

Economic costs of protistan and metazoan parasites to global mariculture

A. P. SHINN^{1,2*}, J. PRATOOMYOT³, J. E. BRON², G. PALADINI², E. E. BROOKER²
and A. J. BROOKER²

¹ *Fish Vet Group Asia Limited, 99/386, Chaengwattana Building, Chaengwattana Road, Kwaeng Toongsonghong, Khet Laksi, Bangkok 10210, Thailand*

² *Institute of Aquaculture, School of Natural Sciences, University of Stirling, Stirling FK9 4LA, UK*

³ *Institute of Marine Science, Burapha University, Chonburi, Thailand*

(Received 4 May 2014; revised 4 July 2014; accepted 6 July 2014)

SUMMARY

Parasites have a major impact on global finfish and shellfish aquaculture, having significant effects on farm production, sustainability and economic viability. Parasite infections and impacts can, according to pathogen and context, be considered to be either unpredictable/sporadic or predictable/regular. Although both types of infection may result in the loss of stock and incur costs associated with the control and management of infection, predictable infections can also lead to costs associated with prophylaxis and related activities. The estimation of the economic cost of a parasite event is frequently complicated by the complex interplay of numerous factors associated with a specific incident, which may range from direct production losses to downstream socio-economic impacts on livelihoods and satellite industries associated with the primary producer. In this study, we examine the world's major marine and brackish water aquaculture production industries and provide estimates of the potential economic costs attributable to a range of key parasite pathogens using 498 specific events for the purposes of illustration and estimation of costs. This study provides a baseline resource for risk assessment and the development of more robust biosecurity practices, which can in turn help mitigate against and/or minimise the potential impacts of parasite-mediated disease in aquaculture.

Keywords: Aquaculture, production, mortality, finfish, Crustacea, Mollusca, ornamentals, economic cost, review.

INTRODUCTION

Pre-harvest mortalities in marine aquaculture result from the complex interplay of a broad range of factors that include stock source/genotype, developmental defects, predation and cannibalism, impaired nutrition, physical damage, sub-optimal/hostile environmental conditions and disease. Economic losses accrue not only from mortalities but also from impacts on growth and food conversion, post-harvest downgrading or rejection of carcasses and derived products, fish escapes, management decisions that impact on profitability, e.g. protracted decisions to treat, grade or harvest, and the costs and effects of particular husbandry and management practices, e.g. fallowing, grading, vaccination, treatment and stock handling.

Parasitic diseases attributable to obligate or opportunistic eukaryotic pathogens continue to have a major impact on global finfish and shellfish aquaculture, and in many regions they represent a key constraint to production, sustainability and economic

viability. Although robust data can often be generated concerning the general patterns of stock loss within a typical production cycle, obtaining accurate figures for the impacts of disease can be more problematic due to a number of considerations, which include production scale, available resources, difficulties in making rapid or accurate parasite identifications at the farm level and poor record keeping. The frequent association of disease with other pre-disposing factors, such as poor water quality and the broad variety of precipitating events that may act to stress the farm population, also means that untangling the impacts of disease from those attributable to other causes may be difficult or impossible. Thus, in many cases, the economic impact of parasitic diseases can only be estimated.

Parasite-induced impacts in marine aquaculture can be divided into two broad categories: unpredictable/sporadic and predictable/regular. For both, there may be costs in treating and managing infections once established, but for predictable infections there will also be costs associated with prophylactic treatment/management. For example, the management costs associated with controlling infections of caligid copepods, e.g. *Lepeophtheirus salmonis* and *Caligus* spp. in farmed Atlantic salmon (*Salmo salar*) can largely be predicted within a production cycle as

* Corresponding author: Fish Vet Group Asia Limited, 99/386, Chaengwattana Building, Moo 2, Chaengwattana Road, Kwaeng Tungsonhong, Khet Laksi, Bangkok 10210, Thailand. E-mail: andy.shinn@fishvetgroup.com

these parasites pose a perennial threat to captive reared stocks. The infection dynamics of these species are well understood, and they can be controlled through the employment of an integrated pest management strategy (IPMS) involving the use of a broad range of management tools in addition to direct treatment intervention. The global Atlantic salmon production industry is well established (>40 years) and can draw upon the long-term, shared experiences of parasite control and management that have led to the development of effective strategies to minimise mortalities, damage and loss of profit (see Frenzl *et al.* 2013, 2014). For many other new or less-established industries, however, particularly those restricted to a small number of production sites, the parasite threats may be largely unknown and emerging, and new infections can have a devastating impact. The impact of *Paramoeba perurans* (syn. *Neoparamoeba perurans*), the causative pathogen of amoebic gill disease (AGD), on the early Tasmanian production of rainbow trout, *Oncorhynchus mykiss*, serves as an appropriate example (Munday *et al.* 1990).

Many parasite infection events are complicated by the complex interplay of numerous factors making it difficult to calculate the precise costs attributable to the parasite. The Chilean crash in national salmon production from 385 086 tonnes (t) in 2006 to 230 678 t in 2010 (FAO FishStatJ, 2013), for example, appears to have been multifactorial with the key pathogens involved being ISA v (infectious salmon anaemia virus) and the caligid copepod *Caligus rogercresseyi*. Major contributing factors included a large number of marine farms in production, the high stocking densities employed, a concentration of farms within a small area (~40% of the salmon production around Chiloé Island), a lack of biosecurity measures, weak disease surveillance, poor sanitary control and a failure to employ zone management (Ibieta *et al.* 2011). Many farm sites were subject to infection from other disease agents, such as salmonid rickettsial septicaemia (SRS) caused by *Piscirickettsia salmonis* (see Olivares and Marshall, 2010), infectious pancreatic necrosis (IPN) caused by the pancreatic necrosis virus (IPNV) (Ibieta *et al.* 2011), *P. perurans* (see Bustos *et al.* 2011; Rozas, 2011) and rising *C. rogercresseyi* infections (Rozas and Asencio, 2007), which were suggested to predispose salmon stocks to the ISA v infection. By the end of 2008, 105 Chilean sites were confirmed as ISA v positive with a further 44 suspect sites; a quarter of the positive sites were owned by a single company who declared losses of US\$ 81.2 billion for the second half of 2007 (Marine Harvest, 2007). As can be seen from the above, the calculation of the proportion of this loss that could be deemed to be attributable to *C. rogercresseyi* is not possible.

Although immediate losses to production can often be estimated, it is usually difficult to calculate the

full magnitude of the downstream socio-economic effects of major disease events on the livelihoods and associated industries centred around primary producers. While insurance claims may provide some guidance as to losses, these may be overinflated, based on 'best price' or on an estimated loss of trade/income. In addition, the costs of remedial action (e.g. treatment, disposal and monitoring) and/or changes to management practices and infrastructure also need to be considered. The resilience of the Chilean salmonid aquaculture industry, which rapidly implemented improved infectious disease control measures and was able to fall back on well-established coho salmon, *Oncorhynchus kisutch*, and rainbow trout industries (24 and 38% of national salmonid production in 2011, respectively), arguably minimised the full potential economic impact of the 2007 crisis (Alvial *et al.* 2012a, b). Although there are a number of studies that have attempted to estimate the full economic consequences of parasite infections, both through the documentation of a specific disease event (Roberts *et al.* 1994; Torgerson and MacPherson, 2011; Charlier *et al.* 2012) and through the estimation of the potential impact of disease introduction (Paisley *et al.* 1999; Riddington *et al.* 2006; Voort van der *et al.* 2013), the data and resources required to undertake such studies generally preclude the accurate estimation of the cost of parasite-associated impacts. For example, teasing out the role and the precise economic impact attributable to parasitic agents, e.g. *P. perurans* and *C. rogercresseyi*, from that due to the other contributing factors leading to the Chilean 2007 crisis, is a near-impossible task, and therefore, the costs can only be speculated upon. For this reason, the economic impact of these parasites in the Chilean 2007 crisis, is not included in this summary.

In this review, we provide an overview of the world's major marine and brackish aquaculture production industries and assess the impacts of the major parasite species that affect production or otherwise impose an economic cost. As discussed above, it is not possible to provide a comprehensive review of all parasite-related losses in aquaculture, but we provide estimates for some of the more serious loss-related events and provide brief details relating to each. For each event resulting in a notable loss, i.e. either mortality or deviation from projected revenue, we cite either figures given in the original publication or have applied a simple formula to determine the likely loss to the stock only at the point in the production cycle when the disease event occurred. In the absence of details within the original report, many of the costs provided here are assumption-based and so a degree of caution should be exercised. It is anticipated that this study will prove informative for risk assessment by new aquaculture-based enterprises and will aid an appreciation of the sporadic nature and impact of some parasite-induced

Table 1. Aquaculture production in brackish and marine environments in 2011 presented in tonnes (t) for each broad class of aquatic species. Figures are calculated from the FAO FishStatJ databases (2011)

	Brackish		Marine		Combined	
	Tonnage	%	Tonnage (t)	%	Tonnage (t)	%
Algae	635 654	11.55	20 265 356	51.66	20 901 010	46.722
Aq. inverts. (unspec.)			74 664	0.19	74 664	0.167
Ascidacea			12 369	0.03	12 369	0.028
Cnidaria			69 749	0.18	69 749	0.156
Crustacea	2 829 894	51.41	705 530	1.80	3 535 424	7.903
Echinoidea			6 791	0.02	6 791	0.015
Holothuroidea			138 186	0.35	138 186	0.309
Mollusca	99 341	1.80	14 069 022	35.86	14 168 363	31.672
Pisces	1 939 393	35.24	3 888 446	9.91	5 827 844	13.028
Total	5 504 282		39 230 113		44 734 400	

Table 2. The top 30 marine and brackish finfish aquaculture industries ranked by the value of production. Figures are predominantly derived from the FAO Fisheries and Aquaculture FishStatJ databases (2011)

Common name	Latin name	Tonnage (t)	Value (US\$ × 1000)	Largest producer
Atlantic salmon	<i>Salmo salar</i>	1 711 455	9 628 336	Norway (1 059 958 t; 61.93%)
Rainbow trout	<i>Oncorhynchus mykiss</i>	298 186	2 118 922	Chile (208 482 t; 69.92%)
Milkfish	<i>Chanos chanos</i>	823 781	1 427 587	Indonesia (467 044 t; 56.70%)
Japanese amberjack ^a	<i>Seriola quinqueradiata</i>	146 274	1 375 841	Japan (146 240 t; 99.98%)
Gilthead sea bream	<i>Sparus aurata</i>	154 771	928 666	Greece (70 600 t; 45.62%)
Coho salmon	<i>Oncorhynchus kisutch</i>	159 694	916 647	Chile (144 120 t; 90.25%)
European seabass	<i>Dicentrarchus labrax</i>	144 315	860 939	Turkey (47 013 t; 32.58%)
Turbot	<i>Scophthalmus maximus</i>	75 413	600 332	China (64 000 t; 84.87%)
Silver sea bream	<i>Pagrus auratus</i>	64 684	572 754	Japan (61 186 t; 94.59%)
Groupers	<i>Epinephelus</i> spp.	95 153	550 585	China (59 534 t; 62.57%)
Bastard halibuts	<i>Paralichthys olivaceus</i>	44 280	484 846	Rep. Korea (40 805 t; 92.15%)
Pompano	<i>Trachinotus</i> spp.	115 133	461 032	China (115 000 t; 99.88%)
Flathead grey mullet	<i>Mugil cephalus</i>	125 678	441 875	Egypt (84 001 t; 66.84%)
Barramundi	<i>Lates calcarifer</i>	55 685	258 645	Malaysia (17 607 t; 31.62%)
Cyprinids ^b	<i>Cyprinus carpio</i> , <i>Hypophthal michthys</i> <i>molitrix</i> , <i>Ctenophary-</i> <i>ngodon idellus</i> ^c	100 000	235 992	Egypt (100%)
Red seabream	<i>Pagrus major</i>	124 799	166 241	China (122 964 t; 98.53%)
Korean rockfish	<i>Sebastes schlegelii</i>	17 338	137 136	Rep. Korea (100%)
Chinook salmon	<i>Oncorhynchus tshawytscha</i>	15 131	103 140	New Zealand (13 362 t; 88.31%)
Large yellow croaker	<i>Larimichthys crocea</i>	80 212	95 452	China (100%)
Red drum	<i>Sciaenops ocellatus</i>	67 339	91 878	China (64 838 t; 96.29%)
Tiger pufferfish	<i>Takifugu rubripes</i>	14 906	85 986	China (11 632 t; 78.04%)
Porgies, sea breams		58 029	86 732	China (56 313 t; 97.04%)
Atlantic bluefin tuna	<i>Thunnus thynnus</i>	3 410	72 832	Croatia (1610 t; 47.21%)
Cobia	<i>Rachycentron canadum</i>	40 863	66 258	China (37 210 t; 91.06%)
Meagre	<i>Argyrosomus regius</i>	13 742	55 428	Egypt (12 092 t; 87.99%)
Atlantic cod	<i>Gadus morhua</i>	16 126	50 508	Norway (15 249 t; 94.56%)
Lefteye flounders ^d	<i>Psettina brevirectis</i>	47 589	47 589	China (100%)
Southern bluefin tuna	<i>Thunnus maccoyii</i>	1 987	44 261	Australia (100%)
Tilapia ^b	<i>Oreochromis niloticus</i> / <i>O. mossambicus</i>	617 533	42 033	Egypt (580 617 t; 94.02%)
Mangrove red snapper	<i>Lutjanus argentimaculatus</i>	5 259	32 113	Malaysia (5 237 t; 99.58%)
Total		5 238 765	22 040 586	

^a Australian production figures are missing from FAO FishStatJ databases.^b As this table includes all species cultured in brackish and marine waters, species cultured in inland brackish waters are included for completeness.^c See Sadek (2011).^d In the absence of a Latin name it is not known whether this generic term refers the culture of *Psettina brevirectis* or *Paralichthys olivaceus*, which is known as the bastard halibut, olive or Japanese flounder, given this uncertainty it is listed separately in the table.

Table 3. The top 25 farmed marine molluscan species ranked by the value of their respective industry. Data are extracted from the FAO FishStatJ database of production figures for 2011 and are based on the identifiable stocks. Although a number of additional high value, general classes of mollusc that are listed by country also appear within the database, the species composition of some of these cannot be determined and so are not included in the listing below

Common name	Latin name	Tonnage (t)	Value (US\$ × 1000)	Largest producer
Pacific cupped oyster	<i>Crassostrea gigas</i>	4 533 804	3 620 764	China (3 756 310 t; 82·85%)
Japanese carpet shell	<i>Ruditapes philippinarum</i>	3 681 436	3 438 190	China (3 546 502 t; 98·15%)
Yesso scallop	<i>Patinopecten yessoensis</i>	1 426 179	2 072 999	China (1 306 124 t; 91·58%)
Chilean mussel	<i>Mytilus chilensis</i>	288 583	1 148 561	Chile (100%)
Abalone species	<i>Haliotis</i> spp. ^a	86 445	676 004	China (76 786 t; 88·83%)
Constricted tagelus	<i>Sinonovacula constricta</i>	744 794	670 315	China (100%)
Peruvian calico scallop	<i>Argopecten purpuratus</i>	63 231	623 870	Peru (52 213 t; 82·58%)
Blood cockle	<i>Anadara granosa</i>	443 686	468 045	China (293 200 t; 66·08%)
Mediterranean mussel	<i>Mytilus galloprovincialis</i>	1 019 420	447 630	China (707 401 t; 69·39%)
Blue mussel	<i>Mytilus edulis</i>	176 445	348 965	France (61 800 t; 35·03%)
Green mussel	<i>Perna canalicula</i>	222 633	219 689	New Zealand (101 311 t; 45·51%)
Pearl oyster shells	<i>Pinctada</i> spp. ^b	55	216 255	Japan (20 t; 36·36%)
Japanese hard clam	<i>Meretrix</i> spp. ^c	119 771	165 460	Vietnam (~ 60 000 t; ~ 50·1%)
Sea snails	<i>Babylonia/Hemifusus</i> spp. ^d	203 266	123 992	China (100%)
Asian clam	<i>Corbicula fluminea</i>	36 983	65 756	China (22 327 t; 60·37%)
Northern quahog	<i>Mercenaria mercenaria</i>	28 841	62 873	USA (100%)
American cupped oyster	<i>Crassostrea virginica</i>	71 355	58 219	USA (67 975 t; 95·26%)
Grooved carpet shell	<i>Ruditapes decussatus</i>	41 38	52 834	Portugal (2315 t; 55·94%)
Sydney cupped oyster	<i>Saccostrea commercialis</i>	5 700	41 638	Australia (100%)
Korean mussel	<i>Mytilus coruscus</i>	70 416	29 247	Rep. Korea (100%)
Penguin wing oyster	<i>Pteria penguin</i>	48 449	27 471	Indonesia (100%)
Pacific geoduck	<i>Panopea generosa</i>	607	21 415	USA (100%)
Inflated ark	<i>Scapharca broughtonii</i>	2 110	20 248	Rep. Korea (100%)
Pen shells	<i>Atrina</i> spp.	30 126	19 281	China (100%)
S. Amer. rock mussel	<i>Perna perna</i>	15 970	12 776	Brazil (100%)
Total		13 324 443	14 652 497	

^a Consists of a number of species, including *Haliotis discus hannai*, *H. diversicolor*, *H. midae*, *H. rubra* and *H. rufescens*.

^b Principally *Pinctada fucata martensii* and *P. margaritifera*.

^c Figures consist of ~50% Taiwanese *Meretrix lusoria* and the 50% Vietnamese *M. lyrata* and *M. meretrix*.

^d Consists of a number of species, including *Babylonia areolata*, *B. lutosa*, *B. formosae* and *Hemifusus ternatanus* and *H. tuba*.

infections, and can assist in the development of more robust risk assessments and biosecurity practices, which can help mitigate against and/or minimise the potential impacts of parasite-mediated disease in aquaculture.

MATERIALS AND METHODS

Ranking of aquaculture industries

FAO's FishStatJ has been used as the principal source of data for ranking industries according to their commercial value and tonnage (see Tables 1–6); however, it should be emphasised that some caution needs to be exercised in the interpretation of both the original figures and those provided here given that (1) the species in FishStatJ are listed by common names rather than by a specific Latin binomial nomenclature or by a generic group, e.g. groupers, creating potential errors in correctly identifying and allocating a species to its true class and ascribing

an accurate tonnage and value; (2) the values returned for some species are estimated in the absence of precise data and are based on either best knowledge at the time of data submission, the returns submitted for previous years or represent cautious projections based on either national growth trends for each industry or nominal figures intended to demonstrate growth and development; and (3) aquaculture activities in some countries appear to have been omitted, e.g. Australia's production figures for Japanese yellowtail *Seriola quinqueradiata*. In the case of the latter, the figures may have been included and are listed under another category but are not easily identifiable. We have, therefore, attempted to identify national production figures for some of these industries and provide amended values.

Table 1 provides a breakdown of brackish/marine aquaculture by class. In addition to providing production figures and values for the major marine/brackishwater finfish (Table 2), molluscan (Table 3),

Table 4. The top 10 farmed crustacean species ranked by the value of their respective industry. Figures are generated from the FAO FishStatJ databases and are based on identifiable stocks

Common name	Latin name	Tonnage (t)		Value (US\$ × 1000)		Total value (US\$ × 1000)
		Brackish	Marine	Brackish	Marine	
Whiteleg shrimp ^a	<i>Litopenaeus vannamei</i>	2032 416	278 858	8 113 926	1 054 008	9 167 934
Giant tiger prawn ^b	<i>Penaeus monodon</i>	644 376	117 809	2 926 750	541 915	3 468 665
Indo-Pacific swamp crab ^c	<i>Scylla serrata</i>	36 668	121 641	166 966	289 015	455 981
Indian white prawn ^d	<i>Fenneropenaeus indicus</i>	20 851	21 007	99 411	252 000	351 411
Swimming crab ^e	<i>Portunus trituberculatus</i>	–	92 907	–	326 104	326 104
Kuruma prawn ^f	<i>Marsupenaeus japonicus</i>	275	52 628	3778	309 247	313 025
Fleshy prawn ^g	<i>Fenneropenaeus chinensis</i>	41 646	16	166 584	267	166 851
<i>Metapenaeus</i> spp. ^h	<i>M. monoceros</i> + <i>M. ensis</i>	35 047	–	124 760	–	124 760
Mud crab ⁱ	<i>Scylla (paramanosain?)</i>	109	17 401	431	70 300	70 731
Blue shrimp ^j	<i>Litopenaeus stylirostris</i>	–	1929	–	19 771	19 771
Total		2811 388	704 196	11 602 606	2 862 627	14 465 233

^a China is the largest brackish producer at 665 588 t; Mexico the largest marine producer at 106 886 t. FAO FishStatJ allocate 76 507 t of Chinese marine-farmed shrimp to 'marine *Penaeus*', it is believed that this may represent China's *L. vannamei* production and has been included within the marine figures.

^b Vietnam is the largest brackish producer at an estimated 300 000 t; China the largest marine producer at 60 691 t.

^c Philippines is the largest producer of crabs in brackish waters at 15 731 t; China the largest marine producer at 121 458 t.

^d Bangladesh is the largest brackish producer at 2364 t; Saudia Arabia the largest marine producer at 21 000 t.

^e China is the only producer listed.

^f Taiwan is the largest brackish producer at 178 t; whilst China the largest marine producer at 50 991 t.

^g China is the largest brackish producer at 41 646 t; Korea the largest marine producer at 16 t.

^h Bangladesh is the largest producer of identifiable *Metapenaeus* shrimp at 17 777 t.

ⁱ Taiwan is the only brackish producer at 109 t; China the only nation producing mud crabs in marine waters in 2011 with an annual production of 17 401 t.

^j Limited information available.

Table 5. Other important cultured marine species. These figures are generated from the FAO FishStatJ databases and are based on identifiable stocks

Common name	Latin name	Tonnage (t)	Value (US\$ × 1000)	Largest producer
Ascidacea				
Sea squirts	<i>Halocynthia roretzi</i>	12 369	20 842	Rep. Korea (11 676 t; 94·40%)
Cnidaria				
Jellyfish	<i>Rhopilema esculentum</i>	69 749	164 608	China (100%)
Echinodermata				
Echinoidea				
Sea urchins ^a	<i>Strongylocentrotus intermedius</i> , <i>S. nudus</i>	6791	23 514	China (6756 t; 99·48%)
Holothuroidea				
Japanese sea cucumber	<i>Apostichopus japonicus</i>	138 186	482 712	China (137 754 t; 99·69%)

^a Limited information available on species that are being cultured.

crustacean (Table 4), ascidian, cnidarian and echinoderm (Table 5) industries, we also consider farmed marine ornamental species (Table 6). Identifying global figures for the most popular traded ornamental species, however, has proven more difficult, and here we use information presented by Rhyne *et al.* (2012) on the number of fish imported into the USA during the period May 2004–May 2005 as an indicator. The shipment data, which was extracted from the Law Enforcement Management Information Systems (LEMIS) database maintained by the United States Fish and Wildlife Services (USFWS), was used to list the 20 most popular fish entering the USA. The

retail value of each ornamental species, for which a commercial aquaculture production unit could be identified, was determined from the average retail price in the USA determined from a minimum of three outlets.

Estimates of parasite-induced loss

The species rankings provided in Tables 2–6 were then used as a basis for identifying the principal parasite threats that have been reported to have resulted in economic loss. In addition to reporting the estimates of loss cited in peer-reviewed and

Table 6. The top 12 marine ornamental fish species, for which some commercial production units could be identified that were imported into the USA with details on their average retail price and estimated trade value. As numbers for global production are unknown, here we use numbers imported into the USA during the period May 2004–May 2005 as presented by Rhyne *et al.* (2012) as an indication of the potential size of each industry. Species are ranked by their estimated economic value based on their average retail prices (min. $n = 3$). Common names are taken from the Ornamental Fish International website (<http://www.ornamental-fish-int.org/>)

Common name	Latin name	Av. price US\$ (range)	No. fish p.a. USA ^a	Est. value imported US\$
Green Chromis	<i>Chromis viridis</i>	10.99 (6.99–12.99)	923 423	10 148 419
False percula	<i>Amphiprion ocellaris</i>	20.85 (14.99–39.99)	427 177	8 906 641
Flame angel	<i>Centropyge loricula</i>	44.32 (37.99–49.99)	133 634	5 922 659
Blue devil	<i>Chrysiptera cyanea</i>	7.32 (4.99–11.99)	729 730	5 341 624
Domino damsel	<i>Dascyllus trimaculatus</i>	7.59 (4.99–11.99)	624 625	4 740 904
Banggai cardinal	<i>Pterapogon kauderni</i>	27.99 (25.99–29.99)	164 414	4 601 948
Maroon clownfish	<i>Premnas biaculeatus</i>	32.84 (23.99–44.99)	117 868	3 870 785
Mandarinfish	<i>Synchiropus splendidus</i>	21.49 (18.99–23.99)	141 892	3 049 259
Three-stripe damsel	<i>Dascyllus aruanus</i>	6.74 (5.99–6.99)	438 438	2 955 072
Yellowtail damsel	<i>Chrysiptera parasema</i>	4.99 (4.99)	393 393	1 963 031
Tomato clownfish	<i>Amphiprion frenatus</i>	18.32 (14.99–23.99)	100 601	1 843 010
Royal gramma	<i>Gramma loreto</i>	17.49 (13.99–19.99)	91 592	1 601 944
Est. total			4 286 787	54 945 296

^a Figures extrapolated from the graphics presented in Rhyne *et al.* (2012).

grey literature accounts, we also provide estimates of direct fish loss based on the information given in the published record (i.e. number, size or age class of aquatic animals, tonnage, number of net-pens/culture systems affected, etc.) and the value (US\$ kg⁻¹) of the mariculture stock using a combination of the FAO FishStatJ statistics (value divided by tonnage) for the corresponding year of the loss. These latter figures, however, are for harvested, unprocessed stock. The age of stock affected is then corrected for using one of the relevant species listed in Table 7 and assuming a linear increment in the value of stock from the cost per juvenile to harvest-sized stock and a 100% probability of survival to harvest. Additional assumptions relevant to the loss, where appropriate, are provided as part of the entry in Tables 8–12.

RESULTS

Ranking of aquaculture industries

The world's leading aquaculture industries derived from FishStatJ for 2011 are ranked in Tables 2–6 by their market value. These tables include 69 of the 222 unique aquatic animal categories listed in the database and account for approximately 94% (i.e. 22 305 887 t) of the total weight of marine animals produced (i.e. 23 832 381 t) and 93% (i.e. US\$ 51 849.962 M) of their total value (i.e. US\$ 55 781.97 M). The database, however, also includes a number of general categories, e.g. 'aquatic invertebrates nei', 'marine fishes nei', which potentially embrace a large number of species that are produced in small volumes. The total number of aquatic

animals that are reared under brackish and marine aquaculture conditions is, therefore, likely to have been substantially underestimated here. Table 6 ranks the 12 most popular marine aquarium species, for which a commercial production unit could be identified, by their estimated retail value based on numbers imported into the USA (see Rhyne *et al.* 2012).

Tables 8–12 provide a summary of notable parasite-induced events with estimates of the costs incurred.

In 2011, global aquaculture production was 83 729 313 t (all species) of which 44 734 400 t (i.e. 53.43%) were produced in brackish or marine waters (see Table 1). Global aquaculture has increased at a rate of 6.4% year-on-year over the past five years and by 7.2% over that produced in 2010. By comparison, brackish–marine production has increased by 5.7% (over the past five years) and 6.6%, respectively. Of this, Asia produces 39 542 391 t (i.e. 88.39%), although brackish–marine algae represent over half (i.e. 52.46%, 20 743 150 t) of this. In this study, the top 30 fish species listed in Table 2 account for 89.89% by production tonnage of all brackish/marine aquaculture species that were produced in 2011. Likewise, the top 25 molluscs provided in Table 3 account for 94.04% of what was produced and the top 10 crustacean species given in Table 4 for 99.44%.

DISCUSSION

The data presented in Tables 8–12 provide a basic review of some of the major parasite events experienced by these production industries and, where possible, ascribe an estimate of the resultant loss

Table 7. Production details for a range of fish species that are used, in part, for the estimations of loss

Species	Harvest size (kg)	Time to harvest (month)	Stocking density (kg m ⁻³)	Juvenile size (g)	Price juveniles (US\$ fish ⁻¹)	Country	Reference
<i>Dicentrarchus labrax</i>	0.4–0.5	18–24	35–50	2.5	0.39	Across Eur prod	Lupatsch <i>et al.</i> (2010); www.fao.org (2014a)
<i>Diplodus puntazzo</i>	0.4–0.5	18–22	15	15	0.02	Spain	García García and García García (2010)
<i>Epinephelus</i> spp.	0.93–3.1	24	11.5–18.5	34–168	1.02–2.33	Vietnam	Petersen <i>et al.</i> (2011a)
<i>Lates calcarifer</i>	0.25–3.0	3–18	1–16 fish m ³	4 (?)	0.12–0.13	Vietnam	Petersen <i>et al.</i> (2011b)
<i>Oreochromis</i> sp.	0.55	5–6	1.1–2.3 fish m ³	20–25	0.04–0.07	Vietnam	Petersen <i>et al.</i> (2010)
<i>Scophthalmus maximus</i>	0.7–2.0	18–20	10 kg m ²	5	1.48–1.57	Across Eur prod	Watanabe and Daniels (2011); www.fao.org (2014b)
<i>Rachycentron canadum</i>	6–10	12–18	28 (juv) – 10–15 (ad)	1–1.5	1–2.50	USA	Kaiser and Holt (2005); www.fao.org (2014c)
<i>Salmo salar</i>	> 4	24–40	12.5–15	60–100	0.35–0.60	Global	Marine harvest (2013)
<i>Seriola</i> spp.	15–70	12–18	20–30	<50	4.8 (50 g)–14.3 (600 g)	Japan	Nakada (2008)
<i>Sparus aurata</i>	0.35–0.4	12–16	10–15	2–5	0.14–0.38	Across Eur prod	Koçak and Tathdil (2004); www.fao.org (2014d)
<i>Takifugu rubripes</i>	> 0.8	18–20	2.0–3.0	2–5	0.02	Japan	Kikuchi (2006); www.was.org

Across Eur prod, figures derived from European producers; ad, adult; juv, juvenile.

where not given in the original work. The tables show that there are elements of both predictability and unpredictability in infection events with elements of both certainty and uncertainty in the consequential economic losses. There are, for example, perennial costs associated with the management of and mitigation against salmon lice, *L. salmonis*, and other caligid species. Despite the apparent rise in the cost of 'sea lice control' in Norway, for example, from US\$ 33.4 M p.a. in 1996 (Kvenseth, 1997) to US\$ 206 M p.a. in 2009 (Costello, 2009), production over the same period increased by 248% from 297 557 t (1996) to 737 694 t (2008), and yet the price of salmon increased by only 20% (US\$ 3.08–3.71 kg⁻¹). During this time, there have also been significant improvements in salmon welfare, and the number of sea lice-related mortalities and the number of fish downgraded as a consequence of sea lice damage has fallen dramatically. To achieve this, the basic costs of sea lice control have increased from 3.64% (1996) of total production costs to 7.53% (2008). As sea lice pose a relatively consistent threat to sea-caged populations of Atlantic salmon, infection can be predicted and, therefore, factored into farm business plans and farm-level treatment strategies. In addition, area management agreements, which form part of national strategies for the control of lice in some countries, e.g. Scotland and Norway, can prompt additional treatments for the area-level management of lice infections. If these are not conducted or the mean lice burden exceeds agreed national threshold levels, then penalties can be imposed in some countries, e.g. Norway (Tallaksen, 2013).

An analysis of unpublished data by the current authors suggests that parasites account for an annual loss of 5.8–16.5% (i.e. US\$ 62–175 M; assuming £1 = US\$ 1.6) of the value of UK aquaculture production (across all species in both freshwater and marine systems). Although the FishStatJ database lists 17 farmed species for the UK in 2012, parasite-associated losses were largely dominated by the impact of AGD and mitigation strategies against sea lice on the Atlantic salmon industry. Although these estimates are only for UK losses and do not include losses due to bacterial and viral pathogens, they can still be considered low when compared with anecdotal reports suggesting that typical disease losses for certain aquaculture industries in Asia range from 30 to 50%. A significant proportion of these latter losses is attributable to the loss of juvenile stock within the hatchery/nursery phases of production, and for many industries, such losses are commonly factored into and accepted as part of typical production cycles. As a consequence of such fatalistic acceptance of their inevitability, the ubiquity of low-specificity pathogens in some warm-water aquaculture systems and the small-scale nature/lack of diagnostic capacity of some production enterprises,

Table 8. The estimated economic cost of notable protistan and metazoan parasite events on some of the world's leading marine and brackish water finfish production industries. For each industry, protistans and metazoans are grouped separately and are listed alphabetically within each. The order in which the finfish aquaculture industries are listed is based on their global economic value in 2011 according to FAO statistics and the rank order provided in Table 2. For each parasite-induced event, brief details are provided and an estimate of loss, where possible, has been calculated. Although 'cyprinids' cultured in brackish water are listed in Table 2, it is not possible to identify all the species that are embraced within this and as such are not considered here

Parasite	Impact	Estimated loss (US\$)	Reference
(a) Atlantic salmon, <i>Salmo salar</i> L.			
<i>Desmozoon lepeophtherii</i> (syn. <i>Paramucleospora theridion</i>)	Mortalities (~ 600–1000 fish net-pen ⁻¹) in 6 net-pens were seen throughout March–May 2002 in northern Norway. Fish were bleeding from the eyes and each fish had a mass of myxozoan trophozoites covering their pseudobranchs. A value of US\$ 2.11 kg ⁻¹ for harvest-sized Norwegian salmon in 2002 is used to estimate loss	2532	Karlsbakk <i>et al.</i> (2002)
	35% of the 200 000 smolts transferred to sea pens in northern Norway in Sept. 2001 were lost by March. An average weight of 250 g fish ⁻¹ and a value of (US\$ 2.11 kg ⁻¹ for the price of harvest-sized Norwegian salmon in 2002 is used to estimate loss	36 925	Sterud <i>et al.</i> (2003)
	Mortalities in northern Norway re-emerge in 2003, with mortalities ranging from low grade to significant (40%). The study uses material collected from 13 <i>S. salar</i> sites and 2 <i>O. mykiss</i> sites. Loss is estimated on the assumption that a single 1575 m ³ cage at each site stocked at 12.5 kg m ³ was affected with resultant mortalities of between 5 and 10%. The harvest price of Norwegian salmon in 2003, i.e. US\$ 2.28 kg ⁻¹ is applied	33 664–67 328	Nylund <i>et al.</i> (2005)
	Study looked at 55 Norwegian farms in 8 counties, 43 of which had either proliferative gill disease (PGI), pancreas disease (PD), heart and skeletal muscle inflammation (HSMI) and cardiomyopathy syndrome (CMS). Analysis found high prevalence and densities of the parasite in farms in southern Norway where PGI and PD predominate. Heavy infections were found in fish with PGI suggesting that parasitic infection may be the primary cause of mortality in fish with PGI	–	Nylund <i>et al.</i> (2011)
	A production site in the Scottish Highlands experienced mortalities due to a seasonal gill disease – fish were lethargic and had pale, thickened gill filaments. S1 smolts were stocked into 10 cages in Feb 2009 and mortalities were observed throughout Sept 2009–Jan 2010 with weekly mortalities peaking at 0.36% in Oct 2009. Total site mortality was 8.7%. Loss is based on the assumption that cages measured 15 × 15 × 7 m ³ , were stocked at 15 kg m ³ , that the size of smolts were 100 g (i.e. 2 362 500 fish stocked), at the time of the event the fish were 400 g and were not graded out prior to the event, and, the harvest price of Scottish salmon in 2010, i.e. US\$ 4.42 kg ⁻¹ , applied to the total lost weight of fish (i.e. 82 215 kg)	363 390	Matthews <i>et al.</i> (2013)
<i>Hexamita salmonis</i>	Increased mortality and morbidity in 200–250 g stock held at a farm in northern Norway in Nov 1989 was reported. The smolts had been transferred to sea in August 1989. Impression smears from the kidney, cardiac atrium, spleen, pyloric caeca, posterior gut, eye and brain revealed a large number of flagellates. One month later, another case of systemic hexamitosis was reported in a second Norwegian farm site. The fish at each site had originated from the same freshwater farm but at different time periods. No details relating to the magnitude of losses were provided	–	Mo <i>et al.</i> (1990)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
	Infections are associated with a severe systemic granulomatous disease resulting in increased mortality of stock and the rejection of up to 60% of the harvested stock in northern Norway. Loss is based on the assumption that at least three 20 t production units were affected, a loss/rejection of 30% of stock, and, the harvest price of Norwegian salmon in 1992, i.e. US\$ 4·81 kg ⁻¹	86 580	Poppe and Mo (1993)
<i>Hexamita</i> sp.	Infections were found in 4 northern Norwegian sites throughout 1989–1992. Large granulomatous boils were typically found in all the internal organs in 200–250 g post-smolts. At 1 site, boils within the caudal muscle, liver and kidney of 4–5 kg fish were found in 60% of the stock filleted from 1 cage site. The fish from all 4 farm sites originated from the same hatchery in 1988–1989, although the hatchery did bring in several thousand alevins from the south of Norway in 1987. A subsequent infection in 3 classes of fish at a fifth site, situated 200 m from one of the farms and within 100 m of a slaughtering facility, was found in 1992. Infection differs from the gut infections typically seen with <i>Hexamita salmonis</i> . Loss is estimated on a 50% downgrade of the infected stock (i.e. 60%) reported from 1 cage of fish (750 m ³ ; 12·5 kg m ³ stocking density) and the harvest price of Norwegian salmon in 1991, i.e. US\$ 4·20 kg ⁻¹	11 813	Poppe <i>et al.</i> (1992)
<i>Ichthybodo necator</i>	Large mortalities were seen in S2 smolts 6 weeks after their transfer into seawater cages in July 1977. The source of infection was believed to be from parasites surviving a malachite green treatment in freshwater 3 weeks prior to fish transfer. Low summer salinities (min 23‰) and rising sea temperatures (max 16 °C) are considered contributory factors	–	Ellis and Wootten (1978)
<i>Kudoa thyrssites</i>	A mortality rate of 0·57% day ⁻¹ was seen in smolts ($n = 1200$; av. ~55 g) 5 months following their transfer to a marine site. Although no figures of the total loss are provided, if an av. loss of 0·4% day ⁻¹ and an av. weight of 70 g fish ⁻¹ is assumed over 120 days, then a 50% cumulative mortality might have resulted, i.e. the loss of 42 kg of fish. Loss estimates are based on the value harvest-sized fish in the USA in 1986 (first figures available), i.e. US\$ 8·33 kg ⁻¹ . <i>Kudoa</i> as the cause of mortality has, however, been questioned	350	Harrell and Scott (1985) Moran <i>et al.</i> (1999)
	Processors reported that the prevalence of myoliquefaction in smoked fillets prepared from pen-reared stock can range from 2 to 25%. Loss is based on the assumption that all infected fillets were rejected and on the value of harvest-sized fish produced on the Pacific NE of Canada in 1991, i.e. US\$ 5·33 kg ⁻¹ . Canadian Pacific production in 1991 was 3651 t, industry-wide losses would have been between ~75 and 913 t	5330 t ⁻¹ of rejected fish or 107–1333 t ⁻¹ of production. ~0·4–4·8 M across the entire industry	Whitaker and Kent (1991)
	Infection leading to a reduction in fillet quality costs the industry in British Columbia est. US\$ 15 M in 2010	15 M	Martell <i>et al.</i> (2013)
<i>Myxobolus cerebralis</i>	>90% mortalities in net-pen-reared smolts in Ireland over the summers 1992–1994 were attributed to intracellular presporogonic multicellular developmental stages of	8135 per cage	Rodger <i>et al.</i> (1995);

	the parasite-inducing encephalitis. Foci of non-suppurative encephalitis in the myelencephalon significant; appears 26 days post-introduction. Assuming that cages were 125 m ³ and stocked at a min. 15 kg m ³ , then loss estimates are based on the value of harvest-sized Irish fish in 1994, i.e. US\$ 4.82 kg ⁻¹		Scullion <i>et al.</i> (1996); Frasca <i>et al.</i> (1998, 1999)
<i>Paramoeba perurans</i> (Amoebic Gill Disease, AGD)	Mortality of infected smolts in Tasmania may reach 10% week ⁻¹ , 2–4% week ⁻¹ in fish weighing 1–2 kg and 1–2% week ⁻¹ in fish over 2 kg	–	Foster and Percival (1988)
	Mortality variable but can reach 2% day ⁻¹ and up to 50% in untreated cages. Juvenile fish in the first season usually affected when water temps >12 °C and salinities approach 35‰	–	Munday <i>et al.</i> (1990)
	Infection confirmed at 8 Irish farms in post-smolts (450–800 g) in Oct–Nov 1995 resulting in >10% mortality of stock at 2 sites, <5% at a further 3 sites and no significant losses at the remaining 3 sites. Loss is based on the value of harvested Irish salmon in 1995, i.e. US\$ 5.00 kg ⁻¹	250–500 t ⁻¹	Rodger and McArdle (1996)
	Infections account for an estimated 10–20% of production (i.e. 1200–2400 t) in Tasmania in 2001. Production losses are subsequently suggested to be ~14% (i.e. ~1780 t). Annual loss of ~60% stock at 1 Irish site since 1996. Assuming that an unaffected site would harvest 75 000 fish at 4 kg fish ⁻¹ , and that the smolts av. 70 g fish ⁻¹ transferred in Oct would have an expected min weight of ~450 g fish ⁻¹ by the following July (i.e. 1.136 g day ⁻¹ weight gain), a summer loss of 60% of stock would be ~20.25 t. The loss is estimated on the value of harvest-sized fish in Ireland in 2004, i.e. US\$ 4.54 kg ⁻¹	5–10 M 7.2 M 91 935	Munday <i>et al.</i> (2001); Morrison <i>et al.</i> (2006) Adams and Nowak (2001) Birmingham and Mulcahy (2004)
	Outbreaks lasting 7–12 weeks at 4 Norwegian farms in the spring of 2006 resulted in mortality rates of 12–20% at 3 of the farms and 82% at the fourth site in 1.0–1.6 kg salmon. Loss is est. on av. 1.3 kg salmon being lost from at least 4 cages (50 × 50 × 20 m ³) stocked at 12.5 kg m ³ at each site with mortality rates of 12, 16, 20 and 82% and using a value of US\$ 3.86 kg ⁻¹ for Norwegian salmon in 2006	12.55 M	Steinum <i>et al.</i> (2008)
	Losses at 3 Chilean farm sites in 2007 (all with concomitant <i>Caligus rogercresseyi</i> infections) are recorded as 7.7, 5.4 and 1.8%, i.e. 4.9 ± 4.5 loss across all farms with an av. 66% prevalence of amoebae. Highest mortalities coincide with highest salinities and temps. Low feed intake led to a 25% reduction in growth. Loss estimates are based on the value of harvest-sized fish in Chile in 2007, i.e. US\$ 7.90 kg ⁻¹	142–608 (387 ± 356) t ⁻¹ of production	Bustos <i>et al.</i> (2011)
	Oct 2011, the first loss of Scottish Atlantic salmon (279 000 fish) to AGD is reported	–	Vass (2013)
	Typical losses in Scotland are put at between 10 and 20% but are as high as 70% at certain Scottish sites	–	Marine Scotland (2012)
	Estimate of loss put at >US\$ 48 M (i.e. ~9000 t)	>48 M	Vass (2013)
	According to the Scottish Environmental Protection Agency (cited in FAO Globefish; http://www.globefish.org/salmon-june-2013.html), 13 600 t of dead salmon were disposed of (cost of Scottish salmon=US\$ 5.96 kg ⁻¹ in 2011)	~81 M	FAO Globefish (2013)
<i>Spiroplasma bairdii</i>	Systemic infections resulted in increased mortality and morbidity in a cage of 200–250 g post-smolts in northern Norway in Nov 1989 when seawater temperatures were 6 °C. Infections also found as boils in the caudal muscle causing severe lesions in adults (4–5 kg) at 1 site in June 1991; approx. 60% of the stock had lesions. A total of 4 fish farms were affected	–	Mo <i>et al.</i> (1990); Poppe <i>et al.</i> (1992); Poppe and Mo (1993)
	Fish mortality occurs in both the blood and tissue phases of the disease	–	Guo and Woo (2004)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
<i>Tetrahymena</i> sp.	An increased rate of mortality was recorded in ~1-year old salmon stocked in 8 m diameter concrete tanks. No details relating to the pattern or magnitude of loss are provided	–	Ferguson <i>et al.</i> (1987)
<i>Trichodina</i> sp.	In July 1982 and 1983, there was a 20% mortality of broodstock in an Irish sea cage when sea temperatures rose to 23 °C and when many of the male fish had begun sexual maturation. There were no appreciable losses in an adjacent cage of younger salmon. Assuming that cages were min. 125 m ³ , stocked at 12.5 kg m ³ , then the loss of fish would have been >312.5 kg. Figures of loss are based on the harvest price of Irish salmon in 1984, i.e. US\$ 6.50 kg ⁻¹	2031 per cage	McArdle (1984)
<i>Caligus</i> spp.	Infections and the delousing of harvest fish in Chilean processing plants incur costs of US\$ 0.30 kg ⁻¹ . Loss is based on the reported production of Chilean salmonids of 97 000 t in 1995	29.1 M	Carvajal <i>et al.</i> (1998)
<i>Caligus elongatus</i>	Infections on Scottish stock reared in cages are seasonal and peak at 25+ lice fish ⁻¹ in July–Sept 1991. Infections commonly co-occur with <i>L. salmonis</i> (see the entry below)	–	Bron <i>et al.</i> (1993)
<i>Caligus elongatus</i> + <i>Lepeophtheirus salmonis</i>	Infections on Irish cage-reared salmon were reported to peak at an av. 11.9 ± 5.2 s.d. lice (2:1 <i>C. elongatus</i> : <i>L. salmonis</i>) in August 1987 declining to 3.3 ± 3.3 s.d. by mid-Sept. Lice numbers are controlled by organophosphate treatments	–	Tully (1989)
	Infections that were monitored at several Scottish cage sites over the period July 1990–March 1992 were observed to peak at 100+ lice fish ⁻¹ in Oct 1990 and March 1991. Lice burdens necessitated 9 treatments at single-year class sites and 11 treatments for multi-year class sites across the 20 months monitoring period. Average cost of cage treatment alone is est. to be ~ US\$ 10 000 per 200 000 fish (see Mustafa <i>et al.</i> 2001)	0.09–0.11 M per 200 000 fish over 20 months	Bron <i>et al.</i> (1993)
	92% of salmon at 1 site at Eastport, Maine throughout 1993–1994 were infected with an av. 3.1 lice fish ⁻¹ in Sept 1993, 99.6% of which were <i>C. elongatus</i> . In addition, 87% of smolts were infected with an av. 2.1 lice fish ⁻¹	–	Shaw and Opitz (1993)
<i>Caligus longirostris</i>	Reported with a prevalence of 1.5% on av. 5.51 kg salmon in 1 Tasmanian cage site and 1.9% on 5.62 kg fish ⁻¹ in a second, experimental cage site. Low-level infection	Negligible	Nowak <i>et al.</i> (2011)
<i>Caligus rogercresseyi</i>	The costs of removing lice from Chilean salmon in processing plants costs ~ US\$ 0.30 kg ⁻¹ according to the Asociación de Productores de Salmón y Trucha de Chile. Loss is based on the 96 675 t of Atlantic salmon produced in 1997	29 M	Carvajal <i>et al.</i> (1998)
	Cost to Chilean industry ~ US\$ 193.6 M p.a.	193.6 M	Costello (2009)
<i>Caligus teres</i>	Infections were monitored on sea-pen-reared stock in Puerto Montt, Chile over April 2000–Feb 2001. Infections were observed to peak in mid-December with an av. 27 mobile lice and 9 chalimus on 2.3 kg fish. Delousing the 4000 t of stock with ivermectin was conducted at a cost of US\$ 0.004 kg ⁻¹	16 000	Bravo (2003)
<i>Ceratothoa gaudichaudii</i>	Prevalence of isopod infection on 0.9–1.1 kg salmon in Chile rose from 33.1% (1.37 parasites fish ⁻¹) in May 1993 to 99.9% (6.05 parasites fish ⁻¹) in Aug 1994. Infection	1571 per 1000 harvest-sized fish	Sievers <i>et al.</i> (1996)

	had a significant impact on growth across the study period with average terminal weights determined as 4.427 kg (0–2 isopods fish ⁻¹), 4.151 kg (3–7 isopods fish ⁻¹) and 3.763 kg (> 8 isopods fish ⁻¹). This equates to loss of 413.4 kg or US\$ 1571 (assuming US\$ 3.8 kg ⁻¹ based on 1996 prices for Chilean salmon) per 1000 salmon		
<i>Ergasilus labracis</i>	Ten floating cages (3–6000 fish cage ⁻¹), 14‰, 18–20 °C, 100% infection with av. 84.6 parasites fish ⁻¹ (range 44–2229), severe gill hyperplasia, 90% gill damage, loss of 4000 fish (85–125 mm long) over 4 days. Loss is based on US\$ 1 smolt ⁻¹	4000	Hogans (1989); O'Halloran <i>et al.</i> (1992)
<i>Eubothrium crassum</i>	Cestodes within the pyloric caeca result in a direct economic loss of food uptake impacting detrimentally on growth (infection of av. 71 worms fish ⁻¹ effects loss of 800 g in 5.2 kg males, whilst 115 worms fish ⁻¹ effect a loss of 440 g in 4.3 kg females), i.e. a ~10% loss in the size of market fish. This equates to a minimum loss of ~ US\$ 1848 per 1000 harvest-sized fish (assumes US\$ 4.20 kg ⁻¹ based on Norwegian prices in 1991)	1848 per 1000 harvest-sized fish	Bristow and Berland (1991)
	Potential loss of growth owing to chronic low worm burdens est. at between 10 and 20%. This equates to an average loss of between US\$ 2253 and 4506 per 1000 harvest-sized fish (assumes US\$ 4.506 kg ⁻¹ based on Scottish salmon prices in 1993)	2253–4506 per 1000 harvest-sized fish	Mitchell (1993)
	Impact on growth and haematocrit confirmed; no correlation between worm burdens and fish weight	–	Saksvik <i>et al.</i> (2001)
<i>Gyrodactyloides bychowskii</i>	Infections of up to several hundred flukes per fish (post-smolts to 2.5 kg fish) were reported at 20 Norwegian sea-cage farm sites between 1989 and 1991. Hypertrophy and hyperplasia of the gill epithelium was commonly observed associated with infections	–	Mo and MacKenzie (1991)
	Parasite burdens of up to 200 flukes per gill arch were observed on 20% of fish examined in June 1999 from a farm site in the Shetland Isles, Scotland. Prevalences of between 0.5 and 10% were observed throughout 1999–2000 on 3 Scottish farm sites. Mild to marked epidermal hyperplasia, slight oedema and hypertrophy was seen at the sites of parasite attachment	–	Bruno <i>et al.</i> (2001)
<i>Lepeophtheirus salmonis</i>	Direct losses, treatments and lost growth imposed by 'sea lice', i.e. <i>C. elongatus</i> and <i>L. salmonis</i> are estimated to cost the Norwegian industry US\$ 33.4 M	33.4 M	Kvenseth (1997)
	Fish growth reduced by 5–15%. 5% more feed needed. Total tonnage produced in Scotland in 1997 was 110 917 t suggesting loss production may have been in the region of 5546–16 638 t. Estimates of loss are based on the value of harvest-sized fish in Scotland in 1997, i.e. US\$ 2.94 kg ⁻¹	16.3–48.9 M	Sinnott (1998)
	Suggested that a Canadian farmer can expect to lose US\$ 162 000 per 200 000 fish. Infection accounts for a 5% loss in food conversion necessitating the provision of additional feed; a 1–3% downgrading of carcasses; a 1% loss due to secondary infections; treatment costs (i.e. ~ US\$ 9000 per hydrogen peroxide treatment and US\$ 10 800 per azamethiphos treatment) and resultant losses	0.16 M per 200 000 fish	Mustafa <i>et al.</i> (2001)
	Cost to Scottish industry US\$ 52.9 M in 2000 equal to ~7–10% of total production value, i.e. control costs US\$ 0.39 kg ⁻¹	52.9 M	Rae (2002)
	Sea lice are estimated to cost the Canadian salmon industry ~ US\$ 16 M	16 M	Roth (2000)
	Up to 15% of Scottish fish downgraded		Michie (2001)
	Parasiticides, staff and equipment represent 17–30% of the total sea louse control costs	–	Mustafa <i>et al.</i> (2001)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
	Estimated that the stress of sea louse burdens account for 5% lost growth (i.e. US\$ 20.8 M based on the value of the Scottish 130 000 t industry in 2000). Medicines account for a further US\$ 6.4–8 M, whilst the remainder is associated with treatment costs (i.e. labour, accidental mortalities during treatment, the downgrading of lice-damaged stock). Costs are based on £1=US\$ 1.6	32–48 M	Rae (2002)
	Cost to Norwegian industry ~ US\$ 206 M p.a.	206 M	Costello (2009)
	If the estimated costs of sea lice control remain unchanged (i.e. € kg ⁻¹) from those given in Costello (2009), then based on salmon production in 2012, sea louse costs for the main producing nations are estimated at US\$ 468 M for Norway (0.19 € kg ⁻¹ ; 1.18 M tonnes), US\$ 60.5 M for Chile (0.19 € kg ⁻¹ ; 232 700 t) and US\$ 55.5 M for Scotland (0.25 € kg ⁻¹ ; 162 223 t)	468 M (Norway) 60.5 M (Chile) 55.5 M (Scotland)	
	Norwegian Food Safety Authority ordered 5 producers operating north of Vikna, Norway to slaughter of 8000 t of fish (3.5–4 kg class) because of treatment failure/resistance/lice burdens exceeding permitted national threshold of 0.5 adult female lice fish ⁻¹ . Failure to comply before the imposed deadline could have incurred a daily penalty of ~ US\$ 16 250. Loss estimates here are due to the premature harvest of stock and can be speculated at 0.5–1 kg fish ⁻¹ (i.e. 1143–2000 t) and are based on the value of harvest-sized fish in Norway in 2011, i.e. US\$ 4.58 kg ⁻¹	5.23–9.16 M	Tallaksen (2013)
Sea lice ^a	The costs of sea lice control in Chile are estimated at US\$ 0.022 kg ⁻¹ . Loss is estimated on the Chilean production of 253 607 t in 2001	5.58 M	Bravo (2003)
	US\$ 480 M p.a. cost to global salmonid industry (incl. costs of parasiticide, impact on growth and food conversion), i.e. av. cost of sea lice control is US\$ 0.30 kg ⁻¹	480 M	Costello (2009)
(b) Rainbow trout, <i>Oncorhynchus mykiss</i> (Walbaum, 1792)			
<i>Kudoa thyrsites</i>	A 2.6% infection was found within the cardiac muscle of fresh, dead fish collected from a single hatchery on the coast of British Columbia, Canada in Sept 1987	–	Kabata and Whitaker (1989)
<i>Paramoeba perurans</i>	Mortality of Tasmanian salmonids, including <i>O. mykiss</i> in the period 1987–1988, variable – can reach 2% day ⁻¹ and up to 50% in untreated cages. Details relating to the impact of AGD on the production of Atlantic salmon (53 t in 1987; 240 t in 1988) and rainbow trout (207 t in 1987; 890 t in 1988) not given. Juvenile fish in their first season are usually affected when water temps >12 °C and salinities approach 35‰. Loss is based on an estimated 20% loss of fish numbers both in 1987 (i.e. 17 250 fish) and in 1988 (i.e. 74 166 fish) that first season fish were an av. 200 g, that stock are harvested at 3 kg and on the value of harvest-sized fish in 1987 as US\$ 7.49 kg ⁻¹ and US\$ 5.49 kg ⁻¹ in 1988	Loss of 3.45 t (1987) to 14.83 t (1988) worth 25 840–81 430	Munday <i>et al.</i> (1990)
	Chronic loss of fish at 1 Irish sea farm beginning in July 1982 resulting in a >50% loss of stock. Recorded as the driest July on record. Loss is based on the value of harvest-sized Irish rainbow trout in 1984, i.e. US\$ 3 kg ⁻¹	1500 t ⁻¹	Rodger and McArdle (1996)
	Sub-optimal osmoregulatory performance of infected stock in Tasmania in full strength seawater has led to their culture in brackish water	–	Findlay <i>et al.</i> (1995)
<i>Trichodina</i> sp.	In July 1982, there was a 20% mortality of 1+ trout in an Irish sea cage when sea temperatures rose to 23 °C and many of the male fish had begun sexual maturation.	600 t ⁻¹	McArdle (1984)

