

Maternal pregnancy C-reactive protein predicts offspring birth size and body composition in metropolitan Cebu, Philippines

C. W. Kuzawa^{1,2*}, R. L. Fried¹, J. B. Borja³ and T. W. McDade^{1,2}

¹*Department of Anthropology, Northwestern University, Evanston, IL, USA*

²*Institute for Policy Research, Northwestern University, Evanston, IL, USA*

³*Office of Population Studies Foundation, University of San Carlos, Cebu City, Philippines*

The gestational milieu is an important influence on fetal development and long-term disease risk. Here we assess relationships between maternal pregnancy inflammation, indicated by C-reactive protein (CRP), and offspring anthropometric outcomes measured soon after birth. Data come from female participants ($n = 327$, age 24.4–30.2 years) in a longitudinal study located in Metropolitan Cebu, Philippines. Between 2009 and 2014, pregnancy interviews ($n = 429$) were conducted during which questionnaire and anthropometric data were obtained along with dried blood spot cards for CRP measurement. Offspring body weight, length, head circumference and five skinfold thickness measures were obtained soon after birth. Maternal pregnancy CRP was borderline (-1.11 ± 0.64 days/log-mg/l; $P < 0.1$) inversely related to gestational age at delivery, but did not increase the likelihood of preterm delivery. After adjusting for maternal pre-pregnancy body mass index, height, pregnancy adiposity, age, parity and other covariates, CRP was significantly, inversely related to offspring body weight (-0.047 ± 0.017 kg/log-mg/l), length (-0.259 ± 0.092 cm/log-mg/l) and sum of skinfolds (-0.520 ± 0.190 mm/log-mg/l) (all $P < 0.05$), and borderline inversely related to offspring head circumference (-0.102 ± 0.068 cm/log-mg/l; $P < 0.1$). Notably, relationships were continuous across the full CRP range, and not limited to unusually high levels of inflammation. These findings point to an important role of maternal non-specific immune activation as a predictor of offspring birth outcomes. In light of evidence that early life microbial, nutritional and stress experiences influence adult inflammatory regulation, these findings point to inflammation as a potential pathway for the intergenerational transmission of maternal experience to offspring health.

Received 2 December 2016; Revised 29 May 2017; Accepted 6 June 2017; First published online 19 July 2017

Key words: body fat, growth, inflammation, intergenerational, newborn

Introduction

Evidence that birth outcomes predict a range of later outcomes related to health and human capital has heightened interest in the maternal factors that shape the gestational environment.^{1,2} Although much of this work has focused on the importance of nutrient supply to the fetus, more recent work has highlighted the influence of the maternal immune system on fetal growth and development.^{3,4} Acute infections, including malaria and sexually transmitted diseases, have long-been recognized as important risk factors for giving birth to a smaller baby, operating through both fetal growth restriction and shortened gestational duration.^{5–7} Research has also shown that endogenous immune processes, notably chronic inflammation, can have a negative influence on birth outcomes.^{3,4,8,9} Normal pregnancy is associated with a modest and sustained increase in inflammation, reflected in elevated C-reactive protein (CRP), and by marked changes in immune priorities that suppress maternal rejection of the fetus by decreasing the ratio of T helper 1 to T helper 2 cells.^{10–14} However, abnormally elevated CRP levels during pregnancy are associated with preeclampsia, preterm birth and impaired fetal growth.^{9,15}

Despite the predominant focus on clinical outcomes in prior research, a small but growing number of studies demonstrate an association between maternal CRP levels and offspring outcomes across the full range of maternal inflammatory status.^{4,16,17} Among mothers who experience clinically normal pregnancy, CRP has been found to be negatively associated with birth weight for gestational age, explaining up to 25.9% of the variance in birth weight.^{4,18} Women with increased pregnancy CRP have also been shown to give birth to smaller and shorter babies with reduced adiposity and moderately decreased head circumference.¹⁷

Although the pathways linking maternal inflammation with suboptimal fetal growth are not fully understood, elevated maternal CRP can indicate cellular damage and endothelial dysfunction characterized by dysregulated vasoconstriction and vasodilation.^{19,20} The resulting poor vascular function may contribute to restricted fetal growth by decreasing nutrient flow across the maternal–fetal interface.²¹ In addition, there are feedbacks between maternal and placental immune regulation that can lead to dysregulated inflammation.²² For example, poor endometrial decidualization and/or placental perfusion can lead to placental hypoxia, ischemia and tissue death.^{20,23,24} These conditions may lead to release of inflammatory stimuli by the placenta into maternal circulation which in turn stimulates placental production of proinflammatory cytokines, thus further inhibiting maternal–fetal nutrient delivery.^{21,22,25}

