



RESEARCH ARTICLE

Silicon dioxide nanofertilizers improve photosynthetic capacity of two Criollo cocoa clones (*Theobroma cacao* L.)

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Summary

Studies on the effect of nanofertilizers (NF) in physiological performance of plants is scarce, especially that related to substances encapsulated into silicon dioxide (SiO₂) nanoparticles in cocoa plants. The effect of foliar application of SiO₂-NF on nutrient contents, gas exchange, photochemical activity, photosynthetic pigments, total soluble protein (TSP), photosynthetic nitrogen use efficiency (PNUE), and growth in seedlings of two cocoa clones (OC-61 and BR-05) in a greenhouse was assessed. Spraying with SiO₂-NF increased net photosynthetic rate (A) by 16 and 60% and electron transport rate (J) by 52 and 162% in clones OC-61 and BR-05, respectively, without changes in photosynthetic pigment concentration in either clone. The SiO₂-NF caused a decrease of 37 and 22% in stomatal conductance in OC-61 and BR-05, respectively; a similar trend was observed in transpiration rate, causing an increase of 42 and 100% in water use efficiency in OC-61 and BR-05, respectively. In both clones, diameter of graft increased on average 28% with SiO₂-NF. Higher photosynthetic capacity was related to an increase in leaf N, P, and TSP. A significant reduction in PNUE (A/N ratio) was found in OC-61, whereas in BR-05 PNUE increased after spraying with SiO₂-NF. Overall, spraying with SiO₂-NF had a positive effect on photosynthetic processes in both cocoa clones, associated with an increase in nutrients content, which translated into improved growth. A differential physiological response to spraying with SiO₂-NF between clones was also found, with BR-05 being the clone with a better physiological response during the establishment and development stages.

Keywords: Cocoa; Fluorescence; Foliar fertilizer; Nanotechnology; Nutrients; SiO₂; photosynthesis

Abbreviations: A, net photosynthetic rate; BR-05, Bromelia 05; C, control plants; DG, diameter of graft; DM, dry mass; E, transpiration rate; FM, fresh mass; F_v/F_m, maximum quantum yield PSII; g_s, stomatal conductance; H, total height; HF, high frequency application; HG, height of the main branch of graft; J, electron transport rate; LF, low frequency application; NF, nanofertilizers; OC-61, Ocumare 61; PNUE, photosynthetic nitrogen use efficiency; PPF, photosynthetic photon flux density; q_N, non-photochemical quenching; q_P, photochemical quenching; RWC, relative water content; SiO₂-NF, SiO₂ nanofertilizers; T_a, air temperature; T_L, leaf temperature; TSP, total soluble protein; WUE, water use efficiency; Δ_w, leaf-to-air vapor pressure deficit; Φ_{PSII}, relative quantum yield of PSII

Introduction

Cocoa (*Theobroma cacao* L., Malvaceae) is a native tree from tropical rainforest of South America (Motamayor *et al.*, 2002). Cocoa represents one of the commercially most important perennial crops worldwide, with an estimated global output of 4.75 million tons in 2019 and a yield greater

than 600 kg/ha (ICCO, 2020). Venezuela is part of a select group of countries in America producing fine aroma cocoa, which together account for about 7% of total world production (ICCO, 2020). Cocoa activities and production in Venezuela are very low; therefore, the search for new strategies to recover cocoa productivity is required.

The increasing food demand due to the growth of world population has driven the large-scale use of traditional fertilizers. Due to resource limitations and the low efficiency of fertilizer usage, the cost to farmers is dramatically increased (Raliya *et al.*, 2017). Foliar fertilization has been used to supply nutrients, hormones, biostimulants, and other substances that are beneficial to plants, which usually results in an increase in growth, yield, and quality, and greater resistance to pests and diseases (Fernández *et al.*, 2015). One of the benefits of the applications of foliar nutrients is that the constant application, which allows adjusting the formula and dosing the fertilizer, avoiding the use of high concentrations that could generate an environmental impact (Morales-Díaz *et al.*, 2017).

Foliar fertilizer applications imply that nutrients supplied must be absorbed and exported from the leaf surfaces to the mesophyll tissue (Monreal *et al.*, 2015). Leaf fertilizer nutrients move through leaf cuticle, stomatal pore, trichomes, or others epidermis structures; however, only small amounts of nutrients move through stomatal pores (<10%), because they represent a small proportion of the total leaf area (Fernández *et al.*, 2015).

Nanotechnology is defined as the research and technological development on a scale of 1 to 100 nm (nanoparticles) using atoms, molecules, or macromolecules (Raliya *et al.*, 2017; Ram *et al.*, 2018). Nanotechnology may be the key to the evolution of the science of precision agriculture, it is emerging as a promising strategy that offers great potential to adapt the production of fertilizers with the desired chemical composition, improve water and nutrient use efficiency, reduce environmental impact, and increase productivity of crops (Morales-Díaz *et al.*, 2017; Raliya *et al.*, 2017; Wang *et al.*, 2020).

Nanoparticles (NPs) have exhibited significant potential for promoting photosynthesis and enhancing crop productivity, therefore understanding the fundamental interactions between NPs and plants is crucial for the sustainable development of agriculture (Wang *et al.*, 2020). Recently, it has been shown that NPs are an attractive alternative for the manufacture of nanofertilizers (NF), which are nutrient compounds of nanostructured formulations that can be applied to plants, allowing efficient absorption or slow release of micronutrients, macronutrients, fungicides and insecticides, and thus are more effective and efficient than traditional fertilizers due to their larger impact on the nutritional quality and stress tolerance of crops (Elsheery *et al.*, 2020; Morales-Díaz *et al.*, 2017; Raliya *et al.*, 2017).

A large number of studies related to the use of NF in an exhaustive number of crops species have been reported (Elsheery *et al.*, 2020; Raliya *et al.*, 2017); however, plant physiological responses to NF application are scarce, especially, the effects of application of SiO₂ nanofertilizers (SiO₂-NF) in cocoa. Application of foliar Si in cocoa plants subjected to water deficit caused an increase in photochemical efficiency, whereas stomatal conductance (g_s), transpiration rate (E), and relative water content (RWC) decreased in non-irrigated plants at specific SiO₂ concentrations (Zanetti *et al.*, 2016). Nano-silica confers tolerance to salt stress on tomato (Haghighi and Pessaraki, 2013), tolerance to drought in wheat and SiO₂ nanoparticles mitigate chilling effects on photosynthesis and photoprotection in sugarcane, contributing to the increase of net photosynthetic rate (A), biomass production, photosynthetic pigments and TSP concentration, and RWC (Ahmed *et al.*, 2016; Elsheery *et al.*, 2020; Raliya *et al.*, 2017).

There is an interest in studying ecophysiological traits in some Venezuelan cocoa clones (Ávila-Lovera *et al.*, 2016, 2021; De Almeida *et al.*, 2018; Tezara *et al.*, 2016, 2020). However, the effect of specific NF, using SiO₂ nanoparticles as nutrients vehicle (SiO₂-NF), on the physiological traits of cocoa plants is largely unknown. In order to gain knowledge on the effect of SiO₂-NF spray frequency on photosynthetic activity of two cocoa clones, we evaluated gas exchange, photosynthetic pigments and nutrient concentration, photochemical activity of PSII, and photosynthetic nitrogen and phosphorus use efficiency of two Criollo cocoa clones (OC-61 and BR-05), and whether there is a

differential physiological response to spraying with SiO₂-NF between the two clones. We expected positive effects of SiO₂-NF spray on photosynthetic activity in both clones, which may contribute significantly to the incorporation of nutrients, especially those related to photosynthesis.

Materials and Methods

Plant material and growth conditions

The study was carried out in a greenhouse at the ‘Las Bromelias’ farm in Cumboto, Central coast, Venezuela (10° 25' 10.3" N and 67° 46' 48.2" W) at 500 m asl. Saplings of the two cocoa clones, Ocumare 61 (OC-61), and Bromelia 05 (BR-05) tested in this study were obtained by cuttings grafted onto OC-61 rootstock and planted in 5 kg bags of sandy soil from the banks of a nearby river. All plants were fertilized with N:P:K 15:15:15 once a week to ensure their growth and establishment before starting the experiments. A new exhaustive classification of cocoa germplasm has been proposed in ten genetically differentiated groups, where Criollo cocoa is one of them (Motamayor *et al.*, 2008). Cocoa clones were selected based on their high productivity, presence of Criollo cocoa traits, and their presence within the study area: OC-61 is a modern Criollo, that is, a hybrid between Criollo and Forastero cocoa that differs in quality, vigor, and yield, and has many Criollo morphological traits such as high quality and fine flavor (Motamayor *et al.*, 2002); BR-05 is a new selection with characteristics of Criollo generated in the “Las Bromelias” farm.

