cambridge.org/ssr

Research Paper

Cite this article: Hepp J, Gómez M, León-Lobos P, Montenegro G, Vilalobos L, Contreras S (2021). Characterisation of seed dormancy of 12 Chilean species of *Nolana* (Solanaceae) from the coastal Atacama Desert. *Seed Science Research* **31**, 20–29. https://doi.org/10.1017/ S0960258520000434

Received: 3 February 2020 Revised: 13 August 2020 Accepted: 16 November 2020 First published online: 22 December 2020

Key words:

endosperm; funicular plug; germination; lomas vegetation; mericarp; physiological dormancy

Author for Correspondence: Samuel Contreras, E-mail: scontree@uc.cl

© The Author(s), 2020. Published by Cambridge University Press



Characterisation of seed dormancy of 12 Chilean species of *Nolana* (Solanaceae) from the coastal Atacama Desert

Josefina Hepp^{1,2}, Miguel Gómez^{1,2}, Pedro León-Lobos³, Gloria Montenegro^{1,2}, Luis Vilalobos⁴ and Samuel Contreras^{1,2}

¹Departamento de Ciencias Vegetales, Facultad de Agronomía e Ingeniería Forestal, Pontificia Universidad Católica de Chile, Santiago, Chile; ²Centro del Desierto de Atacama, Pontificia Universidad Católica de Chile, Santiago, Chile; ³Centro Regional de Investigacion La Platina, Instituto de Investigaciones Agropecuarias, INIA, Santiago, Chile and ⁴Laboratorio de Fisiología del Estrés en Plantas, Facultad de Ciencias Agronómicas, Universidad de Chile, Santiago, Chile

Abstract

The genus Nolana (Solanaceae) comprises numerous species endemic to the coastal Atacama Desert of Chile and Peru of high ornamental potential and conservation value. The environments in which these species have evolved and are present today correspond to particular conditions in the midst of a hyper-arid habitat, so the study of their germination requirements and characterisation of seed dormancy becomes important in terms of conservation but also for ecological and evolutionary purposes. Different treatments were performed on mericarps of 12 species of Nolana: control (intact seeds imbibed in distilled water), scarification in funicular plug and distilled water and scarification in funicular plug and addition of GA3 (500 ppm); their permeability to water was also tested. It was determined that the species did not present physical dormancy, as had been previously reported, but rather physiological dormancy (PD). Germination results after treatments were not homogeneous among all 12 species, indicating differences in their dormancy levels. Also, the important role of the endosperm in the prevention of germination for the studied Nolana species was highlighted. Regarding the relationship between the level of PD (expressed as the percentage of germination for the most successful treatment) and the latitudinal distribution of the species or their phylogenetic closeness, it was determined that, for the studied species, their proximity in terms of clades was more relevant than their latitudinal distribution.

Introduction

Seed dormancy in environments with high temporal and spatial variability, such as deserts, is often described as a bet-hedging strategy (Eberhart and Tielbörger, 2012). Not all mature seeds germinate immediately after dispersal; in many cases, a significant fraction remains dormant for a period that can last for years (Fenner, 1985). These 'persistent seed banks' are considered one of the main strategies of desert plants (Figueroa et al., 2004; Facelli et al., 2005; Baskin and Baskin, 2014). Larger proportions of dormant species are found in environments with distinctive unfavourable seasons (Jurado and Flores, 2005); the percentage increase from 40% in tropical rainforests to over 80% in hot deserts (Gutterman, 2002).

The coastal Atacama Desert of northern Chile and southern Peru has a temperate climate with constant presence of fog, a decisive ecological factor to which the maintenance of formations known as 'fog oases' (Ellenberg, 1959, cited in Rundel et al., 1991) or 'lomas vegetation' (Rundel et al., 1997) is attributed. These are isolated ecosystems, separated by a hyper-arid habitat (Rundel et al., 1991) located along 3500 km of coastal desert (Dillon et al., 2009); lomas sustain over 1400 plant species with an endemism of around 40% (Dillon et al., 2009; Schulz et al., 2011).

The genus *Nolana* L. ex L. f. (Solanaceae) currently comprises 90 species, mostly distributed in the costal Atacama Desert, of which 46 are present exclusively in Chile. They are annual or perennial herbaceous plants or small shrubs (Dillon et al., 2007; Tu et al., 2008) and present foliar succulence and trichomes that capture moisture and restrict transpiration (Dillon et al., 2009). *Nolana* is the only genus found in the whole range of lomas, where it stands out as the most conspicuous and diverse floristic element (Tago-Nakazawa and Dillon, 1999). Some species are important components of the blooming desert phenomenon, associated with El Niño Southern Oscillation events of heavy rainfall and relatively high temperatures (Dillon, 2005). Several of these species have high ornamental potential (Freyre et al., 2005; Riedemann et al., 2006; Fig. 1), and also high conservation value (Tu et al., 2008; Dillon et al., 2009) given their endemism to habitats with extreme conditions of aridity and salinity. Dillon et al. (2009), using molecular markers, identified several strongly supported clades within *Nolana*, with geographic and morphological



Fig. 1. Context, flowers and fruits of four Nolana species included in this study. (A–C) N. divaricata (clade G); (D–F) N. intonsa (clade F); (G–I) N. carnosa (clade C); (J–L) N. jaffuelii (clade B).

fidelity, four of which are confined to Chile and three are mainly Peruvian with some presence in Chile.

Physiological non-deep dormancy, apparently induced by endosperm and testa, has been detected in Solanaceae (Finch-Savage and Leubner-Metzger, 2006; Freyre et al. (2005) and Douglas and Freyre (2006), studying *Nolana paradoxa* and *N. aplocaryoides* among others, found that germination occurs when opening a 'funicular plug' in mericarps and increases when adding gibberellic acid. Douglas (2007) also reported an increased germination effect on nine *Nolana* species with gibberellic acid and dry storage for 2 years, suggesting physiological dormancy (PD). Cabrera et al. (2015) obtained a higher percentage of germination in *N. jaffuelii* with scarification in the funicular area and application of gibberellic acid, reporting physical and PD. No reference was found regarding dormancy for any of the other 11 *Nolana* species included in this research.

