

Molecular phylogenetic analysis of the genus *Gyrodactylus* (Platyhelminthes: Monogenea) inferred from rDNA ITS region: subgenera versus species groups

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SUMMARY

Analyses of small subunit ribosomal RNA gene sequences of representatives of major taxa of Monopisthocotylea were performed to identify the sister group of *Gyrodactylus*. Nuclear ribosomal DNA sequences from the complete internal transcribed spacer (ITS) region were used to infer phylogeny of 37 *Gyrodactylus* species and *Gyrodactyloides bychowskii*, *Macrogyrodactylus polypteri* and *Gyrdicotylus gallieni*, using maximum likelihood, parsimony and Bayesian inference. The genus *Gyrodactylus* appeared to be a monophyletic group in all analyses, based on the present data set. Within the genus, there were 3 major groups recognized by high bootstrap values and posterior probabilities. None of the 6 subgenera appeared to be monophyletic, and the most basal subgenus *G.* (*Gyrodactylus*) was paraphyletic. Characteristics of the excretory system of *Gyrodactylus* do not seem to be conservative enough to reveal subgenera within *Gyrodactylus* and we suggest abandoning existing subgenera as indicators of phylogeny. The grouping of species based on the morphology of the ventral bar and marginal hooks seems to have sufficient power to infer relationships between the *Gyrodactylus* species.

Key words: Monogenea, *Gyrodactylus*, SSU, ITS, phylogeny.

INTRODUCTION

Members of the family Gyrodactylidae (Monogenea: Monopisthocotylea) are flatworm parasites with a large range of host organisms, predominantly teleost fish (William & Jones, 1994). Within the Gyrodactylidae, *Gyrodactylus* is the most diverse and widespread genus.

Based on the morphology of the excretory system, 6 *Gyrodactylus* subgenera were proposed by Malmberg (1956, 1964); *G.* (*Gyrodactylus*), *G.* (*Mesonephrotus*), *G.* (*Metanephrotus*), *G.* (*Paranephrotus*), *G.* (*Neonephrotus*), and *G.* (*Limnonephrotus*). Difficulties in the use of excretory system characters for systematic studies have arisen because the components of the protonephridial system are only clearly visible in live parasites, rendering fixed specimens all but

useless for differentiating subgenera. For this reason, not all described *Gyrodactylus* species may be readily assigned to subgenera. The shape of the marginal hook was proven to be another character suitable to group *Gyrodactylus* species. Using this character, groups such as the *G. elegans*- or *G. wagneri*-group were established (see Malmberg, 1964, 1970 for more details) including other species of similar marginal hook morphology.

Due to advances in molecular biology, genetic markers for species identification of gyrodactylids have been investigated, based mainly on the ribosomal RNA (rRNA) genes and the associated internal transcribed spacers (ITS) (Cunningham *et al.* 1995a; Cunningham, 1997). The ITS sequences from approximately 55 described *Gyrodactylus* species are known (Cunningham, 1997; Cable *et al.* 1999; Zietara *et al.* 2000; Cunningham *et al.* 2001; Matejusová *et al.* 2001; Huysse & Volckaert, 2002; Zietara & Lumme, 2002) with other sequences obtained from as-yet unidentified species (Matejusová & Cunningham, unpublished data; Zietara & Lumme, 2002).

Although members of the genus *Gyrodactylus* show high species diversity, with 402 valid species descriptions (Bakke, Harris & Cable, 2002), there

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¶ Nucleotide sequence data reported in this paper are available in the GenBank, DDBJ and EMBL databases under the accession numbers AJ566375–79, AJ566768, AJ567670–74, AJ581657.

Table 1. Monopisthocotylean parasite sequences used for ML analysis of the SSU rRNA gene for outgroup comparison (Olson & Littlewood, 2002)

Species	Family	GenBank/EMBL
<i>Anoplodiscus cirrusspiralis</i>	Anoplodiscidae	AJ287475
<i>Benedenia</i> n. sp.	Capsalidae	AJ228774
<i>Capsala martinieri</i>	Capsalidae	AJ276423
<i>Encotyllabe chironemi</i>	Capsalidae	AJ228780
<i>Pseudodactylogyrus</i> sp.	Dactylogyridae	AJ287567
<i>Pseudohaliotrema sphincteroporos</i>	Dactylogyridae	AJ287568
<i>Gyrodactylus carassii</i>	Gyrodactylidae	AJ566377§
<i>Gyrodactylus sedelnikowi</i>	Gyrodactylidae	AJ566378§
<i>Gyrodactylus gobiensis</i>	Gyrodactylidae	AJ566375§
<i>Gyrodactylus rhodei</i>	Gyrodactylidae	AJ567670§
<i>Gyrodactylus rutilensis</i>	Gyrodactylidae	AJ566376§
<i>Gyrodactylus salaris</i>	Gyrodactylidae	Z26942
<i>Gyrodactyloides bychowskii</i>	Gyrodactylidae	AJ566379§
<i>Macrogyrodactylus polypteri</i>	Gyrodactylidae	AJ567671§
<i>Leptocotyle minor</i>	Microbothriidae	AJ228784
<i>Calicotyle affinis</i>	Monocotylidae	AJ228777
<i>Dictyocotyle coeliaca</i>	Monocotylidae	AJ228778
<i>Troglocephalus rhinobatidis</i>	Monocotylidae	AJ228795
<i>Pseudomurraytrema (alabarrum?)</i>	Pseudomurraytrematidae	AJ228793
<i>Sundanonchus micropeltis</i>	Sundanonchidae	AJ287588
<i>Udonella caligorum</i>	Udonellidae	AJ228796

§ Sequences obtained in the present study.