	Similar losses were also seen during the summer of 1983. Although <i>Trichodina</i> was predominant, some of the infections were a mix of <i>Trichodina</i> and <i>Ichthyobodo</i> . Loss is based on value of harvest-sized Irish rainbow trout in 1984, i.e. US\$ 3 kg ⁻¹		
<i>Caligus orientalis</i>	Heavy burdens of up to 188 lice fish ⁻¹ were found on a cage of 320 fish (av. wt. 597 g) reared in Lake Mokoto on the coast of the Okhotsk Sea, NE Hokkaido, Japan during the summer of 1988 when surface water temperatures ranged from 16 to 23 °C. The fish lost their appetite and most died within a month. Similar infections on stocks reared in Lake Saroma, NE Hokkaido throughout 1986 and 1987 resulted in the stop of production. Loss is based on the assumption that 90% of the Lake Mokoto stock was lost and the harvest price for Japanese freshwater rainbow trout (no figures for marine produced stock) in 1988, i.e. US\$ 4.19 kg ⁻¹	720	Urawa and Kato (1991)
<i>Caligus teres</i>	Infections were monitored on sea-pen-reared stock in Puerto Montt, Chile over April 2000–Feb 2001. Infections were observed to peak in mid-December with an av. 27 mobile lice and 8 chalimus on 1.7 kg fish	–	Bravo (2003)
<i>Lepeophtheirus salmonis</i>	The lice burdens on stock (300–700 g) reared in sea cages sited in Katsurakoi Harbor near Kushiro on the Eastern Pacific coast of Hokkaido, Japan in 1988 were 0.02 ± 0.15 lice fish ⁻¹ (prev. 2.2%) in Sept 1988 rising to 3.8 ± 2.5 lice fish ⁻¹ (prev. 92%) in Nov 1988. Peri-anal haemorrhages were observed but not thought to cause serious disease	–	Urawa <i>et al.</i> (1998a)
	In Nov 1992, 20 000 juvenile fish (<1-year old; 103–161 g) were transferred to net-pens in Onmae Bay, Miyagi Prefecture, Japan. One month later, 60% of the stocks were infected. By Feb 1993, all fish were infected with an av. 3.0 lice fish ⁻¹ . Thereafter, lice burdens rose sharply from 5.2 lice fish ⁻¹ in April to 36.8 lice fish ⁻¹ (range 19–51 lice fish ⁻¹) in July 1993. Fish had visible abrasions and haemorrhages. Loss is based on the assumption that fish harvested in July would have been downgraded (1–3%, see Mustafa <i>et al.</i> 2001), fish weighed ~ 300 g; harvest price for stock in 1993 would have been US\$ 6.24 kg ⁻¹	374–1123	Ho and Nagasawa (2001)
(c) Milkfish, <i>Chanos chanos</i> (Forsskål, 1775)			
<i>Amyloodinium ocellatum</i>	An infection resulted in the complete loss of hatchery stock (14 days old; ~ 8 mm long) in two 10 m diameter tanks at one establishment in Iloilo, the Philippines in June 2001. Loss is estimated on a stocking density of 10 kg m ⁻³ (Yap <i>et al.</i> 2007) and a value of US\$ 0.01 fish ⁻¹ (~ 0.1 g), i.e. the loss of 2 million fry	20 000	Cruz-Lacierda <i>et al.</i> (2004)
<i>Trichodina</i> sp.	Infected juvenile stock sampled from ponds at Pagbilao, Quezon Province and from the Laguna de Bay, Binangonan, Rizal Province, Philippines (salinity range from 2 to 30 ppt) in Sept 1985–April 1986 were found at 5 sites with a prevalence of 8–60%. Flagged as a concern	–	Regidor and Arthur (1986)
<i>Alitropus typus</i>	Infections have been reported to result in the mortality of Filipino held stock	–	Velasquez (1979; cited in Regidor and Arthur 1986)
<i>Caligus</i> sp.	Wild-caught stocks (5–12 kg) caught in Feb–May 1976 were grown on in 10–100 t tanks at a Research Station at Mag-aba, Pandan, Antique, Philippines. When a single fish in this group died in June 1976, the stock was found to be infected with lice. Fish with heavy burdens were emaciated. The infection (av. lice burdens not specified) were controlled by a single 0.25 ppm treatment of Neguvon (2,2,2,-trichloro-1-hydroxyl-phosphoric acid dimethylethol)	–	Laviña (1977, 1978)
<i>Caligus chanos</i>	Heavy lice burdens on Taiwanese stock in the winter of 1980–1981 were reported to cause severe lesions resulting in the death of fish. No details provided	–	Lin (1989)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
<i>Caligus epidemicus</i>	Infections were recorded on juvenile fish sampled from 5 sites (salinity range from 2 to 30 ppt) from ponds at Pagbilao, Quezon Province, from others at Molo, Iloilo Province, and, from the Laguna de Bay, Binangonan, Rizal Province, Philippines in Sept 1985–April 1986. Prevalence of infection ranged from 4 to 36% with up to 8.8 ± 13.2 lice fish ⁻¹	–	Regidor and Arthur (1986)
<i>Diergasilus kasaharai</i>	Report of a mass mortality of stock within a brackish water pond in a village in Tainan County, Taiwan in Dec 1993 stocked with 20 000 milkfish. Mortalities began with 10 fish day ⁻¹ rising to 100 fish day ⁻¹ after a week, each fish had ~ 130 parasites. No details are provided on the size of fish. Loss is estimated on the assumption that fish were on the lower side of their harvestable size range (i.e. 250 g), and, on their value in Taiwan in 1993, i.e. US\$ 1.60 kg ⁻¹ . Furthermore, it is assumed that a minimum 20% of stock was lost, i.e. 4000 fish	1600	Lin and Ho (1998)
Digenea unidentified sp.	Metacercariae were recorded on juvenile fish sampled from 5 sites (salinity range from 2 to 30 ppt) from ponds at Pagbilao, Quezon Province and from the Laguna de Bay, Binangonan, Rizal Province, Philippines in Sept 1985 to April. Prevalences of infection ranged from 8 to 90% with up to 5.2 ± 5.0 metacercariae fish ⁻¹	–	Regidor and Arthur (1986)
<i>Ichthyoxenus</i> sp.	Infections have been reported to kill cultured Filipino held stock. Details unknown	–	Velasquez (1979; cited in Regidor and Arthur 1986)
<i>Transversotrema laruei</i>	Infections were observed at 6 pond sites (2–30 ppt) at Pagbilao, Quezon Province, Philippines in Sept 1985–April 1986 with prevalences of between 8 and 76% with up to 5.2 ± 5.2 parasites fish ⁻¹	–	Regidor and Arthur (1986)
Unidentified cymothoid isopod	Reported to have resulted in the mortality of juvenile stock in the Philippines. Details unknown	–	Ronquillo and Caces-Borja (1960; cited in Regidor and Arthur, 1986)
(d) Japanese amberjack/yellowtail, <i>Seriola quinqueradiata</i> (Temminck et Schlegel, 1845) and other seriolids			
<i>Ceratomyxa buri</i>	A 100% infection was found in the bile of 0+ aged sea-pen reared stock sampled from Oita Prefecture, Japan in Nov 1999. The relationship between the occurrence of this parasite and the green livers observed in this host requires establishing	–	Yokoyama and Fukuda (2001)
<i>Ceratomyxa seriolae</i>	Examination of the bile from 0+ aged sea-pen reared stock sampled from Oita Prefecture, Japan in Feb 2000 found 70% prevalence. Whether this parasite is responsible for the green livers observed in this host requires investigating	–	Yokoyama and Fukuda (2001)
<i>Ichthyophonus hoferi</i>	Juvenile fish in Mie Prefecture were reported as being infected within the first year of culture. A low incidence of infection was reported but resulting in mortality. No details given	–	Egusa (1983)
<i>Kudoa amamiensis</i>	Infections have impacted on the commercial value of harvested stock. In June 1952, 63.6% of the 0.6–1.3 kg fish ($n = 11$) that were sampled from Nagasaki Prefecture, Japan had infected trunk muscles. In 1970, in Ehime Prefecture, 70 000 fish were reared on a trash fish diet until 400–500 g; almost all the valuable tail sections of the fish were infected.	63 000–80 000	Egusa and Nakajima (1978); Egusa (1983)

	The subsequent examination of a further 100 000 (~ 400 g) tail muscle sections found that ~ 30% were infected. Figures based on a back projected historical price of harvest stock in 1970 of US\$ 2 kg ⁻¹		
	A survey of 10 farm sites between 1994 and 1998 indicates that between 90 and 100% of stock in the Motobu area (2 sites) of Okinawa Prefecture, Japan were infected, whereas elsewhere in the Prefecture infection rates are in the region of 0–9% (12 sites). Loss is estimated on the assumption that each site produces 20 t, that infected fish are not sold, and, on the price of Japanese amberjack in 1998, i.e. US\$ 7.36 kg ⁻¹ . Loss in the Motobu area is calculated to be 37.954 t (i.e. 94.89% of stock), whilst losses for the rest of Okinawa are 1.52 t (i.e. 0.95% of stock)	279 341 for the 2 Motobu sites 11 187 for the remaining 8 Okinawa sites	Sugiyama <i>et al.</i> (1999)
<i>Kudoa megacapsula</i>	Disease outbreaks in Oct 1999–Jan 2000 were reported within 4 prefectures in Western Japan. The prevalence of infection was recorded at 12%. Infections appear as unsightly filamentous black cysts in skeletal muscle without consequential myoliquefaction. Loss is estimated on the basis of four, 20 t production units, a 12% loss of market-sized fish and the value of harvest stock in Japan in 2000, i.e. US\$ 9.71 kg ⁻¹	93 216	Yokoyama <i>et al.</i> (2006)
<i>Kudoa pericardialis</i>	Trophozoites were found in the pericardial cavity of stock at numerous farms in Kagoshima, Kouchi, Mie, Miyazaki, Shizuoka and Tokushima prefectures, Japan. Does not affect the harvest price because infections within the heart are inconspicuous	–	Nakajima and Egusa (1978)
<i>Kudoa yasunagai</i>	Infection results in abnormal swimming behaviour (whirling, lying on the bottom of the cage) and scoliosis of the spine. As a consequence feeding and swimming performance is impaired; fish suffer skin damage from swimming into the net. In 2011, a mortality event in a cage of juveniles situated at Wakayama, Japan was recorded 2 months post-transfer. Over 50% of the fish showed signs of infection ultimately resulting in mortalities; prev. of infection >80%. The precise number of fish lost is not specified but experimental trials suggest a low level of loss (<1%). Loss is estimated on a single cage (3 × 3 × 3 m ³) of av. 2.5 g fish stocked at 25 kg m ³ (i.e. 270 000 fish) having an 80% survival through to harvest (min. harvest size of 15 kg fish ⁻¹) but with a 50% reduction (accounts for poor growth, feed inefficiency, specimen condition) on the 2012 market price, i.e. US\$ 9.86 kg ⁻¹	1.6 M	Shirakashi <i>et al.</i> (2012)
	Alternatively, assuming that the 50% of stock that developed scoliosis and fed poorly died, then loss is estimated on the assumption that 270 000 fish were transferred, each with a value of US\$ 2.00 fish ⁻¹ (extrapolated from Table 7)	270 000	Shirakashi <i>et al.</i> (2012)
<i>Microsporidium seriolae</i> ('Beko disease')	Disintegration of massive cysts within the trunk muscles causes a shrunken appearance, emaciation and mortality in farm stocks. Cysts appear 2 weeks post-transfer to sea cages, prevalence of infection rising to >90% 6–25 weeks post-transfer. Most fish with light infections recover; suggested that heavily infected juvenile should be disposed of as soon as possible. No details of loss or subsequent downgrading are provided	–	Sano <i>et al.</i> (1998); Yokoyama <i>et al.</i> (2011)
<i>Myxobolus acanthogobii</i> (syn. <i>M. buri</i>)	Infections cause severe scoliosis in up to 30% of 1+ age stock in farms in Shizuoka Prefecture, Japan in 1981. Cysts are frequently found in the cerebral cavities. Loss is based on the per ton assumption that up to 30% of stock may be lost, that fish in their first year are valued at 50% of the harvest price per ton of fish, and, harvest prices in Japan in 1984, i.e. US\$ 3953 t ⁻¹	up to 593 t ⁻¹	Egusa (1985); Yokoyama <i>et al.</i> (2005a, b)
<i>Myxobolus spirosulcatus</i>	Monitoring of net-caged stock (0+ aged fish) in Oita Prefecture, Japan between Aug 1999 and Feb 2000 found a peak of infection (i.e. 80% prevalence) in the bile in Oct 1999. No pathological changes in the host were observed but this parasite may have a relationship with the green livers observed in this host	–	Yokoyama and Fukuda (2001)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
	Heavy mortalities were recorded in 100 g to 1 kg stock in farms near Uwajima, Ehime Prefecture Japan. Fish were emaciated, had skin ulcerations, and at post-mortem were found to have a severe encephalomyelitis, nerve cell necrosis, blood congestion, encephalomeningitis, granulomatous peritonitis, and, a redness to the brain and spinal cord. Unidentified “exotic parasite-like bodies” were found in the seriously damaged spinal cords of 12 of the 65 (i.e. 18%) fish examined. These parasites are later suggested to be <i>M. spirosulcatus</i> (Yokoyama <i>et al.</i> 2010). Loss is est. on the basis of at least four, 20 t production units, an 18% loss of market sized fish as indicated by the post-mortem study (i.e. 14.4 t), and the value of harvest stock in Japan in 2007, i.e. US\$ 6.04 kg ⁻¹	86 976	Katagiri <i>et al.</i> (2007)
<i>Benedenia seriola</i>	Considered a serious problem since the advent of cage culture in Japan in the 1950s	–	Harada (1966); Egusa (1983)
	Infections result in severe skin damage, lesions and infections of up to 570 flukes fish ⁻¹ are reported. Although alludes to mortalities as a consequence of direct and secondary infection, fish with heavy burdens are emaciated and have poor weight gain. The av. weight of fish at 2 heavily infected sites, Kuki and Shiroura, Japan in Oct and Nov, 1963 were 650–750 g and 700–800 g, whilst the av. weight of stock at Lake Shiraishi, where infections were absent, were 800–900 g. No details relating to the total economic cost of this parasite on the Japanese industry are given but here an estimated loss of 10% is assumed. Figures are calculated on the size of the industry in 1963, i.e. 5038 t and an est. harvest price in 1963 of US\$ 2.00 kg ⁻¹ (no figures are available but 50% of the harvest price in 1984, i.e. US\$ 3.95 kg ⁻¹ , the first costs available, is applied)	1 M	Kubota and Takakuwa (1963)
	Parasite accounts for 22% of production costs, i.e. ~ US\$ 214 M (based on Japanese production of 153 075 t in 2001 worth ~ US\$ 973.9 M). This figure includes production of <i>S. quinquerediata</i> , <i>S. dumerili</i> and <i>S. lalandi</i>	214 M	Ernst <i>et al.</i> (2002); Whittington <i>et al.</i> (2001)
<i>Caligus</i> sp.	Heavy parasite burdens on the gills (10–240 lice fish ⁻¹ ; <i>n</i> = 5) and within the buccal cavities (2–54 lice fish ⁻¹) are reported on stock reared at a site in Kuki, Japan. Infections cause fish to rub against nets to the extent that the upper and lower jaw bones become exposed. No details of loss are given but this parasite causes the greatest damage to its host. Loss is estimated on a 10% loss or downgrade of stock at the Kuki site which is assumed to be a 50 t production site and an est. harvest price in 1963 of US\$ 2.00 kg ⁻¹ (no figures are available but 50% of the harvest price in 1984, i.e. US\$ 3.95 kg ⁻¹ , is used)	10 000	Kubota and Takakuwa (1963)
<i>Caligus spinosus</i>	Of the 30 000 fish that were ranched around Goto, Japan in April/May 1967, ~ 10 000 fish were subsequently lost to lice. Loss is estimated on the suggested av. size of fish as being 6 kg and the harvest price of fish in Japan in 1984 (no figures exist for 1967 and a back extrapolation 1984–1994 ascribes a negative cost for 1967), i.e. US\$ 3.95 kg ⁻¹	237 000	Fujita <i>et al.</i> (1968)
	Caligids usually infect the branchial arches. With heavy infections, fish become emaciated, anaemic and can develop rubbing-induced ulcerations around the mouth which are subject to secondary infection	–	Egusa (1983)
<i>Callotetrarhynchus nipponica</i>	Stock fed raw fish, notably anchovies, <i>Engraulis japonica</i> , became infected with larval cestodes (pleurocercoids) which reduced the marketability of the flesh. No details of infections provided	–	Nakajima and Egusa (1972); Ogawa (1996); Hutson <i>et al.</i> (2007)

	Trypanorhynch plerocercoid larvae in the abdominal cavity causes physical disturbance impacting on growth	–	Egusa (1983)
<i>Heteraxine</i> (syn. <i>Axine heterocerca</i>)	Infections result in anaemia and fish death. Study reports almost all the farm sites with only 2 or 3 exceptions within Mie Prefecture, Japan. All sites reported severe damage notably those at Hayada, Kuki and Shiroura. No details of loss are provided but here an estimate of 10% is assumed. Figures are calculated on the size of the Japanese industry in 1963, i.e. 5038 t and an estimated harvest price for 1963 of US\$ 2.00 kg ⁻¹ (no figures are available but 50% of the harvest price in 1984, i.e. US\$ 3.95 kg ⁻¹ , the first costs available, is applied)	1 M	Kubota and Takakuwa (1963)
Isopoda unidentified sp.	Heavy gill infections result in anaemia, emaciation and death	–	Egusa (1983)
<i>Philometroides seriolae</i>	Fish reared in Kuki Bay, Japan with infections on the gills and within the buccal cavity are reported to be emaciated. No details of loss are given	–	Kubota and Takakuwa (1963)
Greater amberjack, <i>Seriola dumerili</i> (Risso, 1810)	Nematodes bore through the trunk muscle or reside in the muscle reducing the commercial value of the fish. Although the overall price of the carcass may be downgraded, the affected lesion can be removed during processing	–	Egusa (1983)
<i>Cryptocaryon irritans</i>	100% mortality of broodstock held in land-based seawater tanks (6 or 10 m ³) at a stocking density of 8.8 kg m ³ . Loss is based on the assumption that 88 kg of fish were lost and the harvest price of Spanish greater amberjack in 1997 as no prices for Greece are listed, i.e. US\$ 9.48 kg ⁻¹	834	Rigos <i>et al.</i> (2001)
<i>Kudoa amamiensis</i>	93% of stock (1+ to 3+ aged fish; $n = 93/100$ fish) at one culture facility on the Spanish Mediterranean coast died 30 days after the first signs of infection. Loss is based on the price of harvest-sized fish in Spain in 2007, i.e. US\$ 10 kg ⁻¹ and an av. weight of 5 kg	4650	Montero <i>et al.</i> (2007)
	A survey of 10 farm sites between 1994 and 1998 indicates that between 30 and 90% of stock in the Motobu area (2 sites) of Okinawa Prefecture, Japan were infected, whereas elsewhere in the Prefecture infection rates are in the region of <1% (8 sites). Loss is estimated on the assumption that each site produces 20 t, that infected fish are not sold, and, on the price of Japanese amberjack in 1998, i.e. US\$ 7.36 kg ⁻¹ . Loss in the Motobu area is calculated to be 24.6 t (i.e. 61.5% of stock), whilst losses for the rest of Okinawa are 0.2 t (i.e. 0.125% of stock)	181 056 for the 2 Motobu sites 1472 for the remaining 8 Okinawa sites	Sugiyama <i>et al.</i> (1999)
<i>Microsporidium seriolae</i> ('Beko disease')	Stock transferred to sea cages in Miyazaki Prefecture, Japan in June 2006 were monitored over 2006–2009. Infections in a sub-sample of 20 fish were 100% (av. 16 cysts fish ⁻¹) in July–Aug 2006 and decreased thereafter but were still 80% by Sept 2007. Most fish recover, although it is suggested that heavily infected fish should be disposed of. No details of loss or fish disposed of are provided	–	Yokoyama <i>et al.</i> (2011)
<i>Benedenia seriolae</i>	Parasite accounts for 22% of production costs, a figure which includes the production of <i>S. quinqueradiata</i> , <i>S. dumerili</i> and <i>S. lalandi</i> *	214 M*	Ernst <i>et al.</i> (2002); Whittington <i>et al.</i> (2001)
<i>Benedenia seriolae</i> / <i>Zeuxapta seriolae</i>	Mixed infections on farmed <i>Seriola</i> sp. in Australia in 2003 were responsible for the subsequent loss of 39 t (i.e. ~15 000 fish) of stock	580 000	Kolkovski and Sakakura (2004)
<i>Neobenedenia melleni</i> (syn. <i>girellae</i>) ^b	A heavy fluke infection results in the 100% mortality of 0-year-old fish in Okinawa, Japan in July 1992. Inspection of 5 fish consignments entering the country from Hong Kong and Hainan, China which are shown to be infected with a prevalence of 7.7–70% are	192 000	Ogawa <i>et al.</i> (1995)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
	believed to be the source of infection in Japan. Loss is based on the assumption that a single $10 \times 10 \times 10 \text{ m}^3$ cage of juvenile fish ($\sim 50 \text{ g}$) stocked at 20 kg m^3 was lost; that 50 g juveniles are each valued at US\$ 4.80 each (see Table 7) assuming no change in prices of stock between 1992 (price of harvest fish = US\$ 6.96 kg^{-1}) and 2007 (price of harvest fish = US\$ 6.04 kg^{-1})		
	Economic losses due to <i>N. melleni</i> may now exceed those of <i>Benedenia seriolae</i> with <i>N. melleni</i> being more of a problem during the summer months	> 200 M	<i>Pers. comm.</i> Dr S. Shirakashi
<i>Paradeontacylix</i> -like trematodes	Approx. 80–85% of the Spanish wild-caught juveniles ($\sim 100 \text{ g}$) that were transferred to at least 3 sea-cage sites along the Tarragona coast and around the Balearic Islands in Sept–Oct began dying over Dec–May. Mortalities were attributed in part to epitheliocystis found in 30% of the Balearic stock and in 90% of those from Tarragona but also to heavy digenean fluke infections in 50% of the fish. Eggs were found in the heart and mortalities were attributed to the accumulation of eggs in the blood vessels of the gills. Loss is based on the assumption of an av. loss of 80% stock across all 3 sites, that each was a 20 t production unit, and, a US\$ 4.00 value assigned to each fish est. 250 g	768 000 across 3 Spanish sites	Crespo <i>et al.</i> (1994)
<i>Paradeontacylix grandispinus</i> / <i>Paradeontacylix kampachi</i>	Sudden outbreak of disease at several Japanese sites leading to a mass mortality of stock (cumulative 50 to >80% in a month; highest recorded daily mortality $\sim 15\%$) in juvenile 0+ fish in March–May 1993 several months after their importation from Hainan, China. Heavy eggs burdens within afferent gill arteries despite low number of parasites ($1\text{--}5 \text{ fish}^{-1}$) detected. Loss is estimated on at least 3 farms losing an av. 70% of stock, that each fish was valued at US\$ 4.00, and, that each site had 5000 fish as part of their starting stock	42 000	Ogawa and Fukudome (1994)
	Mass mortalities of stock imported from Hainan Island and Taiwan into Japan (12 stocks; 6 shipments; 2 from Hong Kong, 4 from Hainan) first seen in Dec 1983–Mar 1984	–	Ogawa and Egusa (1986); Ogawa and Fukudome (1994)
Sanguinicoliasis	The mass mortality of 0+ stock in sea-cage sites at Majorca, Catalonia and Murcia Spain has been occurring since 1989, typically between Dec and Mar each year. Losses were attributed to a combination of prokaryotic organism inducing epitheliocystis and heavy blood fluke burdens. Losses amounting to 3% of stock, due only to blood flukes, are also reported in 1+ aged stock. No stock or farm details are provided. Loss is based on the death of fish from the single infection, the assumption that cages ($5 \times 5 \times 5 \text{ m}^3$) were stocked at 20 kg m^3 and the harvest price of market sized fish in Spain in 1992 was US\$ 12.90 kg^{-1}	968 per cage	Crespo <i>et al.</i> (1992)
<i>Zeuxapta seriolae</i>	Mass mortalities ($0.8\text{--}1.5\% \text{ day}^{-1}$; 50% final) of 400–500 g stock in sea cages sited at Mallorca, Spain across the period Jan–Mar, 2002 were attributed to heavy infections causing anaemia and damage to the gills impairing respiratory function. No stock or farm details are provided. Loss is based on the assumption that cages ($5 \times 5 \times 5 \text{ m}^3$) were stocked at 20 kg m^3 . No data for Spain exists in FishStatJ for 2002, but using a linear scale of values between 1994 (US\$ 13 kg^{-1}) and 2010 (US\$ 19.28 kg^{-1}), the harvest price of market sized fish is est. at US\$ 16.12 kg^{-1}	20 150 per cage	Grau <i>et al.</i> (2003)
	Recurrent outbreaks of mortality ($33.3\text{--}47.2\%$; 686.7 ± 125.4 parasites fish^{-1}) in 0+ to 3+ aged fish reared in experimental culture tanks in Spain throughout 1998–2000. Loss is based on the 40 fish (av. wt. 558 g) lost during the mortality event and the estimated	212	Montero <i>et al.</i> (2004)

	harvest price of Spanish fish in 2000 of US\$ 9.50 kg ⁻¹ (only figures for 1997, i.e. US\$ 9.48 kg ⁻¹ , and 2007, i.e. US\$ 10.00 kg ⁻¹ are available)		
Kingfish, <i>Seriola lalandi</i> (Valenciennes, 1833)			
<i>Kudoa</i> (= <i>Pentacapsula neurophila</i>)	In Nov 2008, 55% of 1 kg stock in 1 cage (i.e. 13 750 out of 25 000) and 80% (i.e. 20 000 out of 25 000 fish) in an adjoining cage in western Australia were lost. Fish were found to have mixed infections: 76.6% of stock had <i>K. neurophila</i> in the brains, whilst 66.1% of stock had an infection of <i>Unicapsula seriolae</i> within their skeletal muscle. The loss estimate is that provided at the time of the outbreak (i.e. AUS\$ 300 000)	280 668	Stephens and Savage (2010)
	A second mortality event in the same site as above, resulted in the loss of 12% of starting stock (i.e. 6480 juveniles out of 54 000); 86.6% of these (i.e. 99/114 fish sampled) had infections in the brain. Loss is estimated on the cost of juveniles given in Table 7, i.e. US\$ 4.80 fish ⁻¹	31 100	Stephens and Savage (2010)
<i>Unicapsula seriolae</i>	Study suggests that fish on the open market are priced at more than US\$ 1.50 kg ⁻¹ (assuming AUS\$ 1=US\$ 1) but infected fish sold through Brisbane markets are lower priced because the flesh disintegrates on cooking and releases an unpleasant odour	–	Lester (1982)
	The loss of 33 750 1 kg fish in Nov 2008 was attributed to an infection of <i>U. seriolae</i> in the skeletal muscle of 66.1% of stock and an infection of <i>Kudoa neurophila</i> in 76.6% of stock. Losses were recorded at AUS\$ 300 000	280 668	Stephens and Savage (2010)
	A second infection in juvenile stock at the same site in western Australia in Nov 2008 resulted in the loss of 6480 fish. A sample of 114 fish revealed that 7 of these had skeletal muscle infections. Loss is estimated on estimated on the price of juvenile fish (see Table 7)	31 100	Stephens and Savage (2010)
<i>Benedenia seriolae</i>	Parasite accounts for 22% of production costs – a figure which includes the production of <i>S. quinquerradiata</i> , <i>S. dumerili</i> and <i>S. lalandi</i> *	214 M*	Ernst <i>et al.</i> (2002); Whittington <i>et al.</i> (2001)
	Complications associated with a hydrogen peroxide bath administered in 4 cages for the treatment of parasitic infections in Australia in 2010 resulted in the death of 80 t of fish. No figures for Australian kingfish are listed within FAO FishStatJ and so the market price of Japanese amberjack in 2010 is used as an estimator, i.e. US\$ 9.66 kg ⁻¹ . In addition to the loss of fish, the incident also wiped US\$ 3.71 M (AUS\$ 4 M) off the farm's market value (loss of share price)	772 800 (fish losses) 3.71 M	McIlwraith (2010)
<i>Caligus chiestos</i>	Lice burdens do not pose a significant threat to the health of captive reared stock, their numbers controlled by the administration of hydrogen peroxide treatments to control monogenean infections (treatment costs approx. US\$ 4–5 K cage ⁻¹ including labour)	4500	Hayward <i>et al.</i> (2007); Nowak <i>et al.</i> (2011)
<i>Paradeontacylix</i> -like spp.	Infections in juvenile stock in New Zealand are reported from the heart, brain and internal organs resulting in granulomatous pathology in the heart and gills. Infections are associated with low-level mortalities. No details provided in the original report	–	Diggles and Hutson (2005)
<i>Zeuxapta seriolae</i>	Mortality (12.8%) of 223.5 ± 21.7 g experimentally infected fish (<i>n</i> = 91) of Australian sea cage stock between weeks 4 and 8	–	Mansell <i>et al.</i> (2005)
Longfin amberjack, <i>Seriola rivoliana</i> (Valenciennes, 1833)			
<i>Neobenedenia melleni</i> (syn. <i>girellae</i>) ^b	Heavy infection of 1-year-old stock reared in cages Ishigaki Island, Japan. No details on the losses are provided	–	Ogawa <i>et al.</i> (1995)
(e) Gilthead sea bream, <i>Sparus aurata</i> L.			
<i>Amyloodinium ocellatum</i>	75% mortality (i.e. 30 000) of 10 g juveniles over 48 h in a sea cage in Greece in the summer of 1997; water temperatures 23 to 26 °C. Loss is estimated on US\$ 0.10 fish ⁻¹	3000	Rigos <i>et al.</i> (1998)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
	An infection in stock (av. 481 ± 92.9 g) held in 2500 m^3 earthen ponds at 0.75 kg m^3 with an equal weight of meagre resulted in a 29% mortality in 2 days (i.e. $271.88 \text{ kg pond}^{-1}$). Loss is estimated on the assumption that a minimum of 2 ponds were affected and on the market price of Portuguese fish in 2011, i.e. US\$ 7.67 kg^{-1}	4170	Soares <i>et al.</i> (2012)
<i>Brooklynella hostilis</i>	An infection is reported from the gills of moribund stock (av. 150–200 g) held in a floating net-pen in the Gulf of Eilat. Details on the prevalence of infection and mortality are not provided but the fish were co-infected with an enteric bacterium, <i>Enteromyxum leei</i> and <i>Furnestinia echeensis</i>	–	Diamant (1998b)
<i>Ceratomyxa sparusaurati</i>	Prevalence of infection across 3 sites on the western Atlantic coast of Spain ranged from 0 to 59.9%, whilst that at a single site on the southern Atlantic coast of Spain was 3%. Infection resulted in trickling mortalities	–	Palenzuela <i>et al.</i> (1997)
	Accounted for low levels of loss ($0.5\% \text{ day}^{-1}$) in Greece	–	Rigos <i>et al.</i> (1999)
	Infections rates of between 6.66–20% were found at 3 separate cage sites within the Adriatic	–	Mladineo (2003)
<i>Cryptocaryon irritans</i>	56–100% mortality of broodstock held in land-based seawater tanks stocked at $6\text{--}10.2 \text{ kg m}^3$. Loss is based on the price of harvest-sized fish in Greece in 2000, i.e. US\$ 4 kg^{-1}	$2240\text{--}4000 \text{ t}^{-1}$	Rigos <i>et al.</i> (2001)
<i>Cryptosporidium molnari</i>	Between 1998 and 2000, a total of 346 sea bream sampled from various culture sites along the Atlantic, Cantabric and Mediterranean coasts of Spain, as part of a larger survey of mortality and morbidity events found a 24.4% prevalence of infection within the stomach epithelium. Infected hosts had abdominal swelling, infected zones of epithelium were necrotic which subsequently sloughed and detached from the lumen. Infection is also linked to an early mortality event of fingerlings	–	Sitjà-Bobadilla and Alvarez-Pellitero (2001); Alvarez-Pellitero and Sitjà-Bobadilla (2002)
<i>Enteromyxum leei</i>	Chronic mortalities of $5\text{--}10 \text{ fish day}^{-1}$ in stock tanks holding 5000 fish (av. 150 g) are reported to have occurred Dec 1991–Jan 1992 in a culture facility in southern Cyprus. Loss is est. on a 7 fish day^{-1} mortality rate, that the event lasted 60 days, and, the price of Cypriot fish in 1992, i.e. US\$ 14.44 kg^{-1}	910	Diamant (1992)
	29% loss of stock observed over numerous Greek cage sites covering several geographical regions during the period Sept 1996–Sept 1997. Estimate of loss is based on the harvest price of Greek fish in 1997, i.e. US\$ 6.33 kg^{-1}	1836 t^{-1}	Rigos <i>et al.</i> (1999)
<i>Henneguya</i> sp.	An infection within the hearts of stock (av. 260 g) reared in seawater, earthen ponds resulted in a low-level of mortality (i.e. $4\text{--}5 \text{ fish day}^{-1}$)	–	Caffara <i>et al.</i> (2003)
<i>Ichthyophonus</i> sp.	Two farm sites in southern Greece had reported mortalities of 25% (farm A) and 15% (farm B). Sampling throughout 1981–1987 found that 15% of the fish from farm A ($n = 40$; stocked at 15 kg m^3 and fed a mixed diet of pellets and trash fish) and 23.5% of those from farm B ($n = 51$; stocked at 30 kg m^3 and fed a pellet diet only) were infected. Infection was a common granulomatous response in the liver, heart, kidneys and intestines. Loss is based on the assumption that each site produced 10 t, the size of fish lost	$53\,160 \text{ p.a. across the 2 sites}$	Athanassopoulou (1992)

	<p>were harvest size, i.e. 0.5 kg, the total mortality at each site can be attributed to <i>Ichthyophonus</i>, and, the harvest price of Greek fish in 1987, i.e. US\$ 13.29 kg⁻¹</p> <p>Three Spanish culture sites were sampled throughout 1990–1991. A prevalence of 75% was found in stock (3.9–27.9 cm long) in a closed system near Valencia ($n = 185$), 20.8% of hatchery and nursery stock (2–15 cm long) from a site near Cadiz ($n = 49$), and, in 70% of wild stock (26–31 cm long) caught from the wild and then reared on at a site near the River Ebro delta ($n = 10$). No mortalities were reported but the presence of this parasite in stock is considered a threat</p>	–	Franco-Sierra <i>et al.</i> (1997)
<i>Kudoa</i> sp.	High mortalities associated with infection reported. No details available	–	Arfara <i>et al.</i> (1995 cited in Rigos <i>et al.</i> 1999)
Microsporidian: <i>Pleistophora</i> -like	Constant low-level mortalities of stock held in floating sea cages in Malta, i.e. 0.06% week ⁻¹ in fish ~ 10 g 6–8 weeks post-transfer in Jan–Feb 1994 rising to 0.2% week ⁻¹ in 1995–1996. In larger fish, infection appears as large white to yellow nodules within the musculature, occasionally granulomatous decreasing marketability of fish. Loss is est. on a single 5 × 5 × 5 m ³ cage of fish stocked at 15 kg m ³ (i.e. 187, 500 10 g fish), a linear incremental increase in mortality from 0.06 to 0.2% week ⁻¹ across a 12-month culture period, a linear 35 g weight gain per month, and, the harvest price of Maltese fish in 1996, i.e. US\$ 6.24 kg ⁻¹ . Final estimated losses were 11 840 fish representing a final mortality of approx. 6.3% and 2860 kg of fish	17 846	Abela <i>et al.</i> (1996)
<i>Myxidium</i> sp.	Occasional mortalities were attributed to <i>Myxidium</i> infections	–	Diamant (1992)
<i>Pleistophora</i> sp.	Muscular infection of av. 75 g stock in Greece in 4 pens of fish resulted in a daily mortality of 0.2% for a period of ~ 7 days in Feb–Mar 1996. Infected fish were lethargic, emaciated, had discoloured skin and had scoliosis. Mortalities were arrested following a fumagillin treatment. Four cages (assuming 5 × 5 × 5 m ³) were stocked at 15 kg m ³ , loss equates to ~ 410 fish cage ⁻¹ (i.e. 30.75 kg) or ~ US\$ 270 cage ⁻¹ (based on US\$ 8.767 kg ⁻¹ harvest price in 1996 for Greek sea bream). Total loss est. at ~ US\$ 1080	1080	Athanassopoulou (1998)
<i>Polysporoplasma sparis</i>	Infection rates of between 7.14 and 100% were recorded in ~ 256 g stock held at 4 different cage sites within the Adriatic Sea	–	Mladineo (2003)
<i>Sphaerospora</i> sp.	Prevalence of infection can reach 80% and cause serious kidney damage, including glomerular destruction and haemorrhagia	–	Sitjà-Bobadilla and Alvarez-Pellitero (1992)
<i>Trichodina</i> sp.	An infection of newly stocked fingerlings (1 fish L ⁻¹ ?) from Bardawil Lagoon, Israel in a 1000 L (?) tank in 1976(?) at a research facility in Eilat, Israel resulted in morbidity and mortality of up to 40–50%	–	Paperna <i>et al.</i> (1977)
<i>Ceratothoa oestroides</i>	Infection is reported to have caused high losses in juvenile populations reared in Croatia as a consequence of gill damage and respiratory failure. In older fish, infection results in a 20% reduction in growth. Assuming the 2012 market price of Croatian fish, i.e. US\$ 6.50 kg ⁻¹ infection represents a loss of US\$ 1.30 kg ⁻¹	1.30 kg ⁻¹	Šarušić (1999)
<i>Ceratothoa parallela</i>	Cymothoid isopods were found in the branchial and buccal cavity of 2–2.3 g juveniles when being stocked into cages in Greece. The cumulative mortality over the subsequent 2 months was >50% (i.e. the loss of >120 000 fish; ~ 258 kg). Loss is based on the value of Greek harvest-sized stock in 2001, i.e. US\$ 3.88 kg ⁻¹	1000+	Papapanagiotou and Trilles (2001)
<i>Furnestinia echeensis</i>	The study suggests that infections, of up to 239 flukes fish ⁻¹ , on hatchery produced stock (80–130 mm) and on fingerlings from Bardawil Lagoon reared in single 1000 L (?) tanks at a research facility in Eilat, Israel in 1976(?) resulted in the morbidity and mortality of 40–	13.5	Paperna <i>et al.</i> (1977)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
<i>Gyrodactylus oreochiae</i>	50% of stock. Although some details are provided, specifics relating to the mortality event are not clear. It is assumed that fish were stocked at 1 fish L ⁻¹ . Loss assumes the mortality of 450 (10 g) fish in 2 tanks (i.e. total loss of 9 kg stock) and is based on the harvest price of Israeli freshwater tilapia in 1984 (first figures available), i.e. US\$ 1.50 kg ⁻¹	1406	Paladini <i>et al.</i> (2009)
	Approx. 2–10% mortality in juvenile stock (5–10 g) held in inshore-floating cages in Albania and Croatia; infection of gills and all external surfaces with 1000+ gyrodactylids fish ⁻¹ . Fish were hypermelanotic, lethargic, anorexic with a progressive loss of weight. Loss is based on the assumption that at least 3 cages measuring 5 × 5 × 5 m ³ , each stocked at 15 kg m ³ were affected (i.e. 187 500 fish cage ⁻¹), av. mortality was 5% (i.e. 9375 fish cage ⁻¹), and, a price of US\$ 0.05 juvenile ⁻¹ (see Table 7)		
<i>Lernanthropus kroyeri</i>	Infection reported from farms in Bosnia-Herzegovina and Italy		Paladini <i>et al.</i> (2011)
	Infection at a single 2000 m ³ , 1500 t cage production site in Greece producing <i>D. labrax</i> and <i>S. aurata</i> . Cages are stocked at 12–15 kg m ³ (50–300 g sized fish). Infection accounts for <1% loss over an 8-month period in 200(1?). Losses for each species are not given but assuming an equal production of both species, then mortality is <7 t. Loss figures are based on the value of Greek stock in 2001, i.e. US\$ 3.88 kg ⁻¹	<27 160	Vagianou <i>et al.</i> (2006)
<i>Microcotyle</i> sp.	Anorexia, lethargy, anaemic gills and increased mortalities attributed to a <i>Microcotyle</i> infection reported in 200–300 g stock reared in sea cages in southern Spain in Feb 1992	–	Sanz (1992)
<i>Microcotyle</i> sp. & <i>Lamellodiscus</i> sp.	Continuous low mortalities in 0+ sea bream in floating cages in NE Spain with mild to severe parasite burdens and proliferative epitheliocystis during the winter of 1993–1994	–	Padrós and Crespo (1995)
Unidentified sanguinicolid	An infection (prev. 82.6%) in net caged stocked (50–400 g body weight) in north-eastern Spain throughout Dec 1998–April 1999 resulted in a low level, chronic pattern of mortality (0.01–0.1% daily). The prevalence of infection through the cold season the following year was found to be 100%. Stocks also were infected with the gill monogeneans <i>Furnestinia echeneis</i> and <i>Microcotyle chrysophrii</i> (prev. of 35% in 1999 and 20% in 2000). Loss is based on the assumption the site consisted of four 5 × 5 × 5 m ³ cages stocked with 50 g fish at 15 kg m ³ (i.e. 180 000 fish as a 9 t production site). Assuming that the mortality event occurred over a 120-day period (i.e. mid-Dec to mid-April), then using the pattern of losses provided a total of 11 880 fish were lost or 6.6% of starting fish stock. The figure of loss is based on an av. fish wt. of 100 at the time of mortality and the harvest price of Spanish fish in 1999, i.e. US\$ 6.00 kg ⁻¹	7128	Padrós <i>et al.</i> (2001)
<i>Sparicotyle chrysophrii</i>	Infections cause anaemia and high mortality at low intensities (8–10 parasites gill arch ⁻¹) in fish weighing 10–300 g. No details provided	–	Paladini <i>et al.</i> (2011)
(f) Coho salmon, <i>Oncorhynchus kisutch</i> (Walbaum, 1792)			
<i>Kudoa thyrssites</i>	A 1.0–11.2% infection within the cardiac muscle of fresh morts collected from 5 different hatcheries along the coast of British Columbia, Canada was found between Aug 1986 and Jan 1988	–	Kabata and Whitaker (1989)

