Field astrobiology research in Moon–Mars analogue environments: instruments and methods

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Abstract: We describe the field demonstration of astrobiology instruments and research methods conducted in and from the Mars Desert Research Station (MDRS) in Utah during the EuroGeoMars campaign 2009 coordinated by ILEWG, ESA/ESTEC and NASA Ames, with the contribution of academic partners. We discuss the entire experimental approach from determining the geological context using remote sensing, in situ measurements, sorties with sample collection and characterization, analysis in the field laboratory, to the post sample analysis using advanced laboratory facilities.

We present the rationale for terrestrial field campaigns to strengthen astrobiology research and the link between *in situ* and orbital remote sensing data. These campaigns are supporting the preparation for future missions such as Mars Science Laboratory, ExoMars or Mars Sample Return. We describe the EuroGeoMars 2009 campaign conducted by MDRS crew 76 and 77, focused on the investigation of surface processes in their geological context. Special emphasis was placed on sample collection and pre-screening using *in-situ* portable instruments. Science investigations included geological and geochemical measurements as well as detection and diagnostic of water, oxidants, organic matter, minerals, volatiles and biota.

EuroGeoMars 2009 was an example of a Moon–Mars field research campaign dedicated to the demonstration of astrobiology instruments and a specific methodology of comprehensive measurements from selected sampling sites. We discuss in sequence: the campaign objectives and trade-off based on science, technical or operational constraints. This includes remote sensing data and maps, and geological context; the monitoring of environmental parameters; the geophysical context and mineralogy studies; geology and geomorphology investigations; geochemistry characterization and subsurface studies.

We describe sample handling (extraction and collection) methods, and the sample analysis of soils and rocks performed in the MDRS laboratory using close inspection, initial petrological characterization, microscopy, Visible-NIR spectrometry, Raman spectrometry, X-ray diffraction/X-ray fluorescence spectrometry, soil analysis, electrochemical and biological measurements.

The results from post-mission analysis of returned samples using advanced facilities in collaborator institutes are described in companion papers in this issue. We present examples of *in-situ* analysis, and describe an example investigation on the exploration and analysis of endolithic microbial mats (from reconnaissance, *in-situ* imaging, sampling, local analysis to post-mission sample analysis). Received 22 January 2011, accepted 24 January 2011, first published online 14 March 2011

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Introduction

Terrestrial analogue studies are used to better understand the nature and rates of geological and biological processes on Earth in order to interpret and validate information from orbiting or surface missions on extraterrestrial bodies. These terrestrial analogue data complement the interpretation of missions such as Mars-Express, SMART-1, Chandrayaan-1, Lunar Reconnaissance Orbiter (LRO), Mars Exploration Rovers and Mars Reconnaissance Orbiter (MRO), and help to prepare for future lunar and planetary Lander missions. International cooperation in terrestrial analogue activities provides a logical early step to implementing international Moon-Mars missions (see ILEWG Reports and ICEUM Declarations 2006-2010 (ICEUM 9, 10, 11); Foing 2008; Foing et al & ICEUM participants (2008b, c, d, e); MEPAG Report 2007; COSPAR Planetary Exploration Committee (PEX) Report 2010).

Surface science versus remote sensing

Surface science is one of the primary objectives of recent and future Mars and Moon missions. The geological record of Mars indicates a diversity of water-modified environments, including potential ancient habitable environments. Hydrated minerals on Mars trace the history of surface water and the global atmosphere and a long-term climate cycle (Christensen et al. 2001; Bibring et al. 2006). Recent Moon missions advanced our knowledge on surface composition (Lucey et al. 1998; Jolliff et al. 2000) and the bombardment history and indicated the presence of H₂O and hydroxyl species on the lunar surface (Feldman et al. 2001; Pieters et al. 2009). Science investigations include a wide range of activities from global mapping to microscopic scale. Significant new science results will be obtained from coordinated multi-instrument operations on the surface. In-situ investigations of rocks and soil or sample return missions both require the development of systematic multi-instrument protocols, characterization diagnostics and methods to merge data from various instruments. Remote sensing/ground truth validation will enhance the science exploitation of future missions.

Mars research context

Orbital remote sensing has revealed a complex geologic record of planet Mars that formed in response to processes that include volcanism, weathering/erosion, sedimentation, glaciation, polar ice cap processes, fluid/rock interactions and tectonism and others. Six spacecraft have unveiled a new face of Mars history (Mars Global Surveyor (MGS), Mars Odyssey, Mars-Express (MEX), the two Mars Exploration Rovers (MER) and MRO). Various minerals have been identified both from the orbit or from the Martian surface (Klingelhofer et al. 2004; Squyres et al. 2004; Bibring et al. 2005; Gendrin et al. 2005; Poulet et al. 2005). For instance, the Gusev area has been studied both by the MER rover and MEX orbiter (Greeley et al. 2005; Parker et al. 2010). Recent results revealed the timing and duration of hydrologic activity on Mars and the evolution of sedimentary processes through time. Water, an important ingredient for life, could also be trapped as underground ice. MEX high-resolution stereo camera HRSC images have been used to determine that volcanic activity continued until recent times (Neukum *et al.* 2004). They indicate recent periglacial tropical activity (Murray *et al.* 2005; Head *et al.* 2005a, b), possibly the result of erratic variations of Mars obliquity. The past conditions of Mars may have eventually allowed life to develop (McKay & Stoker 1989). However, today, a combination of solar ultraviolet radiation, the extreme dryness of the soil and the oxidizing nature of the soil chemistry provides a toxic environment to biological and organic material on the surface or the near subsurface. Understanding the complex interactions between organic compounds and the soil mineralogy is vital for the potential detection of past or present life on Mars.

Moon research context

On the Moon we can study geological processes shaping the surface due to impacts, volcanism and space weathering. Recent lunar orbiters SMART-1 (Foing et al. 2006, 2008a, b, c, d, e), Selene Kaguya (Kato et al. 2008; Haruyama et al. 2008, 2009; Ono et al. 2009), Chandrayaan-1 (Goswami et al. 2008; Pieters et al. 2009), Chang'E1 and LRO (Chin et al. 2007; Vondrak et al. 2010) have studied impact processes and surface morphologies such as terraces, ejecta, central peaks for a number of craters of various sizes and ages in different locations. Bulk crustal composition provides constraints on the origin and evolution of the Moon, the lunar crusts and the large basins (such as the South Pole-Aitken Basin, SPA) (Jolliff et al. 2000). Measurements from orbit and existing lunar samples will enhance our knowledge on absolute chronology of the Moon and on the early or late heavy bombardment in the Solar System. The survival of exogenous ices and organics at lunar poles is also relevant in the astrobiology context.

Rationale for terrestrial campaigns

Extreme environments on Earth often provide similar terrain conditions to landing/operation sites on the Moon and Mars. In order to maximize scientific return of space missions, it is important to rehearse mission operations in the field and through simulations. Terrestrial field research campaign in support of future planetary missions often include investigations of the geological, geochemical, biological and environmental context of a site; *in-situ* analysis, drilling of cores and sampling. This approach allows the demonstration of remote control field rovers; improvement of instrument performance; and evaluating crew operations and Extra Vehicular Activity (EVA) technologies. In this paper, we describe the planning and protocol development for both *in-situ* and post-mission lab-analysis for the astrobiology research campaign at MDRS (MDRS website; http://desert.marssociety.org).

EuroGeoMars astrobiology field demonstration

The campaign EuroGeoMars 2009 was conducted in Utah (MDRS crew 76 and 77) and had four sets of objectives:

• Technology demonstration: a set of instruments were deployed, tested and assessed, and training was provided to scientists using them in subsequent rotations.

- Research aspects: a series of field science and exploration investigations were conducted in geology, geochemistry, biology and astronomy, with synergies with space missions and research from planetary surfaces and Earth extreme environments.
- Human crew-related aspects, i.e. (a) evaluation of the different functions and interfaces of a planetary habitat, (b) crew time organization in this habitat, (c) evaluation of man-machine interfaces of science and technical equipment.
- Education, outreach, communications, multi-cultural and public relations aspects.

