

# Landscape composition modulates population genetic structure of *Eriosoma lanigerum* (Hausmann) on *Malus domestica* Borkh in central Chile

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## Abstract

Landscape genetics have been particularly relevant when assessing the influence of landscape characteristics on the genetic variability and the identification of barriers to gene flow. Linking current practices of area-wide pest management information on pest population genetics and geographical barriers would increase the efficiency of these programs. The woolly apple aphid, *Eriosoma lanigerum* (Hausmann), an important pest of apple orchards worldwide, was collected on apple trees (*Malus domestica* Borkh) from different locations in a 400 km north-south transect through central Chile. In order to determine if there was population structure, diversity and flow were assessed. A total of 215 individuals from these locations were analysed using Inter Simple Sequence Repeat (ISSR) markers. Four ISSR primers generated a total of 114 polymorphic loci. The percentage of molecular variation among locations was 18%. As the algorithm used by STRUCTURE may be poorly suited for inferring the number of genetic clusters in a data set that has an IBD relationship, the number of genetic clusters in the samples was also analyzed using a Bayesian clustering method implemented in software BAPS version 4.14. We inferred the presence of four genetic clusters in the study region. Clustering of individuals followed a pattern explained by some geographical barriers. Using partial Mantel tests, we detected barriers to gene flow other than distance, created by a combination of main rivers and mountains. Although landscape genetics are rarely used in pest management, our results suggest that these tools may be suitable for the design of area-wide pest management programs.

**Keywords:** *Eriosoma lanigerum*, genetic structure, barriers to gene flow, pest, isolation-by-distance

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## Introduction

Landscape genetics that combine high resolution genetic markers with spatial data analysis have been particularly relevant when assessing the influence of landscape characteristics on the genetic variability and the identification of barriers to gene flow (Storfer *et al.*, 2007). Geographical barriers can interrupt gene flow between neighbouring populations independently from geographical distances and, thus, must be tested properly (Bossart & Prowell, 1998; Storfer *et al.*, 2007).

Several studies on landscape genetics of insect species, as endangered organisms or biodiversity indicators in forest and agricultural landscapes, have been performed during the last decade (Darvill *et al.*, 2006; Sander *et al.*, 2006a,b; Herrmann *et al.*, 2007; Vandergast *et al.*, 2007). However, much less attention has been devoted to forest and agricultural pests, although molecular markers are available for many species (Behura, 2006), and area-wide pest management programs provide valuable information about landscape attributes (Calkins & Faust, 2003; Carrière *et al.*, 2004; Beckler *et al.*, 2005; Park *et al.*, 2006). Linking current practices of area-wide pest management information on pest population genetics and geographical barriers would increase the efficiency of these programs.

The woolly apple aphid, *Eriosoma lanigerum* (Hausmann) (Hemiptera: Aphididae), is an important pest of apple orchards worldwide (Blommers, 1994). In North America, this aphid seems to be holocyclic (i.e. cyclic parthenogenesis alternating sexual reproduction with parthenogenesis), with sexual forms on elm trees (*Ulmus americana* L.) and asexual forms on apple (*Malus domestica* Borkh) (Blackman & Eastop, 1994). In other regions of the world, this aphid is anholocyclic (i.e. with loss of the sexual phase), with year round asexual forms occurring on apple trees (Blackman & Eastop, 1994). Production of sexual forms in areas with mild winters and the absence of the primary host has been reported; however, the genetic contribution of sexual genotypes to populations infesting apple trees has not been clarified (Sandanyaka & Bus, 2005; Timm *et al.*, 2005).

*E. lanigerum* can disperse through infested plant material and wind (Asante *et al.*, 1993). In regions where holocyclic genotypes are present, the production of sexual alates allows long distance aerial dispersal (Sandanyaka & Bus, 2005). There geographic barriers, such as mountains and rivers, could limit wind flow, therefore acting as barriers to dispersal and potentially modulating the population genetic structure (Westbrook & Isard, 1999). Genetic diversity of *E. lanigerum* has been studied only in South Africa, where a low genetic variation within and between locations was found, probably associated with predominantly asexual reproduction and dispersal through infested plant material (Timm *et al.*, 2005).

