

# Patterns of trunk spine growth in two congeneric species of acanthocephalan: investment in attachment may differ between sexes and species

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## SUMMARY

Acanthocephalans have evolved a hooked proboscis and some taxa have trunk spines to attach to their definitive hosts. These structures are generated before being used, thus a key question is how investment in attachment could optimally be allocated through the ontogeny. The number and arrangement of hooks and spines are never modified in the definitive host, but it is unclear whether these structures grow during adult development. A comparison of the size of trunk spines between cystacanths and adults of *Corynosoma cetaceum* and *C. australe* indicated that spines grow in both species, but only in females, which also had significantly larger spines than males. This sexual dimorphism did not result from pure allometry because the body of females was smaller, and did not grow more than that of males. However, having a longer lifespan, females would need to withstand the extreme flow conditions prevailing in marine mammals for longer, inducing different investment and development schedules for spines. Patterns of spine growth also differed between species: fore-trunk spines grew in both species, but hind-trunk spines did only in *C. cetaceum*. In conclusion, investment strategies on attachment may differ, not only between congeneric species of acanthocephalan, but also between sexes of the same species.

**Key words:** Acanthocephala, *Corynosoma australe*, *Corynosoma cetaceum*, trunk spine, investment strategy, ontogeny, attachment.

## INTRODUCTION

Parasites have evolved a wide array of holdfast mechanisms that maximize the likelihood of successful attachment upon recruitment to their hosts and minimize the risk of subsequent dislodgment (Poulin, 2009; Randhawa and Poulin, 2010). Selective pressures on morphology are especially strong in parasites living in the lumen of the gastrointestinal tract, where physical disturbance in the form of peristalsis and food movement can exert powerful drag on attached parasites (Poulin, 2009). Acanthocephalans, in particular, have developed a proboscis armed with hooks that anchor to the gut of their definitive host (Taraschewski, 2000). Many species also have trunk spines that engage on the gut surface, sometimes playing a significant role in attachment (Van Cleave, 1952; Aznar *et al.* 1999a,

2002a). It has been argued that investment in these primary holdfast structures is optimized for the species of host and the particular microhabitat where each species of acanthocephalan lives (Poulin, 2007). A possible reason is that attachment structures are costly to produce and, therefore, it would not be advantageous for a worm to produce them larger than the size necessary to ensure attachment (Poulin, 2007). Also, depending on the size of the animal, the size of holdfast structures should also be bounded within certain limits to ensure that attachment performance is functional (Van Cleave, 1952; see also Koehl, 1996).

Interestingly, both the proboscis and trunk spines of acanthocephalans are generated prior to being used for attachment, and this raises the question of how investment in such structures could optimally be allocated through ontogeny. The first larval stage, the acanthor, hatches from the egg and passes through 2 subsequent stages, the acanthella and the cystacanth, within an intermediate arthropod host; many acanthocephalans may also use a paratenic host (usually a vertebrate) in which the cystacanth gets

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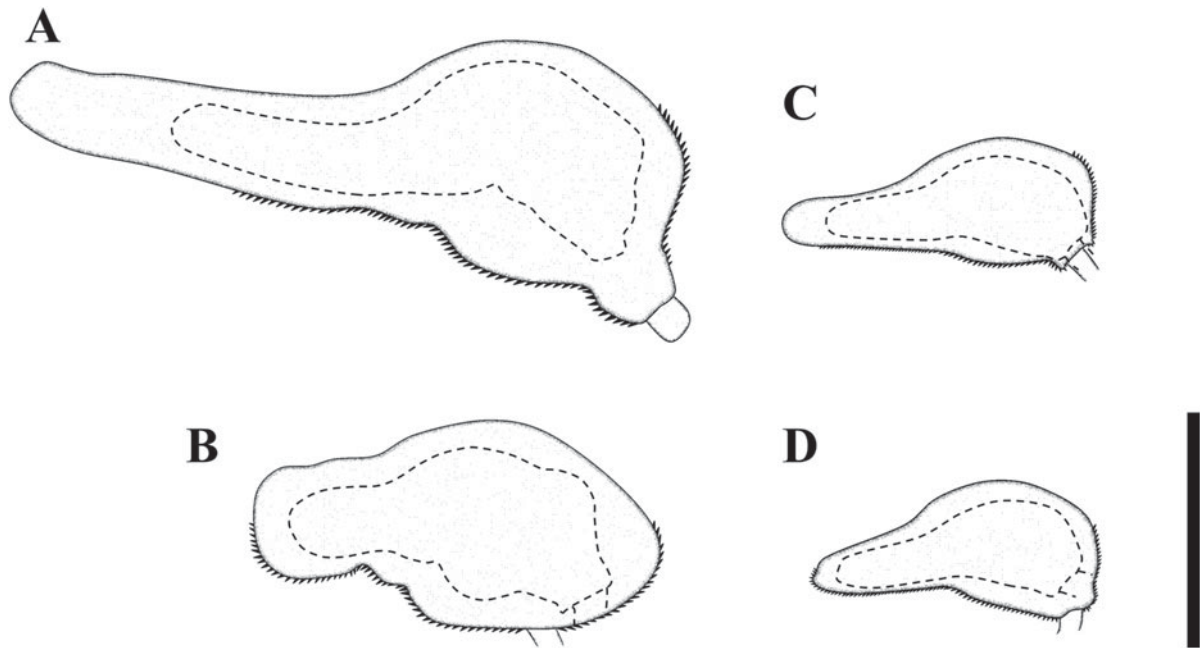


Fig. 1. Diagrammatic comparison of the body size and spine coverage in two species of *Corynosoma*. (A) Male *Corynosoma cetaceum*, (B) Female *C. cetaceum*, (C) Male *C. australe*, (D) Female *C. australe*. Dashed lines indicate the relative body size of cystacanths. Scale bar = 2 mm.

encysted in the mesentery without further development (Schmidt, 1985). The cystacanth is the infective stage that is consumed by the definitive vertebrate host and already has all the primary attachment structures of the adult. Van Cleave (1952) and Petrochenko (1956) suggested that, in most species, attachment structures are fully formed at the cystacanth stage, perhaps as an investment priority of the developing worm to secure successful establishment upon arrival to the definitive host. Indeed, to the best of our knowledge, the number and arrangement of hooks in the proboscis and the extension of spines on the trunk are never modified in the definitive host (Van Cleave, 1952). However, the extent to which the proboscis, proboscis hooks, and trunk spines grow during the adult development is an open question. Some authors reported no changes in the size of proboscis and/or proboscis hooks between cystacanths and adults of some species (Podesta and Holmes, 1970; Amin *et al.* 1995, 2004). Other authors, however, noted an increase in the size of proboscis hooks or trunk spines in adults of different species compared to cystacanths (Podesta and Holmes, 1970; Amin *et al.* 1995), or juveniles *i.e.* recently recruited worms in the definitive host (Amin, 1986, 1987).

In any of the above studies it is difficult to separate the putative growth of the holdfast from measurement error because none used inferential statistics. However, it seems likely that the timing of growth of attachment structures may differ among species of acanthocephalans depending on their body size. Adult acanthocephalans are subject to the unsteady flow of digested food generated by peristalsis (Poulin, 2007).

