

Environmental parameters as risk factors for human and canine *Leishmania* infection in Thessaly, Central Greece

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SUMMARY

Thessaly, Central Greece, is an endemic area for leishmaniasis with higher incidence rate during the last years. We herein investigated the geographical distribution of human leishmaniasis cases and *Leishmania* infected dogs in relation to environmental parameters to identify high-risk areas. All the human leishmaniasis cases ($n = 82$) reported to Hellenic Centre for Disease Control and Prevention from 2007 to 2014 and 85 *Leishmania* polymerase chain reaction positive dogs were included in this study. To analyse the data geographical information system (GIS) together with the Ecological Niche Model (ENM) were used. The most important findings of the study were: (i) Central plain of Thessaly together with the coast line and the western and eastern lowlands were identified as high-risk geographical areas. (ii) The highest percentage of the high-risk areas was found in low altitude (<200 m above sea level) and in irrigated and cultivated agricultural areas. (iii) A total of 20% of the human settlements was found in high-risk areas. (iv) The maximum temperature of the warmest month contributes the highest per cent to define both environmental niche profiles for humans and dogs. (v) The ENM could be a useful tool for the epidemiological study of leishmaniasis. Spatial analysis may allow the design of entomological studies and identify target population in order to implement preventive measures.

Key words: *Leishmania*, human leishmaniasis cases, dogs, GIS, Greece.

INTRODUCTION

Leishmaniasis is a mandatory notifiable disease in Greece, endemic in most islands and coastal regions of Greece and constitutes a veterinary and public health issue of major concern. From 1998 to 2011, human leishmaniasis cases were recorded in the mainland as well as in the islands of Greece with a mean annual incidence of 0.36 cases per 100 000 population (Gkolfinopoulou *et al.* 2013). Regarding the canine population, the overall reported seroprevalence in seven regions of the Greek mainland was nearly 20% ranging from 2.05% in Florina to 30.12% in Attiki (Athanasiou *et al.* 2012). Studies that have been conducted in

Greece using geographical information system (GIS) mapped the occurrence of human leishmaniasis and related it to dog seropositivity (Ntais *et al.* 2013; Sifaki-Pistola *et al.* 2014). Several studies conducted in other countries produced environmental risk mapping and spatial analysis of canine and human leishmaniasis using GIS, thus showing how an ecological approach can help improve our understanding of the spatial distribution of leishmaniasis (Chamaillé *et al.* 2010; Franco *et al.* 2011; Barón *et al.* 2013; Tsegaw *et al.* 2013; Seid *et al.* 2014).

The previous years increase in the incidence rate of human leishmaniasis cases in Thessaly, Central Greece, prompted us to investigate possible risk factors. The aim of the study was to analyse together human cases and *Leishmania* infected dogs taking into consideration environmental parameters. The overall objective was to provide evidence for target interventions.

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MATERIALS AND METHODS

Study area

Thessaly is located in the central part of Greece and has a total area of 14 036 km², which roughly represents 11% of the whole country. Thirty six per cent of the land is flat and 17% is semi-mountainous, whereas the remaining 45% is mountainous (Domenikiotis *et al.* 2005). The administrative region of Thessaly consists of four prefectures (Larissa, Trikala, Karditsa and Volos). These prefectures include 26 municipalities which are further divided in 545 municipality districts.

Human leishmaniasis cases

All the officially notified human leishmaniasis cases ($n = 82$) reported during 2007–2014 to Hellenic Centre for Disease Control and Prevention (HCDCP) from the treating physicians were included in this study (Supplementary Table S1). The human cases reported to HCDCP were considered and recorded as a human leishmaniasis case when presenting clinical picture compatible with human leishmaniasis and when the protozoan parasite was detected in cytological examination of clinical samples (whole blood and/or bone marrow) (Islam, 2013) or when at least one clinical sample (whole blood and/or bone marrow) was polymerase chain reaction (PCR) positive according to the PCR assays performed in the two different reference laboratories in Greece (Spanakos *et al.* 2002; Christodoulou *et al.* 2012) or when the patient was seropositive (Indirect Fluorescent Antibody Test or Enzyme-linked Immunosorbent Assay or Latex or immunochromatographic test) (with serology titre $\geq 1/160$ for IFAT). In overall, all the human leishmaniasis cases were PCR positive to at least one sample examined or the protozoan parasite was detected in cytological examination of clinical samples except five human cases which were seropositive only. The geo-references for the human cases were obtained by geocoding and they were resolved to municipality district level.

Leishmania infected dogs

Dogs submitted to veterinary clinics in Thessaly, after clinical examination by the private practicing veterinarians, if suspected to be *Leishmania* infected, they were subjected to sample collection. Exfoliative epithelial cells, lymph node aspirates and whole blood in ethylenediamine tetra-acetic acid (EDTA) were collected from the dogs suspected to be *Leishmania* infected and they were sent still frozen to the Laboratory of Microbiology and Parasitology, University of Thessaly, Karditsa, Greece, for molecular investigation. The canine samples were stored at -20°C for pending DNA extraction.