Experimental design

Three blocks (one control and two spray frequencies with SiO₂-NFs) of 30 m² area shaded to 30% of sunlight with the use of neutral polythene sheets were established in the greenhouse, and each block consisting of 30 plants: 15 plants per clone 50 cm apart (3 m × 2.5 m), with a distance of 2 m between blocks. Plants were rotated weekly in each treatment within and between blocks, thus avoiding possible block and edge effects.

Plant treatments

Seven months after grafting, saplings of both clones of similar number of leaves and height (approximately 50–60 cm) were subjected to a 35-day long treatment of two foliar spray frequencies with 25 mL NF per plant with a constant concentration of 11 μmol mol⁻¹ nanostructured SiO₂-NFs suspended in water, as follows: (1) low frequency application, LF (spray SiO₂-NFs every 15 days); (2) high frequency application, HF (spray SiO₂-NFs every 7 days), and (3) control (C) plants without application of SiO₂-NFs.

The fertilizer is a nanocomposite of silicon dioxide (SiO₂) nanoparticles doped with different elements. The SiO₂ nanoparticles encapsulated fertilizer composites (SiO₂-NF) consisted of SiO₂ doped with 0.025% Co, 0.025% B, 0.025% Mo, 0.35% Mg, 0.15% Fe, 0.15% Cu, 0.10% Mn, 0.25% Zn, 0.35% Ca, and 0.20% S, and coated in a chitosan matrix (N:P:K 15:15:15).

The SiO₂ NPs were synthesized from rice husks (RHs). The RHs were washed and lixiviated in an aqueous solution of HCl 1% m/v for 24 h. Then, the lixiviated RHs were rinsed with water until pH 7, and dried in an oven at 110°C. The dried RHs were heated to 500°C for 1 h and 700°C for 9 h with a ramp of 500°C/h, and the resulting white solid was ball-milled until obtaining nanometric distribution (1–100 nm), which was verified by dynamic light dispersion (DLS) technique.

Nanocomposites were prepared by wet impregnation methods. SiO₂ NPs were impregnated with different metal ion salts (FeCl₃, CaCl₂, MgCl₂, and CuSO₄) in the minimum quantity of water. Once the impregnation process ended the mixture was dried and heated to 350°C. The salt concentration warranted 30% of weight of metal in the SiO₂ nanoparticle.

Figure 1 shows images obtained by scanning electron microscopy (SEM) and DLS data for the raw SiO₂ nanoparticles. The DLS signal data is more accurate and showed a wide size distribution

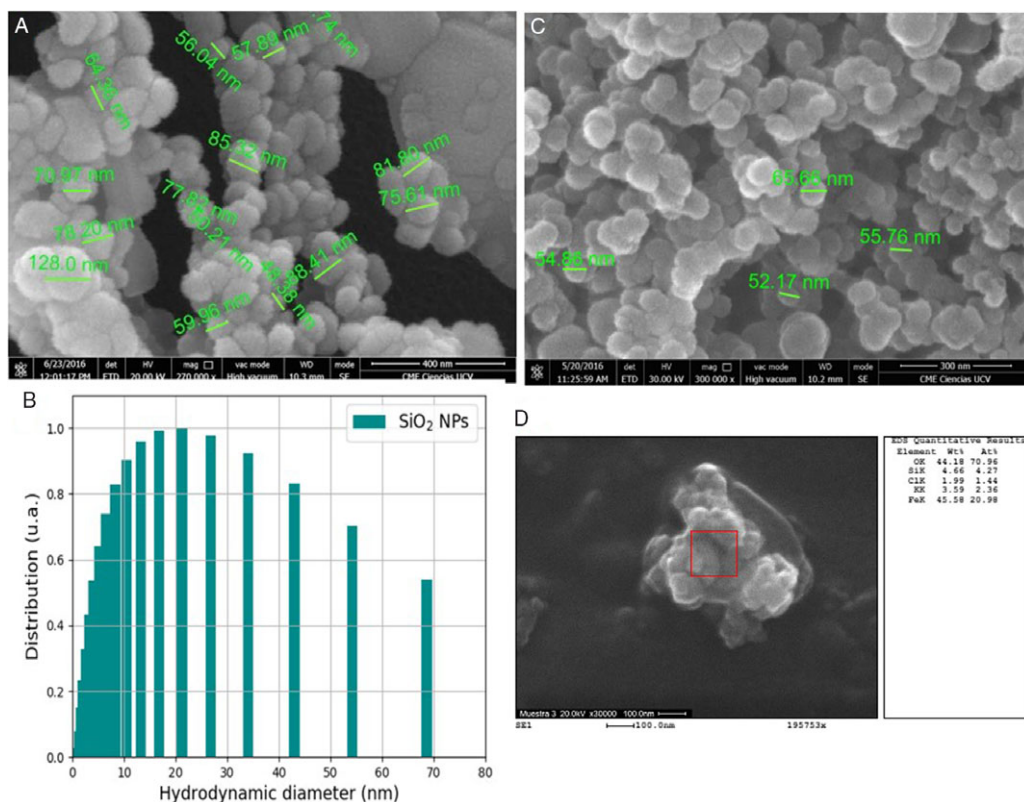


Figure 1. A, Scanning electron microscopy (SEM) images of SiO₂ NPs from rice husks ash; B, SiO₂ NPs DLS signal; C, SiO₂ NPs with Chitosan; D, SiO₂ NPs doped with iron.

for the NPs of an approximate size of 34 nm and SiO₂ NPs stabilized with chitosan. The variation in the size distribution is too small to be considered further.

Silicon nanoparticles and mesoporous aluminosilicates have important properties such as highly ordered channels with high accessible porosity (with diameters between 2 and 50 nm, according to the chemical compound nomenclature system, IUPAC), large surface areas, active sites for adsorption, ion exchange, and catalysis (Monreal *et al.*, 2015; Wu *et al.*, 2008).

All plants were placed on benches at a height of 50 cm above the ground to avoid soil nutrient uptake by roots.

Soil characteristics

Soil from the banks of a nearby river used in the study was sandy loam, slightly acidic (pH = 6.4) and not saline (cation exchange capacity of 1.3 mΩ, and composed of 77.5% sand, 12.5% loam, and 10% clay). The soil had high concentration of Si (13.2 mol kg⁻¹). High concentration of K, Mg, and Ca (42, 215 and 148 mmol kg⁻¹, respectively), low concentration of P (0.13 mmol kg⁻¹), and no N were detected in the irrigation water.

Microclimatic measurements

Air temperature and relative humidity in the greenhouse were measured throughout the experiment with two HOBO Pro V2 loggers and data stored in a HOBO Waterproof Shuttle (Onset

Computer Corporation, Pocasset, MA, USA). Photosynthetic photon flux density (PPFD) was measured with a quantum sensor and a light meter (LI-250, LI-COR Inc., Lincoln, NE, USA). Leaf and soil temperature were measured with thermistors (YSI 04B0618-409B) connected to a Switch Craft LN4153-405 (8402-10, Cole-Parmer Instrument Company, IL, USA) and connected to a telethermometer (Yellow Springs Instruments Co, Texas, USA). With data of temperature and relative humidity, leaf-to-air vapor pressure deficit (Δ_w) values were calculated. Microclimatic variables were measured every hour between 07:00 and 17:00 h.

Leaf and soil nutrients concentration

Nutrient concentration was determined in the leaves used for the determination of photosynthetic pigments. Fresh mass (FM) of the leaves was recorded when they were collected, next they were washed with deionized water and dried in a ventilated oven at 60 °C until reaching a constant dry mass (DM). The leaves were ground to a powder and acid digestions (sulfuric and perchloric acid with a 4:1 ratio) were done on a digester block (Digestion Kjeltac System 40–1016, Tecator, Sweden). Total phosphorous concentration (P) was determined according to Murphy and Riley (1962); major cations (Na^+ , K^+ , Ca^{+2} , and Mg^{+2}) were determined by flame atomic absorption (SpectrAA 55B, Varian, Australia). The total organic nitrogen concentration was determined according to the microKjeldahl method (Jackson, 1958). The leaf and soil Si concentration was determined by X-ray diffraction using a Quanta FEI 2501 Scanning Electron Microscope with an EDAX/EDX detector.

Physiological variables

Physiological measurements were made on the third mature, fully expanded leaf of seven plants per clone per treatment. Measurements were carried out at the beginning of the experiment (day zero) and every 7 days after the application of sprays with SiO_2 -NFs during 35 d. Samples of leaf, soil, and water used for irrigation were collected to determine macronutrient concentrations at the beginning (day zero) and at the end of the experiment (day 35). Aluminum sheets were placed in the base of the stem of the plant (on the ground), to avoid the incorporation of nutrients from the SiO_2 -NFs to the soil throughout the study.

