SHORT COMMUNICATION

Underdeveloped embryos and kinds of dormancy in seeds of two gymnosperms: *Podocarpus costalis* and *Nageia nagi* (Podocarpaceae)

Shun-Ying Chen¹, Carol C. Baskin^{2,3}, Jerry M. Baskin² and Ching-Te Chien^{1*}

¹Division of Silviculture, Taiwan Forestry Research Institute, 53 Nan-Hai Road, Taipei 10066, Taiwan; ²Department of Biology, University of Kentucky, Lexington, Kentucky 40506-0225, USA; ³Department of Plant and Soil Sciences, University of Kentucky, Lexington, Kentucky 40546-0312, USA

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Abstract

Although it has been speculated that seeds of the gymnosperm family Podocarpaceae have an underdeveloped embryo, no detailed studies have been done to definitively answer this question. Our purpose was to determine if embryos in seeds of two species of Podocarpaceae, Podocarpus costalis and Nageia nagi, from Taiwan are underdeveloped and to examine the kind of dormancy the seeds have. Embryos in fresh seeds of *P. costalis* were 4.6 ± 0.5 mm long, and they increased in length by about 54% before radicle emergence (germination), demonstrating that the embryo is underdeveloped at seed maturity. Seeds germinated to >90% at 30/20, 25/15 and 25°C in light in \leq 4 weeks, without any cold stratification pretreatment. Thus, seeds of *P. costalis* have morphological dormancy (MD). Embryos in fresh seeds of N. nagi were 7.4 \pm 0.8 mm long and they increased in length by about 39% before radicle emergence (germination) occurred, indicating that the embryo is underdeveloped at seed maturity. Seeds germinated to <25% at 30/20 and 25° C in light in 4 weeks but to >90% at the same temperatures in 12 weeks. Thus, most seeds of N. nagi have morphophysiological dormancy (MPD). Although underdeveloped embryos are considered to be a primitive condition in seed plants, they also occur in the most advanced orders. The occurrence of underdeveloped embryos in Podocarpaceae documents that they are not restricted to a basal clade in gymnosperms.

*Correspondence Email: chien@tfri.gov.tw Keywords: embryo growth, morphological dormancy, morphophysiological dormancy, *Nageia*, *Podocarpus*, radicle emergence, shoot emergence

Introduction

The occurrence of an underdeveloped embryo in a seed means the embryo must grow inside the seed before the radicle emerges. If seeds with an underdeveloped embryo are incubated at a suitable temperature, light:dark and moisture conditions, and the delay in emergence of the radicle is only about 1 month or less, seeds have morphological dormancy (MD) (Nikolaeva, 1977; Baskin and Baskin, 1998). In contrast, if the delay in emergence of the radicle is longer than about 1 month and warm and/or cold moist treatments are required to promote germination, seeds have morphophysiological dormancy (MPD). Various levels of MPD have been identified, depending on requirements to break dormancy and promote germination, temperature requirements for embryo growth and response to gibberellins (GA₃) (Nikolaeva, 1977; Baskin and Baskin, 1998, 2004; Baskin et al., 2008).