Several studies report a decline in lomas vegetation of the coastal desert (Muñoz-Schick et al., 2001; Pinto et al., 2001; Egaña et al., 2004; Pinto and Luebert, 2009). Since projections indicate that rainfall will continue to decrease in arid and semiarid areas, while average temperatures could rise (Santibañez et al., 2017), lomas vegetation is expected to continue deteriorating (Schulz et al., 2011). It is crucial to establish the germination requirements and dormancy mechanisms of these species, so that their storage in seed banks and subsequent propagation for *in situ* reintroduction or *ex situ* cultivation can be successful (León-Lobos et al., 2012; León-Lobos et al., 2020).

Seed germination and dormancy of phylogenetically closely related species have not been extensively studied, but there are reports of similar germination requirements and local adaptations mediated by species relatedness (Carta et al., 2016) and of the major role of phylogeny in determining seed dormancy occurrence (Dayrell et al., 2016). Also, a relationship between dormancy level and habitat for certain species (Meyer et al., 1995) has been found. However, for *Nolana*, such relationships have not been established. Therefore, the objective of this study was to characterise dormancy of 12 *Nolana* species from the coastal desert of Chile, to determine their germination requirements, and to evaluate if there is any relationship between dormancy level and geographical distribution or phylogeny.

Material and methods

Plant material

Between October 2014 and April 2016, mericarps of 12 *Nolana* species (Fig. 1) were collected at different locations (Table 1) from individual plants at maturity and kept in labelled paper bags. Whenever possible, mericarps were collected randomly within the population from at least 50 different individuals, to obtain a representative sample for each species (León-Lobos et al., 2003). Fruits were then kept in paper bags, partially dried (they were put inside jars with equal weight of silica gel, for 2–3 d) and stored at 20and 40% RH until used for analysis. Selection of species is the result of availability of suitable fruits (i.e. mature mericarps, black/brown in colour and with dry or senescent calyx) in the field, which explains the overrepresentation of certain clades.

In *Nolana*, dispersal units are fruits called mericarps, which can vary in number between species from 2 to 30 per schizocarp and from laterally united and multi-seeded to completely free and single-seeded (Tago-Nakazawa and Dillon, 1999; Knapp, 2002). Seeds within the mericarp remain in independent chambers (Saunders, 1936) and are firmly embedded within the fruit. To facilitate analysis, in this study, we consider only one seed per mericarp, so 'seed' will be the unit in which results are expressed.

Seed viability

For evaluation of viability, a tetrazolium test with 2,3,5triphenyl-2H-tetrazolium chloride (Tz) was performed on a sample of at least 25 mericarps for each seed lot (according to disponibility). This was performed in a separate sample of mericarps, and not in the same fruits after finishing the germination experiments, given the extension of the experiments and the possibility of fungal damage. Mericarps were scarified (one funicular plug per mericarp was removed) and stained with 1% Tz solution for approximately 24 h at 30°C. After that time, mericarps were cut in halves and surfaces were observed using a magnifier. Only seeds that were completely stained red (embryo and endosperm) were considered viable; one seed per mericarp was evaluated.

Seed imbibition

Imbibition in methylene blue $(1 \text{ g } 100 \text{ ml}^{-1})$ was used to evaluate if water was able to penetrate and reach the embryo in intact mericarps. Twenty mericarps of each species were left at room temperature in metal containers with enough dye to cover them. Five mericarps were evaluated after 3, 24, 48 and 196 h. They were washed with distilled water and dried with paper towel, then allowed to air dry for 20 min at room temperature. Finally, stained seed were sectioned using a scalpel and observed under a stereomicroscope system (SZX7, Olympus Corporation, Tokyo, Japan), making sure that the cut allowed to see the plug and half the embryo (longitudinal cut).

Evaluation of germination and characterisation of dormancy

Evaluation of germination for the 12 species was performed in three experiments. Mericarps were placed in 9 cm diameter Petri dishes, over three layers of filter paper, saturated in distilled water or a solution of gibberellic acid (GA₃). Plates were sealed with Parafilm to prevent drying and, during evaluation, water or GA₃ solution was added when needed in order to keep moisture. Seed was considered germinated if radicle emergence was over 2 mm. Results were reported as total germination percentage.

Experiment 1. Role of funicular plug on germination

Germination tests were carried out using mericarps from ten Nolana species within 1-3 months from their collection. In the case of N. crassulifolia, N. divaricata and N. sedifolia, mericarps from different collection dates and/or sites were evaluated separately. Mericarps were randomly assigned to one of the following treatments: (1) control (Ct), intact dry mericarps imbibed in distilled water; (2) scarification (Sc), removal of funicular plug and imbibition in distilled water; (3) Sc and imbibition in a solution of 500 ppm GA₃ (Sc + GA500). A scalpel was used to dissect the pericarp at the funicular scar area avoiding radicle damage, as this proved to be the most effective treatment for the germination of mericarps of N. jaffuelii (Cabrera et al., 2015). Scarification was performed on one funicular plug, chosen randomly, per mericarp. The presence of an embryo was verified for that selected seed; if there was no embryo, a second funicular plug was removed. Germination was evaluated 3 d a week during 45 d in a chamber at constant 20°C, with constant fluorescent light (10 to 20 μ mol m⁻² s⁻¹ photosynthetically active radiation), in four replicates of 25 seeds each.