Gas exchange

Measurements of leaf gas exchange were made in intact leaves with a portable infrared gas analyzer (CIRAS-II, PP Systems Inc., Amesbury, MA, USA) connected to a leaf chamber PLC (B) at a CO_2 concentration of $410 \pm 10 \mu\text{mol mol}^{-1}$, a leaf chamber temperature of $28.0 \pm 0.5^\circ\text{C}$, a Δ_w of $1.25 \pm 0.017 \text{ kPa}$ and a PPFD of $1000 \mu\text{mol m}^{-2} \text{ s}^{-1}$ (light provided by a LED-based light unit from the same manufacturer). Water use efficiency was calculated as $\text{WUE} = \text{A/E}$. Measurements were taken from 10:00 to 12:00 h because it was previously determined that maximum rates of photosynthesis are attained during this period (De Almeida *et al.*, 2018).

Photosynthetic pigment and total soluble protein concentration

Leaf chlorophyll and carotenoid concentrations were determined in 80% chilled acetone extracts of leaf discs of known area in leaves ($n = 7$), after Wellburn (1994). Total soluble protein concentration (TSP) was determined in leaves ($n = 7$) using a standard protocol (Bradford, 1976).

Specific leaf area

Specific leaf area (SLA) was calculated as the ratio between leaf area and dry mass in seven individuals per clone using a protocol described by Tezara *et al.* (2020).

Photochemical activity

Photochemical activity of PSII was determined by fluorescence measurements of chlorophyll in intact leaves of seven plants (one leaf per plant in seven different plants per treatment), using a fluorometer (PAM 2100, Walz, Effeltrich, Germany) after the protocol described by Genty *et al.* (1989). Measurements of maximum quantum yield PSII (F_v/F_m) were made at pre-dawn (dark-adapted leaves), where F_v = variable fluorescence, and F_m = maximum fluorescence in dark-adapted leaves, relative quantum yield of PSII at steady state photosynthesis (Φ_{PSII}), electron transport rate (J), photochemical (q_p), and non-photochemical (q_N) quenching coefficients were measured in light-adapted leaves at 10:00–12:00 h at a PPFD of $1000 \pm 10 \mu\text{mol m}^{-2} \text{s}^{-1}$. Values of J were estimated as $J = \Phi_{PSII} \times \text{PPFD} \times 0.84 \times 0.5$, where $\Phi_{PSII} = (F'_m - F_s)/F'_m$ in light-adapted leaves, F'_m = maximum fluorescence and F_s = steady-state fluorescence in light-adapted leaves.

Growth variables

Total height (H), height of the main branch of graft (H_G), and diameter of graft (D_G) were measured with a measuring tape and a vernier caliper, respectively, in four plants ($n = 4$) per clone and treatment after 14, 21, and 35 days of spray with SiO_2 -NFs.

Statistical analysis

Results are presented as mean \pm standard error (SE). Significance was tested at $p < 0.05$ and assessed by one-way ANOVA using the Statistica 10 package (StatSoft Inc., Tulsa, OK, USA). All regressions and correlations were tested for significance at $P < 0.05$. Curves were fitted using the Sigmaplot 11.0 package (Systat Software, Inc., San Jose, CA, USA).

Results

Microclimatic parameters

Mean maximum PPFD outside the greenhouse was reached at noon ($1600 \pm 100 \mu\text{mol m}^{-2} \text{s}^{-1}$); the neutral mesh effectively reduced 70% of the PPFD inside the greenhouse. The highest T_a was also reached at noon ($30 \pm 2 \text{ }^\circ\text{C}$). Air, soil, and leaf temperature ranged from 20 to 32 $^\circ\text{C}$, and had a similar trend throughout the day, the lowest values of temperature being found in soil. Relative humidity decreased from 90% in the early morning to 60% in the afternoon (Figure 2). Maximum leaf Δ_w , though, was relatively low during the experiments ($1.6 \pm 0.3 \text{ kPa}$).

Leaf and soil nutrients

No significant differences in leaf ($p > 0.63$) and soil ($p > 0.75$) nutrient concentration were observed between day 0 (start the experiments) and day 35 (end of the experiments). In OC-61, leaf N concentration in LF and HF was 30 and 31% higher than in control plants. Also, leaf P concentration was 26% higher in HF in OC-61 than in control plants. For BR-05, there was a significant increase of 22 and 10% and of 38 and 23% in leaf N and K concentration, respectively in LF and HF (Table 1). No significant differences in Mg and Ca concentration in the leaves of both clones were observed among treatments. For BR-05, there was a significant increase of two-fold on leaf Si concentration in LF and HF, whereas no changes were observed in OC-61. In soil, differences were found in Ca concentration between the LF and HF in OC-61. A similar result was found in BR-05 with P concentration; also, a significant reduction in K concentration in HF compared to C was found (Table 1).

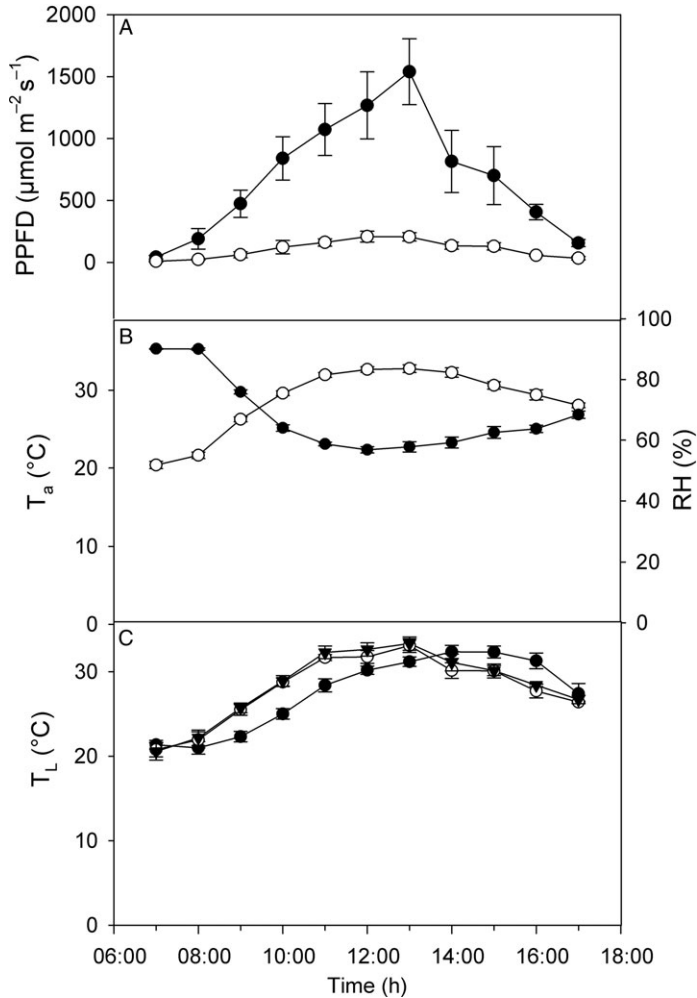


Figure 2. Time course of changes in microclimatic parameters during the experiment: A, photosynthetic photon flux density outside (●) and inside the greenhouse (○); B, air temperature and relative humidity; and C, leaf temperature in the greenhouse. Control plants (●), low frequency (○), and high frequency (▼). Values are mean \pm SE (n = 5).

Gas exchange

In both cocoa clones, significant changes in gas exchange variables due to treatment were observed (Figure 3). An increase of 16 and 20% in A was found in OC-61 in LF and HF, respectively, whereas in BR-05 increases of 56 (LF) and 60% (HF) on day 35 were observed. At day 35, clone OC-61 showed a 26% reduction in E in LF and HF compared to C, while in BR-05, E decreased by 23 and 36% in HF and LF, respectively. Stomatal conductance had a similar trend for E; in OC-61, a decline of g_s of 11 and 37% was found in HF and LF, respectively; whereas in BR-05, a 22% decrease in HF was observed compared to C plants. This resulted in an increase in WUE by 42% in LF in OC-61, while BR-05 showed an increase in WUE of 100% in HF and LF.