have been no analyses that included exemplar species of all defined subgenera in order to display their phylogenetic relationship or to test their proposed monophyly. A phylogenetic analysis inferred from the combined 5.8S and ITS2 sequences of 10 gyrodactylids has given us partial information about the relationships among 3 subgenera and demonstrated a separation of *G. (Limnonephrotus)* from *G. (Mesonephrotus)* and *G. (Metanephrotus)* (Cable *et al.* 1999). Also, certain species belonging to the subgenus *G. (Gyrodactylus)* were suggested to be distant from species of the subgenus *G. (Limnonephrotus)* based on differences in the ITS1 and the morphology of haptor structures, in particular the marginal hooks and ventral bar (Matejusová *et al.* 2001). Subsequently, using 10 *Gyrodactylus* species belonging to 4 subgenera, maximum likelihood analysis inferred from the 5.8S sequences presented a monophyletic origin of *Gyrodactylus* with *G. (Mesonephrotus)* and *G. (Metanephrotus)* being a sister group to *G. (Paranephrotus)* and *G. (Limnonephrotus)* species (Zietara *et al.* 2002). The same authors also demonstrated that each of the subgenera possessed a unique 5.8S gene sequence.

Here we present phylogenetic analyses inferred from the ITS sequences of members of all *Gyrodactylus* subgenera to elucidate their relationships. Prior to this analysis, sequences of the small subunit (SSU) ribosomal RNA (rRNA) gene were used to identify a sister group of *Gyrodactylus*. SSU rRNA has been used successfully for determining the interrelationships of monopisthocotylean monogeneans and a substantial database is now available (e.g. Olson & Littlewood, 2002).

MATERIALS AND METHODS

Sequence alignment

The small subunit (SSU) ribosomal RNA gene of 6 *Gyrodactylus* species, *Gyrodactyloides bychowskii* and *Macrogyrodactylus polypteri* were sequenced. DNA extraction and PCR was carried out according to Matejusová *et al.* (2001), the primers used to amplify the SSU region were as described by Cunningham *et al.* (1995b).

SSU sequences together with sequences of the complete internal transcribed spacers (ITS1 and ITS2) and 5.8S ribosomal DNA of 37 *Gyrodactylus* species, *G. bychowskii*, *Gyrdicotylus gallieni* Vercaemmen Grandjean, 1960 (AJ001843) and *M. polypteri* were aligned in CLUSTAL X (Jeanmougin *et al.* 1998), using default parameters. The full list of taxa used in the SSU and ITS rDNA alignments are shown in Tables 1 and 2 respectively, with GenBank accession numbers and specimen details for new sequences. The extraction, PCR, and sequencing methods for the ITS region of newly sequenced species were as described by Matejusová *et al.* (2001). Alignments were refined by eye using MacClade v. 4.05 (Maddison & Maddison, 2000). Regions of ambiguity were recorded and approximately 400 bp from the 5' end of ITS1 were removed prior to analysis. Although the majority of positions were alignable among all taxa, it was difficult to satisfactorily align some ITS positions. All analyses were carried out using only positions that were unambiguously alignable across all taxa.

New SSU sequences were aligned to the monopisthocotylean portion of an existing published