Experiment 2. Alternatives to overcome PD

This experiment was conducted in mericarps of *N. linearifolia*, which was selected due to the number of available mericarps. The following germination treatments were evaluated: (1) control (Ct), intact mericarps imbibed in distilled water; (2) plug scarification (Sc), removal of funicular plug and imbibition in distilled water; (3) plug scarification with partial removal of endosperm (Sc + en), imbibed in distilled water and (4) Sc + en and imbibition in a solution of 500 ppm GA₃ (Sc + en + GA500). Germination was evaluated three times a week in four replicates (25 seeds each) per treatment, during 45 d in a chamber at constant 20°C.

Experiment 3. Role of endosperm on germination

Based on observations after experiments 1 and 2, and to better understand the role of endosperm on germination, a third experiment was performed using mericarps from four species that had high germination percentage in Exp. 1, and two additional species (*N. aplocaryoides* and *N. paradoxa*) that were collected and included afterwards. Three of the treatments defined in Exp. 2 were evaluated: Ct, Sc and Sc + en. Germination was evaluated three times a week in four replicates (25 seeds each) per treatment, during 45 d in a chamber at constant 20°C. Table 1. Species of the genus Nolana used in the study, latitudinal distribution in Chile, collection site and clade (Flora del Conosur, 2014 and Enciclopedia de la Flora Chilena, 2014)

| Species | Latitudinal distribution ^a | Collection site and date (month/year) | Clade ^b |
|--|---------------------------------------|---|--------------------|
| Nolana aplocaryoides (Gaudich.) I.M. Johnst. | 22°04'S/27°04'S | Pan de Azúcar, Atacama (26°09'S, 70°40'W); 04/16 | G |
| Nolana crassulifolia Poepp. | 26°21'S/33°55'S | Llanos de Challe, Atacama (28°11'S, 71°10'W); 10/14 | _ |
| | | Punta de Choros, Coquimbo (29°15'S, 71°28'W); 01/15 | |
| | | Los Molles, Valparaiso (32°14'S, 71°31'W); 04/16 | _ |
| Nolana divaricata (Lindl.) I.M. Johnst. | 23°31'S/30°54'S | Llanos de Challe, Atacama (28°12'S, 71°10'W); 01/15 and 04/16 | _ |
| | | Punta de Choros, Coquimbo (29°15'S, 71°28'W); 01/15 | _ |
| Nolana linearifolia Phil. | 21°56'S/26°02'S | Hills of Antofagasta, Antofagasta (23°30'S, 70°23'W); 08/15 | _ |
| Nolana onoana M.O. Dillon & Nakazawa | 23°01'S/23°52'S | Hills of Antofagasta, Antofagasta (23°30'S, 70°22'W); 08/15 | _ |
| Nolana sedifolia Poepp. | 19°56'S/33°06'S | Alto Patache fog oasis, Tarapacá (20°50'S, 70°09'W); 12/15 | |
| | | Los Molles, Valparaíso (32°14'S, 71°31'W); 01/15 and 04/16 | _ |
| Nolana intonsa I.M. Johnst. | 19°56'S/21°25'S | Alto Patache fog oasis, Tarapacá (20°49'S, 70°09'W); 12/15 | F |
| Nolana rostrata (Lindl.) Miers ex Dunal | 18°50'S/30°03'S | Llanos de Challe, Atacama (28°07'S, 71°09'W); 01/15 | С |
| Nolana carnosa (Lindl.) Miers ex Dunal | 26°09'S/28°35'S | Llanos de Challe, Atacama (28°03'S, 71°07'W); 04/16 | _ |
| Nolana jaffuelii I.M. Johnst. | 20°13'S/26°09'S | Alto Patache fog oasis, Tarapacá (20°49'S, 70° 10'W); 12/15 | В |
| Nolana paradoxa Lindl. | 29°54'S/43°21'S | Los Molles, Valparaíso (32°14'S, 71°31'W); 04/16 | _ |
| Nolana parviflora (Phil.) Phil. | 27°04'S/28°13'S | Hills of Antofagasta, Antofagasta (23°30'S, 70°23'W); 08/15 | |

^aLatitudinal distribution for all species was obtained from an extensive study of all Herbaria specimens (SGO and CONC), see Hepp (2019) for a detailed description. ^bClades (and letters) used in this study are those determined by Dillon et al., 2009.

A constant temperature of 20°C was chosen for several reasons: (1) it was necessary to have the same set of experiments to apply to all species; (2) the moderate temperatures of the Chilean coast, on average, are around 20°C for the months in which germination usually occurs (Fick and Hijmans, 2017); and (3) the constant temperature of 20°C was effective in evaluations conducted by Cabrera et al. (2015).

The mean germination time (MGT) was calculated by the following equation (Ellis and Roberts, 1980): MGT = $\sum (n \times d)/N$, where *n* is the number of seeds germinated in each evaluation day, *d* is the number of days from sowing, and *N* is the total number of seeds germinated.

Statistical analysis

Data analysis was performed using R software (version 3.3.3, R core team 2017). The effects of treatments on germination percentages were analysed for each species using generalised linear models (GLM) with binomial distribution and logit link function in the *glm* function. In cases in which overdispersion for binomial errors occur, quasibinomial distribution and *F* tests with an empirical scale parameter were used instead of chi-square. When significant differences were detected (P < 0.05), the least significant difference (LSD; $\alpha = 0.05$) *post hoc* test was used from *glht* package to detect significant differences in the comparison between pairs of treatments. Pearson's correlation coefficient was used to evaluate the relationship between germination and latitudinal distribution of species.

Results

Seed imbibition

When mericarps from the 12 Nolana species included in this study were embedded in a solution of methylene blue, we found

that water was able to enter until reaching the embryo for all the mericarps and species, usually within 48 h since imbibition started. Images of mericarps from four representative *Nolana* species after 3 h, 48 h and 8 d of imbibition are presented in Fig. 2. The blue staining allows to see that water entrance to the embryo occurred through the funicular plug.