	Processors report that a 60% prevalence of myoliquefaction in smoked fillets prepared from 1 net-pen of stock. Assuming that all infected fillets were rejected, loss is based on the value of harvest-sized fish in Canada in 1992, i.e. US\$ 4.44 kg ⁻¹	2664 t ⁻¹ of production or 4440 t ⁻¹ of rejected fish	Whitaker and Kent (1992)
<i>Loma salmonae</i>	<i>Loma</i> combined with a bloom of <i>Corethron</i> -like diatoms led to severe inflammatory lesions associated with ruptured xenomas leading to 60% mortality in 100 000 net-pen held smolts (~ 40 g) over a 7-day period in British Columbia in Oct 1987. Estimate of loss is based on price of harvest-sized fish in Canada in 1987, i.e. US\$ 4.77 kg ⁻¹	11 448	Speare <i>et al.</i> (1989)
	Significant gill pathology seen in smolts transferred to net-pens in Washington State; parasite prevalent on 33–65% of gills leading to severe lesions and mortalities throughout the summer	–	Kent <i>et al.</i> (1989)
<i>Paramoeba pemaquidensis</i>	25% mortality in stock (no size details) reared net-pens in Washington in 1985. Estimate of loss is based on the value of harvest-sized fish in the USA in 1985, i.e. US\$ 3.50 kg ⁻¹	875 t ⁻¹	Kent <i>et al.</i> (1988)
<i>Parvicapsula</i> sp.	Kidney infections resulted in mortalities in juvenile fish held in net-pens in Washington State, USA in 1979	–	Hoffman (1984)
	A 30% mortality observed in marine stock in Oregon State. No details available	–	Johnstone (1984)
<i>Lepeophtheirus salmonis</i>	Lice burdens on 150 g juveniles stocked in sea cages sited in Shizugawa Bay on the Pacific coast of north-eastern Honshu, Japan in Oct 1991 were followed until they were harvested in Aug 1992. Lice burdens peaked in late July to early Aug 1992 at 2.6 ± 2.4 lice fish ⁻¹ (prev. 84.6%). Peri-anal haemorrhages were observed but not thought to cause serious disease	–	Urawa <i>et al.</i> (1998a)
	10 000 juvenile fish (~ 1-year old; 420–430 g) were transferred to net-pens in Onmae Bay, Miyagi Prefecture, Japan. Lice infections determined on stock over the period March–July 1993 were low at an av. 3.5 lice fish ⁻¹ (prev. 73.3%)	–	Ho and Nagasawa (2001)
(g) European seabass, <i>Dicentrarchus labrax</i> L.			
<i>Amyloodinium ocellatum</i>	An infection at 2 Italian farms in July 1979 resulted in the loss of at least 600 (av. wt. 80.6 g, 18.5 cm long) fish from 1 tank at 1 site. No specific details relating to the mortalities at the second farm are provided. Loss is estimated on the assumption that at least 1200 fish in total were lost and the price of harvest-sized Italian fish in 1984, i.e. US\$ 10.51 kg ⁻¹ (date at which first figures are available)	1016	Ghittino <i>et al.</i> (1980)
<i>Ceratomyxa</i> spp. (<i>C. diplodae</i> & <i>C. labracis</i>)	The examination of 340+ fish found <i>C. diplodae</i> in 10.2% of <1-year-old fish and 3.33–6.97% in 1+ to >4-year-old fish and <i>C. labracis</i> in 46.93% of <1-year-old stock and 21.05–71.05% in 1+ to >4-year-old fish. Variable degrees of damage to the gall bladders were seen	–	Alvarez-Pellitero and Sitjà-Bobadilla (1993)
<i>Cryptosporidium molnari</i>	Between 1998 and 2000, a total of 151 fish were sampled from various culture sites along the Atlantic, Cantabric and Mediterranean coasts of Spain, as part of a larger survey of mortality and morbidity events found a 4.64% prevalence of infection within the stomach epithelium. Infection associated losses were suggested.	–	Alvarez-Pellitero and Sitjà-Bobadilla (2002)
<i>Enteromyxum lei</i>	42% loss of stock observed over numerous Greek cage sites covering several geographical regions during the period Sept 1996–Sept 1997	–	Rigos <i>et al.</i> (1999)
<i>Ichthyophonus</i> sp.	Infection, with a prevalence of 24.4%, was found in stock (size not specified; <i>n</i> = 127) from a Spanish farm near the River Ebro delta, Spain. Infections, notably in the heart and muscle, appeared as granulomas with an intense fibrotic reaction. Severe tissue damage was noted in the trunk kidney. No mortality or morbidity of stock was commented upon	–	Sitjà-Bobadilla and Alvarez-Pellitero (1990)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
	Three Spanish culture sites were sampled throughout 1988–1991. Infections were found with prevalences of 90% in stock (2.4–29.6 cm long) at a grow-out facility near Valencia ($n = 82$); 78.8% of 2.3–52.3 cm long stock caught from the wild and then reared on at a site on the western Spanish Mediterranean coast ($n = 33$), and, in 70% of 24–34 cm sized stock ranched from the wild and then grown on in sea cages near the River Ebro delta ($n = 10$). No losses were reported but the presence of this parasite in stock is considered a threat	–	Franco-Sierra <i>et al.</i> (1997)
<i>Loma</i> spp.	Intestinal infections of a microsporidian resulted in the emaciation, poor growth rate and low mortality of stock at 3 Italian inland marine farm sites. No details provided	–	Caffara <i>et al.</i> (2010)
<i>Oodinium</i> sp.	[see entry for <i>Amyloodinium ocellatum</i>]	1016	Ghittino <i>et al.</i> (1980)
<i>Philasterides dicentrarchi</i>	A low level of mortality (i.e. ~ 5 morts day ⁻¹) was seen in 250 g stock reared in tanks at a French farm site within a Mediterranean laguna in March 1993. Moribund fish had swollen urogenito-anal papillae and congestion of the internal body organs	–	Dragesco <i>et al.</i> (1995)
<i>Sphaerospora dicentrarchi</i>	Infection rates of between 7.14 and 100% were found at 5 separate cage sites within the Adriatic	–	Mladineo (2003)
<i>Sphaerospora testicularis</i>	Infections caused significant damage to the testes with significant losses (5–10% day ⁻¹) at certain Greek sites during the period Sept 1996–Sept 1997	–	Rigos <i>et al.</i> (1999)
<i>Anilocra physodes</i>	A suggested serious parasite problem of juvenile, caged reared stock. No details available	–	Athanassopoulou <i>et al.</i> (2001a)
<i>Caligus minimus</i>	30 g–2 kg Italian fish infected with ~ 40 copepods fish ⁻¹ were anorexic and lethargic; infection resulted in $\sim 9\%$ mortality. No details on which size of fish were lost	–	Pavoletti <i>et al.</i> (1999) cited in Johnson <i>et al.</i> (2004)
<i>Ceratothoa oestroides</i>	Infection is reported to cause high losses in juvenile populations reared in Croatia as a consequence of gill damage and respiratory failure. In older fish, infection results in a 20% reduction in growth. Assuming the 2012 market price of Croatian fish, i.e. US\$ 6.74 kg ⁻¹ , infection represents a loss of US\$ 1.35 kg ⁻¹	1.35 kg ⁻¹	Šarušić (1999)
	A survey of 109 cages at a Turkish site in the Aegean Sea in July 2000 found a $23.11 \pm 16.71\%$ prevalence of infection. A later survey of 53 cages in Sept 2000 found $13.51 \pm 17.51\%$ prevalence. The study found that there was a 20.1% (i.e. 14 g) difference in the weight of infected ~ 290 -day-old fish (i.e. >2 parasites fish ⁻¹) than in uninfected or those with low parasite burdens (i.e. <2 parasites fish ⁻¹). This translates into a 27–46.2 kg t ⁻¹ difference in production at this stage. Figures of loss are based on the price of Turkish harvest-sized fish in 2000, i.e. US\$ 4.13 kg ⁻¹	111–191 t ⁻¹ by day 290 of production	Horton and Okamura (2001)
<i>Diplectanum aequans</i>	Approx. 5–10% of Italian juvenile fish are lost each spring. Loss is estimated on harvest-sized fish being 0.5 kg, that production in 2006 was 8835 t, and, that the price of each juvenile was US\$ 0.5, i.e. between 1 850 000 and 3 700 000 fish are lost	0.925–1.85 M	Dezfuli <i>et al.</i> (2007)
<i>Emetha audouini</i>	Infection of a cage of 60 000 juvenile (~ 30 g, 3 months old) in Nov, 1997 in the northern part of Greece resulted in the loss of 10.75% of stock over a 2-week period. Isopods caused extensive, deep skin damage to the cranium and eyes. Adjoining cages of sea bream were unaffected. Loss is estimated on the value of each juvenile, i.e. US\$ 0.70 fish ⁻¹ .	4515	Papapanagiotou <i>et al.</i> (1999)

<i>Lernanthropus kroyeri</i>	Infection is reported to be responsible for the mortality of small-sized fish (<10 g) by asphyxia and anaemia in Greek culture systems. Loss is est. on a 1% mortality rate in stock reared in 5 × 5 × 5 m ³ cages, each stocked at 40 kg m ³ were affected (i.e. 500 000 fish cage ⁻¹), and, an est. price of US\$ 0.40 juvenile ⁻¹ (see Table 7)	2000	Athanassopoulou <i>et al.</i> (2001b)
	Infection at a single, mixed species (<i>D. labrax</i> and <i>S. aurata</i>), 2000 m ³ , 1500 t cage production site in Greece stocked at 12–15 kg m ³ (50–300 g sized fish) accounts for a <1% loss over an 8-month period in 200(1?). Losses for each species are not given but assuming a 50:50 production of both species, then mortality is <7 t. Loss figures are based on the value of Greek stock in 2001, i.e. US\$ 4.55 kg ⁻¹	< 31 850	Vagianou <i>et al.</i> (2006)
<i>Nerocila orbignyi</i>	Mortality of cage-reared stock in Corsica, France rises from 7 to 18% over the period July–Dec 1981; parasite prevalence is ~90%. Wild mullet are the source of the haematophagous isopods infection establishing in the cages. No details relating to the system or the size of the fish are provided but are assumed to be av. 50 g stocked in a single 125 m ³ cages at a stocking density of 40 kg m ³ (i.e. 100 000 fish cage ⁻¹). Loss is est. on a final mortality of 18% of starting stock (i.e. 18 000 fish) but assumes a linear 20 g per month weight gain, and, the price of harvest-sized French fish in 1985 (first figures available), i.e. US\$ 11.23 kg ⁻¹ . Final weight of fish lost is 1450 kg	16 284	Bragoni <i>et al.</i> (1984)
(h) Turbot, <i>Scophthalmus maximus</i> L.			
<i>Amyloodinium ocellatum</i>	The mortality of 3 tanks of 27 g fish (<i>n</i> = 27) within a research facility in Portugal is reported	–	Ramos and Oliveira (2001)
<i>Enteromyxum</i> -like sp.	High levels of mortality, reaching 100% in all infected land-based systems, were seen at a commercial culture site in northern Galicia, Spain during the spring and summer of 1997. A range of stock sizes from 50 g to 2 kg were lost from the systems, each of which typically measured 160 m ² and were stocked at 25 kg m ³ (i.e. 4000 kg system ⁻¹). Figures of loss are based on Spanish prices for harvest-sized fish in 1998, i.e. US\$ 9.1 kg ⁻¹	36 400 per system	Branson <i>et al.</i> (1999)
<i>Ichthyophonus</i> sp.	Three Spanish culture sites along the Cantabric coast were sampled throughout 1989–1991. The study found prevalences of 36.8% in nursery stock (2.9–5.2 cm long) at the first site (<i>n</i> = 95), 21% in hatchery and nursery fish (1.7–7.8 cm long) at a second site (<i>n</i> = 181), and, 42.4% in nursery, hatchery and grow-out stock at a third, undisclosed site (<i>n</i> = 118). No losses were reported but the presence of this parasite in stock is considered a threat	–	Franco-Sierra <i>et al.</i> (1997)
<i>Paramoeba</i> sp.	Amoebae caused severe gill damage resulting in the loss of between 5 and 20% of 500 g stock in 20 out of 150 tanks at a culture facility in north-west Spain over a 3-month period (Oct–Dec 1996). No tank sizes were given but loss is based on the assumption that each tank measured 160 m ² , stocked at 25 kg m ³ (i.e. 4000 kg system ⁻¹), an av. mortality of 10%, and, the price of Spanish harvest-sized fish in 1996, i.e. US\$ 9.78 kg ⁻¹	78 240	Dyková <i>et al.</i> (1998)
<i>Philasterides dicentrarchi</i>	During the summer of 1999 and the spring of 2000, 2 outbreaks of infection resulted in mortalities. The first event occurred in 2 tanks of >500 g fish, the second in 6 tanks of fish ranging from 150 to 1500 g, with 20–30 morts day ⁻¹ , rising to 100–150 fish day ⁻¹ . The infection resulted in the total loss of stock in some tanks. Assuming a 25 kg m ³ stocking density for this species, 4 m ³ culture tanks and a 50% loss across all infected tanks, then the loss of stock would have been ~400 kg. Figures are based on the price of Spanish harvest-sized fish in 2000, i.e. US\$ 8.50 kg ⁻¹	3400	Iglesias <i>et al.</i> (2001)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
<i>Philasterides</i> -like	Several outbreaks at a grow-out facility (av. fish wt. 300 g) in the north of Portugal were reported throughout March 2004–Feb 2005. Mortality reached 3–6% over the period May–Aug 2004. No specific details of loss are provided but it is assumed that the loss occurred in a single 160 m ² tank stocked at 25 kg m ³ (i.e. 4000 kg system ⁻¹). A normal distribution pattern of loss is assumed resulting in the loss resulting in a 21.3% loss of stock (i.e. 2875 fish with a total weight of 862.5 kg). The harvest price of Portuguese stock in 2005, i.e. US\$ 8.75 kg ⁻¹ is applied	7547	Ramos <i>et al.</i> (2007)
<i>Tetramicra brevifilum</i>	A chronic, low-level pattern of mortality in juvenile stock (~30 g) held in 4 m ³ concrete tanks in Galicia, Spain, resulted in the loss of ~3450 fish (11.5% of stock) over ~100 days, i.e. ~103.5 kg of fish. Infection was evident as small xenomas in a range of host tissues, where aggregations of spores occasionally elicited an inflammatory reaction. Figures of loss are based on the harvest price of Spanish turbot in 1990, i.e. US\$ 15.59 kg ⁻¹	1614	Figueras <i>et al.</i> (1992)
<i>Tetramicra brevifilum</i> + a histophagous ciliate	A 22.5% (i.e. ~675 fish) mortality of stock occurred in a single-holding unit within a grow-out facility holding ~3000 juveniles in north-west Galicia, Spain. The histophagous ciliates were tentatively identified as <i>Miamiensis avidus</i> and <i>Uronema marinum</i> . No details relating to the size of the fish are provided but av. wt. is assumed to be 50 g. Loss is estimated on the value of harvest-sized Spanish fish in 1993, i.e. US\$ 8.39 kg ⁻¹	283	Dyková and Figueras (1994)
Unidentified protistan	Seven out of 8 batches of eggs were infected with a large multinucleated protistan (1–2 per embryo; prev. 20.0–21.5%) in the embryos and yolk-sac larvae, which subsequently breaks up into numerous mononucleated organisms. No associated mortality was recorded in this study, unlike the infections observed in wild cod eggs (Pedersen <i>et al.</i> 1993)	–	Pedersen (1993)
<i>Uronema</i> -like ciliates	Between 1989 and 1996, a farm in Vest-Agder County, southern Norway had 3 outbreaks in their fry units (av. <0.3 g; 18 °C) in the autumn of each year resulting in the near complete loss on each occasion of stock. It is assumed that 100% of stock, valued at US\$ 0.10 fish ⁻¹ , in a single 160 m ³ tank stocked at 10 kg m ³ was lost	533 000	Sterud <i>et al.</i> (2000)
	The same farm also lost 30% of 500–1000 g stock held in on-grower units (15 °C) in Aug, 1998. Loss is est. on the assumption that an av. 30% of stock (av. 750 g) in three 160 m ³ tanks was lost. No prices for Norwegian turbot are available in FishStatJ and so the value of French turbot in 1998, i.e. US\$ 8.00 kg ⁻¹ , is used	11 520	Sterud <i>et al.</i> (2000)
(i) Silver sea bream, <i>Pagrus auratus</i> (Forster, 1801)			
<i>Anoplodiscus cirruspiralis</i>	Infections are reported to affect the fins and nasal lamellae of Australian stock. Approx. 10% of experimental stock died when held at higher stocking densities (av. 26.0–43.3 parasites fish ⁻¹) despite having similar parasite burdens as those held at lower stocking densities (av. 33.8–36.5 parasites fish ⁻¹)	–	West and Roubal (1998)
<i>Anoplodiscus tai</i>	Infections were reported at several cultures sites within Fukui and Nagasaki Prefecture, Japan throughout 1989–1993. Heavily infected fish were emaciated and monogenean activity on the fins had led to tissue damage and the partial loss of fins. The study reports that 5-year-old fish with heavy infections were rejected at market. Examination of stock found an av. 41.2 monogeneans per fish (range 26–64 flukes fish ⁻¹). No details relating to mortality or the economic loss associated with infections were given	–	Ogawa (1994)

(j) Groupers (general)			
Fish diseases (general)	Over 80% of farmers remarked that disease, including parasites, in small scale cage culture in Thailand (i.e. 1–5 cages) results in the 30–50% loss of stock. Loss is est. on fish being grown in 5 × 5 × 5 m ³ cages at a stocking density of 15 kg m ³ , and, the value of each 150 g fish is US\$ 2.00 (see Table 7)	7500–12 500 per cage	Bondad-Reantaso <i>et al.</i> (2001); Kanchanakhan <i>et al.</i> (2001)
Parasite (general)	This review suggested that the following parasite species cause problems in grouper culture throughout South-east Asia: <i>Amyloodinium</i> sp., <i>Brooklynella</i> sp., <i>Cryptocaryon irritans</i> , <i>Ichthyophonus</i> sp., <i>Trichodina</i> sp., <i>Benedenia</i> sp., <i>Megalocotyloides epinepheli</i> and <i>Pseudorhabdosynochus epinepheli</i>	–	Arthur and Ogawa (1996); Bondad-Reantaso <i>et al.</i> (2001)
<i>Benedenia</i> sp.	Reported to infect stock of <i>Epinephelus</i> spp. reared in sea cages off Pingtung, Taiwan. Flukes caused haemorrhages which were subject to secondary bacterial infection. No details of fish losses were provided	–	Chang and Wang (2000)
<i>Caligus lalandei</i>	Stocks of <i>Epinephelus</i> spp. reared in marine cages in the Pingtung area of Taiwan were commonly found infected. Lice activity resulted in lesions, inflammation and ulceration by secondary bacterial infections. No details of loss or downgrading provided	–	Chang and Wang (2000)
Hong Kong grouper or red spotted grouper, <i>Epinephelus akaara</i> (Temminck et Schlegel, 1842)			
<i>Pseudorhabdosynochus epinephali</i>	Hatcheries in the southwest Japan produce seed for stock enhancement and aquaculture Broodstock (0.5–1.0 kg) within a hatchery facility in Kagawa Prefecture reported gill infections as being common place and requiring treatment as heavy infections resulted in lower egg production. Moderate mortalities in juvenile stock were reported as a consequence. Loss is estimated on the basis that ‘moderate’ infers a 5–10% mortality and uses the values provided in Table 7 for juvenile fish, i.e. US\$ 1.02–2.33 fish ⁻¹	51–117 (5% loss) to 102–117 (10% loss) per 1000 juveniles	Isshiki <i>et al.</i> (2007)
Orange-spotted grouper, <i>Epinephelus coioides</i> (Hamilton, 1822)			
<i>Zeylanicobdella arugamensis</i>	A heavy infection of leeches on juveniles (83% infected) and adults (17%) reared in brackish water (22‰) tanks subsequently resulted in the mortality of the adult ($n = 1$) 3 days post-infection. The cause of death was suggested to be blood loss, secondary infection and loss of appetite	–	Cruz-Lacierda <i>et al.</i> (2000)
Malabar grouper, <i>Epinephelus malabaricus</i> (Bloch et Schneider, 1801)			
<i>Cryptocaryon irritans</i> + <i>Trichodina</i> sp.	Mixed infections were suggested to be the primary cause of disease outbreaks in juvenile stock ($n = 100$; av. 3.6 cm) imported from Thailand to Penang, Malaysia. Disease outbreaks occurred 2–3 weeks post-transfer; 97% of the healthy and 90% of the diseased grouper had <i>Trichodina</i> sp. infections	–	Leong and Wong (1990)
<i>Sphaerospora epinepheli</i>	Disease outbreaks in cage cultured stock in 100–800 g sized fish and ~15–20 kg broodstock between May–Aug each year at sites situated on both the Gulf of Thailand and Andaman Sea coastlines. Fish have loss of equilibrium, haemorrhagic regions across the body, the lumen of renal tubules contained masses of spores whilst large numbers of pseudo-plasmodia were seen attached to the epithelium of the tubules, which had become necrotic with evident peritubular fibrosis. Details relating to the magnitude of loss not given	–	Supamattaya <i>et al.</i> (1990)
<i>Ergasilus borneoensis</i>	Approx. 18.5% of stock ($n = 254$; av. wt. 579.3 g; range 260–920 g) sampled from floating cages in Penang Malaysia were infected with a mean intensity of 177.4 copepods fish ⁻¹ . No details relating to the impact of this parasite on the gills of host fish were provided	–	Leong and Wong (1988)
<i>Ergasilus lobus</i>	Approx. 9000 juveniles were imported from Thailand to brackish water pond site in	3308	Lin and Ho (1998)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
<i>Proserhynchus pacificus</i>	Tainan County, Taiwan in June 1993. Subsequently ~ 3–400 fish day ⁻¹ died on a daily basis over the following 15 days (i.e. approx. 5250 fish); all fish were infected, the gills had numerous ‘vacuoles’ and parasites. Loss is estimated on the adjusted value for harvest-sized fish in Taiwan between 1993 (i.e. US\$ 4.98 kg ⁻¹) and 2011 (i.e. US\$ 13.04 kg ⁻¹) and the adjusted value for juveniles given in Table 6 (i.e. 38% of their 2011 value). It is assumed that the juveniles weighed 100 g at the time of loss; loss is extrapolated from the value of fish in 2011, i.e. US\$ 1.68 fish ⁻¹ , and a value of US\$ 0.63 fish ⁻¹ is used Approx. 97.2% of stock ($n = 254$; av. wt. 579.3 g; range 260–920 g) sampled from floating cages in Penang Malaysia were infected with a mean intensity of 81.2 parasites fish ⁻¹ . No details relating to the impact of this parasite on stocks were provided	–	Leong and Wong (1988); see also study of Leong and Wong (1990)
<i>Pseudorhabdosynochus epinepheli</i>	All the stock (100% prevalence of infection; $n = 254$; av. wt. 579.3 g; range 260–920 g) sampled from floating cages in Penang, Malaysia were infected with a mean intensity of 335 flukes fish ⁻¹ The infection levels in wild fish were three times lower (mean intensity of 126.6 flukes fish ⁻¹ ; prevalence of infection 94.3%; $n = 35$; av. wt. 415.3 g; range 170–900 g) Juvenile stock ($n = 100$; av. 3.6 cm) imported from Thailand into Malaysia in Dec 1986 were infected (prev. 92.9%) with a mean intensity of 29.5 flukes fish ⁻¹ . Similarly, 92% of a shipment importation from the Philippines ($n = 50$; av. 15.4 cm) were infected with 51.1 flukes fish ⁻¹	–	Leong and Wong (1988) Leong and Wong (1990)
<i>Epinephelus coioides</i> + <i>E. malabaricus</i>			
Leeches (unspecified)	A survey of small scale grouper culture in the Philippines (72 farmers +1 co-operative of 65 families) in July–Sept 1999 found a 15.3% occurrence of infection resulting in an av. mortality rate of 30% (up to 100% in some ponds) in nursery and grow-out stock. Loss is est. on a 125 m ³ pond, stocked at 15 kg m ³ with juvenile (av. 100 g) fish, i.e. 18 750 fish cage ⁻¹ , and, the harvest price of Filipino fish in 1992, i.e. US\$ 13.00 kg ⁻¹ (NB: in FAO FishStatJ this is given as a price for seabass and grouper)	~ 7310 per cage	Somga <i>et al.</i> (2002)
(k) Olive flounder/Japanese flounder/bastard halibut, <i>Paralichthys olivaceus</i> (Temminck et Schlegel, 1846)			
<i>Ichthyobodo</i> sp.	Approx. 17 200 metamorphosed juveniles (av. 0.14 g) were lost over a 4-day period in March 1989 from two 15 m ³ tanks in Okayama Prefecture, Japan which were supplied with 17 °C, 30‰ seawater at a rate of 20 L min ⁻¹ . Loss is est. on each fish being valued at US\$ 0.05, however, Tomiyama <i>et al.</i> (2008) suggest that the price of a 10 cm juvenile trawled juvenile is 100 yen (=US\$ 1)	860	Urawa <i>et al.</i> (1991)
<i>Miamiensis avidus</i> (syn. <i>Phiasterides dicentrarchi</i>)	The causative agent of scuticociliatosis in Korean stock at several farm sites is identified from collected brain, gill and ulcerated skin samples. No details of loss throughout the Korean industry are provided	–	Kim <i>et al.</i> (2004a); Jung <i>et al.</i> (2007)
<i>Pseudocohnilembus persalimus</i>	Ciliates causing scuticociliatosis were collected from the brains, gills and ulcerated skin from stock at several Korean farm sites are identified. No details of loss are provided	–	Kim <i>et al.</i> (2004b)

Unidentified scuticociliatid ciliate	Study makes reference to this parasite as being commonly responsible for the mass mortality of fry and juveniles within Japanese hatcheries. Also reported to occur in commercial-sized fish. No details relating to the economics of loss throughout the Japanese industry are given	–	Yoshinaga and Nakazoe (1993)
<i>Neobenedenia melleni</i> ² (syn. <i>girellae</i>)	A heavy infection of 2000 flukes (fish ⁻¹ ?) recorded on 310–340 g stock held in floating cages in Oita Prefecture, Japan in Oct 1991. Details of loss not provided. Loss is est. on a 20 kg m ² (see http://www.lib.noaa.gov/retiredsites/korea/main_species/flounder.htm) stocking density, a cage area of 100 m ² (see Yoon, 2008), the assumption that heavy burdens resulted in a 50% loss of stock, and, the price of Japanese stock in 1991, i.e. US\$ 20·32 kg ⁻¹ (see the entry in FAO FishStatJ under bastard halibut)	20 320	Ogawa <i>et al.</i> (1995)
<i>Neoheterobothrium hirame</i>	Infections in the buccal cavity cause anaemia in spawning stock maintained in tanks and in fish reared in off-shore nets	–	Michine (1999); Ogawa (1999)
<i>Uronema marinum</i>	Mass mortality of farmed fry and high cumulative mortality of juveniles frequently observed in numerous farms. Loss is based on at least one 2000 t cage (i.e. 10 × 10 m ²) at 5 sites each stocked with 100 000 juveniles experiencing a loss of 50%. The value of each fish is est. at US\$ 0·10	50 000	Jee <i>et al.</i> (2001)
(l) Pompano, <i>Trachinotus</i> spp.			
<i>Lepeophtheirus spinifer</i>	Lice on market-sized (~ 350 g) reared in floating cage-reared <i>Trachinotus blochii</i> in Bataan Province Central Luzon, Philippines, caused lesions and scale loss reducing the market value of fish	–	Cruz-Lacierda <i>et al.</i> (2011)
(m) Flathead grey mullet, <i>Mugil cephalus</i> L.			
<i>Amyloodinium</i> -like	Infection within a culture facility resulted in 6 mortalities; the remaining 3 moribund fish (40–51 cm) were sampled to determine the cause of mortality	–	Baticados and Quintio (1984)
<i>Myxobolus parvus</i>	Mild to heavy gill and visceral infections seen in stock kept in ponds (11–13%); no morts	–	Paperna (1975)
<i>Benedenia monticelli</i>	Skin and mouth infections caused mass mortality of cultured grey mullet (80–100 mm long) in Eilat; captured fish with typically <6 parasites fish ⁻¹ stocked in 0·75–1 m ³ open, seawater culture tanks, die within 2 weeks of stocking. It is assumed that a 50% mortality occurred in three 1 m tanks stocked at 10 kg m ³ with av. 5 g (10–12 cm) and the value of individual wild-caught fish was US\$ 0·05 fish ⁻¹	150	Paperna (1975); Paperna <i>et al.</i> (1984)
<i>Ergasilus lizae</i>	Mortalities in brackish ponds (11–13%) in Israel, 90% infection with av. 10–70 parasites fish ⁻¹ on fish >50 mm long. Loss is assumed that the loss occurred in similar 250 m ³ ponds referred to in Paperna <i>et al.</i> (1977), stocked at 3 fish m ³ , that the av. size of infected fish was av. 5 g (10–12 cm), av. mortality was 12%, and, the value of individual wild-caught fish was US\$ 0·05 fish ⁻¹ . It is assumed that at least 10 ponds were affected and were the basis of the report	45	Paperna (1975)
<i>Pseudocaligus apodus</i>	Open sores on dead fish held in a seawater pond at the Dor Fisheries Research Station situated in the coastal plain of Israel. The entire stock was lost. It is assumed that the single pond measured 250 m ³ in capacity, stocked at 3 fish m ³ (av. 5 g), each valued at US\$ 0·05 fish ⁻¹	38	Paperna (1975)
(n) Barramundi, <i>Lates calcarifer</i> (Bloch, 1790)			
Fish diseases gen. spp.	Over 80% of farmers remarked that disease, including parasites, in small scale cage culture in Thailand (i.e. 1–5 cages) results in the 30–50% loss of stock. Loss is est. on fish being grown in 5 × 5 × 5 m ³ cages at a stocking density of 15 kg m ³ , and, the value of each 150 g fish is US\$ 2·00 (see Table 7)	7500–12 500 per cage	Bondad-Reantaso <i>et al.</i> (2001); Kanchanakhan <i>et al.</i> (2001)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
<i>Cryptosporidium</i> -like	A total of 12 000 out of 35 000 (i.e. ~34%) 9-week-old fry (av. 35–45 mm) were lost from an Australian hatchery over a 2-week period. Loss is based on the US\$ 0.12 fish ⁻¹ value given in Table 7	1440	Gabor <i>et al.</i> (2011)
	A low-level mortality (10–20 fish per 10 000 day ⁻¹) and emaciation in 2% of the hatchery stock (30–40 mm long) prompted an investigation which found a <i>Cryptosporidium</i> -like infection in 29% of the fish that were sampled. Shortly thereafter, moderate lymphocytic and plasmacytic gastroenteritis in the fish resulted in losses rising to 1100 fish day ⁻¹ with a total loss of 20 000 fish. Loss is based on the US\$ 0.12 fish ⁻¹ value provided in Table 7	2400	Gabor <i>et al.</i> (2011)
<i>Eimeria</i> sp.	A cumulative mortality of up to 30% was reported in 2.5–7 cm juveniles stocked in 5 nurseries (each stocked with 3000–5000 fish) in Ca Mau, Vietnam. <i>Eimeria</i> infections were seen in more than 60% of the diseased fish sampled from each site. Loss is est. on an av. 30% mortality within 20 000 fish valued at US\$ 0.12 fish ⁻¹ (see Table 7)	720	Gibson-Kueh <i>et al.</i> (2011)
<i>Trypanosoma</i> sp.	High mortalities were recorded in sea cage reared stock in the Northern Territory, Australia in July, 2005. Fish were lethargic, anaemic and blind with intra-ocular haemorrhages, there were large haemorrhagic ulcers on the skin, fish had massively enlarged spleens with huge numbers of trypanosomes in the blood. No details relating to the number and size of fish lost are given; however, an image provided in the report would suggest the fish were 30–40 cm in size (est. av. 1.5 kg). It is assumed that a 'high' mortality infers ~10%, that at least four 5 × 5 × 5 m ³ cages stocked at 10 kg m ³ were affected, and, the fish were valued at the av. harvest price for Australian fish in 1995, i.e. US\$ 6.69 kg ⁻¹	3345	Schipp <i>et al.</i> (2007)
<i>Caligus epidemicus</i>	Infections on juvenile fish (100% prevalence; av. 12.7 lice fish ⁻¹ ; range 6–35) resulted in the loss of appetite, lethargy and poor growth performance. Fish were stocked in floating net cages within earthen ponds at Dumangas, Iloilo, Philippines	–	Cruz-Lacierda <i>et al.</i> (2011)
	The prevalence of infection of stock across 6 Malaysian sites was determined to be 55.85% (i.e. 267/456 fish) with a mean intensity of 14.3 ± 23.8 lice fish ⁻¹ . Although <i>C. epidemicus</i> is one of many <i>Caligus</i> species found on stock, it is the most prevalent species and is reported to be a serious problem in production	–	Muhd-Faizul <i>et al.</i> (2012)
<i>Cirolana fluviatilis</i>	An infection on fingerlings (8–12 cm, 12–22 g) stocked in 3 cages 5 × 2 × 2 m ³ at a density of 2 kg m ³ (i.e. 130 fingerlings m ³) at a site in Cochin, India resulted in a 35% mortality after 2 months and 45% after 6 months (i.e. 1127 fish, Nov 2006–May 2007). Loss is est. on a value of US\$ 0.5 fish ⁻¹	564	Sanil <i>et al.</i> (2009)
<i>Diplectanum latesi</i>	The mortality of 4.5–7.0 kg broodstock held at an experimental station at Muttukkadu, India in Nov–Dec 1989(?) is attributed to heavy monogenean burdens on the gills. No details regarding the number of fish lost are provided. Loss is estimated on the mortality of 50 fish av. wt. 5 kg and the harvest price of Thai fish in 1999 (as no figures exist for India), i.e. US\$ 2.69 kg ⁻¹	807	Rajendran <i>et al.</i> (2000)
<i>Neobenedenia melleni</i>	The infection of stock in sea cages within Hinchinbrook Channel, Australia in Aug 2000 resulted in the loss of ~50 t of fish (i.e. ~200 000) worth an est. US\$ 277 000	277 000	Deveney <i>et al.</i> (2001)

	Fluke numbers were observed at >400 fish ⁻¹ . Low seawater temps (i.e. ~19 °C) regarded a contributory factor	–	
<i>Pseudorhabdosynochus latesi</i>	This monogenean is reported to occur at high intensities on stock sampled from floating cages in Penang, Malaysia	–	Leong and Wong (1986, 1988, 1990)
(o) Cyprinids			
From FAO FishStatJ the 100 000 t of ‘cyprinids’ cultured in Egyptian brackish waters ranks as the world’s 15th largest brackish/marine finfish aquaculture industry, however, this figure results from the production of multiple species (possibly <i>Cyprinus carpio</i> , <i>Hypophthalmichthys molitrix</i> , <i>Ctenopharyngodon idellus</i>). As the precise tonnage of each species cannot be identified, they are not considered here as part of this review			
(p) Red seabream <i>Pagrus major</i> Temminck et Schlegel, 1843			
NB: In the absence of Latin binomials within the FAO FishStatJ database to identify species, finding accurate data for this species has proven difficult			
<i>Enteromyxum lei</i>	Low but consistent mortality due to enteromyxosis seen in cages of ~350 g fish in Greece in 1994. The study comments that most of the fish were removed and removed during post-processing. This event occurred alongside a mortality event in 2 cages of <i>Puntazzo puntazzo</i> which resulted in the loss of 30% of stock over a 6-month period. In the absence of details, it is assumed that at least two 125 m ³ cages were affected with an equivalent 30% loss (a combination of direct mortality and subsequent carcass rejection; i.e. 1125 kg), that cages were stocked at 15 kg m ³ , and, that the fish would have achieved the av. harvest price for Greek fish in 1994, however, this species is not readily identifiable from the FAO FishStatJ database and so a value of US\$ 8.66 kg ⁻¹ is used (i.e. the av. value of red porgy at US\$ 7.83 kg ⁻¹ and gilthead sea bream at US\$ 9.49 kg ⁻¹)	9743	Le Breton and Marques (1995)
<i>Henneguya pagri</i>	Infections followed after the importation of Chinese seabass seed into Japan in the early 1990s. A chronic pattern of mortality is reported in juveniles (50–90 g). Infected fish which typically have anaemic gills, an enlarged <i>bulbus arteriosus</i> and suffer internal haemorrhages in the pericardial cavity. It is assumed that at least two 125 m ³ cages stocked at 15 kg m ³ with 50 g fish (i.e. 12 500 fish cage ⁻¹) were affected each year, that the price of juveniles followed those given in Imai (2005), i.e. ¥4000–15 000 kg ⁻¹ live fish (=US\$ 39.10–146.62) equivalent to US\$ 1.96 to 7.33 fish ⁻¹ . Loss is based on the assumption that chronic infections resulted in 2.5% of starting stock (i.e. 625 fish across 2 cages)	1225–4581 p.a.	Yokoyama <i>et al.</i> (2005a, b)
<i>Kudoa iwatai</i>	Infection identified from farm specimens collected in Kinko Bay, Japan	–	Egusa and Shiomitsu (1983)
<i>Microsporidium seriola</i>	Juvenile sea bream (2.5–3.5 cm) collected from off shore cages; muscle liquefaction around cysts masses; morbidity ~20% but mortality lower than this. Loss is based on 2 cages of 125 m ³ cages stocked at 15 kg m ³ with 2 g fish (i.e. 937 500 fish cage ⁻¹) that the value of each fish is 70% of that given in Imai (2005), i.e. US\$ 0.003–0.01 fish ⁻¹ . A lower figure based on 5% mortality (i.e. loss of 46 875 fish cage ⁻¹) and an upper figure of 20% morbidity is given (i.e. subsequent loss of 1 87 500 fish cage ⁻¹). The upper and lower value of the fish for each is provided	282–938 (5% mortality) 1125–3750 (20% morbidity)	Egusa <i>et al.</i> (1988)
Unidentified scuticociliatid ciliate	Study makes reference to this parasite as being responsible for the mortality of Japanese fry and fingerlings in Hiroshima Prefecture, and also of commercial sized fish (unpublished data). Loss is based on an est. 10% mortality in both 2 g juveniles and 350 g harvest-sized fish stocked in a 125 m ³ cage stocked at 15 kg m ³ , and, based on the juvenile price suggested by Imai (2005) and in the absence of a harvest price in FA FishStatJ, an av. price based on other Japanese sea breams produced and sold in 1992, i.e. US\$ 9.85 kg ⁻¹ (i.e. blackhead sea bream, <i>Spondyliosoma cantharus</i> *, at US\$ 13.53 kg ⁻¹ ; crimson sea bream, <i>Evynnis japonica</i> , at US\$ 8.04 kg ⁻¹ ; and, silver sea bream, <i>Diplodus argenteus</i> spp., at US\$ 7.99 kg ⁻¹)	282–938 (for 2 g juveniles) 1847 (harvest size)	Yoshinaga and Nakazoe (1993)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
*Again the lack of Latin binomials in the FAO FishStatJ is a source of confusion as it is believed that this species should be black sea bream, <i>Acanthopagrus schlegeli</i>			
<i>Anoplodiscus tai</i>	5-year-old farm fish rejected at market. Heavily infected fish (26–64 parasites fish ⁻¹ ; 98.5% on the fins) are emaciated with partial fin loss. Parasites induce epithelial hyperplasia and loss of epithelium at sites of attachment	–	Ogawa (1994)
<i>Caligus sclerotinosus</i>	A survey of sea ranched stock reared in net cages at a farm site at Tongyeong, Gyeongsangnamdo, South Korea throughout June to Nov. 2011 found severe infections (100% prev., mean intensity 7.06 lice fish ⁻¹ , range 2–49 lice fish ⁻¹). No mortalities were reported in this study the authors make reference to winter mortalities within farms at Yeosu, Jeollanamdo but it is not clear if these can be directly attributed to <i>C. sclerotinosus</i> . This parasite was reported to have been introduced to Japan from Hong Kong in a shipment of brood stock (see Miyazaki <i>et al.</i> 1986)	–	Venmathi Maran <i>et al.</i> (2012)
Larval gnathiids	21 broodfish (0.45–1.1 kg; 2–4 yrs old) reared in concrete tanks in Italy, were killed by 3 episodes of infection. Average infection was ~30 parasites fish ⁻¹ with up to 146 parasites recorded on 1 fish	–	Patarnello <i>et al.</i> (1995)
(q) Korean rockfish, <i>Sebastes schlegelii</i> (Hilgendorf, 1880)			
<i>Microcotyle sebastis</i>	High seasonal water temperatures, high prevalences of infection (77.8–100%) and parasite burdens (0–2120 fish ⁻¹) are suggested to be factors underlying the mass mortality of cultured stocks in South Korea. Estimates of loss are based on the Korean price of harvest-sized fish in 2011, i.e. US\$ 7.91 kg ⁻¹	7910 t ⁻¹	Yoon <i>et al.</i> (1997)
	The in-feed administration of praziquantel (20 g kg ⁻¹ commercial feed fed to 14 g fish as 1% ration on alternate days for a total of 4 treatments) was effective in the near complete removal of gill monogeneans. Assuming that failure to treat would otherwise result in the complete loss of stock, then 800 g praziquantel would be required for four 1-day treatments (assuming 1 kg of drug costs ~ US\$ 110; Xian Wango Biopharm Co. Ltd.)	88 t ⁻¹ treated	Kim and Cho (2000)
(r) Chinook salmon, <i>Oncorhynchus tshawytscha</i> (Walbaum, 1792) and other species of <i>Oncorhynchus</i>			
<i>Hexamita salmonis</i>	18–43% mortality in 1–2 kg stock at a net-pen farm in British Columbia in Sept 1991, fish were anaemic, had enlarged kidneys, spleens and congested livers, and, swollen abdomens with serosanguinous ascites. Loss is based on the value of harvest-sized fish in Canada for 1991, i.e. US\$ 3.74 kg ⁻¹ . Assumed cage volume is min 125 m ³ stocked at ~12.5 kg m ³ , i.e. 1562.5 kg biomass	1050–2510 per infected cage	Kent <i>et al.</i> (1992)
<i>Kudoa thyrsites</i>	An infection rate of 4% was found within the cardiac muscle of fresh mortis in Oct. 1987 at 1 of the 3 hatcheries that were sampled along the coast of British Columbia, Canada	–	Kabata and Whitaker (1989)
<i>Loma salmonae</i>	An infection within a stock of 337 869 hatchery fish at a hatchery in Anchorage, Alaska throughout March to mid-May 1980 resulted in the mortality of 9.95% (i.e. 33 618 fish). No details relating to the size of the fish are provided but it is est. that the fish were 2–5 g valued at US\$ 0.10 fish ⁻¹	3362	Hauck (1984)
<i>Nucleospora salmonis</i> (previously <i>Enterocytozoon salmonis</i>)	Elston <i>et al.</i> (1987) found that an intranuclear microsporidium of haematopoietic cells (47% of haemoblast nuclei were infected) was responsible for acute anaemia in 3-year-old salmon reared in net pens in Washington State, USA. Features of this infection were similar to a case plasmacytoid leukaemia in adult chinook salmon reared in seawater reported by Kent and Dawe (1990) that resulted in severe mortalities (no details provided) that occurred at several sites in British Columbia. An experimental infection by Hedrick	6615	Elston <i>et al.</i> (1987); Kent and Dawe (1990); Hedrick <i>et al.</i> (1991)

	<i>et al.</i> (1991) resulted in 100% mortality 53–60 days post-injection. An estimate of loss is provided for the study of Kent and Dawe (1990) which assumes that at least two $10 \times 10 \times 10 \text{ m}^3$ cages stocked at 15 kg m^{-3} were infected resulting in a 10% mortality, and, the value of Canadian fish in 1990, i.e. US\$ 4.41 kg^{-1}		
<i>Sphaerothecium destruens</i>	80% mortality of 3-year-old broodstock held in a net-pen Puget Sound, US over an 8-month period between 1983 and 1984. All fish had enlarged kidneys and spleens. Loss is based on the value of harvest-sized fish in Canada for 1984 as no figures are available for the US, i.e. US\$ 3.59 kg^{-1} . Assumed cage volume is min 125 m^3 stocked at $\sim 12.5 \text{ kg m}^{-3}$, i.e. $1562.5 \text{ kg biomass}$	4490	Harrell <i>et al.</i> (1986)
<i>Gilquinia squali</i>	Trypanorhynch metacestodes within the vitreous humor (1–2 parasites eye^{-1}) resulted in blindness and a 10% mortality in a single net-pen. Loss is based on the value of harvest-sized fish in Canada for 1988, i.e. US\$ 5.40 kg^{-1} . Assumed cage volume is min 125 m^3 stocked at $\sim 12.5 \text{ kg m}^{-3}$, i.e. $1562.5 \text{ kg biomass}$	845	Kent <i>et al.</i> (1991)
Chum salmon, <i>Oncorhynchus keta</i> (Walbaum, 1792) ^c			
<i>Ichthyobodo necator</i>	Parasite survives transfer to seawater (33%), can cause severe epidermal destruction, drastically reduced tolerance to seawater and juvenile mortality (60–70%, 4–6 weeks after transfer).	–	Urawa and Kusakari (1990); Urawa (1995)
	Experimental trial demonstrates that infected fry ($n = 1000$; av. wt. 0.75 g) can be transferred from freshwater into 33 ppt seawater and that the flagellates survive and can reproduce. A total of 17 426 flagellates were found on 10 fry 1 week post-transfer. After two 2 weeks, the fish began to die and final mortality exceeded 40%. In March and April 1986, approx. 30 million hatchery reared fry were released into the Chitose River, Hokkaido, Japan. The prevalence of infection was 34% in the hatchery but following formalin treatments was reduced to 2%. The prevalence of infection on fry in the river in April and May was found to be 20 and 34% respectively, whilst samples taken from the mouth of the Ishikari River were determined to be 32 and 27% in the same months. Loss estimates are based on an av. infection of 28.25% on the fry released into the river, the assumption that 40% of these infected fish will die (i.e. 3 390 000 fry), and, the individual est. value of released fry in 1990, i.e. US\$ 0.002 fish^{-1} . An upper estimate is also provided based on the assumption that all infected fish die, i.e. $8 475 000 \text{ fry}$	67 800–169 500	Urawa and Kusakari (1990)
	Trial demonstrates that 60–70% of infected fish transferred to seawater die within 4–6 weeks. Treatment with 250 ppm formalin (1 h) is effective in reducing mortalities. Without treatment $\sim 30\%$ of transferred fry die. Following treatment, a total of 9 million hatchery fish were subsequently released 4 weeks post-treatment. Loss is based on the assumption, that in the preceding year, the Yoichi River Hatchery in western Hokkaido, lost 30% of fish (i.e. $\sim 3 850 000 \text{ fry}$), and, the the individual est. value of released fry in 1987–1988, i.e. US\$ 0.002 fish^{-1}	77 000	Urawa (1993, 1995)
Pink/humpback salmon, <i>Oncorhynchus gorbuscha</i> (Walbaum, 1792) ^c			
<i>Kudoa thyrssites</i>	An infection rate of 1% was found in the cardiac muscle of fresh mortis in Sept 1987 at a single hatchery situated on the coast of British Columbia, Canada	–	Kabata and Whitaker (1989)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
(s) Large yellow croaker, <i>Larimichthys crocea</i> (Richardson, 1846)			
<i>Cryptocaryon irritans</i>	Study infers that infections are a problem in culture but does not provide specific details	–	Lin <i>et al.</i> (2013)
(t) Red drum, <i>Sciaenops ocellatus</i> L.			
<i>Enteromyxum leei</i>	Stock (23–117 g) held in an Israeli culture facility are shown to be susceptible to infection by co-habitation with infected <i>S. aurata</i> (45·8%) or by exposure to effluent water from the tanks holding infected stock (35·0%)	–	Diamant (1998a)
(u) Tiger pufferfish, <i>Takifugu rubripes</i> (Temminck et Schlegel, 1850)			
<i>Enteromyxum fugu</i> (syn. <i>Myxidium fugu</i>) + <i>Leptotheca fugu</i>	Infections in the intestine which result in emaciation disease have been reported since 1996 resulting in mortalities, typically of 1-year-old stock, between May/June through to Aug/Sept). The resultant mortality is variable (prev. <20 to >60% at different farm sites). A 10% mortality in a single 125 m ³ cage stocked at 2·5 kg m ³ is assumed, and, the harvest price of Japanese fish between 1996 to 2002 (the latter date being that of the report), i.e. US\$ 22·00 (virtually constant throughout this period), is used to estimate loss	688 per cage	Tun <i>et al.</i> (2000, 2002); Ogawa and Yokoyama (2001)
<i>Enteromyxum leei</i>	Reported to be highly pathogenic and one of the causative agents of emaciation disease	–	Yanagida <i>et al.</i> (2005)
<i>Ichthyobodo</i> sp.	Infections were said to frequently occur in spring in net reared stock (av. 226–463 g) along the western coast of Okayama Prefecture, Japan	–	Urawa <i>et al.</i> (1998b)
<i>Kudoa shiomitsuui</i>	Infection first described from the pericardial cavity of farm reared fish (135–360 g) collected from Kagoshima prefecture, Japan		Egusa and Shiomitsu (1983)
<i>Leptotheca fugu</i> hyperparasitised with a microsporean	The host inflammatory response in individuals infected with hyperparasitized <i>L. fugu</i> is reported to be more severe than with infection of <i>L. fugu</i> alone	–	Tun <i>et al.</i> (2002)
<i>Heterobothrium okamotoi</i>	Infections in the gills and wall of the branchial cavity lead to anaemia. This parasite was believed to be the cause of major mortalities seen in Japanese farms throughout the 1950s–1960s. No details of loss are provided but it is assumed that major mortality events allude to the loss of >20% stock	–	Okamoto (1963)
	Losses due to heavy infections are reported to be rising with expansion of the industry. Infection was first noted 5 months after the fish were transferred (i.e. in Nov). No details relating to the magnitude of loss are provided but losses of 10% in 125 m ³ cages stocked at 2·5 kg m ³ , with a harvest price of US\$ 22·00 (1996–2002), are assumed	688 per cage	Ogawa and Inouye (1997); Ogawa <i>et al.</i> (2005b)
	Parasites produce eggs in strings that can exceed 2 m in length. These become caught in the net mesh, accumulating and representing a source of infection. Regular net changes are required as part of parasite management	–	Ogawa and Yokoyama (1998)
	Recurrent problems seen in early 2000s in floating net-pens	–	Ogawa (2002)
Bullseye pufferfish, <i>Sphoeroides annulatus</i> (Jenyns, 1842)			
<i>Amyloodinium ocellatum</i>	Severe infections can result in the sig. mortality of cultured stocks – no losses are provided but a 5% mortality, in a 125 m ³ cage stocked at 2·5 kg m ³ , and a harvest	78 per cage	Fajer-Ávila <i>et al.</i> (2003)