*Address for correspondence: C. W. Kuzawa, Department of Anthropology, Northwestern University, 1810 Hinman, Evanston, IL 60208, USA.
 (Email kuzawa@northwestern.edu)

Through these pathways, elevated maternal CRP can serve as a marker of restricted maternal–fetal nutrient exchange. Furthermore, there is evidence that inflammatory regulation in adulthood is related to prenatal and early postnatal conditions, such that being born small predicts elevated CRP in adulthood.^{26–30} To the extent that both relationships are causal, they suggest that inflammation may serve as an intergenerational pathway linking adverse health across generations.^{28,31}

In this study, we aim to clarify the role of maternal inflammation as an influence on offspring birth outcomes. To that end, we report maternal CRP measured during pregnancy, which we relate to offspring weight, length, head circumference and adiposity measured soon after birth. Data come from a well-characterized cohort in the Philippines, in which the original cohort are now of reproductive age and having offspring.

Methods

Study population

Data come from the Cebu Longitudinal Health and Nutrition Survey, a longitudinal survey of 3080 singletons whose mothers were recruited during pregnancy between 1983 and 1984 in Metropolitan Cebu, Philippines.³² Of the 1447 original female cohort participants, 823 were interviewed in the 2009 survey (at ages 25–26). A special survey tracked new pregnancies among these women between 2009 and 2014. There were 383 who reported pregnancies (28% with 2–3 pregnancies) within the tracking period, yielding 507 pregnancy episodes. Women were visited in-home during pregnancy for anthropometric and questionnaire assessments, along with collection of a dried blood spot (DBS) – capillary whole blood collected on filter paper – for CRP measurement. A second visit was arranged right after delivery to obtain additional information from the mothers along with anthropometric measures of their newborns. This research was conducted under conditions of written informed consent, and with approval of the Institutional Review Boards of Northwestern University (Evanston, IL, USA), and the Office of Population Studies Foundation (Cebu, Philippines).

Sample inclusion criteria

Newborn anthropometric outcomes in these analyses included weight, length, head circumference and sum of five skinfold thicknesses (triceps, subscapular, suprailiac, bicep and calf), which were measured in-home by trained interviewers using standardized procedures.³³ Efforts were made to obtain these measurements as soon after birth as possible. The median and mean interval (in day) between birth and newborn anthropometry measurements were 3 and 4.5 days, respectively, with a range from 1 to 44 days. To minimize any impacts of the postnatal environment and postnatal growth on infant anthropometry, we limited analyses to infants who were measured within 2 weeks of birth, and adjusted for age at measurement in models (this excluded 17 individuals measured

more than 2 weeks after birth). We further limited analyses to newborns born with gestational ages between 32 and 44 weeks, which excluded five very premature births and two implausibly late deliveries of around 46 weeks. Finally, we predicted newborn weight as a function of gestational age at birth and postnatal age at anthropometry measurement and excluded three individuals whose residuals were ≥ 3 s.d. away from their predicted weights (e.g. individuals who were implausibly light for their gestational age at delivery and/or postnatal age at anthropometry measurement). After these exclusions, the final sample with all necessary biological and questionnaire data included 429 relatively healthy singletons born to 328 women. Regression models were clustered on mother to account for non-independence among siblings (see below).

Maternal covariates

We adjusted for mother's age, parity and triceps skinfold thickness, at the time that the pregnancy interview and DBS collection were completed, and adult stature that had been collected during previous assessments. Because both CRP and birth outcomes are potentially impacted by the mother's adiposity, we also adjusted for the mother's pre-pregnancy body mass index (BMI). Maternal socio-economic status was measured using a pregnancy household assets scale reflecting whether the family had electricity, owned their home, owned an air conditioner, refrigerator, TV, vehicle and other appliances assessed, and a measure of household income that tallies all sources of income within the household (Adair et al. 2011). Because women were enrolled in the birth outcome sub-study after they were pregnant, we used height and weight measurements collected during prior surveys to estimate pre-pregnancy BMI. We used 2009 BMI when available, and then used 2007 and 2005 data as necessary. Under the assumption that women will tend to maintain a stable position within the population BMI distribution even as the population mean increases with age, we converted all BMIs to age-specific within-sample Z-scores before pooling into a single pre-pregnancy BMI variable. Supporting the validity of this approach, the correlation between Z-scores for BMI measured in 2005 and 2009 was very high ($r=0.84$).

CRP measurement

Samples were analysed for CRP in the Laboratory for Human Biology Research at Northwestern University using a modified high sensitivity enzyme immunoassay protocol previously developed for use with DBS.³⁴ Prior validation of assay performance indicates that the DBS CRP method produces results that are comparable to gold standard plasma-based clinical methods, with a lower limit of detection of 0.03 mg/l.³⁴ To minimize between-assay variation, all samples were analysed by the same technician using a single lot of capture antibody, detection antibody and calibration material. Between-assay CVs for low, mid and high control samples included with all runs were 11.8, 9.6, and 8.1%, respectively.