Photosynthetic pigments and total soluble protein

No significant differences in photosynthetic pigment concentration or in Chl_{a+b}/C_{x+c} ratio were observed between treatments in any clone. A significant increase of 15 and 54% of TSP was

Table 1. Changes in leaf and soil nutrients concentrations of cocoa clones OC-61 and BR-05, control plants (C), low frequency (LF), and high frequency (HF) of spray with SiO₂-NF, after 35 days of treatment. Values are mean ± standard error (n = 7). Different letters indicate significant differences among treatments at the end of the trial (one-way ANOVA, *p* < 0.05 and Duncan's post hoc test)

NUTRIENT CONCENTRATION (mmol Kg ⁻¹)	CLONE	TREATMENT		
		C	LF	HF
Leaf				
N	OC-61	951.9 ± 88 a	1237.2 ± 82 b	1249.1 ± 44 b
	BR-05	1071.3 ± 29 a	1311.4 ± 83 b	1180.9 ± 50 b
P	OC-61	66.5 ± 3 a	65.5 ± 6 a	80.3 ± 3 b
	BR-05	74.4 ± 3 a	80.8 ± 5 a	96.7 ± 11 a
K	OC-61	476.2 ± 37 a	476.3 ± 64 a	546.2 ± 54 a
	BR-05	486.5 ± 48 a	670.2 ± 65 b	599.7 ± 52 b
Mg	OC-61	385.3 ± 7 a	378.2 ± 22 a	396.9 ± 21 a
	BR-05	377.9 ± 31 a	398.3 ± 25 a	406.4 ± 15 a
Ca	OC-61	325.3 ± 13 a	295.4 ± 21 a	358.2 ± 41 a
	BR-05	291.2 ± 22 a	334.2 ± 21 a	414.4 ± 66 a
Si (mg g ⁻¹)	OC-61	723.8 ± 54 a	637.9 ± 21 a	749.3 ± 58 a
	BR-05	415.1 ± 33 a	906.9 ± 19 b	742.6 ± 84 b
Soil				
N	OC-61	38.8 ± 2 a	36.6 ± 3 a	51.2 ± 7 a
	BR-05	38.4 ± 1 a	39.9 ± 4 a	38.7 ± 2 a
P	OC-61	17.3 ± 1 a	18.1 ± 2 a	19.9 ± 2 a
	BR-05	18.8 ± 1 ab	21.9 ± 2 b	14.0 ± 1 a
K	OC-61	165.1 ± 17 a	168.8 ± 6 a	144.1 ± 1 a
	BR-05	175.8 ± 20 b	179.7 ± 10 b	141.8 ± 37 a
Mg	OC-61	332.4 ± 17 a	337.6 ± 6 a	327.1 ± 1 a
	BR-05	333.9 ± 20 a	333.4 ± 10 a	253.8 ± 37 a
Ca	OC-61	150.2 ± 2 ab	155.2 ± 3 b	139.1 ± 4 a
	BR-05	160.5 ± 5 a	160.1 ± 4 a	123.6 ± 25 a
Si (mg g ⁻¹)	OC-61	372.2 ± 20 a	383.7 ± 40 a	374.6 ± 39 a
	BR-05	359.5 ± 53 a	413.8 ± 45 a	424.7 ± 52 a

observed in OC-61 in LF and HF, respectively; while in BR-05, an increase of 26% and 33% in HF and LF, respectively, was found (Table 2).

Specific leaf area

Clone OC-61 showed significantly lower values of SLA in LF and HF compared to C but no change was observed in BR-05 (Table 2).

Photochemical activity

Clone OC-61 on average had higher F_v/F_m (0.81 ± 0.002 HF, 0.81 ± 0.01 BF and 0.78 ± 0.005 C) compared to clone BR-05 (0.79 ± 0.006 HF, 0.80 ± 0.007 LF, 0.77 ± 0.010 C). Significant changes in variables of chlorophyll *a* fluorescence were found between clones and across treatments (Figure 4). Clone OC-61 showed a significant increase of 44% in *J* in HF compared to LF and C at the end of the experiment; whereas BR-05, showed an increase of 162 and 94% in *J* in LF and HF, respectively. In OC-61, an increase of 70% in q_p was observed in HF compared to LF and C. In BR-05, an increase in q_p was found of 221 and 45% in LF and HF compared to C. High values of q_N in both cocoa clones was observed throughout the experiment, and a slight decrease of q_N in HF and LF compared to C in BR-05 was found.

Growth

In both cocoa clones, neither *H* nor H_G of plants sprayed with SiO₂-NF were affected significantly, but there was a significant increase of 27 and 32% of D_G in clone OC-61 in LF and HF, respectively, and 24% increase in BR-05 in LF with no change in HF compared to C (Table 3).

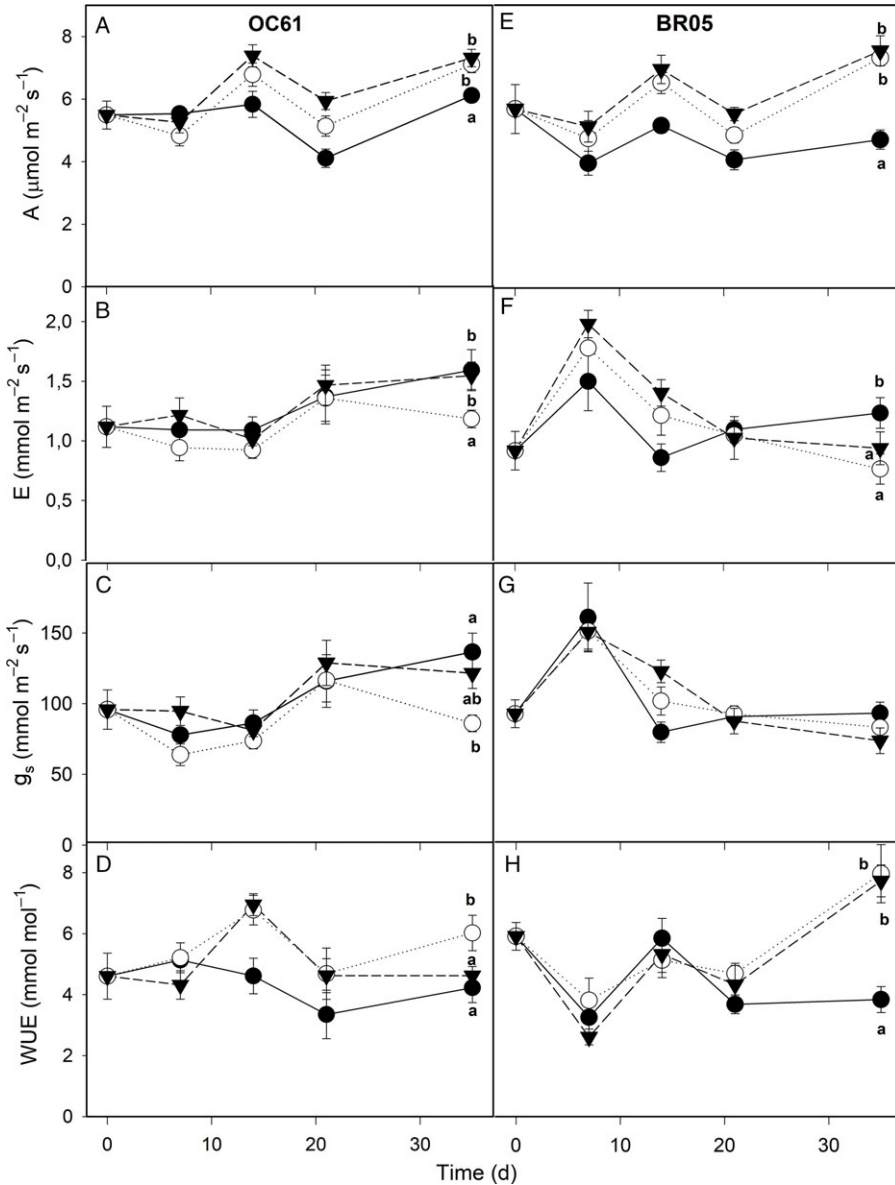


Figure 3. Time course of changes in gas exchange in cocoa clones subjected to low frequency (○) and high frequency (▼) of spraying with SiO₂-NF and in control plants (●); A–E, net photosynthetic rate; B–F, transpiration rate; C–G stomatal conductance; and D–H, water use efficiency. Values are mean ± SE (n = 7). Clone name indicated in the center panels.

Photosynthetic N and P use efficiency

A significant reduction in PNUE was found in OC-61 after 35 of LF and HF treatment, whereas in BR-05 an increase was observed. The PPUE remaining unchanged under treatment in both cocoa clones (Table 4).

