

Low gains in ecosystem carbon with woody plant encroachment in a South African savanna

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Abstract: Total ecosystem carbon storage has frequently been found to increase with woody encroachment in savannas. However the loss of grass roots associated with woody encroachment can lead to a decrease in below-ground carbon storage which is not compensated for by an increase in above-ground carbon. To investigate how the extent of total woody cover affected ecosystem carbon, soil and above-ground carbon storage along eight thicket–savanna and five forest–grassland boundaries were measured. To investigate whether changes in soil carbon concentrations were related to the percentage of C4 (grass) roots to total roots and root quantity and quality, we measured fine-root biomass, root C:N ratios, root N, and % C4 roots at three different depths across thicket patches of different ages (n = 189). Forests contained significantly more carbon than adjacent grasslands in both above-ground carbon (mean difference 12.1 kg m⁻²) and in the top 100 cm of the soil (mean difference 4.54 kg m⁻²). Thickets contained significantly more above-ground carbon than adjacent savannas (3.33 kg m⁻²) but no significant differences in soil carbon were evident. Total fine-root biomass appeared to be more important than root quality (root C:N) in determining soil carbon concentrations during the encroachment process (i.e. in thicket of different ages). Similarly for thickets, the % C4 roots had no significant effect on soil carbon concentrations. In conclusion, thicket invading into open savanna vegetation did not lead to significant gains in ecosystem carbon at this study site. Significant gains were only evident in mature forest, suggesting that the process may take place very slowly.

Key Words: above-ground carbon, ecosystem functioning, fine-root biomass, root quality, soil carbon concentration, soil carbon storage

INTRODUCTION

Many African savannas are gaining woody biomass despite differing management practices such as variable fire regimes and stocking rates (Eckhardt *et al.* 2000, Higgins *et al.* 1999, 2007, Wigley *et al.* 2010). In fact woody encroachment, also referred to as woody plant invasion or the expansion of woody species into grassland and savanna, has been reported at locations worldwide in the past few decades (see Eldridge *et al.* 2011 for a review). The causes of woody encroachment are not clear and could include local drivers such as variable land management or a global driver such as increasing atmospheric CO₂ (Bond 2008, Bond & Midgley 2012, Bond *et al.* 2003, Hoffmann *et al.* 2000). Regardless of the drivers, the implication of the widespread woody

expansion is that large areas of grassland and savanna are in the process of ‘thickening up’ to become woodland or forest. Increasing woody encroachment influences rangeland use, biodiversity and ecosystem functioning, including hydrology and nutrient cycling at both the local and landscape scale (Archer *et al.* 2000, Parr *et al.* 2012). At the regional scale, changes in tree cover could have large feedbacks to the earth–atmosphere system (Archer *et al.* 2000, Asner *et al.* 2004, Beerling & Osborne 2006).

Woody encroachment often leads to increases in ecosystem carbon (C) storage (Geesing *et al.* 2000, Hibbard *et al.* 2001, Hudak *et al.* 2003). Jackson *et al.* (1996) found significant variation in SOC storage in ecosystems with different dominant plant functional types (i.e. grass, shrub and tree) and explained the differences as an effect of varying plant production and decomposition rates which ultimately determine SOC storage. Subsequent work has found significant changes in SOC profiles among vegetation types caused by differences in root distribution and above- and

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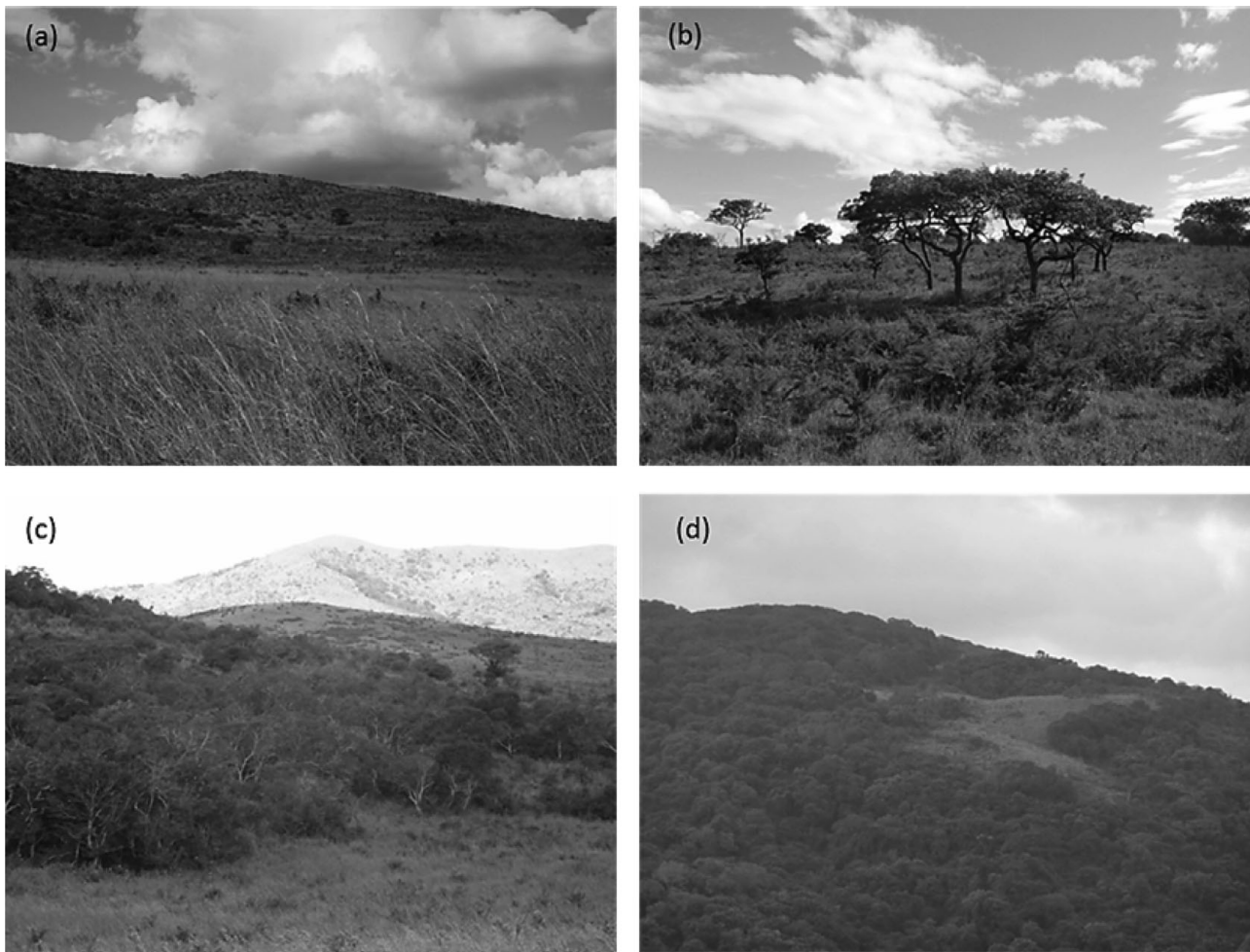


Figure 1. Grassland (a), savanna (b), thicket (c) and forest (d) in the Hluhluwe section of the Hluhluwe-iMfolozi Park, South Africa.

below-ground allocation patterns (Jobbágy & Jackson 2000). Expansion of shrubs in North American rangeland in Texas has led to huge increases in SOC as a result of increased biomass below ground and decreases in turnover and decomposition (Archer *et al.* 2000, Boutton *et al.* 1998). Hibbard *et al.* (2001) mention that the contribution of below-ground inputs to observed increases in soil C is not clear but that inputs from fine roots far exceeded inputs from litter in their study which investigated the invasion of *Prosopis* into Texan savannas.