Evaluation of seed germination

Experiment 1

There were significant differences in germination percentages between control and some of the treatments in nine of the ten *Nolana* species evaluated in Exp. 1 (Table 2). In four species, there was a significant increment of germination when the funicular plug was removed, and in seven species, the imbibition of scarified mericarps in gibberellic acid had a positive effect on germination.

Experiment 2

Results from Exp. 2, carried out with *N. linearifolia*, confirm that some of the treatments had a significant effect in promoting seed germination (P < 0.001). While intact mericarps only reached 2% germination, treatments that combine removal of the funicular plug with partial removal of endosperm reached 54% germination when imbibed in water (Sc + en) and 55% when imbibed in a GA₃ solution (Sc + en + GA500) (Fig. 3). When the funicular plug was removed and mericarps were imbibed in water (Sc), germination was 3%, similar to control.

Experiment 3

Germination and the viability percentages of the six *Nolana* species evaluated in Exp. 3 are presented in Table 3. In five species, there was a significant promotion of germination when the funicular plug was removed, only in *N. linearifolia*, this effect



Fig. 2. Longitudinal sections of intact mericarps (with funicular plug) of *N. divaricata, N. jaffuelii, N. linearifolia* and *N. sedifolia* at 3 h, 24 h, 48 h and 8 d after imbibition in methylene blue. Abbreviations: em: embryo; fp: funicular plug; ra, radicle; * indicates regions which have imbibed.

was not observed. Additionally, in the six *Nolana* species, the treatment that combines the removal of the plug with a partial removal of the endosperm had a significantly higher germination than only removing the plug.

A summary of germination values from intact mericarps and after the best germination treatment for the 18 accessions evaluated in this study is presented in Table 4. In this case, germination is expressed as a percentage of live seed, which was calculated from the estimations of seed viability that are presented in Tables 2 and 3. The MGT, calculated from the mericarps under the best germination treatment, is also presented in Table 4.

Figure 4 shows the results from the analysis of germination values after the best treatment (Table 4) of *Nolana* species grouped by clades. In this case, there was a significant difference among the clades, with clade G having higher germination than clades B and C.

Table 2. Viability and germination percentages of mericarps from 13 accessions of 10 *Nolana* species in response to three treatments: control, intact mericarps imbibed in distilled water; scarification (Sc), removal of funicular plug and imbibition in distilled water; scarification and imbibition in a solution of 500 ppm GA_3 (Sc + GA_3)

| | | | | | Germination (%) ¹ | | |
|------------------|-------|-------------------|------------------|---------|------------------------------|-------------|------------------------------|
| Species | Clade | Collection site | Viable seeds (%) | Control | Sc | $Sc + GA_3$ | <i>P</i> -value ² |
| N. crassulifolia | G | Llanos de Challe | 65 | 1 b | 40 a | 62 a | 0.0035 |
| N. crassulifolia | G | Punta de Choros | 60 | 16 | 16 | 34 | 0.0630 |
| N. divaricata | G | Llanos de Challe | 50 | 0 b | 23 a | 35 a | <0.0001 |
| N. divaricata | G | Punta de Choros | 91 | 0 b | 4 b | 37 a | 0.0009 |
| N. linearifolia | G | Antofagasta hills | 96 | 3 b | 3 b | 55 a | <0.0001 |
| N. onoana | G | Antofagasta hills | 75 | 15 b | 25 b | 54 a | 0.0007 |
| N. sedifolia | G | Alto Patache | 61 | 0 c | 25 b | 53 a | <0.0001 |
| N. sedifolia | G | Los Molles | 38 | 1 b | 24 a | 30 a | 0.0033 |
| N. intonsa | F | Alto Patache | 40 | 0 | 0 | 0 | |
| N. carnosa | С | Llanos de Challe | 96 | 0 b | 2 b | 13 a | 0.0134 |
| N. rostrata | С | Llanos de Challe | 76 | 0 b | 0 b | 17 a | <0.0001 |
| N. jaffuelii | В | Alto Patache | 88 | 0 b | 5 b | 27 a | <0.0001 |
| N. parviflora | В | Antofagasta hills | 84 | 0 b | 19 a | 25 a | 0.0040 |

Percentage of seed viability was determined by tetrazolium test in a separate mericarp sample.

¹For each species and collection, germination values with different letters are significantly different according to a LSD test ($\alpha = 0.05$).

²P value from a generalised linear models (GLM).

When germination results of the studied *Nolana* species (corrected by the percentage of seed viability) were correlated with the latitudinal distribution of each species (Fig. 5), no evident relationship was obtained (r = 0.45; *P*-value = 0.06).

Discussion

The role of the funicular plug on germination

A thick layer of sclereids was identified by Cabrera *and* colleagues (2015) in *Nolana jaffuelii* mericarps, and authors suggested that this species presented physical dormancy. However, our results show that water was able to enter the mericarps until reaching the embryo in the 12 *Nolana* species studied and confirm that the entrance route of the water is the funicular plug (Fig. 2). Douglas (2007) also reports that the imbibition path in *Nolana* mericarps occurs through tracheid tubes in the funicular plugs. Since physical dormancy is defined as impermeability of the fruit or seed to water (Baskin and Baskin, 2004, 2014), these results indicate that dormancy in *Nolana* species would not be physical.

Seeds with PD are permeable to water and possess a physiological inhibiting mechanism that prevents radicle emergence (Baskin and Baskin, 2014). In hot semideserts and deserts, PD is the most common type of seed dormancy for shrubs, perennial succulents, herbaceous perennials and annuals (Baskin and Baskin, 2014). Taking into account our imbibition data (Fig. 2) and reports by previous studies (Douglas and Freyre, 2006; Douglas, 2007), and following the definition by Baskin and Baskin (2014), our results (Tables 2 and 3) indicate that seeds of the studied *Nolana* species present PD. Morphological or morphophysiological dormancy was discarded as a set of mericarps of all species were dissected prior to imbibition (Hepp, 2019); embryo size was then fully developed and did not change after imbibition.



