

Short Communication

Assessment of genetic diversity within and among sage (*Salvia*) species using SRAP markers

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Abstract

Salvia (sage) is the most important and largest genus of the *Lamiaceae* family. High similarities among species in this genus lead to difficulty in its systematic identification. Despite its economic importance, limited molecular studies have been conducted to evaluate the genetic diversity among and within *Salvia* species. In this study, SRAP (sequence-related amplified polymorphism) markers, which targeted ORFs (open reading frames) as functional regions in the genome, were used to detect the genetic diversity of five *Salvia* species (*S. virgata* Jacq., *S. nemorosa* L., *S. officinalis* L., *S. cereal* L. and *S. sclarea* L.). Fourteen primer combinations (PCs) were amplified by 265 fragments on 54 genotypes, in which 255 (96%) were polymorphic. The average polymorphism information content (PIC) value was 0.308 over all PCs. The genetic distance among species ranged from 0.126 (between *S. virgata* Jacq. and *S. nemorosa* L.) to 0.568 (between *S. nemorosa* L. and *S. sclarea* L.). Based on Jaccard's similarity coefficient and UPGMA algorithm, cluster analysis separated different species ($r = 0.920$). The results showed high genetic differentiation ($F_{st} = 0.337$) and negligible gene flow ($N_m = 0.750$) among species. Owing to the high genetic variation among and within *Salvia* species, it serves as a rich source of germplasm with potential for use in breeding programmes.

Keywords: AMOVA; genetic variation; *Salvia*; SRAP

Introduction

Salvia is an important and also the largest genus of the *Lamiaceae* family, which includes nearly 1000 species (Walker and Sytsma, 2007). Fifty-eight annual or perennial species of the genus have been found in Iran, 17 of which are endemic (Walker and Sytsma, 2007). Some of the *Salvia* species are considered as a valuable spice in

food industries (Gali-Muhtasib *et al.*, 2000), and grown in parks and gardens as ornamental plants.

Identification of *Salvia* species is complicated due to the morphological similarity and common occurrence of natural hybridization within species (Reales *et al.*, 2004; Walker *et al.*, 2004). Molecular markers have been widely used in the identification of species and genotypes (Skoula *et al.*, 1999; Karaca *et al.*, 2004, 2008; Berteau *et al.*, 2005). Numerous SSR markers have also been developed for the most important *Salvia* species, *S. officinalis* L. (Mader *et al.*, 2010;

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Radosavljević *et al.*, 2011, 2012; Wang *et al.*, 2011). Among the different molecular marker systems, sequence-related amplified polymorphism (SRAP) is a relatively simple and highly reproducible DNA marker (Li and Quiros, 2001).

Within the genus *Salvia*, there are few species of significant economic importance, and numerous studies have focused on cultivation, effective ingredients and pharmacological properties of these species (Gali-Muhtasib *et al.*, 2000; Delamare *et al.*, 2007; Kelen and Tepe, 2008). According to the literature, no report has been recorded regarding genetic diversity at the inter-specific level among *Salvia* species using SRAP markers. Therefore, this study aimed at the utilization of SRAP markers in assessing the genetic diversity of *Salvia* species including *S. virgata* Jacq., *S. nemorosa* L., *S. officinalis* L., *S. sclarea* L. and *S. cereal* L.

Experimental procedure

Aerial parts of 54 sage samples belonged to *S. virgata* (12), *S. nemorosa* L. (14), *S. officinalis* L. (15), *S. sclarea* L. (3) and *S. cereal* L. (10) species, which were collected from different regions in Iran (Supplementary Table S1 and Fig. S1, available online). Genomic DNA was

extracted from the ground powder using a HiYield genomic DNA mini kit (HiYield™ Genomic DNA Mini Kit, Real Biotech Corporation, Banqiao City, Taiwan) following the manufacturer's instructions.

PCRs and amplifications were performed according to Li and Quiros (2001). The amplified products were separated on 8% non-denatured polyacrylamide gel electrophoresis and visualized by silver staining (Bassam *et al.*, 1991).

Based on the presence of reproducible polymorphic bands on the gel, DNA fragments were scored in all the 54 *Salvia* species samples. A dendrogram was constructed based on Jaccard's similarity coefficient using the UPGMA (unweighted pair group method with arithmetic mean) algorithm, and genetic relationships among genotypes were further analysed by the principal coordinate analysis (PCoA) of a similarity matrix using NTSYS-pc version 2.02 (Rohlf, 1998).