Since genetic variability and structure of insect pest populations, associated with life cycle characteristics, can provide some insights into the dispersal mechanisms of these, in the present study, we tested whether geographical parameters affected genetic diversity and gene flow of *E. lanigerum* in central Chile (33° to 37° latitude south). Geographical variables as possible barriers to gene flow were then individually tested in order to explain the genetic structure found.

## Materials and methods

Samples from ten orchards in different locations were collected in central Chile during spring and summer of

2005/06. Sampling locations were: Ancoa (S 35.91943 W 71.33041, 404 m), Colín (S 35.46507 W 71.73353, 47.5 m), El Colorado (S 35.62074 W 71.27864, 405 m), Villa Alemana (S 33.19079 W 71.37163, 106 m), Los Niches (S 35.04175 W 71.18500, 235 m), Panguilemo (S 35.37091 W 71.59524, 121 m), Pencahue (S 35.38667 W 71.81009, 65.3 m), San Fernando (S 34.57442 W 70.98117, 314 m), Colbún (S 35.68596 W 71.47453, 234 m) and Chillán (S 36.59805 W 72.08192, 133 m). From each location 30–40 wingless adult females of *E. lanigerum* were collected in 95% alcohol to avoid DNA degradation (Sunnucks & Hales, 1996). To avoid collecting specimens from the same asexual lineage (clones), all individuals were collected from different trees. Individuals were examined under the microscope, and parasitized aphids were discarded to avoid amplification of parasitoid DNA.

### DNA extraction and PCR amplification

The genomic DNA was obtained following the 'salting out' procedure from Sunnucks & Hales (1996). DNA template was suspended in 40 µl of distilled sterile water. A total of 18 ISSR primers (Biotechnology Laboratory from University of British Columbia, Vancouver) were tested. The inter-simple sequence repeats (ISSR) are a very useful tool for studies of genetic diversity (Wolfe *et al.*, 1998), which are based on the use of the microsatellites sequences as primers to generate multiple band patterns. This is a dominant genetic marker, which does not allow differentiating between homozygote and heterozygote genotypes, which are necessary to calculate allele frequencies. However, its main advantage is that no previous knowledge of the genome is necessary, which makes this technique a quick and economic alternative with higher reliability than random amplification of polymorphic DNA (RAPDs) when more informative co-dominant markers are not available (Zietkiewicz *et al.*, 1994; Abbot, 2001).

PCR reactions were carried out in a Mastercycler<sup>®</sup> gradient Eppendorf thermocycler with a 0.5 ng µl<sup>-1</sup> DNA template, 2.5 mM MgCl<sub>2</sub>, 0.2 mM dNTP, one U *Taq* DNA polymerase Invitrogen, 0.75 µM primers in a final volume of 20 µl. PCR consisted of a program of 5 min initial denaturation at 94°C and then 35 cycles of a 1 min denaturation at 94°C, 1.5 min annealing (see table 1 for annealing temperatures), 7 min extension at 74°C and a final extension at 74°C for 4 min. PCR products were separated in a 2% agarose gel with a GeneRuler<sup>™</sup> 100 bp DNA ladder plus (Fermentas). Band weight, presence and absence were determined for all samples.

### Statistical analysis

A total of 114 loci generated from the best four ISSR primers, based on the consistency of the resulting PCR products, were considered for analysis (table 1). Analysis of molecular variance (AMOVA), pairwise genetic difference between loci and a Mantel test to check for isolation by distance (IBD) was done using ARLEQUIN 3.1 software (Excoffier *et al.*, 2005). Nei's (1973) gene diversity for each location was calculated with software POPGENE version 1.31 (Nei & Li, 1979). Using GenAlEx version 6 (Peakall & Smouse, 2006),  $\Phi_{PT}$ , which is the proportion of the variance among populations relative to the total variance and pairwise  $\Phi_{PT}$  between locations, was calculated in order to assess total variance between all populations and the proportion of

Table 1. Primer sequence, number of bands, polymorphic bands and annealing temperatures for *E. lanigerum*.