Before data analysis, DBS results were converted to plasma equivalent values using a conversion formula based on 69 matched DBS and plasma samples.³⁵ DBS samples were analysed using the same procedures, lot number of reagents and technician as applied to the study DBS samples. Plasma samples were analysed for high sensitivity CRP in a high throughput clinical laboratory on the Beckman Coulter Synchron DXC platform. The correlation between DBS and plasma values was high (Pearson's $R=0.98$) and the resulting Deming regression conversion formula was as follows: plasma (mg/l) = $1.64 \times$ DBS (mg/l).

Statistical analysis

All statistical analyses were conducted using Stata 13.0 (College Station, TX). We reported unadjusted means and standard deviations (or % for count variables) for the full sample and stratified on a median split of pregnancy CRP. We then ran a sequence of multivariate regression models (either linear or logistic) designed to assess relationships between maternal CRP and offspring gestational timing and weight, length, head circumference and sum of skinfolds measured soon after birth. All models were run with the *cluster* option in Stata, with clustering on mother's ID, to account for the non-independence of siblings among women with multiple births in the analysis sample. CRP was right skewed and was therefore log-transformed before analysis. Because body fat can have

both positive effects on birth outcomes (via nutrient supply) and negative effects on birth outcomes (via effects on proinflammatory cytokines) we assessed relationships between CRP and offspring outcomes before and after adjusting for maternal adiposity measures. Models predicting postnatal outcomes were adjusted for days after birth of anthropometry measurement, offspring gender, maternal parity and age during pregnancy visit and the mother's adult height (coefficients not shown). Neither household assets nor income were close to significantly related to maternal CRP and adjusting for them did not substantially change model coefficients; we thus report each SES variable in the descriptive statistics for reference but omit them from models. To clarify the extent to which CRP influences offspring outcomes via effects on growth rate *v.* gestational duration, we evaluated models before and after adjustment for gestational age at delivery. Finally, to evaluate a possible role of preeclampsia, we assessed whether coefficients relating CRP to offspring outcomes were attenuated after further adjustment for the mother's systolic blood pressure measured during the pregnancy interview.

Results

The women in this sample were an average of 27 years old during their pregnancy interviews (Table 1). When compared to women with below-median pregnancy CRP, women with

Table 1. Characteristics of mothers during pregnancy and offspring after birth stratified on a median split of maternal pregnancy C-reactive protein (CRP)^a

	All (<i>n</i> = 429)	CRP < 1.14 mg/l (<i>n</i> = 215)	CRP ≥ 1.14 mg/l (<i>n</i> = 214)	<i>P</i> -value ^b
Mother's pregnancy traits				
Height (cm) ^c	151.2 ± 5.3	151.2 ± 5.5	151.3 ± 5.2	0.895
Mother's age (years)	27.0 ± 1.5	27.1 ± 1.5	26.8 ± 1.5	0.128
Household income (pesos)	518 ± 660	536 ± 810	499 ± 464	0.633
Assets scale (0–16)	4.2 ± 3.6	4.2 ± 3.4	4.1 ± 3.7	0.803
Parity	2.9 ± 1.6	2.7 ± 1.6	3.0 ± 1.6	0.036
Triceps SF (mm)	22.2 ± 5.6	20.8 ± 5.1	23.6 ± 5.6	0.000
CRP (mg/l)	2.0 ± 3.6	0.65 ± 0.29	3.3 ± 4.7	0.000
Systolic blood pressure (mmHg)	101.0 ± 10.4	100.2 ± 9.7	101.7 ± 11.1	0.167
Gestational day of measure	205.8 ± 26.5	206.0 ± 24.9	205.7 ± 28.2	0.921
Offspring neonatal traits				
Male offspring [% (<i>n</i>)]	51.1% (219)	49.8% (107)	52.3% (112)	0.604
Gestational age at birth (weeks)	39.3 ± 1.8	39.4 ± 1.6	39.2 ± 2.0	0.254
Weight (kg)	3.06 ± 0.39	3.06 ± 0.39	3.05 ± 0.39	0.832
Length (cm)	48.6 ± 2.0	48.6 ± 2.0	48.5 ± 2.0	0.825
Head circumference (cm)	33.0 ± 1.3	33.0 ± 1.4	33.0 ± 1.3	0.861
Sum of skinfolds (mm)	22.4 ± 4.4	22.7 ± 4.4	22.2 ± 4.3	0.203
Age at measurement (days)	3.8 ± 2.5	3.7 ± 2.3	3.9 ± 2.7	0.315

^aMean ± S.D.

^bFrom bivariate linear or logistic regression clustered on mother.

^cPre-pregnancy height (*n* = 327 women).