In certain ecosystems, C and nitrogen (N) storage have been found to decrease as above-ground gains are offset by below-ground soil organic matter losses when grass roots disappear at very high woody densities (Hudak *et al.* 2003, Jackson *et al.* 2002). At mesic (but not semi-arid) woody-dominated sites in North America, losses of soil C stocks with encroachment were of such magnitude when compared with grass-dominated sites that the above-ground gains in plant biomass could not compensate for losses of soil C (Jackson *et al.* 2002). This pattern

has also been observed in a southern African semi-arid savanna where soil C storage decreased at very high woody densities because the growth of understorey grass was inhibited (Hudak *et al.* 2003). The main aim of this study was to test whether soil C storage will increase with woody encroachment. If so, we would expect old established forest and encroaching thicket to contain the most C, and the more grassy savannas and grasslands to contain the least. Our second aim was to investigate which factors are important in driving changes in observed C concentrations and whether these changes are related to the percentage of grass roots (C4) to total roots (C4 and C3, hereafter % C4 roots) in the soil. If soil C concentrations are positively related to % C4 roots, then we expect that C concentrations should decrease in the oldest thicket as only a small percentage of total roots were grass roots (~10%). As previous work has not offered a mechanism to explain the correlation between low % C4 roots and low soil C concentrations, we also tested whether changes in C concentrations were related to changes in fine-root biomass, root N as well as root C : N ratios.

Table 1. Information on extensively sampled paired boundary sites from Hluhluwe-iMfolozi Park, including position and soils; F–G refers to forest–grassland boundaries and T–S to thicket–savanna boundaries. Classification of soil form follows Fey (2010); that of soil series follows MacVicar *et al.* (1977). WRB = World reference base.

Site code	Latitude	Longitude	Soil group	Soil form	Soil series	WRB soil classification
F–G 1	–28.0792	32.04254	Oxidic	Hutton	Doveton	Ferralsol
F–G 2	–28.0630	32.05212	Oxidic	Hutton	Doveton	Ferralsol
F–G 3	–28.0609	32.05182	Oxidic	Hutton	Doveton	Ferralsol
F–G 4	–28.0595	32.04981	Oxidic	Hutton	Doveton	Ferralsol
F–G 5	–28.0721	32.03355	Oxidic	Hutton	Doveton	Ferralsol
T–S 1	–28.1206	32.03911	Melanic	Mayo	Mayo	Phaeozem
T–S 2	–28.1216	32.03329	Oxidic	Hutton	Doveton	Ferralsol
T–S 3	–28.0554	32.13183	Melanic	Mayo	Mayo	Phaeozem
T–S 4	–28.0650	32.11923	Oxidic	Hutton	Doveton	Ferralsol
T–S 5	–28.0668	32.12049	Oxidic	Hutton	Doveton	Ferralsol
T–S 6	–28.0738	32.13079	Oxidic	Hutton	Doveton	Ferralsol
T–S 7	–28.0887	32.08183	Melanic	Mayo	Mayo	Phaeozem
T–S 8	–28.1059	32.04944	Oxidic	Hutton	Doveton	Ferralsol

METHODS

Study site

This study was conducted in South Africa, in the Hluhluwe-iMfolozi Park (HiP) complex (28°00′–28°26′S; 31°43′–32°09′E, Figure 1), which consists of the Hluhluwe Game Reserve (225 km²) to the north, the iMfolozi Game Reserve (447 km²) to the south and a corridor (227 km²) joining the two (Whateley & Porter 1983). The terrain of the park is hilly with altitudes between 60 and 450 m. Rainfall in the park increases with altitude and the higher-altitude Hluhluwe section receives from 700–1000 mm y⁻¹ on the highest hilltops and supports a mesic savanna (Balfour & Howison 2001).

The sampling was undertaken in two steps; an extensive approach to compare C stocks and an intensive approach to explore rooting differences and their contributions to C concentrations. During the extensive sampling, we sampled soils along eight thicket-savanna boundaries, the ages of the thicket patches ranging from 30–70-y-old. We also sampled along five forest–grassland boundaries. For the purposes of investigating the effect of changes in root distribution on soil C concentration (i.e. intensive approach), sampling took place at three different sites representing a chronosequence of time since invasion; 10-y-old thicket, 40-y-old thicket and 70-y-old thicket. The soils at the study sites are derived from basalts. They vary from oxidic soils (Fey 2010) with black topsoil overlying a red subsoil, to melanic soils (Fey 2010) with a black structured topsoil and similar subsoil of varying depth onto weathered rock. Forest and grassland sites were always on oxidic soils. Savanna and thicket sites had more variable soils with both oxidic and melanic soils, with the latter more common on steeper sloping terrain (Table 1). The thicket sites sampled in the intensive sampling approach were situated on oxidic soils.

The vegetation is subtropical and consists predominantly of Northern Zululand Sourveld (Mucina & Rutherford 2006). The Hluhluwe-iMfolozi Park complex provided an ideal situation to conduct this study as the area is characterized by a mixture of savanna (from open grassland to open woodland) and forest (including thicket which is also referred to as dry seasonal forest in the neotropics (Pennington *et al.* 2009, Ratnam *et al.* 2011); a single-layer woodland typically with a dense non-grassy understorey). There are sharp boundaries between the grassy and forested biomes (see Figure 1 for examples of different vegetation types), which are often unrelated to the underlying soils.

The forest patches we sampled are classified as Scarp Forest (FOz 5, Mucina & Rutherford 2006) and although these may burn under unique conditions, their core areas seem to be quite stable. Mucina & Rutherford (2006) define these forests as a tall (15–25 m), species-rich and structurally diverse, multi-layered vegetation, with well-developed canopy and understorey tree layers, but with a poorly developed herb layer. The forested sites were dominated by large canopy species such as *Protorhus longifolia* (Bernh.) Engl., *Harpephyllum caffrum* Bernh., *Combretum kraussii* Hochst., *Celtis africana* Burm.f., and a mid-storey of *Englerophytum natalense* (Sond.) Heine & J.H. Hemsl. and *Maytenus mossambicensis* (Klotzsch) Blakelock, and had little or no herbaceous understorey growth. West *et al.* (2000) used ¹³C isotopes to establish whether the forests in Hluhluwe are remnants of much larger forests or whether the forest invaded into what was previously grassland. Although they did not date the C in their study, it was shown that the forest used to be grassland or grass-dominated savanna sometime in the distant past (>2000 y, L. Gillson, unpubl. data). This led us to assume that the forest–grassland boundaries we sampled on were temporally stable. Our grassland sites had a low percentage of woody species present (~5% of the total area).

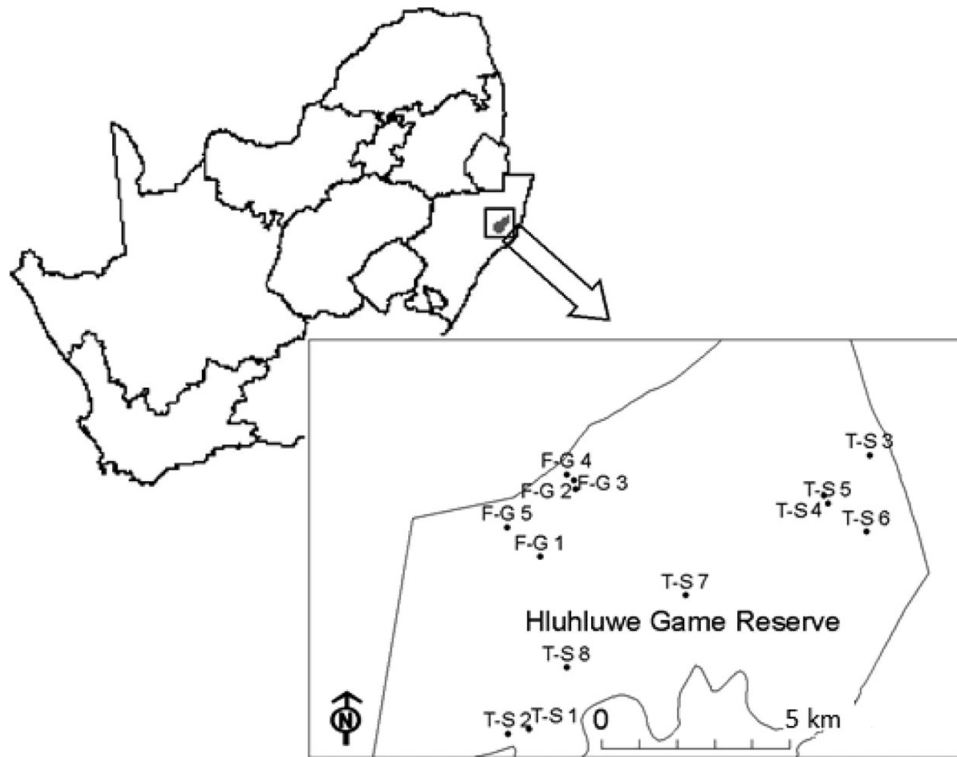


Figure 2. A map of South Africa indicating the position of Hluhluwe-iMfolozi Park with the Hluhluwe section in the inlay, and the 26 extensively sampled sites; (F-G refers to five paired forest and grassland sites and T-S to eight paired thicket and savanna sites).

