-222
- 60 Hattori, T. et al. (2004) Long-term inhibition of Rho-kinase suppresses left ventricular remodeling after myocardial infarction in mice. *Circulation* 109, 2234-2239

- 61 Bao, W. et al. (2004) Inhibition of Rho-kinase protects the heart against ischemia/reperfusion injury. *Cardiovascular Research* 61, 548-558
- 62 Rikitake, Y. et al. (2005) Decreased perivascular fibrosis but not cardiac hypertrophy in ROCK1 + /- haploinsufficient mice. *Circulation* 112, 2959-2965
- 63 Zhang, Y.M. et al. (2006) Targeted deletion of ROCK1 protects the heart against pressure overload by inhibiting reactive fibrosis. *FASEB Journal* 20, 916-925
- 64 Hunter, J.J. et al. (1995) Ventricular expression of a MLC-2v-ras fusion gene induces cardiac hypertrophy and selective diastolic dysfunction in transgenic mice. *Journal of Biological Chemistry* 270, 23173-23178
- 65 Zheng, M. et al. (2004) Sarcoplasmic reticulum calcium defect in Ras-induced hypertrophic cardiomyopathy heart. *American Journal of Physiology. Heart and Circulatory Physiology* 286, H424-H433
- 66 Kai, H. et al. (1998) Expression of proto-oncogenes and gene mutation of sarcomeric proteins in patients with hypertrophic cardiomyopathy. *Circulation Research* 83, 594-601
- 67 Jugdutt, B.I. (2003) Ventricular remodeling after infarction and the extracellular collagen matrix: when is enough enough? *Circulation* 108, 1395-1403
- 68 Porter, K.E. and Turner, N.A. (2009) Cardiac fibroblasts: at the heart of myocardial remodeling. *Pharmacology and Therapeutics* 123, 255-278
- 69 Zhou, Q. and Liao, J.K. (2010) Pleiotropic effects of statins. *Basic research and clinical perspectives. Circulation Journal* 74, 818-826
- 70 Brown, R.D. et al. (2005) The cardiac fibroblast: therapeutic target in myocardial remodeling and failure. *Annual Review of Pharmacology and Toxicology* 45, 657-687
- 71 Camelliti, P., Borg, T.K. and Kohl, P. (2005) Structural and functional characterisation of cardiac fibroblasts. *Cardiovascular Research* 65, 40-51
- 72 Weber, K.T. (2004) Fibrosis in hypertensive heart disease: focus on cardiac fibroblasts. *Journal of Hypertension* 22, 47-50
- 73 Banerjee, I. et al. (2006) Dynamic interactions between myocytes, fibroblasts, and extracellular matrix. *Annals of the New York Academy of Sciences* 1080, 76-84
- 74 Kohl, P. (2004) Cardiac cellular heterogeneity and remodelling. *Cardiovascular Research* 64, 195-197
- 75 Camelliti, P., Green, C.R. and Kohl, P. (2006) Structural and functional coupling of cardiac myocytes and fibroblasts. *Advances in Cardiology* 42, 132-149
- 76 Mann, D.L. and Bristow, M.R. (2005) Mechanisms and models in heart failure: the biomechanical model and beyond. *Circulation* 111, 2837-2849
- 77 Mital, S. and Liao, J.K. (2004) Statins and the myocardium. *Seminars in Vascular Medicine* 4, 377-384
- 78 Oi, S. et al. (1999) Lovastatin prevents angiotensin II-induced cardiac hypertrophy in cultured neonatal rat heart cells. *European Journal of Pharmacology* 376, 139-148
- 79 Nishikimi, T. et al. (2002) Cerivastatin, a hydroxymethylglutaryl coenzyme A reductase inhibitor, inhibits cardiac myocyte hypertrophy induced by endothelin. *European Journal of Pharmacology* 453, 175-181
- 80 Laufs, U. et al. (2002) Impact of HMG CoA reductase inhibition on small GTPases in the heart. *Cardiovascular Research* 53, 911-920
- 81 Morikawa-Futamatsu, K. et al. (2006) HMG-CoA reductase inhibitor fluvastatin prevents angiotensin II-induced cardiac hypertrophy via Rho kinase and inhibition of cyclin D1. *Life Sciences* 79, 1380-1390
- 82 Hordijk, P.L. (2006) Regulation of NADPH oxidases: the role of Rac proteins. *Circulation Research* 98, 453-462
- 83 Takemoto, M. et al. (2001) Statins as antioxidant therapy for preventing cardiac myocyte hypertrophy. *Journal of Clinical Investigation* 108, 1429-1437
- 84 Choi, E.Y. et al. (2010) Rosuvastatin inhibits norepinephrine-induced cardiac hypertrophy via suppression of Gh. *European Journal of Pharmacology* 627, 56-62
- 85 Hwang, K.C. et al. (1996) Alpha 1-adrenergic receptor coupling with Gh in the failing human heart. *Circulation* 94, 718-726
- 86 Wu, L. et al. (2006) Simvastatin attenuates hypertrophic responses induced by cardiotrophin-1 via JAK-STAT pathway in cultured cardiomyocytes. *Molecular and Cellular Biochemistry* 284, 65-71
- 87 Liu, J., Shen, Q. and Wu, Y. (2008) Simvastatin prevents cardiac hypertrophy in vitro and in vivo via JAK/STAT pathway. *Life Sciences* 82, 991-996
- 88 Planavila, A. et al. (2008) Atorvastatin inhibits GSK-3beta phosphorylation by cardiac hypertrophic stimuli. *Biochimica et Biophysica Acta* 1781, 26-35
- 89 Hauck, L. et al. (2007) Critical role for FoxO3a-dependent regulation of p21CIP1/WAF1 in