Discussion

Foliar fertilization with the SiO₂-NF was beneficial for the Criollo cocoa clones. Mainly, SiO₂-NF caused positive effects in physiological processes, resulting in an improvement in the

Table 2. Changes in the total chlorophyll (Chl_{a+b}), carotenoids content (C_{x+c}), chlorophyll to carotenoids ratio (Chl_{a+b}/C_{x+c}), total soluble proteins (TSP) content, and specific leaf area (SLA) of cocoa clones OC-61 and BR-05, under three treatments: control plants (C), low frequency (LF), and high frequency (HF) of spray with SiO_2 -NF, after 35 days of treatment. Values are mean \pm standard error ($n = 7$). Different letters indicate significant differences among treatments at the end of the trial (one-way ANOVA, $p < 0.05$ and Duncan's post hoc test)

Variable	Clone	TREATMENT		
		C	LF	HF
Chl_{a+b} ($\mu g\ cm^{-2}$)	OC-61	72.53 \pm 8.0 a	68.28 \pm 8.7a	77.75 \pm 6.3 a
	BR-05	54.35 \pm 6.1 a	70.92 \pm 6.4 a	63.57 \pm 5.3 a
C_{x+c} ($\mu g\ cm^{-2}$)	OC-61	10.79 \pm 1.7 a	10.50 \pm 1.1 a	11.69 \pm 1.4 a
	BR-05	9.34 \pm 0.9 a	12.15 \pm 1.0 a	10.77 \pm 0.9 a
Chl_{a+b}/c_{x+c}	OC-61	7.05 \pm 0.4 a	6.60 \pm 0.4 a	6.87 \pm 0.4 a
	BR-05	5.78 \pm 0.2 a	5.83 \pm 0.2 a	5.95 \pm 0.2 a
TSP ($\mu g\ cm^{-2}$)	OC-61	18.5 \pm 1.6 a	21.3 \pm 1.5 a	28.5 \pm 2.9 b
	BR-05	18.2 \pm 1.6 a	24.2 \pm 1.6 b	22.9 \pm 1.5 b
SLA ($cm^2\ g^{-1}$)	OC-61	287.2 \pm 11.3 c	207.6 \pm 7.9 a	241.8 \pm 5.7 b
	BR-05	234.0 \pm 8.9 a	222.4 \pm 5.6 a	230.9 \pm 8.5 a

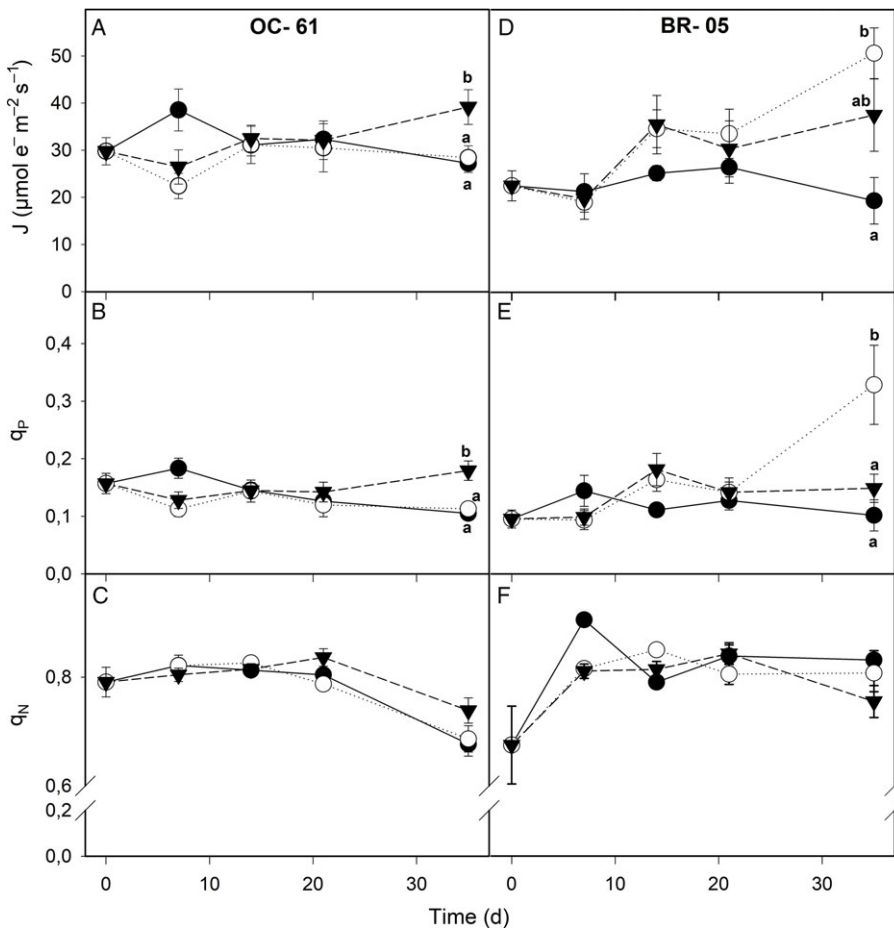


Figure 4. Time course of changes in variables of chlorophyll a in plant of cocoa clones subjected to low frequency (○) and high frequency (▼) of spraying with SiO_2 -NF and in control plants (●): A–D, electron transport rate; B–E, photochemical quenching coefficients; and C–F, non-photochemical quenching coefficients. Values are mean \pm SE ($n = 7$). Clone name indicated in the center of the panels.

Table 3. Time-course of growth variables of plants of cocoa clones (OC-61 and BR-05) subjected to different treatments in the greenhouse: height (H), height of the main branch of graft (H_G), and diameter of graft (D_G). Control (C), low frequency (LF), and high frequency (HF) of spray with SiO_2 -NF. Values are mean \pm standard error ($n = 4$). Different letters indicate significant differences among treatments for each day (one-way ANOVA, $p < 0.05$ and Duncan's post hoc test)

Clone	Day	GROWTH VARIABLES								
		H (cm)			H_G (cm)			D_G (cm)		
		C	LF	HF	C	LF	HF	C	LF	HF
OC-61	14	50.9 \pm 1 a	62.4 \pm 8 a	56.9 \pm 11 a	34.1 \pm 2 a	45.4 \pm 6 a	39.4 \pm 13 a	0.68 \pm 0.02 a	0.82 \pm 0.03 ab	0.91 \pm 0.07 b
	21	58.7 \pm 4 a	64.0 \pm 8 a	59.0 \pm 13 a	45.7 \pm 2 a	47.5 \pm 6 a	47.7 \pm 14 a	0.83 \pm 0.01 a	0.97 \pm 0.03 ab	0.95 \pm 0.07 b
	35	64.5 \pm 1 a	67.4 \pm 5 a	62.0 \pm 15 a	53.7 \pm 1 a	55.1 \pm 4 a	49.5 \pm 14 a	0.82 \pm 0.02 a	1.04 \pm 0.09 b	1.08 \pm 0.11 b
BR-05	14	46.4 \pm 12 a	59.1 \pm 4 a	52.9 \pm 5 a	32.1 \pm 13 a	43.5 \pm 2 a	38.1 \pm 3 a	0.69 \pm 0.09 a	0.76 \pm 0.1 a	0.76 \pm 0.04 a
	21	61.9 \pm 5 a	62.4 \pm 5 a	53.3 \pm 7 a	45.3 \pm 4 a	45.7 \pm 1 a	41.1 \pm 4 a	0.80 \pm 0.09 a	0.99 \pm 0.2 b	0.84 \pm 0.0 a
	35	67.0 \pm 3a	62.8 \pm 5 a	56.8 \pm 3 a	54.8 \pm 3 a	48.0 \pm 3 a	45.3 \pm 4 a	0.91 \pm 0.06 a	1.02 \pm 0.1 a	0.90 \pm 0.04 a

Table 4. Photosynthetic N and P use efficiency (PNUE, PPUE) of cocoa clones (OC-61 and BR-05) in control plants (C) and subjected to low frequency (LF) and high frequency of spraying (HF) after 35 days of treatment. The values represented are averages \pm SE ($n = 7$ per treatment). The letters indicate significant differences obtained among treatments on day 35 through a one-way ANOVA ($P < 0.05$) and a post hoc Duncan test

Variable	Clone	TREATMENT		
		C	LF	HF
PNUE ($\mu\text{mol CO}_2 [\text{mol N}]^{-1} \text{s}^{-1}$)	OC- 61	196 \pm 15 b	131 \pm 13 a	161 \pm 7 ab
	BR- 05	112 \pm 8 a	129 \pm 8 ab	166 \pm 19 b
PPUE ($\mu\text{mol CO}_2 [\text{mol P}]^{-1} \text{s}^{-1}$)	OC- 61	2.7 \pm 0.2 a	2.5 \pm 0.3 a	2.5 \pm 0.2 a
	BR- 05	1.7 \pm 0.1 a	2.1 \pm 0.1 a	2.1 \pm 0.2 a

photosynthetic capacity in both cocoa clones (significant increases in A, J, and q_p), mostly possibly related to an increase in leaf nutrient concentration, such as N and P, and soluble protein content. Also, spraying of SiO₂-NF caused an increase in WUE as a result of both increase in A and reductions in g_s and E. The significant increase in A and leaf nutrient concentration resulted in a greater diameter of the main stem of the graft, suggesting that plants could be more vigorous by the time they would be transplanted to the field. The physiological response to SiO₂-NF was different between clones, with clone BR-05 having a better physiological performance, given the high percentages of increases in many of the variables studied.