Extensive woody or thicket encroachment has occurred throughout the study area over the past half-century (Balfour & Midgley 2008, Skowno *et al.* 1999, Watson 1995, Wigley *et al.* 2010). The vegetation types of these encroached areas are classified by Mucina & Rutherford (2006) as Northern Zululand Sourveld. According to Low & Rebelo (1996) thicket is a type of vegetation transitional between savanna and forest; almost impenetrable, generally not divided into strata and with variable herbaceous cover. In our study area, thicket consisted of savanna and forest species, typically with a single layer of woody plants which seldom exceeded 6–8 m in height. These thickets are often interspersed with small openings in the canopy where shade-tolerant and fire-intolerant grass species are able to persist (Parr *et al.* 2012). In our chronosequence of thickets the older thicket patches are dominated by trees that are generally shorter and smaller in diameter than the forest species, such as *Euclea racemosa* subsp. *schimperi* A.D.C.F. White, *Schotia capitata* Bolle, *Vachellia (Acacia) robusta* Burch and *Ziziphus mucronata* Willd., and thick herbaceous understorey with species of Acanthaceae often prominent. The newly invaded thicket patch is dominated largely by *Vachellia (Acacia) karroo* Hayne and *Dichrostachys cinerea* (L.) Wight & Arn. and has a few small individuals of species typically found in thickets present such as *Euclea racemosa* subsp. *schimperi*.

Sampling

Sampling took place during the growing season of 2009. Extensive sampling took place along eight thicket–savanna boundaries and five forest–grassland boundaries (see Figure 2 for map). We chose vegetation boundaries which occurred on homogeneous soils and landscape positions (Table 1) to ensure that differences in soil would not impact our results. A 1 × 2-m pit was dug 30 m perpendicular to the boundary at each site (13 boundaries = 26 pits). Samples were taken at set depth intervals of 0–2.5 cm, 2.5–7.5 cm, 7.5–15 cm, 15–30 cm, 30–50 cm, 50–75 cm, 75–100 cm (26 pits × 7 depths = 182 soil samples). In order to determine bulk density a known volume was removed from each layer using a core sampler. Known volumes were dried in an oven at 70 °C until constant weight. They were weighed and sieved to remove all roots and stones, the weight and volume of which were measured. The bulk density of each sample was calculated by dividing the dry mass of soil (minus roots and rocks) by the volume of soil (minus roots and rocks). For the purposes of comparisons between vegetation types, soil layers were combined into intervals 0–15 cm, 15–30 cm and 30–100 cm.

In order to make rapid assessments of terrestrial carbon stocks it is necessary to develop a quick method of estimating above-ground biomass, and one way of doing

this is to find a way to convert stand basal area to biomass, as stand basal area can be determined quickly and easily using a Bitterlich wedge (Šálek & Zahradník 2008). At each site, we sampled basal area at four locations 25 m apart on three parallel transects 25 m apart (12 measurements \times 26 sites = 312 basal areas). Basal area is widely used to estimate the biomass of trees, but the only allometric relationships that exist for this region are for individual trees. We followed the approach of Midgley & Seydack (2006) in using an equation developed for neotropical forests by Baker *et al.* (2004) to estimate woody biomass. Baker *et al.* (2004) estimated above-ground biomass (AGB) of single trees using the equation taken from Chambers *et al.* (2001):

$$AGB = \frac{p}{0.67} e^{[0.33(\ln dbh) + 0.93(\ln dbh)^2 - 0.12(\ln dbh)^3 - 0.37]},$$

where dbh is diameter at breast height. We then regressed the stand basal area against stand biomasses reported in Tables 1 and 5 in Baker *et al.* (2004) to get the equation:

$$AGB = \frac{p}{0.67} \times (11.2 \times \text{Stand Basal Area}) + 2.12 \quad (R^2 = 0.96),$$

where ρ is the mean wood density of the stand in our study, and 0.67 is the mean wood density for the neotropical study. This standardization was done in order to remove possible inaccuracy due to differences in mean wood density in our study. However, we determined a mean wood density of 0.672 g cm⁻³ for our study area (H. Beckett, unpubl. data), so the standardization made little difference. This equation has not been verified for the Hluhluwe area, so it must be used with caution. A carbon : biomass ratio of 0.5 was used to estimate how much C was in the above-ground woody biomass (Martin & Thomas 2011, Penman *et al.* 2003).

Grass biomass was estimated using a disc pasture meter (DPM) (Bransby & Tainton 1977). The height measurements were converted into biomass measurements using the method developed by Waldram *et al.* (2008) for the Hluhluwe-iMfolozi area. For the grass, a ratio of 0.37 was used to convert biomass to C (mean of 36.8% for 63 grass samples from Hluhluwe; C. Coetsee, unpubl. data).

Intensive sampling took place at three different sites; 10-y-old thicket, 40-y-old thicket and 70-y-old thicket. The recently invaded thicket patch was part of a burn trial and fire has been excluded from this patch for 10 y. The other thicket patches had not burned since establishment and both were approximately 8 ha in size. Previous studies have measured the distribution of soil C and tree : grass roots through the profile and we combined the methods of Hudak *et al.* (2003), Maclaran & McPherson (1995) and Mordelet *et al.* (1997). At each site, we sampled at seven

locations 10 m apart on three parallel transects, spaced 100 m apart. In order to test whether changes in soil C concentration were related to changes in the overlying vegetation, at each sampling location we noted whether a canopy was present, the height of the lowest and highest canopy and the distance to the nearest woody plant taller than 1 m and taller than 4 m. We also noted the distance to the nearest grass tuft. As the distance to the closest grass tuft or closest tree was respectively very small or very large in the grassland sites, distances to tree and grass tufts were divided into classes. Furthermore, we collected soil samples for soil C measurements, and extracted roots which we used to measure root biomass, root C : N ratios, root N and per cent C₄ roots.

Soil samples were extracted with a stainless steel soil auger (7.2 cm diameter) and four depths were sampled at each site; 0–10 cm, 10–20 cm and 20–30 cm (3 sites \times 21 sampling points \times 3 depths = 189 soil samples). The soil was then sieved using a 2-mm sieve, to remove the bulk of the roots. After soils were sifted, a subsample was taken for the soil C concentration analysis; thereafter soils were submerged in water and floating roots extracted using a 1-mm sieve (after the methods of Aerts *et al.* 1992, Hibbard *et al.* 2001). Fine roots only included roots less than 2 mm in diameter (from both dry sieving and wet sieving) and total root biomass was taken as all the root material collected from each soil core.

Soils for soil C analysis were air-dried and sieved before transportation to the laboratory. Root samples were dried at 60 °C for 3 d and finely ground using a rotary hammer mill (3 sites \times 21 sampling points \times 3 depths = 189 root samples).

Laboratory analyses

$\delta^{13}\text{C}$ values can be used to calculate the percentage of C₃ (i.e. woody and forb) and C₄ (i.e. grass) fine roots in the sample by using the following equation (adapted from Still *et al.* 2003):

$$\% \text{C}_{\text{grass}} = (\delta^{13}\text{C}_{\text{tree}} - \delta^{13}\text{C}_{\text{measured}}) / (\delta^{13}\text{C}_{\text{tree}} - \delta^{13}\text{C}_{\text{grass}}) \times 100$$

where % C_{grass} is the per cent C₄ contribution, $\delta^{13}\text{C}_{\text{tree}}$ is the carbon isotopic composition of C₃ vegetation, $\delta^{13}\text{C}_{\text{grass}}$ is the carbon isotopic composition of C₄ vegetation, and $\delta^{13}\text{C}_{\text{measured}}$ is the isotopic composition of the measured sample. The root samples prepared as described above were combusted in a Carlo-Erba system (Carlo Erba NCS 2500 Elemental Analyser, Carlo Erba Instruments, Milan, Italy), analysed on a GC-IRMS (Finnigan MAT 252 IRMS, Finnigan, Bremen, Germany) and results were reported relative to the internationally accepted carbonate isotope standard PDB (Chicago Pee Dee Belemnite). Carbon

Table 2. Soil bulk density (mean \pm SE, g cm⁻³) for four vegetation types at three depths in the Hluhluwe section of the Hluhluwe-iMfolozi Park.

