

RADIOCARBON ECOLOGY OF THE LAND SNAIL *HELIX MELANOSTOMA* IN NORTHEASTERN LIBYA

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ABSTRACT. Terrestrial gastropods are problematical for radiocarbon (¹⁴C) measurement because they tend to incorporate carbon from ancient sources as a result of their dietary behavior. The ¹⁴C ecology of the pulmonate land snail, *Helix melanostoma* in Cyrenaica, northeastern Libya, was investigated as part of a wider study on the potential of using terrestrial mollusk shell for ¹⁴C dating of archaeological deposits. *H. melanostoma* was selected out of the species available in the region as it has the most predictable ¹⁴C ecology and also had a ubiquitous presence within the local archaeology. The ecological observations indicate that *H. melanostoma* has a very homogenous ¹⁴C ecology with consistent variations in F¹⁴C across sample sites controlled by availability of dietary vegetation. The majority of dated specimens from non-urbanized sample locations have only a small old-carbon effect, weighted mean of 476 ± 48 ¹⁴C yr, with between ~1% and 9% of dietary F¹⁴C from non-organic carbonate sources. Observed instabilities in the ¹⁴C ecology can all be attributed to the results of intense human activity not present before the Roman Period. Therefore, *H. melanostoma* and species with similar ecological behavior are suitable for ¹⁴C dating of archaeological and geological deposits with the use of a suitable offset.

KEYWORDS: land snails, terrestrial mollusks, radiocarbon, ecology, North Africa, dating, isotopes.

INTRODUCTION

Land-snail shells are widely distributed in Quaternary deposits and frequently found on archaeological sites (Prendergast and Stevens 2014). Initial investigations (Goodfriend and Hood 1983; Goodfriend 1987; Pigati et al. 2010; Hill 2015; Hunt et al. 2015) suggest that in suitable circumstances it is amenable to radiocarbon (¹⁴C) dating, opening up the possibility of extending ¹⁴C chronologies into sedimentary facies containing no other source of biotic carbon. For instance, in calcareous oxidized sediments in semiarid and arid environments, land-snail shell is often well preserved but plant macrofossils and wood are not always available and bone collagen is often not preserved (e.g. Hunt et al. 1992; Anketell et al. 1995; Hunt and Garrard 2013). Land-snail shells may offer the only possibility for ¹⁴C dating such sites.

Previous studies (e.g. Goodfriend 1987; Romanellio et al. 2008; Pigati et al. 2010; Hill 2015) have shown that the ecological pathways and uptake mechanisms undergone by ¹⁴C as it is incorporated into the shell of land snails are not straightforward. Some species are detritivores and others graze on the algae encrusting carbonate rocks. In both, old carbon may be consumed and incorporated into shell, causing a well-documented impact on ¹⁴C values, and the shells to have ¹⁴C age values significantly older than their actual age. It is also possible, although thus far unproven, that some taxa may fractionate carbon in their metabolic processes (McConaughy and Gillikin 2008). An understanding of the ¹⁴C ecology at a generic, if not specific, level is therefore required before a species can be used for dating.

This paper presents the ¹⁴C ecology of the Helicid land snail *Helix melanostoma* (Draparnaud 1801) in northeastern Libya. The work was undertaken to provide a baseline for ¹⁴C dating of this species in Quaternary archaeological deposits throughout the Mediterranean and in particular in the Haua Fteah archaeological site lying approximately 1 km south of the

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Mediterranean coast in northeastern Libya, some 7.5 km east of Susa at 22°03'06''E, 32°54'01''N (Barker et al. 2007, 2008, 2009, 2010, 2012; Rabett et al. 2012; Farr et al. 2014).

Background: Radiocarbon Ecology of Land Mollusks

The mechanisms by which ^{14}C is incorporated into the shells of land snails are complex and interconnected (Figure 1). Three key pathways can be clearly defined by which carbon moves through the metabolism of a snail: respiration, diet, and hydration. Of these, diet accounts for the vast majority of carbon laid down in the shell (e.g. Stott 2002; McConnaughey and Gillikin 2008; Xu et al. 2011). Dietary carbon can be from living vegetation, which will be broadly in ^{14}C equilibrium with atmospheric levels. Dietary carbon is also likely to include ancient carbon from a variety of sources, as mentioned above. Similarly, hydration may include precipitation water condensed from the atmosphere, which will be in equilibrium with atmospheric ^{14}C values and also water that has been in contact with, and dissolved, ancient carbon; including atmospheric mineral dust, limestone, and dead organic matter in soils.

The role of $\delta^{13}\text{C}$ in interpreting land snail ecology has been widely explored. The relationship between this isotope and ^{14}C dating land snails has long been an integral element in assessing the impact of “old carbon.” More recent detailed investigations of the ^{14}C dating of land snails (e.g. Pigati et al. 2010) have begun to demonstrate that while $\delta^{13}\text{C}$ is important for fractionation correction in the measurement of ^{14}C values it is not particularly useful in determining “old carbon” impact (e.g. Romaniello et al. 2008; Xu et al. 2011).

Understanding the relationship between the three isotopes of carbon within the biochemistry of the land snail is integral to both understanding why $\delta^{13}\text{C}$ is not as important for identifying old carbon and also why a clear understanding of the life processes of the snail in acquiring carbon are vital to this point (Figure 1). Research has shown that atmospheric sources of carbon tend to have proportionally less impact on the processes involved in shell formation in land snails in comparison to dietary sources (e.g. McConnaughey and Gillikin 2009 and references therein). Thus, it has been argued, based on the biological principles underlying the carbon cycle in mollusks, that CO_2 derived from respiration does not, in normal circumstances, have a significant impact on the $^{14}\text{C}/^{12}\text{C}$ ratio of landsnails (McConnaughey and Gillikin 2009).

The influence of dissolved inorganic carbonate (DIC) in water is understood only for a very few species, e.g. Succinidae, which are generally atypical of land snails (Pigati et al. 2004, 2010). Inference based on biochemistry of similar organisms does suggest that DIC should not in normal circumstances account for a significant portion of land-snail shell carbonate (McConnaughey and Gillikin 2009).

The ingestion of old carbon from various sources has been the subject of a limited number of studies (e.g. De Jorge and Haeser 1968; Tamers 1970; Magaritz et al. 1981; Goodfriend and Hood 1983; Goodfriend 1987; Goodfriend and Ellis 2002; Stott 2002; Balakrishnan and Yapp 2004; Pigati et al. 2004; Baldini et al. 2007; Yanes et al. 2008; Romaniello et al. 2008; Pigati et al. 2010; Xu et al. 2010, 2011; Rakovan et al. 2013). Review of these papers demonstrates that there is little replication or overlap in species analyzed or geographical regions sampled. Only in North America is there any consequential overlap; for Europe, North Africa and elsewhere there is no significant replication. A number of studies examining the use of terrestrial shell carbonate for ^{14}C dating have also indicated, using ^{14}C analysis, that old carbon can account for as much as 30–40% of carbonate in a shell (e.g. Goodfriend and Stipp 1983; Goodfriend and Ellis 2002; Pigati et al. 2004, 2010). However, the quantities ingested appear to hinge on a wide variety of influencing factors in local environments and are taxon-dependent.

