Differential ultrasonic indices of separation distress in the presence and absence of maternal cues in infant rats bred for high and low positive social affect

Iacobucci P, Colonnello V, Fuchs T, D'Antuono L and Panksepp J. Differential ultrasonic indices of separation distress in the presence and absence of maternal cues in infant rats bred for high and low positive social affect.

Objective: Preclinical models of human mood disorders commonly focus on the study of negative affectivity, without comparably stressing the role of positive affects and their ability to promote resilient coping styles. We evaluated the role of background constitutional affect of rats by studying the separation and reunion responses of infants from low and high positive affect genetic lines (i.e., differentially selected for High and Low 50 kHz ultrasonic vocalisations (USVs).

Methods: Infants from Low and High 50 kHz USV breeding lines were isolated from mothers and exposed to either social (familiar or unfamiliar bedding) or neutral (clean bedding) odour cues between two short isolation periods, and tested in homeothermic and hypothermic ambient temperatures. Negative affect was estimated by monitoring separation distress calls (35–45 kHz USVs).

Results: Low Line pups called at higher rates than High Line, and their rates were stable regardless of odour cue. In contrast, High Line pups increased vocalisations during the second compared with the first isolation periods and during exposure to both familiar and unfamiliar odour cues, but not to neutral odour. Furthermore, the greatest increase in USV emission was seen in the second isolation period following exposure to the unfamiliar odour. However, both lines showed comparable elevated distress USVs to the thermal stressor.

Conclusion: High Line animals, selected for a positive affective phenotype (50 kHz USVs), exhibited reduced separation anxiety responses in infancy, making this a promising animal model for the role of constitutional affective states in emotional responsivity and potential resilience against emotional disorders.

Paolo lacobucci^{1,2}, Valentina Colonnello^{1,2}, Thomas Fuchs¹, Laura D'Antuono^{1,2}, Jaak Panksepp¹

¹Department of VCAPP, College of Veterinary Medicine, Washington State University, Pullman, USA; and ²Department of Psychology, University of Rome 'Sapienza', Rome, Italy

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Jaak Panksepp, Department of VCAPP, College of Veterinary Medicine, Washington State University, Pullman, WA 99164-6520, USA. Tel: 509-335-5803; Fax: 509-335-4650; E-mail: jpanksepp@vetmed.wsu.edu

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Significant outcomes

- Selective breeding for low and high levels of positive affective vocalisations in juvenile rats promotes differential responsivity to social but not physical stress, confirming its potential utility for understanding affective temperamental traits.
- Rats selected for high positive affectivity exhibit increased coping with negative affective circumstances, suggesting selection for a stress-resilient phenotype.
- The genetic selection for differential positive and negative affects may provide preclinical models with implications for understanding human psychopathologies, especially depression.

Limitations

- Because only maternal odour was used to evaluate maternal potentiation effects, additional contrasts to the physical presence of the mother would be useful.
- Inclusion of a random selection line would have been desirable, but was unavailable when the research was conducted.
- The genetic characterisation of these selection lines remains to be conducted.

Introduction

In recent years, several genetic models of humantype emotional traits have been developed in rodents in order to facilitate the study of the neural mechanisms of emotionality (1–6). Among these efforts, the characterisation of ultrasonic vocalisations (USVs) in laboratory rats has been a powerful tool to investigate negative as well as positive affective states (7–11) with important implications for psychiatric medicinal development.

Rats emit USVs at the earliest stages of life and the production of USVs varies greatly as a function of age (12). Infant rats exhibit short periods of high rates of 35–45 kHz USVs (13) when separated from their mother and littermates (14). These stress-induced USVs show a developmental curve with a peak at postnatal day (PND) 10–12 and a steep decline after PND 14 (15). Given the aversive circumstances (social isolation, hypothermia, etc.), it has been assumed that these infant USVs are associated with a state of negative affect or distress (hence they are called 'distress calls').