Table 2. Species used for phylogenetic analyses of the ITS

Genus	Species	GenBank/EMBL	Subgenus (Ref.)	Species group (Ref.)	
<i>Gyrodactylus</i>	<i>G. anguillae</i>	AB063293	<i>G. (Neo.)</i> (a)	<i>G. anguillae</i> -group (a)	
	<i>G. aphyae</i>	AJ407865 AJ407915	<i>G. (Limno.)</i> (a)	<i>G. wagneri</i> -group (a)	
	<i>G. arcuatus</i>	AJ001839	<i>G. (Meso.)</i> (a)	<i>G. arcuatus</i> -group (a)	
	<i>G. barbi</i>	AJ407866 AJ407916		<i>G. wagneri</i> -group (b)	
	<i>G. branchicus</i>	AF156669	<i>G. (Meta.)</i> (a)	<i>G. rarus</i> -group (a)	
	<i>G. bullatarudis</i>	AJ011410	<i>G. (Meso.)</i> (a)	<i>G. arcuatus</i> -group (a)	
	<i>G. carassii</i>	AJ407868 AJ407918	<i>G. (Gyro.)</i> (a)	<i>G. phoxini</i> - (a) or <i>G. elegans</i> -group (c)	
	<i>G. derjavini</i>	AJ132259	<i>G. (Limno.)</i> (d)		
	<i>G. elegans</i>	AJ407870 AJ407920	<i>G. (Gyro.)</i> (a)	<i>G. elegans</i> -group (a)	
	<i>G. fossilis</i>	AJ407871 AJ407921		<i>G. wagneri</i> -group*	
	<i>G. gobiensis</i>	AJ407872 AJ566768§		<i>G. wagneri</i> -group (e)	
	<i>G. gobii</i>	AJ407873 AJ407922		<i>G. wagneri</i> -group*	
	<i>G. harengi</i>	AJ309295	<i>G. (Meta.)</i> (a)	<i>G. harengi</i> -group (a)	
	<i>G. hronosus</i>	AJ407876 AJ407924	<i>G. (Limno.)</i> (a)	<i>G. wagneri</i> -group (a)	
	<i>G. jiroveci</i>	AJ567674§	<i>G. (Limno.)</i> ?(a), <i>G. (Para.)</i> ?(f)		
	<i>G. katharineri</i>	AJ407878 AJ407926	<i>G. (Limno.)</i> (a)	<i>G. katharineri</i> -group (a)	
	<i>G. lomi</i>	AJ407882 AJ407929		<i>G. wagneri</i> -group*	
	<i>G. luciopercae</i>	AJ407885 AJ407931	<i>G. (Limno.)</i> (a)	<i>G. wagneri</i> -group (a)	
	<i>G. macronychus</i>	AJ407893	<i>G. (Limno.)</i> (a)	<i>G. wagneri</i> -group (a)	
	<i>G. markakulensis</i>	AJ407886 AJ407932	<i>G. (Gyro.)</i> ?(a)	<i>G. elegans</i> - or <i>G. phoxini</i> -group (a)	
	<i>G. micropsi</i>	AF328868	<i>G. (Para.)</i> (g)	<i>G. rugiensis</i> -group (g)	
	<i>G. nipponensis</i>	AB063295		<i>G. anguillae</i> -group (h)	
	<i>G. poeciliae</i>	AJ001844	<i>G. (Meso.)</i> or <i>G. (Meta.)</i> ?(i)		
	<i>G. prostaе</i>	AJ567673§	<i>G. (Gyro.)</i> (a)	<i>G. elegans</i> -group (a)	
	<i>G. pungitii</i>	AJ001845	<i>G. (Limno.)</i> (a)	<i>G. wagneri</i> -group (a)	
	<i>G. pterygialis</i>	AJ581657§	<i>G. (Meso.)</i> (a)	<i>G. callariatis</i> -group (a)	
	<i>G. rhodei</i>	AJ407889 AJ407933		<i>G. rhodei</i> -group (j) or <i>G. wagneri</i> -group (k)	
	<i>G. rogatensis</i>	AJ011411	<i>G. (Limno.)</i> (l)		
	<i>G. rugiensis</i>	AF328870	<i>G. (Para.)</i> (g)	<i>G. rugiensis</i> -group (g)	
	<i>G. rugiensoides</i>	AJ427414		<i>G. rugiensis</i> -group (m)	
	<i>G. rutilensis</i>	AJ407890 AJ407934		<i>G. wagneri</i> -group (e)	
	<i>G. salaris</i>	Z72477	<i>G. (Limno.)</i> (a)	<i>G. wagneri</i> -group (a)	
	<i>G. sedelnikowii</i>	AJ407891 AJ407935	<i>G. (Gyro.)</i> (a)	<i>G. phoxini</i> -group (a)	
	<i>G. teuchis</i>	AJ249350		<i>G. wagneri</i> -group*	
	<i>G. truttae</i>	AJ132260	<i>G. (Limno.)</i> (n)	<i>G. wagneri</i> -group (e)	
	<i>G. turnbulli</i>	AJ001846	<i>G. (Meta.)</i> (d)	<i>G. eucaliae</i> -group	
	<i>G. vimbi</i>	AJ407892 AJ407936		<i>G. wagneri</i> -group (o)	
	<i>Gyrdicotylus</i>	<i>G. gallieni</i>	AJ001843		
	<i>Gyrodactyloides</i>	<i>G. bychowskii</i>	AJ249348		
	<i>Macrogyrodactylus</i>	<i>M. polypteri</i>	AJ567672§		

(a) Malmberg, 1970; (b) Ergens, 1976; (c) Ergens, 1966; (d) Cable *et al.* 1999; (e) Gläser, 1974a; (f) Ergens & Bychowsky, 1967; (g) Gläser, 1974b; (h) Ernst, Fletcher & Hayward, 2000; (i) Harris & Cable, 2000; (j) Ergens & Yurkimenko, 1975; (k) Zitnan, 1964; (l) Harris, 1985; (m) Huyse & Volckaert, 2002; (n) Zietara *et al.* 2002; (o) Ergens, 1980.

* Included in the *G. wagneri*-group as the shape of the marginal hook is very similar to species included in this group.

§ Sequences obtained in the present study.

alignment of the monogenean SSU (Olson & Littlewood, 2002; EBI accession ALIGN_000146; see Table 1 for list of taxa), using the profile alignment option. The SSU analyses were performed to identify the most basal taxon of *Gyrodactylus* that would be used to root final trees of *Gyrodactylus* spp. based on the complete ITS sequences.

The full alignments for the SSU and ITS data sets (21 and 40 species respectively) have been deposited with EBI and are available by anonymous FTP from ftp.ebi.ac.uk in directory /pub/databases/embl/align and via the EMBLALIGN database via SRS at http://srs.ebi.ac.uk, under the following accessions ALIGN_000604 (SSU) and ALIGN_000605 (ITS). Exclusion sets are added as notes and the alignments may be adapted as NEXUS files.

Phylogenetic analyses

We estimated phylogenies using maximum parsimony (MP), Bayesian inference (BI) and maximum likelihood (ML), rooting the ingroup against *Anoplodiscus cirrusspiralis* for the SSU data set. Following the SSU analyses, ITS phylogenies were rooted against *Gyrodactyloides*, *Macrogyrodactylus* and *Gyrdicotylus*.

MP and ML analyses were conducted with PAUP*4.0b10 (Swofford, 2002), employing a branch-and-bound search strategy for MP and a heuristic search strategy for ML. Modeltest v. 3.06 was used to select the model of evolution of best fit for each data partition (Posada & Crandall, 1998). For the SSU data and for each of the ITS1, 5.8S and ITS2 partitions individually, we employed a GTR+I+G model; this refers to a general-time-reversible model including estimates of invariant sites and gamma distributed among-site rate variation. BI was determined using MrBayes (Huelsenbeck & Ronquist, 2001, ver. 2.01) with the following parameters: nst = 6, rates = invgamma, ncat = 4, shape = estimate, inferrates = yes, and basefreq = empirical, that corresponds to a GTR+I+G substitution model. For the ITS data, each of the data partitions were treated independently, and each using an independently estimated GTR+I+G substitution model. Posterior probabilities were approximated over 1 000 000 generations (ngen = 1 000 000) via 4 simultaneous Markov Chain Monte Carlo chains (MCMC) (nchains = 4) with every 100th tree saved (samplefreq = 100). Default values were used for the MCMC parameters. Consensus trees with mean branch lengths were constructed using the 'sumt' command with the 'contype = allcompat' option and ignoring the initial topologies saved during 'burn in'; the initial *n*-generations before log-likelihood values and substitution parameters plateau (see Huelsenbeck & Ronquist, 2001). MP and ML nodal supports were estimated by bootstrap analyses (heuristic search, 1000 replicates for MP, 100

replicates for ML), and as posterior probabilities in the Bayesian inference analyses (Huelsenbeck *et al.* 2001).