	price of US\$ 5.00 kg ⁻¹ (see García-Ortega <i>et al.</i> 2002) for Mexican fish in 2002 is applied		
<i>Heterobothrium ecuadori</i>	Heavy infections can result in the significant loss of stocks – no details provided	–	Fajer-Ávila <i>et al.</i> (2003)
<i>Lepeophtheirus simplex</i> + <i>Neobenedenia</i> sp.	Report that dual infections can cause skin lesions, anorexia and mortality of juvenile stock. No details are provided	–	Fajer-Ávila <i>et al.</i> (2008)
(v) Porgies, sea breams			
Black porgy, <i>Acanthopagrus schlegelii</i> (Houttuyn, 1782)			
<i>Alella macrotrachelus</i>	Infections on farm stock at a site at Tashima, Japan were followed over Aug. 1978 to Aug. 1980; fish typically reach 100–150 g in 12 months. Infections reached 100% within 3–4 months with up to 15 females fish ⁻¹ resulting in hyperplasia, oedema and haemorrhages of the gill lamellae. These copepods posed the greatest threat in the first winter when the fish were of a smaller size but infections were at their peak. The study does not allude to the loss of stock	–	Muroga <i>et al.</i> (1981)
<i>Caligus acanthopagri</i>	Fish reared in ponds at Chi-Ku village Tainan County, Taiwan in March 1983 had an abnormal feeding condition, were emaciated and black in appearance. Daily mortality of 5–10 fish day ⁻¹ . A total of 713 adult lice were removed from the body surface of 3 moribund hosts. No details relating to the total loss of stock are provided. Loss is estimated on the assumption that the mortality event lasted at least 14 days, occurred in 3 ponds, fish had an av. wt. of 250 g, and, the price of Taiwanese harvest-sized stock was US \$ 6.00 kg ⁻¹ (no figures available for 1983; first figures available are for 1989 when the price was US\$ 7.27 kg ⁻¹)	473	Lin <i>et al.</i> (1994)
<i>Caligus multispinosus</i>	There was a rising daily mortality of 10–30 fish in pond held stock at Pei-Men Village Tainan County, Taiwan in Nov. 1992. A total of 22 adult lice and many juveniles were removed from the gills and buccal cavity of 4 moribund hosts. Loss is estimated on the assumption that the mortality event lasted at least 14 days, occurred in 3 ponds, fish had an av. wt. of 250 g, and, the price of harvest-sized fish in Taiwan was US\$ 7.95 kg ⁻¹	1670	Lin <i>et al.</i> (1994)
Sharp-snout sea bream, <i>Diplodus puntazzo</i> (Walbaum, 1792)			
<i>Amyloodinium ocellatum</i>	100% mortality (i.e. 50000) 12 g juveniles lost over 48 h in a sea cage in Greece in the summer of 1997 when sea water temperatures were 23 to 26 °C. Loss is based on the unchanged value of stock, i.e. US\$ 0.02 fish ⁻¹ (see Table 7)	1000	Rigos <i>et al.</i> (1998)
<i>Ceratomyxa diplodae</i>	A survey of 109 fish (49–650 g) conducted between Dec 2000 and Jan 2002 found infections with a prevalence of 51% in the gall bladder and 2% in the intestines	–	Merella <i>et al.</i> (2005)
	A tank of 32 fish (av. wt. 206 g) began dying 35 days after a treatment with 17b-estradiol for sex inversion. All fish were lost within 90 days. Loss is based on the price of harvest-sized fish in 2011, the first figures available, i.e. US\$ 8.16 kg ⁻¹	54	Katharios <i>et al.</i> (2007)
<i>Ceratomyxa sparusaurati</i>	Low levels of loss, i.e. 0.1% d ⁻¹ , observed in Greece.	–	Rigos <i>et al.</i> (1999)
	An infection rate of 30% was found at a single cage site within the Adriatic	–	Mladineo (2003)
<i>Cryptocaryon irritans</i>	100% of stock (<i>n</i> = 59; 324–410 g) at one culture facility on the Spanish Mediterranean coast died within 15 days of the first observed mortalities. Loss is based on an av. size of 350 g fish ⁻¹ and the av. price of Mediterranean sharp-snout sea bream in 2000,	188	Montero <i>et al.</i> (2007)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
<i>Enteromyxum leei</i>	i.e. US\$ 9.12 kg ⁻¹ (harvest price for Cyprus was US\$ 4.83 kg ⁻¹ ; price in Italy was US\$ 13.41 kg ⁻¹)		
	In July 1994, mortalities of 30–50 fish day ⁻¹ from 2 cages of 15 000 (30–60 g) fish in western Greece. After 6 months 35+ % of stock lost with chronic losses of 20–30 fish week ⁻¹ . Fish were emaciated with swollen abdomens, pale livers with high numbers of spores in the bile; 90% of the fish were found to be carriers of infection. Loss is estimated on the value of 150 g fish, i.e. US\$ 1.50 fish ⁻¹	15 750	Le Breton and Marques (1995)
	Approx. 80% loss of stock reported from 8 farm sites in central and southern Greece during the summer of 1997 when sea temperatures exceeded 22 °C. Fish lost were at the fattening stage and an av. fish wt. of ~400 g is assumed. Loss is estimated on the assumption that each farm produced min. 10 t, and, on the harvest price of Italian fish in 1997, i.e. US\$ 8.23 kg ⁻¹ , as no figures for Greek fish/production are available	526 720	Athanassopoulou <i>et al.</i> (1999)
	9% loss of stock observed over numerous Greek cage sites covering several geographical regions during the period Sept 1996–Sept 1997 when summer seawater temperatures rose to 24–25 °C. Losses at some sites were 1–5% day ⁻¹ with a chronic pattern of mortality over 3–6 weeks. Loss of 30–70% of stock at certain sites. Loss is based on the assumption that at least 5 sites were affected, production at each was a minimum of 10 t, and on the harvest price of Italian fish in 1997, i.e. US\$ 8.23 kg ⁻¹ , as no figures for Greek fish/production are available	~ 37 000	Rigos <i>et al.</i> (1999)
	A total of 13 000 juveniles (av. 25 g) were stocked in to a single 250 m ³ cage in the Gulf of Sardinia in July 2000. A monogenean infection in the winter of 2001 reduced the stock by ~1450. In Aug–Nov 2001, a second major mortality event due to <i>E. leei</i> resulted in daily mortalities of 0.26–0.38% and a final loss of 32% of stock (i.e. ~1800 fish). Parts of the stock were sold in Aug reducing the stock to 50%. Total losses following both events were 25% (i.e. ~3250/13 000). A subsequent survey conducted Dec 2000 to Jan 2002 looked at a total of 109 fish. <i>Enteromyxum</i> was found with a prevalence of 33% in the gall bladder and 41% in the intestines. Loss is based on the assumption that the av. wt of each fish at the time of the event was ~250 g and the price of harvest-sized fish in Italy in 2001, i.e. US\$ 5.13 kg ⁻¹	2309	Merella <i>et al.</i> (2005)
Reference is made to the severe mortalities of Mediterranean stocks that devastated production. A total of 464 t (i.e. Cyprus 64 t, Italy 400 t) were produced in 2001, this fell to 401 t in 2003 (i.e. Cyprus 1 t, Italy 400 t), and then no tonnage recorded for the period 2004 to 2006. If the decline in production is attributed to this one pathogen, then loss can be estimated using the harvest price of Italian fish in 2003, i.e. US\$ 6.56 kg ⁻¹	413 280 (in 2003) 2.63 M (in 2004)	Mladineo (2006)	
<i>Polysporoplasma sparis</i>	An infection of 20% was found at a single cage site within the Adriatic	–	Mladineo (2003)
<i>Atrispinum salpae</i>	A total of 13 000 juveniles (av. 25 g) were stocked into a single 250 m ³ cage in the Gulf of Sardinia in July 2000. In Jan–Feb 2001, high prevalence and intensity of parasites resulted in daily mortalities of 0.19–0.22% and a final cumulative mortality of 11.4% (i.e. ~1450 fish). A subsequent survey conducted Dec 2000 to Jan 2002	930	Merella <i>et al.</i> (2005)

	looked at a total of 109 fish. <i>Atrispinum salpae</i> was found on the gills with a prevalence of 93% (range 1–1370 flukes fish ⁻¹). Loss is based on the assumption that the av. wt of each fish at the time of the event was ~125 g and the price of harvest-sized fish in Italy in 2001, i.e. US\$ 5.13 kg ⁻¹		
<i>Lamellodiscus</i> sp.	43% of stock at 8 farm sites (50 + g fish) infected	–	Athanassopoulou <i>et al.</i> (1999)
<i>Lamellodiscus bidens</i> + <i>L. ergensi</i>	Five wild fish caught off Crete were examined to determine the pathology associated with infections. The prevalence of infection was 100% with each gill arch having an av. 100.2 ± 40.1 flukes (40% <i>L. bidens</i> ; 60% <i>L. ergensi</i>). These burdens resulted in hyperplasia and a proliferation of the gill epithelium notably at the base of the secondary lamellae where there are chloride cells. There was also fusion and destruction of the secondary lamellae. Although no losses were recorded, the study alludes to the risks of infections under intensive aquaculture conditions, which include potential economic loss as a result of reduced growth and/or loss of fish as a consequence of respiratory distress	–	Katharios <i>et al.</i> (2006)
<i>Microcotyle</i> sp.	70% of stock at 8 farm sites (50 + g fish) infected	–	Athanassopoulou <i>et al.</i> (1999)
White sea bream, <i>Diplodus sargus</i> L.			
General parasitic infection	Three Greek farm sites reported total mortalities of 12, 32 and 42%. Nephrocalcinosis infection seen in all sampled fish but mortalities attributed to <i>Kudoa</i> and <i>Myxobolus</i> infections. Samples taken Oct 2002–Oct 2003 had the following: <i>Enteromyxum leei</i> 8.0 ± 12.8 (0–40); prevalence of infection (range) <i>Kudoa</i> sp. 8.0 ± 16.6 (0–60) <i>Myxobolus</i> 33.1 ± 33.6 (0–90) <i>Furnestinia</i> sp. 63.6 ± 30.1 (0–80) <i>Microcotyle</i> sp. 8.4 ± 14.4 (0–40) Loss estimates are based on the value of harvest-sized fish in 2003, i.e. US\$ 5.32 kg ⁻¹	638–2234 t ⁻¹	Golomazou <i>et al.</i> (2006)
Blackspot sea bream, <i>Pagellus bogaraveo</i> (Brünnich, 1768)			
<i>Ceratomyxa sparusaurati</i>	An infection rate of between 6.66 and 33% was found at a single-cage site within the Adriatic	–	Mladineo (2003)
(w) Atlantic bluefin tuna, <i>Thunnus thynnus</i> L.			
<i>Anisakis simplex</i>	21.86% of the 183 tuna (5–12 kg) that were sampled from 3 Adriatic sites throughout 2003–2006 bore infections with a mean abundance of 3.05 nematodes fish ⁻¹ . Although the worms were not responsible for mortalities, the presence of these zoonotic nematodes raises public health concerns for those consuming fresh products	–	Mladineo <i>et al.</i> (2008)
<i>Cardicola forsteri</i>	A sample of 52 harvest-sized fish reared off the Island of Brač in the Adriatic revealed that the eggs of this sanguinicolid caused granulomas and a marked inflammatory response, primarily in the kidney, and to a lesser extent in the heart and gills, of 63.34% of those examined	–	Mladineo and Tudor (2004)
	A total of 62 fish from a site off Brač Island, Croatia were sampled in Jan 2003 and during a mortality event in July 2003. Eggs were found with a prevalence of 63.34% in the gill, heart and kidney. The study comments that there was inflammation associated with infection. No details relating to the mortality event were provided	–	Mladineo (2006)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
<i>Didymocystis wedli</i>	The gills of 73·68% (i.e. 38/52) harvest-sized fish that were sampled and held off the Island of Braè in the Adriatic were infected with an av. 13·26 encysted adults fish ⁻¹ . It was suggested that heavy burdens reduce respiratory function imposing an additional stress upon fish	–	Mladineo and Tudor (2004)
	A total of 183 tuna (5–12 kg) from 3 Adriatic sites were sampled throughout the winter of 2003 to the summer of 2006. Infections were found at a prevalence of 61·75% and an abundance of 28·91 flukes fish ⁻¹ . The presence of these didymozoid digeneans was reported to elicit a marked host reaction but were not attributed to mortality events	–	Mladineo <i>et al.</i> (2008)
<i>Euryphorus brachypterus</i>	Heavy infections have been reported on the pseudobranchs of wild fish result in bleeding and ulceration. Lesions were also reported on the skin and gills of wild fish. Infections may pose a threat to captive ranched stock	–	Williams and Bunkley-Williams (1996)
<i>Hepatoxylon trichiuri</i>	Sixty-two fish from a farm off Brač Island, Croatia were sampled on 2 occasions, i.e. Jan 2003 and during a mortality event in July 2003. Plerocercoids were attributed as the cause of haemorrhages within the mucosa and stomach. This cestode was found in 12·4% of tuna post-mortemed with a mean abundance of 0·12 cestodes fish ⁻¹	–	Mladineo (2006)
<i>Koellikerioides intestinalis</i>	An infection was determined in 54·64% with an abundance 10·96 digeneans fish ⁻¹ in the 183 tuna (5–12 kg) that were sampled from 3 Adriatic sites throughout 2003–2006. There was a marked host inflammatory response at the site of parasite attachment. These parasites were not, however, attributed to observed mortalities throughout the study	–	Mladineo <i>et al.</i> (2008)
<i>Oncophora melanocephala</i>	Stock ($n = 62$) from a farm off Brač Island, Croatia were sampled in Jan 2003 and during a mortality event in July 2003. Adult nematodes were prevalent in 57·89% of hosts with a mean abundance of 1·74 worms fish ⁻¹ . Haemorrhaging was associated with nematode infection in the pyloric caeca and mucosa. The study concludes that this parasite poses a potential threat to the host	–	Mladineo (2006)
(x) <i>Cobia, Rachycentron canadum</i> (Lamarck, 1766)			
Protista gen. spp.	<i>Amyloodinium ocellatum</i> , <i>Coccidia</i> spp., <i>Ceratomyxa</i> , <i>Cryptocaryon irritans</i> , <i>Epistylis</i> spp., <i>Kudoa</i> spp., <i>Myxidium</i> spp. and <i>Trichodina</i> spp. are all identified as potential pathogens to cobia culture	–	McLean <i>et al.</i> (2008); FAO (2014c)
<i>Brooklynella hostilis</i>	Is reported to have caused a Caribbean-wide mass mortality of wild stocks in 1980 but in Oct 2002, a consignment of 30 000 fingerlings (6·0–8·4 cm total length) being sent from a facility in Florida to Puerto Rico died as a consequence of heavy infections. Loss is based on the estimated value of juvenile fish, i.e. US\$ 2·00 (see Table 7)	60 000	Bunkley-Williams and Williams (2006)
<i>Cryptocaryon irritans</i>	A shipment of juveniles ($n = 15$; 8·4–11·2 cm) sent from Florida were stocked in a marine cage culture facility in Puerto Rico before quarantine checks were complete. A moderate infection on the gills resulted. It is suggested that the infection did not result in the loss of stock	–	Bunkley-Williams and Williams (2006)

<i>Ichthyobodo</i> sp.	In June 2003, a shipment of 7 juveniles (3.5–5.2 cm total length) received from a culture facility in Florida were added to cage stock in Puerto Rico before quarantine checks were complete. Some fish died during transit. A moderate infection on the skin and gills of cage stock was subsequently found. No further loss of stock was suggested	–	Bunkley–Williams and Williams (2006)
<i>Sphaerospora</i> -like myxozoan	90% mortality (i.e. loss of 49 500 fish) of 45–80 g juveniles beginning 1 month post-transfer to sea cages in Taiwan. Loss occurred over a 30-day period in 1999. Fish were anaemic, had grossly enlarged kidneys and had ascites; extrasporogonic and sporogonic stages were found in the blood, glomerulus, renal tubules and renal interstitium. Loss is calculated on the value of harvest-sized fish in Taiwan in 1999, i.e. US\$ 5.04 kg ⁻¹ and an av. fish weight of 60 g	14 969	Chen <i>et al.</i> (2001)
	Sporogonic stages were found in the lumen of renal tubules but infections were low and there was no obvious kidney damage	–	Lopez <i>et al.</i> (2002)
<i>Caligus lalandei</i>	Stocks reared in marine cages in the Pingtung area of Taiwan are reported as susceptible to the occasional infection	–	Chang and Wang (2000)
<i>Neobenedenia melleni</i> ^b (syn. <i>girellae</i>)	Oct 2000 and Feb 2001 juvenile stock reared in sea cages off the Penghu islands of Taiwan are reported to have severe ulceration of the cranium as a result of monogenean infection (av. 15–20 flukes fish ⁻¹) resulting in a 40% mortality of stock. No farm production details or size of the fish lost are provided. Loss is estimated on the assumption that av. wt. of fish were 100 g, that at least three 500 m ³ cages on at least 2 production sites were affected, fish were stocked at 20 kg m ³ , and, that value of each fish was US\$ 3.00 (see Table 7)	1.8 M	Lopez <i>et al.</i> (2002)
	A mass mortality of stock on Liu-chiu Hsu Island, Taiwan occurred over the period Oct 2002–Jan 2003. Freshwater baths were given at 1 week intervals to control infections. Subsequent sampling of the 0+ stock (12–37 cm long) in March 2003 found an av. 12.6 parasites fish ⁻¹ (range 1–33) localized principally on the dorsal surface of the cranium (59.7%) and on the eyes (23.7%). No details relating to the mass mortality are provided but loss is estimated on the mortality of 10% of stock, an av. fish wt. of 100 g, that at least three 500 m ³ cages were affected, fish were stocked at 20 kg m ³ , and, that value of each fish was US\$ 3.00	0.9 M	Ogawa <i>et al.</i> (2006)
<i>Parapetalus occidentalis</i>	In 2000, infections are reported in the gill cavities of stock being reared in offshore cage-nets around Penghu Islands, Taiwan. The study reports that 17 females were collected from cage stock to augment existing material for a taxonomy-based study but provides no indication of overall infection levels	–	Ho and Lin (2001)
(y) Meagre, <i>Argyrosomus regius</i> (Asso, 1801)			
<i>Amyloodinium ocellatum</i>	An infection in stock (av. 423 ± 110.5 g) held in 2500 m ³ earthen ponds at 0.75 kg m ³ with an equal weight of gilthead sea bream resulted in 1.2% mortality (i.e. 11.25 kg pond ⁻¹). Loss is estimated on the assumption that a minimum of 2 ponds were affected and on the market price of Portuguese fish in 2011, i.e. US\$ 8.90 kg ⁻¹	200	Soares <i>et al.</i> (2012)
<i>Benedenia sciaenae</i>	In the first season of commercial Turkish production, farmers at Akbüük, western Turkey in April 2005 reported scale loss and skin damage on stock (av. 5.63 ± 0.89 kg). A total of 9088 parasites were recovered from 40 fish (mean intensity 227 ± 77.5; range 84–386 flukes fish ⁻¹). No production figures (tonnage or value) are listed for Turkey in FAO FishStatJ, but figures for France (i.e. US\$ 8.72 kg ⁻¹), Italy (i.e. US\$ 8.41 kg ⁻¹) and for Portugal (i.e. US\$ 12.78 kg ⁻¹) for 2005 are available. Production in the first year is assumed to be 5 t, that infections resulted in a 20% downgrading in price, and, that the price of harvest-sized were similar to those sold in Italy	8410	Toksen <i>et al.</i> (2007)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
<i>Sciaenocotyle panceri</i>	Two batches of meagre (av. wt. 8.3 g) were stocked in adjacent 4000 m ³ floating cages (93 442 fish stocked Aug 2005; 45 000 fish stocked July 2006) at 30 m depth off north-east Sardinia. In May 2007, both stocks were combined (av. wt. 750 and 200 g). In Sept 2007, the fish were lethargic, emaciated and anaemic, mortalities began thereafter. Prevalence of infection was 100% in both age classes, the older stock had a mean intensity of 367 flukes fish ⁻¹ and the younger stock 200 flukes fish ⁻¹ . Mortality in the older stock was 5–10% (i.e. loss of 4670–9340 fish; 3.5–7.0 t lost) and <2% in the younger stock (i.e. <500 fish; 100 kg). Loss is estimated on the value of Italian fish in 2005, i.e. US\$ 8.41 kg ⁻¹	29 435–58 870 (older year class) 841 (younger year class)	Merella <i>et al.</i> (2009)
	A Corsican multi-year class farm in the Gulf of Ajaccio consisting of 8 cages of meagre and 12 of sea bass reported heavy mortalities in Aug 2005 as a consequence of heavy burdens of gill monogeneans. The prevalence of infection on a sample ($n = 47$) of <300 g (6 months old) fish was 95.7% (mean abundance 58.7 ± 30.5 flukes fish ⁻¹ ; mean intensity 61.3 ± 28.4 flukes fish ⁻¹). Although heavy mortalities are reported, no details were provided. It is assumed that each cage measured 125 m ³ , stocked at a min. 15 kg m ³ and that total mortality across all 8 cages was 50%. Loss is estimated on the price of French harvest-sized fish in 2005, i.e. US\$ 8.72 kg ⁻¹	65 400	Ternengo and Katharios (2008); Ternengo <i>et al.</i> (2010)
(z) Atlantic cod, <i>Gadus morhua</i> L.			
<i>Loma morhua</i>	From a sample of 41 (40–52 cm long) cod acquired from a sea ranching facility in Newfoundland in July 1987, 78% had macroscopic xenomas on the gills. Of these, 26 (i.e. 63%) died in Sept 1987 when sea temperatures rose from 11 to 16 °C. Fish had been lethargic, were emaciated and at necropsy all 41 fish had pale gills and a massive infection of xenomas within the gills, heart, spleen and, occasionally, the inner body wall	–	Khan (2005)
	Following the loss of Newfoundland hatchery reared stock (av. 23 cm long; 2 years old) in Sept 2000, the post-mortem examination of 14 specimens revealed that 29% had macroscopic xenomas in the heart; all had microscopic xenomas on the gills and in their hearts. Further losses in Nov 2003, revealed that 21% (i.e. $n = 5$) of the specimens that were post-mortemed had gill and heart xenomas	–	Khan (2005)
	Major outbreaks reported at numerous aquaculture sites including land-based systems Throughout Newfoundland, New Brunswick and New Hampshire. No details provided	“100s of 1000s” –	O’Neill <i>et al.</i> (2011)
Pseudobranchial X-cell	Of a total of 1 795 000 wild cod juveniles (~ 2 g) caught in Isafjardardup fjord, NW Iceland (2002–2004), reared in land-based tanks to ~ 80–100 g in size, <1% of these fish had pseudotumours; however, a prevalence of 2–15% was found in emaciated/moribund fish. Levels in wild-caught juveniles caught in 1998–2000, by comparison, were higher with 7% of 6 months old (6.5–13 cm long), 23% of 22 months old (18.5–27 cm long) and 7% in 2+ year-old fish having pseudotumours	–	Eydal <i>et al.</i> (2010)
<i>Trichodina cooperi</i> + <i>T. murmanica</i>	Infections on wild fish, caught off Isafjordur, West Iceland and then grown on in shore-based tanks, were monitored over a 9-month period. Two cohorts of fish (~ 5 cm) were transferred in Sept of 2002 and 2003. Infections were low on each fish at the time of transfer. The prevalence of <i>T. cooperi</i> reached near 100% on both cohorts within 2 months (60% of the fish had mild infections, i.e. 1–10 parasites in a skin scrape measuring 1.5 cm ² ;	–	Kristmundsson <i>et al.</i> (2006)

	25% medium, i.e. 11–50 parasites; and, 15% heavy, i.e. > 50 parasites). After approx. 4 months post-transfer although, i.e. Jan/Feb, the numbers of <i>T. cooperi</i> began to reduce but the number of <i>T. murmanica</i> was observed to increase towards a peak prevalence of 95–100% with similar parasite burdens to those recorded for <i>T. cooperi</i> in Nov/Dec. The heaviest parasite burdens of <i>T. murmanica</i> , however, were observed in just one cohort in April when <5% of fish had no infection, over 35% had mild infections, 20% had medium infections, and, 40% had heavy infections. Mortalities were recorded but it is difficult to say whether these were due to parasite infections, stress following post-transfer or the subsequent cannibalistic tendencies of the stock		
<i>Trichodina murmanica</i>	A sharp rise in temperature from 6–8 °C to 15–16 °C as a result of defective cooling unit in a cod hatchery in Newfoundland, Canada in Aug 2002 was the stressor linked to the subsequent mortality of >100 000 juvenile cod (av. 4.3 cm body length) over a 4-week period. Each fish was determined to have an av. 62 parasites mm ⁻² . Loss is estimated on the value of each fish, i.e. US\$ 0.50	50 000	Khan (2004)
<i>Trypanosoma murmanensis</i>	By experimental transmission, it is demonstrated that infection can result in the mortality of 65% of 0+, 19% of 1+, 11% of 2+ and 7% of 3+ aged fish with most deaths occurring between 22 and 38 days post-infection	–	Khan (1985)
<i>Lernaeocera branchialis</i>	482 cod (32–76 cm TL) died within 5 days of their transfer from traps to sea cages in Newfoundland in July 1988. 44% of the mortalities were infected with <i>L. branchialis</i> . Assessment of the remaining 1500 cod, 1 month later revealed that 8.5% were infected. Loss is estimated using an av. weight of 1 kg fish ⁻¹ and the harvest price of Canadian cod in 1988, i.e. US\$ 7.17 kg ⁻¹	3456	Khan <i>et al.</i> (1990)
(aa) Lefteye flounders, <i>Psettina brevirectis</i> (Alcock, 1890)			
No records of parasitic infection resulting in the major loss of farmed stocks of this species could be found. There are some concerns surrounding the identity of this species from its common name as listed in the FAO FishStatJ database. It is also possible that Chinese left eye flounder refers to <i>Paralichthys olivaceus</i> , which is also known as the bastard halibut, the olive flounder or the Japanese flounder and not <i>Psettina brevirectis</i> as it is listed here			
(ab) Southern bluefin tuna, <i>Thunnus maccoyii</i> (Castelnau, 1872)			
<i>Uronema nigricans</i>	Infection, seen as parasitic encephalitis, resulted in the loss of between 5 and 10% of the wild-caught fish (~ 15–35 kg) that were fattened on in sea cages for a period of 3–8 months before harvesting. The annual harvest in 1993 was 636 t, assuming that the figures quoted were representative of the entire industry, then this represents a loss of between 31.8 and 63.6 t. Loss estimates are based on the value of harvested fish in Australia in 1993, i.e. US\$ 16.32 kg ⁻¹	518 976–1 037 952	Munday <i>et al.</i> (1997)
	Prior to 1993, the mortality and morbidity rates were suggested to be 5% in captive fish. These were reported to have fallen to 1.34% in 1995 and were <1% in 2001	–	Munday <i>et al.</i> (2003)
	During the 2003 season, total loss was 4%, i.e. 94.9 t valued at US\$ 22 450 t ⁻¹	2.13 M	Deveney <i>et al.</i> (2005)
	Infection was found with a prevalence of 58% in 31 morts and in 2 fish displaying signs of ‘swimmer syndrome’. Fish, weighing 25–50 kg, were from a commercial grow-out facility in Spencer Gulf, east of Port Lincoln, South Australia. Loss (0.5–1 t) is based on the value of Australian stock in 2005, i.e. US\$ 22.45 kg ⁻¹	11 225–22 450	Deveney <i>et al.</i> (2005)
<i>Caligus chiasos</i>	In May 2005, 4–6 weeks after transfer to cages in Spencer Gulf, Australia, 55% of the ranched stock had an av. 5.77 lice per infected fish. Lice activity over the head and eyes and subsequent net collisions as a consequence of irritation-induced flashing resulted in observable eye pathology on 32.5% of stock (av. eye scores were 3.5, i.e. extremely cloudy	–	Hayward <i>et al.</i> (2008, 2009)

Table 8. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
<i>Caligus chiastos</i> and <i>Cardicola forsteri</i>	to erosion of the cornea. Authors estimate the value of each fish at US\$ 500. Consequential losses unknown but are suggested to be significant Tuna (av. 22.5 kg) ranched in the Great Australian Bight looked at the relationship of both parasite species on tuna health for 12 weeks after transfer into and between cages. The <i>Caligus</i> infection was 100%, peaking at 265.8 lice fish ⁻¹ about 4 weeks post-transfer. Lice numbers were positively correlated with gross eye pathology scores and negatively correlated with condition factor. <i>Cardicola</i> infections, which were correlated with lice burdens, reached 100% (highest av. burden 268.3 flukes fish ⁻¹) within 4–6 weeks post-transfer, with epizootics peaking near the start of mortalities ~7 weeks after ranching. The cumulative mortality in the 2 cages were ~40% (<i>n</i> = 474 fish) and 47% (<i>n</i> = 89 fish). Loss in the 2 cages (i.e. 232 fish) is based on the harvest price of Australian stock in 2008, i.e. US\$ 16.37 kg ⁻¹	1.4 M	Hayward <i>et al.</i> (2010)
<i>Caligus elongatus</i>	Suggested that grazing activity of the parasite on captive reared stock may result in ocular damage	–	Rough <i>et al.</i> (1999); Munday <i>et al.</i> (2003)
<i>Cardicola forsteri</i>	The hearts of 84 ranched tuna (size of host and number infected not specified) farmed in sea cages were found to be off Port Lincoln, Australia, were found to be infected with an estimated 19 000 to 1.7 M eggs per heart. Infection resulted in hypertrophy of the ventricle spongiosa. Eggs were also found within the afferent filamentary arteries and lamellae vasculature. The study concluded that infections do not appear to cause mortality A further study of 210 tuna (19.9–31.7 kg) from net pens around Port Lincoln, Australia found that infections peaked 2 months after transfer (May 2004) with a prevalence of 100% and an av. 27 flukes fish ⁻¹ (range 0–99). Infections declined thereafter suggesting that fish are able to control parasite numbers	–	Colquitt <i>et al.</i> (2001) Aiken <i>et al.</i> (2006)
(ac) Mozambique tilapia, <i>Oreochromis mossambicus</i> (Peters, 1852) and Nile tilapia, <i>Oreochromis niloticus</i> L. <i>Amyloodinium ocellatum</i>	An infection in Salton Sea, a 980 km ² hypersaline (i.e. 46 ppt) water body in California, USA throughout May 1997 to Nov 1998 resulted in the mass mortality of juvenile <i>O. niloticus</i> (1.0–13.0 cm length). Infection reached 100% in June/July of each year as water temperatures approached 40 °C. Massive numbers of heavily infected dead fish were noted in shallow waters. The tilapia in Salton Sea are exotic, details relating to the nature and purpose of their introduction are unknown and although this event is not linked to identifiable aquaculture activities, given the global significance of tilapia culture, this event is worth reporting. Loss is estimated on the volume of Salton Sea (9 300 000 dam ³), a tilapia biomass of 11 kg h ⁻¹ reported in 2000 (Riedel <i>et al.</i> 2002), a moderate (20%) and high (50%) level of mortality, and, the value of harvest-sized American stock in 2000, i.e. US\$ 3.31 kg ⁻¹	6.77 M (20% mortality) 16.93 M (50% mortality)	Kuperman and Matey (1999)
<i>Caligus epidemicus</i>	Nile tilapia stock held in ponds in Iloilo (av. 8.6 cm, av. 15.0 g) and in Quezon (av.	–	Natividad <i>et al.</i> (1986)

	11.4 cm, av. 29.1 g), Philippines were found to be heavily infected (prev. 100%, av. 111.7 copepods fish ⁻¹ , range 7–548 on Iloilo stock; prev. 92%, av. 34.3 copepods fish ⁻¹ , range 1–138 on Quezon stock)		
	Infected Mozambique tilapia stock reared in saltwater ponds in Taiwan resulted in mortalities. No details regarding the specific loss of fish are provided	–	Lin and Ho (1993)
<i>Neobenedenia melleni</i> ^b (syn. <i>girellae</i>)	Mozambique tilapia stock (~ 18 cm TL) held in 1 m ³ floating sea cages in Kaneohe Bay Hawaii (1981–1984) were infected with, in certain cases, >400 flukes fish ⁻¹ over the entire body surface and eyes resulting in multi-focal petechial haemorrhages, scale and skin damage, buphthalmos, corneal, ulceration scarring and blindness. Exact losses not specified but infection was sufficiently severe that the patterns of mortality and morbidity impeded preliminary breeding cycles. In estimating loss, it is assumed that a 50% mortality occurred in at least 4 cages, stocked at 15 kg m ³ , and the harvest price of American tilapia in 1985 (first figures available), i.e. US\$ 14.00 kg ⁻¹	420	Kaneko <i>et al.</i> (1988)
(ad) Mangrove red snapper, <i>Lutjanus argentimaculatus</i> (Forsskål, 1775)			
<i>Amyloodinium ocellatum</i>	An infection resulted in 2-month old hatchery stock (av. wt. 0.12 g; 19 mm long) reared in seven 5 t cement tanks in Iloilo, the Philippines in June 2003. Final mortality was 100%, infected fish had an av. 80–100 trophonts on their skin and fins and 40–50 trophonts on their gills. Loss is estimated on a stocking density of 10 kg m ⁻³ and a value of US\$ 0.01 fish ⁻¹ , i.e. the loss of 2.9 million fry	29 000	Cruz-Lacierda <i>et al.</i> (2004)
<i>Caligus epidemicus</i> §	A total of 16 site visits to pond and cage sites in Guimaras and Iloilo, Philippines were made between Dec 1999 and Aug 2003. Stock typically had mixed infections, however, only <i>C. epidemicus</i> was found on 2 visits (<i>n</i> = 14 fish) with prevalences between 2.0 and 12.5 lice fish ⁻¹ (range 1–4 lice per infected host). No details regarding the impact of infections on the host were provided	–	Ho <i>et al.</i> (2004)
<i>Caligus epidemicus</i> + <i>Caligus quadratus</i>	Mixed infections were found on stock (<i>n</i> = 94) during 8 of the 16 visits detailed above (§) The prevalence of infection ranged from 53 to 100% (av. 90.3%), mean intensities 2.5–14.5 lice fish ⁻¹ (av. 6.3 lice fish ⁻¹), range 1–20 lice per infected host	– –	Ho <i>et al.</i> (2004) –
<i>Caligus epidemicus</i> + <i>Caligus quadratus</i> + <i>Pseudocaligus uniartus</i>	A mixed infection was found only once during the study detailed above (§). The prevalence of infection was 100% (<i>n</i> = 15 fish), with a mean intensity of 3 lice fish ⁻¹ (range 1–5)	–	Ho <i>et al.</i> (2004)
<i>Caligus quadratus</i> + <i>Pseudocaligus uniartus</i>	A single mixed infection with a prevalence of 100% (<i>n</i> = 10 fish; mean intensity 16 lice fish ⁻¹ , range 9–39) was found during the study detailed above (§)	–	Ho <i>et al.</i> (2004)
<i>Caligus quadratus</i>	During the study above (§), single infections were found on 3 occasions with the prevalence of infection ranging from 53 to 95% (av. 76%), with mean intensities of between 1.5 and 4.5 lice fish ⁻¹ (av. 2.8 lice fish ⁻¹), range 1–8 lice per infected host	–	Ho <i>et al.</i> (2004)

^a Encompasses a complex of caligid species.

^b There is an ongoing debate as to whether *Neobenedenia girellae* and *N. melleni* represent separate discrete species or should be accepted as *N. melleni*.

^c Given the importance of salmonids in global marine aquaculture Chum and pink salmon are included here.

Table 9. The estimated economic cost of parasitic infections on the world's leading mollusc production industries. For each parasite-induced event, brief details are provided and an estimate of loss, where possible, has been calculated

Parasite	Impact	Estimated loss (US\$)	Reference
(a) Pacific cupped oyster, <i>Crassostrea gigas</i> (Thunberg, 1793)			
<i>Haplosporidium</i> sp.	Isolated occurrences and low levels of infection in California State, Washington State, France and Matsushima Bay, Japan. Low-level infection in 1989–1990 (10% of seed brought to California from Japan were infected). Species unknown, but impacts may be similar to <i>H. nelsoni</i> and <i>H. costale</i> . Sporocysts break down epithelial cells of digestive system resulting in some tissue necrosis. More resistant to infection than <i>Crassostrea virginica</i> , but based on impacts of <i>Haplosporidium</i> sp. on <i>C. virginica</i> , and the exhibition of hemic response similar to that in <i>C. virginica</i> as a result of early systemic infections, authors conclude that this is likely to have negative pathogenic impacts	–	Pereya, (1962); Friedman <i>et al.</i> (1991); Bower <i>et al.</i> (1994); Kamaishi and Yoshinaga (2002); Dégremont <i>et al.</i> (2010)
<i>Marteilia</i> (syn. <i>Marteilioides</i>) <i>chungmuensis</i>	Ovarian protozoan parasite causing nodule-like structures on the gonads and/or enlarged gonads and disfiguring. Substantial economic losses in Korean and Japanese aquaculture as infected oysters cannot be sold. Prevalence increases in summer and decreases autumn–spring. 76% mortality in experimental infection	–	Bower <i>et al.</i> (1994); Itoh <i>et al.</i> (2004); Tun <i>et al.</i> (2008)
	Losses in Okayama Prefecture, Japan in ~2000 estimated at ¥ 2–300 M and later updated in 2009 (?) to an estimated € 0.5 M	2–3 M (2000) 680 000 (2009)	Itoh <i>et al.</i> (2002); Itoh (2009)
<i>Mikrocytos mackini</i> (mikrocytosis or Denman Island disease)	Protistan parasite of unknown taxonomic affiliation. Diseased oysters characterized by green pustules making them unmarketable. Mortality up to 53% but severity fluctuates annually. Infection thought to be via the digestive tract and gills, with subsequent colonization of other tissues. Requires long periods at low temperatures (<10 °C) to cause disease; mostly expressed during the spring in more northerly locations. First reported in 1960, when 17–35% of the oysters in Henry Bay, Denman Island British Columbia died. Experiments indicate mortality occurs approx. 18 weeks after infection. Known range between southern British Columbia to Washington State. Severe infections confined to older oysters (up to 30% at low tide levels). No figures of loss (value or tonnage) are available so loss is est. at 10% of current production which in 2012 was 7165 t at US\$ 1425 t ⁻¹	1.02 M	Quayle (1961, 1982); Farley <i>et al.</i> (1988); Elston (1993); Bower <i>et al.</i> (1994); Hervio <i>et al.</i> (1996); Hine <i>et al.</i> (2001); Bower <i>et al.</i> (2005)
<i>Perkinsus marinus</i>	Widely distributed protistan parasite causing high mortalities in oyster populations. Under intensive cultural conditions, the mortality rate in larval oysters exceeded 90% in Washington State, USA. Occurs at water temperatures of 20–30 °C and salinities of 13–28 ppt. Highest mortalities (up to 95%) occur in the summer months on the East coast of America from Massachusetts to Brazil. American production, for example, in 1987 was 40 449 t, assuming this level of output would have been maintained in the absence of the parasite and that <i>Perkinsus</i> is the only factor responsible for loss, then approx. 2180 t worth an est. US\$ 14.21 M was lost between 1988 and 1992. Production in 1993 had returned to close to the 1987 level at 39 053 t	14.21 M (1988–1992)	Leibovitz <i>et al.</i> (1978); Elston (1980); Andrews (1984, 1988); Perkins (1993); Bower <i>et al.</i> (1994); Burreson (1996); Oliver <i>et al.</i> (1998); La Peyre <i>et al.</i> (2003); Dégremont <i>et al.</i> (2010)

<i>Protista incertae sedis</i> (Humboldt egg parasite)	First observed in 1966 in <i>C. gigas</i> in Humboldt Bay, California. Occurs in the cytoplasm of maturing ova. Superficially resembles <i>Steinhausia</i> , <i>Ovicola</i> and <i>C. mytilovum</i> . Infection in females variable, from <1–77%. Heavily infected females show necrosis in ova and an inflammatory response, possibly reducing reproductive potential. There was no indication, however, that infection kills the mature host. Also recorded in Korea, Northern Territory Australia and Marennes France	–	Becker and Pauley (1968a, b); Bower <i>et al.</i> (1994)
<i>Cliona</i> spp.	Clionid sponges are distributed globally and cause damage by boring through the shell, and if the shell cannot be repaired quickly enough, may weaken the adductor muscle and death may eventually occur. Can affect other oysters, scallops and mussels growing on bottom substrates	–	Bower <i>et al.</i> (1994)
<i>Gymnophalloides tokiensis</i>	Trematode infecting <i>C. gigas</i> and <i>C. virginica</i> in Japan, but other unconfirmed species of same genus recorded in British Columbia and Atlantic Canada. Where infection intensity is high, oysters have a high water content, reduced glycogen, protein and fat, lower growth and survival rates	–	Bower <i>et al.</i> (1994)
<i>Polydora</i> sp. (Spinonidae)	Shell-boring polychaete occurring globally. Causes blistering and shell weakness. Induces oxidative stress, reduces growth and often results in mortality. <i>Polydora haswelli</i> and <i>P. aura</i> found abundantly in cultured <i>C. gigas</i> throughout Asia, and <i>P. uncinata</i> infects <i>C. gigas</i> cultured in Southeastern Korea	–	Chambon <i>et al.</i> (2007); Sato-Okoshi <i>et al.</i> (2012, 2013)
<i>Mytilicola orientalis</i>	Introduced to Pacific coast of the USA and Europe from <i>C. gigas</i> transported from Japan. Considered a serious pest of cultured bivalves. Causes damage to digestive tract epithelium (where it attaches)	–	Elston (1993); Torchin <i>et al.</i> (2002); Kim and Sato (2010)
<i>Myicola ostreae</i>	Severe gill erosion. Found in Europe, eastern and western USA and British Columbia	–	Lauckner (1983); Bower <i>et al.</i> (1994)
(b) Japanese carpet shell, <i>Ruditapes philippinarum</i> (Adams et Reeve, 1850)			
<i>Perkinsus atlanticus</i>	Pathogenic protist, synonymous with <i>P. olseni</i> , that infects <i>Ruditapes</i> sp. In <i>R. decussatus</i> , causes formation of cysts on gills, foot and mantle when infections are high intensity, and associated high mortalities. Laboratory experiments show mortality from heavy infections start to occurs after 4 weeks. Parasitized clams documented to be more susceptible to opportunistic infections. Distributed in the Mediterranean Sea and NW Spain	–	DaRos and Canzonier, (1985); Azevedo (1989); Bower <i>et al.</i> (1994); Almeida <i>et al.</i> (1999); Lee <i>et al.</i> (2001); Montes <i>et al.</i> (2001); Shimokawa <i>et al.</i> (2010)
<i>Perkinsus marinus</i>	Thought to be responsible for decline in harvest of <i>R. philippinarum</i> in Korea since 1990s due to annual mass mortality events. Effects on host include tissue atrophy, reduced energy and growth, reduced reproductive potential, organ necrosis and often mortality (up to 100% when infection is high). Appears to favour high salinity and water temperature. Korean production in 1990 was near its peak at 61 713 t, assuming that the subsequent decline in production can be attributed only to <i>Perkinsus</i> and that year on year tonnage could have been maintained, then it can be est. that across the period 1991–2012, <i>Perkinsus</i> was responsible for the loss of 900 744 t (av. 40 943 t p.a.) worth approx. US\$ 1460 M (av. US\$ 66·385 M p.a.)	1460 M (1991–2014) 66·385 M p.a.	Park and Choi (2001); Ngo and Choi (2004); Park <i>et al.</i> (2005)
<i>Ostrincola koe</i>	Parasitic copepod infecting mantle cavity, causing widespread damage and mortality. Has been responsible for mass mortality events in other cultured molluscs (e.g. <i>Meretrix meretrix</i>)	–	Kim (2004)
<i>Cercaria tapidis</i>	Bucephalid trematode commonly infecting cultured clams in Korea and Japan. High levels of infection causes reduced growth and degeneration or castration of gonads. No obvious seasonality, but prevalence of infection higher in autumn	–	Ngo and Choi (2004)

Table 9. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
(c) Yesso scallop, <i>Patinopecten yessoensis</i> (Jay, 1857)			
<i>Perkinsus qugwadi</i>	Caused significant mortality (up to 98%) in juveniles (< 5 cm shell height, <1-year old) and adults (up to 60%) between 1988 and 1995 in British Columbia, Canada. May be native to British Columbia; not found elsewhere. Pathogenic at low temperatures (8–15 °C). Formerly known as Scallop Protozoan X (SPX). Assuming that infection caused a 20% loss in production, then across the period 1988–1995, an est. 99 t were lost valued at US\$ 92 605	92 605	Bower <i>et al.</i> (1992, 1994, 1998); Blackburn <i>et al.</i> (1998); Itoh <i>et al.</i> (2013)
<i>Cliona</i> sp.	Globally distributed, shell-boring sponges that burrow under shells of bivalves. Scallops able to repair damage; however, if adductor muscle is damaged, impaired feeding may result causing mortality	–	Bower <i>et al.</i> (1994); Lauckner (1983); McGladdery <i>et al.</i> (2006)
<i>Dipolydora alborectalis</i> (Spionidae)	Shell-boring polychaete (Spionidae) that bores visibly into the shells of <i>P. yessoensis</i> ; often highly abundant in Japanese cultures. Causes blistering, shell weakness, reduces growth and often results in mortality	–	Sato-Okoshi <i>et al.</i> (2012)
<i>Pectenophilus ornatus</i>	Highly modified parasitic copepod with brood-pouch that infects gills. Reduces market condition of scallops and thought to infect 100% of Japan's scallop stock. Prevalence also increases with host size. Found to be highly transformed member of the family Mytilicolidae. Loss is est. on a 5–10% downgrading in the value of Japanese stock which in 2012 was 184 287 t worth an est. US\$ 503·76 M	28–56 M	Nagasawa <i>et al.</i> (1988); Nagasawa and Nagata (1992); Bower <i>et al.</i> (1994); Huys <i>et al.</i> (2006); Suzuki and Matsutani (2009)
<i>Polydora brevipalpa</i> (Spionidae)	Highly prevalent, shell-boring polychaete causing large shell defects and some mortality in eastern China and on the west coast of Japan. Host-specific to <i>P. yessoensis</i>	–	Mori <i>et al.</i> (1985); Sato-Okoshi <i>et al.</i> (2012)
<i>Polydora websteri</i> (Spionidae)	Shell-boring polychaete causing up to 84% mortality in British Columbia's scallop grow-out sites from 1989 to 1990. According to FishStatJ, production on the Pacific coast fell from 10 t (US\$ 8565 t ⁻¹) in 1989 to 0 in 1990. If, however, the 10 t in 1989 represents the 16% of viable stock that was sold, then this suggests that 52·5 t of production was lost	85 650 449 660	Bower <i>et al.</i> (1992)
(d) Chilean mussel, <i>Mytilus chilensis</i> (Hupé, 1854) syn. <i>Mytilus edulis</i> (see Gray <i>et al.</i> 1999) – see the entry below			
(e) Abalone, <i>Haliotis</i> spp.			
<i>Candidatus Xenohaliotis californiensis</i> (Rickettsiae)	Withering syndrome appeared in Californian <i>H. cracherodii</i> Leach, 1814 after 1983 El Niño event. Caused high mortality in <i>H. rufescens</i> Swainson, 1822 (90% prevalence) and <i>H. fulgens</i> Philippi, 1845 (37·5% prevalence) in southern Californian and Mexican farms during El Niño event of 1997. Also causes high mortality in <i>H. iris</i> Gmelin, 1791 in New Zealand. Associated with elevated water temperatures >20 °C. Chronic wastage of foot and visceral tissues reduces ability to adhere to substrata. Impairment of digestion due to high infection of pathogen in gut	–	Haaker <i>et al.</i> (1992); Lafferty and Kuris (1993); Kismohandaka <i>et al.</i> (1995); Friedman <i>et al.</i> (1997); Antonio <i>et al.</i> (2000); Cáceres Martínez <i>et al.</i> (2000); Friedman <i>et al.</i> (2000); Moore <i>et al.</i> (2000, 2002, 2009); Diggles <i>et al.</i> (2002); Hine <i>et al.</i> (2002); Braid <i>et al.</i> (2005)
Paua haplosporidiosis	Unknown haplosporidian causing paua (abalone) haplosporidiosis. Causes lethargy, lack of orientation and surface adhesion and chronic mortality of juveniles (av. shell length	–	Diggles <i>et al.</i> (2002); Diggles and Oliver (2005)

	14 mm) in summer months (up to 80–90% in one farm hatchery located on the South Island of New Zealand)		
<i>Labyrinthuloides haliotidis</i>	Protozoan parasite-infecting hatcheries on the west coast of Canada. Destroys head and foot of juvenile abalone. Resulting mortality may be up to 100%. Infection contributed to the closure of the only commercial abalone aquaculture unit	–	Bower (1987a, b); Bower <i>et al.</i> (1994); Bower and Meyer (2005)
<i>Perkinsus olseni</i>	Pathogenic protist, synonymous with <i>P. atlanticus</i> . Spreads rapidly although tissues; thought to cause mortalities in abalone in the Gulf of St. Vincent, Australia	–	Almeida <i>et al.</i> (1999); Bower <i>et al.</i> (1994); Goggin and Lester (1995)
Spionid mudworms (Spionidae) (<i>Boccardia knoxi</i> , <i>Polydora hoplura</i> , <i>Polydora haswelli</i> , <i>Terebrasabella heterouncinata</i> and <i>Boccardia proboscidea</i>)	Shell-boring polychaetes causing blistering of shell. <i>Boccardia knoxi</i> and <i>P. hoplura</i> caused 50%+ mortality in abalone farms in Tasmania and S. Australia in 1995–2000. In South Africa, <i>B. proboscidea</i> , <i>P. hoplura</i> and <i>T. heteruncinata</i> cause high levels of infestation leading to decreased growth, decreased flesh condition and increased mortality in cultured <i>H. midae</i> L. Heavy infestations in cultured <i>H. discus</i> Reeve, 1846 in Asian waters cause blistering, shell weakness, reduced growth and often mortality. Common parasites of <i>H. rufescens</i> in California; increased mortality due to heavy infections or subsequent predation. Only data for blacklip abalone in 1999 (21 t) and 2000 (40 t) are available in FishStatJ. Assuming that only 50% of surviving stock was harvested this suggests that approx. 60 t were lost worth an est. US\$ 26 302 t ⁻¹ (1999) to US\$ 29 117 t ⁻¹	0.55–1.16 M p.a.	Lleonart <i>et al.</i> (2003); Simon <i>et al.</i> (2006, 2010); Sato-Okoshi <i>et al.</i> (2012, 2013); Maguire and Rogers-Bennett (2013)
<i>Evalea tenuisculpta</i>	Pyramidellid snail infecting <i>H. rufescens</i> at certain sites along coast of California where up to 82% have been infected. Although little is known about effects of infection in this host, they can cause shell damage, reduced growth rates, transmission of bacterial disease and mortality	–	Maguire and Rogers-Bennett (2013)
(f) Chinese razor clam, <i>Sinonovacula constricta</i> (Lamarck, 1818)			
<i>Monorchis xiamenensis</i>	Metacercaria of this trematode worm infect both juvenile and mature clams, causing destruction of organs including gonads, digestive gland, gills and mantle	–	Lei (2000)
<i>Vesicocodium solenophagum</i>	Trematode worm completing its life cycle in the razor clam (after eggs have been carried and passed out by gobiid fish). Infects number of tissues including reproductive organs causing serious damage and mortality to second-year clams. Infection seen in first-year clams during July and Aug. Increases in April–June when host damage becomes more serious. Known as ‘black root disease’	–	Chungti and Zhenzu (1979); Shi and Wang (2001)
(g) Peruvian scallop, <i>Argopecten purpuratus</i> (Lamarck, 1819)	No mass mortality events recorded for scallops in Chile and very little information available (Lohrmann, 2009). All known parasites for <i>A. purpuratus</i> have benign effects on the host		
(h) Blood cockle, <i>Anadara granosa</i> L.			
<i>Nematopsis</i> sp.	Oocysts parasitize connective tissue of gills. Heavy infections thought to reduce filtering efficiency and weaken condition in juveniles resulting in loss of culture. Poor environmental conditions may increase vulnerability to infection; several mass mortality	–	Pookasawan <i>et al.</i> (1982); Tuntiwaranuruk <i>et al.</i> (2004); Uddin <i>et al.</i> (2011)