Differences in distress call rates have been used as a procedure for selective breeding of emotional dispositions that may model human temperamental traits: Brunelli and Hofer (16) selectively bred rats based on high and low rates of distress USVs emitted by isolated infants. These two lines exhibited distinct coping styles across the life span, with results suggesting a negative affect phenotype in their high distress USV line. Similarly, Burgdorf et al. (1) developed an animal model for positive affective phenotypes. Whereas Brunelli and Hofer selected for infant distress USVs that may index negative-affect, Burgdorf's group selected for positive USVs: Low and High Lines of Long Evans rats were selectively bred based on the number of positive 50 kHz 'chirping' USVs emitted during a brief hand-play procedure (i.e., 'tickling') conducted in the juvenile period. The number of USVs presumably reflects the intensity of a rat's positive affective social experience during tickling [(1,17),and for a review, see Panksepp and Burgdorf (18)], and it has been validated that the neural circuitry that mediates these calls (our objective measure of affect), has positive reinforcing properties (7,19). Burgdorf's two lines have exhibited variations in behavioural

phenotypes such as social motivation in response to social isolation in infancy (i.e., isolation distress calls) (20), anxiety- and depressive-like responses (21), and cocaine-induced hedonic-like responses (22), suggesting differential affective traits.

In order to extend these results, we investigated potential Line differences in USV emission rates in response to social isolation, ambient temperature and relevant social odours within an isolation-reunionreisolation paradigm first described by Shair (15). This paradigm evinced in rats and mice the so-called 'maternal potentiation' phenomenon, which is considered to be one of the strongest forms of evidence for the social-bonding relevance of infant isolation calls (15,23-25), with a potential resemblance to human infant emotional behaviour (23). Previous studies have shown that isolationinduced USVs increased markedly during the second of two closely spaced social-isolation periods, especially if the rat pups were briefly reunited with their mothers between the two sessions (26). This response was not evident when pups were reunited with littermates or unfamiliar females [see Dichter et al. (6), for a review] suggesting it reflected a mother-specific form of social bonding.

In our study we used the isolation-reunionreisolation paradigm to evaluate differences in social motivation, as measured by isolation USVs, between rats selectively bred for low and high levels of tickleinduced 50 kHz USVs - namely for low and high positive affect, respectively (1). However, rather than uniting pups with mothers and strangers between isolation periods, they were exposed to either social (familiar or unfamiliar), or neutral nest-odour cues, because it has been previously demonstrated that the USV distress emission of infant rats can be differentially attenuated by home-nest odours (14,27–29). We chose this modified paradigm, because rat pups find home nest shavings rewarding and because our target was the pups' vocal activity with no direct influence of maternal interactions. We predicted that both lines would increase rates of USVs in the second isolation, compared with the first isolation, and would decrease rates of USVs in the presence of social odours (both familiar and unfamiliar, but perhaps differentially) compared with neutral (control) odour, as previously found in both rats and mice (3,26). The rate of infant USVs is also affected by environmental temperature: a decrease in temperature is generally associated with elevated rates of USVs, which may reflect age-related thermoregulatory deficits, yielding distress from increasing hypothermia (14,30,31). Thus, we carried out all sessions at two different ambient temperatures: (1) the nest equivalent temperature of 35°C; and (2) a cool temperature of 23°C, in order to evaluate whether the two lines also vocalised differently in response to physical discomfort: we predicted that Low Line rats, which seem more susceptible to negative affect (20), would call more than High Line animals.

Materials and methods

Animals and housing

We used rat pups from the 17th generation of Long Evans rats selectively bred, bidirectionally, for low (Low Line) and high (High Line) rates of 50 kHz USVs emitted in response to a standard repeated (4-day) tickling procedure (1). Parenthetically, we did not use animals from a random-breeding line because those animals were exhibiting apparent inbreeding sterility; thus, we no longer had sufficient animals of that genotype available. The colony was maintained at a constant room temperature ($20 \pm 1^{\circ}$ C and $60 \pm 10\%$ relative humidity) and on a 12/12 light-dark cycle (light on at 07:00 a.m.). Male and female pairs were housed in opaque plastic cages $(73 \times 54 \times 24 \text{ cm})$ with food and water ad libitum. When the females were visibly pregnant, they were individually housed to avoid postpartum pregnancy, so no male odour was present in the bedding at birth. The day of birth was considered PND 0. The litters (Low Line, n = 16; High Line, n = 16) were left undisturbed until testing at PND 12 or 13. On the testing day, no pups had yet opened their eyes. Animals were tested in counterbalanced order across ages.