Depth (cm)	Grassland	Savanna	Thicket	Forest
0–15	1.17 \pm 0.05	1.14 \pm 0.09	1.05 \pm 0.09	1.02 \pm 0.10
15–30	1.17 \pm 0.06	1.09 \pm 0.03	1.06 \pm 0.05	1.15 \pm 0.07
30–100	1.20 \pm 0.06	1.09 \pm 0.04	1.15 \pm 0.05	1.19 \pm 0.03

¹³C/¹²C ratios are calculated relative to this standard from the equation:

$$\delta^{13}\text{C} = ((R_{\text{sample}}/R_{\text{Std}}) - 1) \times 1000$$

where R_{sample} and R_{Std} are the ¹³C/¹²C ratios of the sample and the standard, respectively. Replicate samples were reproducible to 0.25%. Root C concentrations (intensive sampling) and carbon for total ecosystem C (grass, wood and soils – extensive sampling) were measured with the same system as ¹³C.

Soils sampled during the intensive sampling were analysed for total organic C concentration at the Institute for Plant Production, Elsenburg, Stellenbosch, using the Walkley–Black method (Walkley 1947). The coefficient of variation for repeated samples was 0.07. For the extensive sampling, bulk density data (Table 2) were used to convert C concentration to C content.

Statistical analyses

The statistical package R was used throughout for all statistical analyses (R Development Core Team, <http://www.R-project.org>). To investigate whether there was a trend in how soil organic C (SOC) and above-ground carbon (AGC) responded with an increase in trees, we used a linear regression model to predict differences in soil C and AGC between grassy vegetation and the adjacent woody vegetation at different depths (0–15 cm, 15–30 cm, 30–100 cm). Forest–grassland boundaries were compared separately to thicket–savanna boundaries. All boundaries were on homogeneous soils so differences in C amounts could be assumed to be largely due to differences in vegetation type. To investigate how root quality and quantity changed with woody encroachment, we used a two-way ANOVA to predict root C:N ratios and fine-root mass with site, depth and the pairwise interaction. Values were log-transformed before analyses. We used Tukey HSD to test for differences between levels of each factor. The percentage of C₄ grass was compared amongst sites by using the non-parametric, Kruskal–Wallis test.

We used a generalized least squares model in the R package *nlme* to test which factors were important in estimating the response variable, soil C concentration. We only included values to 30 cm as no root data were collected at deeper depths. The original model included all 12 explanatory variables; site, soil depth, distance to

the closest tree over 4 m, distance to the closest shrub over 1 m, presence/absence of a tree canopy, distance to the closest grass tuft, root N, root C, root C:N ratios, per cent C₄ roots, total root weight and fine-root mass. A generalized least-squares model was used as the linear model output revealed that assumptions of normality, and non-constant variance were not met. The results of a VIF test (R package *MuMIn*) revealed no collinearity in the data and the generalized least squares model assumption of non-autocorrelation was met. We used the dredge function in the R package *MuMIn* to validate output of the generalized least-squares model.

RESULTS

We investigated mean differences between paired woody and grassy sites by using linear regression. Forests contained 12.1 \pm 0.73 kg m⁻² more AGC than their adjacent grasslands (Figure 3a, $t = 16.6$, $P < 0.0001$). Thickets contained significantly more (Figure 3a, $t = 6.10$, $P < 0.0001$) AGC (3.33 \pm 0.59 kg m⁻²) than the adjacent savannas, although the difference was not as pronounced as between forests and grasslands.

In the top 15 cm of soil, forests contained 1.32 \pm 0.86 kg m⁻² more soil C than grasslands, this difference was not significantly different from zero (Figure 3b, $t = 1.53$, $P = 0.15$). Thickets contained 1.96 \pm 0.68 kg m⁻² more soil C than savannas in the top 15 cm of soil, significantly different from zero (Figure 3b, $t = 2.87$, $P = 0.02$). Forests contained 0.87 \pm 0.21 kg m⁻² more soil C than adjacent grasslands in the 15–30 cm layer, which was found to be significantly different from zero (Figure 3c, $t = 4.00$, $P = 0.002$). Thickets contained only 0.01 \pm 0.17 kg m⁻² more soil C than adjacent savannas in the 15–30 cm layer, which was not significantly different (Figure 3c, $t = 0.05$, $P = 0.96$). Forests contained 2.35 \pm 1.00 kg m⁻² more soil C than adjacent grasslands at a depth of 30–100 cm, which was found to be significantly different from zero (Figure 3d, $t = 2.34$, $P = 0.04$). On the other hand, thickets contained only 0.52 \pm 0.79 kg m⁻² more SOC than adjacent savannas at a depth of 30–100 cm, which was not significantly different from zero (Figure 3d, $t = 0.66$, $P = 0.18$).

When we investigated differences in the entire soil profile (0–100 cm), we found that forests contained 4.54 \pm 1.70 kg m⁻² more soil C than grasslands, which was significantly higher than a difference of zero ($t = 2.67$, $P = 0.02$). Thickets contained 2.50 \pm 1.34 kg m⁻² more soil C than savannas in the top 100 cm of soil, which was not significantly different from a difference of zero ($t = 1.86$, $P = 0.09$). Soil C:N ratios did not vary in a predictable manner; for instance, forest had the lowest C:N ratios in the surface soil, but the highest C:N ratios at 30–100 cm (Table 3).

Table 3. Soils C : N ratios (means \pm SE) for four vegetation types at three depths in the Hluhluwe section of the Hluhluwe-iMfolozi Park.

Depth (cm)	Grassland	Savanna	Thicket	Forest
0–15	15.9 \pm 0.96	14.6 \pm 0.61	14.1 \pm 0.32	13.2 \pm 0.75
15–30	16.7 \pm 1.91	15.4 \pm 0.67	14.5 \pm 0.50	14.6 \pm 1.49
30–100	17.8 \pm 1.18	13.8 \pm 0.57	13.2 \pm 0.70	18.1 \pm 1.56

To determine whether woody encroachment and the associated changes in woody cover have led to changes in the quality and quantity of roots, we measured root C : N ratios, per cent C₄ roots and fine-root biomass. The output of the ANOVA revealed that root C : N ratios were similar in the 10-y-old thicket (C : N 33.6) and the 40-y-old thicket (C : N 31.7), but lower in the 70-y-old thicket (C : N 25.7; $F_{2,180} = 19.4$, $P < 0.0001$). In general, root C : N ratios increased with depth, root C : N ratios in the top 10 cm were significantly lower than the C : N ratios found at 20- and 30-cm depths (26.9 vs. 31.1 and 33.0, $F_{2,180} = 10.8$, $P < 0.0001$, Figure 4a). Of the fine-root biomass, 55.2% belonged to C₄ grass in the 10-y-old thicket, 22.4% in the 40-y-old thicket and 11.8% in the 70-y-old thicket (Figure 4b). Interactions between site and depth were not significant in the ANOVA for root C : N ratios ($P = 0.91$) and fine-root biomass ($P = 0.32$). The older thicket patches had the highest and similar fine-root biomass (10.3 \pm 0.39 kg m⁻² and 10.9 \pm 0.56 kg m⁻² for the 40-y-old and 70-y-old thickets), while the recently invaded thicket patch had lower fine-root biomass

Table 4. The output of a generalized least squares model is shown with significance levels. Three explanatory variables (site, depth and fine-root biomass, g m⁻²) explained 70% of the variation in soil C concentration for thickets in the Hluhluwe section of the Hluhluwe-iMfolozi Park, significance levels indicated by *** $P < 0.001$, ** $P < 0.01$.