Although the physical properties of the flow of digesta are far from clear (see Schulze, 2006), acanthocephalans are theoretically expected to experience 3 types of dislodging forces *i.e.* frictional drag, pressure drag, and acceleration reaction, which are proportional to the surface area, sectional area, and volume of the body, respectively (see Koehl, 1984, for details). Thus, everything else being equal, dislodging forces should increase disproportionately as the body grows, and larger acanthocephalans could therefore need a finer adjustment of their holdfast structures during the adult growth, particularly if they experience a greater change of body size from the cystacanth to the adult stage (see Poulin *et al.* 2003).

In this study we compared the size of trunk spines between cystacanths and adults of 2 congeneric species of acanthocephalans from the Southern Hemisphere that clearly differ in size, namely *Corynosoma cetaceum* and *C. australe* (Fig. 1). Individuals of *C. cetaceum* inhabit the stomach and upper duodenum of small cetaceans, whereas *C. australe* is found in the intestine, mainly in the ileum and jejunum, of pinnipeds (Aznar *et al.* 2001, 2004, 2012; Sardella *et al.* 2005). We focused on trunk spines because they play a key role in the attachment of species of *Corynosoma* (Van Cleave, 1952; Aznar *et al.* 1999a; 2001) and can be measured in any specimen; the proboscis is rarely found fully evaginated in adult specimens, and cannot be induced to withdraw because worms are collected dead from hosts. The goals of our study were 2-fold. First, we obtained, for the first time, statistical evidence on whether spines grow during the adult development

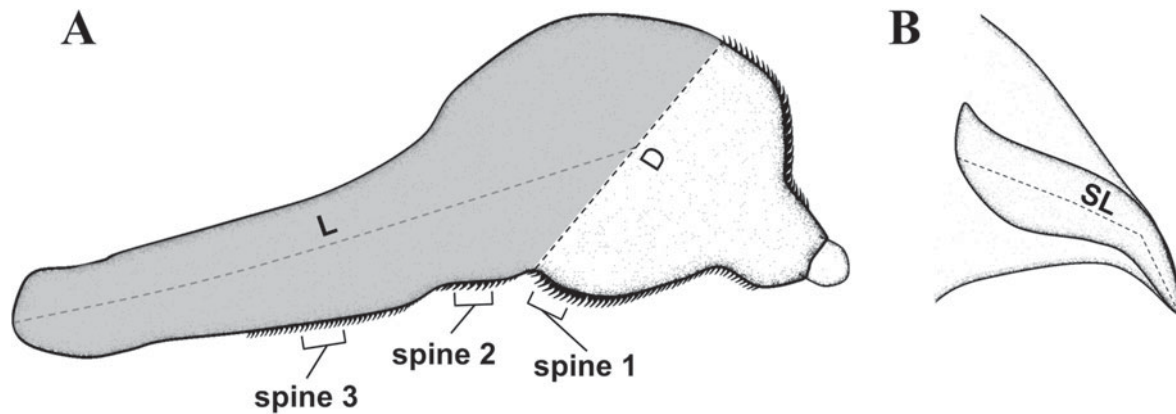


Fig. 2. Morphometric measurements taken in specimens of *Corynosoma cetaceum* and *C. australe*. L, trunk length; D, disk diameter; SL, spine length. The shadowed area is sectional area. Regions where spines were measured are also indicated (see Materials and Methods section for details).

of an acanthocephalan. Second, we investigated the factors that may account for patterns of spine growth, including body size.

#### MATERIALS AND METHODS

##### Data collection

Specimens of *Corynosoma cetaceum* were collected in several localities along the coast of Argentina. Cystacanths (20 females and 26 males) were obtained from the mesentery of 2 individuals of Argentine sandperch *Pseudoperca semifasciata* in the neighbourhood of Península Valdés (42°00'–42°45'S). Adults (43 females and 42 males) were collected from the pyloric stomach of 5 franciscana dolphins, *Pontoporia blainvillei*, that were found drowned in shark fishery gillnets in Necochea (38°27'S, 58°50'W) and Claromecó (38°52'S, 60°05'W). Sampling of *Corynosoma australe* was conducted in the north coast of Patagonia (42°45'S, 62°30'W): cystacanths (33 females and 24 males) were collected from the mesentery of 11 individuals of the flounder *Paralichthys isosceles*, whereas adults (35 females and 35 males) were collected from the intestine of 3 South American sea lions, *Otaria flavescens* stranded on Patagonian beaches. Acanthocephalan specimens were generally washed in saline and fixed and conserved in 70% ethanol. Cystacanths of *C. cetaceum* were fixed in 4% buffered formaldehyde and preserved in 70% ethanol. No significant morphometric differences were found between cystacanths fixed in ethanol or formaldehyde (MANOVA,  $P \gg 0.05$ ).

Acanthocephalans were examined under a stereomicroscope (X100) and identified following the taxonomic criteria of Aznar *et al.* (1999b) and Sardella *et al.* (2005). Then, each specimen was drawn in profile with the aid of a drawing tube (Fig. 2). Trunk length (L) and disk diameter (D) were measured using homologous landmarks that

were unaffected by the degree of fore-trunk invagination (Fig. 2). Four body size variables directly related to attachment performance were obtained from each specimen as follows. (1) Disk area. In species of *Corynosoma*, the disk covered with spines is used as a key attachment device (Van Cleave, 1952; Aznar *et al.* 1999a, 2006). The disk surface is roughly circular, thus its area was estimated as the area of a circle. (2) Sectional area (Fig. 2). This variable is related to pressure drag (Koehl, 1984). To obtain it, the drawing in profile of each specimen was scanned and the area was calculated using Image Tool v. 3.0 (UTHSCSA). (3) Surface area. This variable is related to skin friction drag (Koehl, 1984). The body of species of *Corynosoma* can faithfully be reproduced just by bending a cone (Aznar *et al.* unpublished data; see Fig. 1). Therefore, surface area can be approximated using the formula for a cone surface, without considering the area of the disk (the disk is attached to the intestine so it is not exposed to drag). (4) Body volume. This variable is related to 'virtual buoyancy', a lifting force proportional to the mass of fluid displaced by the body (Koehl, 1984). Volume was calculated assuming a conical body shape.

To measure spines, each specimen was cut with a razor blade through the sagittal plane and one half was temporarily mounted on a slide with lactic acid to clear the tegument. Using this procedure, specimens could be re-accommodated, if necessary, for spines to be drawn in profile minimizing tilt-related error. Three spines were drawn under a light microscope (X1000) from each of the 3 sites indicated in Fig. 2 i.e. the disk border, the interfold area, and the posterior hind-trunk (see Aznar *et al.* 2002a for details). For brevity, we will refer to the spines from these sites as Spines 1, 2 and 3, respectively. Spine length was measured as indicated in Fig. 2, and the values taken from 3 spines randomly selected from each site were averaged to obtain a single value per site and specimen.

Statistical analyses

A preliminary analysis indicated that the factor ‘host individual’ did not have a significant effect on average values of morphometric variables either in paratenic or definitive hosts (MANOVA,  $P > 0.05$  in all 4 tests), thus, this factor was not considered in further analyses.

The effect of developmental stage, sex, and species on body size variables was examined with MANOVA, using disk area, sectional area, surface area and volume as dependent variables. The 3 factors were considered as fixed. Concerning the ‘species’ effect, we were specifically interested in the interaction of ‘species’ with ‘developmental stage’ and ‘sex’ because this analysis allowed investigation of whether patterns of body growth differed between species, a point that was relevant for the interspecific differences observed in spine growth (see the Results section).