RESULTS

SSU rDNA

The new SSU sequence length varied from 1892 bp (*G. rhodei*) to 1974 bp (*G. sedelnikowi*). The complete alignment spanned 2189 positions but only 1579 were included and considered unambiguously aligned. Of these, 1117 positions were constant and 323 informative under the principles of parsimony. Modeltest found that the most appropriate model of substitution was GTR+I+G and we used this for both ML and BI. For ML the following parameters were used: rate matrix, 0.9654 (A-C), 4.0010 (A-G), 2.3835 (A-T), 0.8694 (C-G), 5.4764 (C-T), 1.0000 (G-T); nucleotide frequencies A = 0.2713, C = 0.2069, G = 0.2612, T = 0.2606; assumed proportion of invariable sites = 0.4782; gamma shape parameter (alpha) = 0.6441; 4 rate categories. ML and BI analyses performed on the SSU data set resolved trees with an identical topology, and almost identical (relative) branch lengths. MP found 6 equally parsimonious trees (length = 1019; CI = 0.618; RI = 0.741; RC = 0.458) and the strict consensus was also fully compatible with the single tree topology inferred by ML and BI. The ML tree is shown in Fig. 1, with branch lengths estimated by ML and nodal support from ML (bootstrap, *n* = 100), MP (bootstrap, *n* = 1000) and BI (posterior probabilities).

The BI solution, a consensus of 2560 trees, further resolves *G. rhodei* + *G. rutilensis* and *G. gobiensis* + *G. salaris* (*G. (Limnonephrotus)* subgenus) as sister taxa of *G. carassii* + *G. sedelnikowi* (*G. (Gyrodactylus)* subgenus) with posterior probabilities of 100 in this sample. In addition, the genus *Gyrodactylus* appears monophyletic with *M. polypteri* as its sister group; *G. bychowskii* was resolved as the sister group to *Gyrodactylus* + *Macrogyrodactylus*.

ITS rDNA

MP analysis found 3 equally parsimonious trees (length = 1357; CI = 0.491; RC = 0.338). An incongruence length difference test (Farris *et al.* 1994), as implemented in PAUP*, suggested that the individual data partitions had evolved significantly differently from one another (*P* = 0.007) and were not combinable in a phylogenetic analysis with the same nucleotide substitution model. Modeltest found that the most appropriate model of substitution was GTR+I+G for each of the data partitions and when these partitions were combined. There was little difference in tree topology whether these partitions were modelled separately or combined. We

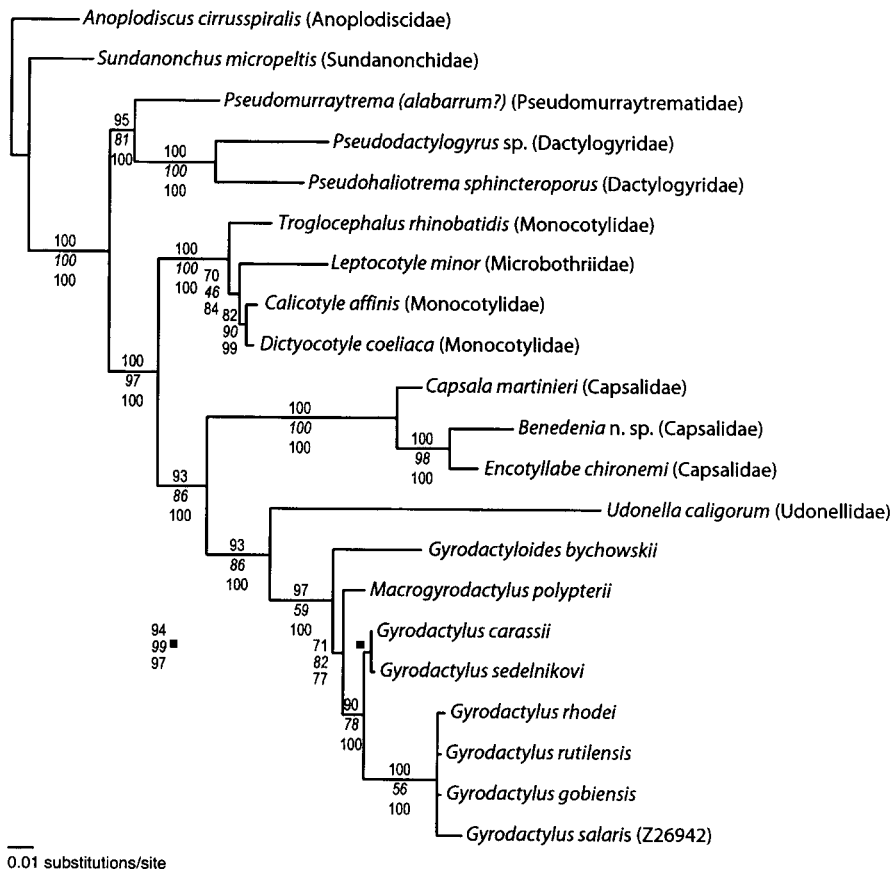


Fig. 1. Phylogeny of monopisthocotylean Monogenea based on SSU rDNA indicating the relative position of *Gyrodactylus* species and potential outgroup taxa. The tree topology is from a maximum likelihood analysis with nodal support indicated, from top to bottom, for maximum likelihood (bootstrap %, $n = 100$), maximum parsimony (bootstrap %, $n = 1000$) and Bayesian inference (posterior probabilities); see text for further details.

used the model for both ML and BI. For BI, where each of the partitions was modelled separately, we estimated that log likelihood values had reached a plateau at approximately 40 000 generations. We ignored results for a further 20 000 generations and summarized trees for the final 940 000 generations (9400 trees). Branch lengths were calculated as means of the branch lengths in the individual topologies saved during Bayesian analysis and summarized using the 'sumt' command of MrBayes.