Primer	N° Bands	5'–3' Primer sequence	Annealing Temperature (°C)
UBC864	27	ATGATGATGATGATGATG	51
UBC841	38	GAGAGAGAGAGAGAYC	56
UBC850	26	GTGTGTGTGTGTGYC	54
UBC848	23	CACACACACACACARG	54

Mixed based positions Y: (C, T) R: (A, G).

variance between the different collecting locations (orchards).  $\Phi_{PT}$  is analogous to  $F_{st}$  when the data are haploid or binary (Maguire *et al.*, 2002).

### Population structure

The use of different methods to study the spatial genetic structure of organisms in a sampled region has been strongly recommended in the literature (Pearse & Crandall, 2005; Frantz *et al.*, 2006; Storer *et al.*, 2007). Analyses methods such as AMOVA have the disadvantage that populations must be defined *a priori*; therefore, the significance of values may change depending on *a priori* grouping. In contrast, Bayesian methods have the advantage of inferring populations based on the frequencies of the alleles, thus clustering individuals based on their genetic values (Bolstad, 2004). Therefore, we selected two Bayesian methods for determining the genetic structure of *E. lanigerum*. In the first method, population structure within *E. lanigerum* samples was inferred using a Bayesian model clustering algorithm implemented in the computer programme STRUCTURE version 2.1 (Pritchard *et al.*, 2000; Pritchard & Wen, 2003). To run the programme, a number  $K$  of genetic clusters, characterized by the matrices of allele frequencies at each locus, is first assumed. Then, for each individual, the proportion of its genome derived from each genetic cluster (proportion of ancestry) is estimated. Five independent runs of the algorithm, assuming values of  $K$  from 1 to 11 with 600,000 Markov chain Monte Carlo (MCMC) repetitions and a burning period of 60,000, were performed, assuming population admixture. The posterior probability (probability of  $K$  given the data) was then calculated for each mean value of  $K$  using the mean estimated log-likelihood of  $K$  in order to choose the optimal  $K$ . The proportion of ancestry in a given cluster was calculated as an assignment rate ( $q$ ), in general a level of 0.8 determines a correct assignment to a single cluster (Vialatte *et al.*, 2005). The algorithm used by STRUCTURE may be poorly suited for inferring the number of genetic clusters in a data set that has an IBD relationship (Pritchard & Wen, 2003; Evanno *et al.*, 2005). Considering the latter, the second method was a Bayesian clustering method described in Corander *et al.*, (2003) and implemented in software BAPS version 4.14 (Corander *et al.*, 2006). In contrast with STRUCTURE, it uses stochastic optimization to infer the genetic structure; and it can use a spatial model that takes into account individual geo-referenced multilocus genotypes to assign the biologically relevant structure, thereby increasing the power to detect correctly the underlying population structure (Corander *et al.*, 2006). Ten independent repetitions for each  $K$  from 1 to 11 were carried out.

### Barriers to gene flow

Models have shown that wind plays a significant role in the distribution of aphids (Loxdale & Lushai, 1999; Zhu *et al.*, 2006) and, thus, their persistence in habitat patches (Parry *et al.*, 2006). In particular, rivers and mountains could alter normal wind conditions (through vertical flow or canyon-like transversal winds) (DGA, 1998) affecting aphid movement. By performing multiple stepwise regressions of gene flow and standardized genetic differentiation ( $\Phi_{PT} \times (1 - \Phi_{PT})^{-1}$ ) (response variables) between locations and the pairwise landscape parameters, we selected possible landscape features (for example, rivers and mountains) that were influencing the population structure from official geographic charts of Chile at a 1:250,000 scale.

The pairwise landscape parameters were defined in two steps. First, for each location, the following variables were measured: distance among populations in meters (d); distance to main rivers (dr) (the minimum distance in meters (m) from each population to a main river); distance to main mountain chains in meters (dm) (the minimum distance (m) from each population to main mountain chains); distance to main populated areas (dp) (the minimum distance in meters from each location to a main populated area); road density (rd) (kilometers of road per 25 km<sup>2</sup> of terrain).

Second, using this information for each location, pairwise values were calculated as the Euclidean distance among them using the following expression:

$$\Delta LB_{i,j} = \sqrt{(LB_i - LB_j)^2}$$

Where LB are the landscape variables: d, dr, dm, dp or rd;  $i, j$  are two different locations; and  $\Delta LB_{i,j}$  is the Euclidean distance between population  $i$  and  $j$  for the landscape variable (D, DR, DM, DP and RD).