Total genomic DNA extraction was performed on the samples collected from each dog using a commercially available DNA extraction kit (Thermo Scientific GeneJET Genomic DNA Purification Kit) according to the manufacturer's protocol. The purified DNA was stored at -20°C . The samples were analysed by ITS-1 nested PCR (ITS-1 nPCR) as described previously (Leite *et al.* 2010). In overall, 85 stray and owned dogs which were PCR positive to at least one of the samples examined were considered *Leishmania* infected and included in the study (Supplementary Table S2). Data on canine samples were located in the field using hand-held Global Positioning System (GPS) Garmin units. The geo-references were resolved to specific houses level.

Ethics statement

The human data which are being analysed in this study, were part of the on-going surveillance of human cases performed by HCDCP and were reported by the treating physicians. The human data were provided at the municipality district level and were completely anonymized to the authors, without being publicly available. The canine samples included in this study were collected by private practicing veterinarians. No animals were euthanized during the study and efforts taken to ameliorate animal suffering. The study did not involve any experimentation, but was based in samples, that had been collected from the dogs for routine diagnostic purposes. Diagnostic veterinary procedures are not within the context of relevant EU legislation for animal experimentations (Directive 86/609/EC) and may be performed in order to diagnose animal diseases and improve animal welfare.

Environmental parameters

Climatic variables were derived from the WorldClim version 1.4. (Hijmans *et al.* 2005). Land uses and population density were derived from the Corine Land Cover 2000 database (European Environment Agency-EEA). The boundaries of the municipalities/district/community were retrieved from the national open data catalogue (<http://www.geodata.gov.gr>). Distance from permanent water and altitude values extracted from a digital elevation model (DEM). To create environmental layers ($n = 32$) for the analysis, we used ArcGIS 10.1 GIS software (ESRI, Redlands, CA, USA). All data layers were converted to a common projection, map extent and resolution. The resolutions (pixels) of the climatic and NDVI (Normalized Difference Vegetation Index) variables were $10 \times 10 \text{ km}^2$ at source and they were converted to $1 \times 1 \text{ km}^2$ when used for the Maxent modelling. All the other environmental

variables were feature data type (land uses, distance from farms etc) which were converted to raster dataset with the same resolution and cell size using the conversion tool from the spatial analyst extension. In order to determine the altitude and the values for each environmental variable of the human cases precisely, the cell statistics Tool from the Spatial Analyst extension of ArcToolbox and the mean centre of the Spatial Statistics Tool, ArcGIS 10.1 were used for each one of the municipality districts where human leishmaniasis cases were recorded. Then we extracted the cell values at the mean feature point location.

Environmental Niche Model (ENM)

In the Maxent modelling, the pixels of the study area define the area where the distribution of the Maxent probability is defined. Pixels with occurrence records constitute the sample points and the features are environmental parameters (climatic, vegetation, topographic etc.). Maxent method requires presence-only data, utilizes both continuous and categorical data and includes efficient deterministic algorithms and mathematical definitions (Phillips *et al.* 2006). Human leishmaniasis cases and dogs that were *Leishmania* positive were used as occurrence points for the ENM procedure. Maximum entropy modelling (MaxEnt software version 3.3.3) was used to predict the appropriate ecological niches for humans and dogs (Phillips *et al.* 2006). The 'bias file' was included in the analysis in order to represent the sampling effort and to reduce the sampling bias. The goodness of fit of the model predictions was evaluated by the mean area under the curve (AUC) of the receiver operating characteristic curve (ROC). We used the Jackknife procedure to reduce the number of environmental variables to only those that showed a substantial influence on the model. According to Ceccarelli *et al.* (2015) we repeated the test with the Jackknife test until all the remaining variables have a positive effect on the total gain.

RESULTS

Human leishmaniasis cases data

The human leishmaniasis cases were recorded in 19 of the 26 Municipalities of Thessaly ranging from one to 19 cases. The 82 human leishmaniasis cases officially notified in our study period are subdivided as follows: four in 2007, five in 2008, four in 2009, five in 2010, four in 2011, 14 in 2012, 32 in 2013 and 14 in 2014 (annual epidemiological report of HCDCP). It becomes obvious that the annual occurrence of human leishmaniasis cases was almost stable from 2007 to 2011; thereafter, the number of cases increased 3-fold in 2012. Subsequently, it reached

the maximum number in 2013 (the number of cases increased almost 8-fold compared with 2007–2011 and 2-fold compared with 2012) and after that, a decline presented in 2014. Thus, 60 out of the 82 human leishmaniasis cases were recorded during 2012–2014. Interestingly, half of the cases recorded in 2012–2014 (30/60), were living in the prefecture of Larissa.