Procedure

On the testing day, eight Low Line litters and eight High Line litters were assigned to the 'nest temperature' condition, while the remaining litters were assigned to the 'cool temperature' condition. For this purpose, we used acoustically isolated temperature-regulated environmental test chambers (Model 352602, Hotpack, Philadelphia, PA, USA), set at either nest or cool temperatures $(35 \pm 0.5^{\circ}C)$ or $23 \pm 0.5^{\circ}C$, respectively), and illuminated with a dim, red light. This value for the nest temperature condition was determined by monitoring the ambient temperature of a litter immediately after the dam vacated it.

From each litter, three pups were arbitrarily selected and assigned to one of the following odour conditions: familiar bedding (FB), unfamiliar bedding (UB), or clean bedding (CB), reflecting which odour cue they were exposed to between successive isolation sessions. For the FB condition we used 400 g of bedding from their own home cage, while for the UB condition we used the same quantity of bedding from a cage housing a different litter of the same age.

Each pup underwent three test sessions: Isolation 1 (Iso1), Bedding exposure (Bedding), Isolation 2 (Iso2). Fifteen minutes before the test, the dam was removed and the home cage of each litter was placed in a separate acoustically isolated temperatureregulated environmental chamber maintained at $35 \pm 0.5^{\circ}$ C. After 15 min of habituation to the move, each test subject was placed alone into an empty opaque plastic test box $(29 \times 19 \times 12 \text{ cm})$ in temperature-regulated isolation its respective. chamber for 2 min (Iso1). Afterwards, each animal was placed into another box with identical features, but containing 400 g of FB, UB, or CB and monitored for 2 min (Bedding). The pup was then re-isolated (Iso2) for 2 min in the empty isolation box (Fig. 1). Following testing, body weight and axillary temperature (measured by insertion of microprobe (Sixth Sense LT300 Infrared а Thermometer, Instrumart, Carlsbad, CA, USA)) were recorded and the test box was cleaned with a diluted alcohol solution. All tests for a particular pup occurred in the same isolation chamber.

For the FB and UB conditions, faecal boli were removed from the soiled bedding before use. During

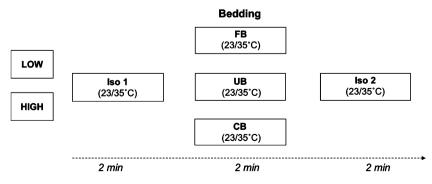


Fig. 1. Experimental design. CB, clean bedding; FB, familiar bedding; Iso, isolation; UB, unfamiliar bedding.

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all the sessions and for all the conditions, the testing boxes were covered with a Plexiglas lid that maintained a constant concentration of odour and also served to prevent odour dispersal into the isolation chamber. Experiments were carried out between 01:00 p.m. and 04:00 p.m. (i.e., starting 6 h into the light cycle), and all pups were handled with nitrile gloves.

The experiment was conducted in compliance with standard institutional guidelines for the care and use of research animals, and all experimental protocols were approved by the Washington State University Institutional Animal Care and Use Committee.

Data collection

USVs were recorded with a high quality ultrasonic microphone (Condenser ultrasound microphone CM16/CMPA; Avisoft Bioacoustics, Berlin, Germany) placed inside the isolation chamber and suspended ~5 cm above the centre of the plastic test box. The signal was then amplified and converted to digital format using a digital audio card (Avisoft USG Ultra Sound Gate; Avisoft Bioacoustics). Digital acquisition and analysis was performed on a PC with SeaPro Ultra software developed by Pavan at CIBRA (http://www.unipv.it/cibra). We report the total number of USVs emitted during each session. The numbers of USVs were scored offline by two trained observers blind to the treatments, and exhibiting high inter-observer concordance (>95%).