SF, skinfold.

Table 2. Maternal pregnancy C-reactive protein (CRP) and adiposity as predictors of gestational age at delivery and preterm delivery

	CRP	Maternal fat	CRP + fat
Gestational age (days) ^a			
Log-CRP	-0.96 ± 0.61		-1.11 ± 0.64-
BMI		0.59 ± 0.86	0.78 ± 0.85
Triceps SF		-0.35 ± 0.73	-0.18 ± 0.73
Preterm delivery ^b			
Log-CRP	1.29 (0.93, 1.79)		1.28 (0.95, 1.74)
BMI		1.37 (0.97, 1.95)-	1.32 (0.94, 1.85)
Triceps SF		0.76 (0.54, 1.06)-	0.73 (0.53, 1.02)-

BMI, body mass index; SF, skinfold.

^aFrom multiple regression models, coefficients $\beta \pm$ s.e.

^bFrom logistic regression models, odds ratio (95% confidence interval).

- $P < 0.1$.

above-median CRP had on average higher parity and thicker triceps skinfold thickness. There were no significant bivariate differences in any offspring trait between high/low maternal pregnancy CRP women.

We next ran models linking maternal pregnancy CRP to offspring birth outcomes. In linear regression models predicting gestational age as a continuous variable, higher CRP during pregnancy was marginally associated with earlier delivery ($P < 0.081$), but only after adjusting for adiposity measures (Table 2). In logistic models adjusting for maternal adiposity measures, CRP was not significantly related to preterm delivery (< 37 weeks; $P < 0.109$).

We next considered CRP as a predictor of offspring weight, length, head circumference and sum of skinfolds measured soon after birth (Table 3). In each instance, negative relationships between CRP and each outcome were strengthened after adjusting for measures of maternal body fat. Relationships between CRP and each outcome were weakened slightly after further adjustment for gestational age. In fully adjusted models (Fig. 1), CRP had statistically significant inverse relationships with sum of skinfolds ($P < 0.006$) and birth length and weight (both $P < 0.005$), while the relationship with head circumference was only borderline significant ($P < 0.095$). Although systolic blood pressure was borderline significantly inversely related to offspring skinfold thickness when added to the full model ($P < 0.055$), adjusting for blood pressure left the coefficients relating CRP to all four outcomes unchanged (results not shown; coefficient relating blood pressure to offspring outcomes all $P > 0.5$ for other three models).

Discussion

In this sample of women from metropolitan Cebu, Philippines, a measure of inflammation, CRP, is inversely related to multiple offspring anthropometric outcomes measured soon after birth, including weight, length, adiposity, and to a lesser extent, head circumference. Although women with higher CRP had

modestly truncated gestational ages at birth, the relationships with CRP were largely independent of gestational age, pointing to slower fetal growth as a likely effect. Furthermore, the inverse relationship between pregnancy CRP and offspring birth outcomes were linear across the full range of CRP, suggesting that any elevation in CRP predicts a proportionately attenuated offspring birth size. These findings point to maternal inflammation as an important influence on the gestational milieu that impacts fetal development, and by implication, long-term health in the next generation.

Our findings are consistent with the limited literature examining the effects of CRP variation in the normal range, in the absence of clinical pathology, as a predictor of offspring birth outcomes. In a subset of 1481 Hyperglycemia and Adverse Pregnancy Outcome study female participants, CRP levels in late pregnancy (24–32 weeks) were found to be negatively associated with offspring weight, sum of skinfolds and percent body fat at birth.¹⁷ Similarly, in a small study, women with normal pregnancies resulting in intrauterine growth restriction had higher CRP in early pregnancy (10–14 weeks) than those with expected fetal growth patterns.⁴ Furthermore, an inverse relationship between maternal CRP across all trimesters and birth weight Z-scores was found among Brazilian women.³⁶ Together these findings demonstrate the negative association of moderately elevated maternal CRP on fetal growth among women experiencing normal pregnancy from diverse populations.

In support of the findings of Lowe *et al.*,¹⁷ we find that higher levels of maternal CRP during pregnancy predict not only a reduction in birth weight and length, but also body fat, as measured by skinfold thicknesses. This finding suggests that maternal inflammation has effects on multiple, distinct domains of fetal growth. Body length reflects chondrocyte differentiation in the growth plates of growing bones, and is driven by nutrients and growth factors. Fat deposition, in contrast, is most rapid during the final trimester of gestation,³⁷ when the fat content of the fetus rises dramatically to the unusually high percentage body fat that characterizes human