In the study site, the general landscape corresponds to an agricultural flat surrounded by mountains of a generally abrupt topography and elevations reaching up to 500 m.a.s.l. In central Chile, main inhabited areas are located in the north-south direction and rivers cut the landscape in the east-west direction. Thus, landscape parameters were extracted from an image mosaic, using a geographical information system in ArcView. The image mosaic was geographically projected using the coordinates systems UTM -WGS84 and all landscape parameter were measured in meters. All coding and analyses were performed with SPLUS version 6.0 (Insightful, 2001), and ArcView version 3.2.

Pairwise gene flow ( $Nm$ ) between locations was calculated using GenALEX version 6. A second matrix was generated from the location coordinates containing the pairwise geographic distances between the individuals.

From the variables that entered the stepwise models (DR and DM), we selected the features that could influence gene flow (rivers and mountains). In order to test whether particular rivers and mountains were impeding gene flow between locations independent from the geographical distance effect, a third matrix was built. To build the third matrix, combinations of individuals from locations on one side of the selected barrier were recorded as 1 and pairs from different sides as 0. Putative barriers from the map were chosen based on the spatial analyses performed by BAPS version 4.14, which were effective interruptions in the landscape (rivers and mountains) between the generated genetic clusters. Using zt version 1.0 (Bonnet & Van de Peer, 2002), a partial

Table 2. Sample size, totals for binary band patterns, percent polymorphisms and Nei's (1973) gene diversity per location.

Location	Ancoa	Chillán	Colín	El Colorado	San Fernando	Los Niches	Panguilemo	Pencahue	Colbún	Villa Alemana
No. Collected	40	30	30	30	30	30	34	35	30	40
No. Individuals with amplification	27	26	18	25	15	15	27	15	22	25
No. Bands	78	67	74	68	64	70	77	68	74	55
No. Bands Freq. $\geq 5\%$	56	47	74	56	64	70	60	68	60	47
No. Private Bands	2	0	0	0	1	1	1	0	2	0
No. LComm Bands ( $\leq 25\%$ )	4	3	7	3	0	2	7	2	4	2
No. LComm Bands ( $\leq 50\%$ )	21	20	23	20	16	18	24	17	23	9
% Polymorphism	0.68	0.59	0.65	0.59	0.54	0.61	0.68	0.60	0.65	0.47
Nei's gene diversity	0.16	0.14	0.17	0.16	0.15	0.17	0.18	0.15	0.17	0.14

Table 3. Pairwise genetic differentiation between populations  $\Phi_{PT}$  values (below diagonal) and corresponding significance after 10,000 permutations (above diagonal) are reported.

Location	Ancoa	Chillán	Colín	Colorado	San Fernando	Los Niches	Panguilemo	Pencahue	Colbún	Villa Alemana
Ancoa	**	**	**	**	**	**	**	**	**	**
Chillán	0.17	**	**	**	**	**	**	**	**	**
Colín	0.05	0.20	**	**	**	**	**	**	**	**
Colorado	0.14	0.14	0.16	**	**	**	**	**	**	**
San Fernando	0.24	0.31	0.20	0.24	**	**	**	**	**	**
Los Niches	0.17	0.22	0.13	0.14	0.12	**	**	**	**	**
Panguilemo	0.12	0.23	0.11	0.18	0.22	0.15	**	**	**	**
Pencahue	0.11	0.21	0.05	0.18	0.12	0.07	0.13	**	**	**
Colbún	0.15	0.11	0.15	0.12	0.31	0.21	0.22	0.18	**	**
Villa Alemana	0.12	0.31	0.10	0.23	0.30	0.20	0.17	0.17	0.24	**

\*\* , highly significant,  $P < 0.01$ .

Mantel test was carried out estimating the correlation and significance for each putative barrier, with 10,000 permutations (Frantz *et al.*, 2006). Thus, a significant correlation would be indicative of a barrier to gene flow between clusters other than distance.