Visceral leishmaniasis was the only form of the disease reported in the region of Thessaly during our study time period. Overall, 54% of the human cases were males. The range for the age was 1–84 years old, with a median of 61 years old. The majority of the human leishmaniasis cases ($n = 37$) were adults older than 65 years while only two cases were children in the first year of age. Only two cases reported had Albanian nationality while the remaining had Greek nationality. It is also worth mentioning that 23 out of the 82 (28%) human leishmaniasis cases reported were immunocompromised. Regarding the presence of dogs, 32% of the human leishmaniasis cases reported the ownership of a dog without known canine leishmaniasis while the remaining 68% reported the absence of dogs in the home. However, 82% of the cases reported the presence of stray dogs in the region. Concerning the presence of vectors, 76% reported the presence of sandflies in their living area.

The majority of the human leishmaniasis cases were recorded in low altitude, 27–200 m above sea level. The mean altitude was 171 m asl (range 27–1083 m \pm 190 329 S.D.). The minority of cases was recorded in broadleaved forests while the majority of cases were recorded in discontinuous urban fabric, cultivated and permanently irrigated land.

Data on *Leishmania* infected dogs

Regarding the canine population, *Leishmania* infected dogs were found in each one of the four prefectures of Thessaly. In particular, 10 dogs in the prefecture of Magnisia, 23 dogs in the prefecture of Karditsa, 38 dogs in the prefecture of Larissa and 14 dogs in the prefecture of Trikala were found *Leishmania* PCR positive to at least one of the samples examined (exfoliative epithelial cells, lymph node aspirates and whole blood in EDTA for each dog).

According to their lifestyle, 38% (32/85) and 62% (53/85) of the dogs used in this analysis were stray and owned dogs, respectively, while 49% of owned dogs were hunting dogs (26/53). Overall, the 58% (49/85) of the dogs were males. Concerning the age groups, 42% (36/85) and 46% (39/85) of the dogs were young adults (>1 and \leq 3 years old) and adults (>3 and \leq 9 years old), respectively, whereas only 4% (4/85) and 3% (3/85) were young (<1 years old) and old (>9 years old), respectively. The age group was unknown for two dogs.

Table 1. The contribution of the environmental variables to the MaxEnt model

Environmental variables	Per cent contribution	Host
Max temperature of warmest month (°C) (Bio_5)	52.8	Humans
Mean diurnal range [mean of monthly (max temp–min temp)] (°C) (Bio_2)	35	Humans
Temperature annual range (Bio_7 = Bio_5–Bio_6) (°C)	6.3	Humans
Farms distance (Farmsdis)	5.9	Humans
Max temperature of warmest month (°C) (Bio_5)	53.1	Dogs
Mean diurnal range [mean of monthly (max temp–min temp)] (°C) (Bio_2)	23.2	Dogs
Population density (Popden)	23.1	Dogs
Max temperature of warmest month (°C) (Bio_5)	52.6	Dogs and humans
Temperature annual range (Bio_7 = Bio_5–Bio_6) (°C)	23.9	Dogs and humans
Temperature seasonality (Coefficient of Variation)	11	Dogs and humans
Farms distance (Farmsdis)	6.3	Dogs and humans
Population density (Popden)	6.2	Dogs and humans

The majority of the *Leishmania* infected dogs were found in low altitude, with a mean altitude of 151 m asl (range 38–538 m \pm 92.715173 s.d.) and in urban and rural areas, in irrigated and non-irrigated land.

Predictive ENM for human cases

The contribution of the environmental variables to the MaxEnt model analysed in this study are shown in Table 1. Jackknife of regularized training gain test for human leishmaniasis in Thessaly is shown in Fig. 1. The environmental variable with the highest gain when used in isolation is Max Temperature of Warmest Month (°C) (Bio_5) while the variable that decreases gain the most when omitted, is distance from farms (Farmsdis). Regularized training gain (sum of the likelihood of the data plus a penalty function) is 2.367, training AUC is 0.964 and unregularized training gain is 2.574.

Predictive ENM for canine cases

The contribution of the environmental variables to the MaxEnt model analysed in this study are shown in Table 1. Jackknife of regularized training gain test for *Leishmania* infection in dogs in Thessaly is shown in Fig. 1. The environmental variable with the highest gain when used in isolation is Max Temperature of Warmest Month (°C) (Bio_5) and the most important when omitted is human population density (Popden). Regularized training gain (sum of the likelihood of the data plus a penalty function) is 3.544, training AUC is 0.989 and unregularized training gain is 3.731.

Predictive ENM for combined human and canine cases

The contribution of the environmental variables to the MaxEnt model analysed in this study are shown in Table 1. Jackknife of regularized training gain test for human leishmaniasis and *Leishmania* infection in dogs in Thessaly is shown in Fig. 1.

Concerning the combined human and canine cases, the environmental variable with the highest gain when used in isolation is Max Temperature of Warmest Month (°C) (Bio_5) while the variable temperature annual range (Bio_7 = Bio_5–Bio_6) also presents high gain when used in isolation. The variable that decreases gain the most when omitted is distance from farms (Farmsdis). Regularized training gain (sum of the likelihood of the data plus a penalty function) is 2.452, training AUC is 0.973 and unregularized training gain is 2.639.

