






RESEARCH ARTICLE

Using cover crops to offset greenhouse gas emissions from a tropical soil under no-till

João Paulo Gonsiorkiewicz Rigon^{1*}, Juliano Carlos Calonego¹, Laércio Augusto Pivetta², Gustavo Castoldi³, Juan Piero Antonio Raphael¹ and Ciro Antônio Rosolem¹

¹São Paulo State University, UNESP, College of Agricultural Sciences, Department of Crop Science, Botucatu-SP, Brazil, ²Federal University of Paraná, UFPR, Department of Agronomic Sciences, Palotina-PR, Brazil and ³Goiano Federal Institute, IF Goiano, Department of Agriculture, Rio Verde-GO, Brazil

*Corresponding author. Email: jp.rigon@unesp.br

(Received 11 September 2020; revised 30 April 2021; accepted 14 July 2021; first published online 21 September 2021)

Abstract

Crop rotations under no-till (NT) have been a strategy to increase soil organic carbon (SOC) and mitigate greenhouse gas (GHG) emissions, enhancing the cropping system efficiency. However, there is still controversy on the role of grasses and legumes, and species diversity and their impacts. This study aimed to assess the GHG emissions, SOC, and Nitrogen (TN) in a soybean production system managed under NT in rotation with different species in the fall–winter and the spring seasons. Main plots during the fall–winter were (1) Triticale (*x Triticosecale*) and (2) Sunflower (*Helianthus annuus*). Subplots established in the spring were (a) Sunn hemp (*Crotalaria juncea*), (b) Sorghum (*Sorghum bicolor*), (c) Pearl millet (*Pennisetum glaucum*), plus a (d) Fallow treatment. Soybean was grown every year in the summer, in sub-subplots. The GHG emission was affected according to crop species. In the spring, Sunn hemp emitted more nitrous oxide (N₂O) (0.82 kg ha⁻¹) than fallow (0.58 kg ha⁻¹); however, the high C and N inputs by the legume and also other cover crop residues reduced the relative emissions compared with fallow. Growing pearl millet or Sunn hemp as a spring cover crop increases SOC by 7% on average compared with fallow. The N₂O emission of Sunn hemp accounted for only 0.28% of the total N accumulated in the legume residues, notably lower than IPCC estimates. In the fall–winter, Triticale increased SOC by 7%, decreased CO₂ emission by 18%, and emitted 20% lower GHG to produce the same soybean yield compared with sunflower. Soybean rotation with triticale in fall–winter and Sunn hemp or pearl millet in spring decreases GHG emissions. Our results indicate that the right choice of species in rotation with soybean under NT increases SOC and may offset GHG emissions from tropical soils. It may be an important tool in mitigating potential global warming.

Keywords: Conservation management; crop rotation; Nitrous oxide

Introduction

In tropical soils, management practices such as no-till (NT) and crop rotation have the potential to increase soil organic carbon (SOC), either through the retention of crop residues on the soil surface (Kaye and Quemada, 2017; Poeplau and Don, 2015) or by changes in the soil caused by roots (Castro *et al.*, 2015; Garcia *et al.*, 2013), especially if legumes such as Sunn hemp (*Crotalaria juncea*) are included in the crop rotations (Raphael *et al.*, 2016).

Under tropical climate, usually, soil C accumulation is lower than in temperate regions because of the fast crop residue decomposition caused by the higher average temperatures (Bolliger *et al.*, 2006; Powlson *et al.*, 2016), leading to higher greenhouse gas (GHG) emission rates. In this case, cover crops play an important role in the supply of C via crop residues (Le *et al.*, 2018), mainly

when under NT (Rigon *et al.*, 2020). Legumes are an important source of N and are recognized as essential for soil C sequestration (Sisti *et al.*, 2004). A recent study observed that Sunn hemp has a lower potential for nitrous oxide (N₂O) emissions than other leguminous crops (Sant'Anna *et al.*, 2018). Conversely, grass residues have greater recalcitrance due to higher lignin and cellulose concentrations than legumes, leading to a lower soil organic matter (SOM) mineralization rate and N₂O emission (Pimentel *et al.*, 2015).

The GHG mitigation potential of conservation agriculture has been frequently ignored (Powlson *et al.*, 2015), and the effect of crops cultivated in each system on GHG emissions has been little studied. As the composition of crop residues and C and N input in the soil are recognized as N₂O (Pugesgaard *et al.*, 2017) and CO₂ emission modulators (Rigon *et al.*, 2018), each species in the rotation has a different potential for GHG emission mitigation and C sequestration (Kaye and Quemada, 2017; Lal, 2015; Rigon *et al.*, 2020). Thus, the hypotheses of this study were: (i) GHG emission modulation depends on the plant species originating the residues on the soil surface; (ii) the use of cover crops with a high C and N input, despite increasing CO₂ and N₂O emissions, has the potential to reduce the emission of these gases and increase soil C and N. The objective of this study was to evaluate GHG emissions, and soil C and N accumulation as affected by species grown in rotation with soybean in a long-term cropping system under NT (2003–2013).

Materials and Methods

Location and edaphoclimatic characterization of the experimental area

The study was conducted in a long-term NT system, where soybean was grown in rotation with fall–winter crops and spring cover crops for over 10 years. Each plot was cropped following the same crop rotation every year. The experiment was started in 2003 in Botucatu, SP (22°49'S and 48°25'W), at an altitude of 780 m, on a Typic Rhodudalf (Soil Survey Staff, 2014), with 655 g clay kg⁻¹, 237 g silt kg⁻¹, and 108 g sand kg⁻¹ at 0–0.2 m soil depth. The soil chemical (Raij *et al.*, 2001) and physical (Smith and Mullins, 1991) characteristics at the 0–0.1 m soil depth were as follows: pH CaCl₂ (5.1), Al³⁺ (0.2 mmol dm⁻³), Ca (43 mmol dm⁻³), Mg (36 mmol dm⁻³), K (1.8 mmol dm⁻³), and P (33 mg dm⁻³); bulk density (1.32 Mg m⁻³), microporosity (0.42 m³ m⁻³), and macroporosity (0.12 m³ m⁻³). The tropical climate is mesothermal with a well-defined dry fall–winter season from May to August. The wet season usually starts in September, extending through late March/April. The average annual rainfall is 1450 mm.

Experimental design

The experiment was laid out as a split-plot in a randomized complete block design with eight treatments and four replications (Figure 1). The main plots consisted of species grown in the fall–winter, after soybean harvest (April–September) and subplots consisted of cover crops grown in the spring (September–December). Soybean was grown in all plots from December to March. The experiment was started in 2003 with the fall–winter crops triticale [*X Triticosecale* (Wittmack)] and sunflower [*Helianthus annuus* (L.)] grown in the fall–winter in 32 m × 5 m plots. After the termination of these fall–winter crops, the cover crops pearl millet [*Pennisetum glaucum* (L.)], Sunn hemp [*Crotalaria juncea* (L.)], and sorghum [*Sorghum bicolor* (L.)] were sown, plus a fallow treatment during the spring, in 8 m × 5 m subplots. Soybean [*Glycine max* (L.) Merrill] was grown each year over the entire area in the summer (December–April). The species grown in the fall–winter, spring, and summer seasons were repeated annually from 2003 to 2013, as shown in Supplementary Table S1.

