Diversity and genetic structure of *Astronium concinnum* Schott ex Spreng. in conservation units

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Abstract

Astronium concinnum Schott ex Spreng. (Anacardiaceae) is a species used in civil construction, naval, luxury furniture, in addition to the potential for recovery and restoration of habitats. The objective of this work was to characterize the diversity and genetic structure of the *A. concinnum* in the Conservation Units, National Forest of Pacotuba and Private Natural Heritage Reserve of Cafundó, located in the south of the state of Espírito Santo. Eight ISSR primers were used, which produced 121 DNA fragments and 73.55% polymorphism. In the analysis of genetic dissimilarity, seven distinct groups were identified, with the majority of individuals (from both Conservation Units) being brought together into a single group. The genetic diversity of Nei (H^*) and the Shannon index (I^*), provided values for the species of 0.312 and 0.473, respectively, indicating the genetic diversity conserved in the species and its potential use for collecting genetically diversified seeds. The analysis of molecular variance revealed that most of the diversity (92.54%) is distributed within populations and the value of gene flow ($N_m = 10.629$) indicates the high rate of genetic exchange between Conservation Units. The results of the genetic structuring indicated the division of individuals into three genetic groups ($K = 3$), however, it was possible to observe a mixture of genetic material with the sharing of alleles between the three groups. The results indicate that *A. concinnum* trees maintain genetic diversity for their maintenance. In addition, the potential of the analysed individuals was certified as future matrixes for seed collection.

Introduction

Deforestation of natural habitats has eliminated much of the world's forest cover for agricultural and urban development, causing a reduction in the available area and the spatial isolation of the remaining populations. As a result, evolutionary processes such as genetic drift have become more striking due to population reduction and limited gene flow between fragments (Haddad *et al.*, 2015). The flow of genes within and between populations is the main influence on patterns of genetic differentiation, being responsible for maintaining variability within populations, reducing allelic differences between them (Hamrick, 2012).

Habitat loss and fragmentation can affect tree species differently. Spatially isolated trees are not always reproductively isolated, as they are capable of maintaining gene flow even hundreds of metres apart (Martins *et al.*, 2016). Thus, the hypothesis is that trees have reproductive characteristics that make them highly resistant to anthropic processes, favouring the maintenance of high levels of genetic variability (Aguilar *et al.*, 2008).

However, if the gene flow is practically absent, or ecological aspects are not finding that facilitated the maintenance of the species, the isolated individuals may disappear in the fragmented populations (Fischer *et al.*, 2010). Thus, assessing the levels of genetic variation within and among populations is essential to understand the genetic structure of species and their pattern of local maintenance and conservation. This knowledge is important for designing strategies for *in situ* conservation of species and for planning seed collections and production of seedlings of native species, which can be used in *ex situ* conservation in forest restoration programmes and, in connection of areas (Schmidt *et al.*, 2019).

Molecular DNA markers stand out as effective tools for studies to assess the diversity and genetic structure of species and populations (Grover and Sharma, 2016). Among the marker classes, Inter Simple Sequence Repeats (ISSR) it has been widely used due to their

reproducibility, universality, high polymorphism, ease of use and low cost (Ng and Tan, 2015). A collection of publications on tree species from different botanical families reveal that ISSR markers have been efficient for studies of characterization, identification of genetic variations, conservation and forest management, allowing to estimate and detect genetic variability and to characterize the divergence between individuals evaluated (Bocanegra-González and Guillemin, 2018; Fajardo *et al.*, 2018; Maheswarappa *et al.*, 2019; Rajalakshmi *et al.*, 2019; Yulita *et al.*, 2020).

Astronium concinnum Schott ex Spreng. (Anacardiaceae), popularly known as Gonçalo-Alves, is one of the species of interest for these studies due to its economic and ecological potential. Its wood is used for confections of luxury furniture, window frames, parquet and boards for floors and external works, being also used in construction civil and naval. The species trees can be used successfully in rural landscaping and in the afforestation of parks and large gardens, and can also be used in the recovery and restoration of degraded areas (Lorenzi, 2002). In addition, some characteristics of *A. concinnum*, such as entomophilic pollination, anemochoric dispersion, seedlings with high germination rate and shade tolerance, suggest that, in addition to the local abundance provided by the formed seed bank, the trees may persist for long periods in affected areas by anthropic processes (Daniel *et al.*, 1987; England *et al.*, 2002).

The species has confirmed occurrence in the states of Bahia, Rio Grande do Norte, Sergipe, Espírito Santo, Minas Gerais and Rio de Janeiro (Luz *et al.*, 2020). In the state of Espírito Santo, among the occurrences of the species, the Conservation Units (CU's) of National Forest of Pacotuba (Flona of Pacotuba) stand out with 449.72 hectares and the Private Natural Heritage Reserve of Cafundó (PNHR Cafundó) of 517 hectares. These Units are located in the south of the State and represent important areas of environmental preservation in the region. Flona of Pacotuba is integrated into the federal government's project, Network of Native Forest Seeds, which aims to identify areas of natural forests with potential for seed production through criteria of matrix selection (ICMBIO, 2011).

The two above mentioned CU's have a history of environmental disturbances due to anthropic processes such as logging for firewood, fires, pasture areas in the surrounding area with the possible penetration of cattle on the edge of forest complexes (ICMBIO, 2011). Thus, the objective was to evaluate the diversity patterns and genetic structure of the species, in these two conservation areas, answering the following questions: (1) How is the genetic diversity of *A. concinnum* partitioned in the Conservation Units? (2) Was the species affected by the processes of environmental modification or does it still preserve local levels of genetic diversity satisfactory for its maintenance? (3) Can the evaluated individuals be selected as future matrices for seed collections?