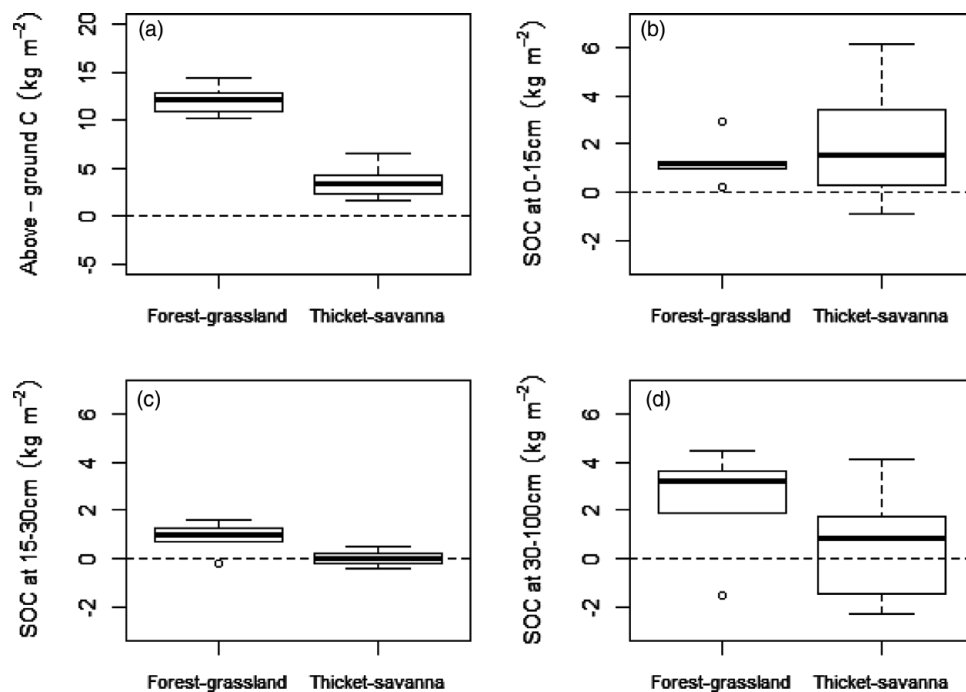
Variables	F-ratios
r^2_{adj}	0.70
Site	5.27**
Depth	233***
Root N	0.001
Fine-root biomass	22.1***

(3.66 \pm 0.12 kg m⁻²; $F_{2,180} = 64.2$, $P < 0.0001$, Figure 5). An interesting pattern that can be seen in Figure 5 is that fine-root biomass is concentrated in the shallower soil depths in recently invaded thicket patches but increases with depth in the older thicket patches.

Our analyses indicated that three explanatory variables explained 70% of the variation within soil C concentration (Table 4); these included sites, soil depth and the mass of fine roots. The amount of C₄ or grass roots had no effect on soil C concentrations.

DISCUSSION

Changes in land use have variable effects on soil C stocks; this is reflected in the findings of two meta-analyses

**Figure 3.** Differences (mean \pm SE, kg m⁻²) in above-ground and below-ground carbon (SOC) between paired forest–grassland and thicket–savanna plots in the Hluhluwe section of the Hluhluwe-iMfolozi Park.

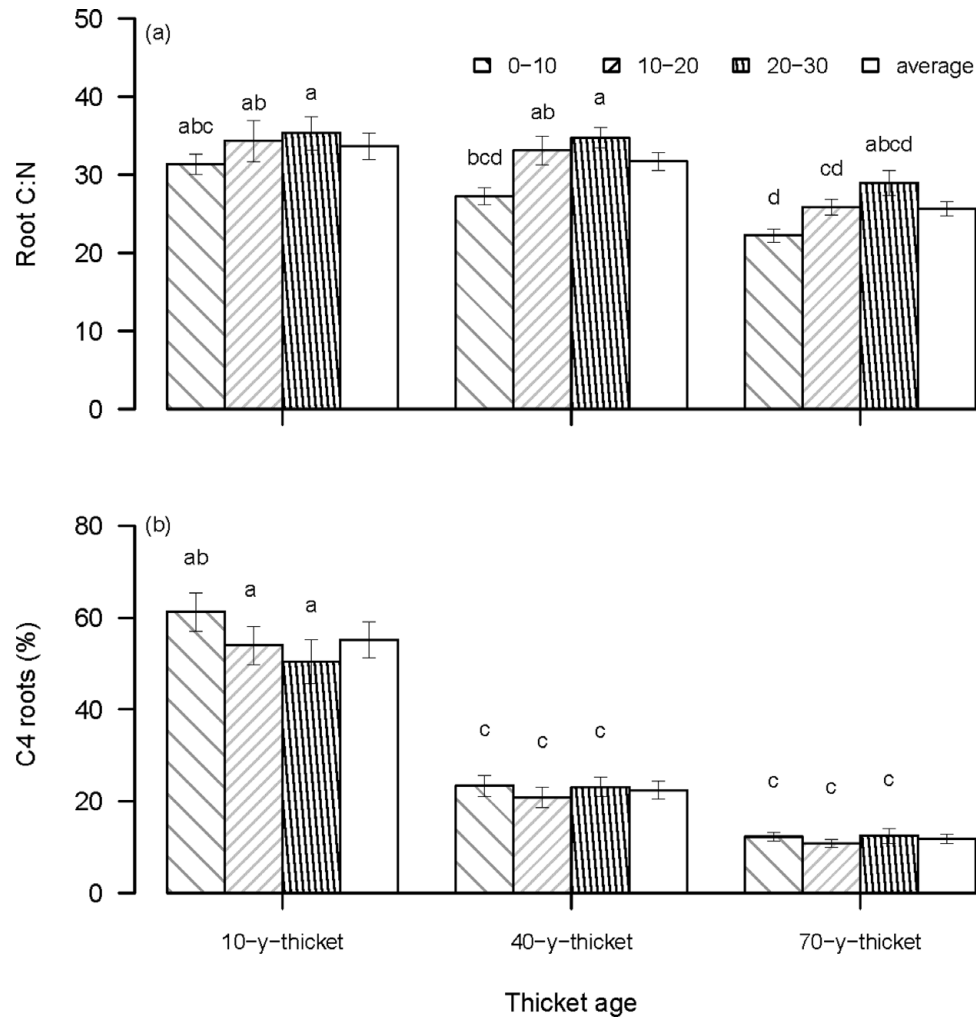


Figure 4. Root C : N ratios and % C4 roots (as a percentage of total roots; both grass and trees, mean \pm SE) for different levels of woody encroachment in the Hluhluwe section of the Hluhluwe-iMfolozi Park, South Africa; 10-y-old thicket, 40-y-old thicket, and 70-y-old thicket, at different depths (0–10 cm, 10–20 cm, 20–30 cm, as well as averaged over depth). Root C : N ratios (a), C4 grass (%); (b). Different lowercase letters indicate significant differences at $P < 0.05$.

on soil C stocks and land-use change. Guo & Gifford (2002) showed that, across 16 countries, soil C stocks increased when native forest was converted to pasture. A more recent meta-analysis found the opposite, that in the tropics and subtropics, forest conversion to grassland leads to an average loss in soil C stocks of 12% (Don *et al.* 2011). At the same time, Eldridge *et al.* (2011) have shown in a global synthesis on shrub encroachment that total soil C increased with woody encroachment in the top 15 cm of soil. This is in contrast to our results which suggest that in the top 15 cm of soil the differences in soil C between forests and grasslands, and thickets and savannas, are not great. Total soil C storage in our study increased with woody encroachment into areas previously dominated by grass, but this was due almost entirely to increases in AGC. Besides the top 15 cm of soil, forests contained consistently more C than

grasslands, and differences in soil C were significant. In contrast, although on average thickets contained slightly more C than adjacent savannas in all components, these differences were only significant in the AGC.

