Prenatal exposure to diethylstilbestrol and long-term impact on the breast and reproductive tract in humans and mice

R. R. Newbold^{*†}

National Institute of Environmental Health Sciences (NIEHS), National Institutes of Health (NIH), Department of Health and Human Services (DHHS), North Carolina, USA

The term 'developmental origins of health and disease' (DOHaD) originally referred to delayed effects of altered maternal factors (e.g. smoking or poor nutrition) on the developing offspring, but it now also encompasses early life exposure to environmental chemicals, which can cause an unhealthy prenatal environment that endangers the fetus and increases its susceptibility to disease later in life. Prenatal exposure to the pharmaceutical diethylstilbestrol (DES) is a well-known DOHaD example as it was associated in the 1970s with vaginal cancer in daughters who were exposed to this potent synthetic estrogen before birth. Subsequently, numerous long-term effects have been described in breast and reproductive tissues of DES-exposed humans and experimental animals. Data reviewed suggest that the prenatal DES-exposed population should continue to be monitored for potential-increased disease risks as they age. Knowledge of sensitive developmental periods, and the mechanisms of DES-induced toxicities, provides useful information in predicting potential adverse effects of other environmental estrogens.

Received 23 August 2011; Revised 14 November 2011; Accepted 16 November 2011; First published online 19 December 2011

Key words: breast cancer, critical windows, epigenetics, fibroids, hormonal carcinogenesis

Introduction

Diethylstilbestrol (DES), a potent synthetic estrogen, was extensively prescribed worldwide to pregnant women from the 1940s to the 1970s, with the mistaken belief that it could prevent miscarriages and other pregnancy complications.¹ It was initially given to at-risk pregnancies, but ultimately also prescribed for normal pregnancies to make them 'healthier'. Estimates ranging upward to 10 million people (mothers, daughters and sons) were exposed to DES in the United States. Subsequently, DES was found therapeutically ineffective in reducing miscarriages, and it was linked to a rare form of cancer termed 'vaginal adenocarcinoma' in a small number (<0.1%) of adolescent daughters of women who took the drug while pregnant.² Although DES was originally identified as a carcinogen based on its effects in the vagina, the extent of its toxicity was later found to be more extensive as it was associated with numerous other medical consequences. DES was later linked with frequent noncancerous (benign) reproductive problems in approximately 90-95% of DES-exposed daughters: reproductive tract malformations and dysfunction, poor pregnancy outcome, ectopic pregnancies and premature labor and births were reported (for review see Giusti et al.1). Furthermore, as

DES-exposed daughters aged, they were at a higher risk for developing breast cancer than their unexposed, age-matched counterparts. DES-exposed daughters > 40 years exhibited a statistically significant increase (2–2.5×) in risk for developing breast cancer;^{3–6} this increased risk was more pronounced (3×) in DES-exposed women over 50 years of age, although the sample size was small.⁵ DES-exposed mothers were also found to be at increased risk for breast cancer.¹ Prenatally DES-exposed sons experienced a range of reproductive tract problems including malformations (hypospadias, microphallus and retained testes) and increased genital/urinary inflammation.^{7–9}

Although reports in the experimental animal literature dating back four decades described the carcinogenic effects of DES, it was not until its effects in humans were reported that its adverse consequences were considered seriously. On the basis of a wealth of accumulated scientific information from humans and experimental animals, DES is now a well-documented 'transplacental carcinogen': it crosses the placenta, reaches the fetus, adversely affects developing tissues/ organs, and causes a myriad of problems including breast and reproductive tract cancer.^{7–10} DES caused a major medical catastrophe that continues unfolding today.

Although DES was banned years ago for use during pregnancy, experimental studies continue to explore mechanism(s) through which DES causes its adverse effects. Questions such as how does DES cause abnormalities (ranging from structural malformations to cellular and molecular defects) in breast and reproductive tissues have led to the development of

^{*}Address for correspondence: R. R. Newbold, National Toxicology Program, National Institute of Environmental Health Sciences (NIEHS), P.O. Box 12233, Mail Drop K2-15, Research Triangle Park, NC 27709, USA. (Email newbold1@niehs.nih.gov)

[†] The author is retired but the research was conducted while employed at NIEHS.

Tissue affected	Category of effect	Developmental effect	♀ Human	♀ Mouse
Reproductive tract	Dysfunction	Subfertility/infertility	+	+
	,	Poor reproductive outcome	+	+
		Altered estrous (menstrual) cycles	+	+
	Malformations	Oviduct, uterus, cervix and vagina	+	+
		Paraovarian cysts of mesonephric origin	+	+
		Retained mesonephric remnants	+	+
	Cellular abnormalities	Lesions (proliferative)		
		Oviduct	+	+
		• Uterus/cervix	+	+
		Uterine fibroids	+	+
		Vaginal adenosis	+	+
		Vaginal adenocarcinoma	+	+
	Molecular abnormalities	Oviduct and uterus	?	+
Breast/mammary gland	Dysfunction	Decreased lactation	?	+
	Malformation	Increased or decreased TEBs and abnormal branching depending on dose	?	+
	Cellular abnormalities	Breast cancer	+	+
	Molecular abnormalities	Abnormal gene expression	?	+

Table 1. Comparative developmental effects of exposure to DES in humans and mice demonstrating the DOHaD phenomena.

DES, diethylstilbestrol; DOHaD, developmental origins of health and disease; TEBs, terminal end buds.

experimental animal models to study the impact of estrogens and other endocrine-disrupting chemicals (EDCs) on differentiating tissues. As the murine model using prenatally DES-exposed outbred mice has successfully duplicated and predicted abnormalities reported in similarly exposed humans (for review see Newbold^{11,12}), this review compares critical windows in breast and reproductive tract differentiation in mice and humans that can be perturbed by DES. For brevity, the focus is on females (Table 1), although DES-induced abnormalities in males are equally important. Investigating DES toxicities has great potential for advancing our scientific knowledge about this chemical and other less-potent environmental estrogens, as well as providing caution in using any drug during pregnancy. Further, studying the vast array of DES health effects will contribute to a better understanding of the role of estrogens in normal and abnormal developmental processes, hormonal imprinting and carcinogenesis.