The PIC was calculated for all selected markers according to Smith *et al.* (1997).

Genetic diversity within and among species was measured by the percentage of polymorphic bands, the effective number of alleles, the observed number of alleles, Nei's gene diversity, Shannon's information index and gene flow. The UPGMA dendrogram of species was constructed using NTSYS-pc version 2.02,

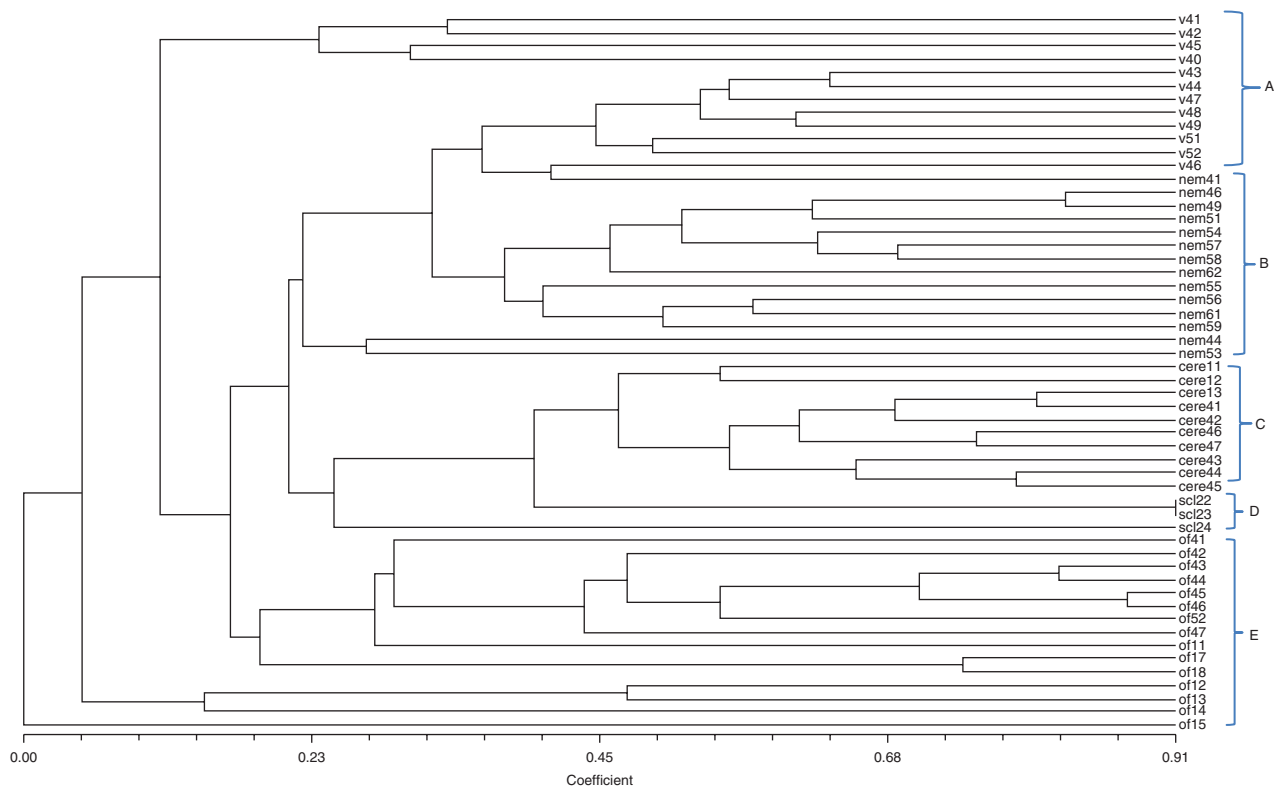


Fig. 1. UPGMA dendrogram of 54 sage genotypes using SRAP markers based on Jaccard's coefficient. A, B, C, D and E groups consist of *S. virgata*, *S. nemorosa* L., *S. cereal* L., *S. sclarea* L. and *S. officinalis* L., respectively.