Table 9. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
Digenetic trematode	events of <i>A. granosa</i> in Phuket, Thailand. Prevalence of <i>Nematopsis</i> sp. is dependent on season and environmental conditions (e.g. salinity) Sporocysts infect gonads, mantle, stomach epithelium, digestive glands and foot. Heavy infection destroys the gonads and reduces glycogen content. Recorded in stocks in Malaysia (Straits of Malacca). No clear seasonality in prevalence of infection, but appears to favour more saline conditions	–	Sayuthi (1993); Uddin <i>et al.</i> (2011)
(i) Mediterranean mussel, <i>Mytilus galloprovincialis</i> (Lamarck, 1819)			
<i>Marteilia refringens</i> (Marteliosis/Aber disease)	<i>M. refringens</i> causes mortality in cultured mussels in Galicia, NW Spain. Also previously observed in Europe in <i>Ostrea edulis</i> , <i>Mytilus edulis</i> and <i>Crassostrea gigas</i> . High levels of parasitism observed in the Thermaikos Gulf (Greece). Extensive mortality in oysters cultured in France and Spain in 1960s–1970s. Mortalities occur from May onwards and peak in June, July and Aug. Causes lesions in digestive tubules, which results in wasting of digestive gland and eventual death. Inhibits gonadal development in <i>M. galloprovincialis</i> and reduced total carbohydrate levels. Mortality often between 50 and 90%. Long-term infections may result in 100% mortality. Assuming that the av. prevalence of infection (12.06 ± 12.06) across 5 sites in Galicia, NW Spain given in Villalba <i>et al.</i> (1997) resulted in mortality, then the loss of production applied to the tonnage in 1988 (i.e. 243 010 t) equates to a loss of 29 307 t worth an est. US\$19.05 M	19.05 M	Lodeiros <i>et al.</i> (1987); Figueras <i>et al.</i> (1991); Villalba (1993); Villalba <i>et al.</i> (1993); Bower <i>et al.</i> (1994); Robledo and Figueras (1995); Robledo <i>et al.</i> (1995); Villalba <i>et al.</i> (1997); Thébault <i>et al.</i> (1999); Zrnčić <i>et al.</i> (2001); Balseiro <i>et al.</i> (2007); Karagiannis and Angelidis (2007); Carella <i>et al.</i> (2010)
	Assuming that the 1988 levels of Spanish production could have been maintained in the absence of parasitic infection and that parasites were the sole cause of loss, then the subsequent drop in production over the period of 1989–1997 equates to a loss of 692 977 t worth an est. US\$ 387.56 M	387.56 M	
<i>Mytilicola intestinalis</i>	Parasitic copepod. Endemic along inshore European coasts. Infection of adults occurs in gut where eggs are laid, hatched and are expelled. Nauplii develop through 2 pelagic stages before infecting new mussels. Mortality occurs when infection is high; thought to be associated with mass mortalities. Causes overall reduction of condition (e.g. lower total carbohydrate levels), which affects the quality of meat in marketable mussels	–	Korringa (1951); Lauckner (1983); Andrews (1984); Dethlefsen (1985); Davey and Gee (1988); Davey (1989); Blateau <i>et al.</i> (1992); Bower <i>et al.</i> (1994); Robledo <i>et al.</i> (1994a, b); Villalba <i>et al.</i> (1997); Buck <i>et al.</i> (2005); Pogoda <i>et al.</i> (2012)
<i>Proctoeces maculatus</i> (syn. <i>Cercaria temuans</i>)	Trematode worm causing a number of symptoms including valve weakness and difficulty attaching to substrates. Main impact is atrophy of reproductive organs and inhibited gametogenesis. Widespread distribution in temperate and tropical waters. Thought to be responsible for mass mortality event of mussels in southern part of Laguna Veneta, Italy	–	Munford <i>et al.</i> (1981); Lauckner (1983); Figueras <i>et al.</i> (1991); Bower <i>et al.</i> (1994); Robledo <i>et al.</i> (1994a, b); Villalba <i>et al.</i> (1997)
	Assuming that the av. prevalence of infection (0.68 ± 0.66) across 5 sites in Galicia, NW Spain given in Villalba <i>et al.</i> (1997) resulted in mortality, then the loss of production applied to the tonnage in 1988 (i.e. 243 010 t) equates to a loss of 1652 t worth an est. US\$ 1.07 M	1.07 M	

(j) Atlantic mussel, <i>Mytilus edulis</i> L.			
<i>Coccidia</i> sp.	Protozoan parasite infecting and damaging kidneys. Mortalities occur when infection is heavy in mussels grown in artificial conditions	–	Bower (1992); Bower <i>et al.</i> (1994)
<i>Marteilia refringens</i>	Causes mortality in cultured mussels in Galicia, NW Spain. Also previously observed in Europe in <i>Ostrea edulis</i> , <i>Mytilus edulis</i> and <i>Crassostrea gigas</i> . Extensive mortality in oysters cultured in France and Spain in 1960–1970. Mortalities occur from May onwards and peak in June, July and August. Causes lesions in digestive tubules, which results in wasting of digestive gland and eventual death. Mortality often 50–90% and long-term infections may result in 100% mortality. Synonymous with <i>Marteilia maurini</i>	–	Villalba (1993); Villalba <i>et al.</i> (1993); Bower <i>et al.</i> (1994); Robledo and Figueras (1995); Thébault <i>et al.</i> (1999); Zrnčić <i>et al.</i> (2001); Balseiro <i>et al.</i> (2007); Carella <i>et al.</i> (2010)
<i>Cliona</i> spp.	Globally distributed sponges causing damage by boring through shell and, if the shell cannot be repaired quickly enough, may weaken the adductor muscle and death may eventually occur. Infection has been recorded for <i>M. edulis</i> on bottom substrates in Scandinavia and North America	–	Lauckner (1983); Bower <i>et al.</i> (1994)
<i>Mytilicola intestinalis</i>	Parasitic copepod found in digestive tract of several molluscs. Endemic along inshore European coasts from Denmark to Italy. Infection of adults occurs in gut where eggs are laid, hatched and are expelled. Nauplii develop through 2 pelagic stages before infecting new mussels. Mortality of mussels occurs when infection is high (5–10 parasites per mussel); thought to be associated with mass mortalities. Causes reduction in feeding rate resulting in overall reduction of condition, which affects quality of meat in marketable mussels	–	Korringa (1951); Lauckner (1983); Andrews (1984); Dethlefsen (1985); Davey and Gee (1988); Davey (1989); Blateau <i>et al.</i> (1992); Elston, (1993); Bower <i>et al.</i> (1994); Buck <i>et al.</i> (2005); Pogoda <i>et al.</i> (2012)
<i>Polydora ciliata</i>	Shell-boring polychaete occurring globally. Causes blistering, weakens and deforms shell, reducing overall condition. In Europe, <i>P. ciliata</i> is associated with reduced market quality and mortalities	–	Kent (1979, 1981); Bower <i>et al.</i> (1994); Buck <i>et al.</i> (2005)
<i>Proctoeces maculatus</i>	Trematode worm causing a number of symptoms including valve weakness and difficulty attaching to substrates. Heavy infections reduce glycogen levels and circulation, and gametogenesis is either impaired or completely stopped. Some cases report associated mortalities. Widespread distribution in temperate and tropical waters; seasonal prevalence with epizootic episodes in autumn and winter	–	Lauckner (1983); Bower <i>et al.</i> (1994); Sunila <i>et al.</i> (2004)
<i>Edotia doellojuradoi</i>	Isopod infecting cultured mussels in Chile and causing gill damage to host. Damage increases with parasite abundance	–	Valencia and George-Nascimento (2013)
(k) New Zealand green-lipped mussel, <i>Perna canalicula</i> (Gmelin, 1791) (syn. <i>Perna canaliculus</i>)			
Apicomplexan parasite X (APX)	Only occurs in NZ. Affects <i>P. canalicula</i> in farms in Marlborough Sound. More common to <i>Ostrea chilensis</i> . Heavy infections cause chronic wastage and reduction of gametes and eventually death	–	Diggles <i>et al.</i> (2002)
<i>Nepinmotheres novaezelandiae</i>	Parasitic pea crab thought to be symbiotic with host; however, recent research has identified detrimental impacts including 30% reduction in shell height and wet mass and erosion of the gills, causing reduced rates of oxygen consumption	Annual loss of US\$ 2·16 M	Trottier and Jeffs (2012); Trottier <i>et al.</i> (2012)
(l) Pearl oyster, <i>Pinctada</i> spp.			
Unknown parasite	Mass mortalities in 1985; 50–80% of cultured <i>P. margaritifera</i> L. in coastal lagoons in French Polynesia. Symptoms include mantle lesions, lack of growth and excessive secretion of mucus. Assuming that in the absence of the parasite production would have increased by 0·21–0·84 t, then loss can be est. at US\$ 8·50–34·02 M	8·50–34·02 M	Cabral (1989a, b)

Table 9. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
Unknown protozoan parasite	Caused high mortalities in cultures of <i>P. margaritifera</i> in the Red Sea in 1969 and 1973	–	Nasr (1982); Cabral (1989b)
<i>Bucephalus</i> sp.	Trematode affecting the formation of pearls. In <i>P. radiata</i> (Leach, 1814), <i>Bucephalus</i> has been reported to destroy female gonads, and overall infections can reduce ability to withstand environmental stress	–	Sakaguchi (1968a, b); Bower <i>et al.</i> (1994); Lee <i>et al.</i> (2001)
Shell-boring worms and sponges	Examples of recorded infections include <i>Polydora vulgaris</i> in <i>P. margaritifera</i> in Kuwait, and <i>Polydora ciliata</i> and <i>Cliona margaritiferae</i> in <i>P. fucata</i> (Gould, 1850) in Sri Lankan pearl banks. Shell-boring parasites are huge problem in pearl oyster culture worldwide, causing damage, reduction in condition and, in some cases, mortality	–	Velayudhan (1982); Bower <i>et al.</i> (1994)
(m) Asiatic hard clam, <i>Meretrix</i> spp.			
<i>Ostrincola koe</i>	Parasitic copepod responsible for mass mortality of <i>Meretrix meretrix</i> L. in 1988–1989 in Southern Jiansu, China. Infects mantle cavity of the clam	–	Ho and Zheng (1994)
<i>Peritrichous</i> ciliate <i>Myxosporozoa</i> <i>Epicomplexa</i>	Protozoan parasite found in diseased clams with mantle hypertrophy disease'. Symptoms include fleshy, swollen mantles, degraded tissues and abnormally increased mucus production	–	Ren <i>et al.</i> (2005)
(n) Sea snails <i>Babylonia/Hemifusus</i> spp.			
<i>Haplosporidium babyloniae</i>	Occurs on <i>Babylonia areolata</i> (Link, 1907) (ivory shell snail) in southern coastal areas of China. Most organs become badly damaged and develop serious lesions as a result of infection	–	Peng <i>et al.</i> (2011)
(o) Asiatic clam, <i>Corbicula fluminea</i> (Müller, 1774)			
Most of the literature describing <i>Corbicula fluminea</i> focuses on its status in many countries as an invasive non-native species and its function as a vector for parasites that impact other wild and cultured species and in humans (Williams Jr. <i>et al.</i> 2001; Graczyk <i>et al.</i> 2003; Sousa <i>et al.</i> 2008a, b, c; Karatayev <i>et al.</i> 2012). Mass mortality events have been recorded for this species; however, these have been attributed to environmental conditions (Werner and Rothhaupt, 2008; Vohmann <i>et al.</i> 2010); the authors have been unable to find any recorded economic impacts of protozoan or metazoan parasites for this species			
(p) Hard clam, <i>Mercenaria mercenaria</i> L.			
Quahog parasite X (QPX) (Phylum: Labyrinthulomycota)	Undescribed parasite but from family Thraustochytridae. Documented in hatcheries in Gulf of St. Lawrence, Canada, Virginia and Massachusetts. Mortalities of 80–90% recorded for juvenile hatchery stock and up to 92% for seed transported from South Carolina to New Jersey during 1995–1998. Infects mantle, gills, foot and connective tissues (and other organs) eventually causing necrosis, decomposition of tissue by bacteria/fungus and mortality; highest prevalence occurs in November and May. Temperature thought to be a determining factor in prevalence of QPX	–	Bower <i>et al.</i> (1994); Calvo <i>et al.</i> (1998); Ragan <i>et al.</i> (2000); Ford <i>et al.</i> (2002); Dahl <i>et al.</i> (2010); Perrigault <i>et al.</i> (2009, 2011); Hégaret <i>et al.</i> (2010)
QPX-like parasite	Observed in 2 sites of <i>M. mercenaria</i> culture in Massachusetts in 1995. Infected clams suffered poor condition and reduced growth followed by mass mortalities in stock 1.5–2 years old	–	Smolowitz <i>et al.</i> (1998)

(q) American cupped oyster, <i>Crassostrea virginica</i> (Gmelin, 1791) <i>Haplosporidium costale</i> (SSO disease) (syn. <i>Minchinia costalis</i>)	Found on East Coast of North America from Chesapeake Bay, Virginia to Nova Scotia. Infections restricted to high salinity (≥ 25 ppt) occur in autumn and remain histologically undetectable until following spring when mortalities occur (sharp increases May–June). Low prevalence but 20–75% mortality in affected populations. Regular sporulation throughout connective tissues causing prompt oyster mortality. Infection site usually epithelia of digestive tract; causes damage to digestive epithelium and affects gonad development	–	Meyers (1981); Andrews (1982, 1984, 1988); Bower <i>et al.</i> (1994); Dégremont <i>et al.</i> (2010); Ford (2011)
<i>Haplosporidium nelsoni</i> (MSX disease)	Occurs on mid-Atlantic Coast of USA: Massachusetts to South Carolina and Florida. Less prevalent in low-salinity water (<18‰), but enzootic in high salinity (>20‰). >90% mortality occurred in Delaware and Chesapeake Bay 1957–1959. During years of drought (1981–1982), <i>H. nelsoni</i> invaded areas of low salinity (where oysters are mostly grown). Life cycle largely unknown. Oysters acquire new infections in spring and mortalities occur in summer. Deaths in infected oysters sometimes delayed by temperature (e.g. late summer infections cause death the following summer). Initial site of infection is epithelial tissues of gills and mortality may begin within 6 weeks of infection and occurs quickly. Continued prevalence of <i>H. nelsoni</i> greatly reduces potential for oyster culture in Chesapeake Bay area although selective breeding improves resistance. Since initial outbreak in the 1950s, MSX has followed cyclical patterns, peaking at 6–8 years	–	Haskin <i>et al.</i> (1966); Andrews (1967, 1984); Sinderman (1976); Ford and Haskin (1982); Haskin and Ford (1982); Ford (1985); Ford and Haskin (1987); Barber <i>et al.</i> (1991); Perkins (1993); Bower <i>et al.</i> (1994); Burreson <i>et al.</i> (2000); Hofmann <i>et al.</i> (2001); Sunila and LaBanca (2003); Dégremont <i>et al.</i> (2010); Wilbur <i>et al.</i> (2012)
	In 1950, 2·881 M bushels (=576·2 M oysters) were landed in Maryland (valued at US\$ 5·53 M) and 3·357 M bushels (671·4 M oysters valued at US\$ 12·02 M) in Virginia. In 1960, although the landings were little changed, i.e. 2·354 M bushels valued at US\$ 3·86 M for Maryland and 3·357 M bushels valued at US\$ 10·88 M for Virginia, if these represent the 10% of the stock that survived, then the combined losses can be estimated at US\$ 132·64 M.	132·64 M	Mackenzie (1996)
<i>Mikrocytos mackini</i> (mikrocytosis)	Protozoan parasite that has been placed within the Class Ascetospora. Known to infect <i>C. virginica</i> and cause mikrocytosis (see <i>C. gigas</i>)	–	Bower <i>et al.</i> (2005)
<i>Perkinsus marinus</i>	Widely distributed protistan parasite causing high mortalities in oyster populations. Disseminates slowly from a centre of infection via snail vector <i>Boonea impressa</i> . Highest mortalities (up to 95%) in summer months on east coast of America from Massachusetts to Brazil. Estimated 50% annual mortality. Increases most readily at 25 °C, but infections recorded in more northerly areas (implications for warmer conditions due to climate change) and at higher salinity. Causes reduction in bodily soft tissue, overall condition and growth rate. Growth rate may be reduced by 60% or more in moderate salinity and 80% at high salinity. Lethal threshold is 10^6 pathogen cells g^{-1} tissue wet weight	–	White <i>et al.</i> (1987); Andrews (1988); Crosby and Roberts (1990); Paynter and Burreson (1991); Burreson (1996); Cook <i>et al.</i> (1998); Oliver <i>et al.</i> (1998); La Peyre <i>et al.</i> (2003); Dungan <i>et al.</i> (2012)
	Infection of oysters in Maryland and Virginia throughout the late 1980s and early 1990s resulted in high mortality rates. American production in 1987 was 80 893 t, assuming that this level of output would have been maintained in the absence of the parasite and that <i>Perkinsus</i> is the only factor responsible for loss, then approx. 2720 t worth an est. US\$ 62·83 M was lost between 1988 and 1991. Production in 1992 had returned pre-1988 levels at 83 544 t	62·83 M (1988–1991)	Burreson and Ragone Calvo (1996)
<i>Bucephalus cuculus</i>	Larval trematode; causes castration, starvation and eventually death. Occurs on East Atlantic coast of USA (Maryland). Infections of <i>Bucephalus</i> sp. documented to result in	–	Tripp (1973); Andrews (1984); Bower <i>et al.</i> (1994)

Table 9. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
<i>Mycicola ostreae</i>	castration and hermaphroditism. Overall infections can reduce ability to withstand environmental stress Generally harmless parasitic copepod, although can cause gill lesions and severe gill erosion. Found in Europe, eastern and western USA and British Columbia	–	Lauckner (1983); Bower <i>et al.</i> (1994)
(r) Grooved carpet shell, <i>Ruditapes decussatus</i> L. (syn. <i>Tapes decussatus</i>)			
<i>Perkinsus atlanticus</i>	Pathogenic protistan, synonymous with <i>P. olseni</i> , infecting <i>Ruditapes</i> sp. Documented in <i>R. decussatus</i> to cause formation of cysts on gills, foot and mantle when infections are high intensity and associated high mortalities. Laboratory experiments show mortality from heavy infections start to occur after 4 weeks. Parasitized clams more susceptible to opportunistic infections. Distributed in the Mediterranean Sea and NW Spain	–	DaRos and Canzonier (1985); Azevedo (1989); Bower <i>et al.</i> (1994); Almeida <i>et al.</i> (1999); Montes <i>et al.</i> (2001); Shimokawa <i>et al.</i> (2010)
<i>Bacciger bacciger</i>	Sporocysts and cecariae heavily infect gonads and visceral mass of <i>R. decussatus</i> in St. Gilla Lagoon, Sardinia. Responsible for mortality of carpet shells (alongside infections of <i>Perkinsus marinus</i>) cultured in Venice Lagoon over 2 years (10% in year 1 and 25% in year 2)	–	Breber (1985); Culurgioni <i>et al.</i> (2006)
<i>Nematopsis</i> sp.	Infects intestinal tissues where it reproduces in host connective tissue and gills causing lysis of the nearby gill cells. Severe mortality attributed to <i>Nematopsis</i> infection in other bivalve species in Asia; however, some discrepancy in literature as to pathological impacts in bivalves	–	Canestri-Trotti <i>et al.</i> (2000); Culurgioni <i>et al.</i> (2006)
<i>Paravortex cardii</i>	Turbellarian found in intestinal lumen, digestive gland and gonads. Also common parasite of cockle <i>Cerastoderma edule</i> . Heavy infection causes widespread host damage and castration	–	Carballal <i>et al.</i> (2001); Culurgioni <i>et al.</i> (2006)
(s) Sydney rock oyster, <i>Saccostrea glomerata</i> (Gould, 1850) (syn. <i>Saccostrea commercialis</i>)			
<i>Bonamia</i> (syn. <i>Mikrocytos</i>) <i>roughleyi</i> (Australian winter disease)	Protozoan causing pustules, ulcerations and abscesses in gills, gonads and mantle. Mortalities up to 70% in mature oysters in third winter before marketing. Linked to low temperatures and high salinities (30–35‰). If an av. mortality of 20% mortality is applied across the industry and to the av. tonnages produced across the period 1988–1994 (i.e. 5760 t; the dates of the 2 reports), then loss can be est. at 1152 t p.a. worth ~ US\$ 4.56 M p.a. or a total loss of ~ 8064 t across the period worth an est. US\$ 31.91 M	4.56 M p.a.	Farley <i>et al.</i> (1988); Bower <i>et al.</i> (1994)
<i>Marteilia sydneyi</i>	Causes QX disease. High mortalities in oyster cultures recorded in eastern Australia. Outbreaks often occur following heavy summer rains in southern Queensland and New South Wales	–	Potter (1983); Lester (1986); Wesche (1995); Hine and Thorne (2000)
<i>Paramarteilia</i> (syn. <i>Marteilioides</i>) <i>branchialis</i>	Protozoan causing lesions in gills. Along with infections of <i>Marteilia sydneyi</i> , high levels of mortalities may occur in tray-cultured oysters throughout autumn	–	Bower <i>et al.</i> (1994)
Korean mussel, <i>Mytilus coruscus</i> (Gould, 1861) (syn. <i>Mytilus crassitesta</i>)			
<i>Dipolydora giardia</i> (Spinoidae)	Shell-boring polychaete infecting Asian cultures of <i>M. coruscus</i> . Causes blistering, weakened shells, reduced growth and often mortality of host	–	Sato-Okoshi <i>et al.</i> (2012)
<i>Modiolicola gracilicaudus</i>	Parasitic copepod infecting <i>M. coruscus</i> in Sea of Japan. Known to cause considerable damage to hosts, which may result in mortality	–	Ho (1980); Kim (2004)

<i>Mytilicola orientalis</i>	Parasitic copepod spread from Eastern Asia to mussel and oyster cultures in Europe and USA. Negative effects on host occur when environmental conditions are extreme and infection is high (>25 copepods per host). Infection occurs in intestine	–	Ho (1980); Pogoda <i>et al.</i> (2012)
(t) Winged pearl oyster, <i>Pteria penguin</i> (Röding, 1798)			
<i>Tylocephalum metacestodes</i>	Larval tapeworm. Encysts in connective tissue of digestive gland	–	Hine and Thorne (2000)
Boring polychaetes, sponges, molluscs and isopods (e.g. <i>Polydora</i> sp., <i>Cliona</i> sp.)	Cultured <i>P. penguin</i> in Philippines suffer from heavy fouling reducing shell quality and survival	–	Bondad-Reantaso <i>et al.</i> (2007)
(u) Geoduck, <i>Panopea generosa</i> (Gould, 1850)			
<i>Isonema</i> -like flagellate	Only found in geoduck larvae in USA hatcheries. Flagellate enters mantle and multiplies within coelom causing death when infection is heavy	–	Kent <i>et al.</i> (1987); Bower <i>et al.</i> (1994); Bower and Blackburn (2003)
(v) Inflated ark or blood clam, <i>Scapharca broughtonii</i> (Schrenck, 1867)			
<i>Perkinsus</i> sp.	Low-level infections were found in specimens taken from Komsoe Bay, S. Korea where Manila clams, <i>Ruditapes philippinarum</i> , were found to be moderately infected (i.e. 1 M+ hypospores clam ⁻¹) at levels that might impair growth and reproduction	–	Park <i>et al.</i> (1999)
(w) Pen shell, <i>Atrina</i> spp.			
The authors have been unable to find any recorded economic impacts of protozoan or metazoan parasites for this species			
(x) South American rock mussel, <i>Perna perna</i> (Linnaeus, 1758)			
Bucephalid (Digenea) sporocysts and <i>Proctoeces</i> sp.	Brown mussels from Hougham Park and Kowie Point, South Africa are commonly infected with digeneans (<i>Proctoeces</i> sp. and bucephalid sporocysts; prev. 50%). <i>Proctoeces</i> infection has an impact on mussel growth whilst the presence of bucephalid sporocysts result in castration of their hosts	–	Calvo-Ugarteburu and McQuaid (1998)

Table 10. The estimated economic cost of notable protistan and metazoan parasite events on the 10 leading crustacean marine and brackish water-based aquaculture industries

Parasite	Impact	Estimated loss (US\$)	Reference
(a) Whiteleg shrimp, <i>Litopenaeus vannamei</i> (Boone, 1931) (syn. <i>Penaeus vannamei</i>)			
<i>Agmasoma penaei</i>	A microsporidian infection was found in 2% of the culture stock at a farm in Yucatan, Mexico	–	Vidal-Martínez <i>et al.</i> (2002)
<i>Apiosoma</i> sp.	Infection found with a 6–57% prevalence on stock held at a farm in Yucatan, Mexico	–	Vidal-Martínez <i>et al.</i> (2002)
<i>Epistylis</i> sp.	Ciliates were prevalent on 2–29% of stock at a site in Yucatan, Mexico	–	Vidal-Martínez <i>et al.</i> (2002)
Haplosporidian sp.	A survey of 149–191-day-old juvenile stock (carapace length 1.47–2.21 cm) at an intensive culture farm at Sisal, Yucatan throughout Dec 2001–Nov 2002 stocked in ponds at 98–136 shrimp m ² found infection on 22–100% of shrimp with a mean intensity of infection of 13 ± 28 to 126 ± 124 parasites shrimp ⁻¹	–	López-Téllez <i>et al.</i> (2009)
	Imported <i>L. vannamei</i> juveniles from Nicaragua into Cuba and held in a quarantine facility between Oct 1985 and March 1986 were subsequently found to have an hepatopancreatic infection which was detected in 31 out of 53 (58.5%) shrimp that were sampled	–	Dyková <i>et al.</i> (1988)
	During the winter season Sept 2004–Feb 2005, stock in ponds in Belize showed slow growth and reduced survival	–	Nunan <i>et al.</i> (2007)
	Progressive mortalities of juvenile (< 1-month-old stock) rising to 60–90% in severely affected Indonesian hatcheries due to infections of intracellular haplosporidians have, since 2007, been estimated to have cost the national industry US\$ >5 M. Infections result in atrophy of the hepato-pancreas, retarded growth, melanization of the cuticle and a flaccid body. Infected shrimp, however, were frequently co-infected with <i>Vibrio</i> spp.	> 5 M	Utari <i>et al.</i> (2012)
<i>Sirolopidium</i> spp.	Study comments that this phycmycete fungus has been responsible for severe mortalities in Larval cultures. No details of loss provided	–	Carr (1996)
<i>Thelohania</i> sp.	Twenty to 25 days post-stocking infections were first observed in shrimp stocked at 125 post-larvae m ² in three 8000 m ² ponds in Thailand. The prevalence of infection rose to 25–28% on day 60 but had decreased to 3–5% when the stocks were harvested. The study demonstrated that the severity of infection impacts on growth performance and survival	–	Prasertsri <i>et al.</i> (2009)
<i>Zoothamnium</i> sp.	Ciliates were observed on 2–65% of farm stock at a site in Yucatan, Mexico	–	Vidal-Martínez <i>et al.</i> (2002)
	A survey of juvenile shrimp (carapace length 1.47–2.21 cm) throughout Dec 2001–Nov 2002 at an intensive culture farm at Sisal, Yucatan found ciliates with a prevalence of between 0 and 87% with a mean intensity of infection of between 0 to 144 ± 132 parasites shrimp ⁻¹	–	López-Téllez <i>et al.</i> (2009)
(b) Giant tiger prawn, <i>Penaeus monodon</i> (Fabricius, 1798)			
<i>Fusarium</i> sp. + luminescent bacteria (<i>Vibrio</i> sp.)	Infections are reported to result in the heavy mortality of mysis and post-larvae in hatcheries on the coast of Andhra Pradesh, India. No details relating to the scale of loss are provided	–	Siva Kamari and Ramesh Babu (2001)
<i>Haliphthoros philippinensis</i>	Suggested that infections within the hatchery production of larvae can be devastating	–	Lio-Po and Sanvictores (1986)

<i>Isochrysis galbana</i>	100% prevalence on the gills and appendages of post-larvae causing fouling. Infection within closed tanks in an Indian hatchery resulted in >80% mortality	–	Aravindan <i>et al.</i> (2007)
<i>Lagenidium</i> sp.	Infections are suggested to be the basis of heavy losses in hatcheries	–	Lio-Po and Sanvictores (1986)
<i>Lagenidium callinectes</i>	Infections are reported to affect the eggs and larvae of marine Crustacea. A culture facility in India stocked six 5000 L tanks with a total of 3.84 M nauplii. There was a 5.33+0.55% mortality between nauplii and zoea, a subsequent 24.68±4.58% mortality of zoea 5 days after stocking, and, a total mortality of 47.89±0.27% of mysis 10 days after stocking, i.e. the mortality of 1.84 M larval stages. No figures for the value of stock at that time are available; however, the current cost of 1 M <i>L. vannamei</i> nauplii in Thailand (as of May 2014) is approx. 10 000 THB (=US\$ 309)	569	Ramasamy <i>et al.</i> (1996)
<i>Lagenidium thermophilum</i>	Fungal infection of eggs and larvae at hatcheries within Chachensao Province, Thailand result in high mortalities. The study provides no details relating to the scale of losses but the economic losses follow those given for <i>L. callinectes</i> above, i.e. US\$ 309 per 1 M nauplii (i.e. 100 nauplii per 1 Thai baht (THB)) or US\$ 3087–4322 per 1 M post-larvae (i.e. 1–1.4 THB per 10 PL)	309–4322 per 1 M juveniles lost	Muraosa <i>et al.</i> (2006)
(c) Indo-Pacific swamp crab, <i>Scylla serrata</i> (Forsskål, 1775)			
<i>Haliphthoros milfordensis</i> + <i>Halocrusticida baliensis</i> + <i>Lagenidium callinectes</i>	Seed production at a research station in Bali, Indonesia has been hampered since 1992 by fungal infections resulting in almost 100% mortality of zoeae. Eggs were derived from spawning adults (200–300 g) brought in from Probolinggo, East Java in July 1997 and held in a 16 m ³ tank. The number of zoeae lost is not detailed but almost all the zoeae were lost on each occasion to fungal agents. It is assumed that 16 female broodstock crabs were held (i.e. 1 crab m ³), that each crab produced an av. 3 million eggs and spawned three times, that the same protocol was used throughout 1992–1997, and, in the absence of livestock prices the current minimum wage for an Indonesian labourer, i.e. US\$ 73.72 month ⁻¹ (=850 000 Indonesian rupiah; see http://en.wikipedia.org/wiki/List_of_minimum_wages_by_country) is used for an animal husbandry for a period of 6 months (based on 3 spawnings with a 41–46-day interval between each; see FAO 2014e)	442 p.a. 2652 over the 6 years programme	Hatai <i>et al.</i> (2000)
<i>Hematodinium</i> sp.	A case of yellow water disease (or milky disease) was recorded in the mud crab reared in the Shanmen area of Zhejiang, China resulting in a high mortality of stock. The infected crabs were thin with heavy parasitic burdens within the gills, heart, muscle and hepatopancreas	–	Xu <i>et al.</i> (2007a)
	A low-salinity culture area (<9 ppt) covering 7000 ha in Guangdong Province, southern China, producing approx. 2000 t of crab, has had recurrent infections of milky disease since 2005. Infections typically breakout in Sept–Nov as the crab approaches maturity with high mortalities resulting (> 60%). Loss is estimated on the 60% of stock lost annually throughout this region (i.e. 3000 t) using the harvest price of Chinese mud crab in 2005, i.e. US\$ 3.85 kg ⁻¹ , to 2008, i.e. US\$ 4.04 kg ⁻¹	11.55–12.12 M p.a. (=46.35 M total loss for the region 2005–2008)	Li <i>et al.</i> (2008)
	If, however, a 60% mortality is typical of the losses throughout the entire industry, then loss can be calculated on the 40% produced in 2005 through to 2008, i.e. 12 075, 13 132, 16 898 and 18 694 t. This represents national losses of 18 112 t (2005), 19 698 t (2006), 25 347 t (2007) and 28 041 t (2008). Estimates are based on the harvest price of crab in each year, i.e. US\$ 3.85, 3.96, 3.60 and 4.04 kg ⁻¹ , respectively	69.73 M (2005) 78 M (2006) 91.25 M (2007) 113.29 M (2008)	
Thraustochytrid sp.	Two cases of egg infection by an epiparasite during a broodstock research programme resulted in 100% mortality. Berried females were collected from the Ross River, South Townsville, Queensland, Australia. Sequencing of the unidentified parasite suggested similarities to <i>Dermocystidium</i> sp. and <i>Rhinosporidium seeberi</i>	–	Kvingedal <i>et al.</i> (2006)
(d) Indian white prawn, <i>Fenneropenaeus indicus</i> (Milne-Edwards, 1837) (syn. <i>Penaeus indicus</i>)			
<i>Amyloodinium ocellatum</i>	Mortality rates can reach 100% under laboratory conditions	–	Aravindan <i>et al.</i> (2007)

Table 10. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
<i>Gymnodinoides caridinae</i>	Infection over the gills and appendages on post larvae results in a heavy accumulation of mucus on the gills with high percentages of resultant mortality	–	Aravindan <i>et al.</i> (2007)
<i>Leptomonas</i> -like parasite	Infections by an amoeboflagellate are reported to cause the mortality of protozoa and mysis in hatcheries at Narakkal, near Cochin, India. A total of 14 incidences throughout a 2-year period were reported with mortalities ranging from 70 to 90%. No mortalities were reported in naupliar and post-larval stages. Assuming that at least a single 1000 L larval culture tank stocked at 500 mysis L ⁻¹ were affected on each occasion, by applying an av. mortality rate of 80% and an est. current cost of US\$ 2500 per 1 M mysis, then the 7 M mysis that were lost can be estimated	17 500	Sahul Hammed (1996)
Microsporidia sp.	The study reports that microsporidian infections represent a significant loss to the industry but does not specify whether reference is being made to aquaculture	–	Ramasamy <i>et al.</i> (2000)
<i>Nitzschia closterium</i>	Nine separate episodes of dinoflagellate infection between Jan and March 1986 in a commercial hatchery at Narakkal, near Cochin, India resulted in mortalities of between 75 and 100% of larval stock. No details relating to the culture system (other than the 1000 L size of the algal culture tanks) are provided. Assuming that at least three 1000 L larval culture tanks stocked at 500 post-larvae L ⁻¹ were affected on each occasion, by applying an av. mortality rate of 87% and current costs of US\$ 3087–4322 per 1 M post-larvae, then the 11·745 M post-larvae that were lost can be estimated	36 256–50 762	Sahul Hammed (1995)
(e) Swimming crab, <i>Portunus trituberculatus</i> (Miers, 1876)			
<i>Hematodinium</i> sp.	A case of milky disease resulting in the high mortality of stock in July–Sept 2004 was reported from Zhoushan, Zhejiang Province	–	Xu <i>et al.</i> (2007b)
	Ten percent ($n = 4$) of the crabs (108–220 g; $n = 40$) sampled from polyculture sites along the Shandong Peninsula, coastal Southern China from July to Nov 2012 were found to be infected. These crabs were lethargic and had watery haemolymph. The experimental inoculation of 15 crabs found that the dinoflagellate was infectious and resulted in 60% mortality. Mortalities in this region of China have been reported since 2004, which contributes approx. a third of the industry's national income. If 10% mortality in the region is assumed, then between 2216 and 3688 t were lost each year between 2004 and 2012. Estimates are based on the harvest price of crab in 2004, i.e. US\$ 3·00 kg ⁻¹ , and 2012, i.e. US\$ 3·51 kg ⁻¹	6·65–12·94 M p. a. (2004–2012)	Li <i>et al.</i> (2013)
	If it is assumed that production in the Shandong Peninsula represents a third of national production, then the total loss in production in the region can be calculated from 2004, when mortalities were first reported, to 2012 (the date of the study), using the harvest price for each year, i.e. a total loss of approx. 9331 t	31·63 M (2004–2012)	
	If losses in this region are representative of the losses throughout the industry, then over the period 2004–2012, a total of 83 982 t of production was lost	284·97 M	
(f) Kuruma prawn, <i>Marsupenaeus japonicus</i> (Spence Bate, 1888) (syn. <i>Penaeus japonicus</i>)			
<i>Cothurnia</i> sp.	Two 9000 m ² ponds stocked with juvenile prawns (1·1–12·5 g) at 40 and 60 prawns m ² were followed over a growth cycle. Infection on stock occurred with a prevalence of 76·7–84·7% with an av. 21·3–35·9 ciliates gill ⁻¹ (range 0–116·2 ciliates gill ⁻¹)	–	Hudson and Lester (1992)
<i>Dermocystidium</i> sp. + <i>Lagenidium</i> sp.	Infection resulted in the large scale mortality of hatchery reared shrimp within 2–3 days	–	Cook (1971 cited in Overstreet, 1973)

<i>Ephelota</i> sp.	Juvenile stocks (1.1–12.5 g) in two 9000 m ² ponds in Brisbane Australia were followed over an extended growth cycle from Feb to Sept 1990. An average 0.3–0.7 ciliates gill ⁻¹ ; prev. 12.0–16.7%; mean intensity 2.0–5.6 ciliates gill ⁻¹ , were recorded. Infections can cause larval stages to continuously flick their tails leading to fatigue and death (see Overstreet, 1987)	–	Hudson and Lester (1992)
<i>Haematonectria haematococca</i> (syn. <i>Fusarium solani</i>)	The agent responsible for black gill diseases can result in serious mortalities of pond cultured stocks	–	Bian and Egusa (1981)
Microsporidia sp.	Four 10 000 L tanks in Australia in 1997 were stocked with the progeny of spawners collected from the wild. Each tank had 1 M larvae (100 larvae L ⁻¹), 92% of which had trailing faecal casts. Following the mortality of 60% first stage nauplii larvae, a subsequent histological investigation found multifocal hepatopancreatic degeneration and necrosis associated with basophilic staining bodies that resembled sporocyst stages of a microsporidian. Loss is est. on the mortality of 2.4 M larva using current value of <i>L. vannamei</i> nauplii in Thailand (as of May 2014), i.e. 10 000 THB (=US\$ 309) per 1 M nauplii	742	Hudson <i>et al.</i> (2001)
<i>Zoothamnium</i> sp.	Infections on juvenile (1.1–12.5 g) stock in two 9000 m ² ponds in Brisbane, Australia, stocked at 40 and 60 prawns m ² , were monitored over a growth cycle from Feb to Sept 1990. An av. 13.5–16.3 ciliates gill ⁻¹ (range 0–85.4 ciliates gill ⁻¹), prev. 67.3–87.3%; mean intensity 18.8–20.6 ciliates per gill ⁻¹ , were found. Although no direct losses were recorded in this study, large infections can cause stress and lead to mortality (see Overstreet, 1973)	–	Hudson and Lester (1992)
(g) Fleishy prawn, <i>Fenneropenaeus chinensis</i> (Osbeck, 1765) (syn. <i>Penaeus chinensis</i>)			
<i>Paranophrys</i> sp.	Ciliate disease has been reported in larvae and overwintering adults frequently causing 100% mortality in infected tanks. No details relating to the numbers of prawn lost or their value are available, however, loss can be estimated by applying contemporary prices for <i>L. vannamei</i> nauplii (1 M=US\$ 309) and post-larvae (1 M=US\$ 3087 to 4322), and, the harvest price for <i>F. chinensis</i> in 2012, i.e. US\$ 4.00 kg ⁻¹ in China and US\$ 16.69 kg ⁻¹ in South Korea	309 per 1 M nauplii; 3087–4322 per 1 M PL; 4000 to 16 685 t ⁻¹	Bower <i>et al.</i> (1994)
(h) <i>Metapenaeus</i> spp., e.g. <i>Metapenaeus monoceros</i> (Fabricius, 1798) + <i>M. ensis</i> (De Haan, 1844)			
<i>Orbione bonnieri</i>	The study suggested that 10% of the shrimp produced in Thailand are infected with bopyrid isopods	–	Printrakoon and Purivirojkul (2012)
(i) Mud crab spp. e.g. <i>Scylla (paramamosain</i> Estampador, 1949)	No details relating to major parasite-induced losses are available		
(j) Blue shrimp, <i>Litopenaeus stylirostris</i> (Stimpson, 1874) (syn. <i>Penaeus stylirostris</i>)			
<i>Pleistophora</i> sp.	In 1987, a culture facility in Baja California, Mexico had infections that rose from 2% to 10% over the period April–Nov 1987 resulting in the destruction of abdominal striated muscle. In the absence of harvest prices for Mexico, an av. price (i.e. US\$ 9.23 kg ⁻¹) is est. from sales in 1987 in Ecuador, i.e. US\$ 7.20 kg ⁻¹ , in El Salvador, i.e. US\$ 12.00 kg ⁻¹ , and, Panama, i.e. US\$ 8.50 kg ⁻¹	923 t ⁻¹	Alarcon-Gonzalez (1990)

Table 11. The estimated economic cost of notable protistan and metazoan parasite events on other large-scale commercial aquaculture industries.

Parasite	Impact	Estimated loss (US\$)	Reference
(a) Ascidiacea: sea pineapple, <i>Halocynthia roretzi</i> (Von Drasche, 1884)			
<i>Azumiobodo hoyamushi</i>	Over the past decade, the mortality of sea squirts attributable to soft tunic syndrome in the Tongyoung region of South Korea has ranged from 60% to 90%, i.e. 13 000–50 000 t of morts, accounting for a loss of 30 M	30 M	Han <i>et al.</i> (2012); Kim <i>et al.</i> (2014)
Kinetoplastid protistan	Production began in Korea in 1982 with 39 t, increasing to 42 800 t in 1994 but then decreasing to 4500 t in 2004 due to the mass mortality of stock during the winter season. Loss is based on the assumption that the production of 42 800 t could have been maintained and the yearly value of Korean stock (i.e. av. loss of 33 216 t p.a. or a total estimated loss of 564 671 t between 2004 and 2011	Est. total loss of 763.65 M from 1995 to 2011; av. loss of 44.92 M p.a.	Kumagai <i>et al.</i> (2010, 2011)
	Outbreaks were observed resulting in the mass mortality (17–100%) of stock at 3 farm sites throughout Miyagi Prefecture, Japan in 2007, 6 sites in 2008, and, 14 sites in 2009. Losses were primarily in 2+ and 3+ year-old individuals, occasionally in 1+. Details relating to the magnitude of each loss were not provided but loss is given the price per tonne of Japanese stock in 2007–2009	Loss of 1102 t ⁻¹ in 2007; 1328 t ⁻¹ in 2008; and, 1465 t ⁻¹ in 2009	Kumagai <i>et al.</i> (2010)
(b) Cnidaria: edible jellyfish, <i>Rhopilema esculentum</i> (Kishinouye, 1891)			
	No details relating to major parasite-induced losses are available, however, the studies of You <i>et al.</i> (2007) and Dong <i>et al.</i> (2009) provide comprehensive overviews of the Chinese <i>R. esculentum</i> aquaculture, with mortality details in the latter relating to the release of stock and thereafter		
(c) Echinoidea: Sea urchins: <i>Strongylocentrotus intermedius</i> (Agassiz, 1863) and the Dalian purple urchin, <i>S. nudus</i> (Agassiz)			
	No major losses due to parasitic agents are recorded for these species; however, the mass mortality of <i>Strongylocentrotus droebachiensis</i> in Nova Scotia in 1982 is documented in the study of Scheibling and Stephenson (1984). Although the authors were unable to identify the specific cause of mortality, they refer to the findings of Li <i>et al.</i> (1982) and their suggestion that an amoeboid protist, tentatively identified as <i>Labyrinthomyxa</i> sp., was the cause of an early mortality event. Further, recent mortality events in the same area have been attributed to the amoeba <i>Paramoeba invadens</i> (see Feehan <i>et al.</i> 2013)		
(d) Holothuroidea: Japanese spiky sea cucumber or Japanese sea cucumber, <i>Apostichopus japonicus</i> (Selenka, 1867)			
Fungi	Although infections between April and Aug each year are common in pond culture, they do not typically result in large-scale mortalities. Infected animals can appear discoloured and develop oedema. No details relating to loss are provided	–	Wang <i>et al.</i> (2004)
<i>Boveria labialis</i>	Large numbers are reported attaching to the inner wall of the respiratory tree of both young and adult specimens in offshore ponds (20 °C, 27 ppt) near Dalian, China. Infected animals are sluggish but infections do not generally cause a serious mortality problem. No figures of loss are provided	–	Wang <i>et al.</i> (2004); Long <i>et al.</i> (2006)
Platyhelminthes	Can cause heavy damage to the skin of both aestivated juveniles (1+ cm) and to adults leading to ulceration and death. Mortality rates of up to 90% within a month are reported. Loss is estimated on the harvest price of Chinese produced sea cucumbers in 2003, i.e. US\$ 3.00 kg ⁻¹	2700 t ⁻¹	Wang <i>et al.</i> (2004)

losses are typically underreported, hiding the severity and true impact of certain parasites, e.g. *Amyloodinium*, *Cryptocaryon* and *Trichodina* spp.

Some parasitic diseases, whose continuous or predictably repeated infection levels and difficulty of treatment have caused major economic impacts, have driven particular industries to the point of near collapse. The impact that a kinetoplastid protist responsible for soft tunic syndrome has had on the

ascidian *Halocynthia roretzi* industry throughout South Korea and Japan is an appropriate example (Kumagai *et al.* 2011). In Korea, infections have resulted in a serious decline in the industry from 42 800 t (valued at US\$ 34.17 M) in 1994 to just 4500 t (worth US\$ 8.92 M) in 2004 (Kumagai *et al.* 2010; FAO FishStatJ, 2013). Assuming that the 1994 levels of production could have been maintained in the absence of the flagellate, then it can be

Table 12. The estimated economic cost of notable protistan and metazoan parasite events on some of the world's leading marine and brackish water ornamental fish production industries

Parasite	Impact	Estimated loss (US\$)	Reference
(a) Blue green damselfish, <i>Chromis viridis</i> (Cuvier) – Pomacentridae			
<i>Cryptosporidium</i> sp.	This is only a report of several infected <i>Chromis viridis</i> specimens, with the identification of new genotypes from aquarium fish	–	Zanguee <i>et al.</i> (2010)
<i>Hysterolecitha nahaensis</i>	<i>Chromis viridis</i> has been recorded to be infected by this digenean, but no details on the pathogenicity are reported	–	Barker <i>et al.</i> (1994)
<i>Kudoa amamiensis</i>	This species was described from several fish species, including <i>Chromis isharai</i> and <i>Chromis notatus</i> . Only one to 3 cysts, 2 mm long, were found to infect the skeletal musculature. Fish were collected from Amami-Ohshima and Okinawa coasts (Japan). No mortalities were reported, however, infected hosts were not marketable, due to the cysts distributed throughout the skeletal muscle. Fish response involved fibrous connective tissue surrounding the cyst	–	Egusa and Nakajima (1980); Moran <i>et al.</i> (1999)
(b) Clown anemonefish, <i>Amphiprion ocellaris</i> (Cuvier) – Pomacentridae			
<i>Amyloodinium ocellatum</i>	Mortality caused by this protozoan in aquarium conditions. Destruction of epithelial cells of the skin and the gills	–	Bower <i>et al.</i> (1987); Woo (2006)
<i>Brooklynella hostilis</i>	Acute mortality, skin discoloration, lethargy, inappetance, mucus hyperproduction	–	Lom and Nigrelli (1970); Fenner (1998); Noga (2010)
<i>Cryptocaryon irritans</i>	Acute mortality, white spots on the skin, ragged fins, skin discoloration, mucus hyperproduction	–	Colorni and Burgess, 1997
(c) Flame angel, <i>Centropyge loricula</i> (Günther) – Pomacanthidae			
<i>Amyloodinium ocellatum</i>	Infection report, no details on pathogenicity	–	Landsberg <i>et al.</i> (1994)
<i>Uronema marinum</i>	Reddish skin lesions with deep ulcers on <i>Centropyge flavissima</i>	–	Bassleer (1983)
(d) Sapphire devil, <i>Chrysiptera cyanea</i> (Quoy and Gaimard) – Pomacentridae			
<i>Amyloodinium ocellatum</i>	Infection report, no details on pathogenicity	–	Landsberg <i>et al.</i> (1994)
<i>Cryptosporidium</i> sp.	<i>Chrysiptera hemicyanea</i> has been found to be infected with <i>Cryptosporidium</i> sp., although no pathogenicity has been reported	–	Zanguee <i>et al.</i> (2010)
(e) Threespot dascyllus, <i>Dascyllus trimaculatus</i> (Rüppell) – Pomacentridae			
No details relating to major parasite-induced losses are available			
(f) Banggai cardinal fish, <i>Pterapogon kauderni</i> Koumans – Apogonidae			
No details relating to major parasite-induced losses are available			
(g) Spinecheek anemonefish, <i>Premnas biaculeatus</i> (Bloch) – Pomacentridae			
<i>Uronema marinum</i>	Skin lesions with deep ulcers and white patches	–	Bassleer (1983)
(h) Mandarinfish, <i>Synchiropus splendidus</i> (Herre) – Callionymidae			
No details relating to major parasite-induced losses are available			
(i) Whitetail dascyllus, <i>Dascyllus aruanus</i> L. – Pomacentridae			
<i>Aponurus</i> sp.	Digenean reported in the stomach with no pathogenicity details	–	Cédrik <i>et al.</i> (1998)
<i>Haliotrema</i> sp.	Monogenean reported in the gills with no pathogenicity details	–	Cédrik <i>et al.</i> (1998)
<i>Hysterolecitha nahaensis</i>	Digenean reported with no pathogenicity details	–	Barker <i>et al.</i> (1994)
<i>Scolex polymorphus</i>	Cestode report in the caecum with no pathogenicity details	–	Cédrik <i>et al.</i> (1998)
<i>Tulinia microrchis</i>	Report of this digenean on <i>Dascyllus aruanus</i> with no pathogenicity details	–	Barker <i>et al.</i> (1994)

Table 12. (Cont.)

Parasite	Impact	Estimated loss (US\$)	Reference
(j) Goldtail demoiselle, <i>Chrysiptera parasema</i> (Fowler) – Pomacentridae			
<i>Kudoa amamiensis</i>	This species was described from several fish species, including <i>Chrysiptera assimilis</i> . Only 1–3 cysts, 2 mm long, were found to infect the skeletal musculature. Fish were collected from Amami-Oshima and Okinawa coasts (Japan). Infected hosts were not marketable, due to the cysts distributed throughout the skeletal muscle. Fish response involved fibrous connective tissue surrounding the cyst	–	Egusa and Nakajima (1980); Moran <i>et al.</i> (1999)
(k) Tomato clownfish, <i>Amphiprion frenatus</i> (Brevoort) – Pomacentridae			
<i>Cladosporium</i> sp.	Deep dermal ulcers reported on stock cultured in North Carolina (USA).	–	Silphaduang <i>et al.</i> (2000)
<i>Uronema marinum</i>	Skin lesions with deep ulcers and white patches	–	Bassleer (1983)
(l) Royal gramma, <i>Gramma loreto</i> (Poey) – Grammatidae			
No details relating to major parasite-induced losses are available			

estimated that infections have cost the Korean industry approximately US\$ 764 M between 1994 and 2011 (Table 10). Infections at three Japanese farms were subsequently reported in 2007, which rose to 14 farms in 2009 (Kumagai *et al.* 2010).

Many of the instances of parasite infection provided in Tables 8–12 are derived from case reports for unexpected mortality events. These represent sporadic parasite infection events that have resulted in significant economic losses at a single or small number of sites or within a single season. For example, a *Uronema nigricans* infection of ranches southern bluefin tuna, *Thunnus maccoyii*, in Australia resulted in the loss of between 5 and 10% of stock, worth an estimated US\$ 0.5–1 M (Munday *et al.* 1997). This illustrates that it is impossible to control parasite exposure in the wild phase, with stock typically belonging to mixed age classes and sizes and ranging over a wide area. As there is no standard crop, disease events are, thus, extremely unpredictable.