In one classical view of extant gymnosperms, there are three classes, Ginkgoopsida, Cycadopsida and Pinopsida (Meyen, 1984). Among the nine families in Pinopsida, only Podocarpaceae and Taxaceae are reported to have underdeveloped embryos. Detailed studies have shown that seeds of species in Taxaceae have an underdeveloped embryo, and deep simple MPD has been documented in seeds of *Cephalotaxus wilsoniana* (Yang *et al.*, 2011), *Taxus baccata* (Devillez, 1978), *T. brevifolia* (Nikolaeva *et al.*, 1985), *T. cuspidata* (Nikolaeva *et al.*, 1985; Cheng *et al.*, 2004) and *T. mairei* (Chien et al., 1998; Zhang et al., 2000; Liu et al., 2011). It also has been observed that embryos in seeds of Podocarpaceae are small in relation to the size of the female gametophyte (Ng, 1992; Ferrandis et al., 2011), and several studies on Podocarpaceae have revealed that varying periods of time are required for seeds to germinate. To reach 50% germination, seeds of Podocarpus angustifolius required 35 d if they first were soaked in water but otherwise >60 d (Ferrandis et al., 2011), while seeds of P. falcatus required a 1-month period of moist storage and then an incubation period of 3 weeks (Negash, 1992). Seeds of P. henkelii germinated to 68% after 160 d (Dodd and Van Staden, 1981). Further, seeds of Dacrydium comosum (Podocarpaceae) germinated over a period of 113-358 d, whereas those of Podocarpus imbricatus germinated over a period of 16-63d and those of *P. neriifolius* over a period of 20–67 d (Ng, 1992). In the temperate rain forest of southern Chile, seeds of *P. nubigena* planted on the soil surface in a canopy gap and in the understorey germinated to 0 and 22% in 2 years, respectively. There was a delay of more than 1 year in the germination of seeds planted in the understorey (Figueroa and Lusk, 2001). Further, cold stratification was ineffective in overcoming dormancy in seeds of this species (Figueroa, 2003). Fresh seeds of Nageia nagi collected from southern Taiwan germinated to 95% after 10 weeks of incubation at 30/20°C and at 25°C with 12 h light, and cold stratification at 5°C shortened germination time (Hong *et al.*, 2009). However, none of these authors reported direct observations or measurements on the embryos to see if they grew prior to radicle emergence.

To our knowledge, no detailed studies have been conducted to determine if embryos of Podocarpaceae grow inside the seed prior to germination, i.e. radicle emergence. Unless embryos grow prior to germination, they are not underdeveloped and the seeds cannot have MD or MPD. Thus, our purpose was to address the following questions. Do embryos in Podocarpaceae seeds grow prior to germination? Do seeds have MD or MPD, and if MPD, which of the nine known levels of MPD do they have?

Materials and methods

Seed collecting and handling

The species selected for this study were *Nageia nagi* (Thunberg) Kuntze and *Podocarpus costalis* Presl. Both species are mainly distributed in tropical and subtropical areas, and both are native to Taiwan, where they are threatened by habitat loss and by overcollection for ornamental uses. In Taiwan, *N. nagi* grows at low elevations in broadleaved evergreen forests of the island, whereas *P. costalis* is found along

the coasts among rocks on Lanyu Island (Orchid Island), south-east Taiwan (Li *et al.*, 1994). *P. costalis* has been widely cultivated in Taiwan.

Mature seeds of *P. costalis* with a deep-blue fleshy sarcotesta and a red-purple fleshy receptacle were collected from Chashan (22°00'N, 120°50'E), Manjhou township, Pingtung County on the southern tip of Taiwan, in September 2010. Mature seeds with a green fleshy sarcotesta were collected from plants of N. nagi growing in Chiavi city (23°29'N, 120°28'E), in southwestern Taiwan in August 2010. In seeds of both species, the embryo is surrounded by the megagametophyte, a stony sclerotesta and a fleshy sarcotesta (see Fig. 1). The sarcotesta was removed from seeds used to test for germination and embryo growth. All cleaned seeds were air-dried at room temperature overnight. There were 2310 seeds l^{-1} and 3725 seeds kg^{-1} in *P. costalis* and 500 seeds l^{-1} and 900 seeds kg^{-1} in N. nagi. The moisture content of fresh seeds without sarcotesta and with sclerotesta was $51 \pm 3\%$ in *P. costalis* and $56 \pm 2\%$ in *N. nagi*, as determined by oven drying for 17 h at 103°C (International Seed Testing Association, 2007). We cut more than ten freshly harvested seeds each of N. nagi and P. costalis and found that all of them had a healthy looking megagametophyte and embryo.

Germination study

Seeds of *P. costalis* and *N. nagi* were mixed with moist sphagnum moss, placed inside polyethylene bags (0.04 mm in thickness) and incubated at 12 h/12 halternating temperature regimes of 30/20, 25/15, 20/10 and 15/6°C and at a constant temperature of 25°C. The sphagnum moss was cut into small pieces, and then wetted to a water content of about 400% of its dry mass. The moist sphagnum provided a good germination medium and prevented the spread of mould because it contains the fungi Trichoderma and actinomycetes which are antagonistic to microorganisms (Wang et al., 1998). The seeds were covered with 3 mm or less of sphagnum moss. The daily photoperiod of 12 h in the incubators $(60-80 \,\mu\text{mol}\,\text{m}^{-2}\text{s}^{-1})$, 400–700 nm) was at the high temperature. Due to the coarse nature of the sphagnum, most seeds received some light, but at any given point in time a few may have been in darkness. However, at weekly intervals the contents of each bag were poured out on a table to facilitate examination of seeds for germination. After germination was monitored, seeds and sphagnum were returned to the bag, resulting in a re-shuffling of seeds with regard to their position in the sphagnum and thus the amount of light they received. Consequently, all seeds were in light part (or all) of the time the lights were on in the incubators. Each treatment consisted of three replicates of 50 seeds.