Materials and methods

Study area and sampling

The study was carried out in two populations of *A. concinnum* located in Conservation Units in the south of Espírito Santo, Brazil: The National Forest of Pacotuba (Flona of Pacotuba – 20°45′ S; 41°17′ O), and the Private Natural Heritage Reserve of Cafundó (PNHR Cafundó – 20°43′ S; 41°13′ O) (Fig. 1). Leaf samples were collected from 48 trees of reproductive age, 25 in Flona of Pacotuba and 23 in PNHR Cafundó. In the sampling, young and healthy leaves were prioritized, and a minimum distance of

50 m between individuals to decrease the probability of collecting related trees (Sebbenn, 2002).

DNA extraction

Genomic DNA was extracted based on the method described by Doyle and Doyle (1990), with the addition of proteinase K. After optimization tests of the species' DNA extraction protocols, the addition of proteinase K allowed obtaining genomic material with better values in quantity and purity. The concentration and purity of the total DNA were determined by spectrophotometry in the NanoDrop apparatus (Thermo Scientifc 2000C) and evaluated according to Aguilar *et al.* (2016).

PCR amplification

Eight ISSR primers developed by the University of British Columbia, Vancouver, Canada (UBC 807; 808; 818; 827; 836; 956; 868; 878), were used for the genetic analysis of the 48 samples of *A. concinnum*. Each amplification reaction via PCR (Polymerase Chain Reaction) presented a final volume of 20 µl containing 1× buffer [10 mmol/l of Tris-HCl (pH 8.5) and 50 mmol/l of KCl]; 2.5 mmol/l of $MgCl_2$; 0.8 mmol/l of dNTP; 0.2 mmol/l of primer, 1 unit of Taq DNA polymerase enzyme and 50 ng of DNA. The amplifications were performed in a thermocycler (Applied Biosystems, Veriti model) programmed for 1 initial denaturation cycle at 94°C for 5 min, followed by 35 cycles consisting of three stages: (a) 45 s at 94°C, (b) 45 s at 52°C and (c) 90 s at 72°C, followed by a final extension of 7 min at 72°C. The DNA amplification products were separated by electrophoresis on a 2% agarose gel containing ethidium bromide (0.50 µg/ml) at 94 volts for 4 h. Subsequently, the gels were photographed under UV light in photodocumentation system (ChemiDoc MP Imaging System – Bio Rad). The molecular size of the fragments was determined with a molecular weight marker (*Ladder*) of 100 base pairs. All of these analyses were performed at the Laboratory of Biochemistry and Molecular Biology, Universidade Federal do Espírito Santo, UFES (Brazil).

Statistical analysis

The polymorphic loci were evaluated by the presence (1) and absence (0) of amplified fragments, building a binary matrix. A descriptive analysis of the data was performed by evaluating the total number of bands (TNB), number of polymorphic bands (NPB) and percentage of polymorphic bands (PPB) per primer and size variation of the fragments generated in base pairs (SVF). The genetic dissimilarity between the pairs of individuals, in the two locations, was evaluated fur the complementarity of the Jaccard coefficient. The number of genetic clusters was determined using the Unweighted Pair Group Method using Arithmetic averages (UPGMA). The cut-off point for the obtained dendrogram was established by the method proposed by Mojema (1977). The cophenetic correlation coefficient (CCC) was estimated to evaluate the fit between the distance matrix and the dendrogram. All these analyses were conducted with the aid of the Genes software (Cruz, 2016). The genetic dissimilarity matrix obtained in the Genes programme was used for the graphical analysis of the dendrogram in the statistical software R (R Core Team, 2016) associated with the use of vegan (Oksanen *et al.*, 2018), cluster (Maechler *et al.*, 2019), dendextend (Galili *et al.*, 2020), factorextra (Kassambara and Mundt, 2020), ggpubr (Kassambara, 2020), cowplot (Wilke, 2019), gridExtra (Auguie and Antonov, 2017).
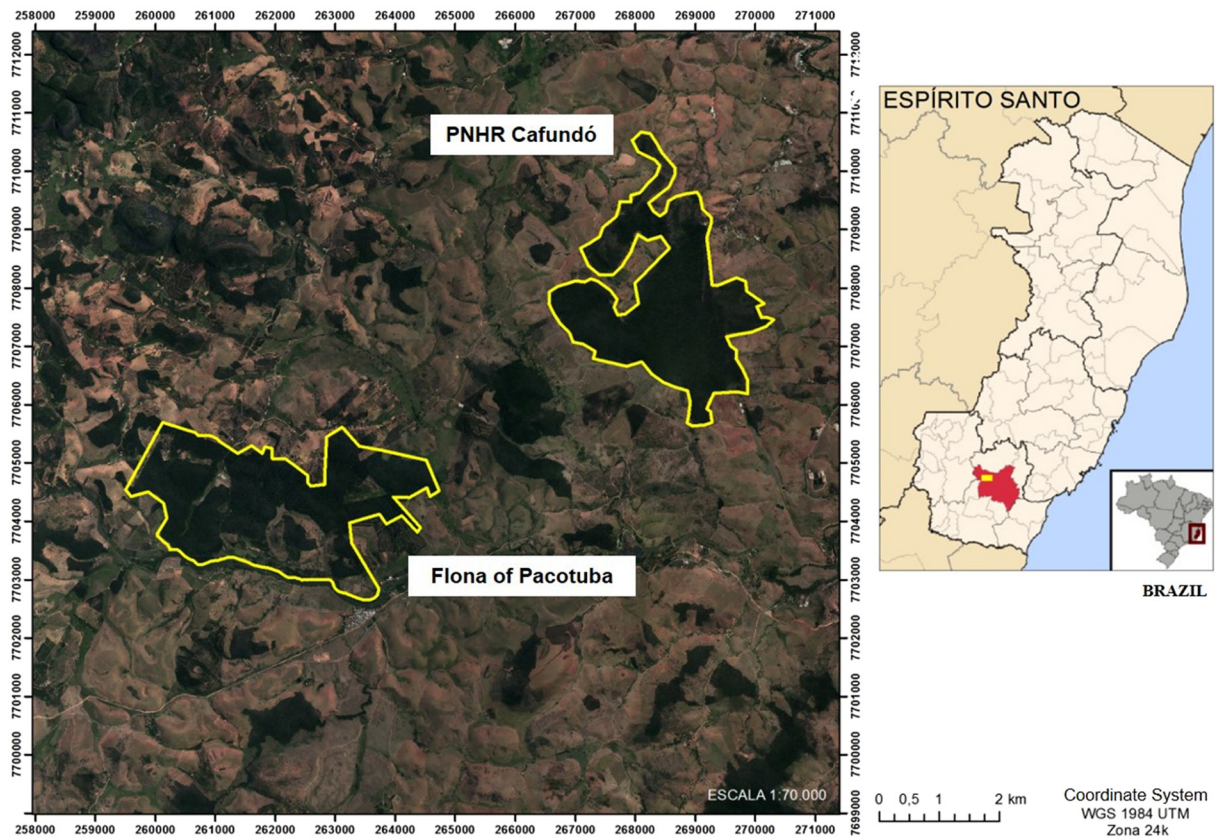
Fig. 1. Location map of the sampling areas of the *Astronium concinnum* species, showing the two Conservation Units evaluated. Espírito Santo, Brazil. *Source*: The author.