Multivariate analysis of covariance (MANCOVA) was used to examine patterns of spine growth within each species. Values of Spines 1, 2 and 3 were treated as dependent variables and ‘developmental stage’ and ‘sex’ as fixed factors. In addition, we used principal component analysis on the 4 body variables to obtain scores on the first axis (PC1) i.e. a multivariate measure of body size (Klingerberg, 1996). The scores in PC1 were then included in the model as a co-variate. The inclusion of PC1 is pertinent to explore the relationships between static and ontogenetic allometry in spine size growth (see Klingerberg, 1996). Static allometry results from co-variation between morphometric traits among individuals of the same age or developmental stage; in our case cystacanths or adults (Fig. 3A). Ontogenetic allometry deals with co-variation between morphometric traits during growth i.e. the population of cystacanths and adults considered as a whole (Fig. 3A). Both allometric patterns are usually, but not necessarily, similar (Cock, 1966; Klingerberg, 1996). In our model, the way to compare allometric patterns was by examining the interaction between PC1 and developmental stage: if the interaction was significant, this would mean that static and ontogenetic allometries did not coincide. In other words, the relationship between body size and spine size would differ between cystacanths and adults, thus indicating changes in relative growth rate during the adult development in the definitive host (Fig. 3B, C). When interaction terms with the co-variate were not significant, they were removed from models to increase the sensitivity of the analysis and to correctly interpret main effects (Engqvist, 2005).

MANCOVA models were also used to explore whether variability in spine size within sites (i.e. the disk border, the interfold area, and the posterior hind-trunk) differed between sexes and developmental stages; PC1 was used as a co-variate. For each

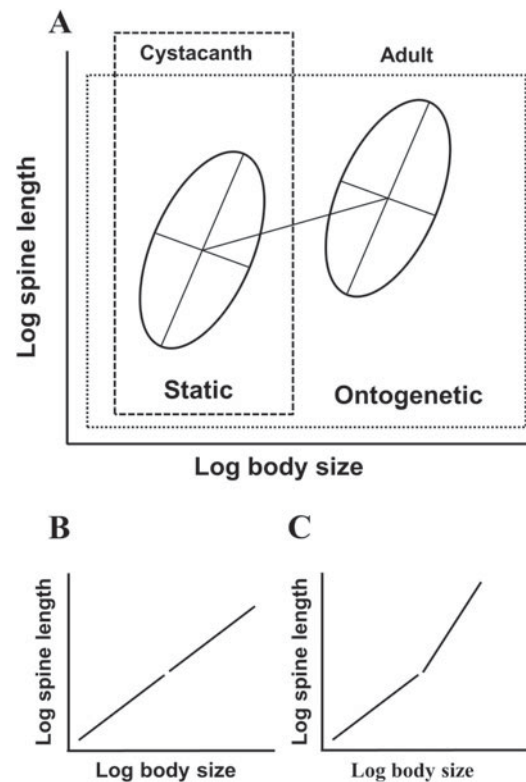


Fig. 3. Theoretical relationships between static and ontogenetic allometry. (A) Levels of co-variation between spine length and body size in 2 developmental stages of an acanthocephalan i.e. cystacanth and adult (redrawn from Klingerberg, 1996). Static allometry (dashed rectangle) refers to co-variation among individuals of the same developmental stage (e.g. cystacanth). Ontogenetic allometry (dotted rectangle) refers to co-variation due to growth from the cystacanth to the adult stage. (B) Hypothetical relationship between static and ontogenetic allometry in which relative growth rate do not change between the cystacanth and the adult stage. (C) Hypothetical relationship between static and ontogenetic allometry in which both levels of allometry differ because the relative growth of spines changes during the adult development.

specimen, the coefficient of variation (CV) of each set of 3 spines was calculated (i.e. for Spines 1, 2 and 3). These CVs were treated as dependent variables in the MANCOVA models.

Statistical analyses were carried out with SPSS v. 17. Statistical significance was set at  $P < 0.05$ .

RESULTS

Patterns of body growth

Data on morphometric variables are shown in Table 1. In *C. cetaceum*, highly significant differences were found in body dimensions, not only between developmental stages, but also between sexes. Also, a highly significant interaction ‘developmental stage \* sex’ was observed (Table 2). Univariate ANOVAs revealed that disk area did not differ

Table 1. Mean values (s.d.) [Coefficient of Variation] of body dimensions and length of trunk spines in cystacanth and adult specimens of the acanthocephalans *Corynosoma cetaceum* and *C. australe*

(Measurements in mm (trunk length and disk diameter), mm<sup>2</sup> (disk, sectional and surface areas), mm<sup>3</sup> (volume) and μm (spine length).)

Species	Group	N	Body dimensions						Spine length		
			Trunk length	Disk diameter	Disk area	Sectional area	Surface area	Volume	Spine 1	Spine 2	Spine 3
<i>C. cetaceum</i>	Cystacanth female	20	1.98 (0.23) [0.12]	1.43 (0.15) [0.10]	1.63 (0.32) [0.20]	1.58 (0.36) [0.23]	3.15 (0.60) [0.19]	1.09 (0.28) [0.26]	60.0 (5.5) [0.09]	53.5 (5.1) [0.09]	49.8 (4.2) [0.08]
	Adult female	43	2.86 (0.37) [0.13]	2.20 (0.36) [0.16]	3.91 (1.32) [0.34]	4.18 (1.20) [0.28]	6.90 (1.93) [0.28]	3.87 (1.85) [0.48]	66.5 (4.6) [0.07]	63.4 (5.5) [0.09]	56.0 (5.8) [0.10]
	Cystacanth male	26	2.55 (0.42) [0.16]	1.33 (0.15) [0.12]	1.42 (0.33) [0.23]	1.96 (0.45) [0.23]	4.15 (0.97) [0.23]	1.21 (0.37) [0.30]	60.1 (3.1) [0.05]	55.1 (3.4) [0.06]	50.6 (3.6) [0.07]
	Adult male	42	5.02 (1.01) [0.20]	2.21 (0.35) [0.16]	3.91 (1.25) [0.32]	5.34 (2.10) [0.40]	14.4 (5.22) [0.36]	6.93 (3.54) [0.51]	59.2 (5.3) [0.09]	54.6 (5.8) [0.11]	49.7 (5.5) [0.11]
<i>C. australe</i>	Cystacanth female	35	1.51 (0.27) [0.17]	0.91 (0.14) [0.16]	0.67 (0.19) [0.30]	0.69 (0.14) [0.20]	1.61 (0.42) [0.26]	0.73 (0.18) [0.24]	46.6 (3.7) [0.09]	37.8 (3.3) [0.09]	40.5 (3.8) [0.10]
	Adult female	34	1.99 (0.31) [0.17]	1.33 (1.17) [0.15]	1.40 (0.33) [0.30]	1.47 (0.40) [0.27]	3.03 (0.80) [0.29]	1.40 (0.35) [0.28]	49.1 (3.0) [0.08]	37.8 (3.3) [0.11]	40.4 (4.0) [0.11]
	Cystacanth male	23	1.49 (0.20) [0.13]	0.95 (0.08) [0.09]	0.70 (0.13) [0.19]	0.66 (0.11) [0.16]	1.62 (0.30) [0.19]	0.74 (0.12) [0.17]	47.0 (2.9) [0.06]	36.3 (2.5) [0.07]	38.3 (3.5) [0.13]
	Adult male	33	2.06 (0.38) [0.19]	1.30 (0.20) [0.16]	1.36 (0.40) [0.30]	1.39 (0.43) [0.33]	3.15 (0.99) [0.33]	1.43 (0.44) [0.32]	47.8 (3.0) [0.09]	36.0 (2.5) [0.07]	37.1 (2.3) [0.06]

Table 2. Results from a multivariate analysis of variance that examines the effects of sex and developmental stage (cystacanth and adult) on 4 body variables i.e. disk area, sectional area, surface area and volume in the acanthocephalans *Corynosoma cetaceum* and *C. australe*

(Statistically significant effects are in bold.)