Leaf and soil nutrients concentration

The values of foliar N, P, K, and Mg concentration obtained in cocoa clones OC-61 and BR-05 coincide with some of the nutritional proposals for different cocoa clones (Puentes-Páramo *et al.*, 2016). In comparison with the most recent reports of nutritional of the leaves (Puentes-Páramo *et al.*, 2016), clones OC-61 and BR-05 had higher values of P, K, and lower values of Ca. The differences in foliar nutrient concentration may be due mainly to the type of substrate, age of the plant, leaf age, and duration of the experiment and the specific nutritional requirement for each clone (Puentes-Páramo *et al.*, 2016).

In OC-61, a significant increase in leaf N concentration was obtained for LF and HF treatments compared to C; also, in the leaf P concentration in HF treatment; while in BR-05, there was a significant increase in leaf N and K concentration in LF and HF treatment. Those increases were most probably due to the source of external nutrient incorporation (sprayed SiO₂-NF), since the samples of their respective soils presented very low N and P concentration; in the case of K for clone BR-05, it may be due to the application of SiO₂-NF and, in turn, to the constant incorporation of K through irrigation, because irrigation water presented high concentrations of K, Mg, and Ca. The critical soil concentration for cocoa plants are below 42.8 mmol kg⁻¹ N, 0.3 mmol kg⁻¹ P, 0.3 mmol kg⁻¹ K, 8 mmol kg⁻¹ Mg, and 22.5 mmol kg⁻¹ Ca (Aikpokpodion, 2010), therefore, the concentration of nutrients (N, P, K, Mg, and Ca) obtained in the soils on day 0 and day 35 for both clones were suitable for cocoa seedlings.

Gas exchange

The average A values obtained in OC-61 and BR-05 agreed with those previously reported in different varieties and cocoa clones in Venezuela and elsewhere (A between 0.5 and 7 $\mu\text{mol m}^{-2} \text{s}^{-1}$) (Ávila-Lovera *et al.*, 2016, 2021; Bertolde *et al.*, 2012; Daymond *et al.*, 2011; De Almeida *et al.*, 2018; Tezara *et al.*, 2016, 2020). The results observed in A coincide with the increase in leaf N and P concentration due to treatment. The suitable supply of nutrients to the plants certainly provided the necessary requirements for specific processes of the metabolism during the development of the plants (Gattward *et al.*, 2012; Latsague *et al.*, 2014) that resulted in increased photosynthetic capacity and

growth of the main stem of the graft. Furthermore, this increased in A was not due to increases in chlorophyll and total carotenoid concentration because no variations due to treatment in their concentrations were observed in any of the clones. Contrary to our results, Chl *a*, Chl *b*, and C_{x+c} concentrations in *Berberidopsis corallina* increased in plants fertilized, contributing to enhanced A (Latsague *et al.*, 2014).

In both cocoa clones a reduction of g_s was obtained after foliar spraying with SiO₂-NF compared to C. The changes in g_s reduced water loss under unchanging Δ_w . The reduction of g_s and E in cocoa plants with foliar spraying of Si subjected to water deficit has been reported before, however, in that study, there was no improvement in photosynthetic capacity during one week of fertilization (Zanetti *et al.*, 2016). Similarly, another study found a significant reduction in E and leaf RWC during the irrigation of SiO₂-NF in wild pear seedlings, but A and g_s did not change with irrigation (Zarafshar *et al.*, 2015).

Considerable WUE enhancement was obtained in both clones after the foliar spraying with SiO₂-NF at the end of the experiment, in accordance with increased A and reduced g_s and E found here. The increase in WUE was of greater magnitude in clone BR-05 than OC-61, so the former clone appears to respond better to the application of SiO₂-NF. Therefore, SiO₂-NF could promote greater tolerance to water loss during dry periods (Ahmed *et al.*, 2016; Raliya *et al.*, 2017; Zarafshar *et al.*, 2015), an aspect of great interest for cocoa since it is a perennial crop whose productivity can be affected by drought, which is why the search for new strategies to overcome this limitation is promoted in Venezuela (De Almeida *et al.*, 2016; Tezara *et al.*, 2020).

Similar to our findings, the application of SiO₂-NF in *Solanum lycopersicum* improved photosynthetic capacity of plants under salt stress, and improved WUE by significantly reducing g_s and E (Haghighi and Pessarakli, 2013). The mitigation of the effects of abiotic stress, such as water deficit, nutrient availability, and chilling on physiological process, has been found by increasing the uptake of water through the roots, maintaining the nutrient balance, reducing E and stimulating A (Elsheery *et al.*, 2020; Wang *et al.*, 2020; Zhu and Gong, 2014). Furthermore, deposition of Si occurs on leaf epidermal cells forming a barrier that can reduce the loss of water through the cuticle (Trenholm *et al.*, 2004).

The increase in A and WUE observed in the clones studied could be due to the beneficial effects of leaf spraying of SiO₂-NF on leaf nutrient concentration, both N and P concentration, as well as the accumulation of Si in leaf tissues. These physiological responses supports the idea that nanoparticles of SiO₂ with a size of 34 nm can be used as a means of supplying and delivering nutrients to cocoa plants. Perhaps, the incorporation of nanoparticles was mediated by cuticular penetration mechanisms such as the 'polar' or 'watery pore' route (Schönherr, 2006; Schreiber and Schönherr, 2009; Schreiber, 2005), since cocoa is a plant with hypostomatic leaves (Hernández *et al.*, 2017).

Photosynthetic pigments and total soluble protein

Chlorophyll and total carotenoid contents in OC-61 and BR-05 clones were within the range of those previously reported in other cocoa clones: 30–112 $\mu\text{g cm}^{-2}$ of total chlorophyll and 5–18 $\mu\text{g cm}^{-2}$ of carotenoids (Bertolde *et al.*, 2012; De Almeida *et al.*, 2018; Tezara *et al.*, 2020). There were no differences in pigment concentrations among treatments in either clone. In contrast, in *Triticum* spp and *Lupinus albus*, a dramatic increase in the concentration of photosynthetic pigments and TSP was reported after irrigation with high concentrations of SiO₂ nanoparticles (Sun *et al.*, 2016). Additionally, it has been proposed that the concentration of photosynthetic pigments may be an indicator of nanoparticles toxicity (Rico *et al.*, 2013), a hypothesis that could be ruled out in our study since there was no negative effect of the SiO₂-NF on photosynthetic pigment concentration.

Leaf TSP content for clones OC-61 and BR-05 were similar to values previously reported for different Trinitario clones of a germplasm bank in Barlovento, Venezuela, in which values between 1.6 and 8.8 mg g^{-1} have been found (Quiñones-Gálvez *et al.*, 2015; Tezara *et al.*, 2020). Clone OC-61 presented higher values of TSP compared to BR-05, with values between 5.1

and 6.9 mg g⁻¹ in OC-61 and between 4.3 and 5.4 mg g⁻¹ in BR-05. The differences obtained between treatments and C in both clones indicate that the SiO₂-NF had a positive effect in the formation of proteins in the photosynthetic tissues of the cocoa plants. Similarly, TSP content was correlated with the increase of the photochemical and photosynthetic activity in wheat and lupin (Sun *et al.*, 2016). These results are in accordance with the increase in leaf N concentration for the clones OC-61 and BR-05 due to spraying of SiO₂-NF, since approximately 40% of the leaf N is usually invested in RubisCO (Spreitzer and Salvucci, 2002). Furthermore, the application of N, P, and K can directly influence the concentration of soluble carbohydrates (glucose), which is directly related to the synthesis of pigments and proteins (Latsague *et al.*, 2014).

Values of SLA in both cocoa clones were on average 237 cm² g⁻¹, similar to those reported in eight Forastero clones in the International Cacao Quarantine Centre at The University of Reading (Daymond *et al.*, 2011), but lower than those reported in Forastero, Trinitario, and Criollo clones in Venezuela (Tezara *et al.*, 2016, 2020). Clone OC-61 showed lower values of SLA in LF and HF, that is, leaves became thicker on a dry mass basis and/or denser, whereas no change was observed in BR-05 leaves.

Photochemical activity

The values of F_v/F_m and Φ_{PSII} obtained in this study agreed with those previously reported in different ecophysiological studies with different varieties of cocoa (Ávila-Lovera *et al.*, 2016; De Almeida *et al.*, 2018; Tezara *et al.*, 2016, 2020). Throughout the experiment, clones OC-61 and BR-05 showed F_v/F_m values of 0.80 and 0.75, respectively, indicating that there was no damage in the photochemical apparatus, and that plants were healthy (Maxwell and Johnson, 2000). In OC-61 and BR-05, the increase of J , Φ_{PSII} , and q_p without changes in q_N showed an improved photochemical activity after the treatment with SiO₂-NF. The application of SiO₂-NF in cocoa plants could have generated a decrease in the emission of fluorescence due to a better stabilization of the photosystems in the membrane structure of the chloroplasts, translating into a greater photochemical activity (Zanetti *et al.*, 2016).