DES exposure provides solid evidence, and indeed proof of principle, that prenatal chemical exposure can have latent effects and lead to diseases later in life in animals and humans. The developmental origins of health and disease (DOHaD) field of scientific investigation points out the vulnerability of the developing fetus to perturbation by DES and other chemicals with estrogenic activity.

The DOHaD concept for environmental chemicals

Although the placenta was once believed to provide complete protection to the unborn child, it is now well established that it only provides partial protection from the external environment, as indicated by the large literature based on the DOHaD paradigm focusing on diet and nutrition, as well as environmental chemicals. Many drugs and chemicals freely pass through the once called 'placental barrier'. As fetal and neonatal developments are characterized by rapid cell division and differentiation, coupled with complex patterns of cell signaling, exposure to chemicals during these active periods can readily disrupt differentiation and result in abnormal cell proliferation and/or differentiation at inappropriate times. These abnormal cells can then give rise to additional generations of altered cells. Often, damage that occurs before birth to actively dividing cells is irreversible (permanent) and likely due to epigenetic programming, whereas similar adult exposures can be reversible, disappearing when the chemical insult is removed. As development is controlled by a myriad of endocrine signals, chemicals with hormonal activity can easily interrupt or alter these signals resulting in changes that are not apparent until much later in life.

The developing organism lacks many of the protective mechanisms that are available to the adult (detoxifying enzymes, a fully competent immune system, DNA repair mechanisms, mature liver metabolism and blood/brain barrier).¹³ In addition, the fetus and neonate have higher metabolic rates as compared with adults, which can cause increased sensitivity to chemicals. Furthermore, prenatal development is an important time of chemical susceptibility as this is when epigenetic marks (programming) are set. Epigenetic events occurring during embryogenesis and development involve heritable changes in gene function and regulation that happen independently from the DNA sequence.

Thus, altered epigenetic programming caused by exposure to DES or other EDCs can persist throughout the life of the organism and manifest as latent disease including cancer.¹⁴

In the fields of nutritional and endocrine disruption, developmental exposure to environmental stressors, drugs and chemicals has been extensively studied and well recognized to interfere with complex differentiating endocrine signaling pathways resulting in adverse consequences later in life.^{15–17} Prenatal DES exposure is the best example of DOHaD involving detrimental effects of chemical exposure during critical windows of differentiation and latent occurrence of disease.

An overriding concept in DOHaD suggests a lag time between time of exposure to environmental chemicals and disease manifestation, but there are other key aspects:

- (1) Time-specific, tissue-specific and dose-specific effects can occur. Chemical exposure may have an entirely different effect on developing organisms as compared with adults. Further, time-specific effects are due to the stage of differentiation of the target tissues when exposure occurred. For example, Hatch *et al.*¹⁸ reported that DES-exposed women are at greater risk of developing high-grade squamous cancer in the reproductive tract if exposure occurs early in differentiation within 7 weeks of gestation rather than later. Regarding tissue-specific effects, DES mainly affects estrogen target tissues such as breast and the reproductive tract. Finally, dose may determine the effect with low doses of DES causing functional changes and tumorigenesis, whereas high doses are teratogenic.
- (2) An environmental chemical such as DES can act alone or with other environmental stressors. Exposure to other environmental stressors could trigger or intensify disease.
- (3) The abnormality can manifest in various ways as the occurrence of a disease that otherwise would not have happened; an increase in risk for a disease that would normally be of lower prevalence; or perhaps an earlier onset of a disease that would normally have occurred; or an exacerbation of the disease. For example, prenatal DES exposure causes vaginal cancer, a disease that otherwise would not have occurred in a young women; increased breast cancer is an example of disease that would normally be of lower prevalence.
- (4) The abnormality can have variable latency from onset in development to early childhood, puberty or adulthood depending on the time of exposure and stage of differentiation of the affected tissue or organ.
- (5) Prenatal exposure to an environmental insult can lead to permanently altered developmental programming and aberrant function of a cell, organ or system, resulting in an individual who is more susceptible to certain diseases later in life. This altered imprinting likely involves epigenetic mechanisms, which are active during differentiation and development, and are known to be influenced by estrogens. With epigenetic alterations, genes are not mutated but

changes occur in how the gene is regulated. The effects of developmental exposures persist because altered epigenetic signaling persists as cells divide throughout life. Thus, early exposure to chemicals can alter epigenetic marks that lead to functional changes in genes, which in turn lead to abnormal tissues that lead to diseases including cancer later in life.

- (6) Prenatal chemical exposure can cause long-lasting adverse effects that may include not only functional, cellular and molecular abnormalities but also structural (malformation) abnormalities (see Table 1 for an example). These different types of effects are probably because of the differences in actual timing and dose of DES exposures with lower doses causing functional changes and higher doses causing structural malformations. These abnormalities may persist and lead to increased diseases such as cancer later in life. Development of new sensitive molecular markers of developmental exposures will contribute useful predictive and preventive information for adult human health.^{14,19–22}
- (7) Extrapolation of risk from environmental exposures can be difficult because abnormalities do not always follow a monotonic dose-response relationship; low-dose effects may be different from those occurring at higher doses because different mechanisms are involved.²³ For example, low doses of DES cause increased weight gain, whereas higher doses do not, and may actually cause weight loss. Low-dose effects are of concern because exposure of the general public occurs in this dose range, whereas occupational and pharmaceutical exposures usually occur at higher doses.^{10,24,25}
- (8) Individual organisms may have different effects to chemicals because of differences in genetic backgrounds. However, identical genotypical twins may have different effects because of different epigenetic programming.

Together, these are important components of DOHaD and apply to DES and other environmental chemicals. These aspects have recently been reviewed by Heindel and Newbold.²⁶ To further examine the DOHaD concept and to understand the unique sensitivity of developing tissues to perturbation by chemicals, sensitive windows of breast and reproductive development in humans and mice are described.

Sensitive windows in perinatal development

Mammalian development involves a complex and wellorganized series of events to grow from a single cell to a fully developed organism at birth. Processes including cell division, proliferation, differentiation and migration are involved and are closely regulated by hormones that communicate information between specialized cells, tissues and organs. For over 50 years, embryonic and fetal development was assumed to occur by 'the unfolding of a rigid genetic program' where the environmental factors played no significant function (for review see Soto and Sonnenschein²⁷). However, this narrow interpretation of developmental events has evolved because experimental and epidemiological studies reveal the developmental plasticity of the fetus and neonate. In fact, it has become increasingly apparent that environmental factors and stressors including nutrition and toxic chemicals can dramatically alter developmental signals.