Table 1. Summary of genetic variation statistics for all loci in the five *Salvia* species

Species name	Sample size	NPB	PPB	H_e	I	N_a	N_e
<i>S. virgata</i>	12	162	63.53	0.221	0.332	1.635	1.381
<i>S. nemorosa</i>	14	177	69.41	0.251	0.375	1.694	1.429
<i>S. cereal</i>	10	128	50.20	0.181	0.270	1.502	1.307
<i>S. sclarea</i>	3	27	10.59	0.090	0.129	1.203	1.162
<i>S. officinalis</i>	15	137	53.73	0.210	0.307	1.537	1.376

NPB, number of polymorphic bands; PPB, percentage of polymorphic bands; H_e , Nei's gene diversity; I , Shannon's information index; N_a , observed number of alleles; N_e , effective number of alleles.

and based on co-ancestry coefficients obtained from the pairwise F_{st} distance matrices using POPGENE 1.32 software (Yeh and Yang, 1999).

Results and Discussion

A total of 32 different SRAP primer combinations (PCs) were evaluated for their ability to prime the PCR amplification of five randomly selected sage samples from different species. Fourteen selected PCs were amplified by 265 bands among the 54 *Salvia* species samples, of which 255 bands were polymorphic (96%). The high polymorphism rate found in this study is in agreement with the previous observations of genetic diversity among *Salvia* species (Song *et al.*, 2010; Sepehry Javan *et al.*, 2012; Zhang *et al.*, 2013; Peng *et al.*, 2014). The mean PIC value for the PCs was 0.308, which ranged from 0.201 to 0.394 (Supplementary Table S2, available online), indicating that SRAP markers showed medium polymorphism and could contribute to the genetic variation of *Salvia* species.

The UPGMA dendrogram with a high cophenetic correlation coefficient ($r = 0.920$) based on SRAP markers grouped the five species into five distinct clusters (Fig. 1), which is in agreement with the previous observations of Sepehry Javan *et al.* (2012). The A, B, C, D and E groups consisted of *S. virgata*, *S. nemorosa* L., *S. cereal* L., *S. sclarea* L. and *S. officinalis* L., respectively. The SRAP analyses showed *S. virgata* Jacq. and *S. officinalis* L. as the most divergent ones.

Genetic relationships among the sage samples were also analysed by the PCoA. The first three principal coordinates explained 26.2% of the total variation, showing that the original data were not highly correlated in the PCoA.

The 54 sage samples were grouped into five groups by their species. On the whole, among the five species, the highest Shannon's information index, Nei's gene diversity index, the observed and effective number of alleles, and the percentage of polymorphic loci were found in *S. nemorosa* L. (Table 1). The largest genetic distance (0.565) was observed between *S. nemorosa* L. and

S. sclarea L. and the smallest one (0.126) occurred between *S. nemorosa* L. and *S. virgata* Jacq. (Supplementary Table S3, available online).

In conclusion, SRAP markers, which targeted ORFs (open reading frames) as functional regions of the sage genome and reached the resulting sufficient polymorphism, can be successfully used for determining the genetic diversity and population structure of *Salvia* species.

Supplementary material

To view supplementary material for this article, please visit <http://dx.doi.org/10.1017/S1479262115000593>

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References

- Bassam BJ, Caetano-Anolles G and Gresshoff PM (1991) Fast and sensitive silver staining of DNA in polyacrylamide gels. *Analytical Biochemistry* 196: 80–83.
- Berte CM, Azzolin CMM, Bossi S, Doglia G and Maffei M (2005) Identification of an *EcoRI* restriction site for a rapid and precise determination of β -asarone-free *Acorus calamus* cytotypes. *Phytochemistry* 66: 507–514.
- Delamare A, Pistorello I, Artico L, Serafini LA and Echverrigaray S (2007) Antibacterial activity of essential oils of *Salvia officinalis* L. and *Salvia triloba* L. cultivated in South Brazil. *Food Chemistry* 100: 603–608.
- Gali-Muhtasib H, Hilan C and Khater C (2000) Traditional uses of *Salvia libanotica* (East Mediterranean sage) and the effects of its essential oils. *Journal of Ethnopharmacology* 71: 513–520.
- Karaca M, Saha S, Callahan FE, Jenkins JN, Read JJ and Percy RG (2004) Molecular and cytological characterization of a cytoplasmic-specific mutant in pima cotton (*Gossypium barbadense* L.). *Euphytica* 139: 187–197.
- Karaca M, Ince AG, Tugrul-Ay S, Turgut K and Onus AN (2008) PCR-RFLP and DAMD-PCR genotyping for *Salvia* species.