Table 3. Multiple regression models predicting offspring birth outcomes^a

	CRP	Maternal fat	CRP + fat	CRP + fat + gestational age ^b
Weight (kg)				
Log-CRP	-0.027 ± 0.018		-0.058 ± 0.019 [†]	-0.047 ± 0.017 [†]
BMI		0.076 ± 0.021 [‡]	0.085 ± 0.021 [‡]	0.081 ± 0.020 [‡]
Triceps SF		0.017 ± 0.022	0.026 ± 0.022	0.026 ± 0.022
Model R ²	0.128	0.172	0.190	0.225
Length (cm)				
Log-CRP	-0.171 ± 0.094 ⁻		-0.301 ± 0.098 [†]	-0.259 ± 0.092 [†]
BMI		0.232 ± 0.093 [*]	0.273 ± 0.093 [†]	0.262 ± 0.092 [†]
Triceps SF		0.125 ± 0.110	0.176 ± 0.108	0.174 ± 0.108
Model R ²	0.157	0.175	0.193	0.209
Head circumference (cm)				
Log-CRP	-0.042 ± 0.059		-0.132 ± 0.064 [*]	-0.102 ± 0.060 ⁻
BMI		0.365 ± 0.068 [‡]	0.383 ± 0.070 [‡]	0.372 ± 0.068 [‡]
Triceps SF		-0.068 ± 0.071	-0.046 ± 0.072	-0.048 ± 0.070
Model R ²	0.149	0.205	0.212	0.225
Sum of skinfolds (mm)				
Log-CRP	-0.358 ± 0.194 ⁻		-0.597 ± 0.209 [†]	-0.520 ± 0.190 [†]
BMI		0.023 ± 0.260	0.104 ± 0.255	0.086 ± 0.250
Triceps SF		0.604 ± 0.266 [*]	0.704 ± 0.262 [†]	0.702 ± 0.263 [†]
Model R ²	0.139	0.151	0.165	0.174

CRP, C-reactive protein; BMI, body mass index; SF, skinfold.

^aAll models also adjust for (not shown) days after birth of anthropometry measurement, offspring gender, maternal parity and age during pregnancy visit and mother's adult height.

^bGestational age coefficient not shown.

⁻ $P < 0.1$; ^{*} $P < 0.05$; [†] $P < 0.01$; [‡] $P < 0.0001$.

newborns.³⁸ Fat deposition accounts for 50% of the energy costs of growth at 27 weeks of gestation, and this value rises to 90% by term.³⁹

We find similarly strong associations between pregnancy CRP and offspring body fat, body length and body weight. In contrast, the relationship with head circumference was only of borderline significance. Although differences in measurement error could contribute to differences in effect size across measures, it is notable that the measurement typified by the lowest technical error of measurement (TEM), head circumference,⁴⁰ exhibited the weakest relationships with CRP, whereas that typically with the highest TEM, skinfolds, showed the strongest relationships with CRP. We interpret our findings as evidence that maternal inflammation reduces fetal fat deposition and skeletal growth. In contrast, the comparably modest effects on head circumference are consistent with brain sparing, which has been shown previously in the context of conditions that limit delivery of substrate or oxygen to the fetus, such as placental insufficiency.^{41,42}

Our prior analyses in Cebu showed that adult CRP is inversely related to that individual's own weight at birth,⁴³ a finding reported in a number of other populations.^{26,31} Viewed alongside these studies, our present findings suggest that inflammation could serve as a pathway linking restricted fetal growth and elevated inflammatory status across generations.

Specifically, factors that lead to chronically elevated inflammation and CRP during pregnancy predict giving birth to a modestly smaller baby, who in turn is predicted to have elevated CRP as a result of their small birth size. At Cebu, we have previously reported similar evidence for reciprocal effects between birth outcomes and adult phenotypes related to hypothalamic–pituitary–adrenal (HPA) axis function at Cebu,^{44,45} and alterations in glucose metabolism, especially with maternal obesity and gestational diabetes, can operate in a similar intergenerational fashion.⁴⁶ Future research will be needed to evaluate the collective impact of these interrelated metabolic, endocrine and immune pathways as pathways linking adverse health across generations.⁴⁷

Several limitations of this study warrant mention. Because our sample is a population-based survey (and not a hospital-based study), interviews and data collection occurred within participants' homes. As a result, our anthropometric measures of offspring were not obtained immediately after birth, but on variable days after birth. Almost all of the data were collected within 4 days of pregnancy, and we adjusted for day of measurement, minimizing any impact of postnatal growth processes. In addition, we rely upon a single measure of CRP as an indication of maternal inflammatory status, where using the multiple of several measures would provide more reliable estimates. As such, our models, which successfully identify