Identification of high-risk areas

The areas presenting probability greater than 80% for the presence of *Leishmania* infection according to the MaxEnt model were considered as high-risk areas (Fig. 2). The model used in this study recognized the high-risk areas for *Leishmania* infection, most of which were concentrated in the central plain of Thessaly. Other high-risk areas were located along the coast line of the region and in the western and eastern areas with low altitudes (Fig. 2). The highest percentage of the high-risk areas was found in low altitude (<200 m asl) and in irrigated and cultivated agricultural areas. In total, 115 out of the 528 villages and small towns were found in high-risk areas.

DISCUSSION

In this study, we used ENM from MaxEnt (Phillips *et al.* 2006) in order to identify the environmental variables related to human leishmaniasis cases and *Leishmania* infection in dogs and to recognize high-risk areas in the region of Thessaly, Central Greece. The analysis revealed that the maximum temperature of the warmest month contributes to the highest per cent to define environmental niche profiles for humans and dogs separately as well as for the combined human and canine cases in the study area. Moreover, our ecological niche modelling approach has produced the first risk map of

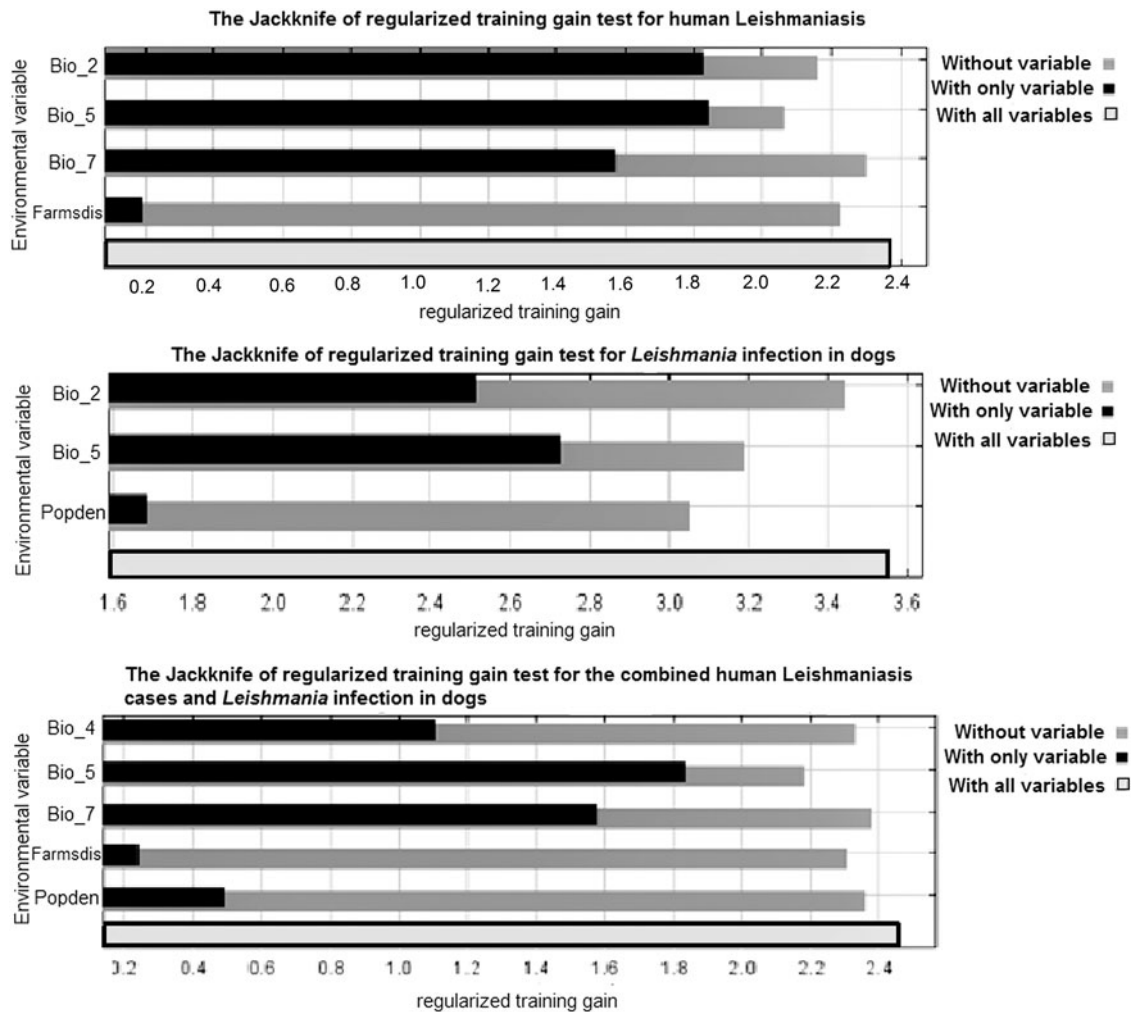


Fig. 1. Jackknife analysis results: (a) The Jackknife of regularized training gain test for human leishmaniasis in Thessaly, Central Greece. (b) The Jackknife of regularized training gain test for *Leishmania* infection in dogs in Thessaly, Central Greece. (c) The Jackknife of regularized training gain test for the combined human leishmaniasis and *Leishmania* infection in dogs in Thessaly, Central Greece. Bio_2: Mean Diurnal Range [Mean of monthly (max temp–min temp)] (°C); Bio_5: Max Temperature of Warmest Month (°C); Bio_7: Temperature Annual Range (BIO7 = BIO5–BIO6) (°C); Farmsdis: Farms Distance; Popden: Population density.