- Journal of the Science of Food and Agriculture* 88: 2508–2516.
- Kelen M and Tepe B (2008) Chemical composition, antioxidant and antimicrobial properties of the essential oils of three *Salvia* species from Turkish flora. *Bioresource Technology* 99: 4096–4104.
- Li G and Quiros C (2001) Sequence-related amplified polymorphism (SRAP), a new marker system based on a simple PCR reaction: its application to mapping and gene tagging in *Brassica*. *Theoretical and Applied Genetics* 103: 455–461.
- Mader E, Lohwasser U, Börner A and Novak J (2010) Population structures of genebank accessions of *Salvia officinalis* L. (Lamiaceae) revealed by high resolution melting analysis. *Biochemical Systematics and Ecology* 38: 178–186.
- Peng L, Ru M, Wang B, Wang Y, Li B, Yu J and Liang Z (2014) Genetic diversity assessment of germplasm collection of *Salvia miltiorrhiza* Bunge. based on morphology, ISSR and SRAP markers. *Biochemical Systematics and Ecology* 55: 84–92.
- Radosavljević I, Jakse J, Javornik B, Satovic Z and Liber Z (2011) New microsatellite markers for *Salvia officinalis* (Lamiaceae) and cross-amplification in closely related species. *American Journal of Botany* 98: e316–e318.
- Radosavljević I, Satovic Z, Jakse J, Javornik B, Greguras D, Jug-Dujakovic M and Liber Z (2012) Development of new microsatellite markers for *Salvia officinalis* L. and its potential use in conservation-genetic studies of narrow endemic *Salvia brachyodon* Vandas. *International Journal of Molecular Sciences* 13: 12082–12093.
- Reales A, Rivera D, Palazon JA and Obon C (2004) Numerical taxonomy study of *Salvia* sect. *Salvia* (Labiatae). *Botanical Journal of the Linnean Society* 145: 353–371.
- Rohlf FJ (1998) *NTSYS-PC Numerical Taxonomy and Multivariate Analysis System, Version 2.02*. Setauket, NY: Exeter Publications, pp. 1–31.
- Sepehry Javan Z, Rahmani F and Heidari R (2012) Assessment of genetic variation of genus *Salvia* by RAPD and ISSR markers. *Australian Journal of Crop Sciences* 6: 1068–1073.
- Skoula M, El-Hilali I and Makris A (1999) Evaluation of the genetic diversity of *Salvia fruticosa* Mill. clones using RAPD markers and comparison with the essential oil profiles. *Biochemical Systematics and Ecology* 27: 559–568.
- Smith JSC, Chin ECL, Shu H, Smith OS, Wall SJ, Senior ML, Mitchell SE, Kresovich S and Zeigler J (1997) An evaluation of the utility of SSR loci as molecular markers in maize (*Zea mays* L.): comparison with data from RFLPs and pedigree. *Theoretical and Applied Genetics* 95: 163–173.
- Song Z, Li X, Wang X and Wang H (2010) Genetic diversity and population structure of *Salvia miltiorrhiza* Bge in China revealed by ISSR and SRAP. *Genetica* 138: 241–249.
- Walker JB and Sytsma KJ (2007) Staminal evolution in the genus *Salvia* (Lamiaceae): molecular phylogenetic evidence for multiple origins of the staminal lever. *Annals of Botany* 100: 375–391.
- Walker JB, Sytsma KJ, Treutlein J and Wink M (2004) *Salvia* is not monophyletic: implications for the systematics radiation and ecological specializations of *Salvia* and tribe Menthaeae. *American Journal of Botany* 91: 1115–1125.
- Wang X, Zhou X, Gao W, Cui G, Huang L and Lui C (2011) New analysis of EST-SSR distribution and development EST-SSR markers in *Salvia miltiorrhiza*. *Zhongguo Zhongyao Zazhi* 36: 289–293.
- Yeh FC and Yang RC (1999) *POPGENE Version 1.31*. Edmonton, Alberta: University of Alberta and Tim Boyle, Center for International Forestry Research.
- Zhang Y, Li X and Wang Z (2013) Diversity evaluation of *Salvia miltiorrhiza* using ISSR markers. *Biochemical Genetics* 51: 707–721.