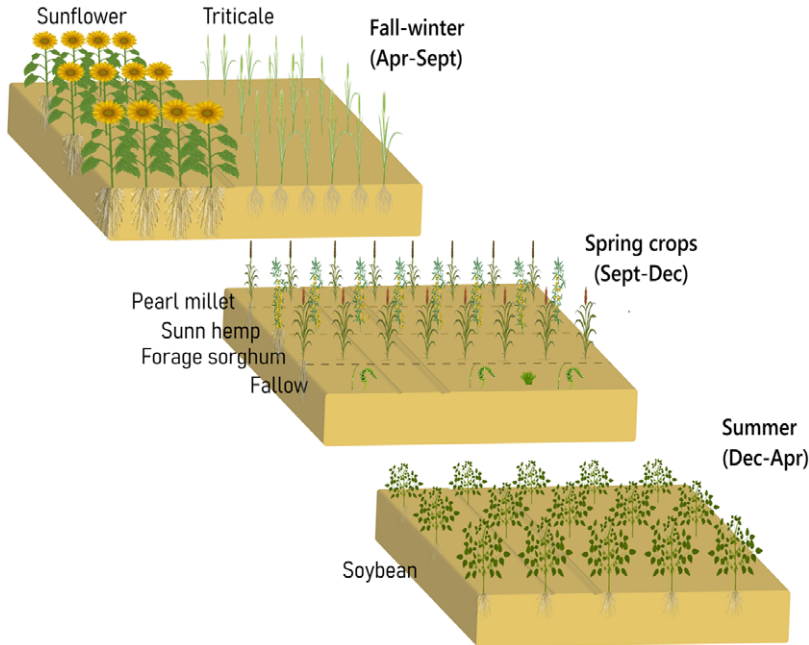


Figure 1. Scheme of experimental design of crop rotation according to fall-winter and spring treatments: Sunflower/Pearl millet, Sunflower/Forage sorghum, Sunflower/Sunn hemp, Sunflower/Fallow, Triticale/Pearl millet, Triticale/Forage sorghum, Triticale/Sunn hemp, Triticale/Fallow. Soybean is the main crop in the summer.

Crop management

Each year triticale and sunflower were sown in the plots in April at 0.17 and 0.34 m spacing, using 165 and 22 kg ha⁻¹ of seeds, respectively, without fertilizers. The harvest was carried out each year in September using a plot harvester. In October, around 1 week after chemical desiccation of the standing plant residues with glyphosate, the spring crops were sown in the subplots over the crop residues, except for the area kept under fallow. The row spacing was 0.17 m, using 15, 25, and 30 kg ha⁻¹ of seeds of sorghum, pearl millet, and Sunn hemp, respectively. The spring species were cultivated without fertilizers up to around 60 days after sowing (DAS) when the plants were desiccated with glyphosate.

Soybean (Monsoy M7211 RR) was sown each year in December, in rows 0.45 m apart from each other, using 430,000 seeds ha⁻¹. Yearly fertilization consisted of 50 kg ha⁻¹ of K₂O and 50 kg ha⁻¹ of P₂O₅, as potassium chloride and triple superphosphate, respectively. Soybean was harvested each April from three 5.0 m long central rows of each subplot using a plot harvester, and the yield was calculated at grain moisture of 13%.

Assessment of C and N in soil and above-ground crop residue (straw)

The assessments for this experiment were made through the 2012–2013 cropping season. At sowing of the fall-winter, spring, and soybean crops, two samples of above-ground crop residues were collected randomly from the soil surface (0.25 m² each) in each subplot. The samples were dried to constant weight at 60 °C. Then the samples were ground and homogenized, and part of them was used to determine the biochemical composition (Silva and Queiroz, 2002), which results are shown in Table 1. The other part of the samples was used to analyze C and N concentrations using an elemental analyzer (LECO-TruSpec[®] CHNS).

Table 1. Biochemical compositions of the crops used in the rotations

| Crop | C | N | Hemicellulose | Cellulose | Lignin | C: N | Lignin: N |
|--------------|------|-----|---------------|-----------|--------|------|-----------|
| | % | | | | | | |
| Triticale* | 44.0 | 0.4 | 20.0 | 49.2 | 17.3 | 94 | 37 |
| Sunflower* | 42.6 | 0.5 | 9.9 | 48.0 | 21.1 | 80 | 31 |
| Soybean* | 36.9 | 0.9 | 12.0 | 44.6 | 30.4 | 41 | 41 |
| Sorghum | 39.7 | 2.6 | 34.7 | 35.4 | 5.4 | 16 | 6 |
| Pearl millet | 42.1 | 3.0 | 27.5 | 24.0 | 10.1 | 14 | 4 |
| Sunn hemp | 42.8 | 3.7 | 11.9 | 33.8 | 14.9 | 12 | 5 |

*Postharvest residue left on the soil.

Before sowing the fall–winter crops, three soil subsamples were collected on 16 April, 2012 from each experimental unit at 0–0.1 m depth using an auger. The samples were air-dried, ground in a ball mill, and analyzed for total organic carbon (SOC) and total soil nitrogen (TN) in an elemental analyzer (LECO-TruSpec® CHNS).

Greenhouse gases sampling and analysis

The GHG were always sampled from 8:00 to 10:00 AM at 1, 3, 8, 15, 30, and 50 DAS for the crops grown in spring, and at 1, 3, 8, 15, 30, 60, and 90 DAS for the plant species grown in fall–winter and summer. Closed chambers were constructed according to Bowden *et al.* (1990); these chambers consisted of a cylindrical steel drum measuring 30 cm in diameter by 13 cm in height, and they were inserted 7 cm into the soil. The drums remained in the area for the entire period of the experiment and were removed only for sowing and harvesting, and they were placed again in the soil at least 24 hours before the next sampling. At the time of sampling, the drums were hermetically sealed with polyvinyl chloride (PVC) lid, 9 cm in height and the same diameter as the drum. The lids were equipped with an 8 mm rubber septum through which air samples were collected from inside the chambers. For determination of the GHG flux, air samples were collected at 0, 10, 20, and 40 minutes after the chamber closure using 20 mL polypropylene syringes (BD20-mL syringe Luer-Lok™, US) with a three-way valve. Immediately after sampling, the samples were placed in coolers with ice and sent to the laboratory where CO₂, N₂O, and CH₄ concentrations were analyzed no more than 24 hours after sampling to ensure the integrity of the samples (Rigon *et al.*, 2017). The gas chromatograph (GC-2014, Shimadzu, Columbia, MD, EUA) used was equipped with a Porapak Q column and two detectors: an electron capture detector (ECD) that quantifies the N₂O and a flame ionization detector (FID) that quantifies the methane and carbon dioxide indirectly. The chromatographic conditions employed were as follows: packed column set at 80 °C, FID set at 250 °C, and EDC detector set at 325 °C, with carrier gas N 5.0 and P5 gas (95% argon and 5% methane) for improved efficiency of EDC; methanator set at 350 °C; and the Porapak Q column set at 80 °C, with N₂ as the carrier gas enriched with N₂O gas in ‘back-flush’ system and with manual injection. The chromatograph was calibrated with the standard gases (White Martins®) (CO₂: 270, 648, 2063, and 7164 μmol mol⁻¹; CH₄: 0.69, 2.06, 3.05, and 9.05 ηmol mol⁻¹; N₂O: 305, 693, 1092, and 1885 ηmol mol⁻¹).

The variation of the gas concentration inside the chamber with time was used to calculate the GHG fluxes (Hutchinson and Mosier, 1981) according to equation 1 (Kim *et al.*, 2002):

$$f = \frac{\Delta C}{\Delta t} x \frac{v}{a} x \frac{m}{Vm} \quad (1)$$

where f is the flux of CO₂ (mg m⁻² h⁻¹), N₂O (μgm⁻² h⁻¹), or CH₄ (μgm⁻² h⁻¹); ΔC is the change in the GHG concentration as a function of the variation in the chamber closure time (Δt), where $\Delta C/\Delta t$ is the slope of the line equation; v is the chamber volume (0.0128 m³); a is the soil area covered by

the chamber (0.071 m^2); m is the molar mass of CO_2 (44.01 g mol^{-1}), CH_4 (16.04 g mol^{-1}), or N_2O (44.01 g mol^{-1}); and V_m is the molar volume of the gases, which was corrected using the air temperature inside the chamber, according to the ideal gas equation.

The accumulated emissions were calculated by trapezoidal integration of the daily emission using Origin software (OriginLab, Ltd., Northampton, MA, USA). The relative C- CO_2 and N_2O -N emissions were obtained by dividing the accumulated C- CO_2 and N_2O -N emissions, by the accumulated C and N in the above-ground crop residues on the soil surface (Qin *et al.*, 2012).