Factor	D.F.	Wilks' lambda	F	P
<i>C. cetaceum</i>				
Stage	4	0.267	85.134	<b>&lt;0.001</b>
Sex	4	0.201	123.256	<b>&lt;0.001</b>
Stage * Sex	4	0.536	26.829	<b>&lt;0.001</b>
Error	124			
<i>C. australe</i>				
Stage	4	0.318	63.152	<b>&lt;0.001</b>
Sex	4	0.863	4.674	<b>0.002</b>
Stage * Sex	4	0.962	1.157	0.333
Error	118			

between sexes ( $F_{(1,127)} = 1.864$ ,  $P = 0.175$ ), but males had a significantly larger sectional area ( $F_{(1,127)} = 14.427$ ,  $P < 0.001$ ), surface area ( $F_{(1,127)} = 81.202$ ,  $P < 0.001$ ) and body volume ( $F_{(1,127)} = 17.469$ ,  $P < 0.001$ ) than females (Fig. 4). Significant univariate differences concerned surface area and volume (interaction 'developmental stage \* sex': surface area,  $F_{(1,127)} = 16.607$ ,  $P < 0.001$ ; body volume,  $F_{(1,127)} = 8.287$ ,  $P < 0.005$ ). These variables grew comparatively faster in males than in females (Fig. 4). In *C. australe*, significant differences in body dimensions were also found between developmental stages and sexes (Table 2). However, sexual dimorphism was slight because none of the univariate ANOVAs was found to be significant (minimum nominal  $P = 0.221$ ) (Fig. 4). Also, the interaction 'developmental stage \* sex' was not significant (Table 2).

As an interspecific comparison, we tested whether the relative amount of growth from cystacanth to adult differed between *C. cetaceum* and *C. australe*. In males, the multivariate interaction 'developmental stage \* species' was highly significant (Wilks' Lambda = 0.279,  $F_{(4,117)} = 75.556$ ,  $P < 0.001$ ), as were interactions of these factors for each dependent variable ( $P < 0.001$ ). Average relative size of adult males compared to cystacanths was as follows (*C. cetaceum* vs *C. australe*): disk area: 175% vs 94%; sectional area: 172% vs 111%; surface area: 247% vs 94%; and volume: 473% vs 93% (see the Table 1). In females, a highly significant interaction 'developmental stage \* species' was also detected (Wilks' Lambda = 0.354,  $F_{(4,125)} = 57.031$ ,  $P < 0.001$ ), but significant univariate differences concerned sectional area and volume only (sectional area,  $F_{(1,128)} = 6.500$ ,  $P < 0.012$ ; volume,  $F_{(1,128)} = 22.603$ ,  $P < 0.001$ ). Sectional area and volume in adult females of *C. cetaceum* increased 164% and 255%, respectively,

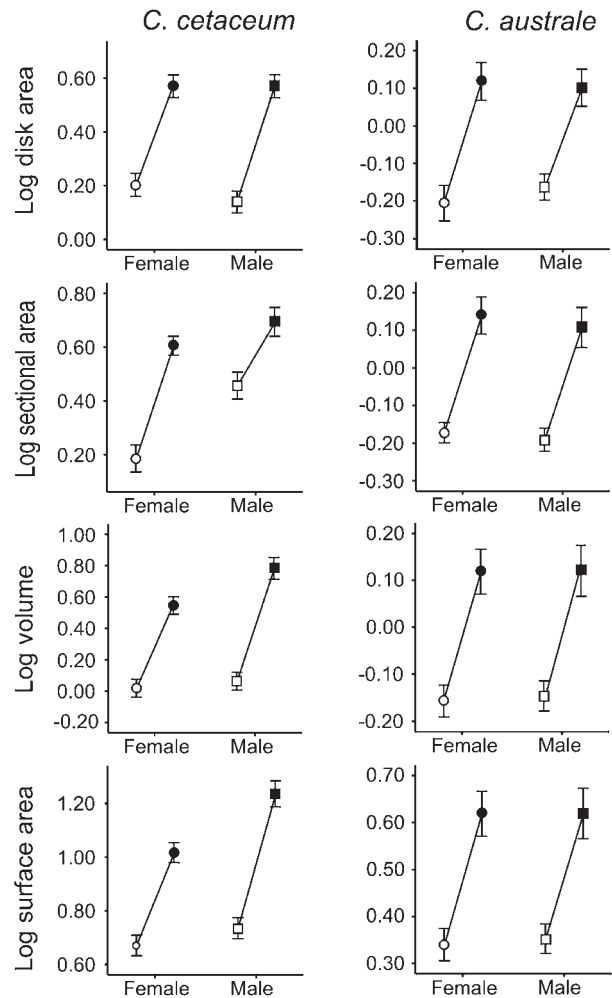


Fig. 4. Mean values (bars: standard error) of 4 body size variables in cystacanths (open symbols) and adults (solid symbols) of individuals from each sex of *Corynosoma cetaceum* and *C. australe*.

compared to cystacanths; however, in *C. australe*, these figures were just 113% and 91% (see the Table 1). In summary, during adult development individuals of *C. cetaceum* grew comparatively more than those of *C. australe*.

Patterns of spine growth

Individuals of *C. cetaceum* had larger spines than those of *C. australe* regardless of developmental stage and sex (Fig. 5; see also the Table 1).