Figure 1. Embryo growth in seeds of *Podocarpus costalis* (A, B) and *Nageia nagi* (C, D). Longitudinal section of fresh seeds with a linear-shaped underdeveloped embryo (A, C) and of seeds with a fully grown embryo (B, D). Fleshy sarcotesta has been removed. The arrow in (A) and in (C) points to the micropyle; em, embryo; mgp, megagametophyte; st, sclerotesta. Scale bar = 2 mm.

Radicle emergence (radicle $\geq 2 \text{ mm}$ long) was recorded weekly for 8 weeks and shoot emergence was studied for an additional 6-week period with weekly recordings of seeds of *P. costalis*. Radicle emergence was recorded weekly for 20 weeks for seeds of *N. nagi*, but shoot emergence was not studied. Results are expressed as mean (\pm SE) germination percentage and as mean (\pm SE) germination time (MGT) in days. MGT = ($\Sigma n_i t_i$)/*N*, where n_i is the number of seeds germinated in t_i days from the beginning of the test, and *N* is the total number of germinated seeds at the end of the test (Naylor, 1981). MGT is a measure of the rate of germination and of the sharpness of the germination peak.

Embryo growth

Lengths of embryo and megagametophyte were measured in ten fresh seeds each of *P. costalis* and *N. nagi* and in ten seeds of each species in which the sclerotesta had split but none of the radicle had emerged (= critical embryo length for germination) (Baskin and Baskin, 2005). Seeds were dissected using a razor blade or scissors, and embryos were measured under a dissecting microscope equipped with a calibrated micrometer; representative sections were photographed.

Statistical analysis

Germination data based on number of seeds in a treatment were converted to percentages, and means (three replications) and standard errors were calculated. Statistical analysis of mean germination percentage and MGT was carried out using the GLM procedure of SAS, and means were compared by Least Significant Difference (LSD) (SAS Institute Inc., Cary, North Carolina, USA) and Microsoft Office Excel 2007. Percentage data were arcsine square-root transformed before analysis, but only non-transformed data are shown in Table 1 and Figs 2 and 3.

Table 1. Mean germination time (MGT) of fresh seeds of *Podocarpus costalis* and *Nageia nagi* incubated at various temperatures. MGT was calculated after 8 weeks' incubation of *P. costalis* seeds and after 20 weeks' incubation of *N. nagi* seeds. Means $(n = 3) \pm$ SE within a column followed by different letters differ significantly (LSD, $\alpha = 0.05$)

Incubation temperature (°C)	MGT of <i>Podocarpus</i> seeds (d)	MGT of <i>Nageia</i> seeds (d)
30/20	21.9 ± 0.8d	$53.4 \pm 1.6c$
25/15	$24.2 \pm 0.6c$	$68.3 \pm 4.7 bc$
20/10	$30.4 \pm 0.8b$	80.6 ± 9.2ab
15/6	$46.0 \pm 0.4a$	94.6 ± 14.7a
25	19.3 ± 0.6e	$50.2 \pm 2.4c$



Figure 2. Percentage (means \pm SE) radicle (A) and shoot (B) emergence of *Podocarpus costalis* seeds incubated at alternating temperatures of 30/20, 25/15, 20/10 and 15/6°C and at a constant temperature of 25°C. Incubation time 0 for shoot emergence starts after radicle emergence. Final percentages of radicle and shoot emergence among the incubation temperatures followed by different letters differ significantly (LSD, $\alpha = 0.05$).