The gravimetric water content (M_{soil}) and the soil temperature (T_{soil}) at 5 cm depth were measured simultaneously with the GHG samplings, using a 5 TM sensor (Decagon Devices). The soil gravimetric moisture was converted to volumetric moisture according to previous calibration (data not shown), thus enabling calculation of the water-filled pore space (WFPS) using equation 2, according to Linn and Doran (1984):

$$\text{WFSP} = \frac{M_{\text{soil}}}{\text{TP}} \times 100 \quad (2)$$

where WFPS is the water-filled pore space (%); M_{soil} is the volumetric soil water content ($\text{m}^3 \text{ m}^{-3}$); and TP is the total porosity of the soil ($\text{m}^3 \text{ m}^{-3}$), as determined by physical analysis with volumetric rings and reported in the results of Calonego *et al.* (2017).

The CO_2 equivalent ($\text{CO}_2\text{-eq}$) is a metric used to compare GHG emissions based on their global warming potential (GWP), where the CO_2 is typically taken as the reference gas, and an increase or reduction in emission of CH_4 and N_2O is converted into 'CO₂-equivalents' through their GWPs. For CH_4 the GWP (IPCC, 2014) is assumed to be 25, and the GWP for N_2O is 298 over 100 years based on the gas mass and atmosphere lifetime. Thereafter, greenhouse gas intensity (GHGI) (Mosier *et al.*, 2006) was calculated by dividing GWP by soybean grain yield, with the results presented in kg kg^{-1} of grain.

Statistics

Data were analyzed using a split-split plot design in randomized complete blocks with four replicates. After testing for homogeneity and normality ANOVA was performed using SAS version 9.2 (SAS, Inc., 2009), and the mean differences were compared by the t -test (LSD, $p < 0.05$). For soil temperature; WFPS; and C- CO_2 , C- CH_4 , and N_2O -N emissions, the standard deviation of the mean was calculated for each period.

Results

Carbon and nitrogen inputs by crop rotations

The amount of crop residue on the soil surface, as well as its C and N contents, was impacted by the species grown in each crop season. As expected, the amount of residues accumulated with fallow was lower than with the spring crops, which did not differ from each other (Figure 2a and b). Cropping species in the spring season reached, on average, 13.5 Mg ha^{-1} of residue on the soil surface, surpassing in 5.2 Mg ha^{-1} the residues observed under fallow (Figure 2a). The amount of C in by crop residues followed the same trend as the biomass. Regardless of the species, spring crops accumulated an additional 2.1 Mg ha^{-1} of C compared with fallow (Figure 2b).

The N accumulated in the crop residues of the cover crops grown in the spring ranged from 100 to 215 kg ha^{-1} depending on the species, which represented a 57–73% increase in the N accumulated in crop rotations including fallow. Rotations where Sunn hemp was grown in the spring resulted in the highest N accumulation in residues (approximately 290 kg ha^{-1}), regardless of the fall–winter crop. In this case, Sunn hemp contribution was 208 kg ha^{-1} on average. The cover crops grown after sunflower accumulated, on average, 180 kg ha^{-1} of N in the plant residues, surpassing the N accumulated in succession to triticale by 30 kg ha^{-1} (Figure 2c).

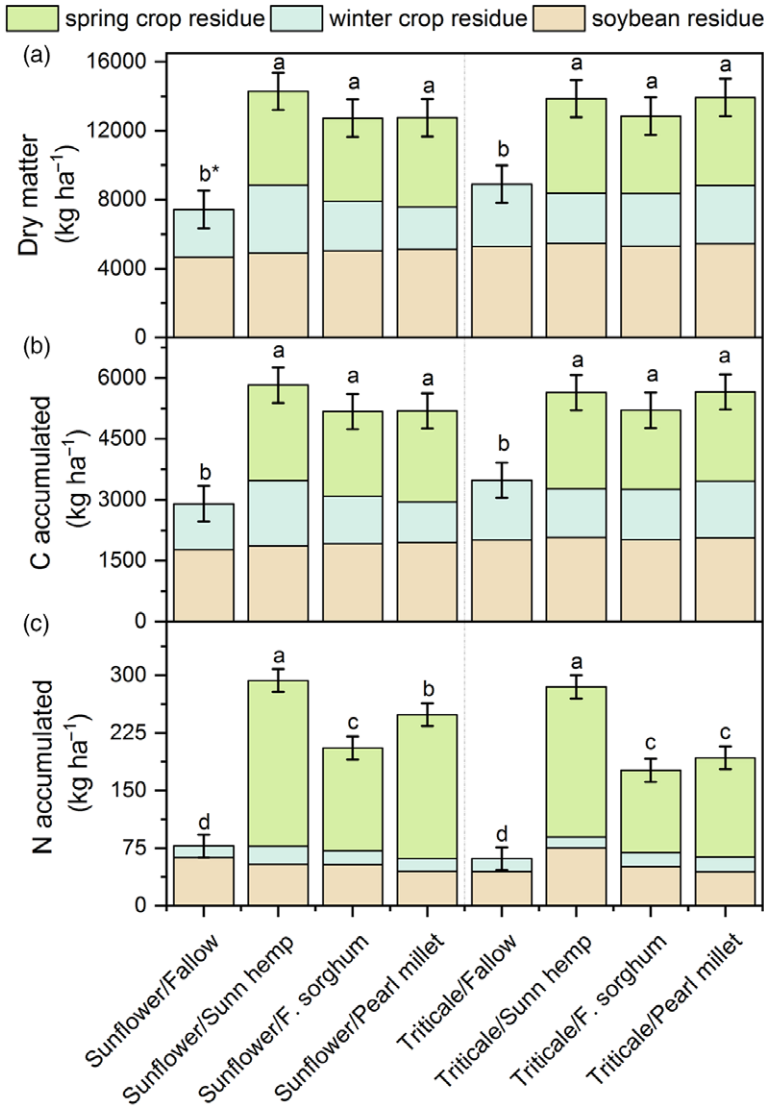


Figure 2. Dry matter (a), carbon (b), and nitrogen (c) are accumulated from residues on the soil surface, according to crop rotations in each growing season. Letters in the columns differ from each other by the *t*-test ($p < 0.05$) of the cumulative values, and the vertical bars correspond to LSD ($p < 0.05$).

Soil organic carbon and nitrogen

The SOC in the uppermost soil layer was affected by the crop rotations, but total N (TN) was not. SOC content was 7% higher when triticale was grown in fall–winter instead of sunflower. The absence of a spring cover crop (fallow) reduced SOC by 8% and 6%, with Sunn hemp and pearl millet, respectively. Although no significant differences were observed in TN ($p = 0.38$), the nominal values proportionally followed the SOC results; that is, TN was 10% higher for triticale than for sunflower (Table 2).

Table 2. Soil organic carbon (SOC) and total soil nitrogen (TN) concentrations at 0–0.1 m soil depth, cumulative greenhouse gases (C-CO₂, C-CH₄, and N₂O-N), relative N₂O-N and C-CO₂ emissions, and greenhouse gas intensity (GHGI) according to the fall-winter and spring crops.

| Crops | SOC | TN | C-CO ₂ | C-CH ₄ | N ₂ O-N | Relative emission | | GHGI |
|-----------------|-------------------------|------|--------------------------------|-------------------|--------------------|---------------------------|-------------------|------------------------|
| | | | | | | N ₂ O-N | C-CO ₂ | |
| | —(g kg ⁻¹)— | | ————(kg ha ⁻¹)———— | | | ——(g kg ⁻¹)—— | | (kg kg ⁻¹) |
| | | | | | | Winter crops | | |
| Sunflower | 21.5 ^b | 1.85 | 7,160 a | −0.84 | 0.76 | 4.60 | 1.58 a | 2.39 a |
| Triticale | 23.1 a | 2.05 | 5,874 b | −0.99 | 0.64 | 4.58 | 1.21 b | 1.93 b |
| LSD | 1.0 | 0.31 | 1,135 | 0.35 | 0.23 | 2.57 | 0.16 | 0.29 |
| | | | | | | Spring crops | | |
| Fallow | 21.3 b | 1.9 | 5,714 | −0.81 | 0.58 b | 8.66 a | 1.82 a | 1.84 |
| Sunn hemp | 23.1 a | 2.0 | 6,708 | −0.79 | 0.82 a | 2.85 b | 1.16 b | 2.16 |
| Sorghum | 22.2 ab | 1.9 | 7,164 | −0.72 | 0.63 ab | 3.28 b | 1.37 b | 2.32 |
| Pearl millet | 22.7 a | 1.9 | 6,483 | −1.34 | 0.78 ab | 3.58 b | 1.22 b | 2.31 |
| LSD | 1.3 | 0.2 | 1,531 | 6.6 | 0.21 | 1.8 | 0.39 | 0.5 |
| Fall/Winter (F) | 0.01 | 0.14 | 0.03 | 0.28 | 0.19 | 0.97 | <0.01 | 0.01 |
| Spring (S) | 0.04 | 0.38 | 0.28 | 0.21 | 0.04 | <0.01 | <0.01 | 0.19 |
| F x S | 0.32 | 0.35 | 0.24 | 0.15 | 0.69 | 0.95 | 0.39 | 0.15 |
| Averaged Mean | 22.33 | 1.95 | 6,517 | −0.91 | 0.70 | 4.59 | 1.39 | 2.16 |

*Means followed by different letters differ from each other by the paired *t*-test (LSD, *p* < 0.05).