The methodology of comprehensive measurements at selected sites includes successive steps (here numbered and later developed in part 3):

- 1. Definition of campaign objectives and trade-off from science, technical or operational constraints.
- 2. Analysis of remote sensing data and existing maps.
- 3. Imaging reconnaissance.
- 4. Monitoring of the local environment and meteorology.
- 5. Geology and geophysical context.
- 6. Field geology and geomorphology investigations.
- 7. Field geochemistry characterization.
- 8. Field subsurface studies.
- 9. Sample handling (extraction and collection) methods.
- 10. Analysis of soils and rocks performed in the station laboratory (physical, mineral, chemical, organic and biological measurements).
- 11. A posteriori sample analysis using advanced facilities in the collaborator institutes.

In order to address these objectives, we developed and adapted tools and methods as described in section 'EuroGeoMars 2009: an example of Moon–Mars astrobiology research campaign'.

EuroGeoMars 2009: an example of Moon-Mars astrobiology research campaign

Science investigations were designed to understand the geological origin of the region through petrological and geochemical study of the constituents (minerals, organic matter, water, chemical compounds and biota). The compiled datasets have been compared to remote sensing data for geological interpretation. Special emphasis was given to the astrobiology objectives of the campaign, and the correlations between mineral, environmental parameters, organics and biota, placed in the geochemistry context.

Definition of campaign objectives and trade-off from science, instruments, technical or operational constraints

In order to assess several human and scientific aspects of future robotic and manned missions on planetary surfaces, the EuroGeoMars campaign was proposed by collaborators from ILEWG, ESTEC and NASA Ames in collaboration with European and US investigators. The campaign was prepared through the ExoGeoLab pilot project (Foing *et al.* 2009; Foing *et al.* 2010a, b, c, d) developed by ILEWG with ESTEC support, to evaluate spin-in of new instrument technologies developed from Earth applications with potential use in space, and spin-off applications of instruments developed from space. The ExoGeoLab pilot project followed a technology programme using breadboard instruments that are attached to an automatic station for remote characterization of selected geological sites as well as sample acquisition and analysis methods. A payload suite (instruments, sensors, data handling system) has been deployed, operated and tested at NASA Ames and at ESTEC. On acceptance, instruments were deployed at Utah MDRS station. It was agreed that the EuroGeoMars campaign would last for 5 weeks and be organized in:

- a technical preparation week (24–31 January 2009) for instrumentation deployment and technology/research field demonstration.
- 1st rotation MDRS crew 76 (1–15 February 2009) conducting preliminary research.
- 2nd rotation MDRS crew 77 (15–28 February 2009) focusing on further research utilization and more in-depth analysis.

The goal of the demonstrated technologies was to start validating a comprehensive set of equipment, instruments and methods that can be used in robotic and human Moon–Mars surface exploration missions. Some technology road-maps have identified required measurements and possible techniques that could be employed (ICEUM Reports and Declarations 2006–2010; MEPAG Report 2007). We selected for this first EuroGeoMars 2009 campaign a subset of relevant instrument breadboards, field sensors or adapted commercial equipment. Several science and exploration instruments were either brought from Europe or lent by US collaborators. Most were deployed and installed during the technical preparation crew week.

Analysis of remote sensing data and maps, and geological context

In preparation for the campaign, we collected geological maps and remote sensing data from the region. We consulted the literature and reports from previous field studies. This included interpretation of aerial photo images and United States Geological Survey geological maps. Traverses were planned using these images and maps and taking into account the time required for *in-situ* measurements and sampling protocol. We developed a method and database to permit a full documentation of samples taken in their geological context. The desert near Hanksville, Utah, includes a range of Mars analogue geological and geochemical features, such as lacustrine and evaporitic sediments, and paleochannels including some with inverted relief. The paper by Clarke & Stoker (2011) in this issue describes the geological context for the samples, and in particular looks at concretions in exhumed channels and their implications for Mars (Figs. 1 and 2).

To support the sampling, the GPS coordinates of samples were collected systematically, together with panoramic imaging to relate to remote sensing, as well as macroscopic and close-up imaging.



Fig. 1. (a) Google Earth satellite image showing the location of the MDRS station (near F) and sampling areas at Kissing Camel Ridge (near G). (b) Geology for the EuroGeoMars campaign, and positions of sampling areas (Wendt *et al.* 2009).



(b)



Fig. 2. (a) Southward view towards MDRS with Henry Mountains background. (b) Landscape and stratigraphy near MDRS station looking North, showing the resistant layer formed by sandstones of the Dakota sandstone formation at the top of the ridge and the shale slopes of the Brushy Basin member of the Morrison formation below.

Imaging and navigation reconnaissance from orbit, aerial view, rover or EVA

Orbital and aerial imagery as well as the geology maps were analysed in order to define the possible sites for *in-situ* investigation. We developed a method for merging different imaging datasets taken from different perspectives (vertical or lateral) and integrated them in an interactive database. In parallel, a technology experiment was conducted on Mars



Fig. 3. Context panoramic imaging of salt wash side view.

navigation using the triangulation of positions of deployed captive helium balloons, in coordination with remote support in order to acquire coordinates (even in absence of a GPS system as would be the case on Mars). A video-cam was lifted by a balloon to provide an aerial view of the field for reconnaissance (Fig. 3).

A microrover (developed by Carnegie Mellon University) was used in the field to perform visualization tests for operation. The rover was equipped with an additional camera system (Hendrikse *et al.* 2010) to provide remote navigation tools. The rover was used to test issues of remote control, locomotion, hazard avoidance and data transfer that are critical in future surface operation missions. The rover was also used to provide remote reconnaissance imaging and geological context of the candidate scenes where samples could later be collected. Prior to sampling, a number of EVA traverses were conducted to specific locations in order to perform reconnaissance of the site and characterize the geology, as well as to select locations for *in-situ* measurements and sampling (Fig. 4).

Monitoring of the local meteorology environment

Measurements of temperature, humidity, radiation, moisture and water activity were derived from sensors available at the MDRS station, or brought to the sampling sites, as well as from nearby local weather stations. Weather statistics and satellite observations can constrain the average and variation of parameters affecting the hydrology, moisture and oxydation level. The region around Hanksville is characterized as arid desert, cold in winter and hot in summer with an average annual temperature of 12 °C. The diurnal range is given as 16–37 °C in July and -7 to +7 °C on 1 Feb. The area is subjected to wind erosion and was shaped by fluvial erosion. Hanksville receives 140 mm of annual average precipitation (Godfrey et al. 2008). Weather station sensors include measurements of the diurnal variations of temperature range, winds median average and gusts. The relative humidity showed minima at 15% and diurnal dawn maxima of 50-80% during the EuroGeoMars campaign. The average barometric pressure was 86 kPa. The wind variations showed a median of 5 kmph and gusts of 40-80 kmph. The Photosynthetic Active Radiation in the range 400-700 nm is at maximum 2000 μ moles of photons m⁻² s⁻¹. At the start of the EuroGeoMars campaign there was snow precipitation of 0.76 cm water equivalent (leading to a snow cover of 5 cm depth) on 26 January, a slight rain equivalent to 0.05 cm on 12 February, of 0.08 cm on 23 February, and fog on 24 February 2009. A more systematic study including the statistics of diurnal and seasonal changes of those quantities, as well as mechanism for eolian dust transport or heterogeneous water activity requires a systematic set of in-situ instruments and data acquisition methods.

Geology and geophysical context

The MDRS is surrounded by a series of early Jurassic to late Cretaceous sediments derived by weathering and erosion from Paleozoic sedimentary rocks to the west. These sediments consist of marine to fluvial and lacustrine deposits that locally contain volcanic ashes. The geology formations and units around the MRDS station are described in Fig. 1(b) (Wendt *et al.* 2009; Clarke & Stoker 2011). The red lines (1–7) in Fig. 1



Fig. 4. Rover used for navigation tests, reconnaissance and EVA assistance (courtesy Carnegie Mellon University/NASA Ames).