Results

Embryo growth and germination

Embryos of fresh *P. costalis* seeds are linear-shaped and a suspensor links the embryo to the micropyle (Fig. 1A). Mean (\pm SE) lengths of embryos and megagametophytes in fresh seeds (n = 10) were 4.6 ± 0.5 mm and 10.6 ± 0.4 mm, respectively. Thus, the embryo length/megagametophyte length (E:M) ratio was 0.43. Embryo length just before radicle emergence was 7.1 ± 0.4 mm (Fig. 1B). Thus, both embryo length and E:M ratio increased to more than 1.5 times their values in fresh seeds.

Fresh seeds of *P. costalis* incubated at 30/20, 25/15, 20/10, 15/6 and 25° C germinated (radicle emergence) to 92, 90, 60, 3 and 99%, respectively, in 4 weeks. Extending the incubation time to 6 weeks increased seed germination to 96% at $20/10^{\circ}$ C and to 42° at $15/6^{\circ}$ C (Fig. 2A). Seeds incubated at $15/6^{\circ}$ C required

8 weeks for 90% germination. In *Podocarpus* seeds incubated continuously at the above five temperatures, cotyledons had emerged from >90% of the seeds at 30/20 and 25°C by 3 weeks following radicle emergence (Fig. 2B). MGT of *P. costalis* seeds ranged from $19.3 \pm 0.6 \text{ d}$ at 25°C to $46 \pm 0.4 \text{ d}$ at 15/6°C (Table 1). The optimal temperatures for maximum germination percentage and rate were 30/20 and 25°C.

Embryos in fresh seeds of *Nageia* are linear-shaped and a suspensor links the embryo to the micropyle (Fig. 1C). Mean (\pm SE) lengths of embryos and megagametophytes in fresh seeds (n = 10) were 7.4 \pm 0.8 and 15.4 \pm 0.6 mm, respectively. Thus, embryo length/megagametophyte length (E:M) ratio was 0.48. Embryo length just before radicle emergence was 10.3 \pm 0.6 mm (Fig. 1D). Thus, both embryo length and E:M ratio increased to 1.4 times their values in fresh seeds. It should be noted that *N. nagi* seeds consistently turned brown during incubation.

Fresh seeds of *N. nagi* incubated at 30/20, 25/15, 20/10, 15/6 and 25°C germinated (radicle emergence) to 24, 13, 0.7, 0 and 16%, respectively, after 4 weeks and to 91, 51, 17, 3 and 99%, respectively, after 12 weeks. Further, the *N. nagi* seeds incubated at 25/15, 20/10 and 15/6°C for 20 weeks germinated to 77, 29 and 7%, respectively (Fig. 3). MGT of *Nageia* seeds ranged from 50.2 \pm 2.4 days at 25°C to 94.6 \pm 14.7 days at 15/6°C (Table 1). The optimal temperatures for maximum germination percentage and rate were 30/20 and 25°C.

Discussion

An underdeveloped embryo is usually relatively small, and in many cases the seed has an embryo length/seed length (E:S) of < 0.5. Also, the small



Figure 3. Percentage (means \pm SE) radicle emergence of *Nageia nagi* seeds incubated at alternating temperatures of 30/20, 25/15, 20/10 and 15/6°C and at a constant temperature of 25°C. Final percentages of radicle emergence among the incubation temperatures followed by different letters differ significantly (LSD, $\alpha = 0.05$).

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embryo must grow inside the seed before the radicle can emerge (Grushvitzky, 1967; Baskin et al., 2006a). The embryo in freshly harvested seeds of P. costalis is small (E:M = 0.43) and increased in length by about 54% inside seeds before the radicle emerged; thus, the embryo is underdeveloped. Fresh Podocarpus seeds germinated to 90-99% at 30/20, 25/15 and 25°C in 3-4 weeks (radicle emergence) and to 97% at 20/10°C after 6 weeks, without any pretreatment. Based on the short delay of 2–3 weeks at suitable temperatures of 30/20, 25/15 and 25°C between time of radicle and shoot emergence (Fig. 2B) and the fact that seeds germinated within 30 d, we conclude that seeds of *P. costalis* have morphological dormancy.