Many biological processes involved in mammalian growth and differentiation are conserved across species including differentiation of breast and reproductive tissues. Although developmental processes are similar in humans and mice, exact timing of events varies with many occurring entirely prenatally in humans, but prenatally and neonatally in mice. The website http://www.endocrinedisruption.com/prenatal. criticalwindows.overview.php compare the timing of developmental events in humans and mice. Extrapolations from experimental animals to humans are most successful when similar periods of differentiation are compared rather than same chronological age,²⁸ and when human exposures are well documented including dose and timing of exposure.

DES and breast/mammary gland development

Critical events in breast development start early in fetal life, continue into postnatal life and are then followed by exponential epithelial outgrowth during puberty; the breast becomes completely competent later in life when it transitions into its lactational role in pregnancy. These well-defined developmental stages occur in humans and mice and are regulated by endogenous hormones, mainly estrogen and progesterone. These hormones mediate their effects by interacting with their receptors (for estrogens, the interaction is with its estrogen receptors (ERs) alpha and beta (α and β) in breast tissue). Excessive estrogen exposure during fetal life is a known risk factor for breast cancer; for example, a positive correlation exists between twin female births (which have increased intrauterine levels of estrogens) and breast cancer later in life of the daughters from these pregnancies.²⁹ ERs can also interact with exogenous estrogens produced outside the body such as DES, which mimic naturally occurring estrogens, bind to their ERs and interact with developing breast tissue.

Precise mechanisms by which prenatal DES exposure causes breast cancer are unclear; however, the carcinogenic process is complex and multistep, and it likely originates in the terminal duct lobular units (TDLUs) in humans and the counterpart terminal end buds (TEBs) in mice. Support for these as carcinogenic sites comes from the 7,12-dimethylben[a]athracene (DMBA)-induced rodent mammary carcinoma model that is well established and appropriate for studying human breast cancer.³⁰ TEBs have very high proliferation rates and remove DMBA adducts less efficiently than the more-differentiated epithelial cells in the mammary gland. Further, the presence of TEBs at the time of carcinogen exposure is positively associated with tumor multiplicity and severity.³¹

Humans

In the first trimester, as early as 4–5 weeks of prenatal life, breast development appears as a thickening of the milk lines on the ventral surface of the fetus. Epithelial budding and branching starts around 6–20 weeks of gestation; breast buds extend, producing cords of epithelium that grow through the underlying mesenchyme (undifferentiated stroma). The mesenchyme develops supporting stroma between weeks 20 and 32. Epithelial cords become hollow and develop a lumen during the last 2 months of gestation; at this time, ductal and lobuloalveolar branching occurs yielding a primitive gland at birth composed of ducts ending in TDLUs. Near birth, the nipple forms by invagination of the breast surface.³²

As DES use during pregnancy often started before 9 weeks of gestation and continued throughout pregnancy, it is clear that the primitive breast structure was present at the time of DES exposure. Furthermore, differentiating breast tissue has the necessary cellular machinery including ERs to respond to DES; ERs were localized in normal mammary epithelial cells as early as gestational week 30 and progesterone receptors by week 41.³³ Epidemiological studies following prenatally DES-exposed women verify that DES acts as a latent breast carcinogen,⁵ thus providing proof of the DOHaD phenomena.

Mice

Developmental events in the rodent mammary gland are similar to the human breast. Mammary gland development begins approximately on gestational day (d.p.c.) 10.5; by 11.5 d.p.c., five placodes appear along each presumptive mammary line as lens-shaped ectodermal structures that invaginate into the dermis by 13.5 d.p.c. The mesenchyme adjacent to the mammary epithelium becomes denser than the surrounding mesenchyme and develops several concentric layers of fibroblasts, which align around the epithelial compartment.³⁴ At 15.5 d.p.c., epithelial buds start to elongate into cords, and on 16 d.p.c. the primary cord undergoes an increase in proliferation as it pushes through the surrounding mammary mesenchyme and grows through the fetal fat pad. By 18 d.p.c., branching of the structure is readily apparent and a lumen starts to form, which differentiates into a mammary duct.³⁵

ER is first expressed in murine mammary tissue around 12.5 d.p.c. in the mesenchyme surrounding the bud.³⁶ Autoradiographic studies show specific binding of labeled DES in the mesenchyme surrounding the epithelial anlagen at 16 d.p.c., suggesting functional receptors at this age and the ability of DES to bind to fetal mammary gland tissue.³⁷ By 18 d.p.c., just before birth, ERs are found predominately in the stroma and some mammary epithelial cells.³⁸ In neonatal life, ER- α is mainly expressed in mammary epithelium. After birth, the mammary gland grows with body growth until puberty, approximately 4 weeks when estrogen levels significantly rise. In response to these elevated estrogen levels at puberty, as in the human, extensive mammary gland growth occurs. At the end of the branched mammary ducts, bulbous epithelial TEBs develop, which have high mitotic (proliferative)

rates and apoptotic (programmed cell death) activity. The cap cells of the TEBs rapidly divide, permitting ductal elongation and the ability of the duct to change direction in the fat pad, whereas apoptosis is responsible for formation of the ductal lumen and extension of the growing duct.³⁹ The branching mammary gland structure invades the stroma until it reaches the edge of the fat pad, establishing a network of ducts and a few alveolar buds.³⁹ This morphology remains quiescent until pregnancy, although fluctuations do occur during different stages of the estrous cycle for mice (v. menstrual cycle for humans). In response to pregnancy, the mammary gland undergoes dramatic proliferation resulting in a massive number of alveolar buds to prepare for lactation. Once lactation is completed, the mammary gland undergoes rapid involution associated with widespread apoptosis and stromal remodeling so that it can return to its pre-pregnancy state. Similar events occur in human breast tissue in response to pregnancy.

