Late Triassic plant microfossils from Miers Bluff Formation of Livingston Island, South Shetland Islands, Antarctica

OUYANG SHU1, DENG XI-GUANG2, SHEN YAN-BIN1, ZHENG XIANG-SHEN2 and LIU XIAO-HAN2

¹Nanjing Institute of Geology and Palaeontology, Chinese Academy of Sciences, Nanjing 210008, China ²Beijing Institute of Geology, Chinese Academy of Sciences, Beijing 100029, China

Abstract: A moderately diverse assemblage of plant microfossils has been recovered from the Johnsons Dock Member of the Miers Bluff Formation, Livingston Island, including spores, pollen, acritarchs, wood fragments and cuticles. Containing a total of c. 45 of miospore taxa, the palynoflora is dominated by non-striate bisaccate pollen, but spores of pteridophytes and pollen of gymnosperms are proportionate in diversity. The palynoflora is similar in composition to those indicative of subzones C+D of the *Alisporites* Zone of Antarctica, and the upper *Craterisporites rotundus* Zone and the lower *Polycingulatisporites crenulatus* Zone of New Zealand, suggesting a Late Triassic (possibly Norian–Rhaetian) age. This determination is supported by the sporadic occurrence of *Aratrisporites* and *Classopollis* in the palynoflora as well as by the absence of striate bisaccate grains.

Received 19 August 1999, accepted 22 March 2000

Key words: Livingston Island, Miers Bluff Formation, palynofloras, plant microfossils, Late Triassic

Introduction

The plant microfossils considered here were recovered from the Miers BluffFormation on Livingston Island, South Shetland Islands, West Antarctica. This formation and its assumed equivalents, such as the Trinity Peninsula Group (TPG) (the Hope Bay, Cape Legoupil and Paradise Harbour Formations) on Graham Land of the northern Antarctic Peninsula, and the Greywacke-Shale Formation (GSF) in the South Orkney Islands, have been usually regarded as "basement rocks" in these areas and interpreted as submarine fan deposits (Arche et al. 1992, Doktor et al. 1994, Smellie et al. 1995, Andreis et al. 1997). The successions are mainly composed of deformed and metamorphosed greywackes, mudstones/shales and sandstones, attaining a thickness of 1000-3000 m. Their geological ages generally have been poorly constrained, ranging from late Palaeozoic to early Mesozoic (Carboniferous-Permian, Permian-Triassic, Early Triassic or pre-Jurassic), because fossils in the turbidites are mostly sparse and too poorly preserved to be identified. Some plant remains have been found in the Miers Bluff Formation on Livingston Island, but their poor preservation has prevented precise identification; (Taylor & Taylor 1989, Arche et al. 1992) suggested a post-Carboniferous, possibly Mesozoic age. Grikurov & Dibner (1968) reported Early and Middle Carboniferous miospores from the Hope Bay Formation of the Trinity Group, but the spores were reasonably doubted by some authors to have resulted from laboratory contamination (Truswell 1991, Playford 1989). Later palynological investigation indicated that the Hope Bay Formation is devoid of sporomorphs (Birkenmajer 1992), but, a few marine fossils obtained from these and related formations in several localities

have provided evidence for dating the rocks. For instance, poorly preserved bivalves, e.g. ?Myalinella sp., Backevellia (Backvelloides) aff. hekiensis, and Neoschizodus halperni have been found from the Cape Legoupil Formation at Cape Legoupil, northern Antarctic Peninsula and inferred to be Triassic in age (Thomson 1975). A few radiolarians and conodonts (Neogondoella sp.) recovered from the GSF indicate a Late Triassic age (Dalziel et al. 1981). This type of lithofacies also extends to Potranca Islands (45°28'S, 74°09'W). the Chonos Archipelago, southern Chile, i.e. the Potranca Formation of the Chonos Metamorphic Complex. The Potranca Formation had been dated as Late Silurian-Early Devonian. but bivalves represented by Monotis (Entomonotis) sp. aff. M. (E.) subcircularis Gabb and Lima sp. recently found from this formation indicate a Late Triassic (Norian) age (Fang et al. 1998). These scattered fossil data demonstrate that these sets of greywacke-shale/mudstone sequences were most likely contemporaneously deposited in Late Triassic times.

The low-grade metasedimentary sequences of the Miers Bluff Formation (MBF), that crop out on the Hurd Peninsula of Livingston Island (Fig. 1), consist of up to 3000 m of mainly greywackes, mudstones, sandstones and pebbly-mudstones. According to the nomenclature of Smellie *et al.* (1995), the formation is composed of, in ascending order, the Johnsons Dock Member (*c.* 1700 m) and the Napier Peak Member (<1300 m), and the overlying Moores Peak breccias (>200 m) which "may be part of the MBF or represent a younger, unrelated unit". Doktor *et al.* (1994) restricted the MBF to the lowermost 1600 m of turbidite rocks in the west of the Hurd Peninsula, but their re-definition was challenged by Willan *et al.* (in Tokarski *et al.* 1996). The MBF mudstones yielded an Rb–Sr errorchron age of 243 ± 8 Ma and were interpreted as probably deposited in Early Triassic times (Willan *et al.* 1994), although other isochron ages (204 ± 19 Ma or 221 ± 34 Ma) have been also reported (Pankhurst 1983, Hervé 1992, *vide* Willan *et al.* 1994).

Palynological investigations of Triassic and Jurassic sediments in Antarctica have been made mainly in the Transantarctic Mountains (e.g. Gair et al. 1965, Norris 1965, Helby & McElroy 1969, Kyle 1977, Tasch 1977, Askin 1981, Tasch & Lammons 1978, Farabee et al. 1989, Shang 1997, Truswell 1991). The results were often compared with those from other parts of Gondwana, particularly Australia. Kyle (1977) proposed an informal palynological zonation for the Permian-Triassic Victoria Group from southern Victoria Land, including an Alisporites Zone with four subzones (A-D) covering the whole Triassic (the uppermost Feather Conglomerate to Lashly Formation). Two phytogeographical provinces of Middle-Late Triassic age in southern Gondwana have been recognized by Australian palynologists and others subsequent to the identification of two distinct palynofloral types by Dolby & Balme (1976): the Ipswich type which appears to have occupied higher palaeolatitudes, and the Onslow type which was roughly latitudinally controlled, reflecting a cooler temperate climate (Zavattieri & Batten 1996). Attempts to recover microfossils from the MBF on Hurd Peninsula have been unsuccessful hitherto (e.g. Doktor et al. 1994). Thus, our discovery of plant microfossils in this formation is of some importance because it provides evidence for biostratigraphic correlation and hence for assisting in tectonic and stratigraphic interpretations of the south-western margin of Gondwana prior to the break-up of Gondwana in Early or early Middle Jurassic times.



Fig. 1. Map showing sampling location.