The parameters of genetic diversity, percentage of polymorphic loci (PPL), Nei genetic diversity index (H^*) and Shannon index (I^*), were estimated using the Popgene 1.32 software (Yeh and Boyle, 1997). The average number of migrants per generation (N_m), which estimates the gene flow between populations, was also calculated using the same software. This analysis was performed indirectly from the estimates of genetic divergence between groups (G_{ST}) as a function of N_m, using the formula $N_m = 0.5 (1-G_{ST})/G_{ST}$. The distribution of genetic diversity between and within Conservation Units was estimated through the analysis of molecular variance (AMOVA), using the software Arlequin version 3.5 (Excoffier and Lischer, 2010). The Structure software (Pritchard *et al.*, 2000), based on Bayesian statistics, was used to define the number of groups (K) in which individuals are structured. To verify the most probable number of groups, K values ranging from $K = 1$ to $K = 5$ were tested. 20 runs were run for each K value, 250,000 burn-ins and 1,000,000 Markov Chains Monte Carlo simulations (MCMC) (Evanno *et al.*, 2005). The most likely K value was defined using the Structure Harvester software (Earl and Vonholdt, 2012), according to the criteria established by Evanno *et al.* (2005).

Results

The eight ISSR primers used in the 48 individuals of *A. concinnum* produced a total of 121 amplified fragments, among which, 89 (73.55%) presented polymorphism. The number of bands per primer varied between nine (UBC 807) and 24 (UBC 878), with an average of 15.12 bands/primer. The most

informative primer was UBC 878 (75% polymorphism) and the least informative was UBC 856 (60% polymorphism) (Table 1).

The genetic dissimilarity obtained between the trees ranged from 0.26 to 0.87. The smallest genetic distance was observed between pairs of individuals 1 and 2 and the longest distance between 40 and 42. The visual analysis of the data in the dendrogram, adopting a cutoff point of 85.23%, allowed the separation of individuals into seven distinct genetic groups (Fig. 2).

Individuals 3, 36, 42 and 45 formed isolated groups on the dendrogram. Individual 3 belongs to Flona of Pacotuba and the others to PNHR Cafundó. Most individuals (from both CU's) were included in a single group (G1). The cophenetic correlation coefficient was 82% (CCC = 0.82).

The parameters of genetic diversity, Nei and Shannon index, showed similar values between the two analysed fragments. The highest values were observed when the two fragments were evaluated together (species) (Table 2).

AMOVA indicated that most genetic diversity (92.54%) occurs within populations and 7.45% of diversity is among populations. The genetic differentiation (Φ_{st}) between the two locations was $\Phi_{st} = 0.074$ and the gene flow of $N_m = 10.629$ (Table 3).

The results of the genetic structuring indicated the division of the individuals of *A. concinnum* into three genetic groups ($K = 3$) identified by three colours (blue, green and red) in the figure below (Fig. 3).