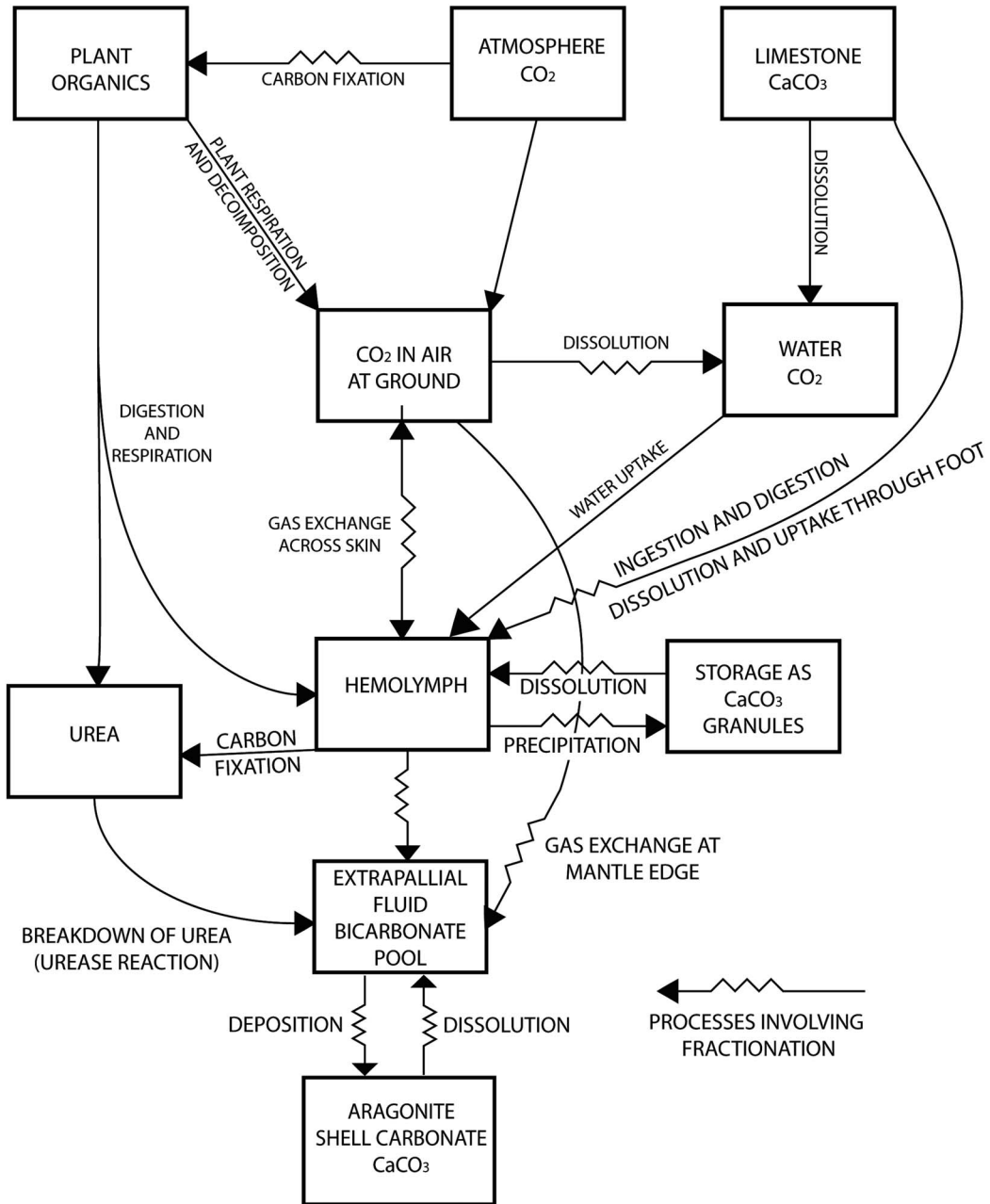


Figure 1 Model for the radiocarbon ecology of land snails after Goodfriend (1987) with modifications based on Stott (2002) and Pigati et al. (2004, 2010).

The mass balance approach pioneered by Goodfriend and Hood (1983) for modeling the isotopic fractions of carbon in terrestrial land-snail shells has been heavily utilized. It has been found to be problematic because it is not possible to separate the sources of ¹⁴C (Figure 1) to the degree required to sufficiently isolate the component parts of the diet for the mass balance equation to be properly calculated (e.g. Romanellio et al. 2008; Pigati et al. 2010; Xu et al. 2011;

Hill 2015). These issues further reinforce the need to build a clear understanding of the modern ecology, as well as the physiology of any species, before the ^{14}C ages can be defined with confidence. This is particularly the case for the current work in Cyrenaica because previous studies of ^{14}C ecology in the Mediterranean (e.g. Goodfriend 1987; Romanellio et al. 2008) have not examined terrestrial gastropods common in this region.

METHODS AND MATERIALS

Study Region

Sampling for modern snail shells and vegetation took place in northeast Libya (Figure 2). Here the Gebel Akhdar massif rises to over 800 m in three great escarpments. The region is an inverted Mesozoic-Tertiary basin, geologically characterized by thick limestones and thin calcareous mudstones (El Hawat and Abdelsamad 2004). Rainfall on the Gebel Akhdar is significantly higher than elsewhere along the northeast African littoral (Sen and Eljadid 1999) and the region is a “habitat island” of Mediterranean vegetation among the generally arid lowlands to the west, south and east. The region bordering the coastal plain with richer soils and less limestone outcropping has noticeably more lush vegetation cover compared with the rising slopes of the Gebel Akhdar, which are heavily grazed. The slope of the first escarpment of the

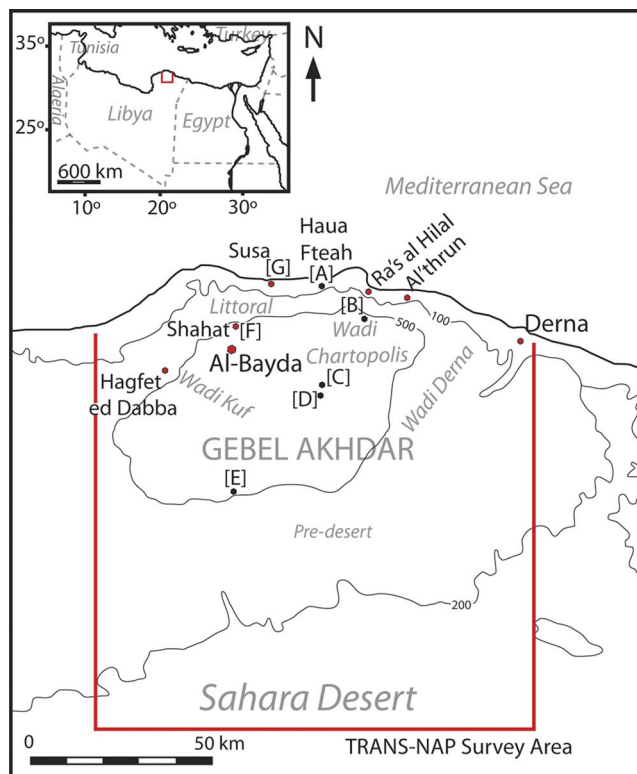


Figure 2 Sample locations in Cyrenaica for this study. [A] = Haul Fteah (HF_Eco), [B] = Wadi Chartopolis (MH12-6), [C] = Gebel Akhdar 1 (MH12-9), [D] = Gebel Akhdar 2 (MH12-10), [E] = Gebel Akhdar 3 (MH12-11), [F] = Shahat project Resthouse (RH12), and [G] = Apollonia (AP12).

Gebel Akhdar is dominated by juniper scrub with large limestone outcroppings and shallow soils. The higher regions of the Gebel were originally covered in Cupressus-Oak forests but are now largely cleared for agriculture. The southern slopes are in rain shadow and largely covered with steppe, grading southward into true desert (Barker 2007; Hegazy et al. 2011).

Ecological Patterning of *H. melanostoma*

Very little information is available on the ecology and distribution of land snails in the Gebel Akhdar. The most informative publication (Brandt 1959), although primarily taxonomic, provides some very short ecological descriptions of the family Helicidae. All Helicidae in the region are xerophytic to varying degrees and the species are found in habitats ranging from Mediterranean woodland to semi-desert.

In the study area, our surveys and conversations with local specialists suggest, that *H. melanostoma* spends most of the year aestivating in shallow, damp shaded soils beneath scrub or small trees such as *Ceratonia siliqua*, *Pistacia lentiscus*, and *Rhus tripartita* in areas of limestone pavement, scrub on slopes with shallow soils, and scrub-forest on deep soils (Hill 2010). It is inferred that low-lying, mostly evergreen, vegetation forms the local primary food source of *H. melanostoma*. These findings were further reinforced by the stable isotope study (Prendergast et al. 2014, 2015) which showed that $\delta^{13}\text{C}$ in *H. melanostoma* shell carbonate was primarily a function of the animals' diet, which is the local available vegetation. Therefore, changes in shell $\delta^{13}\text{C}$ can be used to detect changes in vegetation composition, particularly in relation to the abundance of C3 and C4 vegetation (Prendergast et al. 2014). Our environmental observations concerning this species broadly support ecological findings from southern France (e.g. Kerney et al. 1983; Pflieger 1984) for *H. melanostoma*.

Sample Sites and Methodology

An initial ^{14}C evaluation of a number of surface collected mollusk shells, presumed to be recently dead, carried out in 2010, of the four archaeologically important land snail species known to be abundant in the Haua Fteah archaeological site is shown in Table 1. This evaluation suggests that *Helix melanostoma* has less of an age offset, and less variability than *Trochoidea cretica* or *Sphincterochila* spp., the other ubiquitous species present both in the modern landscape and in the archaeological assemblages and sediments in the Haua Fteah. *Rumina* cf. *decollata* was ruled out based on ecological knowledge of diet and because no modern living specimens could be found in follow-up sampling. Therefore, based on its

Table 1 Results of pilot study evaluating F^{14}C variation of land snails in northeastern Libya at site CPP07 1587. ^{14}C ages are not rounded here to avoid introduction of errors in calculations.