- response to statin signaling in cardiac myocytes. *Circulation Research* 100, 50-60
- 90 Whelan, R.S., Kaplinskiy, V. and Kitsis, R.N. (2010) Cell death in the pathogenesis of heart disease: mechanisms and significance. *Annual Review of Physiology* 72, 19-44
- 91 Webster, K.A. (2007) Programmed death as a therapeutic target to reduce myocardial infarction. *Trends in Pharmacological Sciences* 28, 492-499
- 92 Verma, S. et al. (2004) Novel cardioprotective effects of pravastatin in human ventricular cardiomyocytes subjected to hypoxia and reoxygenation: beneficial effects of statins independent of endothelial cells. *Journal of Surgical Research* 119, 66-71
- 93 Bergmann, M.W. et al. (2004) Statins inhibit reoxygenation-induced cardiomyocyte apoptosis: role for glycogen synthase kinase 3 β and transcription factor β -catenin. *Journal of Molecular and Cellular Cardiology* 37, 681-690
- 94 Vilahur, G. et al. (2009) Induction of RISK by HMG-CoA reductase inhibition affords cardioprotection after myocardial infarction. *Atherosclerosis* 206, 95-101
- 95 Jones, S.P. et al. (2003) Simvastatin attenuates oxidant-induced mitochondrial dysfunction in cardiac myocytes. *Circulation Research* 93, 697-699
- 96 Ito, M. et al. (2004) Statins inhibit beta-adrenergic receptor-stimulated apoptosis in adult rat ventricular myocytes via a Rac1-dependent mechanism. *Circulation* 110, 412-418
- 97 Demyanets, S. et al. (2006) Hydroxymethylglutaryl-coenzyme A reductase inhibitors induce apoptosis in human cardiac myocytes in vitro. *Biochemical Pharmacology* 71, 1324-1330
- 98 Ogata, Y. et al. (2002) Fluvastatin induces apoptosis in rat neonatal cardiac myocytes: a possible mechanism of statin-attenuated cardiac hypertrophy. *Journal of Cardiovascular Pharmacology* 40, 907-915
- 99 Rivera, D.M. and Lowes, B.D. (2005) Molecular remodeling in the failing human heart. *Current Heart Failure Reports* 2, 5-9
- 100 Zheng, X. and Hu, S.J. (2005) Effects of simvastatin on cardiac performance and expression of sarcoplasmic reticular calcium regulatory proteins in rat heart. *Acta Pharmacologica Sinica* 26, 696-704
- 101 Vaquero, M. et al. (2007) Effects of atorvastatin and simvastatin on atrial plateau currents. *Journal of Molecular and Cellular Cardiology* 42, 931-945
- 102 Lee, Y.L., Blaha, M.J. and Jones, S.R. (2011) Statin therapy in the prevention and treatment of atrial fibrillation. *Journal of Clinical Lipidology* 5, 18-29
- 103 Muhlhauser, U. et al. (2006) Atorvastatin desensitizes beta-adrenergic signaling in cardiac myocytes via reduced isoprenylation of G-protein gamma-subunits. *FASEB Journal* 20, 785-787
- 104 Schmechel, A. et al. (2009) Treatment with atorvastatin partially protects the rat heart from harmful catecholamine effects. *Cardiovascular Research* 82, 100-106
- 105 Souders, C.A., Bowers, S.L. and Baudino, T.A. (2009) Cardiac fibroblast: the renaissance cell. *Circulation Research* 105, 1164-1176
- 106 Baudino, T.A. et al. (2008) Cell patterning: interaction of cardiac myocytes and fibroblasts in three-dimensional culture. *Microscopy and Microanalysis* 14, 117-125
- 107 Krenning, G., Zeisberg, E.M. and Kalluri, R. (2010) The origin of fibroblasts and mechanism of cardiac fibrosis. *Journal of Cellular Physiology* 225, 631-637
- 108 Zeisberg, E.M. and Kalluri, R. (2010) Origins of cardiac fibroblasts. *Circulation Research* 107, 1304-1312
- 109 Sun, Y. and Weber, K.T. (2000) Infarct scar: a dynamic tissue. *Cardiovascular Research* 46, 250-256
- 110 Willems, I.E. et al. (1994) The alpha-smooth muscle actin-positive cells in healing human myocardial scars. *American Journal of Pathology* 145, 868-875
- 111 Shiroshita-Takeshita, A. et al. (2007) Effects of simvastatin on the development of the atrial fibrillation substrate in dogs with congestive heart failure. *Cardiovascular Research* 74, 75-84
- 112 Moiseeva, O.M. et al. (2007) Effect of pravastatin on phenotypical transformation of fibroblasts and hypertrophy of cardiomyocytes in culture. *Bulletin of Experimental Biology and Medicine* 143, 54-57
- 113 Haudek, S.B. et al. (2009) Rho kinase-1 mediates cardiac fibrosis by regulating fibroblast precursor cell differentiation. *Cardiovascular Research* 83, 511-518
- 114 Xu, S.W. et al. (2009) Rac inhibition reverses the phenotype of fibrotic fibroblasts. *PLoS One* 4, e7438
- 115 Akhmetshina, A. et al. (2008) Rho-associated kinases are crucial for myofibroblast differentiation and production of extracellular matrix in scleroderma fibroblasts. *Arthritis and Rheumatism* 58, 2553-2564
- 116 Meyer-Ter-Vehn, T. et al. (2008) Lovastatin inhibits TGF- β -induced myofibroblast transdifferentiation in human tenon fibroblasts. *Investigative Ophthalmology and Visual Science* 49, 3955-3960
- 117 Rodrigues-Diez, R. et al. (2008) Pharmacological modulation of epithelial mesenchymal transition