Leishmania infection in Thessaly combining data from humans and dogs.

Among the sandfly species that have been reported in our study area, there are four sandfly species which are considered to be proven vectors of *Leishmania* spp; *Phlebotomus perfliewi*, *P. tobbi*, *P. similis* and *P. neglectus* presenting specific spatial distribution and biological habits. In particular, *P. perfliewii* is related to humid climate of mainland Greece, being scarcer on Greek islands with a distinctly hot and dry Mediterranean climate. In contrast, *P.tobbi* is a sandfly with preference to a more arid and semiarid bioclimate. *P.similis* as well as *P. neglectus* appear to be more common on the islands than in continental Greece (Ivović *et al.* 2007). The presence of these phlebotomine species may be favoured by the climatic conditions in the region of Thessaly but the estimation of the vector abundance and the phlebotomine sandfly species diversity,

distribution and *Leishmania* infection needs to be further investigated in order to reach accurate conclusions.

It is well known that leishmaniasis is a climate-sensitive disease, affected by changes in rainfall, atmospheric temperature and humidity. As it has been repeatedly suggested these changes can strongly impact on the ecology of vectors and reservoir hosts by altering their distribution and influencing their activity, survival and population sizes (Elnaiem *et al.* 1998; Killick-Kendrick, 1999; Aspöck *et al.* 2008; Gage *et al.* 2008; Ready, 2010; Ballart *et al.* 2014). Other environmental parameters including land cover (Colacicco-Mayhugh *et al.* 2010) and the availability of organic matter may also affect the presence and abundance of vectors (Kassem *et al.* 2001; Ozbel *et al.* 2011).

Franco *et al.* (2011) reported that the canine leishmaniasis seroprevalence for the Western Europe was