## Results

### Genetic diversity and population differentiation

A total of 215 individual from the ten localities were considered for analyses. The four ISSR primers amplified a total of 114 loci. There were no repeated haplotypes (for numbers of bands, common and private bands per population see table 2). Mean location polymorphism was 60.53%, ranging from 47.37% to 68.42% (table 2). Nei's gene diversity per site ranged from 0.14 to 0.18 (table 2). Total  $\Phi_{PT}$  was 0.18, with 0.825 of the variation corresponding to within population variation.  $\Phi_{PT}$  values between pairs of locations varied from 0.04 to 0.32 (table 3).

### Population structure

When assigning individuals to clusters using STRUCTURE, log-likelihood values increased, with posterior probability reaching its highest value ( $\sim 1$ ) at  $K=10$ . Clustering did not respond to sampling origin or geographical regions, and individuals were not clearly assigned to a given genetic population. More than 40% of the individuals were assigned

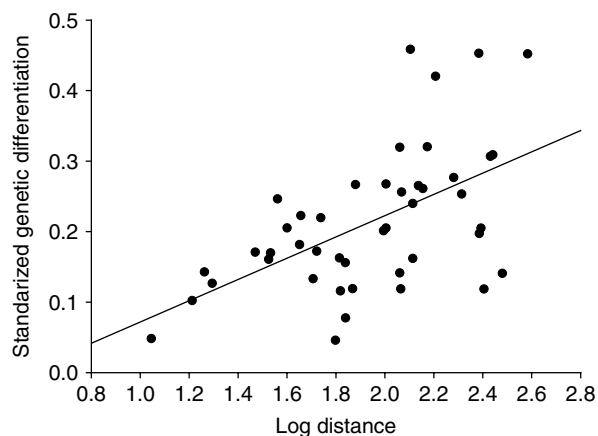


Fig. 1. Correlation between standardized genetic differentiation ( $\Phi_{PT} \times (1 - \Phi_{PT})^{-1}$ ) and the logarithm of the geographical distance (km) between locations ( $R^2 = 0.55$ ; Mantel test:  $r = 0.554$ ,  $P = 0.004$ ).

to more than one cluster, thus the resultant clustering was non reliable. The Mantel test was significant with  $\Phi_{PT} \times (1 - \Phi_{PT})^{-1}$  correlating positively with the pairwise geographical distance ( $r = 0.554$ ,  $P = 0.004$ ) (fig. 1).

The number of clusters in optimal partition assignment with BAPS determined  $K=4$ , with a log -marginal likelihood

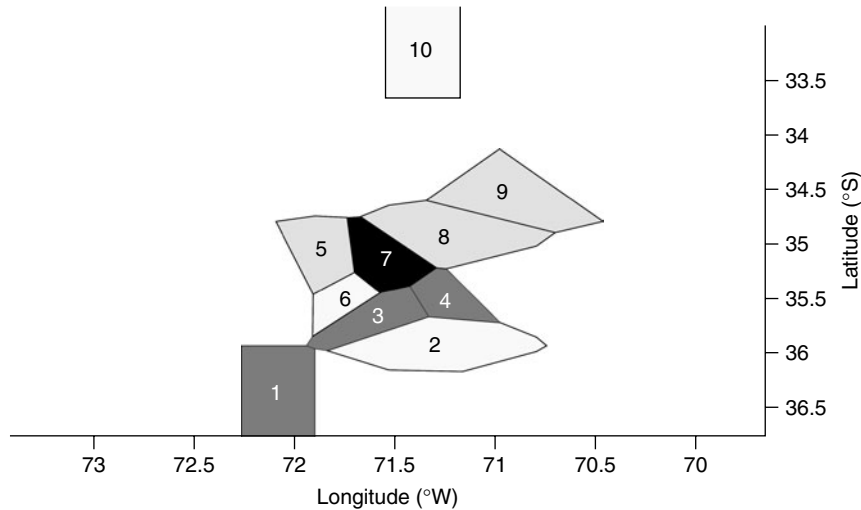


Fig. 2. Voronoi tessellations of the spatial clustering of individuals from ten locations. A cell of the tessellation corresponds to the physical neighbourhood of an observed data point and is shaded according to cluster membership. (1) Chillán, (2) Ancoa, (3) Colbún, (4) Colorado, (5) Pencahue, (6) Colín, (7) Panguilemo, (8) Los Niches, (9) San Fernando and (10) Villa Alemana (■, Cluster 1; □, Cluster 2; □, Cluster 3; ■, Cluster 4).

of optimal partition of  $-8074.93$  and the posterior probability reaching its highest value ( $\sim 1$ ). Aphids from locations Villa Alemana, Colín and Ancoa formed one cluster, whereas aphids from Panguilemo alone formed a second cluster. Aphids from Colbún, Colorado and Chillán formed a third cluster, and aphids from San Fernando, Los Niches and Pencahue formed a fourth cluster (figs 2 and 3).