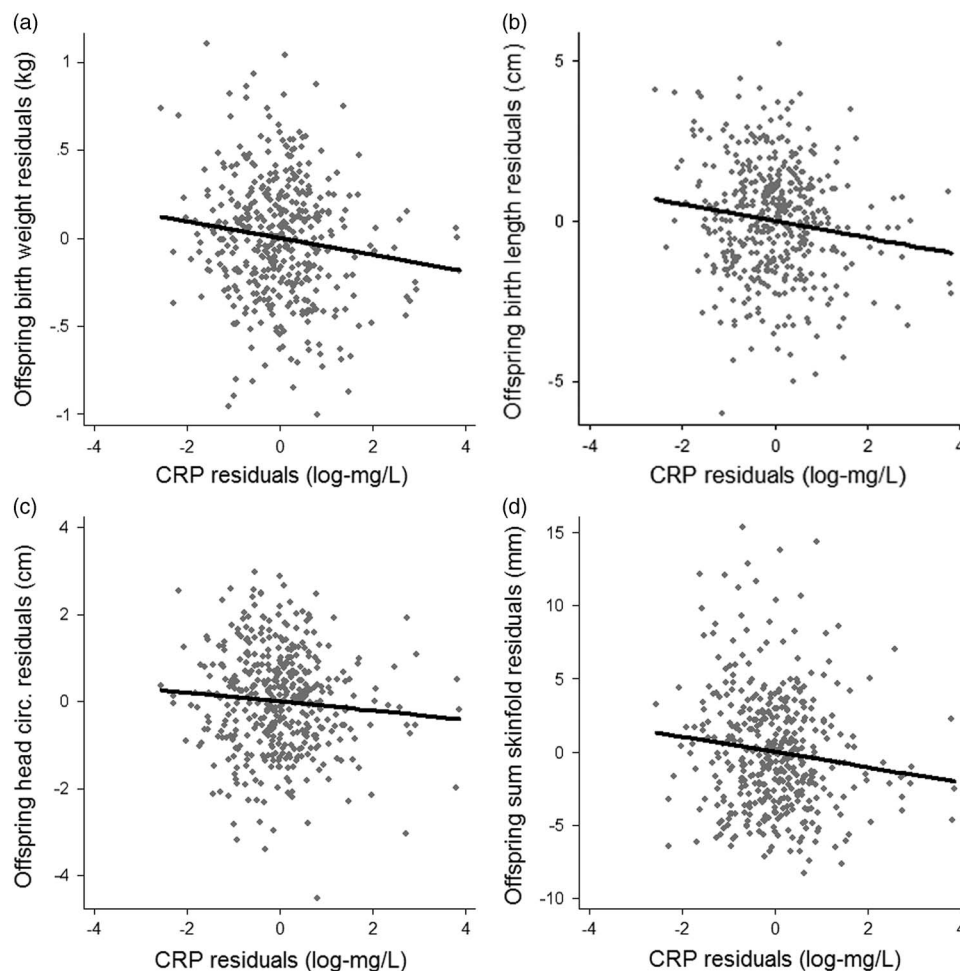


Fig. 1. Offspring birth outcomes predicted by maternal C-reactive protein (CRP) during pregnancy. (a) Offspring birth weight, (b) offspring birth length, (c) offspring head circumference, and (d) offspring sum of skinfolds predicted by maternal CRP. Values represent residuals from models adjusted for gestational age at delivery, postnatal day of offspring anthropometry measurement, pregnancy order, offspring gender, maternal age, height pre-pregnancy body mass index and pregnancy triceps skinfolds (see models in Table 3).

significant relationships with birth outcomes, almost certainly underestimate the true intergenerational impact of maternal inflammation.

In sum, we find that maternal inflammation during pregnancy, as measured by CRP, is inversely related to multiple neonatal measures of offspring anthropometry in this sample of women living in metropolitan Cebu, the Philippines. Relationships were present across the full range of CRP, pointing to negative effects of any incremental increase in maternal inflammation. Maternal CRP was negatively related to measures of skeletal growth and body fat deposition, while an attenuated relationship with head circumference was consistent with the principle of brain sparing. These findings point to endogenous maternal immune processes as an important contributor to the fetal gestational milieu, and highlight inflammation as a potential pathway linking maternal environments and health with development and health in the next generation.

Acknowledgments

The authors thank the researchers at the USC-Office of Population Studies Foundation, Inc., University of San Carlos, Cebu City, Philippines, for their role in the study design and data collection, and the study participants, who generously provided their time.

Financial Support

National Science Foundation (Grant no.: BCS-0746320 and BCS-1440564).

Conflicts of Interest

None.