GHG flux

The weather conditions during the experiment were typical of the region, validating the pattern of GHG emission in the fall–winter, spring, and summer seasons (Figure 3). N₂O emission peaks occurred at the beginning of the fall–winter season (up to 8 DAS) and early summer season (between 8 and 15 DAS), especially in crop rotations with Sunn hemp in spring (Figure 3a). However, in the fall–winter, the emissions did not exceed 60 μg m⁻² h⁻¹, which is much lower than the maximum flux of 140 μg m⁻² h⁻¹ observed with the triticale/sunn hemp rotation in the summer. For the CO₂ fluxes (Figure 3c), emission peaks were observed at 3 and 50 DAS in the spring season. At 3 DAS, the highest CO₂ emissions occurred with the crop rotations sunflower/fallow, sunflower/pearl millet, triticale/sorghum, and triticale/pearl millet, with an average flux of 1300 mg m⁻² h⁻¹. At 50 DAS, the highest CO₂ emissions were observed with pearl millet and sorghum in plots previously cropped to sunflower, with an average mean flux of 1900 mg m⁻² h⁻¹.

In the summer season (soybean), an increase in CO₂ emissions was observed only at 60 DAS, especially in crop rotations with sunflower (regardless of the spring crop used) and with triticale followed by fallow or Sunn hemp, with an average mean flux of 2500 mg m⁻² h⁻¹. Conversely, the crop rotations triticale/sorghum and especially triticale/pearl millet showed the lowest CO₂ emissions, with fluxes 31% and 52% lower than the maximum emissions observed, respectively.

The CH₄ emission was practically nonexistent (Figure 3b), or even negative, during the experiment, regardless of the crop rotation or the soil water-filled pore space (d), and soil temperature (e).

GHG emissions, GWP, and GHGI

Cropping sunflower during the fall–winter increased the cumulative C-CO₂ emission by 21 % compared with triticale. The highest CO₂ emission from sunflower resulted in higher GWP, GHGI, and higher relative CO₂ emissions. Concerning the spring crops, the highest N₂O-N emission was observed with Sunn hemp, which was 30% higher than fallow, but no different from the annual GWP and GHGI. Although the cumulative N₂O and C-CO₂ emissions were

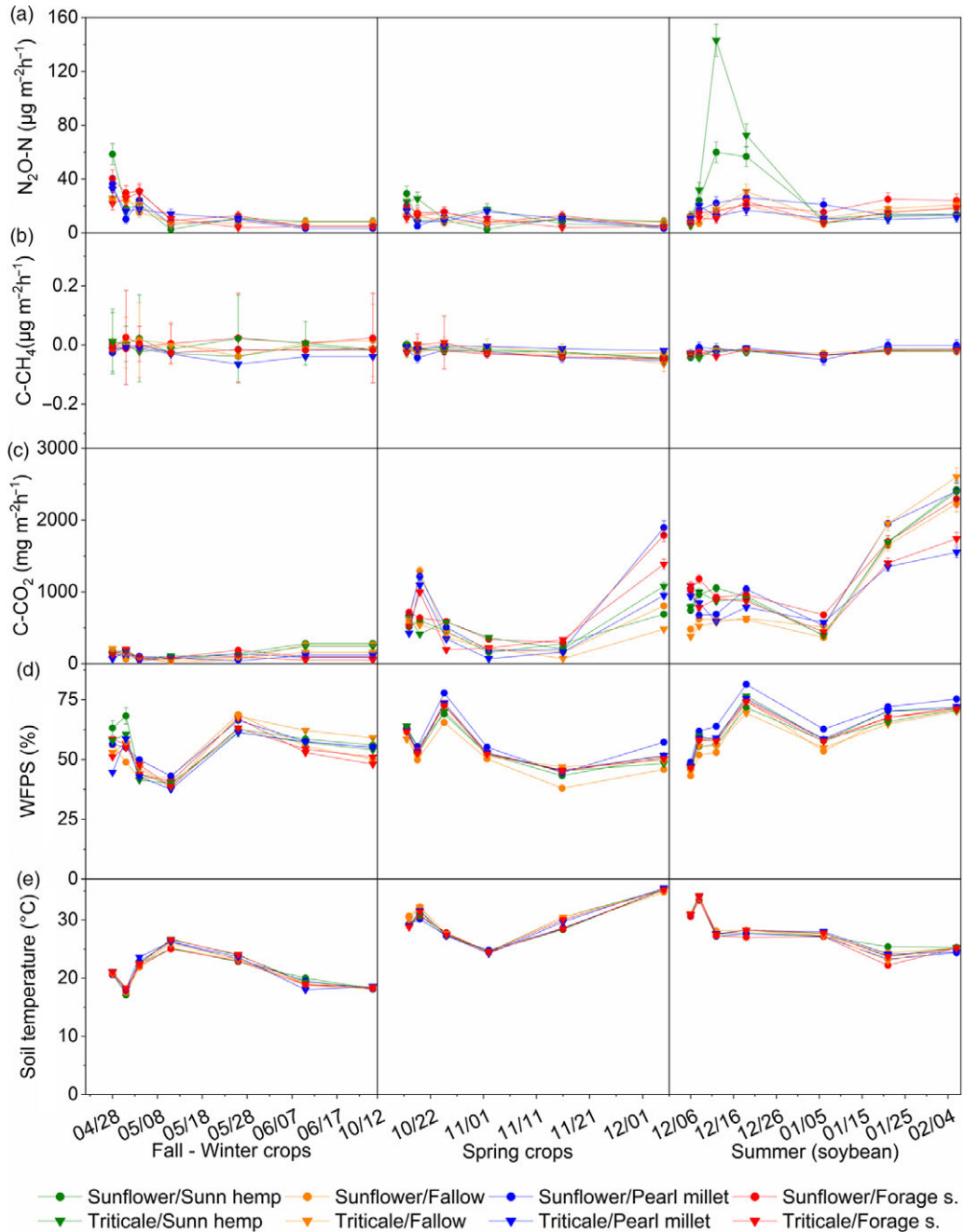


Figure 3. N₂O-N (a), C-CH₄ (b), C-CO₂ emissions (c), water-filled pore space (d), and soil temperature at 5 cm depth (e) of the crop rotations. Vertical bars correspond to the standard error of the mean.

not higher under fallow, the low accumulation of N and C in the residues resulted in relative emissions of 8.66 g kg⁻¹ and 1.82 kg kg⁻¹, respectively, which were approximately 2.5 and 1.5 times higher than the average obtained with millet, Sunn hemp, and sorghum (Table 2).