Many of the smaller magnitude, sporadic mortality events can be attributed to some of the less specific diseases, which can result from low water quality or poor fish handling and welfare and that might, therefore, be ameliorated simply through improved husbandry practices. Although most of these disease events go unreported, either because of the smaller scale of the losses incurred or the general acceptance that they fall within the typical, accepted margins of loss in production, their collective impact on global mariculture production is significant, and the value of fish lost may arguably exceed the economic impact of many of the major parasite pathogens listed in Tables 8–12. Deterioration in water quality can occur through overstocking, inappropriate feeding regimes, low water current speeds/poor flushing or generally poor site hygiene practices. For example, net fouling can result in organic enrichment within culture

systems, facilitating the increase of many opportunistic species, which, if unregulated and unmanaged, can result in health impacts due to infection by low-specificity pathogens, e.g. *Trichodina* spp., *Zoothamnium* spp. Heavily biofouled cage nets can cause reduced flow-through rates resulting in increased retention times of infective stages, net deformation and welfare impacts for caged stock (Lader *et al.* 2008). Decreased flow rates can also result in lower dissolved oxygen and increased ammonia levels and in extreme events, the asphyxiation of stock (Douglas-Helders *et al.* 2003; Madin *et al.* 2010). Fouled nets may also serve as a reservoir for pathogenic agents such as *Paramoeba* (syn. *Neoparamoeba*) *pemaquidensis* (see Tan *et al.* 2002) and the polyopisthocotylean monogenean *Heterobothrium okamotoi*, which infects the gills of the tiger puffer, *Takifugu rubripes*, reared in floating cages. These flukes are extremely fecund with up to 1500 eggs *in utero* producing 50–360 spindle-shaped eggs per day that are extruded in strings of up to 2.8 m in length (Ogawa, 1997; Ogawa and Inouye, 1997; Ogawa *et al.* 2005a). These egg strings readily become entangled within the nets representing a source of reinfection that requires regular net changes to minimise the infection of stock (Ogawa and Inouye, 1997; Ogawa and Yokoyama, 1998; Ogawa, 1999; Ogawa *et al.* 2005a).

Looking through the timeline of parasite episodes, notably those detailed in Table 8, it is interesting to examine some of the underlying husbandry practices that were either directly responsible for or facilitated disease events and to consider whether the magnitude of the subsequent losses may have provided an impetus for change. Some of the earlier reports allude to the use of trash fish feed, e.g. *Kudoa amamiensis* in Japanese amberjack, *S. quinqueradiata* (see Egusa and Nakajima, 1978); the lack of health screening prior to the movement of stocks to new

sites, e.g. the establishment of *Neobenedenia girellae* and *Paradeontacylix* infections on Japanese populations of farmed greater amberjack, *Seriola dumerili* (Ogawa and Egusa, 1986; Ogawa and Fukudome, 1994; Ogawa *et al.* 1995); improvements following the implementation of fallowing and site rotation, e.g. resulting in improved management of *L. salmonis* including a lower number of treatment interventions (Bron *et al.* 1993; Grant and Treasurer, 1993); a shift away from the use of multi-year class sites to an 'all in, all out' single-year class approach (Bron *et al.* 1993); and evident improvements in fish welfare practices, e.g. lower morbidity and mortality rates from *U. nigricans* in ranched tuna (Munday *et al.* 2003). The risks of infection from mixed-species sites have also been previously commented upon (Vagianou *et al.* 2006).

There are, however, a number of concerns associated with the expansion of global aquaculture, much of which is concentrated within the coastal zone, and the potential for an increased incidence of disease events. The human population is set to reach 9.55 billion by 2050 (United Nations, 2012) and there is a general migration towards coastal zones resulting in increasing population density (e.g. in 2000, 53% of the US population resided in 17% of the land area designated as coastal with an expected 24.4% increase in the size of the population residing in the US coastal states between 2000 and 2025 (Boesch *et al.* 2000)), and as a consequence of increased anthropogenic activity along the coastal zone, there will be increase in the levels of nutrients passing into local marine ecosystems and the impacts that these have upon the general marine environment and aquaculture in particular (Halpern *et al.* 2008; Callaway *et al.* 2012). Likewise, the greater globalisation of the trade in aquatic animals and their products, reviewed in Bondad-Reantaso *et al.* (2005), may facilitate the spread of pathogens into new environments. At the same time, in an era of changing climatic conditions, there will be alterations to land run-off, coastal water chemistry/temperature and changes in sea levels and oceanic/coastal currents all of which will have anticipated impacts on current aquaculture production practices, aquaculture systems, the interactions between wild and farmed aquatic stocks, parasite life cycles, transmission pathways and the prevalence and severity of disease events (Overstreet, 2007; Callaway *et al.* 2012). Cook *et al.* (1998) and Hofmann *et al.* (2001), for example, demonstrate links between increasing sea temperatures and the spread of *Perkinsus marinus* and *Haplosporidium nelsoni*, respectively, in the eastern oyster, *Crassostrea virginica*, as do Bermingham and Mulcahy (2004) who indicate that temperature is also an important risk factor in the incidence of amoebic gill disease. Although raised temperature profiles may accelerate the life cycle of some aquatic pathogens, it may also lead to the

decreased prevalence of others as certain parasite species move out of their temperature optima. At the same time, increased water temperatures may preclude the use of certain drug treatment regimes requiring the development of novel strategies for the management and control of parasite pathogens (see Shinn and Bron, 2012).

This study provides a review of the top 69 aquatic species cultured in brackish and marine waters, which accounted for ~94% of the total tonnage derived from mariculture in 2011 and then provides estimates for losses incurred as a consequence of key parasite-associated disease events reported worldwide. Although it has not been possible to provide a single resolved value for the economic impact of parasites on global mariculture, this study clearly demonstrates that parasitic infections remain an important source of economic loss. Without a step-change in management priorities and a concerted move towards more IPMS, it is evident that as the global aquaculture industry grows and intensifies, the level of parasite infections will similarly rise as will the attendant economic costs of parasitism.

ACKNOWLEDGEMENTS

The authors would like to thank Mr Wenbo Zhang from the Institute of Aquaculture, University of Stirling, UK for his kind assistance in providing additional information on Chinese production; Mr Kevin Erickson, the Vice President of the Marine Aquarium Societies of North America (MASNA) for useful discussions on commercially reared ornamental species; and, Ms Jocyntha Joseph from the Asian Fisheries Society (<http://www.asianfisheriesociety.org/>) for providing scientific articles. The views expressed in this article are those of the authors and do not necessarily represent the views of, and should not be attributed to, the Fish Vet Group/Benchmark Holdings PLC, the Institute of Aquaculture, University of Stirling, UK or the Institute of Marine Science, Burapha University, Thailand. There was no funding received to support this study. It was an invited paper and worked on in free time.

REFERENCES

- Abela, M., Brinch-Iversen, J., Tanti, J. and Le Breton, A. (1996). Occurrence of a new histozoic microsporidian (Protozoa, Microsporida) in cultured gilthead sea bream, *Sparus aurata* L. *Bulletin of the European Association of Fish Pathologists* **16**, 196–200.
- Adams, M. B. and Nowak, B. F. (2001). Distribution and structure of lesions in the gills of Atlantic salmon, *Salmo salar* L., affected with amoebic gill disease. *Journal of Fish Diseases* **24**, 535–542.
- Aiken, H. M., Hayward, C. J. and Nowak, B. F. (2006). An epizootic and its decline of a blood fluke *Cardicola forsteri* in farmed southern bluefin tuna *Thunnus maccoyii*. *Aquaculture* **254**, 40–45.
- Alarcon-Gonzalez, C. (1990). Identificación de *Pleistophora* sp. (Microsporida: Nosematidae) como agente infectante del camaron azul *Penaeus stylirostris* sometido a cultivo semi-intensivo en estanqueria rustica en Baja California Sur, Mexico. *Revista Latinoamericana de Microbiología* **32**, 193–196.
- Almeida, M., Berthe, F., Thébaud, A. and Dinis, M. T. (1999). Whole clam culture as a quantitative diagnostic procedure of *Perkinsus atlanticus* (Apicomplexa, Perkinsea) in clams *Ruditapes decussatus*. *Aquaculture* **177**, 325–332.
- Alvarez-Pellitero, P. and Sitjà-Bobadilla, A. (1993). *Ceratomyxa* spp. (Protozoa: Myxosporea) infections in wild and cultured sea bass

- (*Dicentrarchus labrax*) from the Spanish Mediterranean area. *Journal of Fish Biology* **42**, 889–901.
- Alvarez-Pellitero, P. and Sitjà-Bobadilla, A.** (2002). *Cryptosporidium molnari* n. sp. (Apicomplexa: Cryptosporidiidae) infecting two marine fish species, *Sparus aurata* L. and *Dicentrarchus labrax* L. *International Journal for Parasitology* **32**, 1007–1021.
- Alvial, A., Kibenge, F., Forster, J., Burgos, J. M., Ibarra, R. and St-Hilaire, S.** (2012a). The recovery of the Chilean salmon industry: the ISA crisis and its consequences and lessons. *Global Aquaculture Alliance ISA Report*. http://www.gaalliance.org/cmsAdmin/uploads/GAA_ISA-Report.pdf, Published 23rd February 2012.
- Alvial, A., Kibenge, F., Forster, J., Burgos, J. M., Ibarra, R. and St-Hilaire, S.** (2012b). The recovery of the Chilean salmon industry: the ISA crisis and its consequences and lessons. Global Aquaculture Alliance ISA presentation. <http://www.gaalliance.org/update/GOAL11/AdolfoAlvial.pdf>
- Andrews, J. D.** (1967). Interaction of two diseases of oysters in natural waters. *Proceedings of the National Shellfisheries Association* **57**, 38–49.
- Andrews, J. D.** (1982). Epizootiology of late summer and fall infections of oysters by *Haplosporidium nelsoni* and comparison to the annual life cycle of *Haplosporidium costalis*, a typical haplosporidian. *Journal of Shellfish Research* **2**, 15–23.
- Andrews, J. D.** (1984). Epizootiology of diseases of oysters (*Crassostrea virginica*), and parasites of associated organisms in eastern North America. *Helgoländer Meeresuntersuchungen* **37**, 149–166.
- Andrews, J. D.** (1988). Epizootiology of the disease caused by the oyster pathogen *Perkinsus marinus* and its effects on the oyster industry. *American Fisheries Society Special Publication* **18**, 47–63.
- Antonio, D. B., Andree, K. B., Moore, J. D., Friedman, C. S. and Hedrick, R. P.** (2000). Detection of Rickettsiales-like prokaryotes by *in situ* hybridization in black abalone, *Haliotis cracherodii*, with withering syndrome. *Journal of Invertebrate Pathology* **75**, 180–182.
- Aravindan, N., Kalavati, C. and Aravindan, S.** (2007). Protozoan parasites in commercially important shrimp species from northeast coast of Andhra Pradesh, India. *Journal of Experimental Zoology* **10**, 9–20.
- Arfara, S., Bozzetta, E., Prearo, M. and Ghittino, C.** (1995). Cases of kudoosis in cultured juvenile gilthead sea bream. *III Convegno Nazionale Società Italiana Di Patologica Ittica* **7**, 12–17.
- Arthur, J. R. and Ogawa, K.** (1996). A brief overview of disease problems in the culture of marine finfishes in east and Southeast Asia. In *Aquaculture Health Management Strategies for Marine Fishes, Proceedings of a Workshop in Honolulu, Hawaii, October 9th–13th, 1995* (ed. Main, K. L. and Rosenfeld, C.), pp. 9–31. The Oceanic Institute, Hawaii.
- Athanassopoulou, F.** (1992). Ichthyophoniasis in sea bream, *Sparus aurata* (L.), and rainbow trout, *Oncorhynchus mykiss* (Walbaum), from Greece. *Journal of Fish Diseases* **15**, 437–441.
- Athanassopoulou, F.** (1998). A case report of *Pleistophora* sp. infection in cultured sea bream (*Sparus aurata* L.) in Greece. *Bulletin of the European Association of Fish Pathologists* **18**, 19–21.
- Athanassopoulou, F., Bouboulis, D. and Martinsen, B.** (2001a). *In vitro* treatments of deltamethrin against the isopod parasite *Anilocra physodes*, a pathogen of seabass *Dicentrarchus labrax* L. *Bulletin of the European Association of Fish Pathologists* **21**, 26–29.
- Athanassopoulou, F., Prapas, A. and Rodger, H.** (1999). Diseases of *Puntazzo puntazzo* Cuvier in marine aquaculture systems in Greece. *Journal of Fish Diseases* **22**, 215–218.
- Athanassopoulou, F., Ragias, V., Tavla, J., Christophilogannis, P. and Liberis, N.** (2001b). Preliminary trials on the efficacy and toxicity of ivermectin against *Lernathropus kroeyeri* Van Beneden, 1851 in cultured sea bass *Dicentrarchus labrax* L. *Aquaculture Research* **32**, 77–79.
- Azevedo, C.** (1989). Fine structure of *Perkinsus atlanticus* n. sp. (Apicomplexa, Perkinsea) parasite of the clam *Ruditapes decussatus* from Portugal. *Journal of Parasitology* **75**, 627–635.
- Balseiro, P., Montes, A., Ceschia, G., Gestal, C., Novoa, B. and Figueras, A.** (2007). Molecular epizootiology of the European *Marteilia* spp., infecting mussels (*Mytilus galloprovincialis* and *M. edulis*) and oysters (*Ostrea edulis*): an update. *Bulletin of the European Association of Fish Pathologists* **27**, 148–156.
- Barber, B. J., Ford, S. E. and Littlewood, D. T. J.** (1991). A physiological comparison of resistant and susceptible oysters *Crassostrea virginica* (Gmelin) exposed to the endoparasite *Haplosporidium nelsoni* (Haskin, Stauber & Mackin). *Journal of Experimental Marine Biology and Ecology* **146**, 101–112.
- Barker, S., Cribb, T. H., Bray, R. A. and Adlard, R. D.** (1994). Host-parasite associations on a coral reef: pomacentrid fishes and digenae trematodes. *International Journal for Parasitology* **24**, 643–647.
- Bassleer, G.** (1983). *Uronema marinum*, a new and common parasite on tropical salt-water fishes. *Freshwater and Marine Aquarium* **6**, 78–79.
- Baticados, M. C. and Qunitio, G. F.** (1984). Occurrence and pathology of an *Amyloodinium*-like protozoan parasite on the gills of grey mullet, *Mugil cephalus*. *Helgoländer Meeresunters* **37**, 595–601.
- Becker, C. D. and Pauley, G. B.** (1968a). An ovarian parasite (*Protista incertae sedis*) from the Pacific oyster, *Crassostrea gigas*. *Journal of Invertebrate Pathology* **12**, 425–437.
- Becker, C. D. and Pauley, G. B.** (1968b). A parasite from the ova of the Pacific oyster *Crassostrea gigas*. *Proceedings of the National Shellfisheries Association* **58**, 11 (Abstract).
- Bermingham, M. L. and Mulcahy, M. F.** (2004). Environmental risk factors associated with amoebic gill disease in cultured salmon, *Salmo salar* L., smolts in Ireland. *Journal of Fish Diseases* **27**, 555–571.
- Bian, B. Z. and Egusa, S.** (1981). Histopathology of black gill disease caused by *Pisurium solani* (Martius) infection in the Kuruma prawn, *Penaeus japonicus* Bate. *Journal of Fish Diseases* **4**, 195–201.
- Blackbourn, J., Bower, S. M. and Meyer, G. R.** (1998). *Perkinsus qugwadi* sp. nov. (*incertae sedis*), a pathogenic protozoan parasite of Japanese scallops, *Patinopecten yessoensis*, cultured in British Columbia, Canada. *Canadian Journal of Zoology* **76**, 942–953.
- Blateau, D., LeCoguic, Y., Mialhe, E. and Gizel, H.** (1992). Mussel (*Mytilus edulis*) treatment against the red copepod *Mytilicola intestinalis*. *Aquaculture* **107**, 165–169.
- Boesch, D. F., Field, J. C. and Scavia, D.** (2000). *The Potential Consequences of Climate Variability and Change on Coastal Areas and Marine Resources: Report of the Coastal Areas and Marine Resources Sector Team, U.S. National Assessment of the Potential Consequences of Climate Variability and Change, U.S. Global Change Research Program*. NOAA Coastal Ocean Program Decision Analysis Series No. 21. NOAA Coastal Ocean Program, Silver Spring, MD.
- Bondad-Reantaso, M. G., Kanchanakhan, S. and Chinabut, S.** (2001). Review of grouper diseases and health management strategies for groupers and other marine fishes. In *Report of a Workshop held in Bangkok, Thailand, 18th–20th May, October 2000* (ed. Bondad-Reantaso, M. G., Humphrey, J., Kanchanakhan, S. and Chinabut, S.), pp. 121–146. Asia Pacific Economic Cooperation (APEC), Fish Health Section of the Asian Fisheries Society (FHS-AFS), Aquatic Animal Health Research Institute (AAHRI) and Network of Aquaculture Centres in Asia-Pacific (NACA), Bangkok, Thailand.
- Bondad-Reantaso, M. G., McGladdery, S. E., Ladra, D. and Chongming, W.** (2007). Pearl oyster health: experiences from the Philippines, China, the Persian Gulf and the Red Sea. In *Pearl Oyster Health Management: a Manual* (ed. Bondad-Reantaso, M. G., McGladdery, S. E. and Berthe, F. C. J.), pp. 111–121. *FAO Fisheries Technical Paper No. 503*. Rome, FAO, 120 pp.
- Bondad-Reantaso, M. G., Subasinghe, R. P., Arthur, J. R., Ogawa, K., Chinabut, S., Adlard, R., Tan, Z. and Shariff, M.** (2005). Disease and health management in Asian aquaculture. *Veterinary Parasitology* **132**, 249–272.
- Bower, S. M.** (1987a). *Labyrinthuloides haliotidis* n. sp. (Protozoa: Labyrinthomorpha), a pathogenic parasite of small juvenile abalone in a British Columbia mariculture facility. *Canadian Journal of Zoology* **65**, 1996–2007.
- Bower, S. M.** (1987b). Pathogenicity and host specificity of *Labyrinthuloides haliotidis* (Protozoa: Labyrinthomorpha), a parasite of juvenile abalone. *Canadian Journal of Zoology* **65**, 2008–2012.
- Bower, S. M.** (1992). Diseases and parasites of mussels. In *The Mussel Mytilus* (ed. Gosling, E.), pp. 543–563. Elsevier, Amsterdam.
- Bower, S. M., Bate, K. and Meyer, G. R.** (2005). Susceptibility of juvenile *Crassostrea gigas* and resistance of *Panope abrupta* to *Mikrocytos mackinii*. *Journal of Invertebrate Pathology* **88**, 95–99.
- Bower, S. M. and Blackbourn, J.** (2003). Geoduck clam (*Panopea abrupta*): anatomy, histology, development, pathology, parasites and symbionts. *Fisheries and Oceans, Canada Ottawa, Canada*. <http://www.pac.dfo-mpo.gc.ca/science/species-especes/shellfish-coquillages/geopath/intro-eng.html>
- Bower, S. M., Blackbourn, J. and Meyer, G. R.** (1998). Distribution, prevalence, and pathogenicity of the protozoan *Perkinsus qugwadi* in Japanese scallops, *Patinopecten yessoensis*, cultured in British Columbia, Canada. *Canadian Journal of Zoology* **76**, 954–959.
- Bower, S. M., Blackbourn, J., Meyer, G. R. and Nishimura, D. J. H.** (1992). Diseases of cultured Japanese scallops (*Patinopecten yessoensis*) in British Columbia, Canada. *Aquaculture* **107**, 201–210.
- Bower, S. M., McGladdery, S. E. and Price, I. M.** (1994). Synopsis of infectious diseases and parasites of commercially exploited shellfish. *Annual Review of Fish Diseases* **4**, 1–199.
- Bower, S. M. and Meyer, G. M.** (2005). Synopsis of infectious diseases and parasites of commercially exploited shellfish: *Labyrinthuloides haliotidis* of abalone. <http://www.dfo-mpo.gc.ca>

- Bower, C. E., Turner, D. T. and Biever, R. C. (1987). A standardized method of propagating the marine fish parasite, *Amyloodinium ocellatum*. *Journal of Parasitology* **73**, 85–88.
- Bragoni, G., Romestand, B. and Trilles, J.-P. (1984). Parasitoses à cymothoïdiens chez le loup, *Dicentrarchus labrax* (Linnaeus, 1758) en élevage. I. Ecologie parasitaire dans le cas de l'Étang de Diana (Haute Corse) (Isopoda, Cymothoïdidae). *Crustaceana* **47**, 44–51.
- Braid, B. A., Moore, J. D., Robbins, T. T., Hedrick, R. P., Tjeerdema, R. S. and Friedman, C. S. (2005). Health and survival of red abalone, *Haliotis rufescens*, under varying temperature, food supply, and exposure to the agent of withering syndrome. *Journal of Invertebrate Pathology* **89**, 219–231.
- Branson, E., Riaza, A. and Alvarez-Pellitero, P. (1999). Myxosporean infection causing intestinal disease in farmed turbot, *Scophthalmus maximus* (L.) (Teleostei: Scophthalmidae). *Journal of Fish Diseases* **22**, 395–399.
- Bravo, S. (2003). Sea lice in Chilean salmon farms. *Bulletin of the European Association of Fish Pathologists* **23**, 197–200.
- Breber, P. (1985). On-growing of the carpet-shell clam (*Tapes decussatus* L.): two years' experience in Venice Lagoon. *Aquaculture* **44**, 51–56.
- Bristow, G. A. and Berland, B. (1991). The effect of long-term low level, *Eubothrium* sp. (Cestoda: Pseudophyllidae) infection on growth of farmed salmon (*Salmo salar* L.). *Aquaculture* **98**, 325–330.
- Bron, J. E., Somerville, C., Wootton, R. and Rae, G. H. (1993). Fallowing of marine Atlantic salmon farms as a method for the control of sea lice. *Journal of Fish Diseases* **16**, 487–493.
- Bruno, D. W., Collins, C. M., Cunningham, C. O. and MacKenzie, K. (2001). *Gyrodactyloides bychozskii* (Monogenea: Gyrodactylidae) from sea-caged Atlantic salmon in Scotland: occurrence and ribosomal RNA sequence analysis. *Diseases of Aquatic Organisms* **45**, 191–196.
- Buck, B. H., Thielges, D. W., Walter, U., Nehls, G. and Rosenthal, H. (2005). Inshore-offshore comparison of parasite infestation in *Mytilus edulis*: implications for open ocean aquaculture. *Journal of Applied Ichthyology* **21**, 107–113.
- Bunkley-Williams, L. and Williams, E. H., Jr. (2006). New records of parasites for culture Cobia, *Rachycentron canadum* (Perciformes: Rachycentridae) in Puerto Rico. *Revista de Biología Tropical* **54**, 1–7.
- Burreson, E. M. (1996). Epizootiology of *Perkinsus marinus* disease of oysters in Chesapeake Bay, with emphasis on data since 1985. *Journal of Shellfish Research* **15**, 17–34.
- Burreson, E. M. and Ragone Calvo, L. M. (1996). Epizootiology of *Perkinsus marinus* disease of oysters in Chesapeake Bay, with emphasis on data since 1985. *Journal of Shellfish Research* **15**, 17–34.
- Burreson, E. M., Stokes, N. A. and Friedman, C. S. (2000). Increased virulence in an introduced pathogen: *Haplosporidium nelsoni* (MSX) in the Eastern oyster *Crassostrea virginica*. *Journal of Aquatic Animal Health* **12**, 1–8.
- Bustos, P. A., Young, N. J., Rozas, M. A., Bohle, H. M., Ildefonso, R. S., Morrison, R. N. and Nowak, B. F. (2011). Amoebic gill disease (AGD) in Atlantic salmon (*Salmo salar*) farmed in Chile. *Aquaculture* **310**, 281–288.
- Cabral, P. (1989a). Problems and perspectives of the pearl oyster aquaculture in French Polynesia. Workshop on Advances in Tropical Aquaculture, Tahiti, 20th February–4th March 1989. Aquacop. Infremer. Actes de Colloque **9**, 57–66.
- Cabral, P. (1989b). Some aspects of the abnormal mortalities of the pearl oysters, *Pinctada margaritifera* in the Tuamotu Archipelago, (French Polynesia). *Advances in Tropical Aquaculture, Tahiti (French Polynesia)*, 20 February–4 March 1989.
- Cáceres Martínez, J., Álvarez Tinajero, C., Guerrero Rentería, Y. and González Avilés, J. G. (2000). Rickettsiales-like prokaryotes in cultured and natural populations of the red abalone *Haliotis rufescens*, blue abalone, *Haliotis fulgens*, the yellow abalone *Haliotis corrugata* from Baja California, Mexico. *Journal of Shellfish Research* **19**, 503.
- Caffara, M., Marcer, F., Florio, D., Quaglio, F. and Fioravanti, M. L. (2003). Heart infection due to *Henneguya* sp. (Myxozoa, Myxosporea) in gilthead sea bream (*Sparus aurata*) cultured in Italy. *Bulletin of the European Association of Fish Pathologists* **23**, 108–112.
- Caffara, M., Quaglio, F., Marcer, F., Florio, D. and Fioravanti, M. L. (2010). Intestinal microsporidiosis in European seabass (*Dicentrarchus labrax* L.) farmed in Italy. *Bulletin of the European Association of Fish Pathologists* **30**, 237–240.
- Callaway, R., Shinn, A. P., Grenfell, S. E., Bron, J. E., Burnwell, G., Cook, E. J., Crumlish, M., Culloty, S., Davidson, K., Ellis, R. P., Flynn, K. J., Fox, C., Green, D. M., Hays, G. C., Hughes, A. D., Johnston, E., Lowe, C. D., Lupatsch, I., Malham, S., Mendzil, A. F., Nickell, T., Pickerel, T., Rowley, A. F., Stanley, M. S., Tocher, D. R., Turnbull, J. F., Webb, G., Wootton, E. and Shields, R. J. (2012). Review of climate change impacts on marine aquaculture in the UK and Ireland. *Aquatic Conservation: Marine and Freshwater Ecosystems* **22**, 389–421.
- Calvo, L. R., Walker, J. G. and Burreson, E. M. (1998). Prevalence and distribution of QPX, Quahog Parasite Unknown, in hard clams *Mercenaria mercenaria* in Virginia, USA. *Diseases of Aquatic Organisms* **33**, 209–219.
- Calvo-Ugarteburu, G. and McQuaid, C. D. (1998). Parasitism and introduced species: epidemiology of trematodes in the intertidal *Perna perna* and *Mytilus galloprovincialis*. *Journal of Experimental Marine Biology and Ecology* **220**, 47–65.
- Canestri-Trotti, G., Baccarani, E. M., Paesanti, F. and Turolla, E. (2000). Monitoring of infections by protozoa of the genera *Nematopsis*, *Perkinsus* and *Porospora* in the smooth venus clam *Callista chione* from the North-Western Adriatic Sea (Italy). *Diseases of Aquatic Organisms* **42**, 157–161.
- Carballal, M. J., Iglesias, D., Santamarina, J., Ferro-Soto, B. and Villalba, A. (2001). Parasites and pathologic conditions of the cockle *Cerastoderma edule* populations of the Coast of Galicia (NW Spain). *Journal of Invertebrate Pathology* **78**, 87–97.
- Carella, F., Aceto, S., Marrone, R., Maiolino, P. and De Vico, G. (2010). *Marteilia refringens* infection in cultured and natural beds of mussels (*Mytilus galloprovincialis*) along the Campanian coast (Tirrenian Sea, South of Italy). *Bulletin of the European Association of Fish Pathologists* **30**, 189–196.
- Carr, W. H. (1996). Pathogenic organisms of penaeid shrimp in the Hawaiian Islands. *Bishop Museum Occasional Papers* **46**, 15–18.
- Carvajal, J., Gonzalez, L. and George-Nascimento, M. (1998). Native sea lice (Copepoda: Caligidae) infestation of salmonids reared in netpen systems in southern Chile. *Aquaculture* **166**, 241–246.
- Chambon, C., Legeay, A., Durrieu, G., Gonzalez, P., Ciret, P. and Massabuau, J. C. (2007). Influence of the parasite worm *Polydora* sp. on the behaviour of the oyster *Crassostrea gigas*: a study of the respiratory impact and associated oxidative stress. *Marine Biology* **152**, 329–338.
- Chang, P. and Wang, Y. (2000). Studies on the caligusiasis and benedeniiasis of marine cage cultured fish in Pingtung area of Taiwan. In *Proceedings of the First International Symposium on Cage Aquaculture in Asia, 2000* (ed. Liao, I. C. and Lin, C. K.), p. 292. Asian Fisheries Society.
- Charlier, J., Voort van der, M., Hogeveen, H. and Vercruyse, J. (2012). ParaCalc® – a novel tool to evaluate the economic importance of worm infections on the dairy farm. *Veterinary Parasitology* **184**, 204–211.
- Chen, S. C., Kou, R. J., Wu, C. T., Wang, P. C. and Su, F. Z. (2001). Mass mortality with a *Sphaerospora*-like myxosporidean infestation in juvenile cobia, *Rachycentron canadum* (L.), marine cage cultured in Taiwan. *Journal of Fish Diseases* **24**, 189–195.
- Chungti, T. and Zhenzu, X. (1979). The 'black root' disease of the razor clam in estuary of Jiulong River, Fujian. *Acta Zoologica Sinica* **4**, 006.
- Colorni, A. and Burgess, P. (1997). *Cryptocaryon irritans* Brown 1951, the cause of 'white spot disease' in marine fish: an update. *Aquarium Sciences and Conservation* **1**, 217–238.
- Colquitt, S. E., Munday, B. L. and Daintith, M. (2001). Pathological findings in southern bluefin tuna, *Thunnus maccoyii* (Castelnau), infected with *Cardicola forsteri* (Cribb, Daintith and Munday, 2000) (Digenea: Sanguicolidae), a blood fluke. *Journal of Fish Diseases* **24**, 225–229.
- Cook, H. L. (1971). Fungi parasitic on shrimp. *FAO Aquaculture Bulletin* **3**, 13.
- Cook, T., Folli, M., Klinck, J., Ford, S. and Miller, J. (1998). The relationship between increasing sea-surface temperature and the northward spread of *Perkinsus marinus* (Dermo) disease epizootics in oysters. *Estuarine, Coastal and Shelf Science* **46**, 587–597.
- Costello, M. J. (2009). The global economic cost of sea lice to the salmonid farming industry. *Journal of Fish Diseases* **32**, 115–118.
- Crespo, S., Grau, A. and Padrós, F. (1992). Sanguinicoliiasis in the cultured amberjack *Seriola dumerili* Risso, from the Spanish Mediterranean area. *Bulletin of the European Association of Fish Pathologists* **12**, 157–159.
- Crespo, S., Grau, A. and Padrós, F. (1994). The intensive culture of 0-group amberjack in the western Mediterranean is compromised by disease problems. *Aquaculture International* **2**, 262–265.
- Crosby, M. P. and Roberts, C. F. (1990). Seasonal infection intensity cycle of the parasite *Perkinsus marinus* (and an absence of *Haplosporidium* spp.) in oysters from a South Carolina salt marsh. *Diseases of Aquatic Organisms* **9**, 149–155.
- Cruz-Lacierda, E. R., Maeni, Y., Pineda, A. J. T. and Matey, V. E. (2004). Mass mortality of hatchery-reared milkfish (*Chanos chanos*) and mangrove red snapper (*Lutjanus argentimaculatus*) caused by *Amyloodinium ocellatum* (Dinoflagellida). *Aquaculture* **236**, 85–94.
- Cruz-Lacierda, E. R., Pagador, G. E., Yamamoto, A. and Nagasawa, K. (2011). Parasitic caligid copepods of farmed marine fishes in the Philippines. In *Diseases in Asian Aquaculture VII* (ed. Bondad-Reantaso, M. G., Jones, J. B., Corsin, F. and Aoki, T.), pp. 53–62. Fish Health Section, Asian Fisheries Society, Selangor, Malaysia, 385 pp.

- Cruz-Lacierda, E. R., Toledo, J. D., Tan-Fermin, J. D. and Bureson, E. M.** (2000). Marine leech (*Zeylanicobdella arugamensis*) infestation in cultured orange-spotted grouper, *Epinephelus coioides*. *Aquaculture* **185**, 191–196.
- Culurgioni, J., D'Amico, V., De Murtas, R., Trotti, G. C. and Figus, V.** (2006). Parasitological monitoring of commercial native bivalves from St. Gilla lagoon (Sardinia, South Western Mediterranean). *Ittiopatologia* **3**, 243–252.
- Dahl, S. F., Thiel, J. and Allam, B.** (2010). Field performance and QPX disease progress in cultured and wild-type strains of *Mercenaria mercenaria* in New York waters. *Journal of Shellfish Research* **29**, 83–90.
- DaRos, L. and Canzonier, W. J.** (1985). *Perkinsus*, a protistan threat to bivalve culture in the Mediterranean basin. *Bulletin of the European Association of Fish Pathologists* **5**, 23–25.
- Davey, J. T.** (1989). *Mytilicola intestinalis* (Copepoda: Cyclopoida): a ten year survey of infested mussels in a Cornish estuary, 1978–1988. *Journal of the Marine Biological Association of the UK* **69**, 823–836.
- Davey, J. T. and Gee, J. M.** (1988). *Mytilicola intestinalis*, a copepod parasite of blue mussels. Diseases in marine bivalve molluscs. *American Fisheries Society Special Publication* **18**, 64–73.
- Dégremont, L., Bédier, E. and Boudry, P.** (2010). Summer mortality of hatchery-produced Pacific oyster spat (*Crassostrea gigas*). II. Response to selection for survival and its influence on growth and yield. *Aquaculture* **299**, 21–29.
- Dethlefsen, V.** (1985). *Mytilicola intestinalis* parasitism. In *Fiches d'Identification des Maladies et Parasites des Poissons, Crustacés et Mollusques*, No. 24 (ed. Sindermann, C. J.), pp. 1–4. ICES, Copenhagen.
- Deveney, M. R., Bayly, T. J., Johnston, C. J. and Nowak, B. F.** (2005). A parasite survey of farmed southern bluefin tuna, *Thunnus maccoyii* (Castelnau). *Journal of Fish Diseases* **28**, 279–284.
- Deveney, M. R., Chisholm, L. A. and Whittington, I. D.** (2001). First published record of the pathogenic monogenean parasite *Neobenedenia melleni* (Capsalidae) from Australia. *Diseases of Aquatic Organisms* **46**, 79–82.
- Dezfuli, B. S., Giari, L., Simoni, E., Menegatti, M., Shinn, A. P. and Manera, M.** (2007). Gill histopathology of cultured *Dicentrarchus labrax* (L.) infected with *Diplectanum aequans* (Wagener, 1857) Diesing, 1958 (Diplectanidae: Monogenea). *Parasitology Research* **100**, 707–713.
- Diamant, A.** (1992). A new pathogenic histozoic *Myxidium* (Myxosporea) in cultured gilt-head sea bream *Sparus aurata* L. *Bulletin of the European Association of Fish Pathologists* **12**, 64–66.
- Diamant, A.** (1998a). Red drum *Sciaenops ocellatus* (Sciaenidae), a recent introduction to Mediterranean mariculture, is susceptible to *Myxidium leei* (Myxosporea). *Aquaculture* **162**, 33–39.
- Diamant, A.** (1998b). *Brooklynella hostilis* (Hartmannulidae), a pathogenic ciliate from the gills of maricultured sea bream. *Bulletin of the European Association of Fish Pathologists* **18**, 33–36.
- Diggles, B. and Hutson, K. S.** (2005). Diseases of kingfish (*Seriola lalandi*) in Australasia. *Aquaculture Health International* **3**, 12–14.
- Diggles, B. K., Hine, P. M., Handley, S. and Boustead, N. C.** (2002). A handbook of diseases of importance to aquaculture in New Zealand. *NIWA Science and Technology Series No. 49*.
- Diggles, B. K. and Oliver, M.** (2005). Diseases of cultured paua (*Haliotis iris*) in New Zealand. In *Diseases in Asian Aquaculture V* (ed. Walker, P., Lester, R. and Bondad-Reantaso, M. G.), pp. 275–287. Fish Health Section, Asian Fisheries Society, Manila.
- Dong, J., Jiang, L.-X., Tan, K.-F., Liu, H.-Y., Purcell, J. E., Li, P.-J. and Ye, C.-C.** (2009). Stock enhancement of the edible jellyfish (*Rhopilema esculentum* Kishinouye) in Liaodong Bay, China: a review. *Hydrobiologia* **616**, 113–118.
- Douglas-Helders, G. M., Tan, C., Carson, J. and Nowak, B. F.** (2003). Effects of copper-based antifouling treatment on the presence of *Neoparamoeba pemaquidensis* Page, 1987 on nets and gills of reared Atlantic salmon (*Salmo salar*). *Aquaculture* **221**, 13–22.
- Dragesco, A., Dragesco, J., Coste, F., Gasc, C., Romestand, B., Raymond, J. C. and Bouix, G.** (1995). *Philasterides dicentrarchi*, n. sp. (Ciliophora, Scuticociliata), a histophagous opportunistic parasite of *Dicentrarchus labrax* (Linnaeus, 1758), a reared marine fish. *European Journal of Protistology* **31**, 327–340.
- Dungan, C. F., Carnegie, R. B., Hill, K. M., McCollough, C. B., Laramore, S. E., Kelly, C. J., Stokes, N. A. and Scarpa, J.** (2012). Diseases of oysters *Crassostrea ariakensis* and *C. virginica* reared in ambient waters from the Choptank River, Maryland and the Indian River Lagoon, Florida. *Diseases of Aquatic Organisms* **101**, 173–183.
- Dyková, I. and Figueras, A.** (1994). Histopathological changes in turbot *Scophthalmus maximus* due to a histophagous ciliate. *Diseases of Aquatic Organisms* **18**, 5–9.
- Dyková, I., Figueras, A., Novoa, B. and Casal, J. F.** (1998). *Paramoeba* sp., an agent of amoebic gill disease of turbot, *Scophthalmus maximus*. *Diseases of Aquatic Organisms* **33**, 137–141.
- Dyková, I., Lom, J. and Fajer, E.** (1988). A new haplosporean infecting the hepatopancreas in the penaeid shrimp, *Penaeus vannamei*. *Journal of Fish Diseases* **11**, 15–22.
- Egusa, S.** (1983). Disease problems in Japanese yellowtail, *Seriola quinqueradiata*, culture: a review. *Rapports et Procès-verbaux des Réunion Conseil International pour l'Exploration de la Mer* **182**, 10–18.
- Egusa, S.** (1985). *Myxobolus buri* sp. n. (Myxosporea: Bivalvulida) parasitic in the brain of *Seriola quinqueradiata* Temminch et Schlegel. *Fish Pathology* **19**, 239–244.
- Egusa, S., Hatai, K. and Fujimaki, Y.** (1988). Notes on *Microsporidium* species, the etiological agent of 'Beko' disease in Red Sea bream juveniles, *Pagrus major*. *Fish Pathology* **23**, 263–267.
- Egusa, S. and Nakajima, K.** (1978). Kudoasis of cultured yellowtail. *Fish Pathology* **13**, 1–7.
- Egusa, S. and Nakajima, K.** (1980). *Kudoa amamiensis* n. sp. (Myxosporea: Multivalvulida) found in cultured yellowtails and wild damselfishes from Amami-Oshima and Okinawa, Japan. *Bulletin of the Japanese Society of Scientific Fisheries* **46**, 1193–1198.
- Egusa, S. and Shiomitsu, T.** (1983). Two new species of the genus *Kudoa* (Myxosporea: Multivalvulida) from marine cultured fishes in Japan. *Fish Pathology* **18**, 163–171.
- Ellis, A. E. and Wootten, R.** (1978). Costiasis of Atlantic salmon, *Salmo salar* L. smolts in seawater. *Journal of Fish Diseases* **1**, 389–393.
- Elston, R. A.** (1980). Ultrastructural aspects of a serious disease of hatchery reared larval oysters, *Crassostrea gigas* Thünin. *Journal of Fish Diseases* **3**, 1–10.
- Elston, R. A.** (1993). Infectious diseases of the Pacific oyster, *Crassostrea gigas*. *Annual Review of Fish Diseases* **3**, 259–276.
- Elston, R. A., Kent, M. L. and Harrell, L. H.** (1987). An intranuclear microsporidium associated with acute anemia in the chinook salmon. *Journal of Protozoology* **34**, 274–277.
- Ernst, I., Whittington, I., Corneillie, S. and Talbot, C.** (2002). Monogenean parasites in sea-cage aquaculture. *Austasia Aquaculture* February/March, 46–48.
- Eydal, M., Kristmundsson, A. and Bambir, S. H.** (2010). Pseudobranchial X-cell pseudotumors in young wild and farmed Atlantic cod *Gadus morhua* in Iceland. *Diseases of Aquatic Organisms* **91**, 83–88.
- Fajer-Ávila, E. J., Abdo-de la Parra, I., Aguilar-Zarate, G., Contreras-Arce, R., Zaldivar-Ramirez, J. and Betancourt-Lozano, M.** (2003). Toxicity of formalin to bullseye puffer fish (*Sphoeroides annulatus* Jenyns) and its effectiveness to control ectoparasites. *Aquaculture* **223**, 41–50.
- Fajer-Ávila, E. J., Martínez-Rodríguez, I., Abdo de la Parra, M. I., Álvarez-Lajonchere, L. and Betancourt-Lozano, M.** (2008). Effectiveness of freshwater treatment against *Lepeophtheirus simplex* (Copepoda: Caligidae) and *Neobenedenia* sp. (Monogenea: Capsalidae), skin parasites of bullseye puffer fish, *Sphoeroides annulatus* reared in tanks. *Aquaculture* **284**, 277–280.
- FAO (2014a)**. Cultured aquatic species information program: *Dicentrarchus labrax* (Linnaeus, 1758). http://www.fao.org/fishery/culturedspecies/Dicentrarchus_labrax/en
- FAO (2014b)**. Cultured aquatic species information program: *Psetta maxima* (Linnaeus, 1758). http://www.fao.org/fishery/culturedspecies/Psetta_maxima/en
- FAO (2014c)**. Cultured aquatic species information program: *Rachycentron canadum* (Linnaeus, 1766). http://www.fao.org/fishery/culturedspecies/Rachycentron_canadum/en
- FAO (2014d)**. Cultured aquatic species information program: *Sparus aurata* (Linnaeus, 1758). http://www.fao.org/fishery/culturedspecies/Sparus_aurata/en
- FAO (2014e)**. Cultured aquatic species information program: *Scylla serrata* (Forsskal, 1755). http://www.fao.org/fishery/culturedspecies/Scylla_serrata/en
- FAO FishStatJ** (2013). Fisheries and Aquaculture Department, Statistics and Information Service FishStatJ: Universal software for fishery statistical time series. Copyright 2011. Version 2.1.0. (March, 2013). <http://www.fao.org/fishery/statistics/software/fishstat/en>
- FAO Globefish** (2013). Salmon – June 2013. <http://www.globefish.org/salmon-june-2013.html>
- Farley, C. A., Wolf, P. H. and Elston, R. A.** (1988). A long-term study of 'microcell' disease in oysters with a description of a new genus, *Mikrocytos* (g.n.) and two new species *Mikrocytos mackini* (sp. n.) and *Mikrocytos roughleyi* (sp. n.). *US National Marine Fish Service Bulletin* **86**, 581–593.
- Feehan, C., Johnson-MacKinnon, J., Scheibling, R. E., Lauzon-Guay, J. S. and Simpson, A. G. B.** (2013). Validating the identity of

- Paramoeba invadens*, the causative agent of recurrent mass mortality of sea urchins in Nova Scotia, Canada. *Diseases of Aquatic Organisms* **103**, 209–227.
- Fenner, R. M.** (1998). Cyanide collection: deadly truths for reefs, fishermen, and aquarists. *The Conscientious Marine Aquarist*, pp. 165–173. Microcosm, Ltd., Shelburne, VT.
- Ferguson, H. W., Hicks, B. D., Lynn, D. H., Ostland, V. E. and Bailey, J.** (1987). Cranial ulceration in Atlantic salmon *Salmo salar* associated with *Tetrahymena* sp. *Diseases of Aquatic Organisms* **2**, 191–195.
- Figueras, A., Novoa, A., Santarem, B., Martínez, E., Alvarez, J. M., Toranzo, A. E. and Dyková, I.** (1992). *Tetramicra brevisfilum*, a potential threat to farmed turbot *Scophthalmus maximus*. *Diseases of Aquatic Organisms* **14**, 127–135.
- Figueras, A. J., Jardon, C. F. and Caldas, J. R.** (1991). Diseases and parasites of rafter mussels (*Mytilus galloprovincialis* Lmk): preliminary results. *Aquaculture* **99**, 17–33.
- Findlay, V. L., Helders, M., Munday, B. L. and Gurney, R.** (1995). Demonstration of resistance to reinfection with *Paramoeba* sp. by Atlantic salmon, *Salmo salar* L. *Journal of Fish Diseases* **18**, 639–642.
- Ford, S. E.** (1985). Effects of salinity on survival of the MSX parasite *Haplosporidium nelsoni* (Haskin, Stauber and Mackin) in oysters. *Journal of Shellfish Research* **5**, 85–90.
- Ford, S. E.** (2011). SSO Disease of oysters caused by *Haplosporidium costale*. ICES Identification Leaflets for Diseases and Parasites of Fish and Shellfish, Leaflet No. 39. International Council for the Exploration of the Seas, Copenhagen, Denmark.
- Ford, S. E. and Haskin, H. H.** (1982). History and epizootiology of *Haplosporidium nelsoni* (MSX), an oyster pathogen in Delaware Bay, 1957–1980. *Journal of Invertebrate Pathology* **40**, 118–141.
- Ford, S. E. and Haskin, H. H.** (1987). Infection and mortality patterns in strains of oysters *Crassostrea virginica* selected for resistance to the parasite *Haplosporidium nelsoni* (MSX). *Journal of Protozoology* **73**, 368–376.
- Ford, S. E., Kraeuter, J. N., Barber, R. D. and Mathis, G.** (2002). Aquaculture-associated factors in QPX disease of hard clams: density and seed source. *Aquaculture* **208**, 23–38.
- Foster, C. and Percival, S.** (1988). Paramoebic gill disease. Occurrence of *Paramoeba* in Tasmania. *Saltas Aquanote No. 15*, May Salmon Enterprises of Tasmania Pty Ltd, Dover, Tasmania, Australia.
- Franco-Sierra, A., Sitjà-Bobadilla, A. and Alvarez-Pellitero, P.** (1997). *Ichthyophonus* infections in cultured marine fish from Spain. *Journal of Fish Biology* **51**, 830–839.
- Frasca, S., Poynton, S. L., West, A. B. and Van Kruiningen, H. J.** (1998). Epizootiology, pathology, and ultrastructure of the myxosporean associated with parasitic encephalitis of farmed Atlantic salmon *Salmo salar* in Ireland. *Diseases of Aquatic Organisms* **32**, 211–225.
- Frasca, S., Jr., Linfert, D. R., Tsongalis, G. J., Gorton, T. S., Garmendia, A. E., Hedrick, R. P., West, A. B. and Van Kruiningen, H. J.** (1999). Molecular characterization of the myxosporean associated with parasitic encephalitis of farmed caged Atlantic salmon *Salmo salar* in Ireland. *Diseases of Aquatic Organisms* **35**, 221–233.
- Frenzl, B., Migaud, H., Fjellidal, P. G., Shinn, A. P., Taylor, J. F., Richards, R. H., Glover, K. A., Cockerill, D. and Bron, J. E.** (2013). Triploid and diploid Atlantic salmon show similar susceptibility to infection with salmon lice *Lepeophtheirus salmonis*. *Pest Management Science* **70**, 982–988.
- Frenzl, B., Stien, L. H., Cockerill, D., Oppedal, F., Richards, R. H., Shinn, A. P., Bron, J. E. and Migaud, H.** (2014). Manipulation of farmed Atlantic salmon swimming behaviour through the adjustment of lighting and feeding regimes as a tool for salmon lice control. *Aquaculture* **24–425**, 183–188.
- Friedman, C. S., Andree, K. B., Beauchamp, K. A., Moore, J. D., Robbins, T. T., Shields, J. D. and Hedrick, R. P.** (2000). 'Candidatus *Xenohaliotis californiensis*', a newly described pathogen of abalone, *Haliotis* spp., along the west coast of North America. *International Journal of Systematic and Evolutionary Microbiology* **50**, 847–855.
- Friedman, C. S., Cloney, D. F., Manzer, D. and Hedrick, R. P.** (1991). Haplosporidiosis of the Pacific oyster, *Crassostrea gigas*. *Journal of Invertebrate Pathology* **58**, 367–372.
- Friedman, C. S., Thomson, M., Chun, C., Haaker, P. L. and Hedrick, R. P.** (1997). Withering syndrome of the black abalone *Haliotis cracherodii* (Leach): water temperature, food availability, and parasites as possible causes. *Journal of Shellfish Research* **16**, 403–411.
- Fujita, S., Yoda, M. and Ugajin, I.** (1968). Control of an endoparasitic copepod, *Caligus spinosus* Yamaguti, on the cultured adult yellowtail. *Fish Pathology* **2**, 122–127 [In Japanese].
- Gabor, L. J., Srivastava, M., Titmarsh, J., Dennis, M., Gabor, M. and Landos, M.** (2011). Cryptosporidiosis in intensively reared barramundi (*Lates calcarifer*). *Journal of Veterinary Diagnostic Investigation* **23**, 383–386.
- García García, J. and García García, B.** (2010). Econometric model of viability/profitability of on-growing sharp snout sea bream (*Diplodus puntazzo*) in sea cages. *Aquaculture International* **18**, 955–971.
- García-Ortega, A., Hernández, C., Abdo-de-la-Parra, I. and González-Rodríguez, B.** (2002). Advances in the nutrition and feeding of the bullseye puffer *Sphoeroides annulatus*. In *Avances en Nutrición Acuicola VI. Memorias del VI Simposium Internacional de Nutrición Acuicola, 3 al 6 de Septiembre del 2002* (ed. Cruz-Suárez, L. E., Ricque-Marie, D., Tapiá-Salazar, M., Gaxiola-Cortés, M. G. and Simoes, N.), pp. 187–196. Cancún, Quintana Roo, México.
- Ghittino, P. S., Bignami, I. S., Annibaldi, A. and Boni, L.** (1980). First record of serious oodiniasis in seabass (*Dicentrarchus labrax*) intensively reared in brackish water. *Rivista Italiana Piscicoltura e Ittiopatologica* **15**, 122–127.
- Gibson-Kueh, S., Thuy, N. T. N., Elliot, A., Jones, J. B., Nicholls, P. K. and Thompson, R. C. A.** (2011). An intestinal *Eimeria* infection in juvenile Asian seabass (*Lates calcarifer*) cultured in Vietnam – a first report. *Veterinary Parasitology* **181**, 106–112.
- Goggin, C. L. and Lester, R. J. G.** (1995). *Perkinsus*, a protistan parasite of abalone in Australia: a review. *Marine and Freshwater Research* **46**, 639–646.
- Golomazou, E., Athanassopoulou, F., Vagianou, S., Sabatakou, O., Tsadilas, H., Rigos, G. and Kokkokiris, L.** (2006). Diseases of White Sea bream (*Diplodus sargus* L.) reared in experimental and commercial conditions in Greece. *Turkish Journal of Veterinary and Animal Sciences* **30**, 389–396.
- Graczyk, T., Conn, D., Marcogliese, D., Graczyk, H. and De Lafontaine, Y.** (2003). Accumulation of human waterborne parasites by zebra mussels (*Dreissena polymorpha*) and Asian freshwater clams (*Corbicula fluminea*). *Parasitology Research* **89**, 107–112.
- Grant, A. N. and Treasurer, J. W.** (1993). The effects of fallowing on caligid infestations on farmed Atlantic salmon (*Salmo salar* L.) in Scotland. In *Pathogens of Wild and Farmed Fish: Sea Lice* (ed. Boxshall, G. A. and Defaye, D.), pp. 255–260. Ellis Horwood Ltd., Chichester, UK.
- Grau, A., Crespo, S., Pastor, E., González, P. and Carbonell, E.** (2003). High infection by *Zeuxapta seriola* (Monogenea: Heteraxinidae) associated with mass mortalities of amberjack *Seriola dumerili* Risso reared in sea cages in the Balearic Islands (Western Mediterranean). *Bulletin of the European Association of Fish Pathologists* **23**, 139–142.
- Gray, A. P., Lucas, I. A. N., Seed, R., and Richardson, C. A.** (1999). *Mytilus edulis chilensis* infested with *Coccomyxa parasitica* (Chlorococcales, Coccomyxaceae). *Journal of Molluscan Research* **65**, 289–294.
- Guo, F. C. and Woo, P. T. K.** (2004). Experimental infections of Atlantic salmon *Salmo salar* with *Spironucleus barkhamus*. *Diseases of Aquatic Organisms* **61**, 59–66.
- Haaker, P. L., Parker, D. O., Togstad, H., Richards, D. V., Davis, G. E. and Friedman, C. S.** (1992). Mass mortality and withering syndrome in black abalone, *Haliotis cracherodii*, in California. In *Abalone of the World: Biology, Fisheries and Culture. Proceedings of the First International Symposium on Abalone* (ed. Shepherd, S. A., Tegner, M. J. and Gusman del Proo, S. A.), pp. 214–224. Cambridge University Press, Cambridge.
- Halpern, B. S., Walbridge, S., Selkoe, K. A., Kappel, C. V., Micheli, F., D'Agrosa, C., Bruno, J. F., Casey, K. S., Ebert, C., Fox, H. E., Fujita, R., Heinemann, D., Lenihan, H. S., Madin, E. M. P., Perry, M. T., Selig, E. R., Spalding, M., Steneck, R. and Watson, R.** (2008). A global map of human impact on marine ecosystems. *Science* **319**, 948–952.
- Han, H.-S., Nam, B.-H., Kang, J.-H., Kim, Y.-K., Jee, Y.-J., Hur, Y.-B. and Yoon, M.-G.** (2012). Genetic variation in wild and cultured populations of the sea squirt *Halocynthia roretzi* inferred from microsatellite DNA analysis. *Fisheries and Aquatic Sciences* **15**, 151–155.
- Harada, T.** (1966). Studies on propagation of yellowtail (*Seriola quinqueradiata* T. & S.). *Bulletin of the Fisheries Laboratory of Kinki University* **1**, 1–275 [in Japanese].
- Harrell, L. W., Elston, R. A., Scott, T. M. and Wilkinson, M. T.** (1986). A significant new systemic disease of net-pen reared chinook salmon *Oncorhynchus tshawytscha* brood stock. *Aquaculture* **55**, 249–262.
- Harrell, L. W. and Scott, T. M.** (1985). *Kudoa thyrstites* (Gilchrist) (Myxosporaea: Multivalvulida) in Atlantic salmon (*Salmo salar* L.). *Journal of Fish Diseases* **8**, 329–332.
- Haskin, H. H. and Ford, S. E.** (1982). *Haplosporidium nelsoni* on Delaware Bay seed oyster beds: a host–parasite relationship along a salinity gradient. *Journal of Invertebrate Pathology* **40**, 388–405.
- Haskin, H. H., Stauber, L. A. and Mackin, J. A.** (1966). *Minchinia nelsoni* n. sp. (Haplosporida, Haplosporidiidae): causative agent of the Delaware Bay oyster spizootic. *Science (NY)* **153**, 1414–1416.