Fig. 5. Biomarkers on rocks at the base of 'Kissing Camel Ridge' (position G in Fig. 1) (left) field colour image of rock surface with lichens with three main green, yellow and orange constituents. Right: The 'Cyborg astrobiologist' novelty algorithm detects automatically colour special signatures from the same image (McGuire *et al.* 2010; Gross *et al.* 2010).

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(b) indicate some of the geological formations and their member Mb (in clockwise numbered order from top):

Mancos Shale Formation (Cretaceous): Emery sandstone Mb. Blue Gate Shale Mb (1) including carbonaceous pyritic units. Ferron Sandstone Mb (2) formed as fluvial to marginally marine units. Tununk Mb containing bluish carbonaceous pyritic marine shales.

Dakota Sandstone Formation (Early Cretaceous) (3): Made up of non-marine, marginal marine to marine conglomerates, sandstones, shales, coals and oyster reef lime stones.

Morrison Formation (Late Jurassic): Brushy Basin Mb (4) consists of lacustrine and fluvial red brown clays/mudstones, green–bluish–purple beds and sandstone lenses. Interbedded ash layers are weathered to smectite making the soils unfavourable for plant growth. This formation represents an analogue for some Martian terrains. Salt Wash Mb (5) from semiarid alluvial plain with cross bedded or conglomerate

sandstones and local patchy halite and sulphate efflorescence.

Summerville formation (Middle Jurassic): Intercalated siltstone and mudstone (6) locally containing gypsum beds and mud cracks that formed in a tidal flat environment. Location of MDRS habitat is indicated in (7) within the Brushy basin Mb.

The regional landscape has a complex history due to the regional uplift and volcanic activity. The Utah desert is part of the Colorado Plateau, which was uplifted in the Cenozoic as the result of the collision between the Farallon Plate and the North American Plate. Uplift and regional extension was associated with local volcanism that is manifested near the MDRS with diorites such as those found in the Henry Mountains.

The landscape consists of mesas and scarp-bounded surfaces resulting from erosion of the flat-lying succession of alternating units of greater and lesser resistance to erosion. Clay-rich units being more easily eroded and sandstones are less. The sandstone surfaces form smooth plains and the clay-rich materials form dissected slopes.

The Brushy Basin Member forms a dissected plain of cracking clays (Clarke & Pain 2004). Fluvial channels are exposed on the steep slopes or are being exhumed as inverted relief. These features are analogous to those observed from Mars orbiters. The Mars analogue significance of these formations were investigated by Battler *et al.* (2006) and Clarke & Stoker (2011).

Field geology and morphology investigations

The field traverses included an *in-situ* inspection and recognition of the characteristic petrology. A camera system with images at various embedded scales (panoramic, high-resolution, close-up camera) was used in order to document the location, protocol and samples. The soil mechanical properties could be measured *in situ* using penetrometry or by studying the tracks left by rovers or EVA traverses.

A support investigation consisted of an enhanced 'Cyborg astrobiologist' field reporting capability based on a colour

novelty detection algorithm applied to images obtained by a hand-held or rover camera (Gross *et al.* 2010; McGuire *et al.* 2010). The system collects images and detects novelty (see Fig. 5); i.e. unobserved colour ratios compared to previous scenes. We covered a vertical profile in the Brushy Basin Member of the Morrison Formation to test how the system responded to the various clay and sandstone strata. The preliminary results show that the system robustly detects strata not previously recorded.

Field mineralogy characterization

The mineralogy and mineral assemblages of rocks were mostly determined *in situ* by close-up visual inspection. The various minerals identified include quartz, gypsum, clays, calcite and sulfates (Borst *et al.* 2010). Diorites were also sampled from an expedition to Mount Henry. Specific note was made of the original sedimentary processes responsible for the sediment deposition and more recent processes that led to secondary mineral formation such as gypsum and calcite concretions, desert varnish, etc. (Fig. 6(a)).

A Magnetic Susceptibility Meter was used in the field to determine the magnetic susceptibility and conductivity of samples. The Xterra (by InXitu) Field X-ray Diffractometer for mineralogy and X-ray Fluorescence for elemental chemistry and the Raman spectrometer (InPhotonics) were tested in outdoor conditions as the instruments could be transported. As we had installed a geochemical laboratory in the MDRS habitat, we concentrated for this research campaign on fast *in-situ* characterization and sample collection and used the analytical instruments in the laboratory for more accurate and detailed investigations. In some cases, a classical field test for carbonates in the soil was performed using HCl acid and observing the release of CO_2 bubbles.

Field subsurface studies

Drilling equipment included a Milwaukee hand-operated electrical drill that could reach depths down to 1 m. Another manual rotary drill was used to sample soft-clay areas. The drill cores provided information on the vertical structure of soils and the distribution of minerals within rocks. These observations were compared with the lateral variations in rock layers observed from the edges of cliffs to determine the scale of heterogeneity of individual strata.

A comparison was also made with data obtained from Ground Penetrating Radar (GPR) subsurface test measurements. The CRUX GPR developed by JPL (Kim *et al.* 2005), and adapted by NASA Ames was tested to provide information on the stratification of sedimentary structures. The GPR operates at 800 MHz with a penetration of 5 m and a resolution of 15 cm, depending on the soil permittivity and scattering properties. The GPR was only used in few areas to study the clay deposits near the MDRS Morrison Formation and the top of the Dakota Formation.

A later campaign in 2010 (DOMMEX-EuroMoonMars) focused on performing subsurface science-related activities with Mars Underground Mole (MUM, a robotic penetrometry system) and the CRUX GPR (Stoker *et al.* 2010, 2011). Data





Fig. 6. (a) Field inspection of calcite evaporite rocks. (b) Display and documentation of samples from one EVA before laboratory analysis.

collected with the CRUX GPR are reported in Clarke & Stoker (2011).

Sample handling (extraction and collection) methods

The sample context was documented with still and HDTV format cameras for field and lab studies (transported from ESTEC/ILEWG ExoGeoLab). Specific protocols were followed for sterile sampling (using gloves and sterile tools), and for borehole core sampling (to preserve the soil stratification record) (Fig. 6(b)).

Sample analysis of soils and rocks performed in station laboratory

For every EVA, the samples were catalogued and curated. The crew installed a geochemical laboratory in the habitat for analysis of the samples. This included a Raman Spectrometer

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(InPhotonics), a Visible/NIR Spectrometer (OceanOptics), an integrated X-ray diffractometer/X-ray fluorescence meter (Terra 158) as well as an optical microscope. We first performed non-destructive techniques. A physical inspection and imaging was performed on the samples before optical spectroscopy was applied using reflectance and Raman spectrometry to determine the mineral and organic content in the soils or rocks. The biological content of sample equivalents (sample aliquots?) was measured with on-site polymerase chain reaction (PCR) equipment.

Inspection, microscopy and morphology

The morphology of minerals and the microbial relation to the mineral assemblage were studied with an optical microscope (200 power, provided at MDRS). The microscope data (FOV few mm) were linked with close-up imaging data (FOV few cm) to provide the spatial context for the geochemical or biological techniques used with different surface or volume fields of view. Microscopy was used to investigate the water samples. Micro-organisms as well as floating particles were concentrated by centrifugation. Several micro-organisms could be detected, most of them being algae (Thiel *et al.* 2009, 2011).

Visible-NIR spectrometry

An Ocean Optics USB2000 Fibre-optic spectrometer was used to measure the light reflectance in the ultraviolet, visible and NIR spectral regions. This permitted correlation of colour inspection with quantitative reflectance. In a few cases, some signatures of absorption due to organic compounds or red fluorescence could be measured on selected samples.