#### Barriers to gene flow

When regressing the logarithm of gene flow against the landscape pairwise parameters, D and DR entered the model (D:  $\alpha = -0.00107$ ,  $SS = 0.40048$ ,  $P = 0.0021$ ; DR:  $\alpha = -0.00515$ ,  $SS = 0.10716$ ,  $P = 0.0974$ ). When regressing the standardized genetic differentiation ( $\Phi_{PT} \times (1 - \Phi_{PT})^{-1}$ ) values with the landscape parameters D and DM entered the model (D:  $\alpha = 0.00051761$ ,  $SS = 0.08499$ ,  $P = 0.0015$ ; DM:  $\alpha = 0.00262$ ,  $SS = 0.01694$ ,  $P = 0.1357$ ). The only significant variable that entered in the regressions was geographical distance.

Some main rivers and mountains separated clusters on the map, while others did not. For instance, mountains around Villa Alemana, the Claro River, the Lircay River, the Maule River and the mountains around Ancoa separated clusters (see fig. 3). A partial Mantel test confirmed that Villa Alemana mountains were barriers to gene flow other than distance ( $r = -0.44$ ,  $P = 0.01$ ). The Claro River also was confirmed as a significant barrier to gene flow other than distance between clusters ( $r = -0.45$ ,  $P = 0.01$ ). A partial Mantel test also confirmed the Lircay River as a significant barrier to gene flow other than distance ( $r = -0.44$ ,  $P = 0.01$ ) and the same for the Maule River and the Ancoa mountains ( $r = -0.42$ ,  $P = 0.02$  and  $r = -0.45$ ,  $P = 0.005$ , respectively).

## Discussion

### Genetic diversity and population differentiation

Polymorphism percentage and Nei's gene diversity in our study were higher than those from the only other study

on the genetic variation of *E. lanigerum* published to date (Timm *et al.*, 2005). Gene diversity in South African populations was extremely low, with the highest values reaching 0.024, as compared to our study where gene diversity was highest at Panguilemo with 0.18. In fact, the Villiersdorp population in South Africa was invariant (Timm *et al.*, 2005). In contrast, in our data, the lowest gene diversity was 0.14 (Chillán and Villa Alemana). Such values are similar to those reported in studies from other aphid species (Hales *et al.*, 1997) but do not provide a clear insight in terms of sexuality and life cycle predominance in aphid populations (Halkett *et al.*, 2005). Significant absence of linkage disequilibrium using codominant markers would suggest the possibility of sexual reproduction in *E. lanigerum* populations from Chile. Production of sexual males and females able to successfully oviposit on apple trees were reported from New Zealand (Sandanayaka & Bus, 2005), and a similar situation might be occurring in central Chile.

Proper testing of the presence of sexual populations of *E. lanigerum* in Chile will require more informative co-dominant molecular markers (Halkett *et al.*, 2005), as well as biological studies to identify functionality of putative sexual morphs (Sandanayaka & Bus, 2005). Isolation of microsatellites from *E. lanigerum* is underway in order to readily assess the contribution of sexual reproduction to the population genetics of this aphid species in Chile.

Genetic differentiation in our data was modest, with  $\Phi_{PT}$  values reaching 0.18. This estimate is based on genetic distance data and could be comparable with the population divergence estimates obtained for the South African populations (Timm *et al.*, 2005). This latter study also reported a low genetic differentiation probably related with asexual reproduction and spread to other regions through infested plant material from the Elgin area nurseries. The relative isolation of apple producing areas in the Western Cape and the absence of wild host for *E. lanigerum* are further factors of the South African landscape that could explain the variability found. In our study, a higher differentiation between