It has been proposed that Si has an important role in the synthesis of intracellular organic compounds and the maintenance of normal biochemical functions, such as the biosynthesis and degradation of chlorophyll, suggesting that the stimulation of the expression of some genes with the addition of Si can improve the activity of PSII and J , due to an increase in chlorophyll concentration (Li *et al.*, 2015; Matchenkov *et al.*, 2008; Song *et al.*, 2014). Silicon can improve photochemical efficiency in a variety of species subject to water deficit (Chen *et al.*, 2011), because the fluorescence emission of Chl *a* depends on the structures of the thylakoids membrane, which allows for a more stable anchoring of the photosynthetic pigments (Zanetti *et al.*, 2016). Cocoa plants subjected to water deficit after the foliar spraying of non-nanostructured SiO₂ showed improvements in photochemical efficiency by increasing the density of active PSII reaction centers and the index of performance for the conservation of energy until the reduction of the final acceptor of PSII (Zanetti *et al.*, 2016).

Possible mechanism how SiO₂-NF increased photosynthesis

A possible mechanism of how SiO₂-NF foliar spray increased photosynthesis and WUE could be that the cocoa plants nanofertilized with mineral nutrients encapsulated in the nanoparticles, which were released in the foliar tissues, caused a higher incorporation of nutrients (Monreal *et al.*, 2015) that were probably used in the synthesis of proteins (higher concentration of TSP) or other enzymes that intervene in the Calvin–Benson cycle, or in protein complexes associated with the thylakoid membrane, all factors that enhance carboxylation efficiency and therefore A , q_p , and J , without the need to increase photosynthetic pigment concentrations (Elsheery *et al.*, 2020; Morales-Díaz *et al.*, 2017; Raliya *et al.*, 2017; Wang *et al.*, 2020). Furthermore, a dense layer formed by the deposition of Si can be found on the cuticle in many species. A formation of this

layer forming a barrier between leaves and the atmosphere, could have reduced the loss of water through the cuticle; these factors are essential to cope with abiotic stress conditions, and are suggested here as a mechanism for Si-induced improvement of WUE.

Growth in both cocoa clones

Among the growth parameters that were considered in this experiment, only a significant increase in D_G of both cocoa clones was observed. This increase may have been due to both increased in leaf nutrients and photosynthesis, as well as increased in Si concentration. In *Poa pratensis*, the accumulation of Si leads to hardness, can facilitate the erection of the leaves, and, therefore, increase the interception of light, which could contribute to the increase of photosynthetic capacity, photo-assimilation, and growth, which then improve overall plant performance (Saud *et al.*, 2014). Similarly, it has been reported in different species, such as *S. lycopersicum*, *Triticum* spp and *L. albus*, that after the application of SiO_2 -NF without fertilizer formulation, the dry mass of tomato plants increased, and root tissues and shoot dry mass increases by 51 and 26%, respectively, for *Triticum* spp and *L. albus* (Haghighi and Pessarakli, 2013; Raliya *et al.*, 2017; Sun *et al.*, 2016). Also, it has been reported in rice plants that Si irrigation could increase biomass and promote the phytolith concentration (derived from the biomineralization of Si), although in enriched soils, excess Si may cause reduction in biomass (Sun *et al.*, 2016).

The SiO_2 -NF can be an attractive agro-input for cocoa farmers and producers, since it could be applied as a complement to stimulate the photosynthetic apparatus, growth, and tolerance of cocoa plants against water deficit during their most important developmental stage, and without generating negative impacts on the environment. However, future tests with mature cocoa trees under field conditions are recommended to assess whether SiO_2 -NF spraying can improve important agronomic characteristics and yield in different cocoa clones.

Photosynthetic N and P use efficiency

All of the photochemical and biochemical process of photosynthesis involve nitrogen: RubisCO, regeneration of RuBP, chlorophylls, and thylakoids proteins (Ávila-Lovera *et al.*, 2016). The values observed in PNUE were 4–5 times higher than those calculated with data from A and N reported in other cocoa plants (Ávila-Lovera *et al.*, 2016, 2021; Tezara *et al.*, 2020; Daymond *et al.*, 2011). In our study, there was a differential PNUE response to spray with SiO_2 -NF, it increased in BR-05 and decreased in OC-61, suggesting that the BR-05 had a better use of nitrogen, i.e. a greater A per unit of foliar N. Contrary, PPUE was not affected by the treatment in either clone.

Conclusions

Foliar fertilization with SiO_2 -NF was beneficial for both Criollo cocoa clones (OC-61 and BR-05). Foliar spraying of SiO_2 -NF improved leaf gas exchange and photochemical activity of PSII by increasing net photosynthetic rate, electron transport rate, and the photochemical extinction coefficient of both cocoa clones. Spraying with SiO_2 -NF promoted higher concentration of TSPs, higher leaf N and P concentration in clone OC-61, and higher N, K, and Si in clone BR-05. This increased nutrient concentration may have increased both photosynthetic rate and photochemical activity of PSII. The improved water use efficiency was achieved by simultaneous increases in net photosynthetic rate and reduction in stomatal conductance and transpiration rates in both clones, without changes in the concentration of photosynthetic pigments. The significant increase in photosynthesis and leaf nutrient concentration resulted in a greater diameter of the main stem of the graft, suggesting that plants could be more vigorous with a better opportunity of surviving by the time they would be transplanted to the field.

We conclude that SiO_2 -NF has a positive effect on physiological processes related to photosynthesis in both cocoa clones studied, and this was associated with an increase of nutrients

(N, P, and K) and Si concentration, which translates into greater growth (i.e., increase diameter of graft). Therefore, the SiO₂-NF can be a useful tool for cocoa farmers, since they could be applied as a complement to stimulate the photosynthetic apparatus and growth during their most important stage of development, that is, the seedling stage.

A differential physiological response to spraying with SiO₂-NF between clones was found, likely the great genetic variability among cocoa may determine different physiological responses to the foliar application of SiO₂-NF. Clone BR-05 was the clone with a better physiological performance, given the high percentages of increases in many of the variables studied, and hence one that we would recommend for further field experimentation. However, more research on the effect of SiO₂-NFs on adult cocoa trees grown in field conditions is necessary.

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References

- Ahmed M., Qadeer U., Ahmed Z.I. and Hassan F. (2016). Improvement of wheat (*Triticum aestivum*) drought tolerance by seed priming with silicon. *Archives of Agronomy and Soil Science* **62**, 299–315.
- Aikpokpodion P.E. (2010). Nutrients dynamics in cocoa soils, leaf and beans in Ondo State, Nigeria. *Journal of Agricultural Science* **1**, 1–9.
- Ávila-Lovera E., Blanco H., Móvil O., Santiago L.S. and Tezara W. (2021). Shade tree species affect gas exchange and hydraulic conductivity of cacao cultivars in an agroforestry system *Tree Physiology* **41**, 240–253.
- Ávila-Lovera E., Coronel I., Jaimez R., Ulrich R., Pereyra G., Araque O., Chacón I. and Tezara W. (2016). Ecophysiological traits of adult trees of Criollo cocoa cultivars (*Theobroma Cacao* L.) from a germplasm bank in Venezuela. *Experimental Agriculture* **52**, 137–153.
- Bertolde F.Z., Almeida A.F., Pirovani C.V., Gomes F.P., Ahnert D., Baligar V.C. and Valle R.R. (2012). Physiological and biochemical responses of *Theobroma cacao* L. genotypes to flooding. *Photosynthetica* **50**, 447–457.
- Bradford M. (1976). A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry* **72**, 248–254.
- Chen W., Yao X., Cai K. and Chen J. (2011). Silicon alleviates drought stress of rice plants by improving plant water status, photosynthesis and mineral nutrient absorption. *Biological Trace Element Research* **142**, 67–76.
- Daymond A., Tricker P.J. and Hadley, P. (2011). Genotypic variation in photosynthesis in cacao is correlated with stomatal conductance and leaf nitrogen. *Biologia Plantarum* **55**, 99–104.
- De Almeida J., Herrera A. and Tezara W. (2018). Phenotypic plasticity to photon flux density of physiological, anatomical and growth traits in a modern Criollo cocoa clone. *Physiologia Plantarum* **166**, 821–832.
- De Almeida J., Tezara W. and Herrera, A. (2016). Physiological responses to drought and experimental water deficit and waterlogging of four clones of cocoa (*Theobroma cacao* L.) selected for cultivation in Venezuela. *Agricultural Water Management* **171**, 80–88.
- Elsheery N., Sunoj V.S.J., Wen Y., Zhu J.J., Muralidharan G. and Cao K.F. (2020). Foliar application of nanoparticles mitigates the chilling effect on photosynthesis and photoprotection in sugarcane. *Plant Physiology and Biochemistry* **149**, 50–60.
- Fernández V., Sotiropoulos T. and Brown P.H. (2015). *Fertilización foliar: principios científicos y práctica de campo*. Melgar R. and Fernández V. (eds), Francia: Asociación Internacional de la Industria de Fertilizantes IFA, pp. 13–31.
- Gattward J.N., Almeida A.–A.F., Souza J.O., Gomes F.P. and Kronzucker H.J. (2012). Sodium-potassium synergism in *Theobroma cacao*: stimulation of photosynthesis, water-use efficiency and mineral nutrition. *Physiologia Plantarum* **146**, 350–362.
- Genty B., Briantais J.M. and Baker N.R. (1989). The relationship between the quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. *Biochimica et Biophysica Acta* **990**, 87–92.