Discussion

The polymorphism levels for *A. concinnum* followed the pattern found for natural populations and dominant markers, suggesting

Table 1. Primers used and their base sequences, the total number of bands (TNB), the number of polymorphic bands (NPB), percentage of polymorphic bands (PPB) and size variation of the fragments generated in base pairs (SVF) based on 100 bp marker

Primer	Sequences (5′-3′)	TNB	NPB	PPB %	SVF (min–máx)
UBC 807	$(AG)_8T$	9	7	77.77	320–900
UBC 808	$(AG)_8C$	16	11	68.75	200–1550
UBC 818	$(CA)_8G$	15	11	73.33	400–2080
UBC 827	$(AC)_8G$	16	12	75	420–2040
UBC 836	$(AG)_8YA$	18	16	88.88	210–2080
UBC 856	$(AC)_8YA$	10	6	60	300–2080
UBC 868	$(GAA)_6$	13	8	61.53	250–2080
UBC 878	$(GGAT)_4$	24	18	75	300–2080
Total	–	121	89	73.55	–

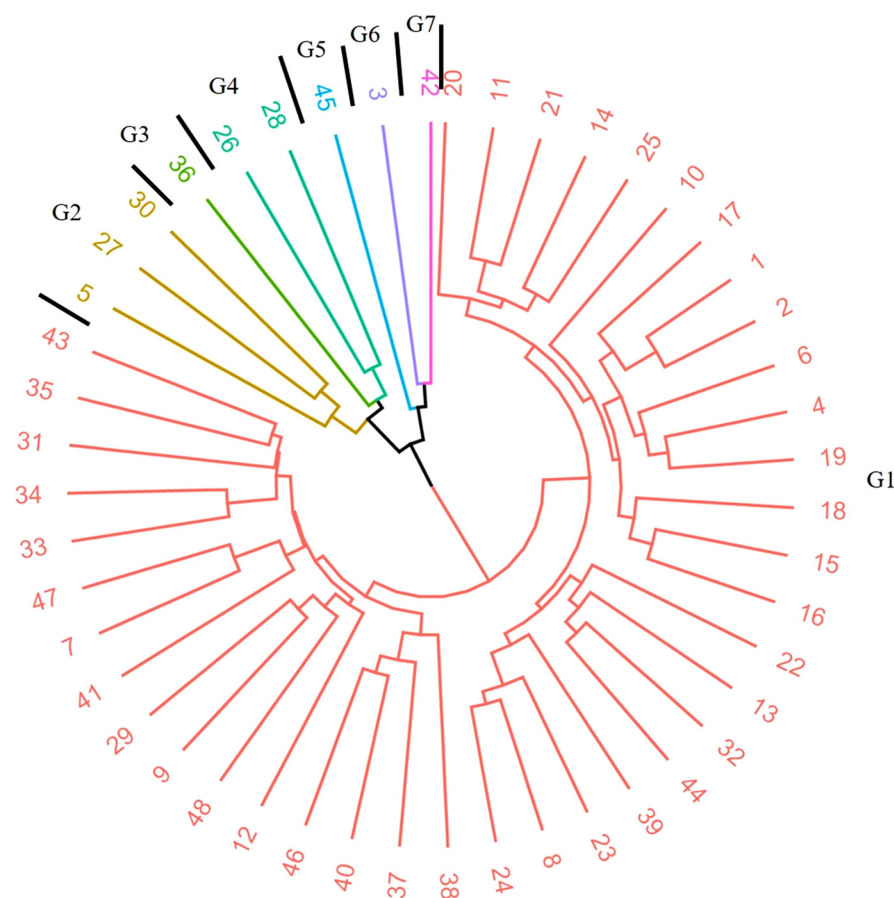A, Adenine; T, Thymine; C, cytosine; G, Guanine; Y, (C or T).

Fig. 2. Dendrogram representative of genetic dissimilarity among 48 individuals of *Astronium concinnum*. Samples 1 to 25 (Flona of Pacotuba, ES), samples 26 to 48 (PNHR Cafundó, ES). The seven groups formed (G1 to G7) are identified by different colours.

Table 2. Genetic diversity of natural populations of *Astronium concinnum* sampled in Flona of Pacotuba and PNHR Cafundó, Espírito Santo

Population	N	H*	I*
Flona of Pacotuba	25	0.298	0.455
PNHR Cafundó	23	0.297	0.456
Species	48	0.312	0.474

Sample size (*n*), Nei Index (*H**), Shannon Index (*I**).

high genetic variability due to the PPB (73.55%). The literature provides variations in the percentage of polymorphism found among different trees studied with ISSR markers, to mention, *Bouea macrophylla* (50.30%) (Ghazalli *et al.*, 2015); *Pistacia atlantica* (74.10%) (Labdelli *et al.*, 2020) and *Spondias mombin* L. (64.65%) (Silva *et al.*, 2017). Thus, one can affirm the efficiency and reproducibility of ISSR markers in detecting genetic variability among individuals evaluated for *A. concinnum*.