Experimental studies with rodents support the carcinogenic effects of DES in the human breast. Fetal DES exposure increases mammary tumorigenesis and tumor multiplicity.^{40–48} In addition, neonatal DES exposure of rodents (a time that corresponds to prenatal development in humans) also increases the incidence of cancer in experimental mammary tumor models^{49–54} (for review see Fenton and Newbold⁷²). Recently, altered gene expression was demonstrated in rat TEBs exposed neonatally to DES, thus showing that differentiation- and development-related genes are involved in altered TEB structural and cellular abnormalities in DES-exposed mammary tissue and its increased propensity to develop cancer.⁵⁵

Similarities/differences

Developmental milestones in human breast and mouse mammary gland development are similar, although timing of events differs slightly. Ductal branching originates in prenatal life for both species, but minor differences in the extent of ductal branching of the mammary gland occur at birth between species, and alveolar buds are prominent in human infants but not mice. The growing mammary ducts end in TEBs in mice v. TDLUs in humans; these are morphologically analogous structures, which contain actively proliferating cells, and potentially stem cells, accounting for their sensitivity to carcinogenic agents. Most importantly, breast and mammary tissue at all stages of differentiation and development have the ability to respond to endogenous estrogens produced in the body, as well as exogenous estrogens produced outside the body (such as DES).

DES and reproductive tract development

Critical events in mammalian reproductive tract differentiation start in prenatal life similar to mammary gland differentiation. Early in embryonic development, there is an undifferentiated stage (sometimes referred to as the indifferent period) in which the sex of the embryo cannot be determined. At this stage, the gonads have not developed into either testis or ovary and all embryos have a double set of genital ducts, Mullerian (paramesonephric) and Wolffian (mesonephric) ducts. In the female, as sex differentiation proceeds, the Mullerian duct differentiates into oviduct, uterus, cervix and upper vagina, whereas the mesonephric duct regresses and largely disappears. In the male, under the influence testicular secretions, the mesonephric duct forms the epididymis, vas deferens, seminal vesicles and ejaculatory ducts, whereas the Mullerian duct regresses.⁵⁶ DES exposure during this critical window of sex differentiation results in alterations in female and male gonads, and reproductive tracts including retention of the opposite duct system in both sexes. Permanent alterations were observed in both sexes, but only those seen in the female will be further discussed.

Humans

Primordial germ cells migrate to the genital ridge to populate the primitive gonad around gestation week 5. During weeks 5-6 of embryonic development, the undifferentiated genital tract is comprised of two set of ducts (Wolffian and Mullerian), which are located in the urogenital ridge. By week 7, as the indifferent gonad starts to differentiate into either the testis or ovary, the duct systems responding to gonadal hormones start their differentiation processes. In the female, at approximately 8 weeks, the mesonephric ducts begin to regress cranially to caudally, leaving only vestiges such as in the rete ovarii, broad ligament and cervical area (Gartner's duct), which may give rise to cysts later in life. In contrast, the Mullerian duct differentiates and grows caudally, lateral to the mesonephric duct, fusing caudally as it grows and ending blindly in the urogenital sinus (UGS). The upper portion of the Mullerian ducts, which remains separated and unfused, differentiates into oviducts, whereas the caudally fused Mullerian tube starts differentiation into the uterus by approximately week 9 and into the upper vagina by week 10. ERs have been localized in the primitive uterus as early as gestation week 13. Although major reproductive tract organogenesis is complete by the start of the second trimester, tissue and cell differentiation continue into the second trimester. Birth occurs at approximately38 weeks in gestation.

Mice

Similar differentiation events occur in the mouse reproductive tract; however, analogous to developmental events in the mammary gland, variations occur in timing between humans and mice. Primordial germ cells migrate to the genital ridge during prenatal life starting on approximately gestational day 9. By 12 d.p.c, the undifferentiated genital tract is formed and is composed of the mesonephric and Mullerian duct systems. Regression of the mesonephric ducts and differentiation of the Mullerian ducts start on approximately late 12 d.p.c. and is complete by 16 d.p.c. Tissue and cell differentiation continue into neonatal life.

Similarities/differences

Although developmental milestones are similar, cell differentiation in various regions of the reproductive tract differ between mice and humans. An ovarian bursa is characteristic of mice that aids in guiding fertilized eggs into the oviduct in a mature female, but this structure is not present in women. Oviductal coiling is seen in mice but not in humans, and the utero-tubal junction varies between the species. Further, the cranial Mullerian ducts fuse in humans to form uterus, cervix and upper vagina; however, in mice, fusion occurs further caudally in the cervical region resulting in two separated uterine horns. Although controversy exists over the origin of the vaginal epithelium, it is generally considered that UGS epithelium grows cranially replacing Mullerian epithelium, but the replacement is more complete in humans than in mice. Vaginal adenosis and adenocarcinoma are thought to result from DES interfering with the UGS replacement of Mullerian epithelium, resulting in pockets of Mullerianderived columnar epithelial cells rather than squamousderived epithelium from UGS origin. At the molecular level, DES interferes with developmental genes that regulate cellular and tissue patterning.⁵⁷

Considering the differences in the reproductive tract development between humans and mice, it is apparent that the mouse is well suited for multiple pups per pregnancy with an ovarian bursa and two uterine horns, whereas the human is ideally suited for a single embryo with a unilateral uterine fundus. However, similar to breast tissues, the reproductive tract in humans and mice, at all stages of differentiation and development, has the ability to respond to estrogens such as DES.

DES and latent effects: adult disease/dysfunction following developmental exposure

Exposure to DES has been reported to cause numerous structural malformations in the oviduct, uterus and vagina of both humans and mice. Further, retained mesonephric remnants and paraovarian cysts of mesonephric origin have also been described in both species. Structural malformations in the mammary gland of mice characterized by altered TEBs and abnormal branching have been reported. In addition to malformations, which are teratogenic changes, dysfunction of the reproductive tract resulting in subfertility/infertility, poor reproductive outcome, altered menstrual (estrus) cycles and dysfunction of the mammary gland including altered lactation have all been described (see Table 1 for a summary). Some of these alterations such as oviductal malformations and altered mammary gland structure are apparent early in life, whereas many other alterations such as cancer require a much longer time to manifest.