Species	UBA nr	F^{14}C	\pm	^{14}C age (yr)	\pm
<i>H. melanostoma</i>	15321	0.274	0.0012	10,395	36
<i>H. melanostoma</i>	15322	1.0246	0.0034	Modern	
<i>H. melanostoma</i>	15323	1.1279	0.0035	Modern	
<i>Rumina</i> cf. <i>decollata</i>	15324	0.8904	0.0029	932	26
<i>Rumina</i> cf. <i>decollata</i>	15325	0.7912	0.0026	1882	27
<i>Rumina</i> cf. <i>decollata</i>	15326	0.8709	0.0029	1110	26
<i>Trochoidea cretica</i>	15327	0.91	0.0029	758	26
<i>Trochoidea cretica</i>	15329	0.7857	0.0026	1938	27
<i>Sphincterochila</i> spp.	15333	0.9923	0.0044	62	35
<i>Sphincterochila</i> spp.	15334	0.8151	0.0037	1642	37
<i>Sphincterochila</i> spp.	15335	0.8866	0.004	967	36

ubiquitous presence in archaeological sediments, and modern ages in this study *H. melanostoma* was selected for intensive work. The outlying sample, UBA 15321, appears to be archaeological material that was present on the landscape at the point of collection, based on comparisons of ages of material from within the Haua Sequence (Hill, 2015) reinforcing the importance of live-collected specimens for studies like this.

The sampling of live collected modern specimens was carried out during 2010 and 2012 field seasons (Hill 2010; Barker et al. 2012; Rabett et al. 2012; Prendergast et al. 2015). The sampling strategy was designed to encompass the widest possible range of modern environments within the study area but focused on five key locations given the constraints of time and security (Figure 2; Table 2). Site sample identifications for sites have been shortened in all tables and figures as follows: HF12_Eco refers to sites around the Haua Fteah; AP12 to Apollonia; RH12 to the Project Resthouse, Shahat; MH12 to Gebel Akhdar massif sample sites.

At these localities transect sampling found live aestivated specimens in leaf litter and soil beneath vegetation and active mollusks on vegetation after very rare rainfall events. Ethnographic observations suggest that all of these sites are extremely heavily grazed by domesticated animals, mostly goats and sheep and vegetation levels are consequently lower than would be expected in habitats grazed by native animals. It is also likely that a number of sites were impacted by the relatively intense anthropogenic activity on the Gebel, near Shahat, Susa and Albayda including mass building work, cement works and atmospheric pollution.

A total of 37 *H. melanostoma* were selected for ^{14}C analysis with a minimum of three specimens analyzed from each sample location. Vegetation and soil samples were gathered from every sample site where live mollusks were collected. Where possible, samples of vegetation were gathered in association with live mollusks. In selecting samples for atmospheric and dietary ^{14}C values species favored by *H. melanostoma* were selected in accordance with existing ecological knowledge of the species (e.g. Brandt 1959) and observations made in the field. Where such species (Table 2) were unavailable, dominant vegetation was sampled. Once collected, mollusks were killed by freezing. Freeze-dried bodies were separated from the shell using forceps.

Radiocarbon

Whole shells were carefully broken apart using forceps or a Dremel tool with a circular carborundum cutting head. An aperture sample was obtained from every adult *H. melanostoma* that was dated (Figure 3). In small specimens with less than one full season of growth the entire shell was sampled and considered as an apex because the aperture had not thickened and so was too insubstantial for effective analysis.

Many previous radiocarbon studies, which were concerned primarily with predicting the “limestone effect,” did not assess lifetime changes in apparent ^{14}C age (e.g. Quarta et al. 2007; Romanellio et al. 2008; Pigati et al. 2010; Xu et al. 2011). Investigations of parts of the shell were carried out to evaluate changes in apparent ^{14}C age over the lifetime of the individual snail (Figure 3). At least two paired aperture/apex dates were obtained from adult specimens for every location sampled except for the environment immediately outside the Haua Fteah where all the apices were used in a related $\delta^{13}\text{C}$ shell study (Prendergast 2013; Prendergast et al. 2015), looking at the suitability of *H. melanostoma* for climatic reconstruction. In some instances, whorl segments were also selected to provide further detail.

Vegetation samples for ^{14}C analysis were rinsed in distilled water, dried, weighed into a precombusted quartz tube with an excess of copper oxide (CuO), sealed under vacuum and

Table 2 Details of sample sites for modern sampling for mollusk and vegetation.*

Site	Location	Site code	Elevation (m)	Dominant vegetation	Est. total vegetation (%)	Est. arboreal (%)	Sampled vegetation taxon	<i>Helix melanostoma</i>	<i>Sphincterochila</i> spp.	<i>Trochoidea cretica</i> (.agg)	Date
Haua Fteah	32°54.017, 22°3.011	HF Eco02	75	Scrub with some herbaceous vegetation: PL, CS, P	70	50	<i>Ceratonia siliqua</i>	5	0	3	April 2010
		HF Eco03	70	Dense scrub with herbaceous vegetation: PL, PH, JO, RT, CS, P	75	60	<i>Juniperus oxycedrus</i>	3	0	2	April 2010
		HF Eco04	66	Scrub woodland on limestone: CS, JO	90	70	<i>Juniperus oxycedrus</i>	1	2	3	April 2010
		HF Eco05	50	Scrub woodland on limestone with light herbaceous vegetation: JO, RT, CS, P, PH	80	60	<i>Juniperus oxycedrus</i>	2	1	4	April 2010
		HF Eco06	49	Scrub woodland with herbaceous vegetation: CS, JO, RT, SS, PL, P, PH	90	60	<i>Ceratonia siliqua</i>	8	2	9	April 2010
		AP12	5	Sparse coastal scrubland: PL, AS	20	5	<i>Artemisia sp.</i>	5	4	10	May 2012
Apollonia (Susa)	32°54.176, 21°58.524	AP12	5	Sparse coastal scrubland: PL, AS	20	5	<i>Artemisia sp.</i>	5	4	10	May 2012
Resthouse (Shahat)	32°49.579, 21°51.730	RH12	590	Urbanized woodland:CS	95	30	<i>Ceratonia siliqua</i>	4	0	12	May 2012
Wadi Chartopolis	32°50.779, 22°09.619	ML12-6	508	Dense Mediterranean woodland: AS, P, JO, ZS, PL	75	45	<i>Pistacia lentiscus</i>	3	1	7	May 2012
Gebel Akhdar	32°40.038, 21°51.789	ML12-9	798	Grassy steppe scrub: SS, PH, ZS	70	40	<i>Zizyphus</i>	3	1	6	May 2012
		ML12-10	853	Steppe scrub: SS, PH, ZS	70	35	<i>Zizyphus</i>	3	2	5	May 2012
		ML12-11	710	Steppe scrub: AS, PH, ZS	50	5	Poaceae	0	0	3	May 2012

*PL = *Pistacia lentiscus*; PH = *Phlomis sp.*; JO = *Juniperus oxycedrus*; RT = *Rhus tripartita*; CS = *Ceratonia silaqua*; SS = *Sarcopoterium spinosum*; AS = *Artemisia sp.*; P = *Poaceae*; Z = *Zizyphus sp.*

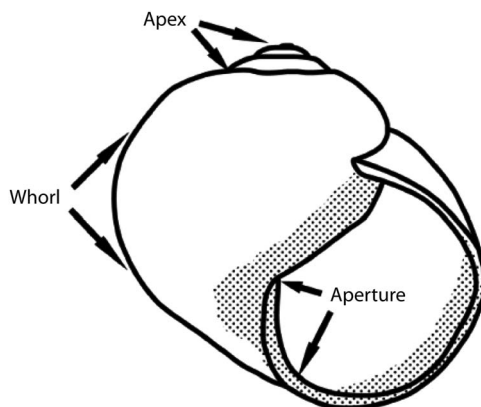


Figure 3 Diagram of *H. melanostoma* showing parts selected for sampling. The stippling shows the dark brown areas on the shell after which the species is named.

combusted to carbon dioxide (CO_2) and converted to graphite using the hydrogen reduction method (Vogel et al. 1987). Shell samples were pretreated using 1% HCl to remove any surface contamination and then rinsed in distilled water, dried and weighed. The samples were then placed in to septa seal containers and converted into CO_2 using 80% phosphoric acid. The resulting CO_2 was converted to graphite on an iron catalyst using the zinc reduction method (Slota et al. 1987). The $^{14}\text{C}/^{12}\text{C}$ and $^{13}\text{C}/^{12}\text{C}$ ratios were measured by accelerator mass spectrometry (AMS) at the 14CHRONO Centre, Queen's University Belfast. The $^{14}\text{C}/^{12}\text{C}$ ratio of the vegetation and shell samples were background corrected using measurements of anthracite and Icelandic Spar Calcite, respectively, and normalized to the HOXII standard (SRM 4990C; National Institute of Standards and Technology). The ^{14}C ages were corrected for isotope fractionation using the AMS measured $^{13}\text{C}/^{12}\text{C}$, which accounts for both natural and machine fractionation. The ^{14}C age and one standard deviation were calculated using the Libby half-life of 5568 yr following the methods of Stuiver and Polach (1977). ^{14}C data for modern samples are presented as $F^{14}\text{C}$ values (Reimer et al. 2004). The 1σ uncertainties are the maximum of the measurement statistics and the variance of seven 2-min runs for each sample. Throughout this paper, ^{14}C ages reported in tables are not rounded as this may introduce errors in calculations (Millard 2014).