Discussion

C and N in crop residue and soil

The amount of C and N accumulated on the soil surface mainly from the spring crop residues is very important in NT cropping systems because the amount and quality of residues are determinants of the success of the system (Büchi *et al.*, 2018; Rigon *et al.*, 2020; Chen *et al.*, 2014), and these species were allowed to grow for only 60 days. These cover crops resulted in the accumulation of 5.1 Mg ha⁻¹ of residues on average, which represents around one-third of the cumulative dry matter input during the entire year. Considering the importance of keeping soil under NT covered most of the time (Büchi *et al.*, 2018; Tonitto *et al.*, 2006), the additional crop residue produced by the spring cover crops before planting soybean in the summer provide an additional ecosystem service to soybean cultivation option under NT (Rigon *et al.*, 2018). This condition is even more important in tropical regions with dry winters combined with the fast decomposition rates under high temperatures (Jantalia *et al.*, 2007; Lal, 2002). In weathered tropical soils, the heterotrophic microorganisms almost always find optimal environmental conditions for intense oxidation of SOM, causing high rates of substrate mineralization and consequent CO₂ emission (Sanchez and Logan, 1992). Hence, C losses must be offset by an increase in the amount of crop residues on the soil surface, which will increase SOC and TN, either by cover or cash crops. Although the similar C accumulation in the residues of the spring species, the highest N input with Sunn hemp explains the increase in SOC content with the use of this legume. Crop residue characteristics are essential to increase SOC under NT (Frasier *et al.*, 2016; Kaye and Quemada, 2017; Lal, 2004; Schipanski *et al.*, 2014), which confirms the importance of N input for SOC sequestration (Boddey *et al.*, 2010; Rigon and Calonego, 2020; Van Groenigen *et al.*, 2017). The high C and N in crop residues is considered one of the most important factors in increasing soil carbon retention (Yang *et al.*, 2018). Sunn hemp was able to supply, on average, 190 kg ha⁻¹ of N in only 60 days, confirming that it is one of the species with a greater capacity to fix N (Chikowo *et al.*, 2004). This result is important considering cropping systems with the absence of N fertilization and may help in enhancing the sustainability of agriculture in tropical regions (Rigon *et al.*, 2020). In addition to being the nutrient most taken up by plants, N is involved in the assimilation of C in stable and humified SOM fractions (Cyle *et al.*, 2016). Furthermore, the crop residue of Sunn hemp has a low C/N ratio, providing an excellent substrate for soil microbial activity and improve soil quality (Ferrari Neto *et al.*, 2020; Raphael *et al.*, 2016; Rigon *et al.*, 2020, 2021; Zhao *et al.*, 2018). The well-defined dry season limited the growth of sunflowers leading to an insufficient cover of the soil surface by residues compared with triticale (Rigon and Calonego, 2020). In addition, the high C/N and lignin/N ratios of the triticale residues compared with sunflower (Table 1) resulted in lower N mineralization rates and higher persistence of these residues (Rigon *et al.*, 2018; Rigon and Calonego, 2020). Grass residues are recognized by the presence of lignocellulose complexes that are acetylated or esterified by coumaric and ferulic acids, hindering their mineralization (del Río *et al.*, 2007). According to several authors (Chen *et al.*, 2014; Palm *et al.*, 2001; Rigon and Calonego, 2020), the biochemical characteristics of the crop residue, such as high levels of lignin and hemicellulose, cellulose, C/N ratio, and lignin/N ratio, are important indicators of its quality. These characteristics allow us to predict the persistence of crop residue on the soil surface and also the potential of C stabilization into SOM (Castellano *et al.*, 2015; Cotrufo *et al.*, 2013).

The high C/N ratio of triticale residue may pose a limitation to decomposers, which maintain soil covered for a longer time (Rigon *et al.*, 2018). Besides, the low residue decomposition rate results in lower C-CO₂ emissions to the atmosphere. This may explain the SOC increase by triticale, even with similar C accumulation by sunflower.

The SOC increase depends on the amount and quality of the residues (Frasier *et al.*, 2016; Rigon *et al.*, 2021), which are essential for conservation agriculture systems (Derpsch *et al.*, 2014). However, when fallowing is introduced in the rotation system, despite the low labile C

and microbial respiration (Bell *et al.*, 2003; Qiao *et al.*, 2014), loss of C-CO₂ is a continuous process (Novelli *et al.*, 2011), decreasing SOM over the years. We found evidence that the maintenance of the soil covered with crops and crop residues throughout the year is a good strategy to mitigate GHG emissions basically because less soil carbon can be respired by soil microorganisms because it is more efficiently maintained in the biomass. Thus, depending on their residue quality, spring cover crops, besides keeping the soil covered, can lead to an increase in SOC in systems under NT.

The balance between the C inputs and outputs in the soil determines C stabilization, which is a parameter that allows assessing the cropping systems management quality from the perspective of the soil conservation and sustainability, and GHG mitigation potential.

In tropical weathered soils, Fe and Al oxides can provide additional protection of SOC against decomposition under low pH (Lal, 2002; Six *et al.*, 2002). Thus, sustainable agricultural intensification, i.e. crop rotations under conservation management focusing on the maintenance of soil cover is an interesting tool to restore and increase SOC (Lal, 2015). It is important to note that the potential of SOC increase varies according to the crop rotation species and its residue characteristics under NT.

GHG emission

The GHG emissions were associated with the weather variations across the seasons, which regulate microbial activity and SOM mineralization (Khalil and Baggs, 2005; Zhang *et al.*, 2017). WFPS and soil moisture and temperature are recognized as the main factors controlling GHG fluxes (Oertel *et al.*, 2016) and help to explain the low GHG emissions during fall–winter (Figure 3). The N₂O emission from the crop residues depends on their N content (IPCC, 2006) and quality, i.e. their biochemical composition (Pimentel *et al.*, 2015). Legume plant residues with high N content, low C/N ratio, and low levels of recalcitrant compounds (Shahbaz *et al.*, 2017) stimulates N₂O emissions (Pugesgaard *et al.*, 2017), explaining the highest cumulative N₂O-N emission by Sunn hemp (Table 2). However, the effect of crop residues on N₂O emission is more complex due to the interactions of soil moisture, temperature, and microbial activity (Gonzaga *et al.*, 2018).

Similarly, crop residue quality may also affect CO₂ emission, as it was observed in this experiment in rotations with triticale and sunflower (Figure 2), and reported previously (Rigon *et al.*, 2018). High C/N and lignin/N ratios in the crop residues such as in triticale, results in a limited source of C for soil decomposers and, consequently, lower emissions of both N₂O (Yang *et al.*, 2018) and CO₂ (Sainju *et al.*, 2012).

Cropping systems that keep soil surface uncovered, such as fallow, may show temporary lower N₂O emissions owing to the lack of substrate. According to Šimek *et al.* (2004), one reason is that the consumption of O₂ during the residue decomposition results in an anaerobic environment prone to N₂O emission. Several studies in tropical regions show that keeping the soil covered is essential for improving the sustainability of the system (Gomes *et al.*, 2009; Jantalia *et al.*, 2008; Raphael *et al.*, 2016; Rigon *et al.*, 2018). Thus, despite the higher N₂O emissions in rotations including legume species (Gomes *et al.*, 2009), it is essential to consider also the effects on SOC, which is increased due to the greater availability of soil N. Thus, the emissions can be eventually offset by the retention of C in the SOM under tropical conditions (Bayer *et al.*, 2016).

Cumulative and relative emissions of GHG

The mean cumulative N₂O-N emission can be considered low (0.7 kg ha⁻¹), given the mean N input from residues of 192 kg ha⁻¹, representing 0.36% of the direct N₂O-N emission rate of 1% considered by the IPCC methodology (IPCC, 2006). In our experiment, the N₂O emission rate was similar to the direct emission of Brazilian agricultural soils, estimated at 0.3% (MCTI, 2010). Studies with crop rotation systems carried out in Brazil also observed the same tendency to

overestimate the N₂O emission when using IPCC emission factors (Gomes *et al.*, 2009; Jantalia *et al.*, 2008; Sant'Anna *et al.*, 2018). In our experiment, even when we considered the highest cumulative N₂O-N emission, which was 0.82 kg ha⁻¹ year⁻¹ in the crop rotation with Sunn hemp (Table 2), the cumulative emission represented only 0.28% of the total N accumulated in the crop residues, and these values were approximately half that observed by Rochette and Janzen (2005), who, in a similar study with this same legume, found an N₂O-N emission of 1.97 kg ha⁻¹ y⁻¹, representing 0.58% of the total N accumulated in crop residues. The low N₂O emissions in Brazilian soils are explained by the good drainage of most agricultural soils, with poor anaerobic activity (Jantalia *et al.*, 2008).