Raman spectrometry

Raman spectroscopy is based on inelastic scattering of light, used to study low-frequency modes of a system such as vibration or rotation. Each mineral has a unique Raman spectral signature, which is compared with standard mineral Raman spectra in a database to identify the mineral composition of the sample (Foing *et al.* 2010c; Som & Foing 2010). For the Raman spectrometer (InPhotonics) used at MDRS, we used an exciting laser at 785.335 nm and measured the Raman spectrum in the range of $160-1900 \text{ cm}^{-1}$. We designed and manufactured a sample holder for Raman and NIR sensor head holder to allow controlled and reproducible sample analysis conditions (Fig. 7).

X-ray diffractometer/X-ray fluorescence meter (XRD/XRF)

The samples were crushed into powders for the XRD/XRF analysis. CheMin is the X-ray diffraction (XRD) instrument aboard NASA's Mars Science Laboratory (MSL) (Blake *et al.* 2007). A commercial instrument called Terra was developed by inXitu, Inc. in 2007 to maximize the ease of use and field deployment. Terra has a similar architecture to CheMin with a smaller CCD for reasons of cost, weight and power. The CCD is cooled to -45 °C with a Peltier cooler. The system includes an onboard computer to control the instrument, acquire and process data in real time and providing a graphical user interface through a wireless link. Li-ion batteries allow 4–5 h of



Fig. 7. Raman spectrometer box (silver) and horizontal Raman fiber feed to be placed on sample holder, the vertical fiber feeds to the Ocean-optics visible NIR spectrometer.

autonomous operation. The entire instrument weighs less than 15 kg including batteries and a rugged housing. XRD data permit mineral identification within a few minutes. XRF data, in the energy range (3–15 keV) allow measurement of specific chemical elements (Fig. 8).

Soil analysis kit and electrochemical measurements

The electro chemical activity was measured using a soil analysis kit providing the content of ions and reactivity. The soil composition of the previously collected soil samples was analysed by using colorimetric chemical reactions (LaMotte Soil Testing System). The pH, nitrogen, potassium, phosphorous, magnesium, calcium and water content of soils originating from areas with and without vegetation were determined. The pH of all soil samples was in the range of 8.2–10.0. The magnesium concentration was very low for all samples (<5 ppm). The range for phosphor was between 5 and 100 ppm (Ehrenfreund *et al.* 2010). Soil conductivity measurements were obtained using a Thermo Orion 135A probe after dilution (1:10) in distilled water and ranged from 1 to 20 mS (Fig. 9(a–e)).

Biological measurements

The crew performed upgrades to augment the biological laboratory at MDRS. The laboratory in the habitat was equipped with the instrumentation shown in Fig. 9. The temperature in the laboratory was slightly below nominal (15-16 °C).

An adenosine tri-phosphate (ATP) meter was used to measure the metabolic activity and microbial content of the samples.

The PCR lab was brought from the ESTEC ExoGeoLab project. An overall set-up was integrated and tested in ESTEC and then transported and reintegrated in the MDRS lab for performing PCR experiments (Thiel *et al.* 2011). This included



Fig. 8. (a) X-ray Diffraction and fluorescence spectrometer XRD/XRF (orange case). The sample is crushed into powder and placed in a container inserted in the central upper slot for X-ray illumination. (b) Diorite sample (left), crushing (right) and sieving (middle) device before XRD-XRF analysis.

a precision balance (Satorius), a vortex and a centrifuge for DNA extraction. Reaction mixtures were performed in a glovebox, and fragment amplification in a thermal cycler Primus 25 advanced (Peqlab). PCR fragments were then analysed using agarose E-gels and visualized. The results of PCR-based analysis of microbial communities during the EuroGeoMars MDRS campaign are described in Thiel *et al.* (2011).

Post-campaign sample analysis using advanced facilities in collaborating institutes

The various soil samples extracted in sterile conditions were divided and sent to various laboratories for a later analysis with advanced techniques:

 Culture-independent molecular analyses directed at ribosomal RNA genes including PCR (with Bacteria-, Archaeaand Eukarya-specific primers), DGGE (Denaturing Gradient Gel Electrophoresis), cloning, sequencing and phylogenetic analysis were performed at VU Amsterdam (Direito *et al.* 2011) to investigate the microbiology of desert samples.

- Solid-phase microextraction, organic solvent extraction and gas chromatography/mass spectrometry at JPL (Orzechowska *et al.* 2011) providing a survey of Polycyclic Aromatic Hydrocarbons (PAHs) abundance in relation to texture, pH and overall organic matter content.
- Infrared spectroscopy (in the range 4000–500 cm⁻¹) and XRD at Leiden University and Imperial College, respectively to analyse the mineral composition of 10 selected soil analogues (Kotler *et al.* 2011).
- Extraction of amino acids, derivatization and GC–MS analysis was performed at Imperial College London (Martins *et al.* 2011).
- Additional analytical studies were performed on a subset of the samples using the Raman spectrometer and visible NIR spectrometer together with microscopy at ESTEC ExoGeoLab facility.
- Scanning electron microscopy was performed at NASA Ames Research Centre on some of the endolithic samples.

A synthesis interpretation of the measurements of selected soil samples collected under sterile conditions and distributed to various laboratories is given by Ehrenfreund *et al.* (2010) (mineralogy, organic content and microbiology). The results are discussed in the context of astrobiology and habitability studies in preparation for future Mars missions (Ehrenfreund *et al.* 2010).

EuroGeoMars scientific research highlights

Field science experiments were started as soon as the corresponding instruments were assembled, tested and deployed. More than 100 documented samples were collected by the MDRS crew 77 for geology (50 samples), astrobiology (11 + 5 samples divided for 8 investigators groups) and biology (30 samples divided into 4 collaborating groups). MDRS crew 76 collected 50 documented samples. Samples were screened/ analysed in the lab at the Habitat. Data were sent to remote science support teams in Europe and the US for further evaluation and detailed analysis. The geoscience investigations concerned mostly geological survey, documenting sample context and geochemical analyses of returned samples from the surrounding rock formations.

In-situ sample analysis

Approximately 40 samples have been analysed in the Habitat laboratory for chemical composition (XRF) and mineralogy content (XRD, Raman, VIS/NIR). Samples included clays, sandstones and volcanic ash layers of the Jurassic Morrison formation, pure crystals such as gypsum and calcite, petrified wood, desert varnish, endoliths and salt efflorescence. The sampling and analyses involved the set-up and maintenance of a detailed database with sample description, context geology and test results (Figs. 10–13). (a)

<image>



(b)





Fig. 9. (a) Installed biology and astrobiology laboratory. (b) Glovebox and sample handling. (c) Precision balance, centrifuge and PCR Peqlab. (d) MDRS microscopes. (e) Soil analysis kit.

Biology sample analysis

The primary goal of the biology investigations was the analysis of microbial communities living in the soil in interesting locations in the MDRS area, using protocols that are relevant to the search for organics and life on Mars, and to planetary protection. This investigation had a field aspect and a laboratory aspect: soil sampling was done in the field at depths of 10, 30 and 60 cm, in and out of EVA working conditions.

DNA extraction and PCR analysis were performed in the *in-situ* laboratory. DNA extracted from nine soil and water

samples of five different sampling sites were analysed in a first PCR run (Primus25 advanced; PeqLab) to detect bacterial DNA. Microscopy was used to investigate water samples for micro-organisms as well as floating particles concentrated by centrifugation.

DNA extraction and PCR analysis were also performed in a laboratory at Grand-Junction immediately after the campaign (Thiel *et al.* 2011) and in laboratories in Europe after the campaign (Direito *et al.* 2011). The microbial communities were studied *in situ* indicating already differences between Archaea and Bacteria in samples, and a later analysis of



Fig. 10. MDRS measured Raman spectrum of gypsum CaSO₄·2H₂O.

returned samples provided a more complete description of the relation of microbial communities' composition and phylogenetic analysis (Direito *et al.* 2011).