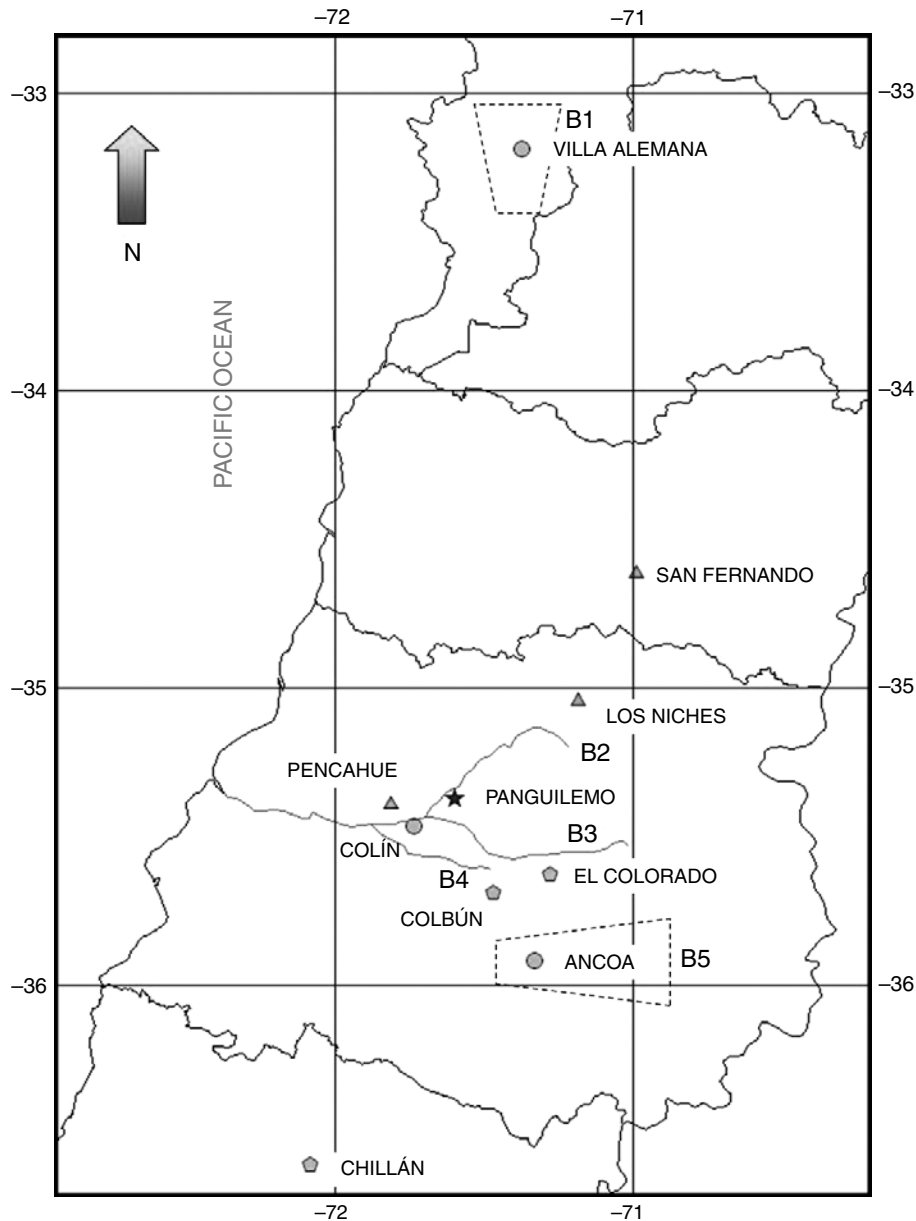


Fig. 3. Schematic diagram of putative barriers and relative position of locations to these barriers. (B1) Villa Alemana Mountains, (B2) Claro River, (B3) Lircay River, (B4) Maule River, and (B5) Ancoa Mountains.  $\blacktriangle$ , Cluster one;  $\star$ , Cluster two;  $\blacklozenge$ , Cluster three and  $\bullet$ , Cluster four.

populations could be expected if holocyclic genotypes are present and abandoned apple orchards could act as reservoirs and corridors between areas.