Statistical analysis

Analyses were performed using a mixed-model analysis of variance (ANOVA) for repeated measures. The model included between factors (Line: Low and High; Temperature: Nest and Cool; Odour condition: FB, UB and CB) and within factors (Sessions: Isol, Bedding, Iso2). The model included the litter as the blocking factor. *Post hoc* comparisons were performed using Tukey's honestly significant differences test, which is permissible in the absence of significant ANOVA effects (32). Statistical analyses were conducted using Statview II software (Abacus Concepts Inc., Berkeley, CA, USA).

Results

Statistical analysis revealed a main effect of Line $[F(1, 28) = 52.8, p < 0.0001, \eta = 1]$, with the Low Line vocalising more than the High Line (Low: 238.3 ± 9.5; High 97.7 ± 4.2) over all sessions. Low Line rates of USV emissions did not change over the three sessions. In contrast, *post hoc* analyses on the

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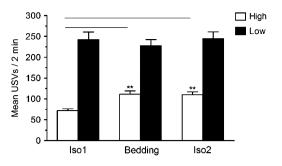


Fig. 2. Line × Session interaction. Mean (±SE) of USVs emitted by Low and High lines during a three-session test: Iso1, Bedding, and Iso2. Iso, isolation; USVs, ultrasonic vocalisations. **p < 0.01.

Line × Session interaction [$F(2, 56) = 4.9, p = 0.01, \eta = 0.7$; Fig. 2] revealed that High Line pups increased USVs during the Bedding (Bedding > Iso1, p < 0.01) and Iso2 sessions (Iso2 > Iso1, p < 0.01).

In addition, there was a significant main effect for Temperature $[F(1, 28) = 5.2, p = 0.03, \eta = 0.6],$ with both lines emitting more vocalisations at the cool temperature compared with the nest temperature (Line \times Temperature interaction: F(1, 28) = 2.0, p = 0.17, $\eta = 0.2$; High: Cool = 106.4 ± 6; Nest = 89.2 ± 5.9 ; Low: Cool = 274 ± 11.8, Nest = 202.5 ± 13.7). No differences were observed for body weight (High: 22.9 ± 0.4 g; Low: 23.8 ± 0.7 g) and axillary temperature (High: $33.8 \pm 0.3^{\circ}$ C; Low: $34.3 \pm 0.2^{\circ}$ C) between lines (p > 0.05). For the odour conditions, the Line \times Odour interaction was not significant [F(2, 56) = 0.2, p > 0.05 overall]. However, a post hoc inspection of the data on the interaction Line × Session × Odour [F(4, 112) = 1.6, p = 0.17, $\eta = 0.3$; Fig. 3] revealed that whereas Low Line USV rates were clearly not affected by exposure to any of the odours during any session, there were odourinduced changes in the High Line animals, which showed lower overall rates of USVs. In particular, High Line pups showed comparable increases of vocalisations both during as well as after the exposure to the soiled bedding (i.e., both FB and UB conditions), but not during and after the exposure to the CB condition (High-Iso1 < High-Bedding in FB, p < 0.01; High-Iso1 < Bedding and High-Iso1 < High-Iso2 in UB, p < 0.01). The elevation effect seemed to be sustained with the UB condition, but restricted to the odour manipulation in the FB condition. The $Line \times Session \times Odour \times Temperature interaction was$ not significant [F(4, 112) = 0.3, p = 0.87]. Descriptive statistics for the two Lines across Sessions and Odour conditions are reported in Table 1.

No differences between lines were found for latencies to emit the first vocalisations (p > 0.05 for main effect and interactions).