- Haghighi M. and Pessaraki M. (2013). Influence of silicon and nano-silicon on salinity tolerance of cherry tomatoes (*Solanum lycopersicum* L.) at early growth stage. *Scientia Horticulturae* **161**, 111–117.
- Hernández C.E., Leiva R. and Ramírez R. (2017). Dinámica estomática en cacao (*Theobroma cacao* L.). *International Symposium on Cocoa Research*, 1–6. Perú: ICCO.
- International Cocoa Organization ICCO. (2020). Quarterly Bulletin of Cocoa Statistics, year 2019/2020. Available at <https://www.icco.org/statistics/other-statistical-data.html>.
- Jackson M.L. (1958). *Soil chemical analysis*. Englewood NJ: Verlag: Prentice Hall, Inc., (eds), 183.
- Latsague M., Sáez P. and Mora M. (2014). Efecto de la fertilización con nitrógeno, fósforo y potasio, sobre el contenido foliar de carbohidratos, proteínas y pigmentos fotosintéticos en plantas de *Berberidopsis corallina* Hook.f. *Gayana Botánica* **71**, 37–42.
- Li P., Song A., Li Z.J., Fan F.L. and Liang Y.C. (2015). Silicon ameliorates manganese toxicity by regulating both physiological processes and expression of genes associated with photosynthesis in rice (*Oryza sativa* L.). *Plant Soil* **397**, 289–301.
- Matichenkov V.V., Bocharnikova E.A., Kosobryukhov A.A. and Biel K.Y. (2008). Mobile forms of silicon in plants. *Doklady Biological Sciences* **418**, 39–40.
- Maxwell K. and Johnson G. (2000). Chlorophyll fluorescence a practical guide. *Journal of Experimental Botany* **51**, 659–668.
- Monreal C.M., DeRosa M., Mallubhotla S.C., Bindraban P.S. and Dimkpa C. (2015). Nanotechnologies for Increasing the Crop Use Efficiency of Fertilizer-Micronutrients. *Biology and Fertility of Soils* **52**, 423–437.
- Morales-Díaz A.B., Ortega-Ortíz H., Juárez-Maldonado A., Cadenas-Pliego G., González-Morales S. and Benavides-Mendoza A. (2017). Application of nanoelements in plant nutrition and its impact in ecosystems. *Advances in Natural Sciences: Nanoscience and Nanotechnology* **8**, 1–13.
- Motamayor J.C., Lachenaud P., Wallace J., Loo R., Kuhn D., Steven J. and Schnell R. (2008). Geographic and Genetic Population Differentiation of the Amazonian Chocolate Tree (*Theobroma cacao* L.). *PLoS One* **3**, 1–8.
- Motamayor J.C., Risterucci A.M., Lopez P.A., Ortiz C.F., Moreno A. and Lanaud C. (2002). Cacao domestication I: the origin of the cacao cultivated by the Mayas. *Heredity* **89**, 380–386.
- Murphy J. and Riley J.P. (1962). A modified single method for the determination of phosphate in natural waters. *Analytica Chimica Acta* **27**, 31–36.
- Puentes-Páramo Y.J., Menjivar-Flores J.C. and Aranzazu-Hernández F. (2016). Concentración de nutrientes en hojas, una herramienta para el diagnóstico nutricional en Cacao. *Agronomía Mesoamericana* **27**, 329–336.
- Quiñones-Gálvez Q., Sosa D., Demey J.R., Aleman S., Sosa M., Parra D., Móvil O., Trujillo R., Capdesuñer Y., Quirós Y., Hernández M. and Infante D. (2015). Caracterización bioquímica de hojas de clones de *Theobroma cacao* y su relación con los tricomas. *Revista Colombiana de Biotecnología* **17**, 33–43.
- Raliya R., Saharan V., Dimkpa C. and Biswas P. (2017). Nanofertilizer for precision and sustainable agriculture: current state and future perspectives. *Journal of Agricultural and Food Chemistry* **66**, 6487–6503.
- Ram P., Kumar R., Rawat A., Pandey P. and Singh V.P. (2018). Nanomaterials for efficient plant nutrition. *International Journal of Chemistry* **6**, 867–871.
- Rico C.M., Hong J., Morales M.I., Zhao L.J., Barrios A.C., Zhang J.Y., Peralta-Videa J.R. and Gardea-Torresdey J.R. (2013). Effect of cerium oxide nanoparticles on rice: a study involving the antioxidant defence system and in vivo fluorescence imaging. *Environmental Science & Technology* **47**, 5635–5642.
- Saud S., Li X., Chen Y., Zhang L., Fahad S., Hussain S., Sadiq A. and Chen Y. (2014). Silicon application increases drought tolerance of Kentucky bluegrass by improving plant water relations and morphophysiological functions. *The Scientific World Journal* **2014**, 1–10.
- Schönherr J. (2006). Characterization of aqueous pores in plant cuticles and permeation of ionic solutes. *Journal of Experimental Botany* **57**, 2471–2491.
- Schreiber L. (2005). Polar paths of diffusion across plant cuticles: new evidence for an old hypothesis. *Annals of Botany* **95**, 1069–1073.
- Schreiber L. and Schönherr J. (2009). *Water and solute permeability of plant cuticles: Measurement and data analysis*. Germany: Springer Verlag, pp. 1–28.
- Song A., Li P., Fan F.L., Li Z.J. and Liang Y.C. (2014). The effect of silicon on photosynthesis and expression of its relevant genes in rice (*Oryza sativa* L.) under high zinc stress. *PLoS One* **9**, 1–21.
- Spreitzer R.J. and Salvucci M.E. (2002). RuBisCO: structure, regulatory interactions, and possibilities for a better enzyme. *Annual Review of Plant Biology* **53**, 449–475.
- Sun D., Hussain H.I., Yi Z., Rookes J.E., Kong L., Cahill D.M. (2016). Mesoporous silica nanoparticles enhance seedling growth and photosynthesis in wheat and lupin. *Chemosphere* **152**, 81–91.
- Tezara W., Pereyra G., Ávila-Lovera E. and Herrera A. (2020). Variability in physiological responses of Venezuelan cacao to drought. *Experimental Agriculture* **56**, 407–421.
- Tezara W., Urich R., Jaimez R., Coronel I., Araque O., Azocar C. and Chacón I. (2016). Does Criollo cocoa have the same ecophysiological characteristics as Forastero? *Botanical Sciences* **94**, 563–574.
- Trenholm L. E., Datnoff L. E. and Nagata R. T. (2004). Influence of silicon on drought and shade tolerance of St. Augustinegrass. *HortTechnology* **14**, 487–490.

- Wang A., Jin Q., Xu X., Miao A.-J., White J.C., Gardea-Torresdey J.L., Ji R. and Zhao L.** (2020). High-Throughput screening for engineered nanoparticles that enhance photosynthesis using mesophyll protoplasts. *Journal of Agricultural and Food Chemistry* **68**, 3382–3389.
- Wellburn A.** (1994). The spectral determination of chlorophylls a and b, as well as total carotenoids, using various solvents with spectrophotometers of different resolution. *Journal of Plant Physiology* **144**, 307–313.
- Wu Z.Y., Wang H.J., Zhuang T.T., Sun L.B., Wang Y.M. and Zhu J.H.** (2008). Multiple functionalization of mesoporous silica in one-pot: direct synthesis of aluminum-containing plugged SBA-15 from aqueous nitrate solutions. *Advanced Functional Materials* **18**, 82–94.
- Zanetti L.V., Milanez C.R.D., Gama V.N., Aguilar M.A.G., Souza C.A.S., Campostrini E., Ferraz T.M. and Figueiredo F.A.M.M.A.** (2016). Leaf application of silicon in young cacao plants subjected to water deficit. *Pesquisa Agropecuária Brasileira* **51**, 215–223.
- Zarafshar M., Akbarinia M., Askari H., Hosseini S.H., Rahaie M. and Struve D.** (2015). Toxicity assessment of SiO₂ nanoparticles to pear seedlings. *International Journal of Nanoscience and Nanotechnology* **11**, 13–22.
- Zhu Y. and Gong H.** (2014). Beneficial effects of silicon on salt and drought tolerance in plants. *Agronomy for Sustainable Development* **34**, 455–472.