Fig. 3. Cumulative germination of *Nolana linearifolia* mericarps after different treatments. Control: intact mericarps imbibed in distilled water; Sc: plug scarification, that is, removal of funicular plug, imbibed in distilled water; Sc + en: plug scarification with partial removal of endosperm, imbibed in distilled water; Sc + en + GA500: plug scarification with partial removal of endosperm, imbibed in gibberellic acid (500 ppm GA₃). Data presented are an average of four replicates of 25 mericarps each and bars represent the 95% confidence intervals. For each evaluation date, germination values with different letters are significantly different according to an LSD test (α = 0.05).

The role of endosperm on germination

While removal of the funicular plug improved germination in mericarps of six of the 12 *Nolana* species studied, an additional and significant effect was observed when this scarification was combined with imbibition in gibberellic acid (Table 2) or partial removal of the endosperm (Table 3). The extraction of the

Table 3. Viability and germination percentages of mericarps from six *Nolana* species in response to three treatments: control, intact mericarps imbibed in distilled water; scarification (Sc), removal of funicular plug and imbibition in distilled water; scarification and partial removal of endosperm (Sc+en) with imbibition in distilled water

| Species | Clade | Collection site | Viable seeds (%) | | Germination (%) ¹ | | |
|------------------|-------|-------------------|------------------|---------|------------------------------|---------|---------|
| | | | | Control | Sc | Sc + en | |
| N. aplocaryoides | G | Pan de Azúcar | 96 | 0 c | 23 b | 63 a | <0.0001 |
| N. crassulifolia | G | Los Molles | 48 | 6 c | 22 b | 55 a | <0.0001 |
| N. divaricata | G | Llanos de Challe | 84 | 0 c | 23 b | 40 a | <0.0001 |
| N. linearifolia | G | Antofagasta hills | 96 | 3 b | 3 b | 54 a | <0.0001 |
| N. sedifolia | G | Los Molles | 60 | 1 c | 22 b | 40 a | <0.0001 |
| N. paradoxa | В | Los Molles | 84 | 2 c | 20 b | 54 a | <0.0001 |

Percentage of seed viability was determined by tetrazolium test in a separate mericarp sample.

¹For each species and collection, germination values with different letters are significantly different according to an LSD test ($\alpha = 0.05$).

²P value from a generalised linear models (GLM).

 Table 4. Germination percentages of control (intact mericarps imbibed in distilled water) and best germination treatment (BGT) for 18 accessions of 12 species of Nolana, corrected according to viability (% of live seed)

| Species | Clade | Collection site and date (month/year) | Control (%) | BGT (%) | MGT | BGT |
|------------------|-------|---------------------------------------|-------------|---------|------|----------------------|
| N. aplocaryoides | G | Pan de Azúcar, 04/16 | 0 | 66 | 18.0 | Sc + en |
| N. crassulifolia | G | Llanos de Challe, 10/14 | 2 | 95 | 16.6 | $Sc + GA_3$ |
| N. crassulifolia | G | Punta de Choros, 01/15 | 27 | 57 | 8.2 | Sc + GA ₃ |
| N. crassulifolia | G | Los Molles, 04/16 | 11 | 100 | 9.5 | Sc + en |
| N. divaricata | G | Llanos de Challe, 01/15 | 0 | 70 | 7.2 | $Sc + GA_3$ |
| N. divaricata | G | Llanos de Challe, 04/16 | 0 | 48 | 10.1 | Sc + en |
| N. divaricata | G | Punta de Choros, 01/15 | 0 | 41 | 11.7 | Sc + GA ₃ |
| N. linearifolia | G | Antofagasta hills, 08/15 | 3 | 57 | 6.8 | $Sc + GA_3$ |
| N. onoana | G | Antofagasta hills, 08/15 | 20 | 72 | 9.0 | Sc + GA ₃ |
| N. sedifolia | G | Alto Patache, 12/15 | 0 | 87 | 15.9 | $Sc + GA_3$ |
| N. sedifolia | G | Los Molles, 01/15 | 0 | 79 | 6.3 | Sc + GA ₃ |
| N. sedifolia | G | Los Molles, 04/16 | 2 | 67 | 10.0 | Sc + en |
| N. intonsa | F | Alto Patache, 12/15 | 0 | 0 | | |
| N. carnosa | С | Llanos de Challe, 04/16 | 0 | 14 | 12.2 | $Sc + GA_3$ |
| N. rostrata | С | Llanos de Challe, 01/15 | 0 | 22 | 14.3 | Sc + GA ₃ |
| N. jaffuelii | В | Alto Patache, 12/15 | 0 | 31 | 18.8 | $Sc + GA_3$ |
| N. paradoxa | В | Los Molles, 04/16 | 2 | 64 | 6.2 | Sc + en |
| N. parviflora | В | Antofagasta hills, 08/15 | 0 | 30 | 5.1 | Sc + GA ₃ |

Data from two experiments; in Exp. 1, BGT corresponded to plug scarification and imbibition in gibberellic acid (Sc + GA₃), while in Exp. 2, the BGT corresponded to plug scarification and partial endosperm removal (Sc + en). Mean germination time (MGT) calculated from BGT data.

funicular plug, therefore, although it is necessary because it is the point from which the radicle emerges, is not as decisive as the removal of the layer immediately below; that is, the endosperm.