Stable Isotopes

Stable carbon analysis of vegetation and soil was carried out at the ^{14}C CHRONO Centre at Queen's University Belfast. Stable isotopes ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$), % carbon, and % nitrogen from vegetation and soil samples were measured on a Thermo Fisher Delta V Advantage with elemental analyzer together with Iso-Analytical Laboratory Standard IA-R041 L-Alanine ($\delta^{15}\text{N}_{\text{AIR}} = 5.56 \text{ ‰}$; $\delta^{13}\text{C}_{\text{VPDB}} = 23.33 \text{ ‰}$) which was measured 11–13 times throughout the runs, bracketing between 6 and 8 samples.

Modeling and Data Analysis

Data analysis was carried out using Microsoft Excel 2013 and SPSS version 19. Simple and exponential linear regression and chi square (χ^2) were used to investigate relationships between variables. Weighted averages of multiple samples and chi square tests were calculated using a VBA coded macro in Excel based on Bevington (1969). The basic relationships between ^{14}C

results from different sites were evaluated using chi-squared ($\chi^2_{i,0.5}$) analysis and analysis of correlations and sample scatter (standard deviation).

$F^{14}C$ values for shells, where apertures, apices, and whorl fragments were measured, and where required, combined for certain analyses of the ^{14}C ecology of *H. melanostoma*. This was done by calculating the weighted averages of the $F^{14}C$ values to simulate analysis of the whole shell (Table 6). Because the aperture and apex $F^{14}C$ values should represent maximum range of $F^{14}C$ values in a land snail across its growth cycles, the resultant average $F^{14}C$ and its aggregated error should represent a reasonable average $F^{14}C$ value for a sample across its whole life span. This was further tested by the measurement of a number of whorl fragments to determine intermediate values.

Predicted $\delta^{13}C$ values were calculated using the method set out in Stott (2002). The Stott equation is based on a linear regression between snail body $\delta^{13}C$ and snail shell $\delta^{13}C$. The values used in this study were first calculated in Prendergast (2013) and snail body $\delta^{13}C$ and shell $\delta^{13}C$ for samples comes from Prendergast et al. (2014). While the equation was originally created for *H. aspersa* rather than *H. melanostoma*, the two species occupy similar niches in different areas around the Mediterranean and ecological observations suggest that *H. melanostoma* is locally being out-competed by introduced *H. aspersa*, particularly in urban areas. It is important to note that there may be further species-specific offsets that may impact detailed comparison with such values.

Calculation of the difference between $F^{14}C$ of mollusk shells and $F^{14}C$ of preferred vegetation diet for *H. melanostoma*, Equation (1), was used to model the potential impact of old carbon on individual shells (e.g. Pigati et al. 2010)

$$F^{14}C_{\text{Shell Average}} - F^{14}C_{\text{diet}} = F^{14}C \text{ from nondietary sources} \quad (1)$$

$F^{14}C_{\text{Shell Average}}$ represents the average $F^{14}C$ value across the lifetime of the shell (Table 6). $F^{14}C_{\text{diet}}$ is derived using $F^{14}C$ atmospheric values for vegetation consumed by *H. melanostoma* gathered from sample sites (Table 5). In instances where more than one value exists for $F^{14}C_{\text{atmosphere}}$ from a site, a weighted average was used for the value of $F^{14}C_{\text{diet}}$.

This method follows the understanding that the total quantity of $F^{14}C$ present in a mollusk shell can only derive from three overall sources, dietary carbon, respiration and hydration. Of these, only dietary carbon is likely to be significant in the vast majority of cases where snails pass most of their lives on or above the soil surface. Dietary carbon will have two components; “young,” derived from living tissue, and “old,” derived from dead tissue or mineral sources. If the $F^{14}C$ of the vegetation eaten by the snail can be established, then the remaining difference between the $F^{14}C$ of the shell and the $F^{14}C$ of the diet should represent the quantity that the shell’s $F^{14}C$ value has been reduced through the incorporation of older carbon, even if the exact source of carbon contributing to the difference cannot be fully known.

The age offset in ^{14}C years, the “old carbon effect” as discussed in this study, where used, was calculated using the following Equation (2)

$$\text{Shell reservoir offset} = -8033 * \ln \left(\frac{F^{14}C_{\text{shell}}}{F^{14}C_{\text{atm}}} \right) \quad (2)$$

This equation was defined after Keaveney and Reimer (2012), $F^{14}C_{\text{shell}} = F^{14}C$ value derived from the land-snail shell. The -8033 is derived from the Libby half-life of ^{14}C ($T_{1/2} = 5568$ yr).

Atmosphere values; $F^{14}C_{atm}$, in this study are based primarily on vegetation samples gathered at sample sites for live land snails in the year of collection (2010 and 2012) as the Northern Hemisphere Zone 2 dataset (Hua et al. 2013) is only current until late 2009 and Levin et al. (2013) does not have any data points with sufficient geographic proximity.

The uncertainty that is associated with each reservoir offset is calculated using the uncertainty associated with the ^{14}C measurement ($F^{14}C_{shell}$) from each modern shell sample and its associated uncertainty $F^{14}C_{sigma}$ using the Equation (3) below (Soulet et al. 2016).

$$\sigma = 8033 * \sqrt{\left(\frac{F^{14}C_{Shell\ Sigma}}{F^{14}C_{Shell\ measured}}\right)^2 + \left(\frac{F^{14}C_{atm\ sigma}}{F^{14}C_{atm\ measured}}\right)^2} \quad (3)$$

RESULTS

Fifty-nine ^{14}C analyses were made from 32 live *H. melanostoma* over the course of the field sampling exercises. The results of the $\delta^{13}C$ on vegetation (Table 3), ^{14}C of vegetation (Table 4), ^{14}C of shells (Table 5), and modeling of ^{14}C shell data and old carbon impact (Table 6) are presented here.

Biometrics

Biometrics were gathered for all *H. melanostoma* specimens sampled for ^{14}C dating. Mean basal diameter (MBD) height and growth checks were measured for all samples. The mean MBD for all specimens was 21.3 mm, Mean height was 18 mm and the average number of growth checks was 2.4. The largest MBD was 28.7 mm, whereas the smallest was 7.3 mm. Those with the smallest MBD and Height were all found in Apollonia (AP12).

Table 3 $\delta^{13}C$ of vegetation and soil samples from across Cyrenaica.

Sample site ID	Genus	Sample type	$\delta^{13}C_{VPDB}$ (‰)
1546*	<i>Quercus</i> sp.	Bulk vegetation	-27.6
1546*	<i>Quercus</i> sp.	Bulk vegetation	-27.6
HF12 Eco02	<i>Juniperus oxycedrus</i>	Bulk vegetation	-22.6
HF12 Eco02	<i>Juniperus oxycedrus</i>	Bulk vegetation	-22.7
HF12 Eco05	<i>Juniperus oxycedrus</i>	Bulk vegetation	-24.4
HF12 Eco05	<i>Juniperus oxycedrus</i>	Bulk vegetation	-24.4
HF12 Eco06	<i>Juniperus oxycedrus</i>	Bulk vegetation	-23.9
HF12 Eco06	<i>Juniperus oxycedrus</i>	Bulk vegetation	-23.9
HF12 Eco06	<i>Ceratonia siliqua</i>	Bulk vegetation	-30.0
HF12 Eco06	<i>Ceratonia siliqua</i>	Bulk vegetation	-30.0
AP12-1	<i>Artemisia</i> spp.	Bulk vegetation	-28.9
MH12-6	<i>Pistacia lentiscus</i>	Bulk vegetation	-27.1
MH12-6	<i>Sarcopoterium spinosum</i>	Bulk vegetation	-28.7
MH12-6	<i>Ziziphus</i> sp.	Bulk vegetation	-26.0
MH12-9	<i>Ziziphus</i> sp.	Bulk vegetation	-25.4
MH12-10	<i>Ziziphus</i> sp.	Bulk vegetation	-25.4
MH12-11	Triticeae	Bulk vegetation	-27.5
HF12 Eco02	N/A	Soil	-25.0
HF12 Eco02	N/A	Soil	-25.0
HF12 Eco06	N/A	Soil	-24.9
HF12 Eco06	N/A	Soil	-24.9

*Sample 1546 was gathered from outside the Haa Fteah.