References

1. Barker D. *Mothers, Babies, and Health in Later Life* (2nd edn) 1998. Churchill Livingstone: Edinburgh, UK.

2. Gluckman PD, Hanson MA, Cooper C, Thornburg KL. Effect of in utero and early-life conditions on adult health and disease. *New Engl J Med.* 2009; 359, 61–73.
3. Guven MA, Coskun A, Ertas IE, et al. Association of maternal serum CRP, IL-6, TNF-alpha, homocysteine, folic acid and vitamin B₁₂ levels with the severity of preeclampsia and fetal birth weight. *Hypertens Pregnancy.* 2009; 28, 190–200.
4. Tjoa ML, van Vugt JMG, Go ATJJ, et al. Elevated C-reactive protein levels during first trimester of pregnancy are indicative of preeclampsia and intrauterine growth restriction. *J Reprod Immunol.* 2003; 59, 29–37.
5. Akum A, Kuoh AJ, Minang JT, et al. The effect of maternal, umbilical cord and placental malaria parasitaemia on the birthweight of newborns from South-western Cameroon. *Acta Paediatr.* 2005; 94, 917–923.
6. Blas MM, Canchihuaman FA, Alva IE, Hawes SE. Pregnancy outcomes in women infected with Chlamydia trachomatis: a population-based cohort study in Washington State. *Sex Trans Infect.* 2007; 83, 314–318.
7. R Romero, Moshe M. Infection and preterm labor. *Clin Obstet Gynecol.* 1988; 31, 553–584.
8. Bullen BL, Jones NM, Holzman CB, et al. C-reactive protein and preterm delivery: clues from placental findings and maternal weight. *Reprod Sci.* 2013; 20, 715–722.
9. Pitiphat W, Gillman MW, Joshupura KJ, et al. Plasma C-reactive protein in early pregnancy and preterm delivery. *Am J Epidemiol.* 2005; 162, 1108–1113.
10. Hwang HS, Kwon JY, Kim MA, Park YW, Kim YH. Maternal serum highly sensitive C-reactive protein in normal pregnancy and pre-eclampsia. *Int J Gynaecol Obstet.* 2007; 98, 105–109.
11. Sacks GP, Seyani L, Lavery S, Trew G. Maternal C-reactive protein levels are raised at 4 weeks gestation. *Human Reprod.* 2004; 19, 1025–1030.
12. Teran E, Escudero C, Calle A. C-reactive protein during normal pregnancy and preeclampsia. *Int J Gynaecol Obstet.* 2005; 89, 299–300.
13. Kuzawa CW, Adair LS, Borja J, McDade TW. C-reactive protein by pregnancy and lactational status among Filipino young adult women. *Am J Human Biol.* 2013; 25, 131–134.
14. McDade TW, Borja JB, Largado F, Adair LS, Kuzawa CW. Adiposity and chronic inflammation in young women predict inflammation during normal pregnancy in the Philippines. *J Nutr.* 2016; 146, 353–357.
15. Redman C, Sacks GP, Sargeant IL. Pregnancy: an excessive maternal inflammatory response to pregnancy. *Am J Obstet Gynecol.* 1999; 180, 499–506.
16. Han YS, Ha EH, Park HS, Kim YJ, Lee SS. Relationships between pregnancy outcomes, biochemical markers and pre-pregnancy body mass index. *Int J Obes.* 2011; 35, 570–577.
17. Lowe LP, Metzger BE, Lowe WL Jr., et al. Inflammatory mediators and glucose in pregnancy: results from a subset of the Hyperglycemia and Adverse Pregnancy Outcome (HAPO) Study. *J Clin Endocrinol Metabol.* 2010; 95, 5427–5434.
18. Sen S, Rifas-Shiman SL, Shivappa N, et al. Dietary inflammatory potential during pregnancy is associated with lower fetal growth and breastfeeding failure: results from project viva. *J Nutr.* 2016; 146, 728–736.
19. Gershov D, Kim S, Brot N, Elkon KB. C-reactive protein binds to apoptotic cells, protects the cells from assembly of the terminal complement components, and sustains an antiinflammatory innate immune response: implications for systemic autoimmunity. *J Exp Med.* 2000; 192, 1353–1364.
20. Lam C, Lim KH, Karumanchi SA. Circulating angiogenic factors in the pathogenesis and prediction of preeclampsia. *Hypertension.* 2005; 46, 1077–1085.
21. Ernst GD, de Jonge LL, Hofman A, et al. C-reactive protein levels in early pregnancy, fetal growth patterns, and the risk for neonatal complications: the Generation R Study. *Am J Obstet Gynecol.* 2011; 205, 132.e1–132.e12.
22. Ali Z, Bokhari FA, Zaki S, et al. Correlation of CRP levels in third trimester with fetal birth weight in preeclamptic and normotensive pregnant women. *J Coll Phys Surg Pak.* 2015; 25, 111–114.
23. Redman CWG. Preeclampsia and the systemic inflammatory response. *Sem Nephrol.* 2004; 24, 565–570.
24. Ertas IE, Kahyaoglu S, Yilmaz B, et al. Association of maternal serum high sensitive C-reactive protein level with body mass index and severity of pre-eclampsia at third trimester. *J Obstet Gynaecol Res.* 2010; 36, 970–977.
25. Malek A, Bersinger NA, Di Santo S, et al. C-reactive protein production in term human placental tissue. *Placenta.* 2006; 27, 619–625.
26. Sattar N, McConnachie A, O'Reilly D, et al. Inverse association between birth weight and C-reactive protein concentrations in the MIDSPAN Family Study. *Arterioscler Thromb Vasc Biol.* 2004; 24, 583–587.
27. deRosset L, Strutz KL. Developmental origins of chronic inflammation: a review of the relationship between birth weight and C-reactive protein. *Ann Epidemiol.* 2015; 25, 539–543.
28. McDade TW. Early environments and the ecology of inflammation. *Proc Natl Acad Sci.* 2012; 109(Suppl. 2), 17281–17288.
29. McDade TW, Rutherford JN, Adair L, Kuzawa C. Population differences in associations between C-reactive protein concentration and adiposity: comparison of young adults in the Philippines and the United States. *Am J Clin Nutr.* 2009; 89, 1237–1245.
30. Skilton MR, Viikari JSA, Juonala M, et al. Fetal growth and preterm birth influence cardiovascular risk factors and arterial health in young adults. *Arterioscler Thromb Vasc Biol.* 2011; 31, 2975–2981.
31. Tzoulaki I, Jarvelin MR, Hartikainen AL, et al. Size at birth, weight gain over the life course, and low-grade inflammation in young adulthood: northern Finland 1966 Birth Cohort study. *Eur Heart J.* 2008; 29, 1049–1056.
32. Adair LS, Popkin BM, Akin JS, et al. Cohort profile: The Cebu Longitudinal Health and Nutrition Survey. *Int J Epidemiol.* 2011; 40, 619–625.
33. Lohman TG, Roche AF, Martorell R. *Anthropometric Standardization Reference Manual.* 1988. Human Kinetics Books: Champaign, IL.
34. McDade TW, Burhop J, Dohnal J. High-sensitivity enzyme immunoassay for C-reactive protein in dried blood spots. *Clin Chem.* 2004; 50, 652–654.
35. McDade TW. Development and validation of assay protocols for use with dried blood spot samples. *Am J Hum Biol.* 2014; 26, 1–9.
36. de Oliveira LC, Franco-Sena AB, Farias DR, Rebelo F, Kac G. Maternal C-reactive protein concentrations during pregnancy and birth weight in a prospective cohort in Rio de Janeiro, Brazil. *J Matern Fetal Neonatal Med.* 2016; 1–8 [epub ahead of print].