Material and methods

More than 130 samples were collected mainly for lithological studies from the Miers Bluff Formation on the Hurd Peninsula, Livingston Island by two of the authors (Deng Xi-guang and Zheng Xiang-shen) in December 1997. Twelve of these, all are black mudstones, were selected for palynological processing. Their figure was redrawn here (Fig. 2) with the sampling horizons marked. Only one macerated sample (A97D002) is derived from another locality about 250 m of southern Johnsons Dock section.

Conventional palynological methods were used: That the samples (each 20-30 g) were digested in HC (38%) and HF (38%) and the organic residues oxidized using HNO₃ followed by treatment with alkali (KClO₃).



Fig. 2. Map showing sampling horizons in the stratigraphical section (modified from Doktor *et al.* 1994, fig. 51). A = thick sandstone, B = stratified siltstone, C = sandy mudstone, D = rhythmical sandy mudstone, E = dark mudstone, F = pebbly mudstone.

All the macerated samples but one (A97D002) yielded poorly preserved plant microfossils. These however, are comparatively diverse only in sample D021-2, and for this sample 200 specimens were counted.

Composition of the palynoflora, and identification remarks for some taxa

The plant microfossils found include moderately diverse, although generally poorly preserved spores, pollen, and a few acritarchs in association with various wood fragments (tracheids) and several cuticle remains. The preservation state of the palynomorphs makes exact species identification impossible in most cases. Some 45 taxa have been recognized; most are illustrated on Figs 3–5. [Explanation of Figs 3–5. All figures x600 unless otherwise indicated. Specimens are curated at Nanjing Institute of Geology and Palaeontology, Chinese Academy of Sciences. The sample number is followed, in brackets, by the slide number and then the coordinates.] The palynoflora is composed of the following taxa:

Spores (18 spp./17 genera): Leiotriletes? sp., Deltoidospora directa, Punctatisporites sp., Dictyophyllidites harrissi, Granulatisporites cf. G. minor, Converrucosisporites sp., Uvaesporites verrucosus, Osmundacidites cf. O. parvus, Lophotriletes cf. L. novicus, Acanthotriletes sp., Apiculatasporites sp., Nevesisporites sp., Duplexisporites? sp., Densosporites? sp., Cingulatisporites? sp., Kraeuselisporites sp., Kraeuselisporites? sp., Aratrisporites sp..

Pollen (22 spp./15 genera): Minutosaccus? sp., Vitreisporites subtilis, V. pallidus, V. microsaccus, Pityosporites sp., Accinctisporites? sp., Densipollenites? sp., Vesicaspora? sp., Alisporites cf. A. australis, A. sp., Pteruchipollenites cf. P. parvus, P. sp., Platysaccus queenslandi, Platysaccus sp., Platysaccus? sp., Podosporites sp., Araucariacites australis, Cycadopites sp. a, Cycadopites sp. b, Exesipollenites? sp., Classopollis cf. meyiana, Classopollis? sp..

Acritarchs (5 spp./5 genera): Leiosphaeridia sp., Micrhystridium sp., Schizosporis? sp., two forms unidentified

Cuticles: Type A of possibly Bennettitales, Type B of unknown affinity

Wood fragments (tracheids): Araucaroid type-Form 1-4; Abietoid type-Form 1-3; Abietoid–araucarioid transitional type-Form 1-2; Tracheid with spiral thickenings-Form 1-2; Tracheid of gymnosperm unknown affiliation; Tracheid with scalariform thickenings of unknown affiliation.

It is impossible to present here a formal description and comparison of the microfossil taxa due to limited space. Brief comparative notes on some important taxa follow.

1. Dictyophyllidites harrisii Couper, 1958 (Fig. 3.3; 35 μ m): D. harrisii is similar to D. mortoni (De Jersey) but shows thinner exine thickenings about the laesurae. The present specimen with a thin (1.5–3.0 μm broad) kyrtome thus stands closer to the former (for discussion see Playford & Dettmann 1965, p. 132; De Jersey & Hamilton 1967, p. 5, pl. 1, fig. 12). Both species are known from the Triassic and Jurassic.

2. Conversucosisporites sp. (Fig. 3.8a, 3.8b; 48 µm): The specimen is somewhat similar to *C. cameroni* (De Jersey) Playford & Dettmann 1965 recorded mainly from the Triassic in Australia (Playford & Dettmann 1965, pl. 12, figs 11–13; De Jersey 1970a, pl. 1, fig. 7; Playford *et al.* 1982, pl. 8, fig. 14), but differs from the latter in having smaller sculpture elements (coni, vertucae, short bacula).

3. Uvaesporites vertucosus (De Jersey) Helby in De Jersey 1971 (Fig. 3.13a, 3.13b; 25 μ m): Among those specific synonyms assigned to this species (De Jersey 1971, p. 6–7), the present specimen stands closest to those described and illustrated from the Bundamba Group (Aberdare Conglomerate and Raceview Formation) of Queensland (De Jersey 1970b, p. 18, pl. 6, figs 4–7) in spite of the fact that the species ranges from Middle Triassic (Moolayember Formation) to Middle Jurassic (Hutton Sandstone) in Australia (De Jersey & Hamilton 1967) or throughout the whole Triassic in Antarctica (Kyle 1977).

4. Osmundacidites cf. O. parvus De Jersey, 1962 (Fig. 3.11, 3.12, 3.16; 32–33 μ m): The specimens characterized by a mixed sculpture of small coni and grana and a trilete with labra are comparable to O. parvus De Jersey, 1962 (p.4, pl. 1, figs. 11 & 12) from the Ipswich coals, the latter (28–39 μ m), however, differs in having "granular–papillate" sculpture and in the absence of well-developed labra about the laesurae although De Jersey mentioned that in the Ipswich specimens "the laesurae are somewhat more distinct" than in O. wellmanii Couper. Osmundacidites wellmanii is larger (40–63 μ m).

5. Lophotriletes cf. L. novicus Singh, 1964 (Fig. 3.10, 27 µm): The specimen is more or less comparable to L. novicus Singh (1964, p. 247, pl. 44, figs 24–25; Foster 1979, p. 37, pl. 5, figs 1–5) from the Permian, especially that recorded from the Esk Beds of Middle Triassic age (De Jersey 1972, p. 6, pl. I, fig. 9), and it is also similar to L. rectus Bharadwaj & Salujha (see Mandal & Maithy 1981, pl.2, fig. 4) from the Upper Permian of India. Lophotriletes bauhinae De Jersey & Hamilton, 1967 differs in having larger size (35–60 µm, 40 µm on average; see Playford et al. 1982, p. 10–13).

6. Nevesisporites sp. (Fig. 3.9; $30.5 \mu m$): In general morphology, the spore is similar to *Limatulasporites limatulus* (Playford) Helby & Foster 1979 (in Foster 1979) (e.g. De Jersey 1970a, pl. 2, fig. 2), a species widely known from Triassic, however, due to the absence of a distal crassitude, it is more appropriate to assign it to *Nevesisporites*.