All 3 analyses resolved the same broad patterns of evolution among the *Gyrodactylus* species. The only topological differences between the phylogenetic solutions were amongst poorly supported clades. We show only the solution derived from Bayesian analysis using the independent GTR+I+G estimates for each of the 3 combined data partitions, which was almost identical to that of BI and ML using a single model, with nodal support from BI and MP (Fig. 2) where posterior probabilities exceed 80% and MP bootstrap exceed 50%; it was computationally impossible to provide bootstrap support for the ML analysis using the most appropriate model of substitution.

At the base of the *Gyrodactylus* clade, *G. markakulensis* was consistently resolved as the most basal

taxon. Next, the remaining taxa within the subgenus *G.* (*Gyrodactylus*) were resolved to be strongly monophyletic, with each of the analyses resolving the same interrelationships as indicated in Fig. 2. The next clade to be resolved was a mixture of taxa in the subgenera *G.* (*Metanephrotus*) and *G.* (*Mesonephrotus*). BI and ML resolved identical topologies within this clade, but MP pulled *G. turnbulli* to the base. The third well-supported clade comprises the remaining taxa, in which some nodes were strong and others indicated poor resolution. BI and ML resolved almost identical topologies; differences concerned only the interrelationships of the most derived taxa where branch lengths were very short. However, within this group, MP resolved the *G. ruginensis*+*G. ruginensoides*, *G. anguillae*+*G. micropsi* clade as the most basal taxa, and *G. rutilensis* and *G. hronosus* as more derived and not as sister taxa; these differences in the MP analysis account for the low bootstrap proportions plotted at the nodes of the Bayesian tree (Fig. 2).

DISCUSSION

The present study analysed exemplar species of all defined subgenera of *Gyrodactylus* and brings more

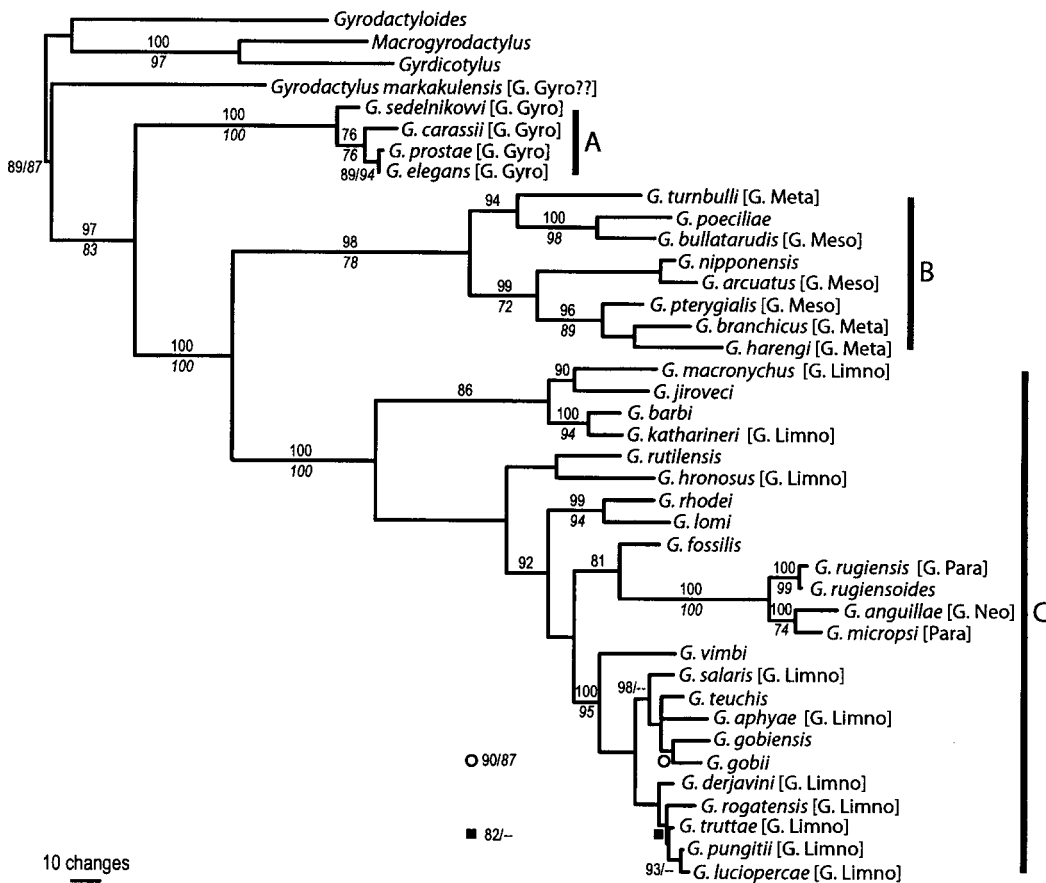


Fig. 2. Phylogeny of *Gyrodactylus* species based on ITS rDNA, rooted against *Gyrodactyloides bychowskii*. The tree topology is from a Bayesian analysis, modelling each data partition separately, with nodal support indicated, from top to bottom, for Bayesian inference (posterior probabilities) and maximum parsimony (bootstrap %, $n=1000$) where these values are $>80\%$ and 50% respectively. Subgenera, where known, are indicated in square brackets as: *G. Gyro* – *G. Gyrodactylus*; *G. Limno* – *G. Limnonephrotus*; *G. Meso* – *G. Mesonephrotus*; *G. Meta* – *G. Metanephrotus*; *G. Neo* – *G. Neonephrotus*; *G. Para* – *G. Paranephrotus* according to terms proposed by Malmberg (1956, 1970); see text for further details.

insights into the phylogenetic relationship within the genus. As the most commonly sequenced region of the genome, the complete ITS rDNA sequences of 37 *Gyrodactylus* species were used to infer the phylogeny. This region has proven valuable within *Gyrodactylus* (Cable *et al.* 1999; Zietara *et al.* 2002) and demonstrated the potential to resolve the phylogeny of different monogeneans (Bentz *et al.* 2001), parasite groups (e.g. Audebert, Durette-Desset & Chilton, 2000) or plants (Steane *et al.* 1999).