The second aim of the study was to examine the relationship between C concentrations and measured variables. Previous work has shown that although soil C increases with woody encroachment, decreases may occur at wetter sites and on specific soils when vegetation reaches the closed-canopy stage (Hudak *et al.* 2003, Jackson *et al.* 2002). Hudak *et al.* (2003) suggested the loss of grass roots as a possible mechanism of declining soil C concentrations at sites where a closed canopy prohibited the growth of grass. Our results, however, showed that soil C was not affected by the per cent C4 roots across sites and high C concentrations were found with both medium and low per cent C4 roots.

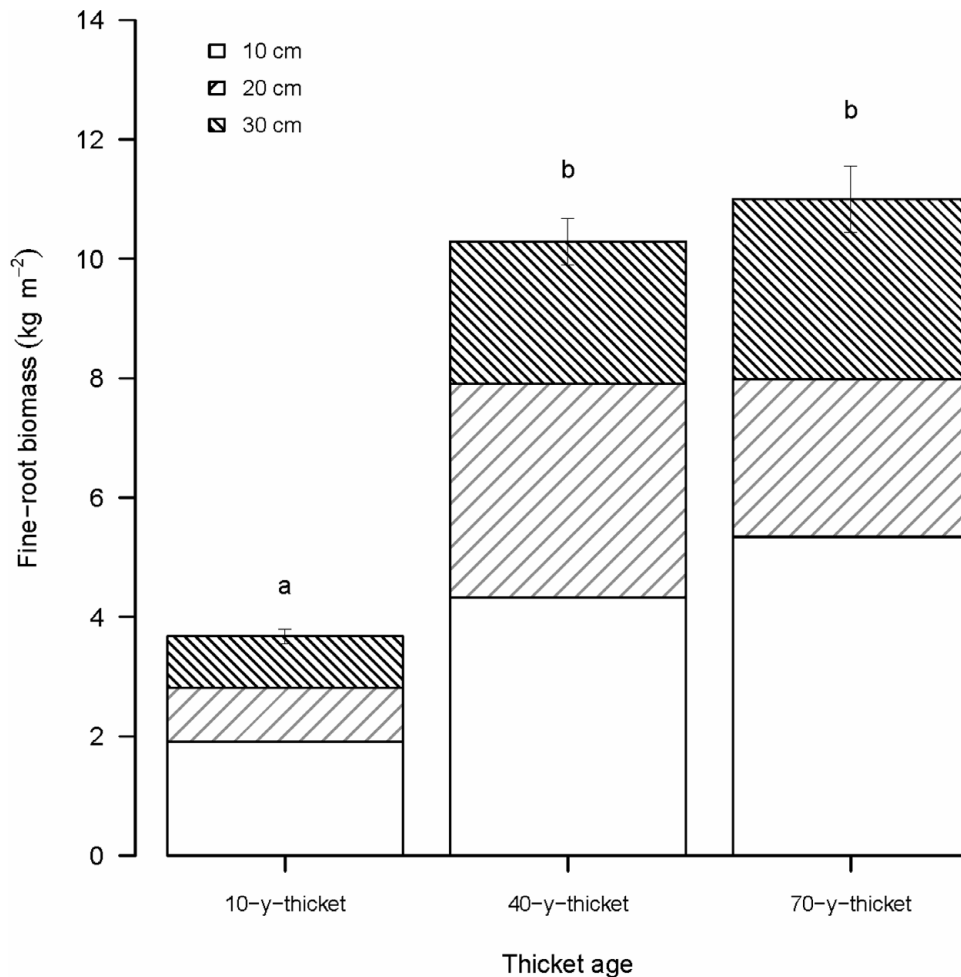


Figure 5. Fine-root biomass (mean \pm SE, g m^{-2} , $n = 189$) for different levels of woody encroachment in the Hluhluwe section of the Hluhluwe-iMfolozi Park, South Africa; 10-y-old thicket, 40-y-old thicket and 70-y-old thicket, at different depths (0–10 cm, 10–20 cm, 20–30 cm). Different lowercase letters indicate significant differences at $P < 0.05$.

Which factors were then affected by changes in woodiness and how did these relate to soil C concentrations? Woody plant encroachment of grassland often increases the sequestration and cycling of ecosystem C (Geesing *et al.* 2000, Hibbard *et al.* 2001, McCulley *et al.* 2004). Cebrián & Duarte (1995) argue that rates of plant turnover affect decomposition and are important in governing soil C storage. Ecosystems dominated by slow-growing plants accumulate large, slowly decomposing detrital pools which act as C sinks. Previous work has shown that woody AGC inputs and low decomposability in forests may increase C storage (Austin & Vitousek 1998, Melillo *et al.* 1982). Similarly, Bird & Pousai (1997) and Kellman (1979) have shown that higher soil C concentrations in tree-dominated areas may be the result of higher C inputs per area and the longer residence time of woody material derived from trees. We have used root C:N ratios as an indicator of root quality as C:N ratios often govern the rate of litter decomposition

and higher C:N ratios often (but not always) mean slower decomposition of litter and as a result higher soil organic C (Enríquez *et al.* 1993, Parton *et al.* 2007). The 70-y-old thicket had the highest root quality (i.e. lowest root C:N ratios) and was also the site with the highest soil C concentrations. This was an unexpected result, as we expected this site with low C:N ratios to have faster decomposition rates and as a result lower C concentrations. Our extensive soil sampling also showed that soil C:N ratios in thickets were generally lower than either grassland or forest (Table 3) and no apparent correlation between soil C:N ratios and C storage.

Apart from altered decomposition rates of above-ground or below-ground inputs, increases of SOC concentrations and storage may be a result of enhanced production below ground (Schlesinger 1977). Our results showed that fine-root biomass was an important predictor of total soil C concentrations. Increases in fine-root biomass were especially noticeable in the deeper (20–30

cm) soil layers when newly invaded grassland turned to established thicket. Jobbágy & Jackson (2000), in a meta-analysis of global root distribution, showed that differing patterns of allocation by vegetation determined vertical distributions of soil C. Other studies have found increased C and N storage with woody encroachment and several mechanisms come into place with higher densities of trees; these include the tree acting as an atmospheric dust trap (Bernhard-Reversat 1988, Escudero *et al.* 1985), and trees and shrubs affecting nutrient cycling by changing soil structure, microbial biomass, soil moisture, microclimate and by changing N fixation (Hibbard *et al.* 2001, Hudak *et al.* 2003, Jacobs *et al.* 2007, Schlesinger *et al.* 1990). As we did not measure root turnover, it is difficult to conclude whether root biomass alone or together with changes in root productivity affects soil C concentration. Hibbard *et al.* (2001) found that higher root biomass together with faster root turnover under woody canopies compared with grassy patches in a North American savanna accounted for higher soil C storage in invaded areas. Our results support this and other work that shows that root biomass is important in governing soil C storage (Jackson *et al.* 1996, 1997; Jobbágy & Jackson 2000).

Our findings showed that it takes a very long period of time before C stocks increase over the entire soil profile in thicket-invaded areas. The magnitude of gains in C stored above ground was larger than that gained below ground; e.g. 4.94 kg m⁻² and 0.46 kg m⁻² contained in thicket and grassland respectively above ground, while below ground thicket and grassland contained similar amounts of C (9.26 kg m⁻² and 8.30 kg m⁻² in the top 15 cm respectively). We suggest that one possible mechanism that explains the low gains in below-ground C is linked with the fact that fine-root biomass appears to be an important driver of soil C in this savanna. Initially, fine-root biomass increases rapidly with woody encroachment, but as there is little difference between fine-root biomass in the 40-y-old and 70-y-old thicket, so it appears to reach some asymptote. In other words, instead of below-ground C increasing linearly over time since woody encroachment, the increases in soil C seems to slow down within 40 y of the initial woody invasion. However, soil C does increase in mature scarp forest. We suggest that a different mechanism is important in driving soil C in these forests. Fine-root biomass may not be an important predictor of soil C, but high litter inputs of low quality and slow turnover may increase soil C storage.