7. Duplexisporites? sp. (Fig. 3.15; 55 μ m): A cinguilum and 4–5 distal parallel ribs are discernible though the spore has been strongly degraded. It is somewhat similar to Contignisporites cooksoniae (Balme) Dettmann as recorded

from the Eagle Mills Formation (Carnian), east-central Texas (Traverse 1988, p. 215, fig. 11-2, j), the latter being about 50 µm in size. But typically this species has more widely spaced muri and clear proximal sculpture (Reiser & Williams 1969, p. 12) which can not be judged from our specimen due to poor preservation. The broad muri, and failure of the muri to join the equatorial cingulum, plus the union of muri (upper right in figure) seem to point to *Duplexisporites (Asseretospora)* (Ian Raine, personal communication 1999). The specimen is comparable to *Duplexisporites problematicus* (Couper) Playford & Dettmann (e.g. De Jersey 1971, p. 8, pl. 2, fig. 3; De Jersey 1972, pl. 3, fig. 1), ranging from Middle Triassic to Early Jurassic (Filatoff 1975), but the latter differs in having distal muri with a tendency of spiral arrangement.

8. Cingulatisporites? sp. (Fig. 4.4; 38 μ m): The specimen is questionably assigned to Cingulatisporites (Thompson) Potonić because it is somewhat similar to C. pallidus De Jersey, 1962 (p. 6, pl. 2, fig. 5) from the Ipswich coals in having a thin and broad cingulum (or zona?), a labrate trilete mark and small size, but differs from the latter in the absence of body ornamentation (an indistinct and incomplete distal reticulum).

9. Aratrisporites sp. (Fig. 3.18; 35 x 21 μ m): Only one specimen has been observed but its generic assignment is beyond question. In spore size, exine ornamentation, it seems to be closer to *A. wollariensis* Helby, 1967 (e.g. De Jersey 1970a, p. 11, pl. 4, figs 8–10, 12, 13) from the Lower-Middle Triassic of Australia but its poor preservation prevented an accurate specific identification.

10. Vitreisporites subtilis (De Jersey) De Jersey, 1962 (Fig. 4.9; 41 x 22 μ m): This pollen grain is undoubtedly conspecific with V. subtilis recorded from the Triassic and Jurassic of Queensland by De Jersey (1962, p. 11, pl. 4, figs 8 & 9; De Jersey 1970b, pl. 5, fig. 7) because it shows all the features of the latter (22–48 μ m in overall breadth). As De Jersey mentioned, V. subtilis differs from V. contectus (De Jersey) in having sacci with lesser overlap on the corpus and "from V. signatus Leschik by the lack of marked saccus ornament and of a short trilete tetrad-scar observed in some of the

European specimens".

11. Vitreisporites pallidus (Reissinger) Nilsson, 1958 (Fig. 4.12; 33 x 16 μ m, side view): The present specimen differs from V. subtilis noted above in having smaller corpus, however, as suggested by De Jersey (*in* Playford & Dettmann 1965), V. subtilis "may well be conspecific with V. pallidus". Vitreisporites pallidus has been widely reported from the Upper Permian and Mesozoic in both Southern and Northern hemispheres.

12. Alisporites cf. A. australis De Jersey (Fig. 4.1; 62.5 x 45 μ m): This pollen grain characterized by a very narrow distal sulcus and a vertically spindle-shaped corpus is quite similar to A. australis De Jersey, 1962 (p. 8, pl. 3, fig. 3; 68 μ m) and an "indeterminate monosaccate pollen" (De Jersey 1962, pl. 3, fig. 6; c. 90 μ m) from the Ipswich coals, but shows more inflated sacci than A. australis. It is also comparable to A. gottesfeldii of Traverse (1988, p. 216, fig. 11-2, x; c. 62.5 x 50 μ m) from the Middle-Late Triassic Chinle Formation, Arizona of the American south-west.

13. *Platysaccus queenslandi* De Jersey 1962 (Fig. 4.2; 52 x 35 μ m): Compared with the holotype of *P. queenslandi* (De Jersey 1962, p. 10, pl. 4, fig. 5; 88 μ m), the present specimen has smaller size and less inflated sacci, but it is quite similar to that identified by Playford *et al.* (1982, pl. 12, fig. 7; *c*. 60 μ m) from the Middle Triassic Moolayember Formation, Queensland.

14. Classopollis cf. meyeriana (Klaus) De Jersey, 1973 (Fig. 3.27, 3.28; 30 μ m and 32.5 μ m): The specimens are characterized by a narrow equatorial girdle and a subequatorial rimula, and on one specimen (Fig. 3.27) a triangular opening is observable. These psilate pollen are similar to *Classopollis meyeriana* (Klaus) De Jersey 1973 (pl. 4, figs 4 & 5) as suggested by Askin & Raine (personal communication 1999), but the latter appears to have a broader distance between the rimula and amb. They are also comparable to *Praecirculina* Klaus, 1960 (see Jansonius & Hills 1976, Card 2138) with type species *P. granifer* (Les.) Klaus of Carnian–Norian age, but differ from the latter in the absence of exine ornamentation.

Fig. 3. (see opposite) 1. Leiotriletes? sp., A97D129 (1), 41.5/99.0; 2. Deltoidospora directa (Balme & Hennelly) Norris, 1965, ARS2 (5), 36.0/101.5, x1000; 3. Dictyophyllidites harrisii Couper, 1958, A97D021-2 (17), 31.0/100.0; 4. Punctatisporites sp., D002 (2), 43.5/106.8, x700; 5, 6. Granulatisporites sp. cf. G. minor De Jersey, 1959, 5. A97D021-2 (27), 45.4/104.7, 6. A97D021-2 (20), 32.0/97.0; 7. Acanthotriletes sp., A97D021-2 (28), 38.0/99.5; 8a-b. Converucosisporites sp., A97D078 (9), 29.2/105.8; 9. Nevesisporites sp., ARS2 (2), 35.0/111.0, x1000; 10. Lophotriletes sp. cf. L. novicus Singh, 1964, A97D021-2 (17), 32.0/104.0, x1000; 11, 12, 16. Osmundacidites sp. cf. O. parvus De Jersey, 1962, 11. A97D021-2 (17), 31.5/99.0; 12. A97D021-2 (30), 40.1/108.0; 16. A97D129 (8), 35.5/100.0; 13a-b. Uvaesporites verucosus (De Jersey) Helby in De Jersey 1971, A97D021-2 (32), 31.3/101.8, x1000; 14, 19. Kraeuselisporites sp., 14. A97D021-2 (30), 42.0/101.0; 19. A97D021-2 (19), 40.8/106.5, x1000; 15. Duplexisporites? sp., A97D021-2 (12), 46.5/104.9; 17. Densosporites? sp., A97D078 (1), 40.0/103.0, x1000; 18. Aratrisporites sp., A97D021-2 (19), 44.8/106.5, x1000; 20. Araucariacites australis Cookson, A97D078(3), 36.8/106.8, X500; 21. Kraeuselisporites? sp., A97D021-2 (19), 39.2/110.1; 22-24. Classopollis? sp., 22. A97D129 (13), 39.6/100.0; 23. A97D021-2 (28), 41.0/107.0, x1000; 24. A97D129 (4), 41.5/106.1, showing a degraded tetrad of Classopollis? sp., the rimula (upper) is discernible; 25, 26. Cycadopites sp. a, 25. A97D021-2 (6), 31.0/102.0; 26. A97D021-2 (12), 39.3/103.0; 27, 28. Classopollis cf. C. meyeriana (Klaus) De Jersey, x1000. 27. ARS2 (3), 29.8/100.0; 28. ARS2 (5), 41.0/105.0; 29. Rimulate, unidentified, A97D129 (9), 36.0/104.5; 30. Cycadopites sp. b, A97D021-2 (27), 42.0/110.0; 31. Rimulate(?), unidentified, A97D078 (1), 34.0/107.0, x1000.