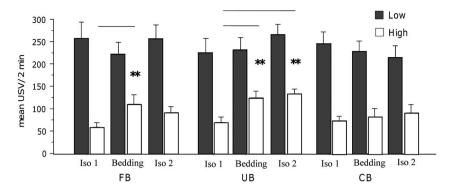


Fig. 3. Line interaction × Session × Odour condition. Mean (\pm SE) of USVs emitted by Low and High Lines during the three-session test and the exposure to the three odour conditions. CB, clean bedding; FB, familiar bedding; UB, unfamiliar bedding; USV, ultrasonic vocalisation. **p < 0.01.

Table 1. Descriptive statistics

	lsol		Bedding		lso2	
	L	Н	L	Н	L	Н
FB	257/143	65/27	221/109	113/79	255/124	99/58
UB	225/133	76/42	231/112	124/57	265/98	132/46
СВ	245/108	79/29	228/92	91/52	214/101	97/49

CB, clean bedding; FB, familiar bedding; UB, unfamiliar bedding.

Mean/SD of USVs emitted by Low (L) and High (H) Lines during the three-session test and the exposure to the three odor conditions.

Discussion

Previous work with these selectively bred lines has found that Low Line pups emit more USVs than High Line pups during a single isolation session (20). Our results confirm those data and extend the findings to new testing conditions using a novel nest-cue potentiation paradigm within the isolation– reunion–reisolation test procedure.

The sensitivity threshold to isolation does not differ between the two lines, as evidenced by a lack of difference in initial USV emission latencies following isolation. However, when comparing the total number of USVs, it becomes clear that the two lines differ in their overall response to a stressful condition, i.e., isolation. Low Line pups in all conditions consistently vocalised at higher levels than High Line pups, suggesting an enhanced responsiveness to negative stimuli in the Low Line animals, which were selectively bred for a lower responsiveness to positive stimuli. In addition, at this age and under these experimental conditions, the two lines responded comparably to variations in our thermal stressor, with both lines vocalising at higher rates at cool temperatures, revealing comparable susceptibility to physical distress, which suggests that these animals were differentially sensitive to social-emotional stress in particular.

Moreover, the lines varied in their responses to the presentation of a social odour. Low Line animals did not alter the amount of separation calls in response to any odours, while High Line animals, contrary to our prediction, increased the rate of calling during exposure to both social odours (FB and UB) and, accordingly with our prediction, the subsequent isolation (Iso2), although the increase from Iso1 to Iso2 was not quite significant in the FB condition. High Line animals did not show this 'potentiation' effect when exposed to a neutral odour (CB). The increase of USVs in the presence of social odours (but not in presence of the control odour) partially resembles the vocal behaviour of another rodent, the Octodon degus, under similar testing conditions (33). In that study, given a choice between FB and UB material, 12-day-old degus easily discriminated between the two kinds of bedding and showed a clear preference for FB. Interestingly, young degus emitted higher rates of separation calls when on FB, as compared with UB (33). Also, when tested in an isolation-reunion-reisolation paradigm, where animals were either exposed to FB or a social partner (mother or unfamiliar female) between isolations, degu pups exposed to bedding continued to vocalise, but those in the social partner conditions did not (T. Fuchs, personal observation). It seems reasonable that the odour cue serves as an indicator of proximity to the nest, stimulating the pups to vocalise, perhaps as an adaptation to request attention from a possibly nearby mother or caregiver. Our results partially corroborate this explanation in that High Line animals increased USVs in the presence of social odour, and this effect also persisted into the second isolation, though the increase was significant only for the UB condition. However, the High Line response to general social odour does not necessarily suggest that they form the maternal bonds that degus form. In fact, their similar response to both familiar and unfamiliar odours may speak against the capacity

of infant rats to 'bond' to their home environments, even though this may happen at older ages when they are motorically mature enough to become lost (34). At that age, they may be more comparable to precocial species that show clear social bonding in infancy when 'proximal separation' of infants from mothers (e.g., being separated by a barrier) typically elevates separation distress type calls (35–37).