In conclusion, fire abatement is often advocated in the literature as a way in which to increase C storage in savannas as fire removal leads to increases in C storage (Grace *et al.* 2006). According to the study by Wigley *et al.* (2010), 44% of the study area in HiP that was dominated by grassy systems 70 y ago is now dominated by woody vegetation. Thickets support an entirely different and

less divergent set of flora and fauna, and nearly half of the plant diversity would be lost with the conversion of savanna to thicket in HiP (Bond & Parr 2010, Parr *et al.* 2012). Furthermore, woody encroachment alters ecosystem functioning by influencing nutrient cycling and hydrology (Asner *et al.* 2004, Bond 2008). Woody plant encroachment may also negatively impact game-viewing experiences in the park, thereby reducing overall visitor numbers in the park (E. Gray, unpubl. data). It is also likely to decrease livestock productivity outside park boundaries (Burrows *et al.* 1990, Mugasi *et al.* 2000). Gains in C should therefore be weighed up against changes in ecosystem function, losses in biodiversity and economic implications before management decisions regarding fire are made in savannas.

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LITERATURE CITED

- AERTS, R., BAKKER, C. & DECALUWE, H. 1992. Root turnover as determinant of the cycling of C, N and P in a dry heathland ecosystem. *Biogeochemistry* 15:175–190.
- ARCHER, S., BOUTTON, T. W. & HIBBARD, K. A. 2000. Trees in grasslands: Biogeochemical consequences of woody plant expansion. Pp. 115–137 in Schulze, E. D., Harrison, S. P., Heimann, M., Holland, E. A., Lloyd, J., Prentice, I. C. & Schimel, D. (eds.), *Global biogeochemical cycles in the climate system*. Academic Press, San Diego.
- ASNER, G. P., ELMORE, A. J., OLANDER, L. P., MARTIN, R. E. & HARRIS, A. T. 2004. Grazing systems, ecosystem responses, and global change. *Annual Review of Environment and Resources* 29:261–299.
- Austin, A. T. & Vitousek, P. M. 1998. Nutrient dynamics on a precipitation gradient in Hawaii. *Oecologia* 113:519–529.
- BAKER, T. R., PHILLIPS, O. L., MALHI, Y., ALMEIDA, S., ARROYO, L., IORE, A. D. I., ERWIN, T., KILLEEN, T. I. J., LAURANCE, S. G., LAURANCE, W. F., LEWIS, S., LLOYD, J., MONTEAGUDO, A., NEILL, D. A., PATINO, S., PITMAN, N. C. A., SILVA, J. N. M. & MARTINEZ, R. V. 2004. Variation in wood density determines spatial patterns in Amazonian forest biomass. *Global Change Biology* 10:545–562.
- BALFOUR, D. A. & HOWISON, O. E. 2001. Spatial and temporal variation in a mesic savanna fire regime: response to variation in annual rainfall. *African Journal of Range and Forage Science* 19:45–53.
- BALFOUR, D. A. & MIDGLEY, J. J. 2008. A demographic perspective on bush encroachment by *Acacia karroo* in Hluhluwe-Imfolozi Park, South Africa. *African Journal of Range and Forage Science* 25:147–151.

- BEERLING, D. J. & OSBORNE, C. P. 2006. The origin of the savanna biome. *Global Change Biology* 12:2023–2031.
- BERNHARD-REVERSAT, F. 1988. Soil nitrogen mineralization under a *Eucalyptus* plantation and a natural *Acacia* forest in Senegal. *Forest Ecology and Management* 23:233–244.
- BIRD, M. I. & POUSAI, P. 1997. Variation of $\delta^{13}\text{C}$ in the surface soil organic pool. *Global Biogeochemical Cycles* 11:313–322.
- BOND, W. J. 2008. What limits trees in C_4 grassland and savannas? *Annual Review of Ecology, Evolution and Systematics* 39:641–659.
- BOND, W. J. & MIDGLEY, G. F. 2012. CO_2 and the uneasy interactions of trees and savanna grasses. *Philosophical Transactions of the Royal Society B* 362:601–612.
- BOND, W. J. & PARR, C. L. 2010. Beyond the forest edge: ecology, diversity and conservation of the grassy biomes. *Biological Conservation* 143:2395–2404.
- BOND, W. J., MIDGLEY, G. F. & WOODWARD, F. I. 2003. The importance of low atmospheric CO_2 and fire in promoting the spread of grasslands and savannas. *Global Change Biology* 9:973–982.
- BOUTTON, T. W., ARCHER, S. R., MIDWOOD, A. J., ZITZER, S. F. & BOL, R. 1998. $\delta^{13}\text{C}$ values of soil organic carbon and their use in documenting vegetation change in a subtropical savanna ecosystem. *Geoderma* 82:5–41.
- BRANSBY, D. I. & TAINTON, N. M. 1977. The disk pasture meter: possible applications in grazing management. *Proceedings of the Annual Congresses of the Grassland Society of Southern Africa* 12:115–118.
- BURROWS, W. H., CARTER, J. O., SCANLAN, J. C. & ANDERSON, E. R. 1990. Savanna ecology and management: Australian perspectives and intercontinental comparisons. *Journal of Biogeography* 17:503–512.
- CEBRIÁN, J. & DUARTE, C. M. 1995. Plant growth-rate dependence of detrital carbon storage in ecosystems. *Science* 268:1606–1608.
- CHAMBERS, J. Q., DOS SANTOS, J., RIBEIRO, R. J. & HIGUCHI, N. 2001. Tree damage, allometric relationships, and above-ground net primary production in central Amazon forest. *Forest Ecology and Management* 152:73–84.
- DON, A., SCHUMACHER, J. & FREIBAUER, A. 2011. Impact of tropical land use change on soil organic stocks – a meta-analysis. *Global Change Biology* 17:1658–1670.
- ECKHARDT, H. C., VAN WILGEN, B. W. & BIGGS, H. C. 2000. Trends in woody vegetation cover in the Kruger National Park, South Africa, between 1940 and 1998. *African Journal of Ecology* 38:108–115.
- ELDRIDGE, D. J., BOWKER, M. A., MEASTRE, F. T., ROGER, E., REYNOLDS, J. F. & WHITFORD, W. G. 2011. Impacts of shrub encroachment on ecosystem structure and functioning: towards a global synthesis. *Ecology Letters* 14:709–722.
- ENRÍQUEZ, S., DUARTE, C. M. & SAND-JENSEN, K. 1993. Patterns in decomposition rates among photosynthetic organisms: the importance of detritus C:N:P content. *Oecologia* 94:457–471.
- ESCUARO, A., GARCIA, B., GOMEZ, J. M. & LUIS, E. 1985. The nutrient cycling in *Quercus rotundifolia* and *Q. pyrenaica* ecosystems ('dehesas') of Spain. *Oecologia* 6:73–86.
- FEY, M. 2010. *Soils of South Africa. Their distribution, properties, classification, genesis, use and environmental significance*. Cambridge University Press, New York. 287 pp.
- GEESING, D., FELKER, P. & BINGHAM, R. L. 2000. Influence of mesquite (*Prosopis glandulosa*) on soil nitrogen and carbon development: implications for global carbon sequestration. *Journal of Arid Environments* 46:157–180.
- GRACE, J., SAN JOSÉ, J., MEIR, P., MIRANDA, H. S. & MONTES, R. A. 2006. Productivity and carbon fluxes of tropical savanna. *Journal of Biogeography* 33:387–400.
- GUO, L. B. & GIFFORD, R. M. 2002. Soil carbon stocks and land use change: a meta analysis. *Global Change Biology* 8:345–360.
- HIBBARD, K. A., ARCHER, S., SCHIMEL, D. S. & VALENTINE, D. W. 2001. Biochemical changes accompanying woody plant encroachment in a subtropical savanna. *Ecology* 82:1999–2011.
- HIGGINS, S. I., SHACKLETON, C. M. & ROBINSON, R. 1999. Changes in woody community structure and composition under contrasting landuse systems in a semi-arid savanna, South Africa. *Journal of Biogeography* 26:619–627.
- HIGGINS, S. I., BOND, W. J., FEBRUARY, E. C., BRONN, A., EUSTON-BROWN, D. I. W., ENSLIN, B., GOVENDER, N., RADEMAN, L., O'REGAN, S., POTGIETER, A. L. F., SCHEITER, S., SOWRY, R., TROLLOPE, L. & TROLLOPE, W. S. W. 2007. Effects of four decades of fire manipulation on woody vegetation structure in savanna. *Ecology* 88:1119–1125.
- HOFFMANN, W. A., BAZZAZ, F. A., CHATTERTON, N. J., HARRISON, P. A. & JACKSON, R. B. 2000. Elevated CO_2 enhances resprouting of a tropical savanna tree. *Oecologia* 123:312–317.
- HUDAK, A. T., WESSMAN, C. A. & SEASTEDT, T. R. 2003. Woody overstorey effects on soil carbon and nitrogen pools in a South African savanna. *Austral Ecology* 28:173–181.
- JACKSON, R. B., CANADELL, J., EHLERINGER, J. R., MOONEY, H. A., SALA, O. E. & SCHULZE, E. D. 1996. A global analysis of root distributions for terrestrial biomes. *Oecologia* 108:389–411.
- JACKSON, R. B., MOONEY, H. A. & SCHULZE, E.-D. 1997. A global budget for fine-root biomass, surface area and nutrient contents. *Proceedings of the National Academy of Science, USA* 94:7362–7366.
- JACKSON, R. B., BANNER, J. L., JOBBÁGY, E., POCKMAN, W. T. & WALL, D. H. 2002. Ecosystem carbon loss with woody plant invasion of grasslands. *Nature* 418:623–626.
- JACOBS, S. M., BECHTOLD, J. S., BIGGS, H. C., GRIMM, N. B., LORENTZ, S., MCCLAIN, M. E., NAIMAN, R. J., PERAKIS, S. S., PINAY, G. & SCHOLLES, M. C. 2007. Nutrient vectors and riparian processing: a review with special reference to African semiarid savanna ecosystems. *Ecosystems* 10:1432–1440.
- JOBBÁGY, E. G. & JACKSON, R. B. 2000. The distributions of soil nutrients with depth: global patterns and the imprints of plants. *Biogeochemistry* 53:51–77.
- KELLMAN, M. 1979. Soil enrichment by neotropical savanna trees. *Journal of Ecology* 67:565–577.
- LOW, A. B. & REBELO, A. G. 1996. *Vegetation of South Africa, Lesotho and Swaziland*. DEAT, Pretoria. 85 pp.
- MACLAREN, M. P. & MCPHERSON, G. R. 1995. Can soil organic carbon isotopes be used to describe grass–tree dynamics at a savanna–grassland ecotone and within the savanna? *Journal of Vegetation Science* 6:857–862.
- MACVICAR, C. N., DE VILLIERS, J. M., LOXTON, R. F., VERSTER, E., LAMBRECHTS, J. J. N., MERRYWEATHER, F. R., LE ROUX, J., VAN