Breast cancer

Administration of DES causes an increased incidence of breast cancer in prenatally DES-exposed daughters^{4,5,58} and

their mothers.^{7,59} Women >40 years of age and exposed *in utero* to DES have an estimated 1.9 times increased risk for developing breast cancer compared with unexposed women at the same age. In addition, the highest risks of breast cancer were correlated with the highest cumulative doses of DES during gestation.⁵

Many studies in rodents show alterations in mammary gland development following perinatal DES exposure. Pregnant rats were treated with DES and their offspring were examined after birth. Although excessive nipple development was observed in male and female offspring, prenatally DESexposed females were subsequently unable to nurse their offspring after they became pregnant and gave birth; this shows that prenatal DES exposure interferes with the later function of the mammary gland. This was likely because of the absence of the nipple sheath indicating failure to form a connection between the mammary ducts and the nipple, rendering the female unable to nurse.⁶⁰ ACI rats demonstrated elongated nipples, extensive lobuloalveolar proliferation, decreased tumor latency and greater multiplicity of tumors following gestational or lactational exposure to DES.44 Prenatal or neonatal DES exposure has also been shown to increase mammary tumorigenesis in Syrian golden hamsters and rats treated with DMBA. DES increased the number of mammary tumors, tumor multiplicity and the grade of tumor malignancy, suggesting that in addition to being carcinogenic, it also increased the sensitivity of the mammary gland to other carcinogens (reviewed in NIH⁷ and Fenton and Newbold⁷²).

Similar effects were seen in mice dosed just after birth with DES. Increased ductal outgrowth was seen in the mammary gland around the time of puberty, dilated mammary ducts were evident at 12 weeks and morphological changes in mice treated with low doses were accompanied by precocious lactation, suggesting that DES altered structure and function of the mammary gland. The association of increased incidence of mammary cancer in mice exposed either prenatally or neonatally to DES, similar to the increased incidence of cancer reported in humans, has been discussed earlier and is reviewed (see, Fenton and Newbold⁷²).

Altered reproduction and menstrual cycle irregularities

DES also causes alterations in the menstrual cycle in prenatally DES-exposed women such as shortened or prolonged length of cycles, abnormal bleeding between cycles, lack of ovulation, absence of menstruation, subfertility and/or infertility.⁷ Similar effects have been seen including abnormal estrus cycles and subfertility/infertility in DES-exposed mice.⁶¹ As menstrual cycle irregularities are not apparent until reproductive maturity, yet are caused by developmental exposure, this abnormality provides another example of DOHaD. Further, DES is also associated with multiple other health problems as it disrupts the endocrine system, which is responsible for communicating cellular signals throughout the body.

Fibroids (Leiomyomas)

Uterine leiomyoma, commonly called fibroids, are tumors of smooth muscle origin, which cause pain, bleeding, infertility and pregnancy complications; they are the leading indication of hysterectomy. Fibroids are the most common type of tumor in women over 30 years of age, and incidence rates have been estimated as high as 77% of women of reproductive age who are afflicted with the disease. In mice, early life exposure to DES causes fibroid development in adult animals⁶² and is another example of DOHaD. Data from the DES mouse model showed that fibroids occurred in approximately 9% of the DES-exposed mice as compared with <1% of unexposed animals. Interestingly, the association of fibroids with prenatal DES treatment in humans has also been shown⁶³. Among white women, 76% who reported prenatal DES exposure had fibroids compared with 52% of unexposed women. Further, DES-exposed women tended to have larger tumors. Thus, in this study, the prenatal DES mouse model predicted fibroid occurrence in DES-exposed women. The data with both DES-exposed mice and humans indicate a role for prenatal estrogen exposure in the etiology of uterine fibroids.

Another animal model is relevant to developmental estrogen exposure and fibroid development. Studies using the Eker rat that contains a defective tumor suppressor gene have shown increased susceptibility to uterine fibroids following early life exposure to DES.⁶⁴ The defective gene resulted in reprogramming of the myometrium leading to an increase in the expression of estrogen-responsive genes. Later in life, these neonatal DES-exposed rats have increased tumor suppressor gene penetrance that correlates with increased tumor size and multiplicity. This effect of DES in the Eker rat is an example of gene–environment interactions during development and provides yet another example of DOHaD.

Together, these data suggest that uterine fibroids observed in women of reproductive age may originate during development and early life, and that altered epigenetic programming in the uterus, due to exposure to environmental estrogens, may play a role in fibroid etiology.

Mechanisms involved in the adverse effects of developmental exposure

ER

Estrogens are linked to the development of breast cancer based on clinical and experimental data; risk factors for breast cancer reflect cumulative exposure of breast epithelium to estrogen.⁶⁵ Two hypotheses involving ER have been proposed to explain this relationship: (1) estrogen binding to ER stimulates proliferation of mammary cells, increasing the target cell number within the tissue, and the increase in cell division and DNA synthesis elevates the risk for replication errors, which may result in the accumulation of detrimental mutations that disrupt normal cellular processes such as cell

proliferation, apoptosis or DNA repair; (2) estrogen metabolism leads to the production of genotoxic by-products that can directly damage DNA, resulting in mutations. Evidence has existed for years to suggest that both mechanisms play a role in breast cancer; however, with DES exposure, genetic mutation and/or damage seem to play less of a role than originally expected.⁶⁶ DES has been shown in some studies to be a non-mutagenic carcinogen, therefore epigenetic mechanisms provide an explanation for the ability of DES to cause cancer in a non-mutagenic manner.⁶⁷

Epigenetics

Starting in the 1990s, this mechanism was progressed to explain developmentally induced carcinogenesis and it has since been vigorously studied. Altered epigenetic programming explains how exposure to exogenous estrogen during prenatal life can result in breast cancer some 40+ years later. The explanation lies at the center of the DOHaD paradigm. Although the link of epigenetics and cancer has received current increasing interest, the field of epigenetics is not new; it was first described in 1939 by Conrad Waddington. Epigenetics, meaning in addition to or above the genome, is defined as the study of heritable changes in gene function that occur without a change in the DNA sequence (there is no gene mutation involved, only a change in how the gene is regulated or expressed). Epigenetic programming is essential to regulate normal development and the maintenance of cell/ tissue differentiation by dictating cell-fate decisions via regulation of specific genes. Among these developmental genes are the Wnt and Hox family members. Thus, disruption in the balance of epigenetic networks can result in severe pathologies including altered structural development of the breast and reproductive tract, altered protein expression and even cancer later in life.