#### Population structure

STRUCTURE software failed to cluster individuals properly and, as noted before, did not perform well under IBD (Frantz *et al.*, 2006). The resulting grouping of individuals in ten clusters did not have any biological meaning, nor correlate with geography, and a great number of individuals were not assigned to a single cluster. The resulting clustering did not agree with AMOVA results. In contrast, BAPS assigned

individuals to four distinct groups. These same grouping were observed when considering the location of individuals and their spatial coordinates. Moreover, the clusters were in partial agreement with the AMOVA results, as locations in each cluster shared low  $\Phi_{PT}$  values; and highest  $\Phi_{PT}$  values were between locations that were assigned to other clusters, with the exception of locations Panguilemu and Colin, which shared low  $\Phi_{PT}$  values. AMOVA has the clear disadvantage of having to define populations *a priori*, thus significance of values may change depending on *a priori* grouping. In contrast, Bayesian methods have the advantage of inferring populations based on the frequencies of the alleles (Bolstad, 2004). Thus, *E. lanigerum* populations seem to consist of four

distinct clusters.  $\Phi_{PT}$  values between populations revealed considerable genetic differentiation, which was correlated with geographical distance between sampling sites.

An important difference with the results reported from South Africa was the significant IBD found in the Chilean populations. A smaller spatial scale used in South Africa (approx. 10–200 km), with a relative isolation of the fruit production areas in the Western Cape, as well as the absence of wild hosts for *E. lanigerum*, could explain this result. On the contrary, apple growing areas in Chile are located almost continuously along the central valley, flanked by the coastal and Andes ranges (Gwynne, 1999). Furthermore, domestic culturing of apple is a widespread practice in the rural Chilean landscape (Gwynne, 1999), and trees are usually maintained without insecticide treatments and act as 'stepping stones' for the dispersal of alates of *E. lanigerum*. The recent spread of modern fructiculture in Chile (the last 50 years for export-oriented production) and the introduction of *E. lanigerum* to the southern cone of America at the middle of the 19th century (Artigas, 1994) could also explain the differences reported with South Africa.

Another explanation for the differences found in the genetic structure of *E. lanigerum* could be attributed to the molecular markers used (i.e. AFLP in South Africa and ISSR in Chile). However, both are dominant markers with high reliability and should not produce different significant results (Wolfe *et al.*, 1998; Abbot, 2001; Nybom, 2004). Simultaneous results have been published only for plant species, with no major difference between the conclusions obtained (Nybom, 2004; Gao *et al.*, 2006), therefore showing the usefulness of these markers.

#### Barriers to gene flow

Clearly the most important parameter affecting gene flow seems to be distance. However, several geographical structures separate the clusters when placed on the map (fig. 3). Some main rivers and mountains effectively separate all clusters. Locations within clusters presented rivers but not mountains with the exception of cluster one (fig. 3). All locations on cluster one were isolated from the other surrounding locations by these putative barriers. As it has been suggested that *E. lanigerum* is mainly dispersed with the wind and with rootstocks, this would explain why some main rivers and mountains could isolate populations. The most geographically isolated locations happened to be in cluster one. Individuals here could be brought to apple orchards with the rootstocks, suggesting a similar origin and, as they are isolated, limited or no flow with other populations from other clusters, might have maintained these populations with no or little change.

The Claro River separates cluster two, from the other clusters; this river is a canyon-like river with a higher north edge (DGA, 1998). As winds move predominantly in a south–north direction during summer and spring (Santibáñez, 1993) and rivers in Chile cut the landscape in a west–east direction (Solervicens, 1995), aphids carried by the wind would not reach the north edge. On the other hand, canyon-like rivers generate wind flows of their own, which could disperse aphids, impeding movement to the north.

Interestingly, other important rivers that were within genetical clusters were not found as barriers. This was the case of the Teno and Tinguiririca rivers within cluster two. This could be due to the fact that these rivers are not

canyon-like rivers and are quite wide with very variable flow from season to season (DGA, 1998). It remains unanswered why some geographical structures that are expected to affect aphid population connectivity do not do so.

Although landscape genetics are rarely used in pest management, our results suggest that area-wide management programs can use this type of information in order to inform management decisions, identifying the source of insect spread and the possible barriers to further dispersal. For example, eradication of a close and small genetic population with limited flow with other areas, such as Panguilemo (cluster four), should be feasible.

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