- Hatai, K., Roza, D. and Nakamura, T.** (2000). Identification of lower fungi isolated from larvae of mangrove crab, *Scylla serrata*, in Indonesia. *Mycoscience* **41**, 565–572.
- Hauck, A. K.** (1984). A mortality and associated tissue reactions of chinook salmon, *Oncorhynchus tshawytscha* (Walbaum), caused by the microsporidian *Loma* sp. *Journal of Fish Diseases* **7**, 217–229.
- Hayward, C. J., Aiken, H. M. and Nowak, B. F.** (2008). An epizootic of *Caligus chistos* on farmed southern bluefin tuna *Thunnus maccoyii* off South Australia. *Diseases of Aquatic Organisms* **79**, 57–63.
- Hayward, C. J., Bott, N. J., Itoh, N., Iwashita, M., Okihira, M. and Nowak, B. F.** (2007). Three species of parasites emerging on the gills of mullet, *Argyrosomus japonicus* (Temminck and Schlegel, 1843), cultured in Australia. *Aquaculture* **265**, 27–40.
- Hayward, C. J., Bott, N. J. and Nowak, B. F.** (2009). Seasonal epizootics of sea lice, *Caligus* spp., on southern bluefin, *Thunnus maccoyii* (Castelnau), in a long-term farming trial. *Journal of Fish Diseases* **32**, 101–106.
- Hayward, C. J., Ellis, D., Foote, D., Wilkinson, R. J., Crosbie, P. B. B., Bott, N. J. and Nowak, B. F.** (2010). Concurrent epizootic hyperinfections of sea lice (predominantly *Caligus chistos*) and blood flukes (*Cardicola forsteri*) in ranched Southern bluefin tuna. *Veterinary Parasitology* **173**, 107–115.
- Hedrick, R. P., Groff, J. M. and Baxa, D. V.** (1991). Experimental infections with *Enterocytozoon salmonis* Chilmonczyk, Cox, Hedrick (Microsporea): an intranuclear microsporidium from chinook salmon *Oncorhynchus tshawytscha*. *Diseases of Aquatic Organisms* **10**, 103–108.
- Hégaret, H., Smolowitz, R. M., Sunila, I., Shumway, S. E., Alix, J., Dixon, M. and Wikfors, G. H.** (2010). Combined effects of a parasite, QPX, and the harmful-alga, *Prorocentrum minimum* on northern quahogs, *Mercenaria mercenaria*. *Marine Environmental Research* **69**, 337–344.
- Hervio, D., Bower, S. M. and Meyer, G. R.** (1996). Detection, isolation and experimental transmission of *Mikrocytos mackini*, a microcell parasite of Pacific oysters *Crassostrea gigas* (Thunberg). *Journal of Invertebrate Pathology* **67**, 72–79.
- Hine, P. M., Bower, S. M., Meyer, G. R., Cochenec-Laureau, N. and Berthe, F. C. J.** (2001). Ultrastructure of *Mikrocytos mackini*, the cause of Denman Island disease in oysters *Crassostrea* spp. and *Ostrea* spp. in British Columbia, Canada. *Diseases of Aquatic Organisms* **45**, 215–227.
- Hine, P. M. and Thorne, T.** (2000). A survey of some parasites and diseases of several species of bivalve mollusc in northern western Australia. *Diseases of Aquatic Organisms* **40**, 67–78.
- Hine, P. M., Wakefield, S., Diggles, B. K., Webb, V. L. and Maas, E. W.** (2002). Ultrastructure of a haplosporidian containing Rickettsiae, associated with mortalities among cultured paua *Haliotis iris*. *Diseases of Aquatic Organisms* **49**, 207–219.
- Ho, J.** (1980). Origin and dispersal of *Mytilus edulis* in Japan deduced from its present status of copepod parasitism. *Publications of the Seto Marine Biological Laboratory* **25**, 293–313.
- Ho, J.-S., Kim, I.-H. and Cruz-Lacierda, E. R.** (2004). Sea lice (Copepoda, Caligidae) parasitic on marine cultured and wild fishes of the Philippines. *Journal of the Fisheries Society of Taiwan* **31**, 235–249.
- Ho, J. S. and Lin, C. L.** (2001). *Parapetalus occidentalis* Wilson (Copepoda, Caligidae) parasitic on both wild and farmed cobia (*Rachycentron canadum*) in Taiwan. *Journal of the Fisheries Society of Taiwan* **28**, 305–316.
- Ho, J. S. and Nagasawa, K.** (2001). Why infestation by *Lepeophtheirus salmonis* (Copepoda: Caligidae) is not a problem in the coho salmon farming industry in Japan. *Journal of Crustacean Biology* **21**, 954–960.
- Ho, J. S. and Zheng, G. X.** (1994). *Ostrincola koe* (Copepoda, Myicolidae) and mass mortality of cultured hard clam (*Meretrix meretrix*) in China. *Hydrobiologia* **284**, 169–173.
- Hoffman, G. L.** (1984). Two fish pathogens, *Parvicapsula* sp. and *Mitraspora cyprini* (Myxosporea) new to North America. *Symposia Biologica Hungarica* **23**, 127–135.
- Hofmann, E., Ford, S., Powell, E. and Klinck, J.** (2001). Modelling studies of the effect of climate variability on MSX disease in eastern oyster (*Crassostrea virginica*) populations. *Hydrobiologia* **460**, 195–212.
- Hogans, W. E.** (1989). Mortality of cultured Atlantic salmon, *Salmo salar* L., parr caused by an infection of *Ergasilus labracis* (Copepoda: Poecilostomatoida) in the lower Saint John River, New Brunswick, Canada. *Journal of Fish Diseases* **12**, 529–531.
- Horton, T. and Okamura, B.** (2001). Cymothoid isopod parasites in aquaculture: a review and case study of a Turkish sea bass (*Dicentrarchus labrax*) and sea bream (*Sparus aurata*) farm. *Diseases of Aquatic Organisms* **46**, 181–188.
- Hudson, D., Hudson, N. B. and Pycroft, S. B.** (2001). Mortalities of *Penaeus japonicus* prawns associated with microsporidian infection. *Australian Veterinary Journal* **79**, 504–505.
- Hudson, D. and Lester, R.** (1992). Relationships between water quality parameters and symbiotic ciliates on prawns (*Penaeus japonicus* Bate) in aquaculture. *Aquaculture* **105**, 269–280.
- Hutson, K. S., Ernst, I. and Whittington, I. D.** (2007). Risk assessment for metazoan parasites of yellowtail kingfish *Seriola lalandi* (Perciformes: Carangidae) in South Australian sea-cage aquaculture. *Aquaculture* **271**, 85–99.
- Huys, R., Llewellyn-Hughes, J., Olson, P. D. and Nagasawa, K.** (2006). Small subunit rDNA and Bayesian inference reveal *Pectenophilus ornatus* (Copepoda *incertae sedis*) as highly transformed Mytilicolidae, and support assignment of Chondranchidae and Xarifiidae to Lichomolgoidea (Cyclopoida). *Biological Journal of the Linnean Society* **87**, 403–425.
- Ibieta, P., Tapia, V., Venegas, C., Hausdorf, M. and Takle, H.** (2011). Chilean salmon farming on the horizon of sustainability: review of the development of a highly intensive production, the ISA crisis and implemented actions to reconstruct a more sustainable aquaculture industry. In *Aquaculture and the Environment: a Shared Destiny* (ed. Sladonja, B.), pp. 1–246. Intech, Croatia. ISBN 978-953-307-749-9.
- Iglesias, R., Parama, A., Alvarez, M. F., Leiro, J., Fernandez, J. and Sanmartin, M. L.** (2001). *Philasterides dicentrarchi* (Ciliophora, Scuticociliatida) as the causative agent of scuticociliatosis in farmed turbot *Scophthalmus maximus* in Galicia (NW Spain). *Diseases of Aquatic Organisms* **46**, 47–55.
- Imai, T.** (2005). Sea farming of red sea bream *Pagrus major* (Temminck et Schlegel) in waters off Kanagawa Prefecture, Japan with special reference to stock enhancement effect. *Bulletin of the Kanagawa Prefecture Fisheries Research Institute* **10**, 65–71.
- Isshiki, T., Nagano, T. and Miki, K.** (2007). Occurrence of a monogenean gill parasite *Pseudorhabdosynochus epinepheli* on red spotted grouper *Epinephelus akaara* and its experimental treatment by hydrogen peroxide bathing. *Fish Pathology* **42**, 71–74.
- Itoh, N.** (2009?). *Marteilioides chungmuensis* and its impact on *Crassostrea gigas*. Presentation available at http://wwz.ifremer.fr/crlmollusc/content/download/39925/545564/file/2_Marteilioides_Itoh.pdf
- Itoh, N., Komiyama, H., Ueki, N. and Ogawa, K.** (2004). Early developmental stages of a protozoan parasite, *Marteilioides chungmuensis* (Paramyxea), the causative agent of the ovary enlargement disease in the Pacific oyster, *Crassostrea gigas*. *International Journal for Parasitology* **34**, 1129–1135.
- Itoh, N., Meyer, G. R., Tabata, A., Lowe, G., Abbott, C. L. and Johnson, S. C.** (2013). Rediscovery of the Yesso scallop pathogen *Perkinsus qugwadi* in Canada, and development of PCR tests. *Diseases of Aquatic Organisms* **104**, 83–91.
- Itoh, N., Oda, T., Ogawa, K. and Wakabayashi, H.** (2002). Identification and development of a paramyxean ovarian parasite in the Pacific oyster *Crassostrea gigas*. *Fish Pathology* **37**, 23–28.
- Jee, B. Y., Kim, Y. C. and Park, M.** (2001). Morphology and biology of parasite responsible for scuticociliatosis of cultured olive flounder *Paralichthys olivaceus*. *Diseases of Aquatic Organisms* **47**, 49–55.
- Johnson, S. C., Treasurer, J. W., Bravo, S., Nagasawa, K. and Kabata, Z.** (2004). A review of the impact of parasitic copepods on marine aquaculture. *Zoological Studies* **43**, 229–243.
- Johnstone, A. K.** (1984). *Pathogenesis and life cycle of the myxosporean Parvicapsula sp. infecting marine cultured coho salmon*. PhD dissertation, University of Washington, Seattle, Washington.
- Jung, S.-J., Kitamura, S.-I., Song, J.-Y. and Oh, M.-J.** (2007). *Miamiensis avidus* (Ciliophora: Scuticociliatida) causes systemic infection of olive flounder *Paralichthys olivaceus* and is a senior synonym of *Philasterides dicentrarchi*. *Diseases of Aquatic Organisms* **73**, 227–234.
- Kabata, Z. and Whitaker, D. J.** (1989). *Kudoa thyrsites* (Gilchrist, 1924) (Myxozoa) in the cardiac muscle of Pacific salmon (*Oncorhynchus* spp.) and steelhead trout (*Salmo gairdneri*). *Canadian Journal of Zoology* **67**, 341–342.
- Kaiser, J. B. and Holt, G. J.** (2005). *Species Profile. Cobia*. Southern Regional Aquaculture Center and the United States Department of Agriculture No. 7202, 6 pp.
- Kamaishi, T. and Yoshinaga, T.** (2002). Detection of *Haplosporidium nelsoni* in Pacific oyster *Crassostrea gigas* in Japan. *Fish Pathology* **37**, 193–195.
- Kanchanakhon, S., Chanratchakool, P. and Direkbusarakom, S.** (2001). The impact of health problems on small-scale coastal cage fish culture in Thailand. In *Proceedings of the Asia Regional Scoping Workshop on Primary Aquatic Animal Health Care for Small Scale Rural Aquaculture, Dhaka, Bangladesh, September 1999*.
- Kaneko, J. J., Yamada, R., Brock, J. A. and Nakamura, R. M.** (1988). Infection of tilapia, *Oreochromis mossambicus* (Trewavas), by a marine monogenean, *Neobenedenia melleni* (MacCallum, 1927) Yamaguti, 1963 in Kaneohe Bay, Hawaii, USA, and its treatment. *Journal of Fish Diseases* **11**, 295–300.

- Karagiannis, D. and Angelidis, P.** (2007). Infection of cultured mussels *Mytilus galloprovincialis* by the protozoan *Marteilia* sp. in the Thermaikos Gulf (N Greece). *Bulletin of the European Association of Fish Pathologists* **27**, 131–141.
- Karatayev, A. Y., Mastitsky, S. E., Burlakova, L. E., Karatayev, V. A., Hajduk, M. M. and Conn, D. B.** (2012). Exotic molluscs in the Great Lakes host epizootically important trematodes. *Journal of Shellfish Research* **31**, 885–894.
- Karlsbakk, E., Sæther, P. A., Høstlund, C., Fjellsøy, K. R. and Nylund, A.** (2002). *Parvicapsula pseudobranchicola* n. sp. (Myxozoa), a myxosporidian infecting the pseudobranch of cultured Atlantic salmon (*Salmo salar*) in Norway. *Bulletin of the European Association of Fish Pathologists* **22**, 381–387.
- Katagiri, T., Hosokawa, A., Maita, M., Hirai, M., Takagi, S. and Endo, M.** (2007). A new disease in cultured yellowtail *Seriola quinqueradiata* characterized by encephalomyelitis. *Fish Pathology* **42**, 223–224.
- Katharios, K., Garaffo, M., Sarter, K., Athanassopoulou, F. and Mylonas, C. C.** (2007). A case of high mortality due to heavy infestation of *Ceratomyxa diplodae* in sharpnose sea bream (*Diplodus puntazzo*) treated with reproductive steroids. *Bulletin of the European Association of Fish Pathologists* **27**, 43–47.
- Katharios, P., Hayward, C., Papandroulakis, N. and Divanach, P.** (2006). Pathology of *Lamnelodiscus* spp. (Monogenea) parasitizing the gills of sharpnose seabream and preliminary results of formalin treatment. *Bulletin of the European Association of Fish Pathologists* **126**, 196–201.
- Kent, M. L. and Dawe, S. C.** (1990). Experimental transmission of a plasmacytoid leukemia of chinook salmon (*Oncorhynchus tshawytscha*). *Cancer Research* **50** (Suppl.), 5679–5681.
- Kent, M. L., Elston, R. A., Nerad, T. A. and Sawyer, T. K.** (1987). An *Isonema*-like flagellate (Protozoa: Mastigophora) infection in larval geoduck clams, *Panope abrupta*. *Journal of Invertebrate Pathology* **50**, 221–229.
- Kent, M. L., Sawyer, T. K. and Hedrick, R. P.** (1988). *Paramoeba pemaquidensis* (Sarcocystidophora: Paramoebidae) infestation of the gills of coho salmon *Oncorhynchus kisutch* reared in sea water. *Diseases of Aquatic Organisms* **5**, 163–169.
- Kent, M. L., Elliott, D. G., Groff, J. M. and Hedrick, R. P.** (1989). *Loma salmonae* (Protozoa: Microspora) infections in seawater reared coho salmon, *Oncorhynchus kisutch*. *Aquaculture* **80**, 211–222.
- Kent, M. L., Ellis, J., Fournie, J. W., Dawe, S. C., Bagshaw, J. W. and Whitaker, D. J.** (1992). Systemic hexamitid (Protozoa: Diplomonadida) infection in seawater pen-reared Chinook salmon *Oncorhynchus tshawytscha*. *Diseases of Aquatic Organisms* **14**, 81–89.
- Kent, M. L., Margolis, L. and Fournie, J. W.** (1991). A new eye disease in pen-reared chinook caused by metacystodes of *Gilquinia squali* (Trypanorhyncha). *Journal of Aquatic Animal Health* **3**, 134–140.
- Kent, R. M. L.** (1979). The influence of heavy infestations of *Polydora ciliata* on the flesh content of *Mytilus edulis*. *Journal of the Marine Biological Association of the United Kingdom* **59**, 289–297.
- Kent, R. M. L.** (1981). The effect of *Polydora ciliata* on the shell strength of *Mytilus edulis*. *Journal du Conseil/Conseil Permanent International pour l'Exploration de la Mer* **39**, 252–255.
- Khan, R. A.** (1985). Pathogenesis of *Trypanosoma murmanensis* in marine fish of the northwestern Atlantic following experimental transmission. *Canadian Journal of Zoology* **63**, 2141–2144.
- Khan, R. A.** (2004). Disease outbreaks and mass mortality in cultured Atlantic cod, *Gadus morhua* L., associated with *Trichodina murmanica* (Ciliophora). *Journal of Fish Diseases* **27**, 181–184.
- Khan, R. A.** (2005). Prevalence and influence of *Loma branchialis* (Microspora) on growth and mortality in Atlantic cod (*Gadus morhua*) in coastal Newfoundland. *Journal of Parasitology* **91**, 1230–1232.
- Khan, R. A., Lee, E. M. and Barker, D.** (1990). *Lernaecocera branchialis*: potential pathogen to cod ranching. *Journal of Parasitology* **76**, 913–917.
- Kikuchi, K.** (2006). Present status of research and production of tiger puffer *Takifugu rubripes* in Japan. In *Avances en Nutrición Acuicola. VIII Simposium Internacional De Nutrición Acuicola, 15th–17th Noviembre* (ed. Cruz Suárez, L. E., Ricque Marie, D., Tapia Salazar, M., Nieto López, M. G., Villarreal Cavavos, D. A., Puello Cruz, A. C. and García Ortega, A.), pp. 20–28. Universidad Autónoma de Nuevo León, Monterrey, Nuevo León, México. ISBN 970-694-333-5.
- Kim, H.-J., Park, J. S., Park, K. H., Shin, Y.-K. and Park, K.-I.** (2014). The kinetoplastid parasite *Azumiobodo hoyamushi*, the causative agent of soft tunic syndrome of the sea squirt *Halocynthia roretzi*, resides in the East Sea of Korea. *Journal of Invertebrate Pathology* **116**, 36–42.
- Kim, I. H.** (2004). Poecilostomatoid copepods associated with bivalves in Korea and their distribution. *Zoological Studies* **43**, 187–192.
- Kim, I. H. and Sato, S. I.** (2010). A review of copepods associated with bivalves in Japan, with description of two new species (Crustacea, Copepoda, Cyclopoidea). *Bulletin of the Tohoku University Museum* **9**, 1–22.
- Kim, K. H. and Cho, J. B.** (2000). Treatment of *Microcotyle sebastis* (Monogenea: Polyopisthocotylea) infestation with praziquantel in an experimental cage simulating commercial rockfish *Sebastes schlegelii* culture conditions. *Diseases of Aquatic Organisms* **40**, 229–231.
- Kim, S. M., Cho, J. B., Kim, S. K., Nam, Y. K. and Kim, K. H.** (2004a). Occurrence of scuticociliatosis in olive flounder *Paralichthys olivaceus* by *Phaenocarpa dicentrarchi* (Ciliophora: Scuticociliatida). *Diseases of Aquatic Organisms* **62**, 233–238.
- Kim, S. M., Cho, J. B., Lee, E. H., Kwon, S. R., Kim, S. K., Nam, Y. K. and Kim, K. H.** (2004b). *Pseudocohnlembus persalinus* (Ciliophora: Scuticociliatida) is an additional species causing scuticociliatosis in olive flounder *Paralichthys olivaceus*. *Diseases of Aquatic Organisms* **62**, 239–244.
- Kismohandaka, G., Roberts, W., Hedrick, R. P. and Friedman, C. S.** (1995). Physiological alterations of the black abalone, *Haliotis cracherodii* Leach, with withering syndrome. *Journal of Shellfish Research* **14**, 269–270.
- Koçak, O. and Tatlıdil, F. F.** (2004). Cost analysis in gilthead sea bream (*Sparus aurata* Linnaeus, 1758) and sea bass (*Dicentrarchus labrax* Linnaeus, 1758) production in Milas District-Muğla Province, Turkey. *Turkish Journal of Fisheries and Aquatic Sciences* **4**, 33–38.
- Kolkovski, S. and Sakakura, Y.** (2004). Yellowtail kingfish, from larvae to mature fish – problems and opportunities. In *Avances en Nutrición Acuicola VII. Memorias del VII Simposium Internacional de Nutrición Acuicola, 16th–19th November* (ed. Cruz Suárez, L. E., Ricque Marie, D., Nieto López, M. G., Villarreal, D., Scholz, U. and González, M.), pp. 1–17. Hermosillo, Sonora, México.
- Korringa, P.** (1951). Le *Mytilicola intestinalis* Steuer (Copepoda Parasitica) menace l'industrie moulière en Zelande. *Revue des Travaux de l'Institut des Pêches Maritimes* **17**, 9–13.
- Kristmundsson, Á., Eydal, M. and Helgason, S.** (2006). Progress of co-infections of *Trichodina cooperi* and *T. murmanica* parasitising farmed Atlantic cod *Gadus morhua* juveniles in Iceland. *Diseases of Aquatic Organisms* **71**, 213–223.
- Kubota, S. S. and Takakuwa, M.** (1963). Studies on the diseases of marine cultured fishes: I. General description and preliminary discussion of fish diseases in Mie Prefecture. *Journal of the Faculty of Fisheries, Prefectural University of Mie* **6**, 107–124. [English translation, Fisheries Research Board of Canada Translation Series, Biological Station, Nanaimo, British Columbia, No. 739].
- Kumagai, A., Suto, A., Ito, H., Tanabe, T., Song, J.-T., Kitamura, S.-I., Hirose, E., Kamaishi, T. and Miwa, S.** (2011). Soft tunic syndrome in the edible ascidian *Halocynthia roretzi* is caused by a kinetoplastid protist. *Diseases of Aquatic Organisms* **95**, 153–161.
- Kumagai, A., Suto, A., Ito, H., Tanabe, T., Takahashi, K., Kamaishi, T. and Miwa, S.** (2010). Mass mortality of cultured ascidians *Halocynthia roretzi* associated with softening of the tunic and flagellate-like cells. *Diseases of Aquatic Organisms* **90**, 223–234.
- Kuperman, B. I. and Matey, V. E.** (1999). Massive infestation by *Amyloodinium ocellatum* (Dinoflagellida) of fish in a highly saline lake, Salton Sea, California, USA. *Diseases of Aquatic Organisms* **39**, 65–73.
- Kvenseth, P. G.** (1997). Best current practice for lice control in Norway. *Caligus* **2**, 4–9.
- Kvingedal, R., Owens, L. and Jerry, D. R.** (2006). A new parasite that infects eggs of the mud crab, *Scylla serrata*, in Australia. *Journal of Invertebrate Pathology* **93**, 54–59.
- Lader, P., Dempster, T., Fredheim, A. and Jensen, Ø.** (2008). Current induced net deformations in full-scale sea-cages for Atlantic salmon (*Salmo salar*). *Aquacultural Engineering* **38**, 52–65.
- Lafferty, K. D. and Kuris, A. M.** (1993). Mass mortality of abalone *Haliotis cracherodii* on the California Channel Islands: tests of epidemiological hypotheses. *Marine Ecology – Progress Series* **96**, 239–248.
- Landsberg, J. H., Steidinger, K. A., Blakesley, B. A. and Zondervan, R. L.** (1994). Scanning electron microscope study of dinospores of *Amyloodinium* cf. *ocellatum*, a pathogenic dinoflagellate parasite of marine fish, and comments on its relationship to the Peridinales. *Diseases of Aquatic Organisms* **20**, 23–32.
- La Peyre, M. K., Nickens, A. D., Volety, A. K., Tolley, G. S. and La Peyre, J. F.** (2003). Environmental significance of freshets in reducing *Perkinsus marinus* infection in eastern oysters *Crassostrea virginica*: potential management applications. *Marine Ecology Progress Series* **248**, 165–176.
- Lauckner, G.** (1983). Diseases of Mollusca: Bivalvia. In *Diseases of Marine Animals, Vol. II*. (ed. Kinne, O.), pp. 805–817. Biologische Anstalt Helgoland, Hamburg.
- Laviña, E. M.** (1977). The biology and control of *Caligus* sp. an ectoparasite of the adult milkfish *Chanos chanos* Forskal. SEAFDEC Research Report 1977, pp. 12–13.

- Laviña, E. M.** (1978). A study on certain aspects on the biology and control of *Caligus* sp., an ectoparasite of the adult milkfish *Chanos chanos* (Forsk.). *Fisheries Research Journal Philippines* **3**, 11–24.
- Le Breton, A. and Marques, A.** (1995). Occurrence of a histozoic *Myxidium* infection in two marine cultured species: *Puntazzo puntazzo* C. and *Pagrus major*. *Bulletin of the European Association of Fish Pathologists* **15**, 210–212.
- Lee, M.-K., Cho, B.-Y., Lee, S.-J., Kang, J.-Y., Jeong, H. D., Huh, S. H. and Huh, M.-D.** (2001). Histopathological lesions of Manila clam, *Tapes philippinarum*, from Hadong and Namhae coastal areas of Korea. *Aquaculture* **201**, 199–209.
- Lei, S.** (2000). Studies on pathogenology and histopathology of disease in *Sinonovacula constricta* caused by metacercariae of *Monorchis xiamenensis*. *Journal of Oceanography in Taiwan Strait* **1**, 60–64.
- Leibovitz, L., Elston, R., Lipovsky, V. P. and Donaldson, J.** (1978). A new disease of larval Pacific oysters (*Crassostrea gigas*). *Proceedings of the Annual Meeting – World Mariculture Society* **9**, 603–615.
- Leong, T.-S. and Wong, S.-Y.** (1986). Parasite fauna of seabass, *Lates calcarifer* Bloch, from Thailand and from floating cage culture in Penang, Malaysia. In *Proceedings of the First Asian Fisheries Forum; 26–31 May 1986, Manila, Philippines* (ed. Maclean, J. L., Dizon, L. B. and Hosillos, L. V.), pp. 251–254. Asian Fisheries Society, Manila, Philippines.
- Leong, T.-S. and Wong, S.-Y.** (1988). A comparative study of the parasite fauna of wild and cultured grouper (*Epinephelus malabaricus* Bloch et Schneider) in Malaysia. *Aquaculture* **68**, 203–207.
- Leong, T. S. and Wong, S. Y.** (1990). Parasites of healthy and diseased juvenile grouper (*Epinephelus malabaricus* (Bloch and Schneider)) and seabass (*Lates calcarifer* (Bloch)) in floating cages in Penang, Malaysia. *Asian Fisheries Science* **3**, 319–327.
- Lester, R. J. G.** (1982). *Unicapsula seriola* n. sp. (Myxosporea, Multivalvulida) from Australian yellowtail kingfish *Seriola lalandi*. *Journal of Parasitology* **29**, 584–587.
- Lester, R. J. G.** (1986). Field and laboratory observation on the oyster parasite *Marteilia sydneyi*. In *Parasite Lives* (ed. Cremin, M., Dobson, C. and Moorhouse, D.), pp. 33–40. University of Queensland Press, Brisbane.
- Li, C., Song, S., Liu, Y. and Chen, T.** (2013). *Hematodinium* infections in cultured Chinese swimming crab, *Portunus triuberculatus*, in northern China. *Aquaculture* **396–399**, 59–65.
- Li, M. F., Cornick, J. W. and Miller, R. J.** (1982). Studies of recent mortalities of the sea urchin (*Strongylocentrotus droebachiensis*) in Nova Scotia. *Conseil International de l'Exploration de la Mer CM 1982/L* **46**.
- Li, Y. Y., Xia, X. A., Wu, Q. Y., Liu, W. H. and Lin, Y. S.** (2008). Infection with *Hematodinium* sp. in mud crabs *Scylla serrata* cultured in low salinity water in southern China. *Diseases of Aquatic Organisms* **82**, 145–150.
- Lin, C. L.** (1989). A new species of *Caligus* Copepoda Caligidae parasitic on milkfish *Chanos chanos*. *Crustaceana* **57**, 225–246.
- Lin, C. L. and Ho, J. S.** (1993). Life history of *Caligus epidemicus* Hewitt, parasitic on the tilapia (*Oreochromis mossambicus*) cultured in salt water. In *Pathogens of Wild and Farmed Fish: Sea Lice* (ed. Boxshall, G. A. and Defaye, D.), pp. 5–15. Ellis Horwood, Chichester, UK.
- Lin, C. L. and Ho, J. S.** (1998). Two new species of ergasilid copepods parasitic on fishes cultured in brackish water in Taiwan. *Proceedings of the Biological Society of Washington* **111**, 15–27.
- Lin, C.-L., Ho, J.-S. and Chen, S.-N.** (1994). Two species of *Caligus* (Copepoda: Caligidae) parasitic on black sea bream (*Acanthopagrus schlegelii*) cultured in Taiwan. *Fish Pathology* **29**, 253–264.
- Lin, Q., Yang, M., Huang, Z., Ni, W., Fu, G., Guo, G., Wang, Z. and Huang, X.** (2013). Cloning, expression and molecular characterization of a 14-3-3 gene from a parasitic ciliate, *Cryptocaryon irritans*. *Veterinary Parasitology* **197**, 427–435.
- Lio-Po, G. D. and Sanvictores, E. G.** (1986). Tolerance of *Penaeus monodon* eggs and larvae to fungicides against *Lagenidium* sp. and *Haliphthoros philippinensis*. *Aquaculture* **51**, 161–168.
- Leonart, M., Handlinger, J. and Powell, M.** (2003). Spionid mudworm infestation of farmed abalone (*Haliotis* spp.). *Aquaculture* **221**, 85–96.
- Lodeiros, C., Bolinches, J., Dopazo, C. P. and Toranzo, A. E.** (1987). Bacillary necrosis in hatcheries of *Ostrea edulis* in Spain. *Aquaculture* **65**, 15–29.
- Lo, C. M., Morand, S. and Galzin, R.** (1998). Parasite diversity/host age and size relationship in three coral-reef fishes from French Polynesia. *International Journal for Parasitology* **28**, 1695–1708.
- Lohrmann, K. B.** (2009). How healthy are cultivated scallops (*Argopecten purpuratus*) from Chile? A histopathological survey. *Revista de Biología Marina y Oceanografía* **44**, 35–47.
- Lom, J. and Nigrelli, R. F.** (1970). *Brooklynella hostilis*, n.g., n.sp., a pathogenic ciliophore ciliate in marine fish. *Journal of Protozoology* **17**, 224–232.
- Long, H., Song, W., Chen, J., Gong, J., Ji, D., Hu, X., Ma, H., Zhu, M. and Wang, M.** (2006). Studies on an endoparasitic ciliate *Boveria labialis* (Protozoa: Ciliophora) from the sea cucumber, *Apostichopus japonicus*. *Journal of the Marine Biological Association of the UK* **86**, 823–828.
- Lopez, C., Rajan, P. R., Lin, J. H., Kuo, T. and Yang, H.** (2002). Disease outbreak in sea-farmed cobia (*Rachycentron canadum*) associated with *Vibrio* spp., *Photobacterium damsela* ssp. *piscicida*, monogenean and myxosporean parasites. *Bulletin of the European Association of Fish Pathologists* **22**, 206–211.
- López-Téllez, N. A., Vidal-Martínez, V. M. and Overstreet, R. M.** (2009). Seasonal variation of ectosymbiotic ciliates on farmed and wild shrimps from coastal Yucatan, Mexico. *Aquaculture* **287**, 271–277.
- Lupatsch, I., Santos, G. A., Schrama, J. W. and Verreth, J. A. J.** (2010). Effect of stocking density and feeding level on energy expenditure and stress responsiveness in European sea bass *Dicentrarchus labrax*. *Aquaculture* **298**, 245–250.
- MacKenzie, C. L.** (1996). History of oystering in the United States and Canada, featuring the eight greatest oyster estuaries. *Marine Fisheries Review* **58**, 1–78.
- Madin, J., Chong, V. C. and Hartstein, N. D.** (2010). Effects of water flow velocity and fish culture on net biofouling in fish cages. *Aquaculture Research* **41**, e602–e617.
- Maguire, A. K. and Rogers-Bennett, L.** (2013). An ectoparasitic snail (*Evalea tenuisculpta*) infects red abalone (*Haliotis rufescens*) in northern California. *California Fish and Game* **99**, 80–89.
- Mansell, B., Powell, M. D., Ernst, I. and Nowak, B. F.** (2005). Effects of the gill monogenean *Zeuxapta seriola* (Meserve, 1938) and treatment with hydrogen peroxide on pathophysiology of kingfish, *Seriola lalandi* Valenciennes, 1833. *Journal of Fish Diseases* **28**, 253–262.
- Marine Harvest** (2007). Annual report. 82 pp.
- Marine Harvest** (2013). Annual report. 208 pp.
- Marine Scotland, The Scottish Government** (2012). Amoebic Gill Disease. Topic Sheet No. 96 V1. <http://www.scotland.gov.uk/Resource/0039/00393037.pdf> on the 30/7/2013.
- Martell, D. J., Duhaim, J. and Parsons, G. J.** (eds) (2013). Canadian aquaculture R&D review 2013. *Aquaculture Association of Canada Special Publication* **23**, 100.
- Mathews, C. G. G., Richards, R. H., Shinn, A. P. and Cox, D. I.** (2013). Gill pathology in Scottish farmed Atlantic salmon, *Salmo salar* L., associated with the microsporidian *Desmozoon lepeophtherii* Freeman et Sommerville, 2009. *Journal of Fish Diseases* **36**, 861–869.
- McArdle, J. F.** (1984). *Trichodina* as a cause of mortalities in cage reared rainbow trout (*Salmo gairdneri*) and salmon (*Salmo salar*). *Bulletin of the European Association of Fish Pathologists* **4**, 3–6.
- McGladdery, S. E., Bower, S. M. and Getchell, R. G.** (2006). Diseases and parasites of scallops. *Developments in Aquaculture and Fisheries Science* **35**, 595–650.
- McIlwraith, I.** (2010). Clean Seas takes a bath on kingfish loss. *The Age* (8th September). <http://www.theage.com.au/business/clean-seas-takes-a-bath-on-kingfish-loss-20100907-14zq0.html>
- McLean, E., Salze, G. and Craig, S. R.** (2008). Parasites, diseases and deformities of cobia. *Ribarstvo* **66**, 1–16.
- Merella, P., Cherchi, S., Garippa, G., Fioravanti, M. L., Gustinelli, A. and Salati, F.** (2009). Outbreak of *Sciaenacotyle panceri* (Monogenea) on cage-reared meagre *Argyrosomus regius* (Osteichthyes) from the western Mediterranean Sea. *Diseases of Aquatic Organisms* **86**, 169–173.
- Merella, P., Cherchi, S., Salati, F. and Garippa, G.** (2005). Parasitological survey of sharpnose seabream *Diplodus puntazzo* (Cetti, 1777) reared in sea cages in Sardinia (western Mediterranean). *Bulletin of the European Association of Fish Pathologists* **25**, 140–147.
- Meyers, T. R.** (1981). Endemic diseases of cultured shellfish of Long Island, New York: adult and juvenile American oysters (*Crassostrea virginica*) and hard clams (*Mercenaria mercenaria*). *Aquaculture* **22**, 305–330.
- Michie, I.** (2001). Causes of downgrading in the salmon farming industry. In *Farmed Fish Quality* (ed. Kestin, S. C. and Warris, P. D.), pp. 129–136. Fishing News Books, Oxford.
- Michine, A.** (1999). *Neoheterobothrium* sp. found on Japanese flounder cultured commercially or maintained as spawners. *Research Report of the Shimane Prefectural Center of Cultural Fisheries* **2**, 15–23. [In Japanese].
- Mitchell, C. G.** (1993). *Eubothrium*. *Aquaculture Information Series*, No. 14, 8 pp. Marine Laboratory, Aberdeen, UK. <http://www.marlab.ac.uk>
- Miyazaki, T., Fujimaki, Y. and Katai, K.** (1986). A light and electron microscopic study on epitheliocystis disease in cultured fish. *Bulletin of the Japanese Society for the Science of Fish* **52**, 199–202.
- Mladineo, I.** (2003). Myxosporidian infections in Adriatic cage-reared fish. *Bulletin of the European Association of Fish Pathologists* **23**, 113–122.
- Mladineo, I.** (2006). Parasites of Adriatic cage reared fish. *Acta Adriatica* **47**, 23–28.

- Mladineo, I. and Tudor, M. (2004). Digenea of Adriatic cage-reared northern bluefin tuna (*Thunnus thynnus thynnus*). *Bulletin of the European Association of Fish Pathologists* **24**, 144–152.
- Mladineo, I., Žilič, J. and Čanković, M. (2008). Health survey of Atlantic bluefin tuna, *Thunnus thynnus* (Linnaeus, 1758), reared in Adriatic cages from 2003 to 2006. *Journal of the World Aquaculture Society* **39**, 281–289.
- Mo, T. A. and MacKenzie, K. A. (1991). Occurrence of *Gyrodactyloides bychowskii* Albova, 1948 on gills of sea-caged Atlantic salmon. *Bulletin of the European Association of Fish Pathologists* **11**, 156–158.
- Mo, T. A., Poppe, T. T. and Iversen, L. (1990). Systemic hexamitosis in salt-water reared Atlantic salmon. *Bulletin of the European Association of Fish Pathologists* **10**, 69–70.
- Montero, F., Cuadrado, M., Padros, F., Crespo, S. and Raga, J. A. (2007). *Cryptocaryon irritans* and *Enteromyxum leei*, two threats for the culture of *Diplodus puntazzo* in the Mediterranean. *Bulletin of the European Association of Fish Pathologists* **27**, 242–247.
- Montero, F. E., Crespo, S., Padrós, F., De la Gándara, F., García, A. and Raga, J. A. (2004). Effects of the gill parasite *Zeuxapta seriola* (Monogenea: Heteraxinidae) on the amberjack *Seriola dumerili* Risso (Teleostei: Carangidae). *Aquaculture* **232**, 153–163.
- Montes, J. F., Durfort, M. and García-Valero, J. (2001). Parasitism by the protozoan *Perkinsus atlanticus* favours the development of opportunistic infections. *Diseases of Aquatic Organisms* **46**, 57–66.
- Moore, J. D., Finley, C. A., Robbins, T. T. and Friedman, C. S. (2002). Withering syndrome and restoration of southern California abalone populations. *Reports of California Cooperative Oceanic Fisheries Investigations* **43**, 112–119.
- Moore, J. D., Juhász, C. I., Robbins, T. T. and Vilchis, L. I. (2009). Green abalone, *Haliotis fulgens* infected with the agent of withering syndrome does not express disease signs under a temperature regime permissive for red abalone, *Haliotis rufescens*. *Marine Biology* **156**, 2325–2330.
- Moore, J. D., Robbins, T. T. and Friedman, C. S. (2000). Withering syndrome in farmed red abalone *Haliotis rufescens*: thermal induction and association with a gastrointestinal Rickettsiales-like prokaryote. *Journal of Aquatic Animal Health* **12**, 26–34.
- Moran, J. D. W., Whitaker, D. J. and Kent, M. L. (1999). A review of the myxosporean genus *Kudoa* Meglitsch, 1947, and its impact on the international aquaculture industry and commercial fisheries. *Aquaculture* **172**, 163–196.
- Mori, K., Sato, W., Nomura, T. and Imajima, M. (1985). Infestation of the Japanese scallop *Patinopecten yessoensis* by the boring polychaetes, *Polydora*, on the Okhotsk Sea coast of Hokkaido, especially in Abashiri waters. *Nippon Suisan Gakkaishi* **51**, 371–380. [In Japanese].
- Morrison, R., Nowak, B., Crosbie, P., Adams, M., Bridle, A. and Rise, M. (2006). Insights into amoebic gill disease pathogenesis. *Aquaculture Health International* **4**–5.
- Muhd-Faizul, H. A. H., Kua, B. C. and Leaw, Y. Y. (2012). Caligidae infestation in Asian seabass, *Lates calcarifer*, Bloch 1790 cultured at different salinity in Malaysia. *Veterinary Parasitology* **184**, 68–72.
- Munday, B. L., Foster, C. K., Roubal, F. R. and Lester, R. J. G. (1990). Paramoebic gill infection and associated pathology of Atlantic salmon, *Salmo salar*, and rainbow trout, *Salmo gairdneri*, in Tasmania. In *Pathology in Marine Science* (ed. Perkins, F. O. and Cheng, T. C.), pp. 215–222. Academic Press, London.
- Munday, B. L., O'Donoghue, P. J., Watts, M., Rough, K. and Hawkesford, T. (1997). Fatal encephalitis due to the scuticociliate *Uronema nigricans* in sea-caged, southern bluefin tuna *Thunnus maccoyii*. *Diseases of Aquatic Organisms* **30**, 17–25.
- Munford, J. G., DaRos, L. and Strada, R. (1981). A study on the mass mortality of mussels in the Laguna Veneta. *Journal of the World Mariculture Society* **12**, 186–199.
- Munday, B. L., Zilberg, D. and Findlay, V. (2001). Gill disease of marine fish caused by infection with *Neoparamoeba pemaquidensis*. *Journal of Fish Diseases* **24**, 497–507.
- Munday, B. L., Sawada, Y., Cribb, T. and Hayward, C. J. (2003). Diseases of tunas, *Thunnus* spp. *Journal of Fish Diseases* **26**, 187–206.
- Muraosa, Y., Lawhavinit, O. and Hatai, K. (2006). *Lagenidium thermophilum* isolated from eggs and larvae of black tiger shrimp *Penaeus monodon* in Thailand. *Fish Pathology* **41**, 35–40.
- Muroga, K., Kawatou, K. and Ichizono, H. (1981). Infestation by *Alella macrotrachelus* (Copepoda) of cultured black sea-bream. *Fish Pathology* **16**, 139–144.
- Mustafa, A., Rankaduwa, W. and Campbell, P. (2001). Estimating the cost of sea lice to salmon aquaculture in eastern Canada. *Canadian Veterinary Journal* **42**, 54–56.
- Nagasawa, K., Bresciani, J. and Lützen, J. (1988). Morphology of *Pectenophilus ornatus*, new genus, new species, a copepod parasite of the Japanese scallop *Patinopecten yessoensis*. *Journal of Crustacean Biology* **8**, 31–42.
- Nagasawa, K. and Nagata, M. (1992). Effects of *Pectenophilus ornatus* (Copepoda) on the biomass of cultured Japanese scallop *Patinopecten yessoensis*. *Journal of Parasitology* **78**, 552–554.
- Nakada, M. (2008). Capture-based aquaculture of yellowtail. In *Highly-Based Aquaculture. Global Overview* (ed. Lovatelli, A. and Holthus, P. F.), pp. 199–215. *FAO Fisheries Technical Paper. No. 508*. FAO, Rome.
- Nakajima, K. and Egusa, S. (1972). Studies on a new trypanorhynchian larva, *Callotetrarhynchus* sp., parasitic on cultured yellowtail – XV. Life cycle. *Fish Pathology* **7**, 6–14. [In Japanese, English summary].
- Nakajima, K. and Egusa, S. (1978). *Kudoa pericardialis* n. sp. (Myxosporidea: Chloromyxidae) from cultured yellowtail, *Seriola quinqueradiata* Temminck et Schlegel. *Bulletin of the Japanese Society of Scientific Fisheries* **44**, 117–120.
- Nasr, D. H. (1982). Observations on the mortality of the pearl oyster, *Pinctada margaritifera*, in Dongonab Bay, Red Sea. *Aquaculture* **28**, 271–281.
- Natividad, J. M., Bondad-Reantaso, M. G. and Arthur, J. R. (1986). Parasites of Nile tilapia (*Oreochromis niloticus*) in the Philippines. In *The First Asia Fisheries Forum* (ed. Maclean, J. L., Dizon, L. B. and Hosillos, L. V.), pp. 255–259. Asian Fisheries Society, Manila, Philippines.
- Ngo, T. T. T. and Choi, K.-S. (2004). Seasonal changes of *Perkinsus* and *Cercaria* infections in the Manila clam *Ruditapes philippinarum* from Jeju, Korea. *Aquaculture* **239**, 57–68.
- Noga, E. J. (2010). *Fish Disease: Diagnosis and Treatment*, 2nd Edn. Wiley-Blackwell, Ames, Iowa, USA, p. 536.
- Nowak, B. F., Hayward, C. J., González, L., Bott, N. J. and Lester, R. J. G. (2011). Sea lice infections of salmonids farmed in Australia. *Aquaculture* **320**, 171–177.
- Nunan, L. M., Lightner, D. V., Pantoja, C. R., Stokes, N. A. and Reece, K. S. (2007). Characterization of a rediscovered haplosporidian parasite from cultured *Penaeus vannamei*. *Diseases of Aquatic Organisms* **74**, 67–75.
- Nylund, A., Karlsbakk, E., Sæther, P. A., Koren, C., Larsen, T., Nielsen, B. D., Brøderud, A. E., Høstlund, C., Fjellsøy, K. R., Lervik, K. and Rosnes, L. (2005). *Parvicapsula pseudobranchicola* (Myxosporaea) in farmed Atlantic salmon *Salmo salar*: tissue distribution, diagnosis and phylogeny. *Diseases of Aquatic Organisms* **63**, 197–204.
- Nylund, S., Andersen, L., Sævareid, L., Plarre, H., Watanabe, K., Arnesen, C. E., Karlsbakk, E. and Nylund, A. (2011). Diseases of farmed Atlantic salmon *Salmo salar* associated with infections by the microsporidian *Paramucleospora theridion*. *Diseases of Aquatic Organisms* **94**, 41–57.
- Ogawa, K. (1994). *Anoplodiscus tai* sp. nov. (Monogenea: Anoplodiscidae) from cultured red sea bream *Pagrus major*. *Fish Pathology* **29**, 5–10.
- Ogawa, K. (1996). Marine parasitology with special reference to Japanese fisheries and mariculture. *Veterinary Parasitology* **64**, 95–105.
- Ogawa, K. (1997). Copulation and egg production of the monogenean *Heterobothrium okamotoi*, a gill parasite of cultured tiger puffer (*Takifugu rubripes*). *Fish Pathology* **32**, 219–223.
- Ogawa, K. (1999). *Neoheterobothrium hirame* sp. n. (Monogenea: Didclidophoridae) from the buccal cavity of Japanese flounder *Paralichthys olivaceus*. *Fish Pathology* **34**, 195–201.
- Ogawa, K. (2002). Impacts of didclidophorid monogenean infections on fisheries in Japan. *International Journal for Parasitology* **32**, 373–380.
- Ogawa, K. and Egusa, S. (1986). Two new species of *Paradeontacylix* McIntosh, 1934 (Trematoda: Sanguinicolidae) from the vascular system of a cultured marine fish *Seriola purpurascens*. *Fish Pathology* **21**, 15–19.
- Ogawa, K. and Fukudome, M. (1994). Mass mortality caused by blood fluke (*Paradeontacylix*) among amberjack (*Seriola dumerili*) imported to Japan. *Fish Pathology* **29**, 265–269.
- Ogawa, K. and Inouye, K. (1997). *Heterobothrium* infection of cultured tiger puffer, *Takifugu rubripes* – a field observation. *Fish Pathology* **32**, 15–20.
- Ogawa, K. and Yokoyama, H. (1998). Parasitic diseases of cultured marine fish in Japan. *Fish Pathology* **33**, 303–309.
- Ogawa, K. and Yokoyama, H. (2001). Emaciation disease of cultured tiger puffer *Takifugu rubripes*. *Bulletin of the National Research Institute of Aquaculture* **5** (Suppl.), 65–70.
- Ogawa, K., Bondad-Reantaso, M. G., Fukudome, M. and Wakabayashi, H. (1995). *Neobenedenia girollae* (Hargis, 1955) Yamaguti, 1963 (Monogenea: Capsalidae) from cultured marine fishes of Japan. *Journal of Parasitology* **81**, 223–227.
- Ogawa, K., Miyamoto, J., Wang, H. C., Lo, C. F. and Kou, G. H. (2006). *Neobenedenia girollae* (Monogenea) infection of cultured cobia *Rachycentron canadum* in Taiwan. *Fish Pathology* **41**, 51–56.