Table 4 Radiocarbon results of vegetation samples taken from across Cyrenaica as a proxy for atmospheric ^{14}C and dietary F^{14}C for land snails. Average dietary F^{14}C of suggested mollusk diet is based on the per site values presented in this table.

Sample location	Genus	Alt. (m)	UBA	F^{14}C vegetation	\pm
<i>Haua Fteah</i>	<i>Ceratonia</i>	70	18083	1.0571	0.003
<i>Haua Fteah</i>	Leaf litter	70	18085	1.1079	0.0029
<i>Apollonia</i>	<i>Artemisia</i>	5	25904	1.0501	0.0031
<i>Wadi Chartopolis</i>	<i>Ceratonia</i>	508	25905	1.0337	0.0034
<i>Wadi Chartopolis</i>	<i>Pistacia</i>	508	25906	1.0373	0.0029
<i>Gebel Akhdar 1</i>	<i>Zizyphus</i>	798	25907	1.0381	0.0029
<i>Gebel Akhdar 2</i>	<i>Zizyphus</i>	853	25908	1.0314	0.0029
<i>Gebel Akhdar 3</i>	<i>Poacae</i>	710	25909	1.0463	0.0043

Within-Shell Radiocarbon Variation

There was a substantial but predictable “age” difference between apices and apertures (Figure 4), with apices being “older” than apertures. This is significant for ^{14}C dating as it suggests that juvenile specimens may have a higher propensity towards old carbon intake than adult specimens. It is also important for archaeologists selecting dating samples because apices and aperture fragments are easily identifiable and preserve well. Whorl fragments, where analyzed, were found to more closely reflect values of apertures (Table 5) rather than more broadly intermediate ages. In ecological terms, it is likely that young snails require calcium (Ca) to start building a shell, obtaining it from small limestone particles in the soil, from outcrops, or even dead snail shell. They may also start life grazing algal biofilms on the soil/rocks. Generally, growth banding suggests lifespans of 2–5 growth seasons. In adults, the shell may grow less rapidly and the snail seems to get a proportionally larger quantity of their calcium through grazing vegetation. More homogenous and higher F^{14}C values in aperture and whorl samples could reflect the high ecological mobility of adult specimens (Kerney et al. 1983) and would also explain why no *H. melanostoma* were collected in proximity to *Juniperus*, since gymnosperms such as this notoriously acidify soils (Augusto et al. 2015) and such environments are not preferred by land snails. This may also account for older average “ages” and wider variation in *H. melanostoma* specimens gathered in some parts of the Gebel Akhdar transect where the primary vegetation is *Juniperus* rather than *Ceratonia* or *Pistacia*, which would not acidify the soil in the same way. In these regions, there is also significantly less vegetation cover and poorer overall diversity potentially driving land snails such as *H. melanostoma* to seek carbonates from non-vegetative sources and exacerbating the “old carbon effect” where it occurs.

Regional Trends in the Radiocarbon Ecology of *Helix melanostoma*

Regional trends in the data are present. Comparison of the differences between $\text{F}^{14}\text{C}_{\text{apertures}}$ and $\text{F}^{14}\text{C}_{\text{apices}}$ values with levels of total vegetation finds weak but significant correlations for both apertures and apices (Table S1). This relation was found to improve noticeably if F^{14}C values were compared specifically with arboreal cover (Table 4). There is an even more pronounced association between vegetation cover and variations in $\text{F}^{14}\text{C}_{\text{wholeshell}}$ values (Figure 5a) that also show a weak but significant correlation and demonstrates a clear positive trend between increased total vegetation cover and higher F^{14}C values. This correlation is further improved when arboreal cover is compared directly to $\text{F}^{14}\text{C}_{\text{wholeshell}}$ values (Figure 5b). The observed relationship between F^{14}C values with vegetation cover suggests that the underlying ecological behavior of juvenile and adult *H. melanostoma* is quite similar in terms of the vegetation they

Table 5 Radiocarbon and stable isotope results and analysis on apertures, whorl fragments, apices and aggregated ages of *H. melanostoma* from the Cyrenaica region of Libya.

UBA aperture	UBA whorl frag.	UBA apices	Sample site ID	F ¹⁴ C apertures	±	F ¹⁴ C whorl frag.	±	F ¹⁴ C apices	±	F ¹⁴ C average	±
16927	17562		HF_Eco02	0.9820	0.0028	1.0070	0.0031			0.9945	0.0030
16928	17563		HF_Eco02	1.0003	0.0026	1.0190	0.0032			1.0097	0.0029
16929	17564		HF_Eco02	1.0180	0.0028	1.0340	0.0030			1.0260	0.0029
16930	17565		HF_Eco02	0.9626	0.0026	0.9650	0.0030			0.9638	0.0028
17566			HF_Eco02	1.0233	0.0033					1.0233	0.0033
17567			HF_Eco03	0.9640	0.003					0.9640	0.0030
17568			HF_Eco03	1.0040	0.0033					1.0040	0.0033
17569			HF_Eco03	1.0227	0.0030					1.0227	0.0030
17570			HF_Eco04	1.0633	0.0032					1.0633	0.0032
17571			HF_Eco05	0.9818	0.0032					0.9818	0.0032
17572			HF_Eco05	0.9759	0.0028					0.9759	0.0028
16931	17573		HF_Eco06	0.9920	0.0026	1.0070	0.0029			0.9995	0.0028
16932	17574		HF_Eco06	1.0167	0.0026	1.0070	0.0034			1.0119	0.0030
17575			HF_Eco06	1.0380	0.0031					1.0380	0.0031
17577			HF_Eco06	1.0227	0.0032					1.0227	0.0032
17578			HF_Eco06	1.0093	0.0033					1.0093	0.0033
17579			HF_Eco06	1.0055	0.0030					1.0055	0.0030
17580			HF_Eco06	1.0021	0.0030					1.0021	0.0030
18082			HF_Eco06	1.0428	0.0040					1.0428	0.0040
21757		21756	HF12 Apollonia	0.8891	0.0025			0.8511	0.0028	0.8701	0.0027
21760		21759	HF12 Apollonia	0.8743	0.0026			0.8561	0.0037	0.8652	0.0032
		21758	HF12 Apollonia					0.8186	0.0032	0.8186	0.0032
		21761	HF12 Apollonia					0.8469	0.0036	0.8469	0.0036
		21762	HF12 Apollonia					0.8773	0.0039	0.8773	0.0039
21749		21748	HF12 Resthouse	0.9536	0.0026			0.8970	0.0030	0.9253	0.0028
21751		21750	HF12 Resthouse	0.9820	0.0026			0.8530	0.0025	0.9175	0.0026
21753		21752	HF12 Resthouse	0.8801	0.0024			0.8396	0.0026	0.8599	0.0025
21755		21754	HF12 Resthouse	0.9863	0.0034			0.9262	0.0038	0.9563	0.0036
		21773	ML12-6 - Chart.					0.9575	0.0038	0.9575	0.0038
21775		21774	ML12-6 - Chart.	0.9934	0.0049			0.9834	0.0041	0.9884	0.0045
21778	21777	21776	ML12-6 - Chart.	0.9659	0.0027	0.9015	0.0027	0.9155	0.0027	0.9276	0.0027
21780		21779	ML12-9-Geb1	0.9876	0.0028			0.9220	0.0033	0.9548	0.0031
21782		21781	ML12-9-Geb1	1.0064	0.0028			0.9975	0.0028	1.0020	0.0028
		21783	ML12-9-Geb1					0.9116	0.0037	0.9116	0.0037
21785		21784	ML12-10-Geb2	0.9651	0.0027			0.9456	0.0027	0.9554	0.0027
21787		21786	ML12-10-Geb2	1.0066	0.0028			0.9353	0.0026	0.971	0.0027
21789		21788	ML12-10-Geb2	0.9270	0.0028			0.8932	0.0029	0.9101	0.0029

Table 6 Table showing the limestone effect observed in sampled mollusks in the form of differences in F¹⁴C values between dietary sources and shells across the Gebel Akhdar.