In *C. cetaceum*, spine length significantly differed between developmental stages and sexes, and the overall relationship between spine size and body size was not significant (Table 3). However, a highly significant interaction 'developmental stage \* sex' was found and, therefore, analyses were carried out for each sex separately to tear apart the effects of developmental stage (ontogenetic allometry) and body size (static allometry). In females, the full factorial MANCOVA indicated that adults had longer spines than cystacanths (Fig. 5A) but neither

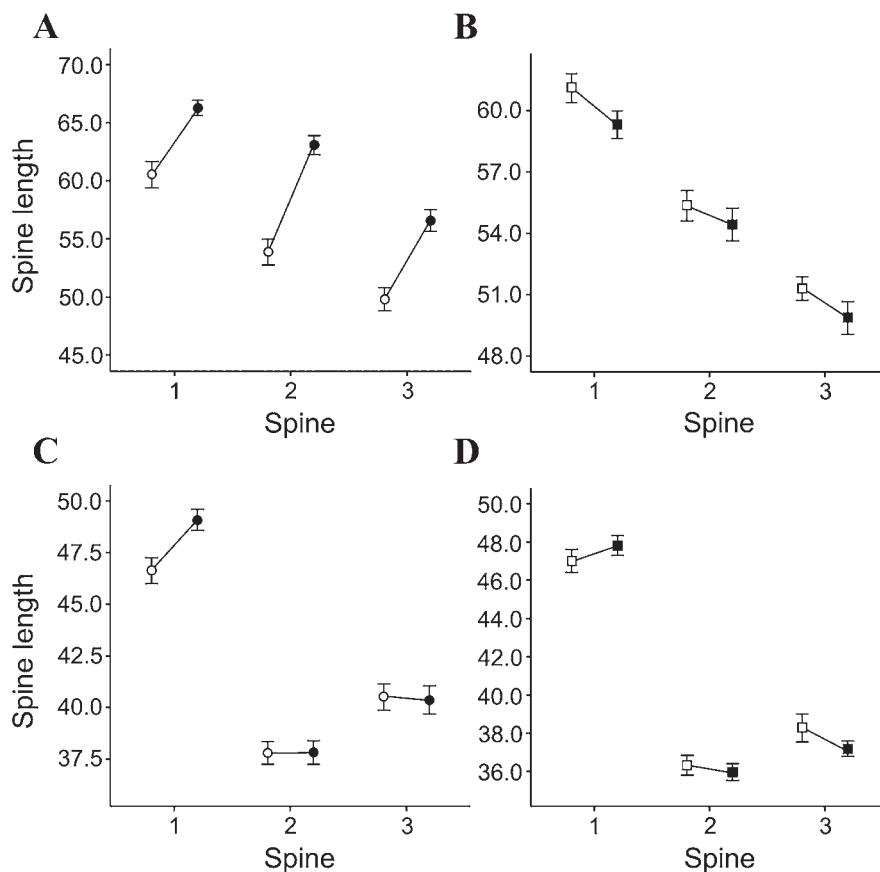


Fig. 5. Mean length (bar: standard error) of spines measured at 3 sites (see Fig. 2) in cystacanths (open symbols) and adults (solid symbols) of both sexes in 2 species of *Corynosoma*. (A) Female *C. cetaceum*; (B) Male *C. cetaceum*; (C) Female *C. australe*; (D) male *C. australe*.

the overall effect of PC1 on spine size nor the interaction 'developmental stage \*PC1' were significant (Table 3). After removing the interaction term, a significant main effect of PC1 was found (Table 3). Univariate ANOVAs indicated that PC1 correlated significantly ( $P < 0.05$ ) with spine length only in Spines 2 and 3 (Fig. 6). In males of *C. cetaceum*, there was no significant difference in spine length between cystacanths and adults, nor was there any indication of a significant relationship between body size and spine size in cystacanths or adults (Table 3, Fig. 5B). In summary, (1) females of *C. cetaceum* had longer spines than males; (2) all spines were longer in adults, but only in females, and (3) there was a significant relationship between spine length and body size only in females (both cystacanths and adults), and only for Spines 2 and 3 (hind-trunk spines).

In *C. australe*, spine length significantly differed between both developmental stages and sexes (Table 4). Again, a significant interaction 'developmental stage \* sex' was found and, therefore, separate analyses were performed for each sex. In females, spine length differed between cystacanths and adults (Table 4); the univariate ANOVAs revealed that only Spine 1 was significantly larger in adults ( $P = 0.003$ ) (Fig. 5C). However, the effect of PC1 on spine size

was not significant, even after removing the interaction 'developmental stage \*PC1' in the model (Table 4). In males, none of the predictors of spine length was significant in any model (Table 4; Fig. 5D). In summary, (1) females of *C. australe* had longer spines than males; (2) disk spines (Spine 1) were longer in adults than in cystacanths, but only in females, and (3) there was no significant pattern of static allometry between spine length and body size in either sex or developmental stage.

None of the MANCOVA models for each species involving CVs of Spines 1, 2 and 3 revealed significant effects of sex or developmental stage on spine variability; an overall MANOVA using 'species' as a single factor also did not (results not shown).

## DISCUSSION

Results from this study provide, for the first time, statistical evidence that trunk spines of 2 species of acanthocephalan grow during the worm development in the definitive host. Unexpectedly, spines appear to grow only in females and exhibit a different pattern of growth depending on the species. A preliminary question that must be addressed is whether there are

Table 3. Models of multivariate analysis of covariance that examine the effects of developmental stage (cystacanth and adult), sex, and a multivariate measure of body size (PC1, the first principal component of the 4 morphometric variables indicated in Table 1) on the length of trunk spines from 3 sites in the acanthocephalan *Corynosoma cetaceum*

(Statistically significant effects are in bold.)

Factor	D.F.	Wilks' lambda	F	P
<i>Full factorial model</i>				
Stage	3	0.832	8.119	<b>&lt;0.001</b>
Sex	3	0.843	7.496	<b>&lt;0.001</b>
PC1	3	0.970	1.253	0.294
Stage * Sex	3	0.742	14.040	<b>&lt;0.001</b>
Stage * PC1	3	0.986	0.552	0.647
Sex * PCA	3	0.985	0.634	0.595
Stage * Sex * PC1	3	0.968	1.329	0.268
Error	121			
Females				
<i>Full factorial model</i>				
Stage	3	0.478	20.745	<b>&lt;0.001</b>
PC1	3	0.924	1.566	0.208
Stage * PC1	3	0.936	1.301	0.283
Error	157			
<i>Main effects model</i>				
Stage	3	0.484	20.639	<b>&lt;0.001</b>
PC1	3	0.854	3.151	<b>0.031</b>
Error	58			
Males				
<i>Full factorial model</i>				
Stage	3	0.962	0.806	0.495
PC1	3	0.987	0.267	0.849
Stage * PC1	3	0.974	0.550	0.650
Error	62			
<i>Main effects model</i>				
Stage	3	0.963	0.803	0.497
PC1	3	0.978	0.483	0.697
Error	63			

sampling and/or measurement artifacts that could confound these results. First, cystacanths and adults of *C. cetaceum* could not be sampled in the same locality, but in places 600 km apart. Since there is evidence of morphological divergence between populations of *C. cetaceum* from South America and Australia (Aznar *et al.* 1999b), perhaps some degree of divergence might also occur at the geographical scale covered in our study, thus potentially affecting the morphometrical comparison between developmental stages. This does not appear to be the case because the morphology of all specimens of *C. cetaceum* thus far collected along the coast of southwestern Atlantic from Uruguay to Patagonia is very uniform (Aznar *et al.* 1999b, 2002b). Second, spines of *C. australe* were clearly smaller than those of *C. cetaceum*, and small structures may exhibit greater levels of variability just because their measurement is less precise