Rimulate (?) grains are also shown on Fig. 3.29 and 3.31 (60 μ m and 33 μ m).

15. Classopollis? sp. (Fig. 3.22, 40 μ m; Fig. 3.23, 26 μ m; Fig. 3.24, a tetrad, 42 μ m): Only a few grains of this type are present in the MBF assemblage. These pollen have been carbonized or degraded after oxidation, however, the thick equatorial band and subequatorial rimula are clearly shown, and the size as well as the general appearance quite resemble *Classopollis* although the identification is made with reservation.

16. Acritarchs: In addition to several specimens of psilate acritarchs, a few spinose acritarchs have been observed. One type (Fig. 4.26, $c.20 \mu$ m) may be identified as *Micrhystridium* sp. The other (Fig. 4.25, 32 μ m) possibly with a tetragonal opening is hard to identify and tentatively ascribed to Acanthomorphic acritarch.

17. Cuticles. Mainly two types of fragmentary cuticles are present in the slides. One is represented by several specimens (Type A: Fig. 5.18, 5.19): This type of cuticle is of some significance because it shows sinuous anticline walls. Sinuous cell walls in Mesozoic plants are almost an exclusively diagnostic feature of bennettitalean cuticles (Meyen 1987). The other (Type B: Fig. 5.17) is rather common, in lacking stomata etc., its cuticle nature and affinity (alga?, higher plant?, even insect?) is difficult to ascertain.

18. Wood fragments (tracheids) (Fig. 5.1–5.16). Various but isolated and incomplete tracheids have been observed in the slides. These are recorded and illustrated here for reference in future palaeobotanical study. Tracheid morphology is an important character both in living and fossil xylotomical studies. However, without evidence of other essential characters, such as primary and secondary xylem, early or late wood, pith, rays, resin canals, parenchyma and pitting in cross-fields etc. as conventionally shown in the three-plane sections (radial, tangential and transverse) (Greguss 1955), tracheids alone are of little use for identifying unknown plants, let alone isolated dispersed and fossil tracheids (Sah & Jain 1964). Many fossil wood genera for Paleozoic and Mesozoic have been proposed under comparative or artificial

nomenclature systems, but they can hardly be applied to the present material, even though in a few cases their applications appear feasible, for example, Araucarioxylon sp. (Fig. 5.3) and Xenoxylon? sp. (Fig. 5.7, 5.12, 5.13). For this reason, the present tracheids are artificially differentiated into several types without using form-genera. The main ones include "araucaroid-type" with contiguous, alternate and typically hexagonal pits (e.g. Fig. 5.3, 5.2), "abietoid-type" with discontinuous, uniseriate or opposite and circular or oval pits (Fig. 5.9, 5.10 with Sanio's bar), and a few are transitional between these two types (Fig. 5.1, 5.16), each containing two or more forms. One type with scalariform thickenings (Fig. 5.14) was possibly derived from cycads or pteridophytes (Greguss 1966), but is uncertain from what group of Gymnospermae the other with spiral thickenings (Fig. 5.4, 5.8) derived.

Due to characters shared by all samples, such as the absence of bisaccate Striatiti and the presence of similar kinds of tracheids, also because of the poverty of the overall recovery, the spores, pollen, and acritarchs are treated as a single palynological assemblage. In the assemblage spores and pollen are proportionate in specific diversity, but quantitatively non-striate bisaccate pollen occupy a dominant position (c.60%as roughly counted in the sample D021-2), and the latter point is somewhat comparable to the Alisporites Zone of Kyle (1977) from the Triassic of southern Victoria Land which "is characterized by abundant non-striate bisaccate pollen of the genus Alisporites" and the Late Triassic Timber Peak assemblage of northern Victoria Land in which the "unidentified bisaccate grains" account for 72.3% (Norris 1965). Except for a few grains of Platysaccus (Fig. 4.2), Pteruchiopollenies (Fig. 4.23) and Alisporites cf. A. australis (Fig. 4.1), most of the bisaccates are rather small in size $(30-52 \mu m, generally)$ 40 µm) and assigned to Vitreisporites (Fig. 4.9, 4.10, 4.12, 4.14) and partly Alisporites (Fig. 4.11) as well as Pteruchipollenites cf. P. parvus (Fig. 4.16).

According to our present knowledge of *in situ* spores (Potonié 1962, Traverse 1988, Balme 1995), the terrestrial parent flora might have been composed of various pteridophytes and gymnosperms, including pteridosperms,

Fig. 4. (see opposite) 1. Alisporites sp. cf. A. australis De Jersey, 1962, A97D107 (1), 36.2/105.8; 2. Platysaccus queenslandi De Jersey, 1962, A97D021-2 (21), 41.0/102.5; 3. Platysaccus sp., A97D021-2 (32), 30.3/103.2; 4. Cingulatisporites? sp., A97D021-1 (1), 33.0/108.0, x1000; 5a-b, 6a-b. Podosporites sp. 5b, 6b, x1000. 5a-b. A97D021-2 (7), 45.0/99.2; 6a-b. A97D021 2 (12)35.0/95.2; 7. Platysaccus sp., ARS2 (5), 33.2/103.0; 8. Vesicaspora? sp., A97D021-2 (17), 33.6/104.3, x700; 9. Vitreisporites subtilis (De Jersey) De Jersey, 1962, A97D021-2 (19), 43.0/108.6; 10. Vitreisporites microsaccus De Jersey, 1964, A97D021-2 (8), 44.0/100.0; 11. Alisporites sp., A97D021-2 (16), 38.0/99.0; 12. Vitreisporites pallidus (Reiss.) Nilsson, 1958, A97D021-2 (2), 31.4/102.4, x1000; 13. Pityosporites sp., A97D021-2 (9), 33.0/96.0; 14. Vitreisporites sp. ef. V. pallidus (Reiss.) Nilsson, 1958, A97D021-2 (6), 38.0/107.0; 15, 16. Pteruchipollenites sp. ef. P. parvus (De Jersey) Foster, 1975, A97D021-2 (11), 44.0/109.0; 15. x550; 17, 20. Accinctisporites? sp., 17. A97D021-2 (32), 36.0/96.0, x550; 20. A97D021-2 (3), 30.5/101.5; 18. Exesipollenites? sp., A97D021-2 (17), 45.0/108; 19. Densipollenites? sp., A97D02102 (2), 43.0/97.5; 21. Minutosaccus? sp. A97D021-2 (19), 37.0/106.8, x1000; 22. Fragment of a microfossil unidentified, ARS2 (2), 39.0/106.0; 23. Pteruchipollenites sp., A97D021-2 (19), 37.0/106.8, x1000; 24. Leiosphaeridia sp., A97D0246 (1), 43.5/102.0; 25. Acanthomorphic acritarch unidentified, A97D021-2 (19), 30.0/104.0; 26. Micrhystridium sp., D002 (5), 31.0/102.8, x800; 27. Schizosporis? sp., A97D021-1 (4), 27.0/100.0; 28. An acritarch unidentified, A97D129 (9), 40.5/95.8.