When analysing the graphical representations of genetic distances (Fig. 2), a pattern of heterogeneity is observed with

Table 3. Analysis of molecular variance and number of migrants per generation in natural populations of *Astronium concinnum*

Sources of variation (%)	Degrees of freedom	Sum of squares	Variance components	Variation (%)
Between populations	1	42.60	1.19	7.45
Within populations	46	664.88	14.84	92.54
Total	47	707.49	16.04	
$\Phi_{st} = 0.074$ $N_m = 10.629$				

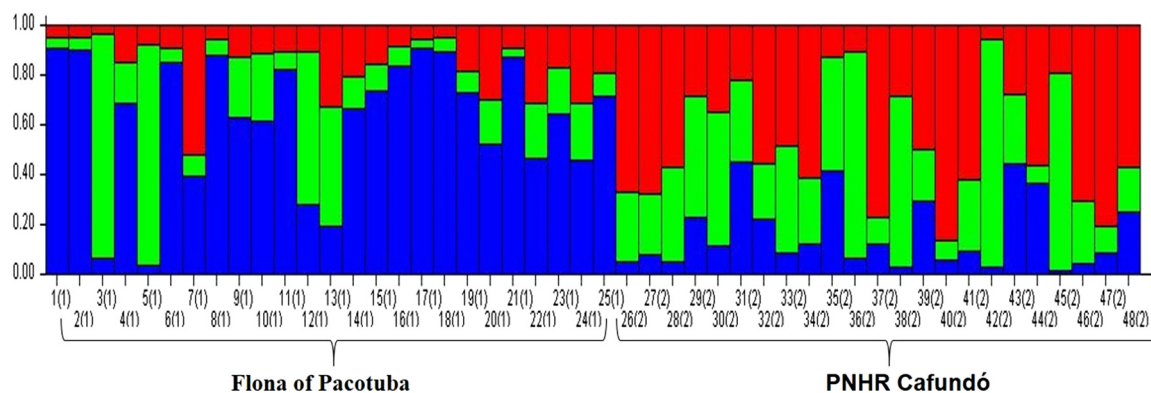Number of migrants per generation (N_m); Genetic differentiation (Φ_{st}).

Fig. 3. Representation of 48 individuals from two natural populations of *Astronium concinnum* using the Structure software. Individuals 1 to 25 (Flona of Pacotuba, ES) and 26 to 48 (PNHR Cafundó, ES) are represented by vertical bars.

the formation of different genetic groups, among them, isolated groups of genetically more distinct trees. However, there was no formation of groups by forest fragment, most of the individuals (from both CU's) were included in a single group, indicating that there is no difference between the two Units evaluated.

The geographical proximity between individuals may have been a factor that influenced the levels of genetic divergence found. Tree species tend to have a pattern of population structure in families, forming demes with homogeneous allele frequencies, due to the dispersion of seeds close to the parent tree (Motta *et al.*, 2004), which increases the probability of collecting related trees among 20–50 m (Sebbenn, 2002). This information explains the results found, the trees with the least divergence belong to Flona of Pacotuba and are at least 50 m apart, on the other hand, the most divergent trees (included in PNHR Cafundó), are at least 100 m away.

The genetic divergence analyses carried out allow to select, among the studied CU's, the most diversified individuals that can be used in the collection and production of seedlings, which besides guaranteeing the *ex situ* maintenance of the species, can be used in the conservation of environments by the processes of forest restoration and recovery (Cortelete *et al.*, 2021).

Studies carried out over the years demonstrate that the ability of a species to adapt in a new environment depends, among other factors, on a broad genetic base (Silva Júnior *et al.*, 2017; Cortelete *et al.*, 2021). Thus, in terms of forest restoration, the sampling of individuals for seed collection must be representative of the genetic variability of the studied population, so that future planting is maintained through crosses between individuals from the population initially established (Sebbenn, 2002).

The minimum number of trees, indicated for seed collection seeking genetic maintenance in natural populations, ranges

from 10 to 20, being suggested that the collection of 13 trees would be sufficient to retain the effective size of 50 (Sebbenn, 2002). The most appropriate strategy, according to the authors, would be the selection of a minimum number of trees, based on prior knowledge about the genetic variability of individuals in a population.

Thus, for the production of seedlings for forest restoration and restoration in the study region, it is suggested the formation of seed lots from trees 3, 5, 7, 9, 10, 12, 13, 18, 20, 22, 26, 28, 30, 32, 34, 36, 38, 40, 42, 45 considered in this study as the most genetically divergent. A seed lot from these 20 trees, which have a high dissimilarity value with each other, will have genetic variability representative of the species in the two Conservation Units evaluated.

The values of the intrapopulation genetic diversity indices can be considered from moderate to high for *A. concinnum*. This information is confirmed when the results are compared to representatives of Anacardiaceae and species with related ecological aspects. In addition, the value of the Shannon index varies from 0 to 1, the closer to 1, the greater the genetic diversity (Lewontin, 1972). In studies with *Spondias mombin* L. (Anacardiaceae) values of 0.26 were found for the Nei index (H^*) and 0.39 for the Shannon index (I^*) (Silva *et al.*, 2017). Analysis of the genetic structure and diversity of *Anacardium excelsum* (Anacardiaceae) revealed values of $H^* = 0.23$ and $I^* = 0.38$ (Bocanegra-González and Guillemin, 2018). Souza *et al.* (2018) found $H^* = 0.38$ and $I^* = 0.55$ in their studies with *Plathymenia reticulata*. In analyses of the genetic diversity of *Dalbergia latifolia*, Yulita *et al.* (2020) obtained values of $H^* = 0.16$ and $I^* = 0.25$.

Although both CU's have a history of landscape modification by anthropization that can promote erosion of genetic diversity (Icmbio, 2011), it is suggested that such events have not affected

the genetic diversity of *A. concinnum*. Longer life cycles, abundant seed bank and highly surviving seedlings prevent tree species from being affected by anthropic processes over a long period of time (England *et al.*, 2002). *A. concinnum* has seedlings with a high germination rate and tolerance to shaded environments, its seeds can overcome strong restrictions of the environment, such as water deficiency, allowing the formation of an abundant seed bank (Daniel *et al.*, 1988). These characteristics probably allowed the species to maintain high genetic diversity for its maintenance in the two Conservation Units.