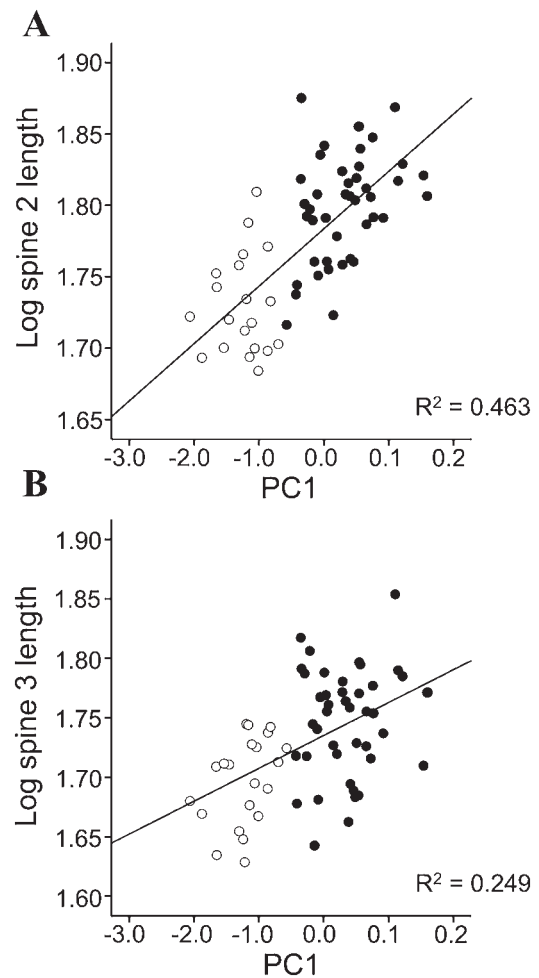


Fig. 6. Regression lines of spine length on the first principal component of 4 body variables (as indicated in Table 2) in cystacanth (open dots) and adult (solid dots) females of *Corynosoma cetaceum*. (A) Spine 2; (B) Spine 3 (see Fig. 2 for location of these spines on the body).

(see Aznar *et al.* 2002a). Although all spines had been measured at the same magnification regardless of species, coefficients of variation were very similar between *C. australe* and *C. cetaceum*. Therefore, the smaller size effect that was observed for spine growth in *C. australe* could hardly be accounted for by higher measurement error.

According to our results, both females and males of *C. australe* are roughly equal in size and grow at a similar rate from the cystacanth to the adult stage, whereas females of *C. cetaceum* are clearly smaller and grow less than males. However, females of both species have longer spines, and only in females do spines grow significantly during the adult development. Therefore, spine growth does not seem to follow simple allometric rules, nor does it conform to simple biomechanical principles i.e. females are not predicted to suffer stronger dislodgment forces than males according to their body size (Koehl, 1984; Poulin, 2007, 2009). So why do spines grow only in females? One hypothesis is that males require no further growth of spines beyond the cystacanth stage



Table 4. Models of multivariate analysis of covariance that examine the effects of developmental stage (cystacanth and adult), sex, and a multivariate measure of body size (PC1, the first principal component of the 4 morphometric variables indicated in Table 1) on the length of trunk spines from 3 sites in the acanthocephalan *Corynosoma australe*

(Statistically significant effects are in bold.)

Factor	D.F.	Wilks' lambda	F	P
<i>Full factorial model</i>				
Stage	3	3.429	3.429	<b>0.020</b>
Sex	3	0.843	6.733	<b>&lt;0.001</b>
PC1	3	0.770	0.770	0.513
Stage * Sex	3	0.875	3.532	<b>0.017</b>
Stage * PC1	3	0.996	0.152	0.557
Sex * PC1	3	0.982	0.695	0.927
Stage * Sex * PC1	3	0.986	0.539	0.657
Error	114			
Females				
<i>Full factorial model</i>				
Stage	3	0.864	3.246	<b>0.028</b>
PC1	3	0.960	0.859	0.467
Stage * PC1	3	0.980	0.416	0.742
Error	62			
<i>Main effects model</i>				
Stage	3	0.865	3.267	<b>0.027</b>
PC1	3	0.955	0.996	0.400
Error	63			
Males				
<i>Full factorial model</i>				
Stage	3	0.941	1.042	0.382
PC1	3	0.964	0.631	0.598
Stage * PC1	3	0.984	0.265	0.850
Error	62			
<i>Main effects model</i>				
Stage	3	0.941	1.062	0.373
PC1	3	0.958	0.741	0.533
Error	63			

because they develop other attachment devices (i.e. the proboscis, the disk) more than females during late ontogeny. We could not provide an overall test for this hypothesis because most adult specimens had an invaginated proboscis. However, our results clearly indicate that the area of the attachment disk does not differ between sexes. Also, information obtained from other datasets indicate that, in both species of *Corynosoma*, the proboscis and hooks are significantly smaller in adult males, and the field of spines covers a roughly similar extension of the trunk in both sexes (Hernández-Orts *et al. unpublished data*; see also Aznar *et al. 1999b*; Sardella *et al. 2005*). Thus, adult males appear to have a less-developed holdfast than females.

A second hypothesis would suggest that factors other than body size exert stronger overall selective pressures on females to develop more efficient

attachment devices, including spines. In this context, Petrochenko (1956) argued that adult females of acanthocephalans need to develop larger attachment structures than males because they must stay in the definitive host for longer to produce and release the eggs. Following this argument, the larger size of spines could be viewed as an adaptation of females to reduce the likelihood of being ripped loose by peristaltic movements and passing food (see Poulin, 2009). Females would also require a fine-tuned adjustment of the spine size to the specific micro-habitat conditions they encounter during the adult development. Note that the latter strategy is not unusual: after recruitment to the definitive host, females, but not males, of the polymorphid *Filicollis anatis* inflate the anchored proboscis as a device that obviously improves attachment performance (Van Cleave, 1952; Petrochenko, 1958).

The hypothesis mentioned above is supported by 2 lines of evidence. First, females of *C. cetaceum* and *C. australe* appear to have indeed a longer lifespan than males, as indicated by the strongly female-biased sex ratios observed in the definitive host (Aznar *et al. 2001, 2004*). A longer lifespan of females has also been recorded in other species of *Corynosoma* using controlled infections in experimental hosts (Valtonen and Helle, 1982; Castro and Martínez, 2004). Unfortunately, we lack direct quantitative data from natural hosts, although information obtained from an allied species of comparable size, *Polymorphus minutus*, suggests that the lifespan of females could be at least 1.5-fold than that of males (see data from Crompton and Whitfield, 1968).

Second, it is likely that lifespan differences between sexes may have a selective impact on attachment devices because carnivorous marine mammals are hosts that impose very harsh conditions for a gut-dwelling helminth (Petrochenko, 1956). Both cetaceans and pinnipeds have higher metabolic rates than terrestrial mammals of comparable size (Williams *et al. 2001*), and high metabolic rates are often associated with high rates of food intake and short transit times of food along the gut (Karasov and Diamond, 1985). With regard to food intake, carnivorous marine mammals need to feed often (Kastelein *et al. 1997a*), and on prey that are patchily distributed in the environment, so that large quantities of food are consumed when the occasion arises (Gaskin, 1978; Williams *et al. 2001*). Accordingly, acanthocephalans must suffer the frequent but unpredictable passing of a great amount of digested food. On the other hand, marine mammals have comparatively long alimentary tracts associated with their elevated metabolic rates (Williams *et al. 2001*), but the transit time of food is generally shorter than that of terrestrial mammals of similar size (Kastelein *et al. 1997b*; Hall-Aspland *et al. 2011*). Therefore, the flow of digesta must be, not only frequent, but fast. In summary, we believe that the need to withstand

extreme flow conditions for periods of different extent might have driven a different investment and development schedule of holdfast structures in males and females of *C. cetaceum* and *C. australe*.