- caused by angiotensin II. Role of ROCK and MAPK pathways. *Pharmaceutical Research* 25, 2447-2461
- 118 He, Y.P. et al. (2008) Involvement of ERK and AKT signaling in the growth effect of arginine vasopressin on adult rat cardiac fibroblast and the modulation by simvastatin. *Molecular and Cellular Biochemistry* 317, 33-41
- 119 Martin, J. et al. (2005) In vitro inhibitory effects of atorvastatin on cardiac fibroblasts: implications for ventricular remodelling. *Clinical and Experimental Pharmacology and Physiology* 32, 697-701
- 120 Tian, B. et al. (2003) Angiotensin II modulates nitric oxide-induced cardiac fibroblast apoptosis by activation of AKT/PKB. *American Journal of Physiology. Heart and Circulatory Physiology* 285, H1105-H1112
- 121 Xu, L. et al. (2006) Effects of simvastatin on DNA synthesis in rat cardiac fibroblasts. *Nan Fang Yi Ke Da Xue Xue Bao* 26, 205-207, 213
- 122 Tian, J.W. et al. (2003) Effects of atorvastatin on the proliferation and collagen synthesis of rat cardiac fibroblasts. *Zhonghua Yi Xue Za Zhi* 83, 118-122
- 123 Chen, J. and Mehta, J.L. (2006) Angiotensin II-mediated oxidative stress and procollagen-1 expression in cardiac fibroblasts: blockade by pravastatin and pioglitazone. *American Journal of Physiology. Heart and Circulatory Physiology* 291, H1738-H1745
- 124 Porter, K.E. et al. (2004) Simvastatin reduces human atrial myofibroblast proliferation independently of cholesterol lowering via inhibition of RhoA. *Cardiovascular Research* 61, 745-755
- 125 Porter, K.E. et al. (2004) Tumor necrosis factor α induces human atrial myofibroblast proliferation, invasion and MMP-9 secretion: inhibition by simvastatin. *Cardiovascular Research* 64, 507-515
- 126 Turner, N.A. et al. (2007) Simvastatin inhibits TNF α -induced invasion of human cardiac myofibroblasts via both MMP-9-dependent and -independent mechanisms. *Journal of Molecular and Cellular Cardiology* 43, 168-176
- 127 Jugdutt, B.I. (2003) Ventricular remodeling after infarction and the extracellular collagen matrix: when is enough enough? *Circulation* 108, 1395-1403
- 128 Adam, O. et al. (2010) Rac1-induced connective tissue growth factor regulates connexin 43 and N-cadherin expression in atrial fibrillation. *Journal of the American College of Cardiology* 55, 469-480
- 129 Shyu, K.G. et al. (2010) Mechanism of the inhibitory effect of atorvastatin on endoglin expression induced by transforming growth factor- β 1 in cultured cardiac fibroblasts. *European Journal of Heart Failure* 12, 219-226
- 130 Porter, K.E. and Turner, N.A. (2002) Statins for the prevention of vein graft stenosis: a role for inhibition of matrix metalloproteinase-9. *Biochem Soc Trans* 30, 120-126
- 131 Luan, Z., Chase, A.J. and Newby, A.C. (2003) Statins inhibit secretion of metalloproteinases-1, -2, -3, and -9 from vascular smooth muscle cells and macrophages. *Arterioscler Thromb Vasc Biol* 23, 769-775