Environmental chemicals, especially those with estrogenic activity, have been documented in experimental animals to alter epigenetic programming that normally occurs during embryogenesis and development, and result in dysfunction/ disease later in life.^{14,19,20,22,68-71} The epigenome is very vulnerable to dysregulation by environmental factors (such as estrogenic chemicals) during critical windows of differentiation because of the high degree of developmental plasticity (especially after fertilization, again at implantation and during organ development) and the high rate of DNA synthesis that is occurring. Most importantly, elaborate epigenetic programming such as DNA methylation patterning required for normal tissue development is established during prenatal life. Alterations in epigenetic events, including changes in DNA methylation, posttranslational modifications of histones involved in chromatin structure and noncoding RNA/ DNA, may be permanent and not readily apparent until later in life. Epigenetic reprogramming by early life exposure to estrogenic chemicals is complex and may involve multiple epigenetic and genetic pathways that combine together to initiate disease/dysfunction later in life. DES is a documented epigenetic toxicant and this mechanism likely plays a role in its DOHaD effects.

Summary and conclusions

It has been over 50 years since DES was first prescribed for use during pregnancy, therefore it is very difficult to get solid information on DES-exposed humans because of lost and incomplete medical records. Important data including DES dose and timing are simply not available. Loss of data is a disadvantage for any DOHaD study involving humans because of the lag time between exposure and adverse effects, an thus animal models with their reduced life spans can play significant roles in studying and defining DOHaD events. The importance of animal models to inform human health and vice versa should not be underestimated. Is there a lesson to be learned from the DES scenario? Unfortunately, DOHaD phenomena are not lessons of past exposures but continue to be of concern as estrogens, progesterone and glucocorticoids are still being prescribed during sensitive prenatal and neonatal time periods, thus potentially sensitizing children to a variety of changes later in life.

In summary, ample scientific evidence from experimental animal and humans documents that prenatal DES exposure is an excellent example of DOHaD as it causes an increased risk for breast cancer and reproductive problems including menstrual irregularities and uterine fibroids, in addition to the originally described vaginal adenocarcinoma. Continued follow-up of the DES-exposed population (mothers, daughters and sons) such as the ongoing NCI studies are essential to further determine disease risks as the cohorts age. Regarding mechanisms, current data point to a role for epigenetic mechanisms involving reprogramming of developmental genes during prenatal life, which subsequently leads to altered responses (at the cell, tissue, organ level) to hormones or other environmental factors later in life. Epigenetic reprogramming suggests that DES effects may be passed on to subsequent generations, thus follow-up of DES grandchildren is also crucial. Finally, information gained from research with DES will help predict and prevent future adverse DOHaD events with other environmental chemicals and stressors.

Acknowledgment

The author thanks Dr Jerry Heindel for his critical reading and comments on this paper.

References

- Giusti RM, Iwamoto K, Hatch EE. Diethylstilbestrol revisited: a review of the long-term health effects. *Ann Intern Med.* 1995; 122, 778–788.
- 2. Herbst AL, Ulfelder H, Poskanzer DC. Adenocarcinoma of the vagina: association of maternal stilbestrol therapy with tumor

appearance in young women. N Engl J Med. 1971; 284, 878–879.

- Hatch EE, Palmer JR, Titus-Ernstoff L, Noller KL, Kaufman RH, Mittendorf R, *et al.* Cancer risk in women exposed to diethylstilbestrol in utero. *JAMA*. 1998; 280, 630–634.
- Palmer JR, Hatch EE, Rosenberg CL, Hartge P, Kaufman RH, Titus-Ernstoff L, *et al.* Risk of breast cancer in women exposed to diethylstilbestrol in utero: preliminary results (United States). *Cancer Causes Control.* 2002; 13, 753–758.
- Palmer JR, et al. Prenatal diethylstilbestrol exposure and risk of breast cancer. Cancer Epidemiol Biomarkers Prev. 2006; 15, 1509–1514.
- Troisi R, Potischman N, Hoover RN. Exploring the underlying hormonal mechanisms of prenatal risk factors for breast cancer: a review and commentary. *Cancer Epidemiol Biomarkers Prev*. 2007; 16, 1700–1712.
- National Institute of Health (NIH). DES Research Update, NIH Publication No: 00-4722. 1999, Bethesda, MD.
- Herbst AL, Bern HA. Developmental Effects of Diethylstilbestrol (DES) in Pregnancy. 1981; pp. 1–193. Thieme-Stratton, Inc., New York.
- Center for Disease Control and Prevention (CDC). Department of Health and Human Services Center for Disease Control and Prevention: DES Update, 2003, http://www.cdc.gov/DES/
- Diamanti-Kandarakis E, *et al.* Endocrine-disrupting chemicals: an Endocrine Society scientific statement. *Endocr Rev.* 2009; 30, 293–342.
- Newbold RR. Cellular and molecular effects of developmental exposure to diethylstilbestrol: implications for other environmnetal estrogens. *Environ Health Perspect.* 1995; 103, 83–87.
- 12. Newbold RR. Lessons learned from perinatal exposure to diethylstilbestrol. *Toxicol Appl Pharmacol.* 2004; 199, 142–150.
- Bern H. The fragile fetus. In *Chemically-induced Alterations in Sexual and Functional Development: the Wildlife Connection*, Vol. XXI (eds. Colborn T, Clement C), 1992; pp. 9–16. Princeton Scientific Publishing Company, Princeton, NJ.
- Newbold R, Kinyamu H. Epigenetic reproductive toxicants. In *Reproductive Toxicology* (eds. Kapp RW, Tyl R), 2010; pp. 317–331. Informa Healthcare, New York.
- Gluckman P, Hanson M, Pinal C. The developmental origins of disease. *Matern Child Nutr.* 2005; 1, 130–141.
- Colborn T, vom Saal FS, Soto AM. Developmental effects of endocrine-disrupting chemicals in wildlife and humans. *Environ Health Perspect*. 1993; 101, 378–384.
- Colborn T, Dumanoski D, Myers JP. Our Stolen Future, 1996. Penguin Books USA Inc., New York, pp. 1–306.
- Hatch EE, Herbst A, Hoover R, Noller K, Adam E, Kaufman R, et al. Incidence of squamous neoplasia of the cervix and vagina in DES-exposed daughters. Ann Epidemiol. 2000; 10, 467.
- Li S, *et al.* Developmental exposure to diethylstilbestrol elicits demethylation of estrogen-responsive lactoferrin gene in mouse uterus. *Cancer Res.* 1997; 57, 4356–4359.
- Li S, *et al.* Neonatal diethylstilbestrol exposure induces persistent elevation of c-fos expression and hypomethylation in its exon-4 in mouse uterus. *Mol Carcinog.* 2003; 38, 78–84.
- Newbold RR, *et al.* Developmental exposure to diethylstilbestrol alters uterine gene expression that may be associated with uterine neoplasia later in life. *Mol Carcinog.* 2007; 46, 783–796.