Three phylogenetic methods revealed trees of almost identical topology, with strong support at some critical nodes and confirmed the ability of the ITS to infer phylogeny in *Gyrodactylus* in our data set. *Gyrodactylus* falls into 3 well-supported clades, suggesting a basal, but not monophyletic, origin of the *G. (Gyrodactylus)* subgenus (*G. markakulensis* and the taxa in clade A). Separation of the *G. (Gyrodactylus)* species from the others is not surprising, considering the molecular and morphological data. Even in the relatively conserved SSU rDNA (V4 region) (see Cunningham *et al.* 1995a), these species varied by up to 12% from those of the

G. (Limnonephrotus) subgenus (Matejusová *et al.* 2001). The basal position of *G. (Gyrodactylus)* in the whole genus might be confirmed by the fact that this subgenus shares some plesiomorphic characters, such as the median junction between the two anterior systems of the excretory system, with the genus *Macroglyrodactylus* (see Malmberg, 1964, 1970), which has been considered the closest ancestor of *Gyrodactylus* (Malmberg, 1998). Based on the morphology of the attachment apparatus, mainly the ventral bar, some similarities can be drawn between the present members of the *G. (Gyrodactylus)* subgenus. Typically, the ventral bar has a long narrow membrane and no lateral processes (*Gyrodactylus elegans*-group) or a tongue-shape membrane with very short or no lateral processes (*Gyrodactylus phoxini*-group) (see Malmberg, 1970). The position of *G. markakulensis* is exceptional as it was consistently resolved as the most basal taxon in all phylogenetic analyses performed. However, the position of this species is controversial, and it was included, albeit with some reservations, in both *G. elegans*- and *G. phoxini*-species groups of *G. (Gyrodactylus)*

(Malmberg, 1970). Based on the morphology of the ventral bar and marginal hooks, *G. markakulensis* seems to fit into the *G. phoxini*-group. However, there are some specific characteristics that may support exclusion of this species from the *G. phoxini*-group or even the *G.* (*Gyrodactylus*) subgenus, such as the specific shape of the marginal hook tip. Moreover, there were differences in penis morphology; the penis of *G. markakulensis* is typified by rows of fine penis spines, finer than those of other *Gyrodactylus* subgenera. However, this is a plesiomorphic character shared with *G. sedelnikowi* and the other species of the *G.* (*Gyrodactylus*) subgenus, typified by a row of these fine spines and a row of larger penis spines.

The monophyletic origin of the other 5 *Gyrodactylus* subgenera, based on the present analyses, is also controversial. They all fall into one well-supported clade as a sister group to *G.* (*Gyrodactylus*) and *G. markakulensis*. Within this clade, 2 groups are recognized, separating species of *G.* (*Mesonephrotus*) and *G.* (*Metanephrotus*) from those of *G.* (*Paranephrotus*), *G.* (*Neonephrotus*) and *G.* (*Limnonephrotus*). None of these subgenera were found to be monophyletic. Within the *G.* (*Metanephrotus*) and *G.* (*Mesonephrotus*) clade, there are 2 well-supported associations; the first consists of *Gyrodactylus turnbulli*, *Gyrodactylus poeciliae* and *Gyrodactylus bullatarudis*. These species are specific parasites of fish from the genus *Poecilia* and their close relationship was previously suggested by Harris & Cable (2000). The excretory system of *G. turnbulli* was described as a *G.* (*Metanephrotus*)-like system (Harris, 1986), and, based on the morphology of the marginal hooks and ventral bar, this species falls into the *G. eucaliae*-group (Cable *et al.* 1999). However, based on the definition of *G.* (*Metanephrotus*) (Malmberg, 1970), there is no strong evidence of convincing autapomorphies to place this subgenus as 'more derived' as presented by Malmberg (1998). Nevertheless, the position of species within this clade might be biased by the fact that only a few species of the *G. eucaliae*-group were sequenced, and also by different mechanisms such as host-parasite coevolution that may play an important role.

The third well-supported clade consists of species of the *G.* (*Limnonephrotus*), *G.* (*Paranephrotus*) and *G.* (*Neonephrotus*) subgenera. Species of the 2 latter subgenera clustered together in a terminal position of the tree and this could be a consequence of the limited number of species sequenced. The morphology of the ventral bar and marginal hook of species of the *G.* (*Paranephrotus*) subgenus is similar to the majority of species of the *G.* (*Limnonephrotus*) subgenus. In addition, the marginal hook of *G. anguillae* is also of similar shape to *G.* (*Paranephrotus*) species but the ventral bar lacks lateral processes. Some of the terminal resolutions, especially the group of (*Gyrodactylus vimbi* – *Gyrodactylus luciopercae*)

are also worth mentioning, as the morphology of the attachment apparatus is very similar and might support the idea of species groups based on the shape of the marginal hook and ventral bar. Close relationships among the majority of species in the group have been discussed already, and the *G. wagneri*-species group to which these species belong was considered as monophyletic, as was the *G.* (*Limnonephrotus*) subgenus (Zietara & Lumme, 2002). A greater number of species of the *G. wagneri*-group were included in the present study (especially species parasitizing cyprinids), and the monophyletic origin of the *G. wagneri*-group and the *G.* (*Limnonephrotus*) subgenus was rejected.