Another non-exclusive hypothesis is also compatible with the observed sexual differences in investment and development schedule of spines in species of *Corynosoma* i.e. males and females differ in sexual behaviour. The mating system of acanthocephalans appears to be polygamous; males have a more active role in copulation than females, seeking and mating with several females (Parshad and Crompton, 1981). In the intestine of Saimaa ringed seals (*Phoca hispida saimensis*), Sinisalo *et al.* (2004) found evidence of significant competition between males of *Corynosoma magdalenii* for the access to females, with large-sized males firstly approaching non-mated females. Therefore, sexual selection could favour strong, permanent attachment in females of *Corynosoma*, but only short-term attachment in males as they need to move in search of mates.

Our study also indicates that patterns of spine growth differ between females of each species of *Corynosoma*. Attempting to infer adaptation in this 2-species comparison inevitably involves the confounding of independent variables (Garland and Adolph, 1994). In other words, each species lives within a different species of host and selects a different microhabitat and, therefore, each species is subject to different ecological regimes, including the degree of physical disturbance and food availability, which have never been quantified in the system under study. Therefore, we have no reasonable clue about the actual factors that account for the differences in morphology and growth patterns between species. Nonetheless, it is worth noting that spines on the disk border are the ones that grow in both species. Perhaps this is not surprising because the disk is a major attachment device in *Corynosoma* (Van Cleave, 1952), with the disk border exerting a wedge-like force against the host tissue (Aznar *et al.* 1999a). In contrast, hind-trunk spines are apparently used only as a secondary holdfast (Aznar *et al.* 2002). On the other hand, it seems clear that females of *C. cetaceum* fine-tune the size of spines during the development in the definitive host more than *C. australe*. All else being equal, this might be adaptive because (i) the relative increment in volume from cystacanth to adult in *C. cetaceum* is almost 3-fold that of *C. australe*, and (ii) females of *C. cetaceum* also achieve a larger adult size, a trait that correlates with stronger dislodging forces (Koehl, 1984; Poulin, 2007, 2009) and, possibly, with a longer lifespan (see Sorci *et al.* 1997).

Rather surprisingly, we found no significant patterns of static allometry between body size and the size of spines on the disk border in females of either *C. cetaceum* or *C. australe*. However, co-variation was significant for hind-trunk spines in females of *C. cetaceum*. This suggests that the final

size of spines may or may not match the adult body size achieved by each individual worm depending on the body region where the spine grows. Again, it seems premature to speculate on the reasons for these differences as we lack information about the factors that control spine morphogenesis (Aznar *et al.* 2002), and the specific attachment performance of disk or hind trunk spines (see Koehl, 1996). It should be pointed out, however, that narrow co-variation between spine size and final body size must not functionally be required if slight increases in spine size suffice for secure attachment within a range of body sizes (see Poulin, 2009).

In conclusion, this study sheds light on the question regarding whether or not the holdfast of acanthocephalans is fully developed prior to entering the definitive host. In particular, it suggests that temporal allocation of investment in attachment structures may differ, not only between congeneric species, but also between sexes of the same species, possibly due to the different selective pressures that each population subset faces. Future studies should address whether life span and body size are also relevant factors affecting development of other attachment structures (e.g. the proboscis) in a multi-species context.

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#### REFERENCES

- Amin, O. M., Heckmann, R. A., Mesa, R. and Mesa, E. (1995). Description and host relationships of cystacanths of *Polymorphus spindlatus* (Acanthocephala: Polymorphidae) from their paratenic fish hosts in Peru. *Journal of Helminthology* **62**, 249–253.
- Amin, O. M., Heckmann, R. A. and Van Ha, N. (2004). On the immature stages of *Pallisentis (Pallisentis) celatus* (Acanthocephala: Quadrigyridae) from occasional fish hosts in Vietnam. *The Raffles Bulletin of Zoology* **52**, 593–598.
- Amin, O. M. (1986). Acanthocephala from Lake Fishes in Wisconsin: Morphometric Growth of *Neoechinorhynchus cylindratu*s (Neoechinorhynchidae) and taxonomic implications. *Transactions of the American Microscopical Society* **105**, 375–380.