Soil sample mineral and organic post-mission analysis

The samples were divided (see Fig. 13) and sent to Earth-based laboratories for sophisticated analysis of PAHs (Orzechowska *et al.* 2010), of mineral matrix composition (Kotler *et al.* 2010) or of amino acids (Martins *et al.* 2011). Post-analysis studies determined the total carbon content (Orzechowska *et al.* 2010). A study of solid phase microextraction (SPME) method for fast screening and determination of PAHs in soil samples was performed, minimising sample handling and preserving chemical integrity of the sample. Complementary liquid extraction was used to obtain information on five- and six-ring PAH compounds. The measured concentrations of PAHs are, in general, very low, ranging from 1 to 60 ng/g (Orzechowska *et al.* 2010).

Core sample analysis

Using a Milwaukee drill (Stoker *et al.* 2009, 2010), we extracted cores down to 70 cm depth in a layered concretion-rich exhumed channel fragment. The drill site can also be analysed from side view near the MDRS habitat. The samples were transferred to a container preserving the stratification. The variation of the mineralogy and chemistry was analysed along the drill core. The samples show layers of quartz, gypsum and clays with some light mixing of those minerals. Visual, reflectance spectrometry, Raman and X-ray analysis was performed on extracts from the drill core (Fig. 12).

After these preliminary investigations, a more comprehensive campaign (DOMEX/EuroMoonMars 2010) was organized in November 2009 and February–March 2010 (Stoker *et al.* 2010, 2011; Clarke & Stoker 2011) using more advanced drilling systems, in conjunction with imaging and GPR reconnaissance.

Analysis of endolithic microbial mats

During the EuroGeoMars campaign we investigated on-site endolithic biota in relation to their environment. Endolithic microbes are extremophile organisms that live inside rocks or in pores between mineral grains. They can be not only lithotrophs but also phototrophs such as cyanobacteria. Phototrophs use light as energy source while lithotrophs oxidize inorganic compounds. They consume reduced elements from rocks, producing energy and free electrons used for ATP production. Litho-autotrophs obtain their carbon from CO₂ included in rocks and litho-heterotrophs from organic material. Endoliths can be slow to grow, due to limited nutrients. Endoliths may be present on Mars, and therefore it is interesting to study them in extreme environments on Earth in the context of life detection. An example is the endolithic, desiccation- and radiation-resistant cyanobacterium Chroococcidiopsis, a model organism for viability studies under Martian conditions. This prokaryote is able to survive in a Martian UV radiation environment when shielded by 1 mm of rock (Cockell et al. 2005). An acidophilic chemolithotroph from Rio Tinto was exposed to simulated Mars UV and atmospheric conditions under the protection of a Mars regolith analogue (Gómez et al. 2010).

We have found, studied *in situ* and sampled some endolithic mats near the MDRS research station. We investigated various areas at the base of 'Kissing Camel Ridge', a geological feature formed by an exhumed palaeo-channel in the Brushy Basin member of the Morrison formation (point G in Fig. 1). In this



Fig. 11. XRD spectra analysed in MDRS with match from database minerals: (a) XRD spectrum of Dakota Formation sandstone sample indicating gypsum and quartz (top), (b) XRD spectrum of Morrison sample indicating quartz and montmorrillonite clay (middle), (c) XRD spectrum of calcite evaporate (bottom).

area, some locations show concretions morphologically similar to the 'blueberries' observed by the Mars Exploration Rover in Meridiani, Mars (Clarke & Stoker 2011). A visual survey was conducted using colour imaging and the Cyborg astrobiologist experiment. The macroscopic pictures and close-up views indicated surface epilithic lichens.

After detachment of the crust, we confirmed the presence of microbial endolith population with green and orange-brown constituents, and the presence of endolith under a purplebrown coating. Samples of endolith attached to the host crust were taken to the MDRS laboratory. The visual and microscopic inspection confirms the presence of different layers: an outer varnish, a cemented crust, a brown microbial mat and a green mat attached to the rocks. Imaging was performed several times: a) before sampling, b) just after



Fig. 12. (top) Extracted drill core in 4 segments of 8–12 cm, (bottom) XRD spectrum of a core extract measured in the Hab, indicating the specific peaks of gypsum (purple), Quartz (yellow) and montmorillonite clay (green).



Fig. 13. Set of samples collected and documented *in situ* for mineral, organic and biology MDRS laboratory and post-mission analysis.

sampling using reference white calibration paper in order to quantify the colours of endolith on first exposure to light, and c) the same scene was revisited 1 week later. After detachment of the crust and varnish layers, the endoliths appear in three different colour units, with variations within 0.1-0.5 mm.



Fig. 14. (a) Context and protocol for endolith imaging and sterile sampling. The white paper was imaged for balance control allowing calibration of quantitative colour information. (b) Zoom of endolith (20 cm field of view) in natural balance colours. (c) Zoom (enhanced colour balance, 15 cm field of view) where endoliths appear as 3 distinct colour units clearly, after detachment of the crust and varnish layers. (d) Zoom of the same 15 cm field of view in black & white, where the endoliths are much harder to distinguish.



Fig. 15. Study of endolithic microbial communities in the MDRS lab: (top) sample view before microscopic inspection (FOV 2.5 cm), (bottom) optical microscope MDRS lab image of green and brown endolith communities in relation to minerals (field of view 3 mm).

Reflectance and Raman spectroscopic studies were performed on the varnish, crust, endolith and on the different adjacent mineral units. The analysis of the varnish coating with the XRF shows an overabundance of manganese, but little potassium, calcium or chromium. This is consistent with reddish iron and manganese oxides precipitates forming a dark and UV protecting layer. The microscopy indicates that the green endolith unit is mostly attached to gypsum grains (Figs. 14 and 15).

Post-mission analysis of the endolith samples was performed using a tabletop with scanning electron microscope (SEM) was conducted at NASA Ames Research Centre. This instrument is portable enough to allow field deployment. In the green area unit, we observed sheet-like structures layers of $100-300 \,\mu\text{m}$ in the interstitial pores between mineral grains. At ×4000, we detect submicron coccoids (Figs. 16–19).

In Fig. 17, autofluorescence microscope images show that the endoliths contain photosynthetic coccoid cells that are approximately 1 μ m in diameter. They form primarily in clusters, perhaps indicating the formation of a biofilm, which is potentially confirmed by the SEM images. The red colour is due to chlorophyll autofluorescence and highlights the clustering of the phototrophic bacteria.



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Fig. 16. Post-campaign study of endolithic microbial communities performed at NASA Ames: sample close-up context, FOV 1 cm (top), SEM magnification \times 250 , FOV 600 µm (middle) and SEM \times 4000 FOV 40 µm (bottom).

In Fig. 18, we see the smooth coccoidal cells that are roughly the same size as those present in the autofluoresence microscopy images. Again, the cells are clustered together and then come into contact with the more jagged, lighter-toned sandstone particles. This transition is not smooth, but rather



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Fig. 18. SEM view of endolith coccoid cells (smooth round features) together with sandstone micron particles (jagged shapes).

Fig. 17. Endolith images taken with an autofluorescence microscope. The red colour is due to chlorophyll autofluorescence. These images indicate that the endoliths are comprised of photosynthetic coccoid cells that are approximately 1 μ m in size. They mainly form in clusters perhaps indicating the formation of a biofilm, potentially confirmed by the SEM images.

the particles are packed in between clusters of cells before only sandstone grains remain in the next layer of the endolith.

In Fig. 19, the abiotic particles surrounding the coccoid cells appear suspended in a matrix. This matrix might be EPS that connects the cells in a biofilm. By forming a biofilm, these endolithic micro-organisms can better survive their nutrient poor environment, as available nutrients can concentrate on the surface. Also, the EPS allows the cells to attach to the surface of the sandstone for a more stable living environment. In addition, it is possible that there is more than one species of micro-organisms present in the biofilm. If so, the community of endolithic micro-organisms would benefit as the different species would be able to break down different types of nutrients, and they would be able to share in the limited nutrients available in the sandstone.

These results on the endoliths from *in situ* measurements to post laboratory sample analysis illustrate a possible astrobiology research avenue that can be performed at the MDRS analogue site.