The observed lack of USV potentiation in response to odour cues in Low Line animals could be due to a diminished responsiveness to social reward (social odour), as indicated by the low rates of 50 kHz USVs emitted in a variety of other, rewarding paradigms (7,21). Since, in our experiment the Low Line pups already had a high rate of vocalisation in Iso1, it would have been reasonable to expect that the social reward of the smell of mother's bedding might result in a reduction of calls. But this did not happen, suggesting a diminished ability for Low Line animals to experience social reward. Similarly to the High Line, we do not wish to suggest that they also showed diminished social bonding because, to the best of our knowledge, infant rats may not really bond to their mothers or homes as robustly as other mammals, perhaps because of reduced selection pressure under historically long-term laboratory breeding, and/or because their altricial status does not permit them to get lost on their own (38).

How do our results relate to the one other socialvocalisation breeding programme that has been undertaken? As mentioned earlier. Brunelli and Hofer (16) developed bidirectional rat lines selected for high and low emissions of social isolation calls. Interestingly, they evidenced that their Lines exhibited differences in play behaviour and 50 kHz USVs during adolescence, with their High Line showing a depression of both pinning and 50 kHz USVs. Although a direct data comparison between the 50 kHz emitted during tickling and the isolation distress calls in response to social isolation is not possible, it is noteworthy that data from Brunelli and Hofer's high-isolation-call rats resemble results from our Low Line animals, especially in terms of sensitivity to first social isolation (i.e., total number of vocalisations emitted during the first isolation). Brunelli's group also found that USV differences detected in early infancy during isolation tests predicted divergent coping styles in later developmental stages (5,39,40). Our results are consistent with that observation: the offspring of animals selected in the juvenile period for a positive social affective trait (high 50 kHz USVs) were more resistant to separation distress. Related studies have indicated that the High Line animals are resistant to negative affective challenges (21).

This suggests that both breeding lines have selected for stable affective-temperamental traits, presumably within presently unidentified brain emotional networks, which may be important for understanding human social-affective temperaments.

Notably, low positive affectivity in children is a key factor in increased risk of developing psychological disturbances such as anxiety and depression (41), particularly following adverse life experiences. Nonetheless, little attention has been devoted to the study of positive affectivity as a protective factor against adversities throughout the lifespan. Along these lines, it is noteworthy that the main classic temperamental theories, while focusing on traits associated with negative affectivity as correlates of psychopathology (42,43), may have underestimated the protective role that constitutional positive affectivity plays in psychopathology during development. Indeed, in recent human temperamental models and emotional problems, high positive affectivity has been considered a stable trait and a protective factor in youth against the emergence of psychopathological symptoms (44). From this perspective, further elucidation of the role of positive emotionality across the life-span, as modelled by our Low and High 50 kHz USV genetically selected lines, may help shed some light on how constitutional positive and negative emotional processes of the brain may regulate susceptibility to affective disorders and other psychiatric problems in later life.

A major limitation of the present work was our inability to include a randomly selected line of animals, since that line was exhibiting inbreeding sterility. We note that in a few more generations, both the High and Low Line animals exhibited similar breeding problems. We harvested brains and other bodily tissues from all lines for further analysis.

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Authors contributions: J.P. and P.I. conceived and designed the experiment, P.I. ran the experiment. All the authors analysed and interpreted the data, drafted the manuscript, and critically revised it.