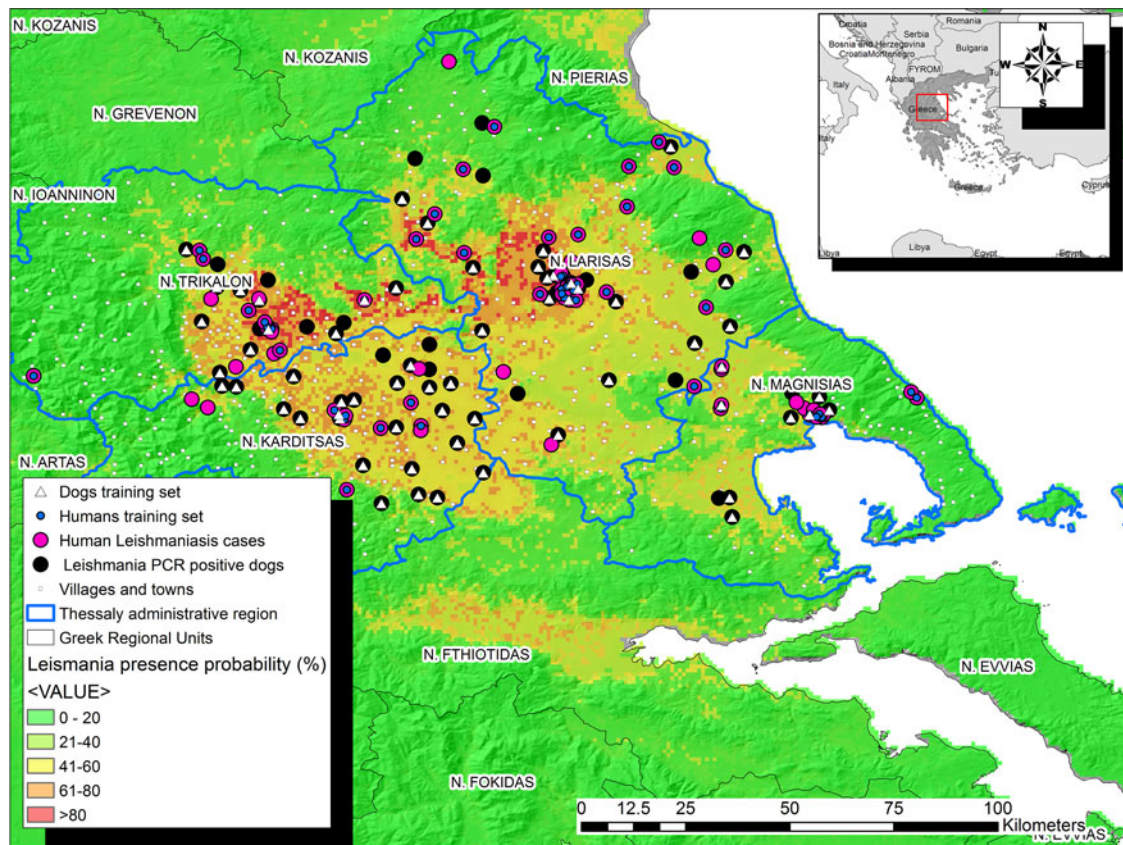


Fig. 2. Map of Thessaly, Central Greece showing the geographical distribution of human leishmaniasis cases and *Leishmania* PCR positive dogs (2007–2014) and the probability for the presence of human leishmaniasis cases and *Leishmania* infected dogs.

inversely U-shaped for the predictors altitude, minimum night-time land surface temperature, night-time land surface temperature amplitude of tri-annual cycle and to a lesser degree the enhanced vegetation index phase of annual cycle. In another study conducted in Greece during 2005–2010 high mean land surface temperature was related to higher risk for dog seropositivity (Ntais *et al.* 2013). Although our study concerns human as well as canine cases of *Leishmania* infection in the region of Thessaly and not in Greece in overall, the importance of the environmental temperature is a common observation with the studies conducted so far in different countries (Chamaillé *et al.* 2010; Franco *et al.* 2011) and in Greece (Ntais *et al.* 2013) probably because it strongly affects the vector distribution and activity. Furthermore, areas of 0–1000 m altitude were estimated to present higher risk than those of >1000 m (Ntais *et al.* 2013) which is in agreement with our findings as the majority of the human leishmaniasis cases as well as the *Leishmania* infected dogs were recorded in low altitude (mean altitude of 171 and 151 m asl, respectively). Moreover, the highest percentage of the high-risk areas was found in low altitude (<200 m asl). Regarding the land cover, the majority of human cases were recorded in discontinuous

urban fabric, cultivated and permanently irrigated land and the majority of the *Leishmania* infected dogs was found in urban and rural areas, in irrigated and non-irrigated land. Most of the high-risk areas were found in irrigated and cultivated agricultural areas. These findings are in accordance with Ntais *et al.* (2013) who reported that the presence of water bodies in an area, the agricultural areas and the semi-natural areas presented the highest risk as well as with previous studies conducted in other countries (Alonso *et al.* 2010; Chamaillé *et al.* 2010; Colacicco-Mayhugh *et al.* 2010; Abdel-Dayem *et al.* 2012).

In another study conducted in Greece during 2007–2010, spatial analysis was used to compare Veterinary Questionnaires results with dog seropositivity data (Sifaki-Pistola *et al.* 2014) retrieved from a previous study (Ntais *et al.* 2013). This analysis showed that the areas of Attiki, Peloponisos, Kavalla, Kerkira and the western regions of Greece are high-risk areas whereas Thessaly was estimated as a low to medium risk area for CanL (Sifaki-Pistola *et al.* 2014). Even though, in our study which was conducted during 2007–2014, an increase in human leishmaniasis cases took place in 2012 and 2013 in the region of Thessaly, a study period which is not included in the previous studies (Ntais *et al.*

2013; Sifaki-Pistola *et al.* 2014). By using the available presence-only data for human cases and *Leishmania* infected dogs; the model used in our study recognized the high-risk areas for *Leishmania* infection in the region of Thessaly, most of which were concentrated in the central plain of Thessaly. Other high-risk areas were located along the coast line of the region and in the western and eastern areas with low altitudes (Fig. 2). It is worth mentioning that in the agricultural areas of Thessaly, 115 out of the 528 villages and small towns were found in high-risk areas.

As it has been already suggested by Franco *et al.* (2011), surveys regarding human and canine leishmaniasis present several limitations in terms of their scope and standardization, which however can be used as guidelines for the collection of prospective data. In our study, extreme altitudes were included for both hosts as the human cases were recorded in altitudes ranging from 27 to 1083 m and the *Leishmania* PCR positive dogs were found in altitudes ranging from 38 to 538 m asl. Data on age and lifestyle have been registered for humans and dogs. However, the dog density has yet to be defined in our study area and this is a limitation of our study.