Conclusions and perspectives

We have described the instruments and methods used for astrobiology research during the EuroGeoMars 2009 campaign. For technology field demonstration, the EuroGeoMars crew used instruments under realistic conditions (cameras, digital microscopy, XRD/XRF Spectrometers, Reflectance spectrometers, Raman spectrometer, GPR, magnetic susceptibility meter, ATP Luminometry meter and other sensors).



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Fig. 19. SEM image of clump of coccoid cells, with abiotic particles suspended in a matrix possibly made of extracellular polymeric substance (EPS) from within an endolith.

Remote sensing maps and geology reconnaissance were compared with surface *in-situ* investigations. A method of sample acquisition, curation and an analysis protocol was developed. The operations of remote rovers or their cooperation with field crew in EVA were investigated. Systems were demonstrated for communication, navigation and positioning. A Cyborg astrobiologist novelty detection algorithm was applied to rocks and landscape in different scenarios. The crew and remote support team used maps and database tools to integrate data and metadata from the sample context, rocks and subsequent measurements.

The EuroGeoMars research investigated processes relevant to Earth–Moon–Mars, in relation to geology and mineralogy. This included the analysis of samples from surface and from drill cores, in the field, in the habitat and later in laboratories. Several advanced and miniaturized instruments representative of those developed for future space missions were used, and provided *in-situ* constraints on mineralogy and organics.

Human crew-related aspects, i.e. (a) evaluation of the different functions and interfaces of a planetary habitat, (b) crew time organization in this habitat (Pletser & Foing, 2010). The evaluation of man-machine interfaces of astrobiology equipment is discussed in Thiel *et al.* (2011). Education, outreach, communications, multi-cultural and public relations aspects have been described in Foing *et al.* (2010a, b, c, d). The campaign experience and data analysis were used for a number of students' projects (bachelor, master and PhD research) and thesis reports.

In conclusion, the goals of EuroGeoMars 2009 field campaign were fulfilled by contributing to:

- (1) testing instruments, rovers, landers, EVA technologies, habitat and field laboratory;
- (2) performing field research in geology, sample analysis, exobiology;
- (3) studying human factors and crew aspects;
- (4) outreach and students' training.

In this paper, we have focused on the description of the instrument technology demonstration and the methodology of sampling and *in-situ* research. The scientific results from subsequent sample analysis are described in companion papers of this special issue. Lessons were learned relevant to human operations, or students training in relation to future robotic and human missions to the Moon or Mars (Pletser & Foing 2010; Thiel *et al.* 2011; De Crombrugghe *et al.* 2011).

As a follow-up of the EuroGeoMars 2009 campaign, ILEWG supported with instruments and experts, a campaign in Eifel Germany on human and robotic cooperation (Foing *et al.* 2010a; Groemer *et al.* 2010), and field campaigns by the CAREX project on 'Life in Extreme Environment' at Rio Tinto in Spain in Sept. 2009 and in Iceland in June 2010 (Direito *et al.* 2010; Gómez *et al.* 2010, 2011), and in Antarctica in December 2009 (De Vera *et al.* 2010) with a specific focus to use research instruments on the field for *in-situ* analysis of bio-organics and minerals in samples.

A EuroMoonMars/DOMEX (Drilling on the Moon and Mars in Human Exploration) campaign was performed in November 2009 and February–April 2010, using analogue missions to develop the approach for using human crews to perform science activities on the Moon and Mars, with the novelty of exploration and sampling of the subsurface using a suite of drills from back-pack carried to large truck-carried systems (Foing *et al.* 2010a, b, c, d; Stoker *et al.* 2010, 2011). A series of EuroMoonMars-DOMEX five crew rotations were deployed for 2 weeks each time performing complementary aspects of this research.

The experience and results from these campaigns in sites representing specific planetary analogue conditions can contribute to the preparation of field tests for Moon and Mars exploration, for missions such as MSL, Exomars, Moon or Mars Sample Return. This will include the investigation of geological and geochemical context, drilling of cores and sampling, remote control of field rovers, cameras and instruments. Also future human missions to the Moon or Mars can be prepared by evaluating crew operations, simulations and EVAs, and interaction with instruments. Terrestrial campaigns including tele-robotics and EVAs enable preparation under both real and simulated conditions for science, technology, research, operational, organizational and communication aspects associated with future robotic and human exploration missions.

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References

- Battler, M.M., Clarke, J.D.A. & Coniglio, M. (2006). Possible analog sedimentary and diagenetic features for Meridiani Planum sediments near Hanksville, Utah: implications for Martian Field studies. In *Mars Analog Research*, ed. Clarke, J.D.A., pp. 55–70. American Astronautical Society Science and Technology Series 111.
- Bibring, J.-P., Langevin, Y., Gendrin, A., Gondet, B., Poulet, F., Berthé, M., Soufflot, A., Arvidson, R., Mangold, N., Mustard, J. & Drossart, P. (2005). Mars Surface Diversity as Revealed by the OMEGA/Mars Express Observations. *Science* **307**(5715), 1576–1581.
- Bibring, J.P., Langevin, Y., Mustard, J.F., Poulet, F., Arvidson, R., Gendrin, A., Gondet, B., Mangold, N., Pinet, P. & Forget, F. (2006). Global mineralogical and aqueous Mars history derived from OMEGA/ Mars express data. *Science* **312**(5772), 400–404.
- Blake, D.F., Sarrazin, P., Bish, D.L., Chipera, S.J., Vaniman, D.T., Collins, S., Elliott, S.T. & Yen, A.S. (2007). Progress in the development of CheMin: a definitive mineralogy instrument on the Mars science laboratory (MSL'09) Rover. LPI Contribution No. 1338, p. 1257.
- Borst, A., Peters, S., Foing, B.H., Stoker, C., Wendt, L., Gross, C., Zavaleta, J., Sarrazin, P., Blake, D., Ehrenfreund, P. *et al.* (2010). Geochemical results from EuroGeoMars MDRS Utah 2009 campaign. LPI Contribution No. 41, p. 2744.
- Chin, G., Brylow, S., Foote, M., Garvin, J., Kasper, J., Keller, J., Litvak, M., Mitrofanov, I., Paige, D., Raney, K., Robinson, M., Sanin, A., Smith, D., Spence, H., Spudis, P., Stern, S.A. & Zuber, M. (2007). Lunar reconnaissance orbiter overview: The Instrument Suite and Mission. *Space Sci. Rev.* 129(4), 391–419.
- Christensen, P.R., Bandfield, J.L., Hamilton, V.E., Ruff, S.W., Kieffer, H. H., Titus, T.N., Malin, M.C., Morris, R.V., Lane, M.D., Clark, R.L. *et al.* (2001). Mars global surveyor thermal emission spectrometer experiment: investigation description and surface science results. *J. Geophys. Res.* **106** (E10), 23823–23872.
- Clarke, J.D.A. & Pain, C.F. (2004). From Utah to Mars: regolithlandform mapping and its application. In *Mars Expedition Planning*,

ed. Cockell, C.C., pp. 131-160. American Astronautical Society Science and Technology Series 107.