Despite N₂O emission peaks with Sunn hemp, the cumulative effect of the legume crop rotations throughout the year did not differ from other crops, as suggested by Peyrard *et al.* (2016), with the addition of plants of this type. Notably, the biological N fixation (BNF) by legumes does not influence the N₂O emission and is not counted as a source of N₂O to the atmosphere in the revised methodology of the IPCC (2006). Therefore, Sunn hemp is an important source of N in agricultural production systems, without impacting the GWP. Our results are in agreement with the approach taken in a recent review of the potential of cover crops to mitigate climate change, mainly due to the increase in SOC, and the reduction in the need for fertilizer use, especially after the cultivation of legumes as cover crops (Kaye and Quemada, 2017). Similarly, Bayer *et al.* (2016) found that under tropical NT with legumes as cover crops, the soil behaves as a sink for the GWP. According to the authors, CO₂ retention rates were approximately six times larger than those of the cumulative N₂O emissions. This outcome confirms our results in which, despite greater N₂O emission by Sunn hemp, no effect on the GWP was observed.

The GHGI values were similar to the values observed in other studies with soybean in the absence of nitrogen fertilization (Langeroodi *et al.*, 2019; Zhang *et al.*, 2017). The different N input from spring crop residues was expected to have a differential effect on GWP, as well as to influence the GHGI, but these effects were not observed. The low C N ratio of sunflower residues was the main driver for the higher GHG fluxes compared with triticale. Hence, crop rotations with triticale emitted a lower GHG to produce the same soybean yield (3.1 Mg ha⁻¹, data not shown), compared with sunflower. The low amount of crop residues in a dry fall–winter season associated with the fast decomposition of sunflower residues helps to explain the high GHGI (Corbeels *et al.*, 2000; Rigon *et al.*, 2018; Saviozzi *et al.*, 1995). According to Wang *et al.* (2004), the decomposition of residues occurs in two distinct phases. Initially, approximately 70% of the C is lost in the form of CO₂, followed by a slower phase, in which more recalcitrant compounds, depending on the residues, are decomposed. Thus, in tropical regions with dry winter, triticale would be a suitable option for SOC increase with low GHG emission in crop rotation under NT.

It is important to highlight the higher relative N₂O and CO₂ emissions under fallow than with the spring crops, regardless of the species. This result confirms the conclusions of a recent meta-analysis on N₂O emissions, which showed that cover crops have lower emissions compared with fallow.

According to Jeuffroy *et al.* (2013), legume crops emit around 5–7 times less GHG per unit area compared with other crops. The introduction of legumes in agricultural rotations helps in reducing the use of fertilizers and energy in arable systems and consequently lowers GHG emissions (Reckling *et al.*, 2014; Tongwane and Moeletsi, 2018). In addition, is important to consider the effects on the C retention and increase of SOC (Van Groenigen *et al.*, 2017; Wieder *et al.*, 2015). Cover crops in production systems are considered a key element in the reduction of C footprints due to the positive effect on SOC (Plaza-Bonilla *et al.*, 2018), in addition to being a potential CO₂ mitigation strategy.

Our findings show that plant residues with high N contents, such as pearl millet and Sunn hemp, even with occasional high short-term N₂O emissions, did not lead to an increase in GWP. It indicates that GHG emissions in agriculture should be relativized because these emissions can be compensated by proportional C inputs from crop residues, as observed in rotations with

triticale/pearl millet, where the C input exceeded GWP. Therefore, in assessing the efficiency of cropping systems as to GHG emissions, it should be always considered the relative emission, i.e. how much is emitted to produce 1 kg ha⁻¹ of grains. These results are important in enhancing the sustainability of agriculture in tropical regions.

Conclusion

Crop rotations under NT affect GHG emissions according to the species. The N₂O emissions increase with the use of legumes in the spring season, although they remain markedly below the standard IPCC emission factor. Growing cover crops in the spring, even for only 60 days, is essential to decrease the relative emissions of N₂O and CO₂, in addition to increasing SOC content on the soil surface compared to those under fallow. From the standpoint of conservation agriculture, triticale would be the best option for the fall–winter season because it increases not only SOC, but also the crop rotation efficiency with lower GHG emission without a penalty in soybean grain yields.

Acknowledgments. This work was supported by the São Paulo Research Foundation - FAPESP [grant number: 11/15361-3 and 11/02117-7].

Supplementary material. To view supplementary material for this article, please visit <https://doi.org/10.1017/S0014479721000156>.

References

- Bayer C., Gomes J., Zanatta J.A., Vieira F.C.B. and Dieckow J. (2016). Mitigating greenhouse gas emissions from a subtropical Ultisol by using long-term no-tillage in combination with legume cover crops. *Soil and Tillage Research* **161**, 86–94.
- Bell J.M., Smith J.L., Bailey V.L. and Bolton H. (2003). Priming effect and C storage in semi-arid no-till spring crop rotations. *Biology and Fertility of Soils* **37**(4), 237–244.
- Boddey R.M., Jantalia C.P., Conceição P.C., Zanatta J.A., Bayer C., Mielniczuk J., Dieckow J., Dos Santos H.P., Denardin J.E., Aita C., Giacomini S.J., Alves B.J.R. and Urquiaga S. (2010). Carbon accumulation at depth in Ferralsols under zero-till subtropical agriculture. *Global Change Biology* **16**(2), 784–795.
- Bolliger A., Magid J., Amado J.C.T., Skóra Neto F., Ribeiro M.de F. dos S., Calegari A., Ralisch R. and de Neergaard A. (2006). Taking stock of the Brazilian “Zero-Till Revolution”: a review of landmark research and farmers’ practice. *Advances in Agronomy* **91**(06), 47–110.
- Bowden R.D., Steudler P.A., Melilo J.M. and Aber J.D. (1990). Annual nitrous oxides fluxes from temperate forest soils in the Northeastern United States. *Journal of Geophysical Research* **95**, 13997–14005.
- Büchi L., Wendling M., Amossé C., Nepalova M. and Charles R. (2018). Importance of cover crops in alleviating negative effects of reduced soil tillage and promoting soil fertility in a winter wheat cropping system. *Agriculture, Ecosystems and Environment* **256**(December 2017), 92–104.
- Calonego J.C., Raphael J.P.A., Rigon J.P.G., Oliveira Neto L. de and Rosolem C.A. (2017). Soil compaction management and soybean yields with cover crops under no-till and occasional chiseling. *European Journal of Agronomy* **85**, 31–37.
- Castellano M.J., Mueller K.E., Olk D.C., Sawyer J.E. and Six J. (2015). Integrating plant litter quality, soil organic matter stabilization, and the carbon saturation concept. *Global Change Biology* **21**(9), 3200–3209.
- Castro G.S.A., Crusciol C.A.C., Calonego J.C. and Rosolem C.A. (2015). Management impacts on soil organic matter of tropical soils. *Vadose Zone Journal* **14**(1), 1–8.
- Chen B., Liu E., Tian Q., Yan C. and Zhang Y. (2014). Soil nitrogen dynamics and crop residues. A review. *Agronomy for Sustainable Development* **34**(2), 429–442.
- Chikowo R., Mapfumo P., Nyamugafata P. and Giller K.E. (2004). Mineral N dynamics, leaching and nitrous oxide losses under maize following two-year improved fallows on a sandy loam soil in Zimbabwe. *Plant and Soil* **259**(1–2), 315–330.
- Corbeels M., Hofman G. and Van Cleemput O. (2000). Nitrogen cycling associated with the decomposition of sunflower stalks and wheat straw in a Vertisol. *Plant and Soil* **218**(1–2), 71–82.
- Cotrufo M.F., Wallenstein M.D., Boot C.M., Deneff K. and Paul E. (2013). The Microbial Efficiency-Matrix Stabilization (MEMS) framework integrates plant litter decomposition with soil organic matter stabilization: Do labile plant inputs form stable soil organic matter? *Global Change Biology* **19**(4), 988–995.
- Cyle K.T., Hill N., Young K., Jenkins T., Hancock D., Schroeder P.A. and Thompson A. (2016). Substrate quality influences organic matter accumulation in the soil silt and clay fraction. *Soil Biology and Biochemistry* **103**, 138–148.