conifers, ginkgoaleans, cycadaleans and possibly bennettitaleans. Tracheids in the assemblage are dominated by araucaroid- and abietoid-types, possibly mainly derived from pteridosperms and conifers (Greguss 1955, 1966, Pant & Singh 1987, Jefferson *et al.* 1983a, 1983b).

The MBF assemblage shows little similarity with those from the Triassic of Carnarvon Basin, Western Australia (Dolby & Balme 1976). Almost all taxa found from the latter have not been recovered from the MBF, including those zonal indices. Combined with the comparison discussed below, it is obvious that the MBF assemblage is essentially of the Ipswichtype.

Correlation and discussion

Bisaccate striate pollen

The absence of bisaccate Striatiti in the present assemblage deserves special attention. In Australia, these kinds of pollen first appeared in the earliest Permian (Playford 1989), and as in other continents of Gondwana (Playford 1989), reached their development acme in Middle–Late Permian and in general gradually decreased in species diversity and abundance in the Early–Middle Triassic (Foster 1979) and dramatically declined or disappeared in the Late Triassic (Playford & Dettmann 1965, Norris 1965, De Jersey & Hamilton 1967, De Jersey 1962, 1970a, 1972, 1979). On the other hand, they generally have not been recorded in the Jurassic assemblages of Australia (e.g. Playford & Dettmann 1965, Reiser & Williams 1969, McKellar 1974, Filatoff 1975). In short, the absence of the Striatiti in the present assemblage suggests an age not older than Late Triassic.

Aratrisporites

One single specimen of definite *Aratrisporites* sp. has been observed in the assemblage. Although sporadically earlier (e.g. the Early Permian Qipan Formation in Tarim Basin of China, Wang 1985) or later (e.g. the Early Jurassic strata in New Zealand, Zhang & Grant-Mackie 1997; the Early Jurassic Beipiao Formation in Liaoning of NE China, Wu & Zhang 1983; the lower Hettangian of NW Europe, Batten & Koppelhus 1996) records have been known in both hemispheres, this genus has been generally taken as an important marker of Triassic. In Antarctica, Kyle (1977) mentioned that *Aratrisporites* is "common to dominant in the older part but infrequent in the younger samples" of her *Alisporites* Zone, although in the Shackleton Glacier area they are abundant in the Middle and Late Triassic (Askin, personal communication 1999). This seems to be in favour of dating the MBF as Late Triassic.

Classopollis

In the present assemblage occur a few grains of *Classopollis*. Classopollis is one of the most important components of the latest Triassic-Middle Cretaceous palynofloras of the world. In the Northern Hemisphere, its first appearance was taken as at the base of the Norian (De Jersey 1970b, Warrington 1996). In Australia, it is first occasionally encountered in Rhaetian deposits (Aberdare Conglomerate; De Jersey 1971, 1973) of the Bundamba Group and became dominant in the Lower Jurassic, such as in the Precipice Sandstone and lower Evergreen Formation in Queensland (Reiser & Williams 1969, McKellar 1974), the upper part of the Leigh Creek Measures in southern Australia (Playford & Dettmann 1965) and the Cockleshell Gully Formation in the Perth Basin of Western Australia (Filatoff 1975). In New Zealand, Classopollis first appears in the upper part (Rhaetian) of the Otapirian Stage and becomes locally abundant in the Hettangian sediments (De Jersey & Raine 1990, Zhang & Grant-Mackie 1997). Zavattieri & Batten (1996) mentioned that ".....it (Classopollis) does not become frequent in Argentinean successions until the Jurassic". In the assemblages from the Ferrar Group at Section Peak of the Rennick Glacier region and at Carapace Nunatak of Transantarctic Mountains, both of Early Jurassic age (Norris 1965, Tasch & Lammons 1978), Classopollis is an overwhelmingly dominant genus. It is believed that the MBF assemblage should be older than the Ferrar Group but can not be older than Norian and is possibly Norian-Rhaetian in age.

Regional correlation

Compared with those of the MBF assemblage, occurrences of some identical or comparable taxa from the Triassic, especially Upper Triassic in Antarctica, Australia and New Zealand are listed in Table I.

Fig. 5. (see opposite) 1. Form 1 of abietoid- araucaroid transitional type, A97D129(8), 40.0/96.5, 142 x 38μm; 2. Form 1 of araucaroid type. ARS2 (2), 28.2/101.5; 87 x 24μm; 3. Form 2 of araucaroid type. ARS2 (1), 32.5/104.1, 95 X 38 μm; 4. Tracheid with spiral thickenings- Form 1. A97D078 (8), 41.0/97.0, 32 x 21 μm; 5. Form 3 of araucaroid type. A97D129 (9), 39.2/108.7, 120 x 48μm;
6. Form 4 of araucaroid type. A97D129 (7), 33.0/102.0, 97 x 42 μm; 7, 12, 13. Form 3 of abietoid type, 7. A97D021-1 (2), 39.0/108.7, 74 x 35 μm; 12. ARS2 (2), 30.0/103.0, 38 x 46 μm; 13. A97D021-2 (27), 36.0/98.0; 76 x 47 μm; 8. Tracheid with spiral thickenings-Form 2, A97D021-2 (18), 31.0/100.0, 57.5 x 24 μm; 9, 10. Form 1 of abietoid type, 9. A97D021-2 (28), 31.2/106.0, 120 x 57 μm;
10. A97D021-2 (24), 37.0/96.5, 90 x 39 μm; 11. Form 1 of gymnosperm unknown affiliation, A97D129 (14), 43.6/101.1, 82 x 55 μm;
14. Tracheid with scalariform thickenings, A97D021-2 (30), 40.0/104.8, 110 x 83 μm; 15. Form 2 of abietoid type, A97D021-2 (27), 39.1/108.0, 62 x 25 μm; 16. Form 2 of abietoid -araucaroid transitional type, A97D246 (1), 33.0/109.0, 96 x 12.5 μm; 17. Cuticle Form B of unknown affinity, A97D021-2 (32), 33.0/104.8, 178 x 135 μm (partly shown); 18, 19. Cuticle Form A of possibly Bennettitales, 18. A97D078 (6), 43.0/108, 75 x 70 μm; 19. A97D078 (7), 39.8/103.8, 80 x 54 μm.