- Clarke, J.D.A. & Stoker, C.R. (2011). Concretions in inverted and exhumed channels near Hanksville Utah: implications for Mars. *Int. J. Astrobiol.* 10, 161–175.
- Cockell, C.S., Schuerger, A.C., Billi, D., Friedmann, E.I. & Panitz, C. (2005). Effects of a simulated Martian UV flux on the cyanobacterium, Chroococcidiopsis sp 029. Astrobiology 5(2), 127–140.
- COSPAR Planetary Exploration Committee (PEX) Report, Ehrenfreund, P. et al. (2010). http://cosparhq.cnes.fr/PEX_Report2010_June22a.pdf
- de Crombrugghe, G., Le Maire, V., Denies, J., Jago, A., Van Vynckt, D., Reydams, M., Mertens, A., de Lobkowicz, Y. et al., in preparation.
- de Vera, J.P.P., Leya, T., Lorek, A., Koncz, A., de La Torre Noetzel, R., Kozyrovska, N., Burlak, O. & Foing, B. (2010). Photosynthesis and its implications for space research, astrobiology science conference 2010: evolution and life: surviving catastrophes and extremes on earth and beyond, LPI No. 1538, p. 5139.
- Direito, S., Foing, B.H., Mahapatra, P., Gomez, F. & Rull, F. (2010). Sample collection and analysis from CAREX Field Workshop at Rio Tinto. LPI Contribution No. 1538, p. 5648.
- Direito, S.O.L., Ehrenfreund, P., Marees, A., Staats, M., Foing, B. & Röling, W.F.M. (2011). A wide variety of putative extremophiles and large beta-diversity at the Mars desert research station (Utah). *Int. J. Astrobiol.* **10**, 191–207.
- Ehrenfreund, P., Foing, B.H., Stoker, C., Zavaleta, J., Quinn, R., Blake, D., Martins, Z., Sephton, M., Becker, L., Orzechowska, G. *et al.* (2010). EuroGeoMars field campaign: sample analysis of organic matter and minerals. LPI Contribution No. 41, p. 1723.
- Ehrenfreund, P., Röling, W., Thiel, C., Quinn, R., Septhon, M., Stoker, C., Direito, S., Kotler, M., Martins, Z., Orzechowska, G.E., Kidd, R. & Foing, B.H. (2011). Astrobiology and habitability studies in preparation for future Mars missions: trends from investigating minerals, organics and biota. *Int. J. Astrobiol.* **10**, 239–253.
- Feldman, W.C., Maurice, S., Lawrence, D.J., Little, R.C., Lawson, S.L., Gasnault, O., Wiens, R.C., Barraclough, B.L., Elphic, R.C., Prettyman, T.H. *et al.* (2001). Evidence for water ice near the lunar poles. *J. Geophys. Res.* **106**(E10), 23231–23252.
- Foing, B.H. (2008). Reports to COSPAR from the International Lunar Exploration Working Group (ILEWG). *Adv. Space. Res.* **42**, 238.
- Foing, B.H., Racca, G.D., Marini, A., Evrard, E., Stagnaro, L., Almeida, M., Koschny, D., Frew, D., Zender, J., Heather, J., Grande, M., Huovelin, J., Keller, H.U., Nathues, A., Josset, J.L., Malkki, A., Schmidt, W., Noci, G., Birkl, R., Iess, L., Sodnik, Z. & McManamon, P. (2006). SMART-1 mission to the Moon: status, first results and goals. *Adv. Res.* 37(1), 6–13.
- Foing, B.H., Racca, G.D., Josset, J.L., Koschny, D., Frew, D., Almeida, M., Zender, J., Heather, D., Peters, S., Marini, A., Stagnaro, L., Beauvivre, S., Grande, M., Kellett, B., Huovelin, J., Nathues, A., Mall, U., Ehrenfreund, P. & McCannon, P. (2008) SMART-1 highlights and relevant studies on early bombardment and geological processes on rocky planets. *Phy. Scr.* 130, 014026.
- Foing, B.H., Richards, R., Sallaberger, C. & ICEUM7 Participants (2008b). Toronto Lunar Declaration 2005. Adv. Space Res. 42, 242.
- Foing, B.H., Wu, J. & ICEUM8 Participants (2008c). Beijing lunar declaration 2006. Adv. Space Res. 42, 244.
- Foing, B.H., Bhandari, N., Goswami, J.N. & ICEUM6 Participants (2008d). Udaipur Lunar Declaration 2004. Adv. Space Res. 42, 240.
- Foing, B.H., Espinasse, S., Wargo, M., di Pippo, S. & ICEUM9 Participants (2008e). Sorrento lunar declaration 2007. Adv. Space Res. 42, 246.
- Foing, B.H., Batenburg, P., Drijkoningen, G., Slob, E., Poulakis, P., Visentin, G., Page, J., Noroozi, A., Gill, E., Guglielmi, M. *et al.* (2009). ExoGeoLab Lander/rover instruments and EuroGeoMars MDRS campaign. LPI Contribution No. 40, p. 2567.
- Foing, B.H., Barton, A., Blom, J.K., Mahapatra, P., Som, S., Jantscher, B., Page, J., Zegers, T., Stoker, C., Zavaleta, J. *et al.* (2010a). ExoGeoLab Lander, rovers and instruments: tests at ESTEC and Eifel volcanic field. LPI Contribution No. 41, p. 1701.

- Foing, B.H., Boche-Sauvan, L., Stoker, C., Ehrenfreund, P., Wendt, L., Gross, C., Thiel, C., Peters, S., Borst, A., Zhavaleta, J. *et al.* (2010b). ExoHab and EuroGeoMars campaigns: human exploration and astrobiology. LPI Contribution No. 1538, p. 5625.
- Foing, B.H., Mahapatra, P., Boche-Sauvan, L., Som, S., Page, J., Stoker, C., Zhavaleta, J., Sarrazin, P., Blake, D., Poulakis, P. *et al.* (2010c). ExoGeoLab test bench for landers, rovers and astrobiology. LPI Contribution No. 1538, p. 5477.
- Foing, B.H., Stoker, C., Zhavaleta, J., Ehrenfreund, P., Quinn, R., Blake, D., Martins, Z., Sephton, M., Becker, L., Orzechowska, G. *et al.* (2010d). Eurogeomars field campaign: sample analysis of organic matter and minerals. LPI Contribution No. 1538, p. 5656.
- Gendrin, A., Mangold, N., Bibring, J.-P., Langevin, Y., Gondet, B., Poulet, F., Bonello, G., Quantin, C., Mustard, J., Arvidson, R. & Le Mouélic, S. (2005). Sulfates in Martian Layered Terrains: The OMEGA/ Mars Express View. *Science* **307**(5715), 1587–1591.
- Godfrey, A.E., Everitt, B.L., & Duque, J.F.M. (2008). Episodic sediment delivery and landscape connectivity in the Mancos Shale badlands and Fremont River system, Utah, USA. *Geomorphology* 102, 242–251.
- Gómez, F., Mateo-Martí, E., Prieto-Ballesteros, O., Martín-Gago, J. & Amils, R. (2010). Protection of chemolithoautotrophic bacteria exposed to simulated Mars environmental conditions. *Icarus* 209, 482.
- Gómez, F., Walter, N., Amils, R., Rull, F., Klingelhöfer, A.K., Kviderova, J., Sarrazin, P., Foing, B., Behar, A., Fleischer, I., Parro, V., Garcia-Villadangos, M., Blake, D., Martin Ramos, J.D., Direito, S., Mahapatra, P., Stam, C., Venkateswaran, K. & Voytek, M. (2011). Multidisciplinary integrated field campaign to an acidic Martian Earth Analogue with astrobiological interest: Rio Tinto. *Int. J. Astrobiol.* 10, 291–305.
- Goswami, J.N. (2010). An Overview of the Chandrayaan-1 Mission, LPI Contribution No. 1533, p 1591.
- Greeley, R., Foing, B.H., McSween, H.Y., Neukum, G., Pinet, P., van Kan, M., Werner, S.C., Williams, D.A. & Zegers, T.E. (2005). Fluid lava flows in Gusev crater, Mars. *J. Geophys. Res.* **110**(E5), E05008.
- Groemer, G., Stumptner, W., Foing, B., Blom, J.K., Perrin, A., Mikolajczak, M., Chevrier, S., Direito, S., Olmedo-Soler, A., Zegers, T.E. *et al.* (2010). ILEWG Eifel 2009 campaign: astronaut extravehicular surface/subsurface activities and human aspects. LPI Contribution No. 41, p. 1680.
- Gross, C., Wendt, L., McGuire, P.C., Bonnici, A., Foing, B.H., Souza-Egipsy, V., Bose, R., Walter, S., Ormö, J., Díaz-Martínez, E. *et al.* (2010). The cyborg astrobiologist: testing a novelty detection algorithm at the Mars desert research station (MDRS), Utah. LPI Contribution No. 41, p. 2457.
- Haruyama, J., Matsunaga, T., Ohtake, M., Morota, T., Honda, C., Yokota, Y., Torii, M., Ogawa, Y. & The Lism Working Group (2008). Global lunar-surface mapping experiment using the Lunar Imager/ Spectrometer on SELENE. *Earth. Planets Space* **60**, 243–255.
- Haruyama, J., Ohtake, M., Matsunaga, T., Morota, T., Honda, C., Yokota, Y., Abe, M., Ogawa, Y., Miyamoto, H., Iwasaki, A. et al. (2009). Long-lived volcanism on the lunar farside revealed by SELENE terrain camera. *Science* **323**(5916), 905.
- Head, J.W., Neukum, G., Jaumann, R., Hiesinger, H., Hauber, E., Carr, M., Masson, P., Foing, B., Hoffmann, H., Kreslavsky, M. *et al.* (2005a). Tropical to mid-latitude snow and ice accumulation, flow and glaciation on Mars. *Nature* **434**(7031), 346.
- Head, J.W., Neukum, G., Jaumann, R., Hiesinger, H., Hauber, E., Carr, M., Masson, P., Foing, B., Hoffmann, H., Kreslavsky, M. *et al.* (2005b). Planetary science: are there active glaciers on Mars? *Nature* 438(7069), 10.
- Hendrikse, J., Foing, B.H., Monaghan, E., Stoker, C., Zavaleta, J., Selch, F., Ehrenfreund, P., Wendt, L., Gross, C., Thiel, C. *et al.* (2010). Highlights from remote controlled rover for EuroGeoMars MDRS campaign. LPI Contribution No. 41, p. 2435.
- ICEUM9 (sci.esa.int/iceum9) and Sorrento Lunar Declaration (2007). http:// sci.esa.int/science-e/www/object/index.cfm?fobjectid=41506
- ICEUM10 (sci.esa.int/iceum10) and Cape Canaveral Lunar Declaration (2008). http://sci.esa.int/science-e/www/object/index.cfm?fobjectid=43654