- del Río J.C., Gutiérrez A., Rodríguez I.M., Ibarra D. and Martínez Á.T. (2007). Composition of non-woody plant lignins and cinnamic acids by Py-GC/MS, Py/TMAH and FT-IR. *Journal of Analytical and Applied Pyrolysis*. <https://doi.org/10.1016/j.jaap.2006.09.003>
- Derpsch R., Franzluebbers A.J., Duiker S.W., Reicosky D.C., Koeller K., Friedrich T., Sturny W.G., Sá J.C.M. and Weiss K. (2014). Why do we need to standardize no-tillage research? *Soil and Tillage Research* **137**, 16–22.
- Ferrari Neto J., Franzluebbers A.J., Crusciol C.A.C., Rigon J.P.G., Calonego J.C., Rosolem C.A., do Nascimento C.A.C. and Ribeiro L.C. (2020). Soil carbon and nitrogen fractions and physical attributes affected by soil acidity amendments under no-till on Oxisol in Brazil. *Geoderma Regional* **24**, e00347.
- Frasier I., Quiroga A. and Noellemeyer E. (2016). Effect of different cover crops on C and N cycling in sorghum NT systems. *Science of the Total Environment* **562**, 628–639.
- García R.A., Li Y. and Rosolem C.A. (2013). Soil organic matter and physical attributes affected by crop rotation under no-till. *Soil Science Society of America Journal* **77**(5), 1724–1731.
- Gomes J., Bayer C., de Souza Costa F., de Cássia Piccolo M., Zanatta J.A., Vieira F.C.B. and Six J. (2009). Soil nitrous oxide emissions in long-term cover crops-based rotations under subtropical climate. *Soil and Tillage Research* **106**(1), 36–44.
- Gonzaga L.C., Carvalho J.L.N., Oliveira B.G. de, Soares J.R. and Cantarella H. (2018). Crop residue removal and nitrification inhibitor application as strategies to mitigate N₂O emissions in sugarcane fields. *Biomass and Bioenergy* **119**(September), 206–216.
- Hutchinson G.L. and Mosier A.R. (1981). Improved soil cover method for field measurement of nitrous oxide fluxes. *Soil Science Society of America Journal* **45**(2), 311–316.
- IPCC. (2006). Guidelines for national greenhouse gas inventories. Available at <https://www.ipcc-nggip.iges.or.jp/public/2006gl/index.html>
- IPCC. (2014). *Climate Change 2014: Synthesis Report. Contribution of Working Groups I, II and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change* (R. K. Pachauri & L. Meyer, Eds.). Ipcc, Geneva, Switzerland: IPCC.
- Jantalia C.P., Dos Santos H.P., Urquiaga S., Boddey R.M. and Alves B.J.R. (2008). Fluxes of nitrous oxide from soil under different crop rotations and tillage systems in the South of Brazil. *Nutrient Cycling in Agroecosystems* **82**(2), 161–173.
- Jantalia C.P., Resck D.V.S., Alves B.J.R., Zotarelli L., Urquiaga S. and Boddey R.M. (2007). Tillage effect on C stocks of a clayey Oxisol under a soybean-based crop rotation in the Brazilian Cerrado region. *Soil and Tillage Research* **95**(1–2), 97–109.
- Jeuffroy M.H., Baranger E., Carrouée B., De Chezelles E., Gosme M., Hénault C., Schneider A. and Cellier P. (2013). Nitrous oxide emissions from crop rotations including wheat, oilseed rape and dry peas. *Biogeosciences* **10**(3), 1787–1797.
- Kaye J.P. and Quemada M. (2017). Using cover crops to mitigate and adapt to climate change. A review. *Agronomy for Sustainable Development* **37**(1). <https://doi.org/10.1007/s13593-016-0410-x>
- Khalil M.I. and Baggs E.M. (2005). CH₄ oxidation and N₂O emissions at varied soil water-filled pore spaces and headspace CH₄ concentrations. *Soil Biology and Biochemistry*. <https://doi.org/10.1016/j.soilbio.2005.02.012>
- Kim D.S., Harazono Y., Baten M.A., Nagai H. and Tsuruta H. (2002). Surface flux measurements of CO₂ and N₂O from a dried rice paddy in Japan during a fallow winter season. *Journal of the Air and Waste Management Association* **52**(4), 416–422.
- Lal R. (2002). The potential of soils of the tropics to sequester carbon and mitigate the greenhouse effect. *Advances in Agronomy* **76**, 1–30.
- Lal R. (2004). Soil carbon sequestration impacts on global climate change and food security. *Science* **304**(5677), 1623–1627.
- Lal R. (2015). Sequestering carbon and increasing productivity by conservation agriculture. *Journal of Soil and Water Conservation* **70**(3), 55A–62A.
- Langeroodi A.S., Osipitan A. and Radicetti E. (2019). Benefits of sustainable management practices on mitigating greenhouse gas emissions in soybean crop (Glycine max). *Science of the Total Environment* **660**, 1593–1601.
- Le K.N., Jha M.K., Jeong J., Gassman P.W., Reyes M.R., Doro L., Tran D.Q. and Hok L. (2018). Evaluation of long-term SOC and crop productivity within conservation systems using GFDL CM2.1 and EPIC. *Sustainability (Switzerland)* **10**(8). <https://doi.org/10.3390/su10082665>
- Linn D.M. and Doran J.W. (1984). Effect of water-filled pore space on carbon dioxide and nitrous oxide production in tilled and nontilled soils. *Soil Science Society of America Journal* **48**(6), 1267–1272.
- MCTI. (2010). Emissões de Óxido Nitroso de Solos Agrícolas e de Manejo de Dejetos. Relatórios de Referência: Agricultura. 2o Inventário Brasileiro de Emissões e Remoções Antrópicas de Gases de Efeito Estufa, Brasília. Retrieved from https://cetesb.sp.gov.br/proclima/wp-content/uploads/sites/36/2014/05/brasil_mcti_solos_agricolas.pdf
- Mosier A.R., Halvorson A.D., Reule C.A. and Liu X.J. (2006). Net global warming potential and greenhouse gas intensity in irrigated cropping systems in Northeastern Colorado. *Journal of Environmental Quality* **35**(4), 1584–1598.
- Novelli L.E., Caviglia O.P. and Melchiori R.J.M. (2011). Impact of soybean cropping frequency on soil carbon storage in Mollisols and Vertisols. *Geoderma*. <https://doi.org/10.1016/j.geoderma.2011.09.015>
- Oertel C., Matschullat J., Zurba K., Zimmermann F. and Erasmi S. (2016). Greenhouse gas emissions from soils—A review. *Chemie Der Erde - Geochemistry* **76**(3), 327–352.