37. Widdowson EM, McCance RA. A review: new thoughts on growth. *Pediatr Res.* 1975; 9, 154–156.
38. Kuzawa CW. Adipose tissue in human infancy and childhood: an evolutionary perspective. *Am J Phys Anthropol.* 1998; 27, 177–209.
39. Haggarty P. Fatty acid supply to the human fetus. *Annu Rev Nutr.* 2010; 30, 237–255.
40. de Onis M, Onyango AW, Borghi E, Garza C, Yang H. Comparison of the World Health Organization (WHO) Child Growth Standards and the National Center for Health Statistics/WHO international growth reference: implications for child health programmes. *Public Health Nutr.* 2006; 9, 942–947.
41. Wladimiroff JW, Tonge HM, Stewart PA, Reuss A. Severe intrauterine growth retardation; assessment of its origin from fetal arterial flow velocity waveforms. *Eur J Obstet Gynecol Reprod Biol.* 1986; 22, 23–28.
42. Baschat AA. Fetal responses to placental insufficiency: an update. *BJOG.* 2004; 111, 1031–1041.
43. McDade TW, Rutherford J, Adair L, Kuzawa CW. Early origins of inflammation: microbial exposures in infancy predict lower levels of C-reactive protein in adulthood. *Proc R Soc B.* 2010; 277, 1129–1137.
44. Thayer ZM, Feranil AB, Kuzawa CW. Maternal cortisol disproportionately impacts fetal growth in male offspring: evidence from the Philippines. *Am J Human Biol.* 2012; 24, 1–4.
45. Lee J, Fried R, Thayer Z, Kuzawa CW. Preterm delivery as a predictor of diurnal cortisol profiles in adulthood: evidence from Cebu, Philippines. *Am J Human Biol.* 2014; 26, 598–602.
46. Benyshek DC. The “early life” origins of obesity-related health disorders: new discoveries regarding the intergenerational transmission of developmentally programmed traits in the global cardiometabolic health crisis. *Am J Phys Anthropol.* 2013; 31, 79–93.
47. Kuzawa CW, Sweet E. Epigenetics and the embodiment of race: developmental origins of US racial disparities in cardiovascular health. *Am J Human Biol.* 2008; 21, 2–15.