- 132 Tousoulis, D. et al. (2005) Effects of combined administration of low dose atorvastatin and vitamin E on inflammatory markers and endothelial function in patients with heart failure. *European Journal of Heart Failure* 7, 1126-1132
- 133 Sola, S. et al. (2006) Atorvastatin improves left ventricular systolic function and serum markers of inflammation in nonischemic heart failure. *Journal of the American College of Cardiology* 47, 332-337
- 134 Zhang, J. et al. (2005) Simvastatin regulates myocardial cytokine expression and improves ventricular remodeling in rats after acute myocardial infarction. *Cardiovascular Drugs and Therapy* 19, 13-21
- 135 Wallace, C.K. et al. (2005) Simvastatin decreases myocardial tumor necrosis factor α content in heart transplant recipients. *Journal of Heart and Lung Transplantation* 24, 46-51
- 136 Turner, N.A. et al. (2007) Mechanism of TNF α -induced IL-1 α , IL-1 β and IL-6 expression in human cardiac fibroblasts: effects of statins and thiazolidinediones. *Cardiovascular Research* 76, 81-90
- 137 Eccles, K.A. et al. (2008) Simvastatin alters human endothelial cell adhesion molecule expression and inhibits leukocyte adhesion under flow. *Atherosclerosis* 200, 69-79
- 138 Arnaud, C., Braunersreuther, V. and Mach, F. (2005) Toward immunomodulatory and anti-inflammatory properties of statins. *Trends in Cardiovascular Medicine* 15, 202-206
- 139 Beltowski, J., Wojcicka, G. and Jamroz-Wisniewska, A. (2009) Adverse effects of statins – mechanisms and consequences. *Current Drug Safety* 4, 209-228
- 140 Marcoff, L. and Thompson, P.D. (2007) The role of coenzyme Q10 in statin-associated myopathy: a systematic review. *Journal of the American College of Cardiology* 49, 2231-2237
- 141 Moosmann, B. and Behl, C. (2004) Selenoprotein synthesis and side-effects of statins. *Lancet* 363, 892-894

- 142 Olson, M.F. (2008) Applications for ROCK kinase inhibition. *Current Opinion in Cell Biology* 20, 242-248
- 143 Dong, M. et al. (2010) Rho-kinase inhibition: a novel therapeutic target for the treatment of cardiovascular diseases. *Drug Discovery Today* 15, 622-629
- 144 Bain, J. et al. (2007) The selectivity of protein kinase inhibitors: a further update. *Biochemical Journal* 408, 297-315

Further reading, resources and contacts

The NIH-supported website provides a searchable database of recent, ongoing and upcoming clinical trials conducted in the USA and around the world:

www.ClinicalTrials.gov

Features associated with this article

Figures

Figure 1. The cholesterol synthesis pathway.

Figure 2. Effect of statins on cardiomyocyte and cardiac fibroblast function.

Figure 3. Proposed mechanisms by which statin-mediated inhibition of GTPases can reduce cardiomyocyte hypertrophy.

Figure 4. Proposed mechanisms by which statins can modulate cardiac fibroblast function.

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