Discussion of Table I

The common occurrences shown in Table I indicate that the MBF assemblage is relatively similar to the ULY, ICM, TCM and NGT assemblages. Compared with subzones A, B, C and D of Kyle's (1977) Alisporites Zone for the Triassic interval of the Victoria Group from the Murlock-Mackay Glacier area, the MBF assemblage is relatively closer to subzones C and D and the Timber Peak assemblage which Kyle considered to be equivalent to her subzone C. Subzone A (Early Triassic) is characterized by various trilete cavate spores (Lundbladispora+Densoisporites) and common or abundant Aratrisporites as well as other taxa such as Grebespora concentrica. Subzone B is defined by the presence of Cadagarsporites senectus, ?Kraeuselisporites verrucifer, Retusotriletes junior, Tigrisporites playfordii, Equisetosporites steevesi and Duplexisporites problematicus. These taxa have not been observed in our assemblage except the common presence of Aratrisporites and Vitreisporites pallidus. If "the lower Lashly assemblage" assigned to subzone B as Kyle did, although some common or similar elements occur in the MBF assemblage as shown in the Table I, the absence of Protohaploxypinus and especially the diagnostic zonal species Cadargasporites senectus defies correlation between the MBF

assemblage and subzone B. The MBF may be temporarily correlated with the upper part of the Ipswich Coal Measures and the Bundamba Group, and seems to be younger than the lower part of the Ipswich Coal Measures because in the latter Aratrisporites is rather diverse. The occurrence of Craterisporites rotundus and bisaccate Striatiti in the Falla Formation suggests a little older age than the MBF. Among the four assemblages recorded from New Zealand (Zhang & Grant-Mackie 1997), the MBF assemblage bears greater resemblance with the assemblages I and II (Norian-Rhaetian), although some index forms have not been observed in the MBF assemblage, such as *Polycingulatisporites crenulatus*, Annulispora microannulata and Foveosporites moretonensis. On the other hand, the assemblages III and IV (Early Jurassic) differ from the MBF assemblage in having some diagnostic Jurassic forms, such as Toripustulatisporites hokonuiensis, Antulsporites varigranulatus, Kyrtomisporites elsenndoonii, Cibotium juncta, Lycopodiumsporites austroclavatidites, Perinopollenites elatoides and abundant Classopollis (assemblage IV). The MBF assemblage is also correlatable with those from the upper *Polycingulatisporites crenulatus* Zone and the Foveosporites more tonensis Zone (the Warepan and Otapirian Stages) of New Zealand, viz. to the more securely dated late Norian and Rhaetian (De Jersey & Raine

Table I. Regional correlation of Triassic taxa.

MBF	ULY	LLY	BST	FLA	ICM	BDB	WOS	тсм	NGT	
Deltoidospora directa			+	+	+		+	sp.	+	
Dictyophyllidites harrisii	+	+	+	+	+	+	+	+	+	
Punctatisporites sp.	+	+		+			+		+	
Granulatisporites sp.					+	+		+		
Osmundacidites sp.	+	+	+	+	+	+	+	+	+	
Convertucosisporites sp.	+	+	+		+		+	+	+	
Uvaesporites verrucosus	+				+	+	+	+	+	
Acanthoriletes sp.			+	+	+	+	+	+	+	
Lophotriletes novicus	+				sp.	sp.	sp.			
Duplexisporites? sp.	+				+					
Cingulatisporites sp.					+					
Densosporites sp.	+				+	+				
Nevesisporites sp.	+	+								
Aratrisporites sp.	+	+			+	÷	+	+	+	
Alisporites spp.	+	+	?		+		+	+	+	
Alisporites parvus	+			+	+					
Alisporites cf. australis	+				+		+			
Vitreisporites pallidus	+	+	+	+	+	+	+			
Vitreisporites subtilis					+					
Pityosporites sp.				+						
Platysaccus queenslandi	+			+	+	+	+	sp.	sp.	
Pteruchipollenites sp.			+	+						
Araucariacites australis			+				+		sp.	
Classopollis sp.						+	+		+	
Cycadopites spp.	+	+	+		+	+	+	+	+	

MBF = Miers Bluff Formation, the present assemblage; ULY = upper Lashly Formation (subzones C+D of the *Alisporites* Zone), Late Triassic, South Victoria Land, Antarctica (Kyle 1977); LLY = "lower Lashly Formation", Ladinian?, Mount Feather, South Victoria Land (Helby & McElroy 1969); BST = "Beacon Sandstone", Late Triassic, Timber Peak, northern Victoria Land (Gair *et al.* 1965, Norris 1965); FLA = Falla Formation, Carnian-Norian, Central Transantarctic Mountains (Farabee *et al.* 1989); ICM = Ipswich Coal Measures, Carnian, Australia (De Jersey 1962, 1971); BDB = Bundamba Group, Aberdale Conglomerate and Raceview Formation, Rhaetian, Australia (De Jersey 1970b, 1973); WOS = Warepan Stage (late Norian) and Otapirian Stage (Norian-Rhaetian) of the Murihiki Supergroup, New Zealand (De Jersey & Raine 1990); TCM = Trechmann Siltstone, Assemblage I, Norian; NGT = Ngutunui Formation and Neucastle Group, Assemblage II, Rhaetian, New Zealand (Zhang & Grant-Mackie 1997).

1990).

To sum up, the MBF assemblage is comparable to those of the C+D subzones of Kyle's *Alisporites* Zone of Victoria Land and the upper *Craterisporites rotundus* Zone and lower *Polycingulatisporites crenulatus* Zone of Australia as well as the *Polycingulatisporites crenulatus* Zone and *Foveosporites moretonensis* Zone of New Zealand, suggesting a Late Triassic (possibly Norian-Rhaetian) age. The absence of Striatiti and the sporadic occurrence of *Aratrisporites* and *Classopollis* lent support to the above dating. In addition, it is interesting to note the presence of a few possibly bennettitalean cuticles in the MBF: Bennettitaleans were represented by a number of taxa only up to Late Triassic in the Northern Hemisphere.