- Tang WY, *et al.* Persistent hypomethylation in the promoter of nucleosomal binding protein 1 (Nsbp1) correlates with overexpression of Nsbp1 in mouse uteri neonatally exposed to diethylstilbestrol or genistein. *Endocrinology*. 2008; 149, 5922–5931.
- Newbold RR, *et al.* Developmental exposure to diethylstilbestrol (DES) alters uterine response to estrogens in prepubescent mice: low versus high dose effects. *Reprod Toxicol.* 2004; 18, 399–406.
- 24. Melnick R, Lucier G, Wolfe M, Hall R, Stancel G, Prins G, *et al.* Summary of the National Toxicology Program's report of the endocrine disruptors low-dose peer review. *Environ Health Perspect.* 2002; 110, 427–431.
- 25. vom Saal FS, Akingbemi BT, Belcher SM, Birnbaum LS, Crain DA, Eriksen M, *et al.* Chapel Hill bisphenol A expert panel consensus statement: integration of mechanisms, effects in animals and potential to impact human health at current levels of exposure. *Reprod Toxicol.* 2007; 24, 131–138.
- 26 Heindel JJ, Newbold RR, (eds.). Developmental origins of health and disease: the importance of environmental exposures. In *Early Life Origins of Human Health and Disease*, 2009; pp. 42–51. Basel, Switzerland, Karger.
- Soto AM, Sonnenschein C. Environmental causes of cancer: endocrine disruptors as carcinogens. *Nat Rev Endocrinol.* 2010; 6, 363–370.
- 28 Hogan MD, Newbold RR, McLachlan JA. Extrapolation of teratogenic responses observed in laboratory animals to humans: DES as an illustative example. In *Developmental Toxicology: Mechanisms and Risk* (eds. McLachlan JA, Pratt RM, Market C), 1987, pp. 257–269. Cold Spring Harbor Laboratory, Cold Spring Harbor: MA, USA.
- Ekbom A, Trichopoulos D, Adami HO, Hsieh CC, Lan SJ. Evidence of prenatal influences on breast cancer risk. *Lancet*. 1992; 340, 1015–1018.
- 30. Russo J, et al. Comparative study of human and rat mammary tumorigenesis. Lab Invest. 1990; 62, 244–278.
- Russo J, Gusterson BA, Rogers AE, Russo IH, Wellings SR, van Zwieten MJ. Developmental, cellular, and molecular basis of human breast cancer. *J Natl Cancer Inst Monogr.* 2000; 27, 17–37.
- Rosso J, Hu YF, Yang X, Russo IH. Anatomy and physiologic morphology. In *Rosen's Breast Pathology*, (ed. Rosen P), 2008, pp. 1–22. Lippincott, Williams, and Wilkins, New York.
- 33. Friedrichs N, Steiner S, Buettner R, Knoepfle G. Immunohistochemical expression patterns of AP2alpha and AP2gamma in the developing fetal human breast. *Histopathology*. 2007; 51, 814–823.
- Robinson GW, Karpf AB, Kratochwil K. Regulation of mammary gland development by tissue interaction. J Mammary Gland Biol Neoplasia. 1999; 4, 9–19.
- Vandenberg LN, Maffini MV, Wadia PR, Sonnenschein C, Rubin BS, Soto AM. Exposure to environmentally relevant doses of the xenoestrogen bisphenol-A alters development of the fetal mouse mammary gland. *Endocrinology*. 2007; 148, 116–127.
- Lemmen JG, Broekhof JL, Kuiper GG, Gustafsson JA, van der Saag PT, van der Burg B. Expression of estrogen receptor alpha and beta during mouse embryogenesis. *Mech Dev.* 1999; 81, 163–167.
- Narbaitz R, Stumpf WE, Sar M. Estrogen receptors in mammary gland primordia of fetal mouse. *Anat Embryol (Berl)*. 1980; 158, 161–166.