PD may be caused by structures that cover the embryo, including endosperm, seed coats and indehiscent fruit walls, among others (Baskin and Baskin, 2014). In seeds of Solanaceae species, the embryo is surrounded by two layers: the endosperm and the testa. The micropylar endosperm, covering the tip of the radicle, has been identified as a limiting factor for germination in Solanaceae species and its weakening seems to be a prerequisite for the protrusion of the radicle during germination (Groot and Karssen, 1987; Sánchez et al., 1990). While gibberellins promote weakening, the removal of the endosperm and testa layers opposite the radicle tip may also assist the process to permit radicle protrusion (Groot and Karssen, 1987; Finch-Savage and Leubner-Metzger, 2006). Both approaches had a positive effect in promoting the germination of scarified mericarps for most of the species included in our study (Tables 2 and 3), which confirms the importance of endosperm in *Nolana* seed dormancy.

In most of *Nolana* species included in this study, there was almost no germination when the funicular plug was not removed. This would indicate a very 'cautious' germination strategy, as



Fig. 4. Germination of *Nolana* species after the best germination treatment (plug scarification and imbibition in gibberellic acid or partial endosperm removal), grouped into clades (B,C,F,G). The number (*N*) of species included for each clade is noted. Data are expressed as the percentage of live seed and correspond to the average value for species in each clade \pm 95% confidence intervals.

suggested by Gutterman (1995), which would be in agreement with the fact that some of these species inhabit places where rainfall occurs every 15 years or more (Orellana et al., 2017; Pliscoff et al., 2017). It is possible to ascertain that the funicular plug not only plays an important role in the control of water uptake (Fig. 2) but also represents a physical barrier that the seed must overcome to complete its germination. How does this happen in nature? During the experiments, we noticed that in the case of several species, by keeping the mericarps moistened, it was later easier to remove the plugs. Douglas (2007) reported that dry storage for 2 years produced significantly more germination than fresh mericarps, so it can be speculated that dormancy is lost over time, and when prolonged rains arrive that 'loosen' the plugs, the radicle could be able to expand and eject the plug.

Germination of Nolana mericarps, expressed as a percentage of live seed, ranged from 0 to 27% in non-treated and from 0 to 100% when treated (Table 4), indicating varying levels of depth in their seed dormancy. In our study, it was not possible to ascribe a specific level or type of PD to each species (Baskin and Baskin, 2004) since the experiments focused not on determining this classification but on establishing comparisons between treatments. However, the relative levels of PD for each species should be related to the MGT or germination percentage after best treatment. MGT values were very variable within the same clade and species (Table 4), so we considered the germination percentage to be a better indicator. Therefore, we suggest a non-deep level of PD in the case of some species which showed germination after imbibition in distilled water, such as N. onoana and N. crassulifolia; others with a more intermediate level of PD (N. divaricata, N. linearifolia, N. sedifolia, N. jaffuelii, N. parviflora), which showed germination after scarification and the addition of gibberellic acid or endosperm remotion; and finally, some that hardly germinate with any of the treatments, which would correspond to a deeper dormancy (N. carnosa, N. intonsa, N. rostrata). Further experiments, considering varying periods of cold stratification and evaluation of germination at different temperatures, need to be performed in order to establish a



Fig. 5. Pearson correlation coefficient (*r*) and *P*-value (*P*) between germination percentage of 19 accessions of 12 species of *Nolana* for the best treatment (plug scarification and imbibition in gibberellic acid or partial endosperm removal) and latitudinal distribution, expressed as the difference between the northernmost and the southernmost latitudinal distribution point (decimal degrees).

more accurate classification of PD levels or types (Baskin and Baskin, 2004).

Relationship between dormancy and latitudinal distribution or phylogeny of species

Similar traits and life cycles are expected for species within a genus, and different germination requirements of species may reflect specific adaptations to the occupied habitat and geographic distributions (Van Assche et al., 2002; Barreto et al., 2016). However, our results showed a non-significant correlation (P =0.06) between germination and latitudinal distribution of the 12 Nolana species studied (Fig. 5). The species that presented higher germination percentages were N. crassulifolia (clade G), a prostrate shrub (Johnston, 1936) with a distribution between 26° and 34°S; and N. sedifolia (clade G), a perennial shrub or subshrub (Mesa, 1981) found from 20° to 33°S. Both species share at least one type of habitat (coastal rocks), although one has a very wide distribution (N. sedifolia) and the other, intermediate. N. onoana (clade G), however, which also presented a high germination percentage, is only found at a few locations in Antofagasta region. A particular case is N. paradoxa (clade B), with the most extensive distribution from the centre to the south of coastal Chile (30° to 43°S), where the climate is temperate humid; N. paradoxa had high germination percentages, but not higher than the other species mentioned. While other studies have also found no evidence of a correlation between dormancy and geographic distribution (Giorni et al., 2018; Dayrell et al., 2016), for Nolana, it remains to be studied if species that occupy a greater diversity of ecological niches present a greater plasticity in their dormancy levels.

In a study of several species from *campo rupestre* grasslands in Brazil, Dayrell et al. (2016) found no significant correlations between seed dormancy and life-history traits, such as growth habit; in our study, we also did not find such a correlation. Clades B and C, which showed similar germination results, are quite different in terms of growth form, and their mericarps are also highly dissimilar (Hepp, 2019). Clade B includes two subclades: rosette-forming, taprooted plants with large flowers, and erect annuals with slightly smaller flowers (Dillon et al., 2009). Clade C, on the other hand, are all large-flowered shrubs (Dillon et al., 2009). A higher level of dormancy would be expected in species with annual habits, since they depend more on seedling survival and/or formation of a seed bank for persistence (Dayrell et al., 2016; Venable, 2007), which was partly the case of *N. paradoxa*; but as mentioned before, its germination percentage was not higher than those of most clade G species. Clade G, for its part, which was found to have the higher germination percentage (Table 4, Fig. 4), is composed of small to moderate shrubs and annuals, all with highly reduced corollas (Dillon et al., 2009).