- Palm C.A., Gachengo C.N., Delve R.J., Cadisch G. and Giller K.E. (2001). Organic inputs for soil fertility management in tropical agroecosystems: Application of an organic resource database. *Agriculture, Ecosystems and Environment* **83**(1–2), 27–42.
- Peyrard C., Mary B., Perrin P., Véricel G., Gréhan E., Justes E. and Léonard J. (2016). N₂O emissions of low input cropping systems as affected by legume and cover crops use. *Agriculture, Ecosystems and Environment* **224**, 145–156.
- Pimentel L.G., Weiler D.A., Pedroso G.M. and Bayer C. (2015). Soil N₂O emissions following cover-crop residues application under two soil moisture conditions. *Journal of Plant Nutrition and Soil Science* **178**(4), 631–640.
- Plaza-Bonilla D., Álvaro-Fuentes J., Bareche J., Pareja-Sánchez E., Justes É. and Cantero-Martínez C. (2018). No-tillage reduces long-term yield-scaled soil nitrous oxide emissions in rainfed Mediterranean agroecosystems: A field and modeling approach. *Agriculture, Ecosystems and Environment* **262**(February), 36–47.
- Poepflau C. and Don A. (2015). Carbon sequestration in agricultural soils via cultivation of cover crops - A meta-analysis. *Agriculture, Ecosystems and Environment* **200**, 33–41.
- Powlson D.S., Stirling C.M., Jat M.L., Gerard B.G., Palm C.A., Sanchez P.A. and Cassman K.G. (2015). Reply to “No-till agriculture and climate change mitigation”. *Nature Climate Change* **5**(June), 489.
- Powlson D.S., Stirling C.M., Thierfelder C., White R.P. and Jat M.L. (2016). Does conservation agriculture deliver climate change mitigation through soil carbon sequestration in tropical agro-ecosystems? *Agriculture, Ecosystems and Environment* **220**, 164–174.
- Pugesgaard S., Petersen S.O., Chirinda N. and Olesen J.E. (2017). Crop residues as driver for N₂O emissions from a sandy loam soil. *Agricultural and Forest Meteorology* **233**, 45–54.
- Qiao N., Schaefer D., Blagodatskaya E., Zou X., Xu X. and Kuzyakov Y. (2014). Labile carbon retention compensates for CO₂ released by priming in forest soils. *Global Change Biology* **20**(6), 1943–1954.
- Qin S., Wang Y., Hu C., Oenema O., Li X., Zhang Y. and Dong W. (2012). Yield-scaled N₂O emissions in a winter wheat-summer corn double-cropping system. *Atmospheric Environment* **55**, 240–244.
- Raij B.V., Andrade J.C., Cantarella H. and Quaggio J.A. (2001). *Chemical analysis for evaluation of the fertility of tropical soils*, Campinas: Instituto Agronômico.
- Raphael J.P.A., Calonego J.C., Milori D.M.B.P. and Rosolem C.A. (2016). Soil organic matter in crop rotations under no-till. *Soil and Tillage Research* **155**, 45–53.
- Reckling M., Preissel S., Zander P., Topp K., Watson C., Murphy-Bokern D. and Stoddard F.L. (2014). Effects of legume cropping on farming and food systems. *Legume Futures Report* **1.6**(245216), 1–37.
- Rigon J.P.G. and Calonego J.C. (2020). Soil carbon fluxes and balances of crop rotations under long-term no-till. *Carbon Balance and Management* **15**(1), 19.
- Rigon J.P.G., Calonego J.C., Guimarães T.M. and Rosolem C.A. (2017). Critical periods of storage of the greenhouse gases in Polypropylene Syringe. *Communications in Soil Science and Plant Analysis* **48**(14), 1726–1732.
- Rigon J.P.G., Calonego J.C., Rosolem C.A. and Scala N.L. (2018). Cover crop rotations in no-till system: Short-term CO₂ emissions and soybean yield. *Scientia Agricola* **75**(1), 18–26.
- Rigon J.P.G., Calonego J.C.C., Capuani S. and Franzluebbbers A. (2021). Soil organic C affected by dry - season management of no - till soybean crop rotations in the tropics. *Plant and Soil* **462**, 577–590.
- Rigon J.P.G., Franzluebbbers A.J. and Calonego J.C. (2020). Soil aggregation and potential carbon and nitrogen mineralization with cover crops under tropical no-till. *Journal of Soil and Water Conservation* **75**(5), 601–609.
- Rochette P. and Janzen H.H. (2005). Towards a revised coefficient for estimating N₂O emissions from legumes. *Nutrient Cycling in Agroecosystems* **73**(2–3), 171–179.
- Sainju U.M., Caesar-TonThat T., Lenssen, A.W. and Barsotti J.L. (2012). Dryland soil greenhouse gas emissions affected by cropping sequence and nitrogen fertilization. *Soil Science Society of America Journal* **76**(5), 1741–1757.
- Sanchez P.A. and Logan T.J. (1992). Myths and Science about the Chemistry and Fertility of Soils in the Tropics, (29). <https://doi.org/10.2136/sssaspecpub29.c3>
- Sant’Anna S.A.C., Martins M.R., Goulart J.M., Araújo S.N., Araújo E.S., Zaman M., Jantalia C.P., Alves B.J.R., Boddey R.M. and Urquiaga S. (2018). Biological nitrogen fixation and soil N₂O emissions from legume residues in an Acrisol in SE Brazil. *Geoderma Regional*, **15**, e00196.
- SAS, Inc. (2009). The SAS System for Windows, Cary, NC.
- Saviozzi A., Scagnozzi A. and Riffaldi R. (1995). Decomposition of crop residues under laboratory conditions. *Soil Use and Management* **11**, 193–198.
- Schipanski M.E., Barbercheck M., Douglas M.R., Finney D.M., Haider K., Kaye J.P., Kemanian A.R., Mortensen D.A., Ryan M.R., Tooker J. and White C. (2014). A framework for evaluating ecosystem services provided by cover crops in agroecosystems. *Agricultural Systems* **125**, 12–22.
- Shahbaz M., Kuzyakov Y., Sanaullah M., Heitkamp F., Zelenev V., Kumar A. and Blagodatskaya E. (2017). Microbial decomposition of soil organic matter is mediated by quality and quantity of crop residues: mechanisms and thresholds. *Biology and Fertility of Soils* **53**(3), 287–301.
- Silva D.J. and Queiroz A.C. (2002). *Food analysis: Chemical and biological methods* (UFV, Ed.), 3rd Edn, Viçosa: UFV.

- Šimek M., Elhottová D., Klimeš F. and Hopkins D.W. (2004). Emissions of N₂O and CO₂, denitrification measurements and soil properties in red clover and ryegrass stands. *Soil Biology and Biochemistry* **36**(1), 9–21.
- Sisti C.P.J., Dos Santos H.P., Kohhann R., Alves B.J.R., Urquiaga S. and Boddey R.M. (2004). Change in carbon and nitrogen stocks in soil under 13 years of conventional or zero tillage in southern Brazil. *Soil and Tillage Research*. <https://doi.org/10.1016/j.still.2003.08.007>
- Six J., Conant R.T., Paul E.A. and Paustian K. (2002). Stabilization of organic matter by soil minerals: Implications for C-saturation of soils. *Plant and Soil* **241**, 155–176.
- Smith K.A. and Mullins E.C. (1991). *Soil Analysis: Physical methods*, New York: Marcel Dekker, Inc.
- Soil Survey Staff. (2014). *Keys to Soil Taxonomy*, 12th Edn, Washington, DC.: USDA-Natural Resources Conservation Service.
- Tongwane M.I. and Moeletsi M.E. (2018). A review of greenhouse gas emissions from the agriculture sector in Africa. *Agricultural Systems* **166**(June), 124–134.
- Tonitto C., David M.B. and Drinkwater L.E. (2006). Replacing bare fallows with cover crops in fertilizer-intensive cropping systems: A meta-analysis of crop yield and N dynamics. *Agriculture, Ecosystems and Environment* **112**(1), 58–72.
- Van Groenigen J.W., Van Kessel C., Hungate B.A., Oenema O., Powlson D.S. and Van Groenigen K.J. (2017). Sequestering Soil Organic Carbon: A Nitrogen Dilemma. *Environmental Science and Technology* **51**(9), 4738–4739.
- Wang W.J., Baldock J.A., Dalal R.C. and Moody P.W. (2004). Decomposition dynamics of plant materials in relation to nitrogen availability and biochemistry determined by NMR and wet-chemical analysis. *Soil Biology and Biochemistry* **36**(12), 2045–2058.
- Wieder W.R., Cleveland C.C., Smith W.K. and Todd-Brown K. (2015). Future productivity and carbon storage limited by terrestrial nutrient availability. *Nature Geoscience* **8**(6), 441–444.
- Yang Y., Huang Q., Yu H., Song K., Ma J., Xu H. and Zhang G. (2018). Winter tillage with the incorporation of stubble reduces the net global warming potential and greenhouse gas intensity of double-cropping rice fields. *Soil and Tillage Research* **183**(February), 19–27.
- Zhang X., Zhang J., Zheng C., Guan D., Li S., Xie F., Chen J., Hang X., Jiang Y., Deng A., Afreh D. and Zhang W. (2017). Significant residual effects of wheat fertilization on greenhouse gas emissions in succeeding soybean growing season. *Soil and Tillage Research* **169**, 7–15.
- Zhao Y., Zhang J., Müller C. and Cai Z. (2018). Temporal variations of crop residue effects on soil N transformation depend on soil properties as well as residue qualities. *Biology and Fertility of Soils* **54**(5), 659–669.