The embryo in fresh seeds of N. nagi is small (E:M = 0.48) and increased in length by about 39% inside the seeds before radicle emergence occurred; thus, the embryo is underdeveloped. The highest germination of N. nagi seeds at 4 weeks of incubation was <25%, at 30/20°C, and it did not reach 50% (at 30/20 or 25°C) until week 7. Germination percentage at these two temperatures reached the maximum 99.3% after 14 (25°C) and 16 (30/20°C) weeks. Hong et al. (2009) also showed that seeds of this species germinated slowly at 25 and at 30/20°C. Thus, seeds of N. nagi required a much longer period of time than 1 month at optimal temperatures to germinate to high percentages, indicating that they have non-deep simple morphophysiological dormancy and need warm stratification to break it. The formula $C_{1b}B-C_{1b}$ describes this level of MPD (Baskin and Baskin, 2008), and it reads as follows. (1) Fresh seeds have an underdeveloped embryo (B) with non-deep physiological dormancy (C_1) of the type requiring warm stratification (b of subscript 1b) for dormancy break. (2) Following (or occurring simultaneously with) breaking of non-deep physiological dormancy, the underdeveloped embryo grows to full size. (3) The seed, now with a fully developed embryo, germinates at warm temperatures $(-C_{1b})$.

At the Taiwan Forestry Research Institute, seeds of P. costalis and N. nagi are generally stored in a moist medium for a short period of time before use. The moisture content of fresh seeds with sclerotesta was 51% in P. costalis and 56% in N. nagi. Hong and Ellis (1996) found that at maturity or at shedding the moisture content was between 36 and 90% for recalcitrant seeds and between <20 and 50% for orthodox seeds. The Royal Botanic Gardens Kew Seed Information Database (Royal Botanic Gardens Kew, 2008) lists seeds of N. nagi (reported as Podocarpus nagi) and several other species of Podocarpus, but not *P. costalis*, as being recalcitrant. However, although no details on the storage behaviour of P. costalis seeds are available, the moisture content of fresh seeds of this species suggests that they may be recalcitrant. retain their viability, and they lose viability rapidly if dried to a moisture content of <20-30% (Hong and Ellis, 1996). Therefore, on the basis of their seed moistures, both species studied may be expected to be recalcitrant, although this matter certainly requires further experimentation.

Underdeveloped embryos are considered to be primitive in seed plants (Martin, 1946; Grushvitzky, 1967; Baskin and Baskin, 1998; Nikolaeva et al., 1999; Forbis et al., 2002; Finch-Savage and Leubner-Metzger, 2006; Linkies *et al.*, 2010). However, this trait is not only present in extant primitive seed plants, but it can be found in advanced angiosperms. For example, an underdeveloped embryo is found in the most basal extant angiosperm Amborella (Baskin and Baskin, 2007) as well as in some species of the advanced order Dipsacales (Baskin et al., 2006b). The most primitive extant gymnosperms, i.e. Ginkgo biloba (West et al., 1970; Del Tredici, 2007; Shepperd, 2008) and cycads (Martin, 1946; Chien et al., 2012) have underdeveloped embryos. Among the extant Pinopsida, underdeveloped embryos occur in some clades but not in others. Quinn et al. (2002) investigated the phylogeny of conifers using the chloroplast genes *rbcL* and *matK* with *Ginkgo* as the outgroup. The first clade to diverge, the Pinaceae, has seeds with fully developed embryos. In addition to the Pinaceae, there are two other large clades in Pinopsida: (1) Araucariaceae and Podocarpaceae, and (2) Taxaceae and Cupressaceae. Now that underdeveloped embryos have been verified in seeds of Podocarpaceae, we can see that they occur in both of these clades, Podocarpaceae and Taxaceae. In their respective clades, both Podocarpaceae and Taxaceae are in advanced positions (Quinn et al., 2002). Thus, as in extant angiosperms, underdeveloped embryos are not restricted to the most basal groups. Unlike the advanced angiosperm clades, however, the Pinaceae do not have underdeveloped embryos.

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References

- Baskin, C.C. and Baskin, J.M. (1998) Seeds: ecology, biogeography, and evolution of dormancy and germination. San Diego, Academic Press.
- Baskin, C.C. and Baskin, J.M. (2005) Underdeveloped embryos in dwarf seeds and implications for assignment to dormancy class. Seed Science Research 15, 357-360.