In conclusion, our results indicate that seeds of the studied species present PD with varying levels of depth. The role of the funicular plug and endosperm in dormancy regulation is emphasised. For the *Nolana* species studied, the results show that similarities in dormancy levels would be more related to phylogenetic proximity than to their latitudinal distribution. Further research should address the mechanism by which the funicular plug is removed under natural conditions, as well as a more accurate classification of PD level for each *Nolana* species.

Acknowledgements. All species were collected with permission and support of the National Forest Commission (CONAF Pan de Azúcar, CONAF Llanos de Challe), the Agricultural Research Institute (INIA) and owners/administrators of private parks (Punta de Choros, Puquén Los Molles) and research stations (Alto Patache Fog Oasis, managed by Universidad Católica de Chile). CONICYT Scholarship (21130176) to Josefina Hepp made this study possible.

References

- Barreto LC, Santos FMG and Garcia QS (2016) Seed dormancy in *Stachytarpheta* species (Verbenaceae) from high-altitude sites in southeastern Brazil. *Flora* 225, 37–44.
- Baskin JM and Baskin CC (2004) A classification system for seed dormancy. Seed Science Research 14, 1–16.
- Baskin CC and Baskin JM (2014) Seeds: ecology, biogeography, and evolution of dormancy and germination (2nd edn). San Diego, CA, USA, Elsevier.
- Cabrera E, Hepp J, Gómez M and Contreras S (2015) Seed dormancy of Nolana jaffuelii I.M.johnst. (Solanaceae) in the coastal Atacama desert. Flora 214, 17–23.
- Carta A, Hanson S and Müller JV (2016) Plant regeneration from seeds responds to phylogenetic relatedness and local adaptation in Mediterranean *Romulea* (Iridaceae) species. *Ecology and Evolution* 6, 4166–4178.
- Dayrell RLC, Garcia QS, Negreiros D, Baskin CC, Baskin JM and Silveira FAO (2016) Phylogeny strongly drives seed dormancy and quality in a climatically buffered hotspot for plant endemism. *Annals of Botany* 119, 267–277.
- Dillon MO, Tu T, Soejima A, Yi T, Nie Z, Tye A and Wen J (2007) Phylogeny of Nolana (Nolaneae, Solanoideae, Solanaceae) as inferred from granuleboundstarch synthase I (GBSSI) sequences. Taxon 56, 1000–1011.
- Dillon MO, Tu T, Xie L, Quipuscoa Silvestre V and Wen J (2009) Biogeographic diversification in *Nolana* (Solanaceae), a ubiquitous member of the Atacama and Peruvian deserts along the western coast of South America. *Journal of Systematics and Evolution* 47, 457–476.
- Dillon MO (2005) Solanaceae of the lomas formations of coastal Peru and Chile, pp. 131–155 *in* Hollowell V; Keating T; Lewis W; Croat T (Eds) A *festschrift for William G. D'Arcy: the legacy of a taxonomist*, Monographs in Systematic Botany from the Missouri Botanical Garden. St. Louis, Missouri.
- Douglas A and Freyre R (2006) Determination of seed germination requirements in Nolana sp. HortScience 41, 1002. Oral session abstracts. 103rd annual international conference of the American Society for Horticultural Science, New Orleans, Louisiana.
- Douglas A (2007) Sexual compatibility and seed germination in Nolana species. Master of science thesis, available at https://scholars.unh.edu/thesis/295/
- **Eberhart A and Tielbörger K** (2012) Maternal fecundity does not affect offspring germination – an empirical test of the sibling competition hypothesis. *Journal of Arid Environments* **76**, 23–29.