Conclusions

- 1)The deposits of the MBF contain various and diverse plant microfossils although they are poorly preserved due to thermal alteration of the rocks. The plant microfossils include a total of about 45 species (forms) assigned to 37 genera of miospores, viz. spores of 18 species/17 genera, pollen of 22 species/15 genera in addition to acritarchs of 5 species /5 genera; wood fragments of 13 forms and cuticles of two types.
- 2)The palynoflora is quantitatively dominated by bisaccate gymnosperm pollen, but with pteridophyte spores and gymnosperm pollen equally diverse. Phytogeographically, it belongs to the Ipswich type.
- 3)The palynoflora is comparable to subzones C+D of Kyle's (1977) Triassic *Alisporites* Zone from southern Victoria Land of Transantarctica and the upper *Craterisporites rotundus* Zone and *Polycingulatisporites crenulatus* Zone from Queensland (viz. the upper part of the Ipswich Coal Measures and Bundamba Group), indicating a Late Triassic (possibly Norian–Rhaetian) age.
- 4)The Late Triassic dating of the MBF lends support correlation with the Potranca Formation in southern Chile, the Greywacke Shale Formation in the South Orkney Islands, and the Cape Legoupil Formation, as well as probably the Paradise Harbour and Hope Bay Formations in the Antarctic Peninsula. All these formations might have been contemporaneously deposited under similar tectonic conditions just prior to the breakup of Gondwana in Early to early Middle Jurassic times.

Acknowledgements

This study was supported by the Science and Technology Commission (98-927-01), the Chinese Academy of Sciences (KZ-95-A1-205) and the NNSF (Grant No. 49572076 and No. 49671176). We are grateful to He Cui-ling for successfully extracting the microfossils from such difficult rocks and to Fang Xiao-yi for printing the microfossil photos. We should like to thank Zhou Zhi-yan, R.A. Askin and especially the reviewers (D.J. Batten and I. Raine) for their critical reading of the manuscript and many valuable and constructive comments.

References

- ANDREIS, P.R., RIBEIRO, A. & TROUW, R.A.J. 1997. Remarks on facies, paleocurrents and petrofacies of Triassic submarine fans from the Antarctic Peninsula, South Shetland Islands and South Orkney Islands. In RICCI, C.A., ed. The Antarctic region: geological evolution and processes. Siena: Terra Antarctica Publication, 401-407.
- ARCHE, A., LÓPEZ-MARTINEZ, J. & DE PISON, E.M. 1992. Sedimentology of the Miers Bluff Formation, Livingston Island, South Shetland Islands. In YOSHIDA, Y., KAMINUMA, K. & SHIRAISHI, K., eds. Recent progress in Antarctic earth science. Tokyo: Terra Scientific Publishing Company, 357-362.
- ASKIN, R.A. 1981. Jurassic-Cretaceous palynology of Byers Peninsula, Livingston Island, Antarctica. Antarctic Journal of the United States, 16(5), 11-13.
- BALME, B.E. 1995. Fossil in situ spores and pollen grains: an annotated catalogue. Review of Palaeobotany and Palynology, 87, 81-323.
- BATTEN, D.J. & KOPPELHUS, E.B. 1996. Biostratigraphic significance of uppermost Triassic and Jurassic miospores in northwest Europe. In JANSONIUS, J. & McGREGOR, D.C., eds. Palynology: principles and applications. AASP Foundation, 2, 795-806.
- BIRKENMAJER, K. 1992. Trinity Peninsula Group (Permo-Triassic?) at Hope Bay, Antarctic Peninsula. Polish Polar Research, 13, 215-240.
- DALZIEL, I.W.D., ELLIOT, D.H., JONES, D.L., THOMSON, M.R.A., WELLS, N.A. & ZINSMEISTER, W.J. 1981. The geological significance of some Triassic microfossil from the South Orkney Islands, Scotia Ridge. Geological Magazine, 118, 15-25.
- DE JERSEY, N.J. 1962. Triassic spores and pollen grains from the Ipswich Coalfield. Publications of the Geological Survey Queensland, 307, 1-18.
- DE JERSEY, N.J. 1970a. Early Triassic miospores from the Rewan Formation. Geological Survey of Queensland Publications, 345, Palaeontological Papers, 19, 1-29.
- DE JERSEY, N.J. 1970b. Triassic miospores from the Blackstone Formation, Aberdare Conglomerate and Raceview Formation. Geological Survey of Queensland Publications, 348, Palaeontological Papers, 22, 1-41.
- DE JERSEY, N.J. 1971. Triassic miospores from the Tivoli Formation and Kholo Sub-group. Geological Survey of Queensland Publications, 353, Palaeontological Papers, 28, 1-40.
- DE JERSEY, N.J. 1972. Triassic miospores from the Esk Beds. Geological Survey of Queensland Publications, 357, Palaeontological Papers, 32, 1-40.
- DE JERSEY, N.J. 1973. Rimulate pollen grains from the lower Mesozoic of Queensland. Special Publications Society of Australia, 4, 127–140.
- DE JERSEY, N.J. 1979. Palynology of the Permian-Triassic transition in the Western Bowen Basin. Geological Survey of Queensland Publications, 374, Palaleontological Papers, 46, 1-39.
- DE JERSEY, N.J. & HAMILTON, M. 1967. Triassic microfloras of the Moolayember Formation. Geological Survey of Queensland Publications, 336, Palaeontological Papers, 10, 1–61.
- DE JERSEY, N.J. & RAINE, J.I. 1990. Triassic and earliest Jurassic miospores from the Murihiku Supergroup, New Zealand. New Zealand Geological Survey Paleontological Bulletin, No. 62, 1-164.
- DOLBY, J.H. & BALME, B.E. 1976. Triassic palynology of the Carnarvon Basin, Western Australia. *Review of Palaeobotany and Palynology*, 22, 105–168.