- Baskin, C.C. and Baskin, J.M. (2007) A revision of Martin's seed classification system, with particular reference to his dwarf-seed type. *Seed Science Research* 17, 11–20.
- Baskin, C.C., Chien, C.T., Chen, S.Y. and Baskin, J.M. (2008) Germination of *Viburnum odoratissimum* seeds: a new level of morphophysiological dormancy. *Seed Science Research* 18, 179–184.
- Baskin, J.M. and Baskin, C.C. (2004) A classification system for seed dormancy. *Seed Science Research* 14, 1–16.
- Baskin, J.M. and Baskin, C.C. (2008) Some considerations for adoption of Nikolaeva's formula system into seed dormancy classification. *Seed Science Research* 18, 131–137.
- Baskin, J.M., Baskin, C.C., Chien, C.T. and Chen, S.Y. (2006a) Seed dormancy in the early diverging eudicot *Trochodendron aralioides* (Trochodendraceae). *Seed Science Research* 16, 71–75.
- Baskin, J.M., Hidayati, S.N., Baskin, C.C., Walck, J.L., Huang, Z.Y. and Chien, C.T. (2006b) Evolutionary considerations of presence of both morphophysiological and physiological seed dormancy in the highlyadvanced euasterids II order Dipsacales. *Seed Science Research* 16, 233–242.
- Cheng, G., Tang, X., Gao, H. and Shen, S. (2004) Dormancy mechanism and relieving techniques of seeds of *Taxus* cuspidata Sieb. et Zucc. Journal – Beijing Forestry University 26, 5–9 (in Chinese with English abstract).
- Chien, C.T., Kuo-Huang, L.L. and Lin, T.P. (1998) Changes in ultrastructure and abscisic acid level, and response to applied gibberellins in *Taxus mairei* seeds treated with warm and cold stratification. *Annals of Botany* 81, 41–47.
- Chien, C.T., Chen, S.Y., Chang, S.H. and Chung, J.D. (2012) Seed germination and storage of *Cycas taitungensis* (Cycadaceae). *Taiwan Journal of Forest Science* **27**, 1–11.
- Del Tredici, P. (2007) The phenology of sexual reproduction in *Ginkgo biloba*: ecological and evolutionary implications. *The Botanical Review* 73, 267–278.
- **Devillez, F.** (1978) Influence de la température sur la postmaturation et la germination des graines de l'if (*Taxus baccata* L.). Bulletin de la Classe des Sciences l'Academie Royale de Belgique **64**, 203–218.
- Dodd, M.C. and Van Staden, J. (1981) Germination and viability studies on the seeds of *Podocarpus henkelii* Stapf. *South African Journal of Science* 77, 171–174.
- Ferrandis, P., Bonilla, M. and Osorio, L.C. (2011) Germination and soil seed bank traits of *Podocarpus angustifolius* (Podocarpaceae): an endemic tree species from Cuban rain forests. *International Journal of Tropical Biology* 59, 1061–1069.
- Figueroa, J.A. (2003) Seed germination in temperate rain forest species of southern Chile: chilling and gapdependency germination. *Plant Ecology* **166**, 227–240.
- Figueroa, J.A. and Lusk, C.H. (2001) Germination requirements and seedling shade tolerance are not correlated in a Chilean temperate rain forest. *The New Phytologist* **152**, 483–489.
- Finch-Savage, W.E. and Leubner-Metzger, G. (2006) Seed dormancy and the control of germination. *The New Phytologist* **171**, 501–523.
- Forbis, T.A., Floyd, S.K. and de Querioz, A. (2002) The evolution of embryo size in angiosperms and other seed plants: implications for the evolution of seed dormancy. *Evolution* **56**, 2112–2125.