- Egaña I, Cereceda P, Pinto R, Larraín H, Osses P and Farías M (2004) Estudio biogeográfico de la comunidad arbustiva del farellón costero de punta patache, iquique, Chile. *Revista de Geografía Norte Grande* 31, 99–113.
- Enciclopedia de la Flora Chilena (2014) Encyclopedia of Chilean flora, available at www.florachilena.cl.
- Ellenberg H (1959) Über den Wasserhaushalt tropischer Nebeloasen in der Küstenwüste Perus. Bericht über das Geobotanisches Forschungsinstitut Rubel in Zürich 1958, 47–74.
- Ellis RH and Roberts EH (1980) Towards a rational basis for testing seed quality, pp. 605–635 *in* Hebblethwaite PD (Ed) *Seed production*, London, Butterworth.
- Facelli JM, Chesson P and Barnes N (2005) Differences in seed biology of annual plants in arid lands: a key ingredient of the storage effect. *Ecology* 86, 2998–3006.
- Fenner M (1985) Seed ecology. New York, NY, Chapman and Hall.
- Fick SE and Hijmans RJ (2017) Worldclim 2: new 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology* 37, 4302–4315.
- Figueroa JA, León-Lobos P, Cavieres LA and Pritchard H (2004) Ecofisiología de semillas en ambientes contrastantes de Chile: Un gradiente desde ecosistemas desérticos a templado-húmedos Fisiología ecológica y evolutiva de plantas, Valparaíso, Ediciones Universidad Católica de Valparaíso, 81–98.
- Finch-Savage WE and Leubner-Metzger G (2006) Seed dormancy and the control of germination. *New Phytologist* 171, 501–523.
- Flora del Conosur (2014) Vascular plants catalog, available at http://www2. darwin.edu.ar/Proyectos/FloraArgentina/FA.asp.
- Freyre R, Douglas A and Dillon MO (2005) Artificial hybridizations in five species of Chilean Nolana (Solanaceae). HortScience 40, 532–536.
- Giorni VT, Bicalho EM and Garcia QS (2018) Seed germination of *Xyris* spp. From Brazilian campo rupestre is not associated to geographic distribution and microhabitat. *Flora* 238, 102–109.
- Groot SPC and Karssen CM (1987) Gibberellins regulate seed germination in tomato by endosperm weakening: a study with gibberellin-deficient mutants. *Planta* 171, 525–531.
- Gutterman Y (1995) Seed dispersal, germination, and flowering strategies of desert plants. *Encyclopedia of Environmental Biology* **3**, 295–316.
- Gutterman Y (2002) Survival strategies of annual desert plants. Berlin/ Heidelberg, Springer-Verlag.
- Hepp J (2019) Characterization of seed dormancy of Nolana (Solanaceae) in the coastal Atacama desert of Chile. Doctoral thesis, available at https://repositorio.uc.cl/handle/11534/26359
- Johnston IM (1936) A study of the Nolanaceae. Proceedings of the American Academy of Arts and Sciences 71, 85–87.
- Jurado E and Flores J (2005) Is seed dormancy under environmental control or bound to plant traits? *Journal of Vegetation Science* 16, 559–564.
- Knapp S (2002) Tobacco to tomatoes: a phylogenetic perspective on fruit diversity in the Solanaceae. Journal of Experimental Botany 53, 2001–2022.
- León-Lobos P, Way M, Pritchad H, Moreira-Muñoz A, León M and Casado F (2003) Conservación *ex situ* de la flora de Chile en banco de semillas. Chloris Chilensis, Año 6, N° 1. http://www.chlorischile.cl.
- León-Lobos P, Way M, Aranda PD and Lima-Junior M (2012) The role of *ex situ* seed banks in the conservation of plant diversity and in ecological restoration in Latin America. *Plant Ecology & Diversity* 5, 245–258.
- León-Lobos P, Bustamante-Sánchez MA, Nelson CR, Alarcon D, Hasbún R, Way M, Pritchard HW and Armesto JJ (2020) Lack of adequate seed supply is a major bottleneck for effective ecosystem restoration in Chile: friendly amendment to Bannister et al. (2018). *Restoration Ecology* 28, 277–281.
- **Mesa A** (1981) *Monographie des Nolanacées*. Thése présentée devant L 'Université de Rennes I pour obtener le Titre de Doctoeur.
- Meyer SE, Kitchen SG and Carlson SL (1995) Seed germination timing patterns in intermountain *penstemon* (Scrophulariaceae). American Journal of Botany 82, 377–389.
- Muñoz-Schick M, Pinto R, Mesa A and Moreira-Muñoz A (2001) "Oasis de neblina" en los cerros costeros del sur de iquique, región de tarapacá, Chile, durante el evento El niño 1997-1998. *Revista Chilena de Historia Natural* 74, 389–405.

- Orellana H, García JL, Ramírez C and Zanetta N (2017) El aluvión del 9 de agosto 2015 en alto patache, región de tarapacá, desierto de Atacama. *Revista de Geografía Norte Grande* **68**, 65–89.
- Pinto R and Luebert F (2009) Datos sobre la flora vascular del desierto costero de arica y tarapacá, Chile, y sus relaciones fitogeográficas con el sur de perú. *Gayana Botánica* 66, 28–49.
- Pinto R, Larraín H, Cereceda P, Lázaro P, Osses P and Schemenauer RS (2001) Monitoring fog-vegetation communities at a fog-site in alto patache, south of iquique, northern Chile, during "El niño" and "La niña" events (1997-2000). 2nd international conference on Fog and Fog collection, Newfoundland, Canadá, St. John's, 293–296
- Pliscoff P, Zanetta N, Hepp J and Machuca J (2017) Efectos sobre la flora y vegetación del evento de precipitación extremo de agosto 2015 en alto patache, desierto de Atacama, Chile. *Revista de Geografía Norte Grande* 68, 91–103.
- Riedemann P, Aldunate G and Teillier S (2006) Flora nativa de valor ornamental: zona norte. Santiago, Chile, Ediciones Chagual.
- Rundel PW, Dillon MO, Palma B, Mooney HA, Gulmon SL and Ehleringer JR (1991) The phytogeography and ecology of the coastal Atacama and Peruvian deserts. *Aliso* 13, 1–49.
- Rundel PW, Palma B, Dillon MO, Sharifi MR and Boonpragob K (1997) *Tillandsia landbeckii* in the coastal Atacama desert of northern Chile. *Revista Chilena de Historia Natural* **70**, 341–349.

- Sánchez RA, Sunell L, Labavitch JM and Bonner BA (1990) Changes in the endosperm cell walls of two *Datura* species before radicle protrusion. *Plant Physiology* **93**, 89–97.
- Santibañez F, Santibañez P, Caroca Cy and González P (2017) Atlas Agroclimático de Chile – Tomo I: Regiones de Arica y Parinacota, Tarapacá y Antofagasta. Centro AGRIMED, Facultad de Ciencias Agronómicas, Universidad de Chile, Santiago de Chile.
- Saunders E (1936) On certain unique features of the gynoecium in Nolanaceae. New Phytologist 35, 423–431.
- Schulz N, Aceituno P and Richter M (2011) Phytogeographic divisions, climate change and plant dieback along the coastal desert of northern Chile. *Erdkunde* 65, 169–187.
- Tago-Nakazawa M and Dillon MO (1999) Biogeografia y evolución en el clado Nolana (Nolaneae – Solanaceae). Arnaldoa 6, 81–116.
- Tu T, Dillon MO, Sun H and Wen J (2008) Phylogeny of Nolana (Solanaceae) of the Atacama and Peruvian deserts inferred from sequences of four plastid markers and the nuclear LEAFY second intron. *Molecular Phylogenetics and Evolution* 49, 561–573.
- Van Assche J, Van Nerum D and Darius P (2002) The comparative germination ecology of nine *Rumex* species. *Plant Ecology* 159, 131–142.
- Venable L (2007) Bet hedging in a guild of desert annuals. Ecology 88, 1086– 1090.