The results of this study should be treated with caution as although human leishmaniasis is a mandatory notifiable disease in Greece, our analysis could be affected by the quality of the collected human cases. It is well-known that both underdiagnoses and underreporting of the human leishmaniasis cases is occurring in most European countries. Thus, our results could be affected by eliminating the power of the study to identify possible risk factors. However, sampling bias in human cases is not justified while for canine samples our methodology and analysis eliminated the sample bias. Another possible limitation of our study is the high percentage of immunocompromised persons in the human cases raising the possibility of reactivation and not of being new cases. The epidemiological pattern of the region was the same as of the whole Greece as it was published before (Gkolfinopoulou *et al.* 2013) and is confirmed by recent data from the National Surveillance Center. The percentage of immunocompromised cases (28%) among the human cases in the study area is similar with the percentage in the whole country. While someone can support that immunocompromised cases could be regarded as reactivation, at the same time could be considered as new cases due to host sensitivity. Moreover, we modelled the human leishmaniasis cases excluding the immunocompromised patients ($n = 23$) and the analysis showed that the environmental variables which contribute to the MaxEnt model do not differ when the immunocompromised patients are included. Thus, their inclusion does not mask the result and the prediction.

Maxent method requires presence-only data, utilizes both continuous and categorical data and includes efficient deterministic algorithms and mathematical definitions (Phillips *et al.* 2006). Thus, this method enhanced the capabilities of results analysis in our study leading in the identification of high-risk areas for *Leishmania* infection in the study area. Further studies concerning the identification of high-risk areas as well as large entomological studies in the mainland and islands of Greece are needed in order to raise awareness and implement targeted control measures for the protection of public health and the improvement of the disease surveillance.

SUPPLEMENTARY MATERIAL

To view supplementary material for this article, please visit <http://dx.doi.org/10.1017/S0031182016000378>

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REFERENCES

- Abdel-Dayem, M. S., Annajar, B. B., Hanafi, H. A. and Obenauer, P. J. (2012). The potential distribution of *Phlebotomus papatasi* (Diptera: Psychodidae) in Libya based on ecological niche model. *Journal of Medical Entomology* **49**, 739–745.
- Alonso, F., Giménez Font, P., Manchón, M., Ruiz de Ybáñez, R., Segovia, M. and Berriatua, E. (2010). Geographical variation and factors associated to seroprevalence of canine leishmaniasis in an endemic Mediterranean area. *Zoonoses and Public Health* **57**, 318–328.
- Aspöck, H., Gersdorfer, T., Formayer, H. and Walochnik, J. (2008). Sandflies and sandfly-borne infections of humans in Central Europe in the light of climate change. *Wiener Klinische Wochenschrift* **120**, 24–29.
- Athanasίου, L. V., Kontos, V. I., Saridomichelakis, M. N., Rallis, T. S. and Diakou, A. (2012). A cross-sectional sero-epidemiological study of canine leishmaniasis in Greek mainland. *Acta Tropica* **122**, 291–295.
- Ballart, C., Guerrero, I., Castells, X., Barón, S., Castillejo, S., Alcover, M. M., Portús, M. and Gállego, M. (2014). Importance of individual analysis of environmental and climatic factors affecting the density of *Leishmania* vectors living in the same geographical area: the example of *Phlebotomus ariasi* and *P. perniciosus* in northeast Spain. *Geospatial Health* **8**, 389–403.
- Barón, S. D., Morillas-Márquez, F., Morales-Yuste, M., Diáz-Sáez, V., Gállego, M., Molina, R. and Martín-Sánchez, J. (2013). Predicting the risk of an endemic focus of *Leishmania tropica* becoming established in south-western Europe through the presence of its main vector, *Phlebotomus sergenti* Parrot, 1917. *Parasitology* **140**, 1413–1421.
- Ceccarelli, S., Balsalobre, A., Susevich, M. L., Echeverria, M. G., Gorla, D. E. and Marti, G. A. (2015). Modelling the potential geographic distribution of triatomines infected by *Triatoma* virus in the southern cone of South America. *Parasites and Vectors* **8**, 153.