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References

- BURGDORF J, PANKSEPP J, BRUDZYNSKI SM, KROES R, MOSKAL JR. Breeding for 50-kHz positive affective vocalization in rats. Behav Genet 2005;35:67–72.
- WÖHR M, DAHLHOFF M, WOLF E, HOLSBOER F, SCHWARTING RK, WOTJAK CT. Effects of genetic background, gender, and early environmental factors on isolation-induced ultrasonic calling in mouse pups: an embryo-transfer study. Behav Genet 2008;38:579–595.
- MOLES A, KIEFFER BL, D'AMATO FR. Deficit in attachment behavior in mice lacking the mu-opioid receptor gene. Science 2004;304:1983–1986.
- STEIMER T, LA FLEUR S, SCHULZ PE. Neuroendocrine correlates of emotional reactivity and coping in male rats from the roman high (RHA/Verh)-and low (RLA/Verh)avoidance lines. Behav Genet 2001;27:503–512.
- HOFER MA, SHAIR HN, MASMELA JR, BRUNELLI SA. Developmental effects of selective breeding for an infantile trait: the rat pup ultrasonic isolation call. Dev Psychobiol 2001;39:231–246.
- 6. DICHTER GS, BRUNELLI SA, HOFER MA. Elevated plus-maze behavior in adult-offspring of selectively bred rats. Physiol Behav 1996;**60**:299–304.
- KNUTSON B, BURGDORF J, PANKSEPP J. Ultrasonic vocalizations as indices of affective states in rats. Psychol Bull 2002;128: 961–977.
- PANKSEPP J. Beyond a joke: from animal laughter to human joy? Science 2005;308:62–63.
- 9. PANKSEPP J. Neuroevolutionary sources of laughter and social joy: modeling primal human laughter in laboratory rats. Behav Brain Res 2007;**182**:231–244.
- BURGDORF J, KROES RA, MOSKAL JR, PFAUS JG, BRUDZYNSKI SM, PANKSEPP J. Ultrasonic vocalizations of rats (Rattus norvegicus) during mating, play, and aggression: behavioral concomitants, relationship to reward, and self-administration of playback. J Comp Psychol 2008;122:357–367.
- BORTA A, WÖHR M, SCHWARTING RK. Rat ultrasonic vocalization in aversively motivated situations and the role of individual differences in anxiety-related behavior. Behav Brain Res 2006;166:271–280.
- NITSCHKE W, BELL RW, BELL NJ, ZACHMAN T. The ontogeny of ultrasounds in two strains of Rattus norvegicus. Exp Aging Res 1975;1:229–243.
- 13. EHRET G. Infant rodent ultrasounds-A gate to the understanding of sound communication. Behav Genet 2005;**35**:19–29.
- 14. HOFER MA. Multiple regulators of ultrasonic vocalization in the infant rat. Psychoneuroendocr 1996;**21**:203–217.
- SHAIR HN. Acquisition and expression of a socially mediated separation response. Behav Brain Res 2007;182:180–192.
- BRUNELLI SA, HOFER MA. Selective breeding for infant rat separation-induced ultrasonic vocalizations: developmental precursors of passive and active coping styles. Behav Brain Res 2007;182:193–207.
- PANKSEPP J. The riddle of laughter: Neural and psychoevolutionary underpinnings of joy. Curr Dir Psychol Sci 2000;9:183–186.
- PANKSEPP J, BURGDORF J. The neurobiology of positive emotions. Neurosci Biobehav Rev 2006;30:173–187.
- BURGDORF J, WOOD PL, KROES RA, MOSKAL JR, PANKSEPP J. Neuro-biology of 50-kHz ultrasonic vocalizations in rats: electrode mapping, lesion, and pharmacology studies. Behav Brain Res 2007;182:274–283.