Zietara *et al.* (2002) claimed that analysis of ITS sequence revealed deep divisions within the genus *Gyrodactylus* that followed Malmberg's (1970) phylogeny. This study has shown that the monophyly of groups demonstrated by Zietara *et al.* (2002) cannot be supported, and may have been a result of the low number of species studied. Their conclusions, based on analysis of only 10 from a genus that contains over 400 species, appear to have been premature, and the close grouping of the *G. wagneri*-group species found by Zietara & Lumme (2002) may be expected from the species studied, which represented restricted host and geographical ranges. Future studies may reveal similar deep divisions within this and other genera and it is likely that analysis of additional species of *Gyrodactylus* will produce more species groups that are difficult to resolve by use of ITS alone.

Finally, we conclude from the results of the present phylogenetic analyses that the characteristics of the excretory system of *Gyrodactylus* as presented by Malmberg (1970) do not seem to be sufficiently conservative or informative to reveal subgenera within *Gyrodactylus*. Moreover, it is impossible to use these characters when the excretory system is unknown in the majority of newly described species. The validity of species groups within this genus is supported, as the morphology of the ventral bar and marginal hooks seem to have power to inform us about relationships between *Gyrodactylus* species. However, we found that some authors do not place species in any species group as part of the species description and that comprehensive revisions might be necessary. The present phylogenetic analyses inferred from the complete ITS region give us satisfactory, although limited, resolution and different topologies may form within the terminal groups when other regions of DNA are analysed.

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REFERENCES

- AUDEBERT, F., DURETTE-DESSET, M.-C. & CHILTON, N. B. (2000). Internal transcribed spacer rDNA can be used to infer the phylogenetic relationships of species within the genus *Nematodirus* (Nematoda: Molineoidea). *International Journal for Parasitology* **30**, 187–191.
- BAKKE, T. A., HARRIS, P. D. & CABLE, J. (2002). Host specificity dynamics: observations on gyrodactylid monogeneans. *International Journal for Parasitology* **32**, 281–308.
- BENTZ, S., LEROY, S., DUPREEZ, L., MARIAUX, J., VAUCHER, C. & VERNEAU, O. (2001). Origin and evolution of African *Polystoma* (Monogenea: Polystomatidae) assessed by molecular methods. *International Journal for Parasitology* **31**, 697–705.
- CABLE, J., HARRIS, P. D., TINSLEY, R. C. & LAZARUS, C. M. (1999). Phylogenetic analysis of *Gyrodactylus* spp. (Platyhelminthes: Monogenea) using ribosomal DNA sequences. *Canadian Journal of Zoology* **77**, 1439–1449.
- CUNNINGHAM, C. O. (1997). Species variation within the internal transcribed spacer (ITS) region of *Gyrodactylus* (Monogenea: Gyrodactylidae) ribosomal RNA genes. *Journal of Parasitology* **83**, 215–219.
- CUNNINGHAM, C. O., MCGILLIVRAY, D. M., MACKENZIE, K. & MELVIN, W. T. (1995a). Discrimination between *Gyrodactylus salaris*, *G. derjavini* and *G. truttae* (Platyhelminthes: Monogenea) using restriction fragment length polymorphisms and an oligonucleotide probe within the small subunit ribosomal RNA gene. *Parasitology* **111**, 87–94.
- CUNNINGHAM, C. O., MCGILLIVRAY, D. M. & MACKENZIE, K. (1995b). Phylogenetic analysis of *Gyrodactylus salaris* Malmberg, 1957 based on the small subunit (18S) ribosomal RNA gene. *Molecular and Biochemical Parasitology* **71**, 139–142.
- CUNNINGHAM, C. O., MO, T. A., COLLINS, C. M., BUCHMANN, K., THIERY, R., BLANC, G. & LAUTRAITE, A. (2001). Redescription of *Gyrodactylus teuchis* Lautraite, Blanc, Thiery, Daniel & Vigneulle, 1999 (Monogenea: Gyrodactylidae), a species identified by ribosomal RNA sequence. *Systematic Parasitology* **48**, 141–150.
- ERGENS, R. (1966). Revision of the helminthofauna of fishes from Czechoslovakia IV. *Folia Parasitologica* **13**, 212–221.
- ERGENS, R. (1976). *Gyrodactylus barbi* sp. n. (Monogenoidea) from the fins of barbels. *Vestník Československé Společnosti Zoologické* **40**, 1961–1962.
- ERGENS, R. (1980). On the problem of three species of the genus *Gyrodactylus*, members of “*G. wageneri*-group” (Gyrodactylidae: Monogenea). *Helminthologia* **17**, 257–267.
- ERGENS, R. & BYCHOWSKY, B. E. (1967). Revision of the species *Gyrodactylus nemachili* Bychowsky, 1936 (Monogenoidea). *Folia Parasitologica* **14**, 225–238.
- ERGENS, R. & YUKHIMENKO, S. S. (1975). *Gyrodactylus* (Monogenoidea) from some Rhodeinae (Cypriniformes). *Folia Parasitologica* **22**, 33–36.
- ERNST, I., FLETCHER, A. & HAYWARD, C. (2000). *Gyrodactylus anguillae* (Monogenea: Gyrodactylidae) from anguillid eels (*Anguilla australis* and *Anguilla reinhardtii*) in Australia: a native or an exotic? *Journal of Parasitology* **86**, 1152–1156.
- FARRIS, J. S., KÄLLERSJO, M., KLUGE, A. G. & BULT, C. (1994). Testing significance of incongruence. *Cladistics* **10**, 315–319.
- GLÄSER, H.-J. (1974a). Sechs neue Arten der *Gyrodactylus-wageneri* Gruppe (Monogenea, Gyrodactylidae) nebst Bemerkungen zur Preparation, Determination, Terminologie und Wirtsspezifität. *Zoologischer Anzeiger* **192**, 56–76.
- GLÄSER, H.-J. (1974b). Eine neue Artengruppe des Subgenus *Gyrodactylus* (Paranephrotus) (Monogenea, Gyrodactylidae). *Zoologischer Anzeiger* **192**, 271–278.
- HARRIS, P. D. (1985). Species of *Gyrodactylus* von Nordmann, 1832 (Monogenea: Gyrodactylidae) from freshwater fishes in southern England, with a description of *Gyrodactylus rogatensis* sp. nov. from the bullhead *Cottus gobio* L. *Journal of Natural History* **19**, 791–809.
- HARRIS, P. D. (1986). Species of *Gyrodactylus* von Nordmann, 1832 (Monogenea, Gyrodactylidae) from poeciliid fishes, with a description of *G. turnbulli* sp. nov. from the guppy, *Poecilia reticulata* Peters. *Journal of Natural History* **20**, 183–191.
- HARRIS, P. D. & CABLE, J. (2000). *Gyrodactylus poeciliae* n. sp. and *G. milleri* n. sp. (Monogenea: Gyrodactylidae) from *Poecilia caucana* (Steindachner) in Venezuela. *Systematic Parasitology* **47**, 79–85.
- HUELSENBECK, J. P. & RONQUIST, F. (2001). MrBayes: Bayesian inference of phylogenetic trees. *Bioinformatics* **17**, 754–755.
- HUELSENBECK, J. P., RONQUIST, F., NIELSEN, R. & BOLLBACK, J. P. (2001). Bayesian inference of phylogeny and its impact on evolutionary biology. *Science* **294**, 2310–2314.
- HUYSE, T. & VOLCKAERT, F. A. M. (2002). Identification of a host-associated species complex using molecular and morphometric analyses, with the description of *Gyrodactylus rugienoides* n. sp. (Gyrodactylidae, Monogenea). *International Journal for Parasitology* **32**, 907–919.
- JEANMOUGIN, F., THOMPSON, J. D., GOUY, M., HIGGINS, D. G. & GIBSON, T. J. (1998). Multiple sequence alignment with Clustal X. *Trends in Biochemical Sciences* **23**, 403–405.
- MADDISON, W. P. & MADDISON, D. R. (2000). MacClade. Version 4. Sinauer Associates, Sunderland, Massachusetts.
- MALMBERG, G. (1956). Om förekomsten av *Gyrodactylus* på svenska fiskar. Skrifter utgivna av Södra Sveriges Fiskeriförening, Årsskrift, pp. 19–76.
- MALMBERG, G. (1964). Taxonomical and ecological problems in *Gyrodactylus* (Trematoda, Monogenea). In *Parasitic Worms and Aquatic Conditions* (ed. Ergens, R. & Rysavy, B.), pp. 203–230. Czech Academy of Science, Prague, Czech Republic.
- MALMBERG, G. (1970). The excretory systems and the marginal hooks as a basis for the systematics of *Gyrodactylus* (Trematoda, Monogenea). *Arkiv för Zoologi, Serie 2, Band, 23* (1). Almqvist & Wiksell, Stockholm.
- MALMBERG, G. (1998). On the evolution within the family Gyrodactylidae (Monogenea). *International Journal for Parasitology* **28**, 1625–1635.