- Chamaillé, L., Tran, A., Meunier, A., Bourdoiseau, G., Ready, P. and Dedet, J.-P. (2010). Environmental risk mapping of canine leishmaniasis in France. *Parasites and Vectors* **3**, 31.
- Christodoulou, V., Antoniou, M., Ntais, P., Messaritakis, I., Ivovic, V., Dedet, J.-P., Pralong, F., Dvorak, V. and Tselentis, Y. (2012). Re-emergence of visceral and cutaneous leishmaniasis in the Greek Island of Crete. *Vector Borne and Zoonotic Diseases (Larchmont, N.Y.)* **12**, 214–222.
- Colacicco-Mayhugh, M. G., Masuoka, P. M. and Grieco, J. P. (2010). Ecological niche model of *Phlebotomus alexandri* and *P. papatasi* (Diptera: Psychodidae) in the Middle East. *International Journal of Health Geographics* **9**, 2.
- Domenikiotis, C., Spiliotopoulos, M., Tsiros, E. and Dalezios, N. R. (2005). Remotely sensed estimation of annual cotton production under different environmental conditions in Central Greece. *Physics and Chemistry of the Earth, Parts A/B/C* **30**, 45–52.
- Elnaiem, D. A., Connor, S. J., Thomson, M. C., Hassan, M. M., Hassan, H. K., Aboud, M. A. and Ashford, R. W. (1998). Environmental determinants of the distribution of *Phlebotomus orientalis* in Sudan. *Annals of Tropical Medicine and Parasitology* **92**, 877–887.
- Franco, A. O., Davies, C. R., Mylne, A., Dedet, J.-P., Gállego, M., Ballart, C., Gramiccia, M., Gradoni, L., Molina, R., Gálvez, R., Morillas-Márquez, F., Barón-López, S., Pires, C. A., Afonso, M. O., Ready, P. D. and Cox, J. (2011). Predicting the distribution of canine leishmaniasis in western Europe based on environmental variables. *Parasitology* **138**, 1878–1891.
- Gage, K. L., Burkot, T. R., Eisen, R. J. and Hayes, E. B. (2008). Climate and vectorborne diseases. *American Journal of Preventive Medicine* **35**, 436–450.
- Gkolfinoupolou, K., Bitsolas, N., Patrinos, S., Veneti, L., Marka, A., Dougas, G., Pervanidou, D., Detsis, M., Triantafyllou, E., Georgakopoulou, T., Billinis, C., Kremastinou, J. and Hadjichristodoulou, C. (2013). Epidemiology of human leishmaniasis in Greece, 1981–2011. *Euro Surveillance: Bulletin Européen Sur Les Maladies Transmissibles = European Communicable Disease Bulletin* **18**, 20532.
- Hijmans, R. J., Cameron, S. E., Parra, J. L., Jones, P. G. and Jarvis, A. (2005). Very high resolution interpolated climate surfaces for global land areas. *International Journal of Climatology* **25**, 1965–1978.
- Islam, A. (2013). *Manual of Bone Marrow Examination*. Trafford Publishing, Bloomington, USA.
- Ivović, V., Patakakis, M., Tselentis, Y. and Chaniotis, B. (2007). Faunistic study of sandflies in Greece. *Medical and Veterinary Entomology* **21**, 121–124.
- Kassem, H. A., Tewfick, M. K. and Sawaf, B. M. El (2001). Evaluation of avermectins as sandfly control agents. *Annals of Tropical Medicine and Parasitology* **95**, 405–411.
- Killick-Kendrick, R. (1999). The biology and control of Phlebotomine sand flies. *Clinics in Dermatology* **17**, 279–289.
- Leite, R. S., Ferreira, S. de A., Ituassu, L. T., de Melo, M. N. and de Andrade, A. S. R. (2010). PCR diagnosis of visceral leishmaniasis in asymptomatic dogs using conjunctival swab samples. *Veterinary Parasitology* **170**, 201–206.
- Ntais, P., Sifaki-Pistola, D., Christodoulou, V., Messaritakis, I., Pralong, F., Poupalos, G. and Antoniou, M. (2013). Leishmaniasis in Greece. *The American Journal of Tropical Medicine and Hygiene* **89**, 906–915.
- Ozbel, Y., Sanjoba, C., Alten, B., Asada, M., Depaquit, J., Matsumoto, Y., Demir, S., Siyambalagoda, R. R. M. L. R., Rajapakse, R. P. V. J. and Matsumoto, Y. (2011). Distribution and ecological aspects of sand fly (Diptera: Psychodidae) species in Sri Lanka. *Journal of Vector Ecology: Journal of the Society for Vector Ecology* **36** (Suppl. 1), S77–S86.
- Phillips, S. J., Anderson, R. P. and Schapire, R. E. (2006). Maximum entropy modeling of species geographic distributions. *Ecological Modelling* **190**, 231–259.
- Ready, P. D. (2010). Leishmaniasis emergence in Europe. *Euro Surveillance: Bulletin Européen Sur Les Maladies Transmissibles European Communicable Disease Bulletin* **15**, 19505.
- Seid, A., Gadisa, E., Tsegaw, T., Abera, A., Teshome, A., Mulugeta, A., Herrero, M., Argaw, D., Jorge, A., Kebede, A. and Aseffa, A. (2014). Risk map for cutaneous leishmaniasis in Ethiopia based on environmental factors as revealed by geographical information systems and statistics. *Geospatial Health* **8**, 377–387.
- Sifaki-Pistola, D., Ntais, P., Christodoulou, V., Mazeris, A. and Antoniou, M. (2014). The use of spatial analysis to estimate the prevalence of canine leishmaniasis in Greece and Cyprus to predict its future variation and relate it to human disease. *The American Journal of Tropical Medicine and Hygiene* **91**, 336–341.
- Spanakos, G., Patsoula, E., Kremastinou, T., Saroglou, G. and Vakalis, N. (2002). Development of a PCR-based method for diagnosis of *Leishmania* in blood samples. *Molecular and Cellular Probes* **16**, 415–420.
- Tsegaw, T., Gadisa, E., Seid, A., Abera, A., Teshome, A., Mulugeta, A., Herrero, M., Argaw, D., Jorge, A. and Aseffa, A. (2013). Identification of environmental parameters and risk mapping of visceral leishmaniasis in Ethiopia by using geographical information systems and a statistical approach. *Geospatial Health* **7**, 299–308.