UBA aperture	UBA whorl frag.	UBA apices	Sample site ID	F ¹⁴ C average* ±	F ¹⁴ C diet ±	F ¹⁴ C average minus F ¹⁴ C diet ±	F ¹⁴ C diff. as %	Age offset (¹⁴ C yr) ±	Measured δ ¹³ C shell ‰ VPDB	Body δ ¹³ C ‰ VPDB	Predicted δ ¹³ C shell	Predicted δ ¹³ C minus measured δ ¹³ C				
16927	17562		HF_Eco02	0.9945	0.003	1.0571	0.0030	-0.0626	0.0042	-6.26%	490	33	-10.3	-23.4	-9.9	0.4
16928	17563		HF_Eco02	1.0097	0.0029	1.0571	0.0030	-0.0475	0.0042	-4.75%	369	32	-10.4	-24.1	-10.6	-0.2
16929	17564		HF_Eco02	1.026	0.0029	1.0571	0.0030	-0.0311	0.0042	-3.11%	240	32	-10.3	24.4	-10.8	-0.5
16930	17565		HF_Eco02	0.9638	0.0028	1.0571	0.0030	-0.0933	0.0041	-9.33%	742	33	-10.8	23.9	-10.4	0.4
17566			HF_Eco02	1.0233	0.0033	1.0571	0.0030	-0.0338	0.0045	-3.38%	261	35	-10.2	-23.6	-10.1	0.1
17567			HF_Eco03	0.964	0.003	1.0571	0.0030	-0.0931	0.0042	-9.31%	741	34	-10.5	-23.8	-10.3	0.2
17568			HF_Eco03	1.004	0.0033	1.0571	0.0030	-0.0531	0.0045	-5.31%	414	35	-10	-23.7	-10.2	-0.2
16932	17574		HF_Eco06	1.0119	0.003	1.0571	0.0030	-0.0453	0.0042	-3.44%	351	33	-11.6	-25.2	-11.6	0
17575			HF_Eco06	1.038	0.0031	1.0571	0.0030	-0.0191	0.0043	0.62%	146	33	-11.8	-25.7	-12	-0.2
17577			HF_Eco06	1.0227	0.0032	1.0571	0.0030	-0.0344	0.0044	-7.53%	266	34	-11.2	-23.3	-9.8	1.4
17578			HF_Eco06	1.0093	0.0033	1.0571	0.0030	-0.0478	0.0045	-8.12%	372	35	-11.5	-23.9	-10.4	1.1
17579			HF_Eco06	1.0055	0.003	1.0571	0.0030	-0.0516	0.0042	-5.76%	402	33	-11.9	-24.9	-11.3	0.6
17580			HF_Eco06	1.0021	0.003	1.0571	0.0030	-0.0550	0.0042	-4.53%	429	33	-11	-24.8	-11.2	-0.2
18082			HF_Eco06	1.0428	0.004	1.0571	0.0030	-0.0143	0.0050	-1.91%	109	38	-10.7	-24.4	-10.9	-0.2
21757		21756	HF12 Apollonia	0.8701	0.0027	1.0501	0.0031	-0.18	0.0041	-3.44%	1510	34	-11.2	-27.3	-13.9	-2.7
21751		21750	HF12 Resthouse	0.9175	0.0026	1.0388	0.0030	-0.1213	0.0039	-4.78%	997	33	-9.9	-29.5	-11.4	-1.5
21753		21752	HF12 Resthouse	0.8599	0.0025	1.0388	0.0030	-0.179	0.0039	-5.16%	1518	33	-10.8	-25.4	-13.8	-3
21755		21754	HF12 Resthouse	0.9563	0.0036	1.0388	0.0030	-0.0826	0.0047	-5.50%	665	38	-10.8	-24.6	-12.4	-1.6
		21773	ML12-6-Chart.	0.9575	0.0038	1.0358	0.0018	-0.0783	0.0042	-1.43%	631	35	-12.3	-24.9	-15.7	-3.4
21775		21774	ML12-6-Chart.	0.9884	0.0045	1.0358	0.0018	-0.0474	0.0032	-18.00%	376	39	-12	-27.5	-14.6	-2.6
21778	21777	21776	ML12-6-Chart.	0.9276	0.0027	1.0358	0.0018	-0.1082	0.0042	-18.49%	886	27	-9.6	-26.0	-11.8	-2.2
21780		21779	ML12-9-Geb1	0.9548	0.0031	1.0381	0.0029	-0.0833	0.0040	-23.15%	672	34	-10.3	-24.2	-10.7	-0.4
21782		21781	ML12-9-Geb1	1.002	0.0028	1.0381	0.0029	-0.0362	0.0047	-20.32%	284	32	-9.7	-25.4	-11.8	-2.1
21785		21784	ML12-10-Geb2	0.9554	0.0027	1.0314	0.0029	-0.0761	0.0040	-17.28%	615	32	-10.5	-25.7	-12.1	-1.6
21787		21786	ML12-10-Geb2	0.971	0.0027	1.0314	0.0029	-0.0605	0.0040	-11.35%	485	32	-10.9	-25.9	-12.3	-1.4
21789		21788	ML12-10-Geb2	0.9101	0.0029	1.0314	0.0029	-0.1213	0.0041	-12.13%	1005	34	-11.1	-25.7	-13.4	-2.3

*F¹⁴C shell average is calculated based on the median F¹⁴C value for samples taken from specific shells. Estimation of old carbon content was based on the methods set out in Pigati et al. (2010). Data paired with δ¹³C_{Shell} and δ¹³C_{Body} values measured by IRMS (not machine measurement from the AMS) on selected samples (Prendergast 2013) and δ¹³C predicted was calculated using the method described in Stott (2002).

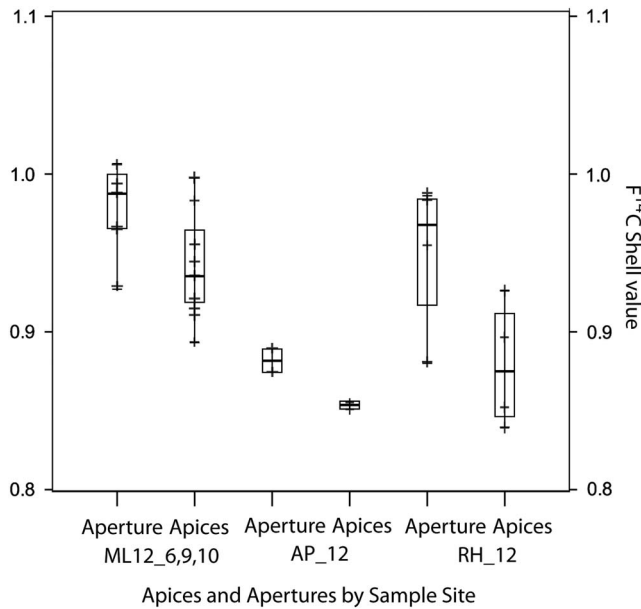


Figure 4 Boxplot of $F^{14}C$ of *H. melanostoma* apices and apertures by site: ML12_6,9,10: wt. average of aperture and apices values ($n = 9$) from sample sites on the Gebel Akhdar. AP_12: Apollonia (Susa) ($n = 2$); RH_12: Project Resthouse (Shahat) ($n = 4$). Individual data points are displayed within the box plot as “+”.

consume across all the sample sites. It is interesting that this correlation is noticeably stronger in the combined $F^{14}C_{\text{shell}}$ values ($F^{14}C_{\text{wholeshell}}$) analysis than for either apertures or apices individually. This is likely to be due to the similarity of the datasets amplifying the observed trend. It is also probable that this approach filters out a significant portion of the local noise reflected in $F^{14}C$ values at individual sites. The stronger correlations between $F^{14}C$ values and tree and shrub cover are almost certainly a reflection of the dietary preferences of *H. melanostoma*, indicating that they prefer taller arboreal vegetation over grasses and low-lying herbaceous vegetation. The availability of such vegetation would appear to be a key variable that molds the ^{14}C ecology of *H. melanostoma* in the study region where the vegetation is largely limited by grazing pressure since rainfall levels are always sufficient to support trees. Therefore, ^{14}C ecology is responding to grazing pressure from domesticated animals and does not reflect a truly natural situation. This conforms well to existing research (e.g. Stott 2002) and could be an important influence if dating shells of this species found in association with Neolithic or later archaeological material where grazing of the local landscape would have been possible.