- HARMON KM, CROMWELL HC, BURGDORF J et al. Rats selectively bred for low levels of 50 kHz ultrasonic vocalizations exhibit alterations in early social motivation. Dev Psychobiol 2008;50:322–331.
- BURGDORF J, PANKSEPP J, BRUDZYNSKI SM et al. The effects of selective breeding for differential rates of 50-kHz ultrasonic vocalizations on emotional behavior in rats. Dev Psychobiol 2009;51:34–46.
- MU P, FUCHS T, SAAL DB, SORG BA, DONG Y, PANKSEPP J. Repeated cocaine exposure induces sensitization of ultrasonic vocalization in rats. Neurosci Lett 2009;453:31–35.
- HOFER MA, MASMELA JR, BRUNELLI SA, SHAIR HN. The ontogeny of maternal potentiation of the infant rats' isolation call. Dev Psychobiol 1998;33:189–201.
- SHAIR HN, MASMELA JR, BRUNELLI SA, HOFER MA. Potentiation and inhibition of ultrasonic vocalization of rat pups: regulation by social cues. Dev Psychobiol 1997;30:195–200.
- HOFER MA, BRUNELLI SA, MASMELA J, SHAIR HN. Maternal interactions prior to separation potentiate isolation-induced calling in rat pups. Behav Neurosci 1996;110:1158–1167.
- HOFER MA, BRUNELLI SA, SHAIR HN. Potentiation of isolation-induced vocalization by brief exposure of rat pups to maternal cues. Dev Psychobiol 1994;27:503–517.
- 27. OSWALT GL, MEIER GW. Olfactory, thermal, and tactual influences on infantile ultrasonic vocalization in rats. Dev Psychobiol 1975;8:129–135.
- BRANCHI I, SANTUCCI D, VITALE A, ALLEVA E. Ultrasonic vocalizations by infant laboratory mice: a preliminary spectrographic characterization under different conditions. Dev Psychobiol 1998;33:249–256.
- CONELY L, BELL RW. Neonatal ultrasounds elicited by odor cues. Dev Psychobiol 1978;11:193–197.
- HOFER MA, SHAIR HN. Independence of ultrasonic vocalization and thermogenic responses in infant rats. Behav Neurosci 1991;105:41–48.
- KRAEBEL KS, BRASSER SM, CAMPBELL JO, SPEAR LP, SPEAR NE. Developmental differences in temporal patterns and potentiation of isolation-induced ultrasonic vocalizations: influence of temperature variables. Dev Psychobiol 2002; 40:147–159.
- 32. WILCOX RR. New Statistical Procedures for the Social Sciences: Modern Solutions to Basic Problems. Hillsdale: Erlbaum, 1987.
- FUCHS T, IACOBUCCI P, MACKINNON KM, PANKSEPP J. Infantmother recognition in a social rodent (*Octodon degus*). J Comp Psychol 2010;124:166–175.
- 34. PANKSEPP J, DEESKINAZI FG. Opiates and homing. J Comp Physiol Psychol 1980;94:650–663.
- WIENER SG, BAYART F, FAULL KF, LEVINE S. Behavioral and physiological responses to maternal separation in squirrel monkeys (Saimiri sciureus). Behav Neurosci 1990;104: 108–115.
- 36. IACOBUCCI P, COLONNELLO V, NEWBERRY RC. Piglets call for mother's attention: Piglet attachment vocal responses are modulated by proximity to their mother. Proceedings of the 41st Annual Meeting of the International Society for Developmental Psychobiology; 12–15 November 2008, Washington DC, USA.
- COLONNELLO V, IACOBUCCI P, PANKSEPP J. Analysis of the disruption of maternal social bonds in *Octodon degus*: separation distress in restricted reunion tests. Dev Psychobiol 2011;53:657–669.

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- PANKSEPP J, NEWMAN JD, INSEL TR. Critical conceptual issues in the analysis of separation distress systems of the brain. In: Strongman KT, editor. International Review of Studies on Emotion, Vol. 2. USA: John Wiley & Sons, 1992.
- 39. BRUNELLI SA, KEATING CC, HAMILTON NA, HOFER MA. Development of ultrasonic vocalization responses in genetically heterogeneous National Institute of Health (N:NIH) rats. I. Influence of age, testing experience and associated factors. Dev Psychobiol 1996;29:507–516.
- BRUNELLI SA. Development and evolution of hidden regulators: selective breeding for an infantile phenotype. Dev Psychobiol 2005;47:243–252.
- FORBES EE, DAHL RE. Neural systems of positive affect: relevance to understanding child and adolescent depression? Dev Psychopathol 2005;17:827–850.
- 42. STELMACK RM. ed. On the Psychobiology of Personality: Essays in Honor of Marvin Zuckerman. Amsterdam: Elsevier Science, 2004.
- ZUCKERMAN M. Psychobiology of Personality, 2nd edn. New York: Cambridge University Press, 2005.
- 44. LONIGAN CJ, PHILLIPS BM, HOOE ES. Relations of positive and negative affectivity to anxiety and depression in children: evidence from a latent variable longitudinal study. J Consult Clin Psychol 2003;71:465–481.