- Saji S, Jensen EV, Nilsson S, Rylander T, Warner M, Gustafsson JA. Estrogen receptors alpha and beta in the rodent mammary gland. *Proc Natl Acad Sci U S A*. 2000; 97, 337–342.
- Richert MM, Schwertfeger KL, Ryder JW, Anderson SM. An atlas of mouse mammary gland development. J Mammary Gland Biol Neoplasia. 2000; 5, 227–241.
- Rustia M, Shubik P. Effects of transplacental exposure to diethylstilbestrol on carcinogenic susceptibility during postnatal life in hamster progeny. *Cancer Res.* 1979; 39, 4636–4644.
- Boylan ES, Calhoon RE. Mammary tumorigenesis in the rat following prenatal exposure to diethylstilbestrol and postnatal treatment with 7,12-dimethylbenz[a]anthracene. J Toxicol Environ Health. 1979; 5, 1059–1071.
- Boylan ES, Calhoon RE. Prenatal exposure to diethylstilbestrol: ovarian-independent growth of mammary tumors induced by 7,12-dimethylbenz[a]anthracene. J Natl Cancer Inst. 1981; 66, 649–652.
- Boylan ES, Calhoon RE. Transplacental action of diethylstilbestrol on mammary carcinogenesis in female rats given one or two doses of 7,12-dimethylbenz(a)anthracene. *Cancer Res.* 1983; 43, 4879–4884.
- Rothschild TC, Boylan ES, Calhoon RE, Vonderhaar BK. Transplacental effects of diethylstilbestrol on mammary development and tumorigenesis in female ACI rats. *Cancer Res.* 1987; 47, 4508–4516.
- 45. Vassilacopoulou D, Boylan ES. Mammary gland morphology and responsiveness to regulatory molecules following prenatal exposure to diethylstilbestrol. *Teratog Carcinog Mutagen*. 1993; 13, 59–74.
- Walker BE. Tumors in female offspring of control and diethylstilbestrol-exposed mice fed high-fat diets. J Natl Cancer Inst. 1990; 82, 50–54.
- Kawaguchi H, Miyoshi N, Miyamoto Y, Souda M, Umekita Y, Yasuda N, *et al.* Effects of fetal exposure to diethylstilbestrol on mammary tumorigenesis in rats. *J Vet Med Sci.* 2009; 71, 1599–1608.
- Nagasawa H, Mori T, Nakajima Y. Long-term effects of progesterone or diethylstilbestrol with or without estrogen after maturity on mammary tumorigenesis in mice. *Eur J Cancer*. 1980; 16, 1583–1589.
- 49. Bern HA, Jones LA, Mori T, Young PN. Exposure of neonatal mice to steroids: longterm effects on the mammary gland and other reproductive structures. *J Steroid Biochem.* 1975; 6, 673–676.
- Warner MR, Warner RL. Effects of exposure of neonatal mice to 17beta-estradiol on subsequent age-incidence and morphology of carcinogen-induced mammary dysplasia. *J Natl Cancer Inst.* 1975; 55, 289–298.
- Mori T, Bern HA, Mills KT, Young PN. Long-term effects of neonatal steroid exposure on mammary gland development and tumorigenesis in mice. *J Natl Cancer Inst.* 1976; 57, 1057–1062.
- 52. Mori T, Nagasawa H, Bern HA. Long-term effects of perinatal exposure to hormones on normal and neoplastic mammary growth in rodents: a review. *J Environ Pathol Toxicol.* 1979; 3, 191–205.
- 53. Hovey RC, Asai-Sato M, Warri A, Terry-Koroma B, Colyn N, Ginsburg E, *et al.* Effects of neonatal exposure to diethylstilbestrol, tamoxifen, and toremifene on the

BALB/c mouse mammary gland. *Biol Reprod.* 2005; 72, 423–435.

- 54. Ninomiya K, Kawaguchi H, Souda M, Taguchi S, Funato M, Umekita Y, *et al.* Effects of neonatally administered diethylstilbestrol on induction of mammary carcinomas induced by 7, 12-dimethylbenz(a)anthracene in female rats. *Toxicol Pathol.* 2007; 35, 813–818.
- 55. Umekita Y, Souda M, Hatanaka K, Hamada T, Yoshioka T, Kawaguchi H, *et al.* Gene expression profile of terminal end buds in rat mammary glands exposed to diethylstilbestrol in neonatal period. *Toxicol Lett.* 2011; 205, 15–25.
- Tuchmann-Duplessis H, Haegel P, (eds.). *Illustrated Human Embrology: Organogenesis*, Vol. 11, 1982. Spring Verlag, New York.
- 57. Taylor HS, Vanden Heuvel GB, Igarashi P. A conserved Hox axis in the mouse and human female reproductive system: late establishment and persistent adult expression of the Hoxa cluster genes. *Biol Reprod.* 1997; 57, 1338–1345.
- Troisi R, Hatch EE, Titus-Ernstoff L, Hyer M, Palmer JR, Robboy SJ, et al. Cancer risk in women prenatally exposed to diethylstilbestrol. Int J Cancer. 2007; 121, 356–360.
- 59. Titus-Ernstoff L, Troisi R, Hatch EE, Palmer JR, Wise LA, Ricker W, *et al.* Mortality in women given diethylstilbestrol during pregnancy. *Br J Cancer.* 2006; 95, 107–111.
- Boylan ES. Morphological and functional consequences of prenatal exposure to diethylstilbestrol in the rat. *Biol Reprod.* 1978; 19, 854–863.
- 61. McLachlan JA, Newbold RR, Shah HC, Hogan M, Dixon RL. Reduced fertility in female mice exposed transplacentally to diethylstilbestrol (DES). *Fertil Steril.* 1982; 38, 364–371.
- 62. McLachlan JA, Newbold RR, Bullock BC. Long-term effects on the female mouse genital tract associated with prenatal exposure to diethylstilbestrol. *Cancer Res.* 1980; 40, 3988–3999.

- 63. Baird DD, Newbold RR. Prenatal diethylstilbestrol (DES) exposure is associated with uterine leiomyoma development. *Reprod Toxicol.* 2005; 20, 81–84.
- Cook JD, Davis BJ, Goewey JA, Berry TD, Walker CL. Identification of a sensitive period for developmental programming that increases risk for uterine leiomyoma in Eker rats. *Reprod Sci.* 2007; 14, 121–136.
- 65. Henderson BE, Feigelson HS. Hormonal carcinogenesis. Carcinogenesis. 2000; 21, 427–433.
- 66. Larson PS, Ungarelli RA, de lasMorenas A, Cupples LA, Rowlings K, Palmer JR, *et al.* In utero exposure to diethylstilbestrol (DES) does not increase genomic instability in normal or neoplastic breast epithelium. *Cancer.* 2006; 107, 2122–2126.
- 67. Szyf M. The dynamic epigenome and its implications in toxicology. *Toxicol Sci.* 2007; 100, 7–23.
- 68. Ho SM, Tang WY. Techniques used in studies of epigenome dysregulation due to aberrant DNA methylation: an emphasis on fetal-based adult diseases. *Reprod Toxicol.* 2007; 23, 267–282.
- Prins GS, Birch L, Tang WY, Ho SM. Developmental estrogen exposures predispose to prostate carcinogenesis with aging. *Reprod Toxicol.* 2007; 23, 374–382.
- 70. Prins GS, Tang WY, Belmonte J, Ho SM. Developmental exposure to bisphenol A increases prostate cancer susceptibility in adult rats: epigenetic mode of action is implicated. *Fertil Steril.* 2008; 89, e41.
- Prins GS, Tang WY, Belmonte J, Ho SM. Perinatal exposure to oestradiol and bisphenol A alters the prostate epigenome and increases susceptibility to carcinogenesis. *Basic Clin Pharmacol Toxicol*. 2008; 102, 134–138.
- 72. Fenton SE, Reed C, Newbold RR. Perinatal environmental exposures affect mammary development, function, and cancer risk in adulthood. *Annu Rev Pharmacol Toxicol.* 2012; 52, 455–479.