- MATEJUSOVA, I., GELNAR, M., McBEATH, A. J. A., COLLINS, C. M. & CUNNINGHAM, C. O. (2001). Molecular markers for gyrodactylids (Gyrodactylidae: Monogenea) from five fish families (Teleostei). *International Journal for Parasitology* **31**, 738–745.
- OLSON, P. D. & LITTLEWOOD, D. T. J. (2002). Phylogenetics of the Monogenea – evidence from a medley of molecules. *International Journal for Parasitology* **32**, 233–244.
- POSADA, D. & CRANDALL, K. A. (1998). Modeltest: testing the model of DNA substitution. *Bioinformatics* **14**, 817–818.
- STEANE, D. A., MCKINNON, G. E., VAILLANCOURT, R. E. & POTTS, B. M. (1999). ITS sequence data resolve higher level relationships among the eucalypts. *Molecular Phylogenetics and Evolution* **12**, 215–223.
- SWOFFORD, D. L. (2002). PAUP*. Phylogenetic Analysis Using Parsimony (*and Other Methods). 4th Edn. Sinauer Associates, Sunderland, Massachusetts.
- WILLIAMS, H. & JONES, A. (1994). Parasitic worms of fish. Taylor and Francis, London.
- ZIETARA, M. S., ARNDT, A., GEETS, A., HELLEMANS, B. & VOLCKAERT, F. A. M. (2000). The nuclear rDNA region of *Gyrodactylus arcuatus* and *G. branchicus* (Monogenea: Gyrodactylidae). *Journal of Parasitology* **86**, 1368–1373.
- ZIETARA, M. S., HUYSE, T., LUMME, J. & VOLCKAERT, F. A. M. (2002). Deep divergence among subgenera of *Gyrodactylus* inferred from rDNA ITS region. *Parasitology* **124**, 39–52.
- ZIETARA, M. S. & LUMME, J. (2002). Speciation by host switch and adaptive radiation in a fish parasite genus *Gyrodactylus* (Monogenea, Gyrodactylidae). *Evolution* **56**, 2445–2458.
- ZITNAN, R. (1964). *Gyrodactylus rhodei* sp. n. – a new monogenean species from skin of *Rhodeus sericeus amarus* (Bloch). *Helminthologia* **5**, 1–4.