The AMOVA indicated moderate genetic differentiation (Φ_{st} = 0.074), according to Wright (1978), between the two CU's, being observed that most of the genetic diversity (92.54%) occurs within populations. In allogamous plants such as *A. concinnum*, the greatest genetic variation has been observed within populations (Brandão *et al.*, 2011; Ferreira *et al.*, 2012; Ramalho *et al.*, 2016) and, may indicate the ability to preserve genetic variability to be explored in forest genetic improvement.

Genetic differences between populations decrease as the gene flow increases via pollen and/or seeds. *A. concinnun* is a dioecious species of entomophilous pollination, and the dispersion of the seeds is carried out by the wind (Daniel *et al.*, 1987). Flona of Pacotuba and PNHR Cafundó show considerable similarity in the characteristics of the landscape and climate and are geographically very close (about 1.5 km) (Alvares *et al.*, 2013). Such characteristics, combined with the reproductive and ecological aspects of the species, are certainly contributing to the exchange of genetic material between the two locations.

Therefore, the moderate differentiation found shows that there is no genetic separation between from Flona of Pacotuba and PNHR Cafundó, nor a reproductive subdivision between the two sampled locations, due to the occurrence of gene flow (N_m). Wright (1951) described that the value of the average number of migrants per generation (N_m) must be greater than 1 ($N_m > 1$) to avoid differentiation between groups. In the species evaluated, a value of 10.62 was found for N_m, indicating a high rate of gene flow between the two locations, and, consequently, less differentiation between them. This result corroborates with those obtained in the analyses of genetic dissimilarity (dendrogram), where there was no formation of groups of individuals separated by forest fragment.

It is important to point out that the genetic material evaluated corresponds to long-lived adult *A. concinnun* individuals, and thus, the calculated gene flow refers to a historical estimate, reflecting past events that led to current patterns of genetic structure. To understand the implications of current environmental changes on genetic diversity indexes, studies with progeny tests would be more appropriate (Kageyama *et al.*, 2003).

The analysis of the genetic structure reinforces the results found and indicates a greater structuring in Flona of Pacotuba, characterized by a greater distribution of the blue group (63.30%), followed by the red (18.50%) and green (18.20%) groups. In contrast, for the PNHR Cafundó the distribution was 45.70% for the red group, 36.20% for the green group and 18.10% for the blue group. Although these three distinct groups were formed (Fig. 3), between the trees and in both forest fragments there is a mixture of genetic material, where all individuals share a genome between the three groups. This structure corroborates that between the two locations there is an exchange of genetic material and that there is no great genetic difference between the Conservation Units, since there was no separation of genetic groups by location.

The moderate differentiation between the locations can also be confirmed by the groups formed. It is possible to observe a contrast of colour distribution in the graph, the blue group was more prominent in the population of Flona of Pacotuba and the red group in PNHR Cafundó. This aspect indicates that there is a moderate genetic differentiation between the two CU's, however, it was not enough to separate the two locations into different groups due to the high rates of gene flow existing between them.

Some inferences can be highlighted, from the results and the pattern of diversity and genetic structure found for *A. concinnun*. The use of ISSR markers can be recommended in similar research, as they were efficient in discriminating and characterizing the individuals evaluated, allowing them to estimate and detect the genetic variability between them.

The conservation units Flona of Pacotuba and PNHR Cafundó have satisfactory levels of local diversity of the species, being important sources of genetic variability that can be exploited in forest genetic improvement. The genetic diversity found can be used for the selection of individuals with the potential for seed collection for recovery and restoration of degraded environments.

The collections must be prioritized in the trees that present greater genetic dissimilarity, since from them, it will be possible to form lots of seeds that will represent the existing genetic variability. It is recommended, in future research studies, the collection of the 20 trees mentioned here, with the highest dissimilarity value, as they are representative of the genetic variability of the species in the two Conservation Units evaluated, they are in considerable geographical distance to decrease the likelihood of kinship and will allow the effective size of populations to remain high.

Conclusion

Astronium concinnum trees maintain high genetic diversity for their maintenance and potential for future use as matrixes for seed collection in the Conservation Units evaluated, these being important sites for *in situ* conservation of the species.

References

Aguilar R, Quesada M, Ashworth L, Herrerias-Diego Y and Lobo J (2008) Genetic consequences of habitat fragmentation in plant populations: susceptible signals in plant traits and methodological approaches. *Molecular Ecology* 17, 5177–5188.

Aguilar LG, López AMS, Aceitun CB, Ávila JAC, Guerreiro JAL and Quesada R (2016) A DNA source selection for downstream applications based on DNA quality indicators analysis. *Biopreservation and Biobanking* 14, 264–270.

Alvares CA, Stape JL, Sentelhas PC, Moraes Gonçalves JL and Sparovek G (2013) Köppen's climate classification map for Brazil. *Meteorologische Zeitschrift* 22, 711–728.

Auguie B and Antonov A (2017) Miscellaneous functions for 'Grid' graphics. R package version 2.3. Available at https://cran.r-project.org/web/packages/gridExtra/gridExtra (Accessed 24 April 2020).

Bocanegra-González KT and Guillemin ML (2018) Guidelines for the restoration of the tropical timber tree *Anacardium excelsum*: first input from genetic data. *Tree Genetics & Genomes* 14, 1–59.