- Grushvitzky, I.V. (1967) After-ripening of seeds of primitive tribes of angiosperms, conditions and peculiarities. pp. 329–336 + 8 figures *in* Borris, H. (Ed.) *Physiologie, ökologie und biochemie der keimung*. Greifswald, Germany, Ernst-Moritz-Arndt Universität.
- Hong, K.Y., Jhuang, P.H., Chien, C.T., Huang, Y.J. and Fan, Y.B. (2009) Germination and storage of seeds of the endangered plant, *Nageia nagi. Scientific Agriculture* 57, 170–175 (in Chinese with English abstract).
- Hong, T.D. and Ellis, R.H. (1996) *A protocol to determine seed storage behaviour.* Rome, International Plant Genetic Resources Institute.
- International Seed Testing Association (2007) *ISTA handbook* on moisture determination (1st edition). Nijënstein, H.; Nydam, J.; Don, R.; McGill, C. (Eds) Bassersdorf, Switzerland, International Seed Testing Association.
- Li, H.L., Keng, H., Yang, Y.P. and Lu, S.Y. (1994) Podocarpaceae. pp. 557–566 *in* Editorial Committee of the Flora of Taiwan, (Ed.) *Flora of Taiwan*, vol. 1, (2nd edition). Taiwan, Editorial Committee of the Flora of Taiwan.
- Linkies, A., Graeber, K., Knight, C. and Leubner-Metzger, G. (2010) The evolution of seeds. *The New Phytologist* 186, 817–831.
- Liu, D., Yu, H.L. and Guo, H.H. (2011) An analysis of dormancy and dormancy release in *Taxus chinensis* var. *mairei* seeds. *Seed Science and Technology* **39**, 29–43.
- Martin, A.C. (1946) The comparative internal morphology of seeds. *The American Midland Naturalist* **36**, 513–660.
- Meyen, S.V. (1984) Basic features of gymnosperm systematics and phylogeny as evidenced by the fossil record. *The Botanical Review* **50**, 1–111.
- Naylor, R.E.L. (1981) An evaluation of various germination indices for predicting differences in seed vigour in Italian ryegrass. *Seed Science and Technology* 9, 593–600.
- Negash, L. (1992) *In vitro* methods for the rapid germination of seeds of *Podocarpus falcatus*. *Ethiopian Journal of Science* **15**, 85–97.
- Ng, F.S.P. (1992) Manual of forest fruits, seeds and seedlings, vol. 2. Kuala Lumpur, Forest Research Institute Malaysia.
- Nikolaeva, M.G. (1977) Factors controlling the seed dormancy pattern. pp. 51–74 *in* Khan, A.A. (Ed.) *The physiology and biochemistry of seed dormancy and germination*. Amsterdam, North-Holland.
- Nikolaeva, M.G., Rasumova, M.V. and Gladkova, V.N. (1985) *Reference book on dormant seed germination*. Danilova, M.F. (Ed.). Leningrad, 'Nauka' Publishers (in Russian).
- Nikolaeva, M.G., Lyanguzova, I.V. and Pozdova, L.M. (1999) *Biology of seeds*. St. Petersburg, V.L. Komarov Botanical Institute, Russian Academy of Sciences (in Russian).
- Quinn, C.J., Price, R.A. and Gadek, P.A. (2002) Familial concepts and relationships in the conifers based on *rbcL* and *matK* sequence comparisons. *Kew Bulletin* 57, 513–531.
- Royal Botanic Gardens Kew (2008) Seed Information Database (SID). Version 7.1. Available at http://data. kew.org/sid/ (accessed May 2008).
- Shepperd, W.D. (2008) Ginkgoaceae–Ginkgo family. Ginkgo biloba L. pp. 559–561 in Bonner, F.T.; Karrfalt, R.P. (Eds) The woody plant seed manual. USDA Forest Service, Agriculture Handbook 727.

- Wang, B.S.P., Lin, T.P. and Chang, T.T. (1998) Control of fungal growth with sphagnum for cold stratification and germination of tree seeds. *Taiwan Journal of Forest Science* 13, 101–108.
- West, W.C., Frattarelli, F.J. and Russin, K.J. (1970) Effect of stratification and gibberellin on seed germination in *Ginkgo biloba*. Bulletin of the Torrey Botanical Club **97**, 380–384.
- Yang, C.J., Chien, C.T., Liao, Y.K., Chen, S.Y., Baskin, J.M., Baskin, C.C. and Kuo-Huang, L.L. (2011) Deep simple morphophysiological dormancy in seeds of the basal taxad *Cephalotaxus*. Seed Science Research 21, 215–226.
- Zhang, Z.-Q., Liao, W.-B., Zhong, L. and Chen, Z.-M. (2000) Biological study on seed germination of *Taxus mairei*. *Forest Research* **13**, 280–285 (in Chinese with English abstract).