These findings also strongly support the hypothesis raised in other studies (e.g. Pigati et al. 2010) that calcium availability could be a contributory factor to the incorporation of old carbon into the diet. In almost all the sites, limestone availability was observed to be essentially inversely proportional to vegetation cover, so the positive trend between vegetation and higher $F^{14}C$ values is also a negative trend between limestone availability and lower $F^{14}C$ values. The most striking example of this is at Apollonia, which has very poor vegetation cover, and $F^{14}C$ values for apices and apertures that are correspondingly much lower (Table 5). It may be, however, that this is a function not of limestone availability but of vegetation availability, which enforces grazing of endolithic algae, with an associated old carbon uptake from the limestone.

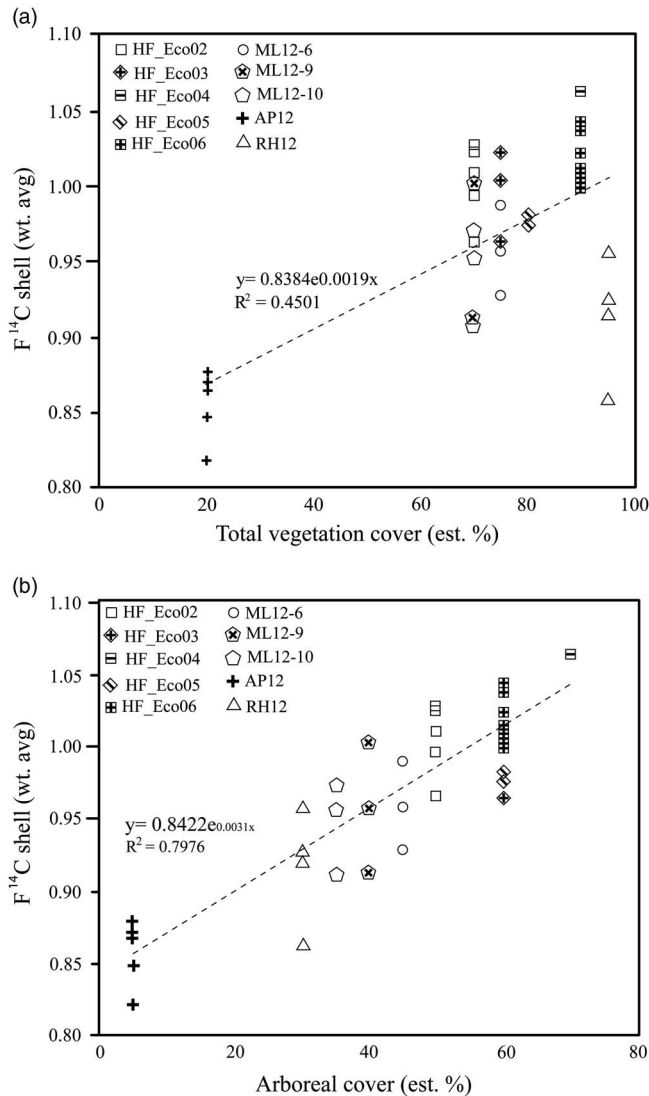


Figure 5 (a) Relationship between $F^{14}C_{\text{wholeshell}}$ and total vegetation cover; (b) relationship between $F^{14}C_{\text{wholeshell}}$ and arboreal vegetation cover.

Previous study has demonstrated that *H. melanostoma* in the sample region have a diet of C3 vegetation (Prendergast et al. 2014, 2015). As part of that study a modeling exercise calculating the dietary $\delta^{13}C$ values for *H. melanostoma* if their diet was 100% vegetation was carried out. The results of this exercise together with the ^{14}C age offset are included in Table 6. The measured $\delta^{13}C$ shell values, when compared with their modeled values, show both a significant observable trend in measured versus predicted $\delta^{13}C$ enrichment in *H. melanostoma* shells (Figure 6a) across the study areas (Table S1). Measured samples from the Haua Fteah are mostly within the margin of error ($\pm 1\%$) when compared. However, samples, from other sites have much greater variation. There is a significant, positive relationship, with a moderate correlation between $\delta^{13}C_{\text{shell}}$ enrichment/depletion and arboreal vegetation cover (Figure 6b). Further comparison of $\delta^{13}C_{\text{shell}}$

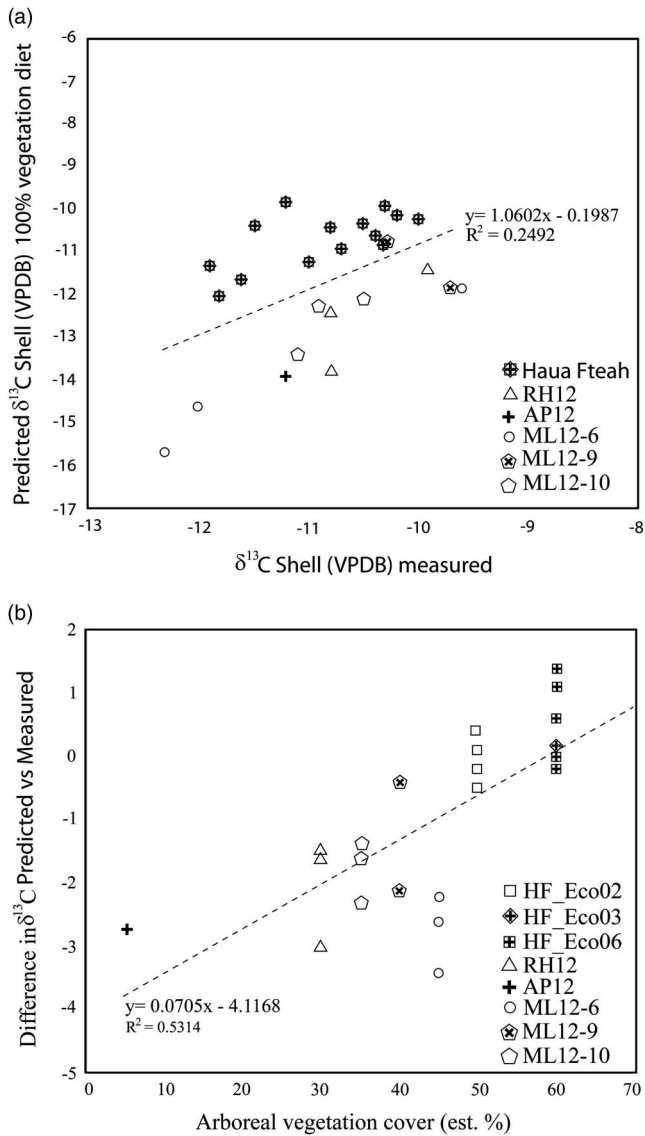


Figure 6 (a) Measured against predicted enrichment $\delta^{13}\text{C}$ across the study area; (b) $\delta^{13}\text{C}$ enrichment and depletion against arboreal cover across the study area. Dashed lines represent the lines of best fit.

variations against other potential variables (Table S1) found that there were no significant trends across the sites sampled and no significant observable relationship between $\delta^{13}\text{C}_{\text{shell}}$ and $\text{F}^{14}\text{C}_{\text{wholeshell}}$ values when compared directly across sampled sites (Table S1).

These findings are not surprising from an ecological or stable isotope perspective since there is a large body of past work that has found that the $\delta^{13}\text{C}$ of land-snail shells has a strong relationship with diet (e.g. Stott 2002; Metref et al. 2003; Colonese et al. 2013; 2014; Yanes et al. 2008; 2014). The changes in enrichment and depletion and their correlation towards F^{14}C values is interesting for ^{14}C dating and the issue of “old carbon.” Sites that present the lowest

$F^{14}C_{\text{shell}}$ values e.g. Apollonia, Shahat (Table 6), also present the most substantial enrichment in $\delta^{13}C$ ($>2\%$) values compared with what would be expected if their diet was entirely vegetation. The Haua Fteah, which has good vegetation cover and higher $F^{14}C$ values, has less enrichment and are in some cases marginal depletion of $\delta^{13}C$ values ($\pm 1\%$). The trend shown in Figure 6b suggests that the magnitude of enrichment of samples may specifically relate to the availability of suitable dietary vegetation at sample sites, e.g. *Ceratonia siliqua* or *Pistacia lentiscus*. Areas lacking in arboreal vegetation in northeastern Libya also tend to have more significant outcroppings of limestone, which would account for lower $F^{14}C$ values for samples from those regions as *H. melanostoma* could change its diet to compensate for the lack of vegetation. When considering the suitability of a species such as *H. melanostoma* for ^{14}C dating this demonstrates the impact of dietary pressure on the $F^{14}C$ value of the shell very clearly in a way that was not picked up by direct comparison of the $\delta^{13}C$ and $F^{14}C$ values. This relationship suggests that such a modeling exercise could be a useful tool in the future to flag specimens that may have an old carbon effect without directly dating ^{14}C dating a sample.