- Amin, O. M.** (1987). Acanthocephala from Lake Fishes in Wisconsin: Morphometric growth of *Pomphorhynchus bulbocollis* (Pomphorhynchidae). *Journal of Parasitology* **73**, 806–810.
- Aznar, F. J., Berón-Vera, B., Crespo, E. A. and Raga, J. A.** (2002b). Presence of genital spines in a male *Corynosoma cetaceum* Johnston and Best, 1942 (Acanthocephala). *Journal of Parasitology* **88**, 403–404. doi: 10.1645/0022-3395(2002)088[0403:POGSIA]2.0.CO;2.
- Aznar, F. J., Bush, A. O., Balbuena, J. A. and Raga, J. A.** (2001). *Corynosoma cetaceum* in the stomach of Franciscanas, *Pontoporia blainvillei* (Cetacea): An exceptional case of habitat selection by an acanthocephalan. *Journal of Parasitology* **87**, 536–541.
- Aznar, F. J., Bush, A. O., Fernández, M. and Raga, J. A.** (1999a). Constructional morphology and mode of attachment of the trunk of *Corynosoma cetaceum* (Acanthocephala: Polymorphidae). *Journal of Morphology* **241**, 237–249.
- Aznar, F. J., Bush, A. O. and Raga, J. A.** (1999b). *Polymorphus arctocephali* Smales, 1986, a synonym of *Corynosoma cetaceum* Johnston & Best, 1942 (Acanthocephala: Polymorphidae). *Systematic Parasitology* **44**, 59–70. doi: 10.1023/A:1006161620990.
- Aznar, F. J., Bush, A. O. and Raga, J. A.** (2002a). Reduction and variability of trunk spines in the acanthocephalan *Corynosoma cetaceum*: the role of physical constraints on attachment. *Invertebrate Biology* **121**, 104–114. doi: 10.1111/j.1744-7410.2002.tb00051.x.
- Aznar, F. J., Cappozzo, H. L., Taddeo, D., Montero, F. E. and Raga, J. A.** (2004). Recruitment, population structure, and habitat selection of *Corynosoma australe* (Acanthocephala) in South American fur seals, *Arctocephalus australis*, from Uruguay. *Canadian Journal of Zoology* **82**, 726–733. doi: 10.1139/Z04-044.
- Aznar, F. J., Hernández-Orts, J., Suárez, A. A., García-Varela, M., Raga, J. A. and Cappozzo, H. L.** (2012). Assessing host-parasite specificity through coprological analysis: a case study with species of *Corynosoma* (Acanthocephala: Polymorphidae) from marine mammals. *Journal of Helminthology*. doi: 10.1017/S0022149X11000149.
- Aznar, F. J., Pérez-Ponce de León, G. and Raga, J. A.** (2006). Status of *Corynosoma* (Acanthocephala: Polymorphidae) based on anatomical, ecological, and phylogenetic evidence, with the erection of *Pseudocorynosoma* n. gen. *Journal of Parasitology* **92**, 548–564. doi: 10.1645/GE-715R.1.
- Castro, M. and Martínez, R.** (2004). Process of the development of *Corynosoma obtusens* (Acanthocephala: Polymorphidae) in *Canis familiaris* and its possible involvement in public health. *Parasitologia Latinoamericana* **59**, 26–30.
- Cock, A. G.** (1966). Genetical aspects of metrical growth and form in animals. *The Quarterly Review of Biology* **41**, 131–190.
- Crompton, D. W. T. and Whitfield, P. J.** (1986). The course of infection and egg production of *Polymorphus minutus* (Acanthocephala) in domestic ducks. *Parasitology* **58**, 231–246.
- Engqvist, L.** (2005). The mistreatment of covariate interaction terms in linear model analyses of behavioural and evolutionary ecology studies. *Animal Behaviour* **70**, 967–971. doi: 10.1016/j.anbehav.2005.01.016.
- Garland, T., Jr. and Adolph, S. C.** (1994). Why not to do two-species comparative studies: Limitations on inferring adaptation. *Physiological Zoology* **67**, 797–828.
- Gaskin, D. E.** (1978). Form and function in the digestive tract and associated organs in Cetacea, with a consideration of metabolic rates and specific energy budgets. *Oceanography and Marine Biology: An Annual Review* **16**, 313–345.
- Hall-Aspland, S., Rogers, T., Canfield, R. and Tripovich, J.** (2011). Food transit times in captive leopard seals (*Hydrurga leptonyx*). *Polar Biology* **34**, 95–99. doi: 10.1007/s00300-010-0862-4.
- Karasov, W. H. and Diamond, J. M.** (1985). Digestive adaptations for fueling the cost of endothermy. *Science* **228**, 202–204. doi: 10.1126/science.3975638.
- Kastelein, R. A., Hardeman, J. and Boer, H.** (1997a). Food consumption and body weight of harbour porpoises (*Phocoena phocoena*). In *The Biology of the harbour porpoise* (ed. Read, A. J., Wiepkema, P. R. and Nachtigall, P. E.), pp. 217–233. De Spil Publishers, Woerden.
- Kastelein, R. A., Nieuwstraten, S. H. and Versteegen, M. W. A.** (1997b). Passage time of carmine red dye through the digestive tract of harbour porpoises (*Phocoena phocoena*). In *The Biology of the Harbour Porpoise* (ed. Read, A. J., Wiepkema, P. R. and Nachtigall, P. E.), pp. 265–275. De Spil Publishers, Woerden, The Netherlands.
- Klingenberg, C. P.** (1996). Multivariate Allometry. In *Advances in Morphometrics*, (ed. Marcus, L. F., Corti, M., Loy, A., Naylor, G. J. P. and Slice, D. E.), pp. 23–49. NATO ASI Series A: Life Sciences, Vol. 284, New York, USA.
- Koehl, M. A. R.** (1984). How do benthic organisms withstand moving water? *American Zoologist* **24**, 57–70. doi: 10.1093/icb/24.1.57.
- Koehl, M. A. R.** (1996). When does morphology matter? *Annual Review of Ecology and Systematics* **27**, 501–542. doi: 10.1146/annurev.ecolsys.27.1.501.
- Parshad, V. R. and Crompton, D. W. T.** (1981). Aspects of acanthocephalan reproduction. *Advances in Parasitology* **19**, 73–138. doi: 10.1016/S0065-308X(08)60266-3.
- Petrochenko, V. I.** (1956). *Acanthocephala of Domestic and Wild Animals*. Vol. I. Izdatel'stvo Akademii Nauk SSSR, Moscow. English translation by Israel Program for Scientific Translations Ltd., 1971.
- Petrochenko, V. I.** (1958). *Acanthocephala of Domestic and Wild Animals*. Vol. II. Izdatel'stvo Akademii Nauk SSSR, Moscow. English translation by Israel Program for Scientific Translations Ltd., 1971.
- Podesta, R. B. and Holmes, J. C.** (1970). The life cycles of three Polymorphids (Acanthocephala) occurring as juveniles in *Hyaella asteca* (Amphipoda) at Cooking Lake, Alberta. *Journal of Parasitology* **56**, 1118–1123.
- Poulin, R.** (2007). Investing in attachment: evolution of anchoring structures in acanthocephalan parasites. *Biological Journal of the Linnean Society* **90**, 637–645. doi: 10.1111/j.1095-8312.2006.00754.x.
- Poulin, R.** (2009). Interspecific allometry of morphological traits among trematode parasites: selection and constraints. *Biological Journal of the Linnean Society* **96**, 533–540. doi: 10.1111/j.1095-8312.2008.01163.x.
- Poulin, R., Wise, M. and Moore, J.** (2003). A comparative analysis of adult body size and its correlates in acanthocephalan parasites. *International Journal for Parasitology* **33**, 799–805. doi: 10.1016/S0020-7519(03)00108-5.
- Randhawa, H. S. and Poulin, R.** (2010). Evolution of interspecific variation in size of attachment structures in the large tapeworms genus *Acanthobothrium* (Tetrathyridae: Onchobothriidae). *Parasitology* **137**, 1707–1720. doi: 10.1017/S0031182010000569.
- Sardella, N. H., Mattiucci, S., Timi, J. T., Bastida, R. O., Rodríguez, D. H. and Nascetti, G.** (2005). *Corynosoma australe* Johnston, 1937 and *C. cetaceum* Johnston & Best, 1942 (Acanthocephala: Polymorphidae) from marine mammals and fishes in Argentinian waters: allozyme markers and taxonomic status. *Systematic Parasitology* **61**, 143–156. doi: 10.1007/s11230-005-3131-0.
- Schmidt, G. D.** (1985). Development and life cycles. In *Biology of the Acanthocephala*, (ed. Crompton, D. W. T. and Nickol, B. B.), pp. 273–286. Cambridge University Press, Cambridge, UK.
- Schulze, K.** (2006). Imaging and modeling of digestion in the stomach and the duodenum. *Journal of Neurogastroenterology and Motility* **18**, 172–183. doi: 10.1111/j.1365-2982.2006.00759.x.
- Sinialo, S., Poulin, R., Högmänder, H., Juuti, T. and Valtonen, E. T.** (2004). The impact of sexual selection on *Corynosoma magdalenae* (Acanthocephala) infrapopulations in Saimaa ringed seals (*Phoca hispida saimensis*). *Parasitology* **128**, 179–185. doi: 10.1017/S003118200300430X.
- Sorci, G., Morand, S. and Hugot, J. P.** (1997). Host parasite coevolution: evidence for covariation of life history traits in Primates and oxyurid parasites. *Proceedings of the Royal Society of London, B* **264**, 285–289.
- Taraschewski, H.** (2000). Host-parasite interactions in Acanthocephala: a morphological approach. *Advances in Parasitology* **46**, 1–179.
- Valtonen, E. T. and Helle, E.** (1982). Experimental infection of laboratory rats with *Corynosoma semerme* (Acanthocephala). *Parasitology* **85**, 9–19. doi: 10.1017/S0031182000054093.
- Van Cleave, H. J.** (1952). Some host-parasite relationships of the Acanthocephala, with special reference to the organs of attachment. *Experimental Parasitology* **1**, 305–330.
- Williams, T. M., Haun, J., Davis, R. W., Fuiman, L. A. and Kohin, S.** (2001). A killer appetite: metabolic consequences of carnivory in marine mammals. *Comparative Biochemistry and Physiology Part A* **129**, 758–796.