- ROOYEN, T. H. & VON HARMSE, H. J. 1977. *Soil classification: a binomial system for South Africa*. The Soil and Irrigation Research Institute, Department of Agricultural Technical Services, Pretoria.
- Martin, A. R. & Thomas, S. C. 2011. A reassessment of carbon content in tropical trees. *PloS One* 6:e23533. doi:10.1371/journal.pone.0023533.
- Mcculley, R. L., ARCHER, S. R., BOUTTON, T. W., HONS, F. M. & ZUBERER, D. A. 2004. Soil respiration and nutrient cycling in wooded communities developing in grassland. *Ecology* 85:2804–2817.
- MELILLO, J. M., ABER, J. D. & MURATORE, J. F. 1982. Nitrogen and lignin control of hardwood leaf litter decomposition dynamics. *Ecology* 63:621–626.
- MIDGLEY, J. J. & SEYDACK, A. 2006. What determines biomass in indigenous forests? An analysis of the Knysna forest, South Africa. *Australian Journal of Botany* 54:701–705.
- MORDELET, P., MENAUT, J.-C. & MARIOTTI, A. 1997. Tree and grass rooting patterns in an African humid savanna. *Journal of Vegetation Science* 8:65–70.
- MUCINA, L. & RUTHERFORD, M. C. 2006. *The vegetation of South Africa, Lesotho and Swaziland*. Strelitzia no. 19. South African National Biodiversity Institute, Pretoria.
- MUGASI, S. K., SABIITI, E. N. & TAYEBWA, B. M. 2000. The economic implications of bush encroachment on livestock farming in rangelands of Uganda. *African Journal of Rangeland and Forage Science* 17:64–69.
- PARR, C. L., GRAY, E. F. & BOND, W. J. 2012. Cascading biodiversity and functional consequences of a global change-induced biome switch. *Diversity and Distributions* 18:493–503.
- PARTON, W., SILVER, W. L., BURKE, I. C., GRASSENS, L., HARMON, M. E., CURRIE, W. S., KING, J. Y., ADAIR, E. C., BRANDT, L. A., HART, S. C. & FASTH, B. 2007. Global-scale similarities in nitrogen release patterns during long-term decomposition. *Science* 315:361–364.
- PENMAN, J., GYTARSKY, M., HIRAISHI, T., KRUG, T., KRUGER, D., PIPATTI, R., BUENDIA, L., MIWA, K., NGARA, T., TANABE, K. & WAGNER, F. 2003. *Intergovernmental Panel on Climate Change Good Practice Guidance for Land Use, Land-Use Change and Forestry*. Institute for Global Environmental Strategies, Kanawaga.
- PENNINGTON, R. T., LAVIN, M. & OLIVEIRA-FILHO, A. 2009. Woody plant diversity, evolution and ecology in the tropics: perspectives from seasonally dry tropical forests. *Annual Review of Ecology and Systematics* 40:437–457.
- RATNAM, J., BOND, W. J., FENSHAM, R. J., HOFFMAN, W. A., ARCHIBALD, S., LEHMANN, C. E. R., ANDERSON, M. T., HIGGINS, S. I. & SANKARAN, M. 2011. When is a 'forest' a savanna, and why does it matter? *Global Ecology and Biogeography* 20:653–660.
- ŠÁLEK, L. & ZÁHRADNÍK, D. 2008. Wedge prism as a tool for diameter and distance measurement. *Journal of Forest Science* 54:121–124.
- SCHLESINGER, W. H. 1977. Carbon balance in terrestrial detritus. *Annual Review of Ecology and Systematics* 8:51–81.
- SCHLESINGER, W. H., REYNOLDS, J. F., CUNNINGHAM, G. L., HUNNENNEKE, L. F., JARRELL, W. M., VIRGINIA, R. A. & WHITFORD, W. G. 1990. Biological feedbacks in global desertification. *Science* 247:1043–1048.
- SKOWNO, A. L., MIDGLEY, J. J., BOND, W. J. & BALFOUR, D. 1999. Secondary succession in *Acacia nilotica* (L.) savanna in the Hluhluwe game reserve, South Africa. *Plant Ecology* 145:1–9.
- STILL, C. J., BERRY, J. A., RIBAS-CARBO, M. & HELLIKER, B. R. 2003. The contribution of C3 and C4 plants to the carbon cycle of a tallgrass prairie: an isotopic approach. *Oecologia* 136:347–359.
- WALDRAM, M. S., BOND, W. J. & STOCK, W. D. 2008. Ecological engineering by a mega-grazer: white rhino impacts on a South African savanna. *Ecosystems* 11:101–112.
- WALKLEY, A. 1947. A critical examination of a rapid method for determining organic carbon in soils: effect of variations in digestion conditions and of inorganic soil constituents. *Soil Science* 63:251–263.
- WATSON, H. K. 1995. Management implications of vegetation changes in Hluhluwe-Umfolozi Park. *South African Geographical Journal* 77:77–83.
- WEST, A. G., BOND, W. J. & MIDGLEY, J. J. 2000. Soil carbon isotopes reveal ancient grasslands under forest in Hluhluwe, KwaZulu-Natal. *South African Journal of Science* 96:252–254.
- WHATELEY, A. & PORTER, R. N. 1983. The woody vegetation communities of the Hluhluwe-Corridor-iMfolozi Game Reserve Complex. *Bothalia* 14:754–758.
- WIGLEY, B. J., BOND, W. J. & HOFFMAN, M. T. 2010. Thicket expansion in a South African savanna under divergent land use: local vs. global drivers. *Global Change Biology* 16:964–976.