Relationships between $F^{14}C_{\text{shell}}$ and $F^{14}C_{\text{diet}}$

The relationship between $F^{14}C_{\text{shell}}$ and $F^{14}C_{\text{diet}}$ was also investigated for *H. melanostoma*. There is only a slight correlation between changing $F^{14}C$ dietary values and $F^{14}C$ shell values (Table 3). This is broadly expected, as there is considerable variation within individual sample sites in $F^{14}C_{\text{shell}}$ and the sample size is not particularly large for some locations. In order to further compare $F^{14}C$ shell and $F^{14}C$ diet, age corrected $\Delta^{14}C$ (Figure 7) values were calculated,

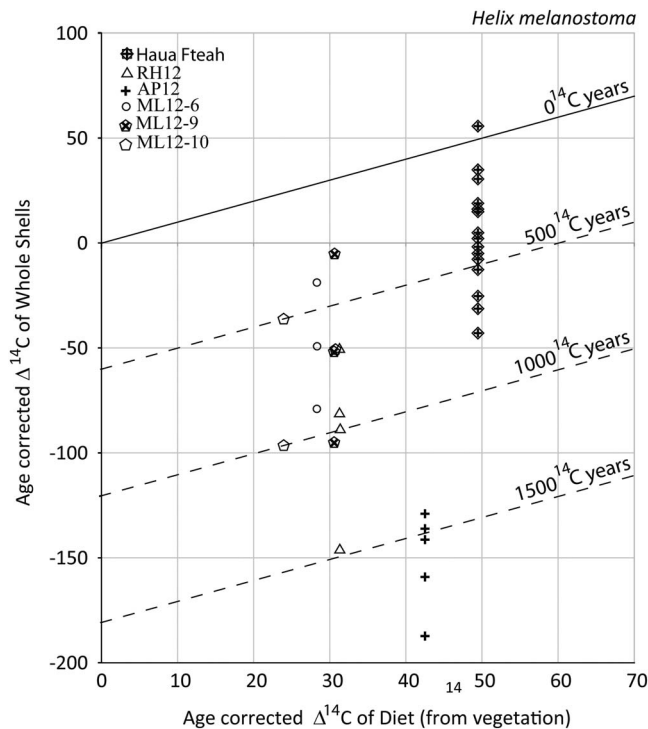


Figure 7 $F^{14}C_{\text{shell}}$ and $F^{14}C_{\text{diet}}$ plotted as age corrected $\Delta^{14}C$, demonstrating the variation and regional clustering of the old carbon effect for *H. melanostoma* in the study region.

and the proportion of old carbon in the diet derived following the methods set out in Pigati et al. (2010) after Stuiver and Polach (1977) to facilitate easier comparison to previous studies.

Findings imply that most specimens are incorporating a proportion of old carbon into their diets as a matter of course. What is significant is that there is a clear delineation that sits well with the earlier observations about vegetation availability and calcium dependency as modulating factors for shell carbonate composition. While the total range of values measured (Table 5) is large; (UBA 18082) 109 ± 38 ^{14}C yr to (UBA 21758) 2001 ± 43 ^{14}C yr with a weighted mean of 659 ± 76 ^{14}C yr, the majority of samples from non-urbanized sample locations have only a small old carbon effect with a weighted mean of 476 ± 48 ^{14}C yr, which approximately relates to between 1% and 9% of their dietary F^{14}C coming from non-organic carbonate sources (Table 6) where this has been measured. The clear exceptions are samples from Apollonia and a single sample from Shahat; the urbanized sample locations manifest old carbon effects of >1000 ^{14}C yr and skew the averages accordingly. These more extreme carbonate environments are most likely to represent the modern extremities of the old carbon effect on *H. melanostoma*. Such environments are highly unlikely to impact archaeological ages significantly since palynological evidence (Hunt et al. 2015; Simpson 2016) shows relatively stable vegetation cover in the region throughout the later Quaternary.

CONCLUSIONS

The ecological observations from all the sampled sites suggest that *H. melanostoma* has a very homogenous ^{14}C ecology in its natural habitats. There is a similar level of variation within F^{14}C values across the majority of sample sites within the study area. Only in localities with drastically anthropogenically altered habitats (Apollonia, Shahat) does *H. melanostoma* exhibit F^{14}C values that are significantly distant from values observed at less impacted sites. This significant deviation from predicted levels of F^{14}C for a species that preferentially consumes fresh, leafy vegetation is attributed to both the lack of choice vegetation and the influence of anthropogenic factors upon available vegetation (e.g. cement works, air pollution). These issues are unlikely to be significant in an archaeological context prior to the Roman Period and therefore these two localities may be regarded as clear outliers. It is significant that F^{14}C values of apices are consistently lower than apertures across all sample sites. The scatter of F^{14}C values for shell apertures, the most widely comparable F^{14}C dataset, is very constrained. There is also a demonstrable trend of enrichment of $\delta^{13}\text{C}$ at coastal and urbanized sample sites, decreasing availability of choice vegetation such as *Ceratonia siliqua* and *Pistacia lentiscus* and lower F^{14}C values. This may suggest that enrichment in $\delta^{13}\text{C}$ is a proxy indicator for the old carbon effect on *H. melanostoma* under certain circumstances.

The most significant of these ecological observations are the very similar, and thus predictable, differences between apertures and apices at all the sites and the predictable behavior of old carbon incorporation, both of which appear to be heavily modulated by the availability of choice vegetation such as *Ceratonia siliqua* and *Pistacia lentiscus*. The scale of differences between apertures and apices are very similar despite the distance between the sample sites, some variation in numbers of growth seasons and size of specimens. This suggests that the ^{14}C ecology of juvenile *H. melanostoma* may differ from that of adults where incorporation of carbonates is concerned. The lack of a significant difference between adult and juvenile $\delta^{13}\text{C}$, even taking into consideration the small number of samples involved ($n = 5$), opens up the possibility that the factor causing this variation is not vegetation availability. The investigation of modern *H. melanostoma* further suggests weaknesses within the existing methods used in the ^{14}C dating of land mollusks, particularly the use of $\delta^{13}\text{C}$ to estimate the impact of the

“limestone effect” within mollusk shells. In this instance this was found to over-emphasize the signature of dietary vegetation.

The similarity of the relationships between $F^{14}C$ values of shell apices and apertures to total vegetation cover suggests that both juvenile and adult *H. melanostoma* consume the same dietary vegetation, and that both are equally affected by this variable. Thus, the increased $F^{14}C$ values of *H. melanostoma* apices should originate in the non-organic carbonate component of diet and has the largest impact on the $F^{14}C$ values of the apices which is the longest growing part of the animal. It may also relate to subtle differences in vital effects due to differing fractionation chemistry between adult and juvenile animals, although only a limited number of juvenile snails were available to test this.

This is further supported by a marked increase in “age” difference between the $F^{14}C$ of apices and apertures in Apollonia and those from the Gebel Akhdar. The landscape around Apollonia is limestone-dominated and has poor vegetation availability, whereas the Gebel Akhdar is comparatively well vegetated with much less outcropping limestone. This reinforces the conclusion that the significantly decreased $F^{14}C$ values at Apollonia are probably an extreme response to dietary pressure. It therefore seems reasonable to infer that dietary calcium availability could be the controlling factor for the incorporation of limestone into the diet of *H. melanostoma*. The difference in the $F^{14}C$ values could be an indicator that juveniles simply consume more calcium carbonate than adults in order to construct their shells quickly during their first season of growth. There is much scope for further work on this hypothesis. Moreover, many of the findings observed for *H. melanostoma* should also be observable for other species with similar ecological behavior and should therefore be replicable for other taxa, elsewhere in the Mediterranean and further afield.

This study demonstrates that, in circumstances where there is a relatively complete vegetation cover including at least some trees and shrubs, the ^{14}C ecology of *H. melanostoma* is stable and predictable. The observed instabilities in the ^{14}C ecology of this species in Cyrenaica can all be attributed to the results of intense human activity which would not have been present before the Roman Period. Therefore, this study opens up the use of *H. melanostoma* and species with similar ecological behavior for ^{14}C dating of archaeological and geological deposits, with the use of a suitable offset to correct for the predictable effects of diet observed here.

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SUPPLEMENTARY MATERIAL

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