Brandão MM, Vieira FA and Carvalho D (2011) Estrutura Genética em Microescala Espacial de *Myrcia splendens* (Myrtaceae). *Revista Árvore* **35**, 957–964.

Cortelete MA, Silva Júnior AL, Pereira MLS, Miranda FD and Caldeira MVW (2021) Molecular characterization as strategy for ex situ conservation of *Anadenanthera peregrina* (L.) Speg. *Scientia Forestalis* **49**, e3443.

Cruz CD (2016) Genes software – extended and integrated with the R, Matlab and Selegen. *Acta Scientiarum* **38**, 547–552.

Daniel O, Reis MGF, Maestri M, Reis GG and Regazzi AJ (1987) Determinação dos padrões de disseminação de *Astronium concinnum* Schot (Gonçalo-alves). *Revista Árvore* **11**, 119–131.

Daniel O, Reis MGF, Maestri M and Reis GG (1988) Germinação de sementes e sobrevivência inicial de plântulas de *Astronium concinnum* Schott (Gonçalo-alves) em condições naturais. *Revista Árvore* **12**, 196–208.

Doyle JJ and Doyle JL (1990) Isolation of plant DNA from fresh tissue. *Focus* **12**, 13–15.

Earl DA and Vonholdt BM (2012) Structure harvester: a website and program for visualizing structure output and implementing the Evanno method. *Conservation Genetics Resources* **4**, 359–361.

England PR, Usher AV, Whelan RJ and Ayre DJ (2002) Microsatellite diversity and genetic structure of fragmented populations of the rare, fire-dependent shrub *Grevillea macleayana. Molecular Ecology* **11**, 967–977.

Evanno G, Regnaut S and Goudet J (2005) Detecting the number of clusters of individuals using the software structure: a simulation study. *Molecular Ecology* **14**, 2611–2620.

Excoffier L and Lischer HEL (2010) Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources* **10**, 564–567.

Fajardo CG, Costa DFD, Chagas KPTD and Vieira FDA (2018) Genetic diversity in natural populations of *Hancornia speciosa* Gomes: implications for conservation of genetic resources. *Ciência e Agrotecnologia* **42**, 623–630.

Ferreira CBB, Lopes MTG, Lopes R, Cunha RNVD, Moreira DA, Barros WS and Matiello RR (2012) Diversidade genética molecular de progênies de dendezeiro. *Pesquisa Agropecuária Brasileira* **47**, 378–384.

Fischer J, Stott J and Law BS (2010) The disproportionate value of scattered trees. *Biological Conservation* **143**, 1564–1567.

Galili T, Benjamini Y, Simpson G, Jefferis G, Gallota M, Renaudie J, Hornik K, Ligges U, Spiess A, Horvath S, Langfelder P, Loo MVD, Vries A, Gu Z, Ma JGK, Hummel M, Clark C, Graybuck L, Ho B, Perreault S, Hennig C and Bradley D (2020) Extending 'dendrogram' functionality in R. R package version 1.13.4. 2020. Available at https://cran.r-project.org/web/packages/dendextend/dendextend (Accessed 24 April 2020).

Ghazalli MN, Yunus MF and Mohammad AL (2015) Assessment of genetic relationships within *Bouea* (Anacardiaceae) accessions in Peninsular Malaysia using inter simple sequence repeats (ISSR) markers. *African Journal of Biotechnology* **14**, 76–85.

Grover A and Sharma PC (2016) Development and use of molecular markers: past and present. *Critical Reviews in Biotechnology* **36**, 290–302.

Haddad NM, Brudvig LA, Clobert J, Davies KF, Gonzalez A, Holt RD, Lovejoy TE, Sexton JO, Austin MP, Collins CD, Cook WM, Damschen EI, Ewe RM, Foster BL, Jenkins CN, King AJ, Laurance WF, Levey DJ, Margules CR, Melbourne BA, Nicholls AO, Orrock JL, Canção DX and Townshend JR (2015) Habitat fragmentation and its lasting impact on Earth's ecosystems. *Science Advances* **1**, 1–10.

Hamrick JL (2012) Tropical breeding systems: one and done? *Heredity* **109**, 330–331.

ICMBIO (2011) *Plano de Manejo da Floresta Nacional de Pacotuba, localizada no estado do Espírito Santo*. Vila Velha: Instituto Chico Mendes De Conservação Da Biodiversidade.

Kageyama PY, Sebbenn AM, Ribas LA, Gandara FB, Castellen M, Perecim MB and Vencovsky R (2003) Diversidade genética em espécies arbóreas tropicais de diferentes estágios sucessionais por marcadores genéticos. *Scientia Forestalis* **64**, 93–107.

Kassambara A (2020) 'ggplot2' Based publication ready plots. R package version 0.4.0. 2020. Available at https://cran.r-project.org/web/packages/ggpubr/ggpubr (Accessed 24 April 2020).

Kassambara A and Mundt F (2020) Factoextra: extract and visualize the results of multivariate data analyses. R package version 1.0.5. 2017. Available at https://CRAN.R-project.org/package=factoextra (Accessed 24 April 2020).

Labdelli A, De La Herrán R, Arafeh R, Resentini F, Trainotti L, Halis Y, Adda A and Merah O (2020) Genetic variation in damaged populations of Pistacia Atlantica Desf. *Plants* **9**, 1541.