- Ogawa, K., Yamabata, N. and Yoshinaga, T. (2005a). Egg-laying of the monogenean *Heterobothrium okamotoi* on experimentally infected tiger puffer *Takifugu rubripes*. *Fish Pathology* **40**, 111–118.
- Ogawa, K., Yasusaki, M. and Yoshinaga, T. (2005b). Experiments of the blood feeding of *Heterobothrium okamotoi* (Monogenea: Diclidophoridae). *Fish Pathology* **40**, 169–174.
- O'Halloran, J., Carpenter, J., Ogden, D., Hogans, W.E. and Jansen, M. (1992). *Ergasilus labracis* on Atlantic salmon. *Canadian Veterinary Journal* **33**, 75.
- Okamoto, R. (1963). On the problems of a monogenetic trematode infection of puffers from the Inland Sea of Japan. *Suisan Zoshoku* **3** (Special issue), 17–29. [In Japanese].
- Olivares, J. and Marshall, S. (2010). Determination of minimal concentration of *Piscirickettsia salmonis* in water columns to establish a fallowing period in salmon farms. *Journal of Fish Diseases* **33**, 261–266.
- Oliver, L.M., Fisher, W.S., Ford, S.E., Ragone Calvo, L.M., Bureson, E.M., Sutton, E.B. and Gandy, J. (1998). *Perkinsus marinus* tissue distribution and seasonal variation in oysters *Crassostrea virginica* from Florida, Virginia and New York. *Diseases of Aquatic Organisms* **34**, 51–61.
- O'Neill, M., Frenette, A., O'Keefe, R., Harrison, G., Neil, S., Trippel, E., Benfey, T. and Duffy, M. (2011). Preliminary experiments to establish *Loma morhua* infections in naïve Atlantic cod (*Gadus morhua*). *Bulletin of the Aquaculture Association of Canada* **109–1**, 13–20.
- Overstreet, R.M. (1973). Parasites of some penaeid shrimps with emphasis on reared hosts. *Aquaculture* **2**, 105–140.
- Overstreet, R.M. (1987). Solving parasite-related problems in cultured Crustacea. *International Journal of Parasitology* **17**, 309–318.
- Overstreet, R.M. (2007). Effects of a hurricane on fish parasites. *Parassitologia* **49**, 161–168.
- Padrós, F. and Crespo, S. (1995). Proliferative epitheliocystis associated with monogenean infection in juvenile sea bream *Sparus aurata* in the North East of Spain. *Bulletin of the European Association of Fish Pathologists* **15**, 42–44.
- Padrós, F., Zarza, C. and Crespo, S. (2001). Histopathology of cultured sea bream *Sparus aurata* infected with sanguinicolid trematodes. *Diseases of Aquatic Organisms* **44**, 47–52.
- Paladini, G., Cable, J., Fioravanti, M.L., Faria, P.J., Di Cave, D. and Shinn, A.P. (2009). *Gyrodactylus oreochiae* n. sp. (Gyrodactylidae: Monogenea) from farmed populations of gilthead seabream (*Sparus aurata* L.) in the Adriatic Sea. *Folia Parasitologica* **56**, 21–28.
- Paladini, G., Hansen, E., Fioravanti, M.L. and Shinn, A.P. (2011). *Gyrodactylus longipes* sp. nov. (Monogenea: Gyrodactylidae) from farmed populations of gilthead seabream (*Sparus aurata* L.) from the Mediterranean. *Parasitology International* **60**, 410–418.
- Palenzuela, O., Sijá-Bobadilla, A. and Alvarez-Pellitero, P. (1997). *Ceratomyxa sparusaureti* (Protozoa: Myxosporea) infections in cultured gilthead sea bream *Sparus aurata* (Pisces: Teleostei) from Spain: aspects of the host-parasite relationship. *Parasitology Research* **83**, 539–548.
- Papapanagiotou, E.P. and Trilles, J.P. (2001). Cymothoid parasite *Ceratothoa parallela* inflicts great losses on cultured gilthead sea bream *Sparus aurata* in Greece. *Diseases of Aquatic Organisms* **45**, 237–239.
- Papapanagiotou, E.P., Trilles, J.P. and Photis, G. (1999). First record of *Emetha audouini*, a cymothoid isopod parasite, from cultured sea bass *Dicentrarchus labrax* in Greece. *Diseases of Aquatic Organisms* **38**, 235–237.
- Paperna, I. (1975). Parasites and disease of the grey mullet (Mugilidae) with special reference to the seas of the near east. *Aquaculture* **5**, 65–80.
- Paperna, I., Colorni, A., Gordin, H. and Kissil, G.W. (1977). Diseases of *Sparus aurata* in marine culture at Elat. *Aquaculture* **10**, 195–213.
- Paperna, I., Diamant, A. and Overstreet, R.M. (1984). Monogenean infestations and mortality in wild and cultured Red Sea fishes. *Helgolander Meeresunters* **37**, 445–462.
- Park, K.-I. and Choi, K.-S. (2001). Spatial distribution of the protozoan parasite *Perkinsus* sp. found in Manila clams *Ruditapes philippinarum*. *Aquaculture* **203**, 9–22.
- Park, K.-I., Choi, K.-S. and Choi, J.-W. (1999). Epizootiology of *Perkinsus* sp. found in the Manila clam, *Ruditapes philippinarum* in Komsong Bay, Korea. *Journal of the Korean Fisheries Society* **32**, 303–309.
- Park, K.-I., Park, J.-K., Lee, J. and Choi, K.-S. (2005). Use of molecular markers for species identification of Korean *Perkinsus* sp. isolated from Manila clams *Ruditapes philippinarum*. *Diseases of Aquatic Organisms* **66**, 255–263.
- Patarnello, P.P., Fioravanti, M.L., Caggiano, M. and Restani, R. (1995). Infection by Gnathiidae (Crustacea: Isopoda) in *Pagrus major*. *Bollettino Società Italiana di Patologia Ittica* **7**, 32–36.
- Pavoletti, E., Fioravanti, M.L., Prearo, M. and Ghitino, C. (1999). Osservazioni sulla Caligosi in spigole d'allevamento. *Bollettino Società Italiana di Patologia Ittica* **11**, 2–9.
- Paynter, K.T. and Bureson, E.M. (1991). Effect of *Perkinsus marinus* infection in the eastern oyster *Crassostrea virginica*: II. Disease development and impact on growth rate at different salinities. *Journal of Shellfish Research* **10**, 425–431.
- Pedersen, B.H. (1993). Embryos and yolk-sac larvae of turbot *Scophthalmus maximus* are infested with an endoparasite from the gastrula stage onwards. *Diseases of Aquatic Organisms* **17**, 57–59.
- Pedersen, B.H., Buchmann, K. and Koie, M. (1993). Baltic larval cod *Gadus morhua* are infested with a protistan endo-parasite in the yolk sac. *Diseases of Aquatic Organisms* **16**, 29–33.
- Peng, J.S., Ge, X.P., Li, M., Zhou, W.C., Zhao, Q.L., Xu, P. and Xie, J. (2011). Study on *Haplosporidium* disease of *Babylonia areolata*. *Acta Hydrobiologica Sinica* **35**, 803–807.
- Pereya, W.T. (1962). Mortality of Pacific oysters, *Crassostrea gigas* (Thunberg), in various exposure situations in Washington. *Proceedings of the National Shellfisheries Association* **53**, 51–63.
- Perkins, F.O. (1993). Infectious diseases of molluscs. In *Pathobiology of Marine and Estuarine Organisms* (ed. Couch, J.A. and Fournie, J.W.), pp. 255–288. CRC Press, Boca Raton, Florida.
- Perrigault, M., Dahl, S.F., Espinosa, E.P., Gambino, L. and Allam, B. (2011). Effects of temperature on hard clam (*Mercenaria mercenaria*) immunity and QPX (Quahog Parasite Unknown) disease development: II. Defense parameters. *Journal of Invertebrate Pathology* **106**, 322–332.
- Perrigault, M., Tanguy, A. and Allam, B. (2009). Identification and expression of differentially expressed genes in the hard clam, *Mercenaria mercenaria*, in response to quahog parasite unknown (QPX). *BMC Genomics* **10**, 377.
- Petersen, E.H., Chinh, D.T.M., Diu, N.T., Phuoc, V.V., Phuong, T.H., Dung, N.V., Dat, N.K., and Giang, P.T. (2011a). Bioeconomics of grouper, Serranidae Epinephelinae, culture in Vietnam. ACE Discussion Paper 2011/1. <http://www.advancedchoiceeconomics.com.au>, 19 pp.
- Petersen, E.H., Hien, T.T., Suc, N.X. and Thanh, D.V. (2010). Tilapia farming in Vietnam – bioeconomics and perceived constraints to development. ACE Discussion Paper 2009/05. <http://www.advancedchoiceeconomics.com.au>, 23 pp.
- Petersen, E.H., Phuong, T.H., Dat, N.K., Tuan, V.A. and Truc, L.V. (2011b). Bioeconomics of Asian seabass, *Lates calcarifer*, culture in Vietnam. ACE Discussion Paper 2011/3. <http://www.advancedchoiceeconomics.com.au>, 15 pp.
- Pogoda, B., Jungblut, S., Buck, B.H. and Hagen, W. (2012). Infestation of oysters and mussels by mytilicolid copepods: differences between natural coastal habitats and two offshore cultivation sites in the German Bight. *Journal of Applied Ichthyology* **28**, 756–765.
- Pookasawan, T., Boonyarattapin, S., Arirat, S., Duongawad, M. and Rernpan, L. (1982). Aquatic diseases. *Thai Fish Gazette* **40**, 41–46.
- Poppe, T.T. and Mo, T.A. (1993). Systemic, granulomatous hexamitosis of farmed Atlantic salmon: interaction with wild fish. *Fisheries Research* **17**, 147–152.
- Poppe, T.T., Mo, T.A. and Iversen, L. (1992). Disseminated hexamitosis in sea-caged Atlantic salmon *Salmo salar*. *Diseases of Aquatic Organisms* **14**, 91–97.
- Potter, M.A. (1983). Growth rates of cultivated Sydney rock oysters *Saccostrea (Crassostrea) commercialis* in two estuaries in subtropical southern Queensland. *Journal of Agriculture and Animal Science* **40**, 137–140.
- Prasertsri, S., Limsuwan, C. and Chuchird, N. (2009). The effects of the microsporidian (*Thelohania*) infection on the growth and histopathological changes in pond-reared Pacific white shrimp (*Litopenaeus vannamei*). *Kasetsart Journal of Natural Sciences* **43**, 680–688.
- Printakoon, C. and Purivirojkul, W. (2012). Infection of speckled shrimp *Metapenaeus monoceros* (Decapoda: Penaeidae) by the brancial parasite *Orbione bonnierii* (Epicaridea: Bopyridae). *Vie et Milieu* **62**, 17–22.
- Quayle, D.B. (1961). Denman Island disease and mortality. *Fisheries Research Board of Canada Manuscript Report* **713**. Ottawa.
- Quayle, D.B. (1982). Denman Island oyster disease 1960–1980. *British Columbia Shellfish Mariculture Newsletter* **2**, 1–5.
- Rae, G.H. (2002). Sea louse control in Scotland, past and present. *Pest Management Science* **58**, 515–520.
- Ragan, M.A., MacCallum, G.S., Murphy, C.A., Cannone, J.J., Gutell, R.R. and McGladdery, S.E. (2000). Protistan parasite QPX of

- hard-shell clam *Mercenaria mercenaria* is a member of Labyrinthulomycota. *Diseases of Aquatic Organisms* **42**, 185–190.
- Rajendran, K. V., Thirunavukkarasu, A. R. and Santiago, T. C.** (2000). Mortality of captive seabass, *Lates calcarifer* (Bloch) due to monogenetic parasite, *Diplectanum lateri* (Tripathi, 1957). *Journal of Aquaculture in the Tropics* **15**, 199–206.
- Ramasamy, P., Jayakumar, R. and Brennan, G. P.** (2000). Muscle degeneration associated with cotton shrimp disease of *Penaeus indicus*. *Journal of Fish Diseases* **23**, 77–81.
- Ramasamy, P., Rajan, P. R., Jayakumar, R., Rani, S. and Brennan, G. P.** (1996). *Lagenidium callinectes* (Couch, 1942) infection and its control in cultured larval Indian tiger prawn, *Penaeus monodon* Fabricius. *Journal of Fish Diseases* **19**, 75–82.
- Ramos, M. F., Costa, A. R., Barandela, T., Saraiva, A. and Rodrigues, P. N.** (2007). Scuticociliate infection and pathology in cultured turbot *Scophthalmus maximus* from the north of Portugal. *Diseases of Aquatic Organisms* **74**, 249–253.
- Ramos, P. and Oliveira, J.** (2001). Amyloodiniosis in turbot, *Psetta maxima* (L.). *Revista Portuguesa de Ciências Veterinárias* **540**, 201–205.
- Regidor, S. E. and Arthur, J. R.** (1986). Parasites of juvenile milkfish, *Chanos chanos*. In *The First Asian Fisheries Forum*, Vol. 1 (ed. Maclean, J. L., Dizon, L. B. and Hosillos, L. V.), pp. 261–264. Asian Fisheries Society, Manila, Philippines.
- Ren, S. L., Yang, X.-C. and Song, W.-B.** (2005). Study on histopathology of the cultured clam *Meretrix meretrix* suffered from the “mantle hypertrophy disease”. *Journal of Ocean University of Qingdao* **6**, 012.
- Rhnye, A. L., Tlustý, M. F., Schofield, P. J., Kaufman, L., Morris, J. A. and Bruckner, A. W.** (2012). Revealing the appetite of the marine aquarium fish trade: the volume and biodiversity of fish imported into the United States. *PLoS ONE* **7**, e35808.
- Riddington, G., Radford, A., Paffrath, S., Bostock, J. and Shinn, A.** (2006). An economic evaluation of the impact of the salmon parasite *Gyrodactylus salaris* (Gs) should it be introduced into Scotland: Summary Report. Prepared for the Scottish Executive Environment and Rural Affairs Department, Project Number SAQ/001/05.
- Riedel, R., Caskey, L. and Costa-Pierce, B. A.** (2002). Fish biology and fisheries ecology of the Salton Sea, California. In *Developments in Hydrobiology: The Salton Sea* (ed. Barnum, D. A., Elder, J. F., Stephens, D. and Friend, M.), pp. 229–244. Kluwer Academic Publishers, Dordrecht, the Netherlands.
- Rigos, G., Christophiliogiannis, P., Giahnishi, M., Andriopoulou, A., Koutsodimoy, M., Nengas, I. and Alexis, M.** (1998). *Amyloodinium ocellatum* infestation on sharpnose sea bream, *Puntazzo puntazzo* Cetti. *Bulletin of the European Association of Fish Pathologists* **18**, 198–200.
- Rigos, G., Christophiliogiannis, P., Yiagnisi, M., Koutsodimoy, M., Andriopoulou, K., Nengas, I. and Alexis, M.** (1999). Myxosporean infections in Greek mariculture. *Aquaculture International* **7**, 361–364.
- Rigos, G., Pavlidis, M. and Divanach, P.** (2001). Host susceptibility to *Cryptocaryon* sp. infection of Mediterranean marine broodfish held under intensive culture conditions: a case report. *Bulletin of the European Association of Fish Pathologists* **21**, 33–36.
- Roberts, T., Murrell, K. D. and Marks, S.** (1994). Economic losses caused by foodborne parasitic diseases. *Parasitology Today* **10**, 419–423.
- Robledo, J. A. F., Caceres-Martinez, J. and Figueras, A.** (1994a). *Mytilicola intestinalis* and *Proctoeces maculatus* in mussel (*Mytilus galloprovincialis* Lmk.) beds in Spain. *Bulletin of the European Association of Fish Pathologists* **14**, 89–91.
- Robledo, J. A. F. and Figueras, A.** (1995). The effects of culture-site, depth, season and stock source on the prevalence of *Marteilia refringens* in cultured mussels (*Mytilus galloprovincialis* LMK.) from Galicia, Spain. *Journal of Parasitology* **81**, 354–363.
- Robledo, J. A. F., Santarém, M. M. and Figueras, A.** (1994b). Parasite loads of rafted blue mussels (*Mytilus galloprovincialis*) in Spain with special reference to the copepod, *Mytilicola intestinalis*. *Aquaculture* **127**, 287–302.
- Robledo, J. A. F., Santarém, M. M., González, P. and Figueras, A.** (1995). Seasonal variations in the biochemical composition of the serum of *Mytilus galloprovincialis* Lmk. and its relationship to the reproductive cycle and parasitic load. *Aquaculture* **133**, 311–322.
- Rodger, H. D. and McArdle, J. F.** (1996). An outbreak of amoebic gill disease in Ireland. *Veterinary Record* **139**, 348–349.
- Rodger, H. D., Turnbull, T., Scullion, F. T., Sparrow, D. and Richards, R. H.** (1995). Nervous mortality syndrome in farmed Atlantic salmon. *Veterinary Record* **137**, 616–617.
- Ronquillo, I. A. and Caces-Borja, P.** (1960). Notes on the infestation of *Chanos chanos* by a parasitic isopod. *Philippines Journal of Fisheries* **8**, 113–117. Reprinted 1975. National Bangos Symposium, 25–26 July 1975, Manila, Philippines, Symp. Pap., SEAFDEC, Makati, Philippines, 3 pp.
- Roth, M.** (2000). The availability and use of chemotherapeutic sea lice control products. *Contributions to Zoology* **69**, 109–118.
- Rough, K. M., Lester, R. J. G. and Reuter, R. E.** (1999). *Caligus elongatus* a significant parasite of cultured southern bluefin tuna *Thunnus maccoyii*. *World Aquaculture* **99**, 26th April–2nd May 1999, Sydney. *World Aquaculture Society*, p. 655.
- Rozas, M.** (2011). Descripción patológica y epidemiológica de Amoebic Gill Disease (AGD) en salmón del Atlántico, *Salmo salar*, en Chile. Tesis de Magister en Ciencias Veterinarias. Universidad Austral de Chile, Valdivia.
- Rozas, M. and Asencio, G.** (2007). Evaluación de la situación epidemiológica de Caligiasis en Chile: hacia una estrategia de control efectiva. *Salmocencia* **2**, 43–59.
- Sadek, S.** (2011). An overview on desert aquaculture in Egypt. In *Aquaculture in Desert and Arid Lands: Development Constraints and Opportunities*. FAO Technical Workshop. 6–9 July 2010, Hermosillo, Mexico. FAO Fisheries and Aquaculture Proceedings No. 20 (ed. Crespi, V. and Vovattelli, A.), pp. 141–158. FAO, Rome.
- Sahul Hameed, A. S.** (1995). Mortality of larvae of *Penaeus indicus* due to the infestation of *Nitzschia closterium* in a hatchery. *Journal of Aquaculture in the Tropics* **10**, 337–342.
- Sahul Hamed, A. S.** (1996). Mortality in protozoa and mysis of *Penaeus indicus* and *P. semisulcatus* by *Leptomonas* like parasite in the hatcheries. *Indian Journal of Fisheries* **43**, 389–391.
- Sakaguchi, S.** (1968a). Studies on the life history of the trematode parasitic in the pearl oyster, *Pinctada fucata*, and on the hindrance for pearl culture. *Bulletin of the National Pearl Research Laboratory* **13**, 1635–1688 [In Japanese, English summary].
- Sakaguchi, S.** (1968b). Studies on a trematode parasitic of the pearl oyster II: its effects on the pearl oyster as the intermediate host. *Bulletin of the National Pearl Research Laboratory* **9**, 1161–1169 [In Japanese, English summary].
- Saksvik, M., Nilsen, F., Nylund, A. and Berland, B.** (2001). Effect of marine *Eubothrium* sp. (Cestoda: Pseudophyllidea) on the growth of Atlantic salmon, *Salmo salar* L. *Journal of Fish Diseases* **24**, 111–119.
- Sanil, N. K., Vikas, P. A., Ratheesh, T. B., George, K. C. and Vijayan, K. K.** (2009). Mortalities caused by the crustacean isopod, *Cirolana fluviatilis*, in tropical, cage-cultured Asian seabass, *Lates calcarifer*: a case study from the southwest coast of India. *Aquaculture Research* **40**, 1626–1633.
- Sano, M., Sata, J. and Yokoyama, H.** (1998). Occurrence of Beko disease caused by *Microsporidium seriola* (Microsporida) in hatchery-reared juvenile yellowtail. *Fish Pathology* **33**, 11–16.
- Sanz, F.** (1992). Mortality of cultured sea bream (*Sparus aurata*) caused by an infection with a trematode of the genus *Microcotyle*. *Bulletin of the European Association of Fish Pathologists* **12**, 186–188.
- Šarušić, G.** (1999). Preliminary report of infestation by isopod *Ceratothoa oestroides* (Risso, 1826), in marine cultured fish. *Bulletin of the European Association of Fish Pathologists* **19**, 110–112.
- Sato-Okoshi, W., Okoshi, K., Abe, H. and Li, J. Y.** (2012). Polydorid species (Polychaeta: Spionidae) associated with commercially important mollusk shells in Korean waters. *Aquaculture* **350–353**, 82–90.
- Sato-Okoshi, W., Okoshi, K., Abe, H. and Li, J. Y.** (2013). Polydorid species (Polychaeta, Spionidae) associated with commercially important mollusk shells from eastern China. *Aquaculture* **406–407**, 153–159.
- Sayuthi, S.** (1993). Fish diseases in Malaysia: status and problems. In *Proceedings of the Aquaculture Workshop for SEAFDEC/AQD Training Alumni, Iloilo, Philippines, 8–11 September 1992*.
- Scheibling, R. E. and Stephenson, R. L.** (1984). Mass mortality of *Strongylocentrotus droebachiensis* (Echinodermata: Echinoidea) off Nova Scotia, Canada. *Marine Biology* **78**, 153–164.
- Schipp, G., Bosmans, J. and Humphrey, J.** (2007). *Northern Territory Barramundi Farming Handbook*. Department of Primary Industry, Fisheries and Mines, Northern Territory Government, Australia. ISBN 0 7245 4727 4, 80 pp.
- Scullion, F. T., Scullion, M. G., Sparrow, D., Rodger, H. D. and Sheahan, B. J.** (1996). Encephalitis and mass mortality of farmed salmon smolts in an isolated sea bay in Ireland. *Veterinary Record* **138**, 161–162.
- Shaw, R. and Opitz, M.** (1993). Preliminary results on sea lice in the Maine aquaculture industry. *Bulletin of the Aquaculture Association of Canada* **93/94**, 102–104.
- Shi, L. and Wang, J.** (2001). Biochemical and immunological characterization of excretory-secretory products of *Vesicocodium solenophagum* and plasma proteins of its bivalve host, *Sinonovacula constricta*. *Journal of Helminthology* **75**, 279–284.
- Shimokawa, J., Yoshinaga, T. and Ogawa, K.** (2010). Experimental evaluation of the pathogenicity of *Perkinsus olseni* in juvenile Manila clams *Ruditapes philippinarum*. *Journal of Invertebrate Pathology* **105**, 347–351.

- Shinn, A. P. and Bron, J. E. (2012). Chapter 8: Considerations for the use of anti-parasitic drugs in aquaculture. In *Infectious Disease in Aquaculture* (ed. Austin, B.), pp. 190–217. Woodhead Publishing Ltd., Cambridge, UK.
- Shirakashi, S., Morita, A., Ishimaru, K. and Miyashita, S. (2012). Infection dynamics of *Kudoa yasunagai* (Myxozoa: Multivalvulida) infecting brain of cultured yellowtail *Seriola quinqueradiata* in Japan. *Diseases of Aquatic Organisms* **101**, 123–130.
- Sievers, G., Lobos, C., Inostroza, R. and Ernst, S. (1996). The effect of the isopod parasite *Ceratothoa gaudichaudii* on the body weight of farmed *Salmo salar* in southern Chile. *Aquaculture* **143**, 1–6.
- Silphaduang, U., Hatai, K., Wada, S. and Noga, E. (2000). *Cladosporiosis* in a tomato clownfish (*Amphiprion frenatus*). *Journal of Zoo and Wildlife Medicine* **31**, 259–261.
- Simon, C. A., Bentley, M. G. and Caldwell, G. S. (2010). 2, 4-decadienal: exploring a novel approach for the control of polychaete pests on cultured abalone. *Aquaculture* **310**, 52–60.
- Simon, C. A., Ludford, A. and Wynne, S. (2006). Spionid polychaetes infesting cultured abalone *Haliotis midae* in South Africa. *African Journal of Marine Science* **28**, 167–171.
- Sinderman, C. J. (1976). Oyster mortalities and their control. *FAO Technical Conference on Aquaculture, Kyoto, Japan*, 25 pp.
- Sinnett, R. (1998). Sea lice – watch out for the hidden costs. *Fish Farmer* **21**, 45–46.
- Sitjà-Bobadilla, A. and Alvarez-Pellitero, P. (1990). First report of *Ichthyophonus* disease in wild and cultured sea bass *Dicentrarchus labrax* from the Spanish Mediterranean area. *Diseases of Aquatic Organisms* **8**, 145–150.
- Sitjà-Bobadilla, A. and Alvarez-Pellitero, P. (1992). Light and electron microscopic description of *Sphaerospora dicentrarchi* n. sp. (Myxosporidia: Sphaerosporidae) from wild and cultured sea bass (*Dicentrarchus labrax* L.). *Journal of Protozoology* **39**, 273–281.
- Sitjà-Bobadilla, A. and Alvarez-Pellitero, P. (2001). *Cryptosporidium* sp. (Apicomplexa: Cryptosporidiidae), an ubiquitous parasite in gilthead sea bream (*Sparus aurata*): epidemiological survey in Spanish cultures. *Tenth International Conference of the EAAP on Diseases of Fish and Shellfish*. Book of Abstracts. Dublin, Ireland, pp. 0–52.
- Siva Kumari, J. and Ramesh Babu, K. (2001). Heavy mortalities in larvae of *Penaeus monodon* in hatcheries of Kakinada and Visakhapatnam coasts due to *Fusarium* and *Vibriosis*. *Journal of the Marine Biological Association of India* **43**, 202–205.
- Smolowitz, R., Leavitt, D. and Perkins, F. (1998). Observations of a protistan disease similar to QPX in *Mercenaria mercenaria* hard clams from the coast of Massachusetts. *Journal of Invertebrate Pathology* **71**, 9–25.
- Soares, F., Quental-Ferreira, H., Moreira, M., Cunha, E., Ribeiro, L. and Pousão-Ferreira, P. (2012). First report of *Amyloodinium ocellatum* in farmed meagre (*Argyrosomus regius*). *Bulletin of the European Association of Fish Pathologists* **32**, 30–33.
- Somga, J. R., Somga, S. S. and Reantaso, M. B. (2002). Impacts of disease on small-scale grouper culture in the Philippines, pp. 207–214. In *Primary Aquatic Animal Health Care in Rural, Small-Scale, Aquaculture Development*. FAO Fisheries Technical Paper No. 406 (ed. Arthur, J. R., Phillips, M. J., Subasinghe, R. P., Reantaso, M. B. and MacRae, I. H.). FAO, Rome.
- Sousa, R., Antunes, C. and Guilhermino, L. (2008a). Ecology of the invasive Asian clam *Corbicula fluminea* (Müller, 1774) in aquatic ecosystems: an overview. *Annales de Limnologie – International Journal of Limnology* **44**, 85–94.
- Sousa, R., Nogueira, A. J., Gaspar, M. B., Antunes, C. and Guilhermino, L. (2008b). Growth and extremely high production of the non-indigenous invasive species *Corbicula fluminea* (Müller, 1774): possible implications for ecosystem functioning. *Estuarine, Coastal and Shelf Science* **80**, 289–295.
- Sousa, R., Rufino, M., Gaspar, M., Antunes, C. and Guilhermino, L. (2008c). Abiotic impacts on spatial and temporal distribution of *Corbicula fluminea* (Müller, 1774) in the River Minho Estuary, Portugal. *Aquatic Conservation: Marine and Freshwater Ecosystems* **18**, 98–110.
- Speare, D. J., Brackett, J. and Ferguson, H. W. (1989). Sequential pathology of the gills of coho salmon with a combined diatom and microsporidian gill infection. *Canadian Veterinary Journal* **30**, 571–575.
- Steinum, T., Kvellestad, A., Rønneberg, L. B., Nilsen, H., Asheim, A., Fjell, K., Nygård, S. M. R., Olsen, A. B. and Dale, O. B. (2008). First cases of amoebic gill disease (AGD) in Norwegian seawater farmed Atlantic salmon, *Salmo salar* L., and phylogeny of the causative amoeba using 18S cDNA sequences. *Journal of Fish Diseases* **31**, 205–214.
- Stephens, F. J. and Savage, A. (2010). Two mortality events in sea-caged yellowtail kingfish *Seriola lalandi* Valenciennes, 1833 (Nannoperidae) from Western Australia. *Australian Veterinary Journal* **88**, 414–416.
- Sterud, E., Hansen, M. K. and Mo, T. A. (2000). Systemic infection with *Uronema*-like ciliates in farmed turbot, *Scophthalmus maximus* (L.). *Journal of Fish Diseases* **23**, 33–37.
- Sterud, E., Simolin, P. and Kvellestad, A. (2003). Infection by *Parvicapsula* sp. (Myxozoa) is associated with mortality in sea-caged Atlantic salmon *Salmo salar* in northern Norway. *Diseases of Aquatic Organisms* **54**, 259–263.
- Sugiyama, A., Yokoyama, H. and Ogawa, K. (1999). Epizootiological investigation on kudoosis amami caused by *Kudoa amamiensis* (Multivalvulida: Myxozoa) in Okinawa Prefecture, Japan. *Fish Pathology* **34**, 39–43.
- Sunila, I. and LaBanca, J. (2003). Apoptosis in the pathogenesis of infectious diseases of the eastern oyster *Crassostrea virginica*. *Diseases of Aquatic Organisms* **56**, 163–170.
- Sunila, I., Williams, L., Russo, S. and Getchis, T. (2004). Reproduction and pathology of blue mussels, *Mytilus edulis* (L.) in an experimental longline in Long Island Sound, Connecticut. *Journal of Shellfish Research* **23**, 731–740.
- Supamattaya, K., Fischer-Scherl, T., Hoffmann, R. W. and Boonyaratpalin, S. (1990). Renal sphaerosporosis in cultured grouper *Epinephelus malabaricus*. *Diseases of Aquatic Organisms* **8**, 35–38.
- Suzuki, H. and Matsutani, T. (2009). Infection of the parasitic copepod, *Pectenophilus ornatus* on juvenile Japanese scallops, *Patinopecten yessoensis*. *Suisan Zoshoku* **57**, 513–514.
- Tallaksen, E. (2013). Salmon farms rushing to slaughter 8,000 t fish due to high lice levels. *Undercurrent News* (2nd October). <http://www.undercurrentnews.com/2013/10/02/salmon-farms-rushing-to-slaughter-8000t-fish-due-to-high-lice-levels/>
- Tan, C. K. F., Nowak, B. F. and Hodson, S. L. (2002). Biofouling as a reservoir of *Neoparamoeba pemaquidensis* (Page, 1970), the causative agent of amoebic gill disease in Atlantic salmon. *Aquaculture* **210**, 49–58.
- Terengo, S., Agostini, S., Quilichini, Y., Euzet, L. and Marchand, B. (2010). Intensive infestations of *Sciaenocotyle panzerii* (Monogenea, Microcotylidae) on *Argyrosomus regius* (Asso) under fish-farming conditions. *Journal of Fish Diseases* **33**, 89–92.
- Terengo, S. and Katharios, P. (2008). Microcotylid parasites: an emerging problem in Mediterranean cage aquaculture. *Fish Farming Expert* **5**, 44–48.
- Thébault, A., Baud, J. P., Le Saux, J. C., Le Roux, F., Chollet, B., Le Coguc, M. J., Fleury, P. G., Berthe, F. and Gerard, A. (1999). Compte rendu sur les mortalité de juillet 1999 des moules (*Mytilus edulis*) en poches dans l'Aber Benoît. *Rapport IFREMER*, 12 p.
- Toksen, E., Buchmann, K. and Bresciani, J. (2007). Occurrence of *Benedenia sciaenae* van Beneden, 1856 (Monogenea: Capsalidae) in cultured meagre (*Argyrosomus regius* Asso, 1801) (Teleost: Sciaenidae) from western Turkey. *Bulletin of the European Association of Fish Pathologists* **27**, 250–253.
- Tomiyama, T., Watanabe, M. and Fujita, T. (2008). Community-based stock enhancement and fisheries management of the Japanese flounder in Fukushima, Japan. *Reviews in Fisheries Science* **16**, 146–153.
- Torchin, M. E., Lafferty, K. D. and Kuris, A. M. (2002). Parasites and marine invasions. *Parasitology* **124**, 137–151.
- Torgerson, P. R. and MacPherson, C. N. L. (2011). The socioeconomic burden of parasitic zoonoses: global trends. *Veterinary Parasitology* **182**, 79–95.
- Tripp, M. R. (1973). Hermaphroditism in *Bucephalus*-infected oysters. *Journal of Invertebrate Pathology* **21**, 321–322.
- Trottier, O. and Jeffs, A. G. (2012). Biological characteristics of parasitic *Nepinnotheres novaezelandiae* within a *Perna canaliculus* farm. *Diseases of Aquatic Organisms* **101**, 61–68.
- Trottier, O., Walker, D. and Jeffs, A. G. (2012). Impact of the parasitic pea crab *Pinnotheres novaezelandiae* on aquacultured New Zealand green-lipped mussels, *Perna canaliculus*. *Aquaculture* **344**, 23–29.
- Tully, O. (1989). The succession of generations and growth of the caligid copepod *Caligus elongatus* and *Lepeophtheirus salmonis* parasitising farmed Atlantic salmon smolts (*Salmo salar* L.). *Journal of the Marine Biological Association of the UK* **69**, 279–287.
- Tun, K. L., Itoh, N., Shimizu, Y., Yamanoi, H., Yoshinaga, T. and Ogawa, K. (2008). Pathogenicity of the protozoan parasite *Marteilioides chungmuensis* in the Pacific oyster *Crassostrea gigas*. *International Journal for Parasitology* **38**, 211–217.
- Tun, T., Ogawa, K. and Wakabayashi, H. (2002). Pathological changes induced by three myxosporidians in the intestine of cultured tiger puffer, *Takifugu rubripes* (Temminck and Schlegel). *Journal of Fish Diseases* **25**, 63–72.
- Tun, T., Yokoyama, H., Ogawa, K. and Wakabayashi, H. (2000). Myxosporidians and their hyperparasitic microsporidians in the intestine of emaciated tiger puffer. *Fish Pathology* **35**, 145–156.

- Tuntiwaranuruk, C., Chalermwat, K., Upatham, E. S., Kruatrachue, M. and Azevedo, C. (2004). Investigation of *Nematopsis* spp. oocysts in 7 species of bivalves from Chonburi Province, Gulf of Thailand. *Diseases of Aquatic Organisms* **58**, 47–53.
- Uddin, M. J., Yasin, Z., Khalil, M. and Shau-Hwai, A. T. (2011). Parasites of blood cockle (*Anadara granosa* Linnaeus, 1758) from the Straits of Malacca. *Journal of Shellfish Research* **30**, 875–880.
- United Nations, Department of Economic and Social Affairs, Population Division (2012). World population 2012. http://www.un.org/en/development/desa/population/publications/pdf/trends/WPP2012_Wallchart.pdf
- Urawa, S. (1993). Effects of *Ichthyobodo necator* infections on seawater survival of juvenile chum salmon (*Oncorhynchus keta*). *Aquaculture* **110**, 101–110.
- Urawa, S. (1995). Marine mortality of chum salmon (*Oncorhynchus keta*) caused by the parasitic flagellate *Ichthyobodo necator*. NPAFC Doc. 147, 7 pp. Research Division, Hokkaido Salmon Hatchery, Fisheries Agency of Japan, Toyohira-ku, Sapporo 062, Japan.
- Urawa, S. and Kato, T. (1991). Heavy infections of *Caligus orientalis* (Copepoda: Caligidae) on caged rainbow trout *Oncorhynchus mykiss* in brackish water. *Fish Pathology [Gyobyō Keokyūg]* **26**, 161–162.
- Urawa, S., Kato, T. and Kumagai, A. (1998a). A status of *Lepeophtheirus salmonis* (Copepoda: Caligidae) on seawater-cultured coho salmon (*Oncorhynchus kisutch*) and rainbow trout (*O. mykiss*) in Japan. *Bulletin of the National Salmon Resources Center* **1**, 35–38.
- Urawa, S. and Kusakari, M. (1990). The survivability of the ectoparasitic flagellate *Ichthyobodo necator* on chum salmon fry (*Oncorhynchus keta*) in seawater and comparison to *Ichthyobodo* sp. on Japanese flounder (*Paralichthys olivaceus*). *Journal of Parasitology* **76**, 33–40.
- Urawa, S., Ueki, N. and Karlsbakk, E. (1998b). A review of *Ichthyobodo* infection in marine fishes. *Fish Pathology* **33**, 311–320.
- Urawa, S., Ueki, N., Nakai, T. and Yamasaki, H. (1991). High mortality of cultured juvenile Japanese flounder, *Paralichthys olivaceus* (Temminck and Schlegel), caused by the parasite flagellate, *Ichthyobodo* sp. *Journal of Fish Diseases* **14**, 489–494.
- Utari, H. B., Senapin, S., Jaengsanong, C., Flegel, T. W. and Kruatrachue, M. (2012). A haplosporidian parasite associated with high mortality and slow growth in *Penaeus (Litopenaeus) vannamei* cultured in Indonesia. *Aquaculture* **366–367**, 85–89.
- Vagianou, S., Athanassopoulou, F., Ragias, V., Di Cave, D., Leontides, L. and Golomazou, E. (2006). Prevalence and pathology of ectoparasites of Mediterranean sea bream and sea bass reared under different environmental and aquaculture conditions. *Bamidgeh* **58**, 78–88.
- Valencia, V. and George-Nascimento, M. (2013). Farming systems as an ecological refuge against the parasitism by *Edotia doellojurdoi* (Isopoda: Idoteidae) on *Mytilus edulis* (Mollusca: Mytilidae) in Aysén, Chile. *Revista Chilena de Historia Natural* **86**, 153–159.
- Vass, S. (2013). Gill disease to cost salmon farmers £30 m. *Herald Scotland* (online), 19th January 2013. <http://www.heraldscotland.com/business/markets-economy/gill-disease-to-cost-salmon-farmers-30m.19956340>
- Velasquez, C. C. (1979). Pest/parasites and diseases of milkfish in the Philippines, p. 65–67. In *Proceedings Technical Consultation on Available Aquaculture Technology in the Philippines*. Aquaculture Department, Southeast Asian Fisheries Development Center, Iloilo City, and Philippine Council for Agriculture and Resources Research, Los Baños, Laguna, Philippines. Also published in *Advances in Milkfish Biology and Culture* (ed. Juario, J. V., Ferraris, R. P. and Benitez, L. V.), pp. 155–159. Island Publishing House, Manila, Philippines, Aquaculture Department, Southeast Asian Fisheries Development Center, Iloilo, Philippines and International Development Research Centre, Ottawa, Canada (1984).
- Velayudhan, T. S. (1982). On the occurrence of shell boring polychaetes and sponges on pearl oyster *Pinctada fucata* and control of boring organisms. *Proceedings of the Symposium on Coastal Aquaculture* **2**, 614–618.
- Venmathi Maran, B. A., Oh, S.-Y., Soh, H. Y., Choi, H. J. and Myoung, J.-G. (2012). *Caligus sclerotinosus* (Copepoda: Caligidae), a serious pest of cultured red seabream *Pagrus major* (Sparidae) in Korea. *Veterinary Parasitology* **188**, 355–361.
- Vidal-Martínez, V., Jiménez, A. and Simá, R. (2002). Parasites and symbionts of native and cultured shrimps from Yucatan, Mexico. *Journal of Aquatic Animal Health* **14**, 57–64.
- Villalba, A. (1993). *Marteiliasis* affecting cultured mussels *Mytilus galloprovincialis* of Galicia (NW Spain). Etiology, phases of the infection, and temporal and spatial variability in prevalence. *Diseases of Aquatic Organisms* **16**, 61–72.
- Villalba, A., Mourelle, S. G., Carballal, M. J. and Lopez, C. (1993). Effects on infection by the protistan parasite *Marteilia refringens* on the reproduction of cultured mussels *Mytilus galloprovincialis* in Galicia (NW Spain). *Diseases of Aquatic Organisms* **17**, 205–213.
- Villalba, A., Mourelle, S. G., Carballal, M. J. and Lopez, C. (1997). Symbionts and diseases of farmed mussels *Mytilus galloprovincialis* throughout the culture process in the Rías of Galicia (NW Spain). *Diseases of Aquatic Organisms* **31**, 127–139.
- Vohmann, A., Borchering, J., Kureck, A., Bij de Vaate, A., Arndt, H. and Weitere, M. (2010). Strong body mass decrease of the invasive clam *Corbicula fluminea* during summer. *Biological Invasions* **12**, 53–64.
- Voort van der, M., Charlier, J., Lauwers, L., Vercruyse, J., Huylenbroeck Van, G. and Van Meensel, J. (2013). Conceptual framework for analysing farm-specific economic effects of helminth infections in ruminants and control strategies. *Preventive Veterinary Medicine* **109**, 228–235.
- Wang, Y.-G., Zhang, C.-Y., Rong, X.-J., Chen, J.-J. and Shi, C.-Y. (2004). Diseases of cultured sea cucumber, *Apostichopus japonicus*, in China. *FAO Fisheries Technical Paper* **463**, 297–310.
- Watanabe, W. O. and Daniels, H. (2011). Chapter 18: Summary and conclusions. In *Practical Flatfish Culture and Stock Enhancement* (ed. Daniels, H. V. and Watanabe, W. O.), pp. 323–357. Wiley Blackwell, Iowa, USA.
- Werner, S. and Rothhaupt, K. O. (2008). Mass mortality of the invasive bivalve *Corbicula fluminea* induced by a severe low-water event and associated low water temperatures. *Hydrobiologia* **613**, 143–150.
- Wesche, S. J. (1995). Outbreaks of *Marteilia sydneyi* in Sydney rock oysters and their relationship with environmental pH. *Bulletin of the European Association of Fish Pathologists* **15**, 23–27.
- West, A. P. and Roubal, F. R. (1998). Population dynamics of the monogenean *Anoplo-discus cirrusspiralis* on the snapper, *Pagrus auratus*. *International Journal for Parasitology* **28**, 571–577.
- Whitaker, D. J. and Kent, M. L. (1991). Myxosporean *Kudoa thyrsites*: a cause of soft flesh in farm-reared Atlantic salmon. *Journal of Aquatic Animal Health* **3**, 291–294.
- Whitaker, D. J. and Kent, M. L. (1992). *Kudoa thyrsites* (Myxosporea) and soft flesh in pen-reared coho salmon. *American Fisheries Society/Fish Health Sector Newsletter* **20**, 4–5.
- White, M. E., Powell, E. N., Ray, S. M. and Wilson, E. A. (1987). Host-to-host transmission of *Perkinsus marinus* in oyster (*Crasostrea virginica*) populations by the ectoparasitic snail *Boonea impressa* (Pyramidellidae). *Journal of Shellfish Research* **6**, 1–5.
- Whittington, I. D., Corneillie, S., Talbot, C., Morgan, J. A. T. and Adlard, R. D. (2001). Infections of *Seriola quinqueradiata* Temminck & Schlegel and *S. dumerili* (Risso) in Japan by *Benedenia seriola* (Monogenea) confirmed by morphology and 28S ribosomal DNA analysis. *Journal of Fish Diseases* **24**, 421–425.
- Wilbur, A. E., Ford, S. E., Gauthier, J. D. and Gomez-Chiari, M. (2012). Quantitative PCR analysis to determine prevalence and intensity of MSX (*Haplosporidium nelsoni*) in North Carolina and Rhode Island oysters *Crasostrea virginica*. *Diseases of Aquatic Organisms* **102**, 107–118.
- Williams, E. H. and Bunkley-Williams, L. (1996). *Parasites of Offshore Big Game Fishes of Puerto Rico and the Western Atlantic*. Puerto Rico Department of Natural Environmental Resources and the University of Puerto Rico, Mayaguez.
- Williams, E. H., Jr., Bunkley-Williams, L., Lilyestrom, C. G. and Ortiz-Corps, E. A. R. (2001). A review of recent introductions of aquatic invertebrates in Puerto Rico and implications for the management of nonindigenous species. *Caribbean Journal of Science* **37**, 246–251.
- Woo, P. T. K. (2006). *Fish Diseases and Disorders. Volume 1: Protozoan and Metazoan Infections*, 2nd Edn. CABI, Wallingford, UK, p. 800.
- Xu, W., Sheng, X., Xu, H., Shi, H. and Li, P. (2007a). Dinoflagellates *Hematodinium* sp. parasitizing the mud crab *Scylla serrata*. *Periodical of Ocean University of China* **37**, 916–920.
- Xu, W., Shi, H., Xu, H. and Small, H. J. (2007b). Preliminary study on the *Hematodinium* infection in cultured *Portunus trituberculatus*. *Acta Hydrobiologica Sinica* **31**, 637–642.
- Yanagida, T., Freeman, M. A., Nomura, Y., Takami, I., Sugihara, Y., Yokoyama, H. and Ogawa, K. (2005). Development of a PCR-based method for the detection of enteric myxozoans causing the emaciation disease of cultured tiger puffer. *Fish Pathology* **40**, 23–28.
- Yap, W. G., Villaluz, A. C., Soriano, M. G. G. and Santos, M. N. (2007). Milkfish production and processing technologies in the Philippines. *Milkfish Project Publication Series No. 2*, 96 pp.
- Yokoyama, H., Ayado, D., Miyahara, J., Matsukura, K., Takami, I., Yokoyama, F. and Ogawa, K. (2011). Infection dynamics of *Microsporidium seriola* (Microspora) causing Beko disease of *Seriola* spp. *Fish Pathology* **46**, 51–58.

- Yokoyama, H., Freeman, M. A., Itoh, N. and Fukuda, Y.** (2005a). Spinal curvature of cultured Japanese mackerel *Scomber japonicus* associated with a brain myxosporean, *Myxobolus acanthogobii*. *Diseases of Aquatic Organisms* **66**, 1–7.
- Yokoyama, H. and Fukuda, Y.** (2001). *Ceratomyxa seriola* n. sp. and *C. buri* n. sp. (Myxozoa: Myxosporea) from the gall-bladder of cultured yellowtail *Seriola quinqueradiata*. *Systematic Parasitology* **48**, 125–130.
- Yokoyama, H., Itoh, N. and Tanaka, S.** (2005b). *Hemeguya pagri* n. sp. (Myxozoa: Myxosporea) causing cardiac henneguyosis in red sea bream, *Pagrus major* (Temminck & Schlegel). *Journal of Fish Diseases* **28**, 479–487.
- Yokoyama, H., Yanagida, T., Freeman, M. A., Katagiri, T., Hosokawa, A., Endo, M., Hirai, M. and Takagi, S.** (2010). Molecular diagnosis of *Myxobolus spirosulcatus* associated with encephalomyelitis of cultured yellowtail, *Seriola quinqueradiata* Temminck & Schlegel. *Journal of Fish Diseases* **33**, 939–946.
- Yokoyama, H., Yanagida, T. and Takemaru, I.** (2006). The first record of *Kudoa megacapsula* (Myxozoa: Multivalvulida) from farmed yellowtail *Seriola quinqueradiata* originating from wild seedlings in South Korea. *Fish Pathology* **41**, 159–163.
- Yoon, G. H.** (2008). Aquaculture in Korea. *Aquaculture News* **34**, 16–17.
- Yoon, G. H., Shinn, A. P., Sommerville, C. and Jo, J. Y.** (1997). Seasonality and the microhabitat of *Microcotyle sebastis* Goto 1894, a monogenean gill parasite of farmed rockfish, *Sebastes schlegeli* Hilgendorf 1880. *Korean Journal of Aquaculture* **10**, 387–394.
- Yoshinaga, T. and Nakazoe, J.-I.** (1993). Isolation and *in vitro* cultivation of an unidentified ciliate causing scuticociliatosis in Japanese flounder (*Paralichthys olivaceus*). *Fish Pathology* **28**, 131–134.
- You, K., Ma, C., Gao, H., Li, F., Zhang, M., Qiu, Y. and Wang, B.** (2007). Research on the jellyfish (*Rhopilema esculentum* Kishinouye) and associated aquaculture techniques in China: current status. *Aquaculture International* **15**, 479–488.
- Zanguee, N., Lymbery, J. A., Lau, J., Suzuki, A., Yang, R., Ng, J. and Ryan, U.** (2010). Identification of novel *Cryptosporidium* species in aquarium fish. *Veterinary Parasitology* **174**, 43–48.
- Zrnčić, S., Le Roux, F., Oraić, D., Šoštarić, B. and Berthe, F. C. J.** (2001). First record of *Marteilia* sp. in mussels *Mytilus galloprovincialis* in Croatia. *Diseases of Aquatic Organisms* **44**, 143–148.