- DOKTOR, M., SWIERCZEWSKA, A. & TOKARSKI, A.K. 1994. Lithostratigraphy and tectonics of the Miers Bluff Formation at Hurd Peninsula, Livingston Island (West Antarctica). Studia Geologica Polonica, 104, 41–104.
- FANG, Z.J., BOUCOT, A., COVACEIVICH, V. & HERVE, F. 1998. Discovery of Late Triassic fossils in the Chonos metamorphic complex, southern Chile. Revista Geologica de Chile, 25, 165-173.
- FARABEE, M.J., TAYLOR, T.N. & TAYLOR, E.L. 1989. Pollen and spore assemblages from the Falla Formation (Upper Triassic), Central Transantarctic Mountains, Antarctica. *Review of Palaeobotany and Palynology*, 61, 101–138.
- FILATOFF, J. 1975. Jurassic Palynology of the Perth Basin, Western Australia. Palaeontographica, B154, 1-113.
- FOSTER, C.F. 1979. Permian plant microfossils of the Blair Athol coal measures, Baralaba coal measures and basal Rewan Formation of Queensland. Geological Survey of Queensland Publications, 372, Palaeontological Papers, 45, 1-244.
- GAIR, H.S., NORRIS, G. & RICKER, J. 1965. Early Mesozoic microfloras from Antarctica. New Zealand Journal of Geology and Geophysics, 8, 231-235.
- GREGUSS, P. 1955. Identification of living gymnosperms on the basis of xylotomy. Budapest: Akadamie Kiado, 263 pp.
- GREGUSS, P. 1966. The relationships of Cycadales on the basis of their xylotomy, branching and leaf epidermis. The Palaeobotanist, 14, 94-101.
- GRIKUROV, G.E. & DIBNER, A.F. 1968. New data on the Trinity Series (C1-2) in West Antarctica. Dokladai Akademy Nauk SSSR, 179, 410-412. [In Russian].
- HELBY, R.J. & MCELROY, C.T. 1969. Microfloras from the Devonian and Triassic of the Beacon Group. New Zealand Journal of Geology and Geophysics, 12, 376–382.
- JANSONIUS, J. & HILLS, L.V. 1976. Genera file of fossil spores and pollen. Department of Geology, University of Calgary, Special Publication.
- JEFFERSON, T.H., SIDERS, M.A. & HABAN, M.A. 1983. Jurassic trees engaged by lavas of the Kirkpatrick Basalt Group, northern Victoria Land. Antarctic Journal of the United States, 18(5), 14-16.
- JEFFERSON, T.H. & TAYLOR, T.N. 1983. Permian and Triassic woods from the Transantarctic Mountains: palaeoenvironmental indicators. Antarctic Journal of the United States, 18(5), 55-57.
- KYLE, R.A. 1977. Palynostratigraphy of the Victoria Group of South Victoria Land, Antarctica. New Zealand Journal of Geology and Geophysics, 20, 1081–1102.
- MANDAL, J. & MAITHY, P.K. 1981. Palaeobotany of Tittitola Bed in Pachwara Coalfield, Santhal Pargana, Bihar with remarks on the age of the bed. *The Palaeobotanist*, **27**, 12-20.
- MCKELLAR, J.L. 1974. Jurassic miospores from the Upper Evergreen Formation, Hutton Sandstone, and basal Injune Creek Group, northeastern Surat Basin. Geological Survey of Queensland Publications, 361, Palaeontological Papers, 35, 1–89.
- MEYEN, S.V. 1987. Fundamentals of palaeobotany. London: Chapman & Hall, 432 pp.
- NORRIS, G. 1965. Triassic and Jurassic miospores and acritarchs from the Beacon and Ferrar groups, Victoria Land, Antarctica. New Zealand Journal of Geology and Geophysics, 8, 236-277.
- PANT, D.D. & SINGH, V.K. 1987. Xylotomy of some woods from Raniganj Formation (Permian), Raniganj Coalfield, India. Palaeontographica, B203, 1-82.
- PLAYFORD, G. 1989. Proterozoic and Palaeozoic palynology of Antarctica: a review. In TAYLOR, T.N. & TAYLOR, E.L., eds. Antarctic paleobiology: its role in the reconstruction of Gondwana. New York: Springer-Verlag, 51-70.

- PLAYFORD, G. & DETTMANN, M.E. 1965. Rhaeto-Liassic plant microfossils from the Leigh Creek Coal Measures, South Australia. Senckenbergiana Lethaea, 46, 127-181.
- PLAYFORD, G., RIGBY, J.F. & ARCHIBALD, D.C. 1982. A Middle Triassic flora from the Moolayember Formation, Bowen Basin, Queensland. Publications of the Geological Survey Queensland, 80, 1-52.
- POTONIE, R. 1962. Synopsis der sporae in situ. Beihefte zum geologische Jahrbuch, 52, 1-204.
- REISER, R.F. & WILLIAMS, A.J. 1969. Palynology of the Lower Jurassic sediments of the Northern Surat Basin, Queensland. Geological Survey of Queensland Publications, 339, Palaeontological Papers, 15, 1-24.
- SAH, S.C.D. & JAIN, K.P. 1964. Some fossil woods from the Jurassic of Rajmhal Hill, Bihar, India. The Palaeobotanist, 12, 169–179.
- SHANG, Y.K. 1997. Middle Jurassic palynology of Carapace Nunatak, Victoria Land, Antarctica. Acta Palaeontologica Sinica, 36, 170–186. [In Chinese with English abstract]
- SMELLIE, J.L., LIESA, M., MUNOZ, J.A., SABAT, F., PALLAS, R. & WILLAN, R.C.R. 1995. Lithostratigraphy of volcanic and sedimentary sequences in central Livingston Island, South Shetland Islands. *Antarctic Science*, 7, 99-113.
- SINGH, H.P. 1964. A miospore assemblage from the Permian of Iraq. Palaeontology, 7, 240-265.
- TASCH, P. 1977. International correlation by conchostracan and palynomorphs from Antarctica, Western Australia, India and Africa. Antarctic Journal of the United States, 12(4), 121.
- TASCH, P. & LAMMONS, J.M. 1978. Palynology of some lacustrine interbeds of the Antarctic Jurassic. *Palinologia*, Numero extraordinaire No. 1, 455-458.
- TAYLOR, T.N. & TAYLOR, E.L. eds. 1989. Antarctic paleobiology: its role in the reconstruction of Gondwana. New York: Springer-Verlag, 261 pp.
- THOMSON, M.R.A. 1975. New palaeontological and lithological observations on the Legoupil Formation, north-west Antarctic Peninsula. British Antarctic Survey Bulletin, No. 41(2), 16-85.
- TOKARSKI, A.K., DOKTOR, M., WILLAN, R.C.R., SMELLIE, J.L., SABAT, F. & MUNOZ, J.A. 1996. Debate: Lithostratigraphy of volcanic and sedimentary sequences in central Livingston Island, South Shetlan Islands. Antarctic Science, 8, 207–08.
- TRAVERSE, A. 1988. Paleopalynology. Boston: Unwin Hyman, 600 pp.
- TRUSWELL, E.M. 1991. Antarctica: a history of terrestrial vegetation. In TINGEY, R.J., ed. Geology of Antarctica. Oxford: Oxford University Press, 499–537.
- WANG, H. 1985. New genera and species in the Early Permian palynological assemblage of Qipan-Duwa area, Tarim Basin. Acta Palaeontologica Sinica, 24, 663-671. [In Chinese with English abstract]
- WARRINGTON, G. 1996. Triassic spores and pollen. In JANSONIUS, J. & MCGREGOR, D.C., eds. Palynology: principles and applications. AASP Foundation, 2, 755-766.
- WILLAN, R.C.R., PANKHURST, R.J. & HERVÉ, F. 1994. A probable Early Triassic age for the Miers Bluff Formation, Livingston Island, South Shetland Islands. Antarctic Science, 6, 401–408.
- WU, H.ZH. & ZHANG, X.L. 1983. Palynological assemblage from the upper coal-bearing member of the Beipiao Formation of Liaoning. Acta Palaeontologica Sinica, 22, 564-569.
- ZAVATTIERI, A.M. & BATTEN, D.J. 1996. Miospores from Argentinian Triassic deposits and their potential for intercontinental correlation. In JANSONIUS, J. & McGREGOR, D.C., eds. Palynology: principles and applications. AASP Fundation, 2, 767-778.
- ZHANG, W.P. & GRANT-MACKIE, J.A. 1997. The sporo-pollen assemblages of the Late Triassic and Early Jurassic in New Zealand and their correlation between China and New Zealand. Beijing: Seismological Press, 1-70. [In Chinese with English abstract].