Lewontin RC (1972) The apportionment of human diversity. *Evolutionary Biology* **6**, 381–398.

Lorenzi H (2002) *Árvores Brasileiras: manual de identificação e cultivo de plantas arbóreas do Brasil*. Nova Odessa: Instituto Plantarum, p. 368.

Luz CLS, Mitchell JD, Pirani JR and Pell SK (2020) Anacardiaceae in Flora do Brasil 2020 em construção. Jardim Botânico do Rio de Janeiro. Available at http://reflora.jbrj.gov.br/reflora/floradobrasil/FB4383 (Accessed 6 April 2020).

Maechler M, Rousseeuw P, Struyf A, Hubert M, Hornik K, Studer M, Roudier P, Gonzalez J, Kozlowski K, Schubert E, Murphy K (2019) Finding groups in data: cluster analysis extended Rousseeuw *et al* R package version 2.1.0. 2019. Available at https://cran.r-project.org/web/packages/cluster/cluster (Accessed 24 April 2020).

Maheswarappa V, Vasudeva R, Hegde R and Ravikanth G (2019) ISSR Analysis of genetic diversity in *Acrocarpus fraxinifolius* from three landscape elements of transition forest belt of Kodagu district, Karnataka, India. *International Journal of Current Microbiology and Applied Sciences* **8**, 1611–1624.

Martins K, Kimura RK, Francisconi AF, Gezan S, Kainer K and Christianini AV (2016) The role of very small fragments in conserving genetic diversity of a common tree in a hyper fragmented Brazilian Atlantic forest landscape. *Conservation Genetics* **17**, 509–520.

Mojema R (1977) Hierarchical grouping methods and stopping rules: an evaluation. *The Computer Journal* **20**, 359–363.

Motta JS, Teixeira MLM and Sebbenn AM (2004) Autocorrelação espacial em população natural de *Terminalia argentea* Mart et Succ. no cerrado de Selvíria, MS. *Scientia Forestalis* **66**, 94–99.

Ng WL and Tan SG (2015) Inter-Simple Sequence Repeat (ISSR) markers. *ASM Science Journal* **9**, 30–39.

Oksanen J, Blanchet FG, Friendly M, Kindt R, Legendre P, Mcglinn D, Minchin PR, O'hara RB, Simpson GL, Solymos P, Stevens MHH, Szoecs E and Wagner E (2018) Vegan: community ecology package. R package version 2.4-5. 2018. Available at https://CRAN.R-project.org/package=vegan (Accessed 24 April 2020).

Pritchard JK, Stephens M and Donnelly P (2000) Inference of population structure using multilocus genotype data. *Genetics* **155**, 945–959.

Rajalakshmi R, Rajalakshmi S and Parida A (2019) Genetic diversity, population structure and correlation study in *Moringa oleifera* Lam. using ISSR and SRAP markers. *Proceedings of the National Academy of Sciences* **89**, 1361–1371.

Ramalho AB, Rossi AAB, Dardengo JFE, Zortéa KÉM, Tiago AV and Martins KC (2016) Diversidade genética entre genótipos de *Bertholletia excelsa* por meio de marcadores moleculares ISSR. *Floresta* **46**, 207–214.

R Core Team (2016) *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing.

Schmidt IB, De Urzedo DI, Piña-Rodrigues FCM, Vieira DLM, De Rezende GM, Sampaio AB and Junqueira RGP (2019) Community-based native seed production for restoration in Brazil–the role of science and policy. *Plant Biology* **21**, 389–397.

Sebbenn AM (2002) Número de árvores matrizes e conceitos genéticos na coleta de sementes para reflorestamentos com espécies nativas. *Revista do Instituto Florestal* **14**, 115–132.

Silva B, Rossi AAB, Tiago AV, Schmitt KFM, Dardengo JFE and Souza SAM (2017) Genetic diversity of Cajazeira (*Spondias mombin* L.) in three geographic regions. *Genetics and Molecular Research* **16**, 1–11.

Silva Júnior AL, Souza LC, Pereira AG, Caldeira MVW and Miranda FD (2017) Genetic diversity of *Schizolobium parahyba* var. *amazonicum* (Huber ex. Ducke) Barneby, in a forest area in Brazil. *Genetics and Molecular Research* **16**, gmr16039774.

Souza LC, Silva Júnior AL, Miranda FD, Souza MC, Kunz SH and Pereira AG (2018) Validação do marcador molecular ISSR para detecção de

diversidade genética em *Plathymenia reticulata* Benth. *Revista Brasileira de Ciências Agrárias* **13**, 1–6.

Wilke CO (2019) Streamlined plot theme and plot annotations for 'ggplot2'. R package version 1.0.0. 2019. Available at https://cran.r-project.org/web/packages/cowplot/cowplot (Accessed 24 April 2020).

Wright S (1951) The genetical structure of populations. *Annals of Eugenic* **15**, 395–420.

Wright S (1978) *Evolution and Genetics of Populations.* Chicago: The University of Chicago Press, p. 465.

Yeh FC and Boyle TJB (1997) Population genetic analysis of co-dominant and dominant markers and quantitative traits. *Belgian Journal of Botany* **29**, 157–163.

Yulita KS, Atikah TD, Wardani and W and Sulita (2020) Unraveling genetic variations of *Dalbergia latifolia* (Fabaceae) from Yogyakarta and Lombok Island, Indonesia. *Biodiversitas: Journal of Biological Diversity* **21**, 833–841.