ICEUM11 (http://sci.esa.int/science-e/www/object/index.cfm?fobjectid= 46028) & Beijing Lunar Declaration (2010). http://sci.esa.int/science-e/ www/object/index.cfm?fobjectid=47170

- ILEWG Reports and ICEUM Declarations 2006-2010 (http://sci.esa.int/ ilewg)
- Jolliff, B.L., Gillis, J.J., Haskin, L.A., Korotev, R.L. & Wieczorek, M.A. (2000). Major lunar crustal terranes: surface expressions and crust-mantle origins. JGR105 4197J.
- Kato, M., Sasaki, S., Tanaka, K., Iijima, Y. & Takizawa, Y. (2008). The Japanese lunar mission SELENE: science goals and present status. *Adv. Space Res.* 42(2), 294–300.
- Kim, S.S., Carnes, S., Haldemann, A., Ulmer, C., Ng, E. & Arcone, S. (2005). Miniature ground penetrating radar, CRUX GPR, IEEEAC paper #1365.
- Klingelhöfer, G., Morris, R.V., Bernhardt, B., Schröder, C., Rodionov, D.S., de Souza, P.A., Yen, A., Gellert, R., Evlanov, E.N., Zubkov, B., Foh, J., Bonnes, U., Kankeleit, E., Gütlich, P., Ming, D.W., Renz, F., Wdowiak, T., Squyres, S.W. & Arvidson, R.E. (2004). Jarosite and Hematite at Meridiani Planum from Opportunity's Mössbauer Spectrometer. *Sci.* 306(5702) 1740–1745.
- Kotler, M., Quinn, R., Martins, Z., Foing, B. & Ehrenfreund, P. (2011). Analysis of Mineral Matrices of planetary soils analogs from the Utah Desert. *Int. J. Astrobiol.* **10**, 221–229.
- Lucey, P.G., Blewett, D.T. & Hawke, B. (1998). Mapping the FeO and TiO₂ content of the lunar surface with multispectral imagery. J. Geophys. Res. 103, 3679L.
- Martins, Z., Sephton, M.A., Foing, B.H. & Ehrenfreund, P. (2011). Extraction of amino acids from soils close to the Mars Desert Research Station (MDRS), Utah. Int. J. Astrobiol. 10, 231–238.
- McGuire, P.C., Gross, C., Wendt, L., Bonnici, A., Souza-Egipsy, V., Ormö, J., Díaz-Martínez, E., Foing, B.H., Bose, R., Walter, S. *et al.* (2010). The cyborg astrobiologist: testing a novelty detection algorithm on two mobile exploration systems at Rivas Vaciamadrid in Spain and at the Mars Desert Research Station in Utah. *Int. J. Astrobiol.* 9, 11.
- McKay, C.P. & Stoker, C.R. (1989). The early environment and its evolution on Mars: implications for life. *Rev. Geophys.* 27, 189–214.
- MEPAG Report (2007). http://www.lpi.usra.edu/pss/presentations/200707/ arvidson_mepag.pdf
- Murray, J.B., Muller, J.-P., Neukum, G., Werner, S.C., van Gasselt, S., Hauber, E., Markiewicz, W.J., Head, J.W., Foing, B.H., Page, D. *et al.* (2005). Evidence from the Mars express high resolution stereo camera for a frozen sea close to Mars' equator. *Nature* **434**, 352.
- Neukum, G., Jaumann, R., Hoffmann, H., Hauber, E., Head, J.W., Basilevsky, A.T., Ivanov, B.A., Werner, S.C., van Gasselt, S., Murray, J.B., McCord, T., HRSC Co-Investigator Team (2004). Recent and episodic volcanic and glacial activity on Mars revealed by the High Resolution Stereo Camera. *Nature* 432(7020), 971–979.
- Ono, T., Kumamoto, A., Nakagawa, H., Yamaguchi, Y., Oshigami, S., Yamaji, A., Kobayashi, T., Kasahara, Y., Oya, H. (2009). Lunar radar sounder observations of subsurface layers under the nearside Maria of the Moon. *Science* **323**(5916), 909.

- Orzechowska, G.E., Kidd, R., Foing, B.H., Kanik, I., Stoker, C. & Ehrenfreund, P. (2011). Analysis of mars analog soil samples using solid phase microextraction, organic solvent extraction and Gas Chromatography/Mass Spectrometry. *Int. J. Astrobiol.* **10**, 209–219.
- Parker, M., Zegers, T., Kneissl, T., Ivanov, B., Foing, B. & Neukum, G. (2010). 3D structure of the Gusev Crater region. *Earth Planet. Sci. Lett.* 294, 411.
- Pieters, C.M., Goswami, J.N., Clark, R.N., Annadurai, M., Boardman, J., Buratti, B., Combe, J.-P., Dyar, M.D., Green, R., Head, J.W. *et al.* (2009). Character and spatial distribution of OH/H₂O on the surface of the Moon seen by M3 on Chandrayaan-1. *Science* **326**, 568.
- Plester, V. & Foing, B. (2011). European contribution to human aspect investigations for future planetary habitat definition studies: field tests at MDRS on crew time utilisation and habitat interfaces. *Micrograv. Sci. Technol.* 23(2), 199.
- Poulet, F., Bibring, J.-P., Mustard, J.F., Gendrin, A., Mangold, N., Langevin, Y., Arvidson, R.E., Gondet, B. & Gomez, C. (2004). Phyllosilicates on Mars and implications for early martian climate. *Nature* 438(7068), 623–627.
- Som, S.M., Foing, B.H. & Exogeolab Team. (2010). Testing portable Raman spectrometry for astrobiology. LPI Contribution No. 1538, p. 5085.
- Squyres, S.W., Grotzinger, J.P., Arvidson, R.E., Bell, J.F., Calvin, W., Christensen, P.R., Clark, B.C., Crisp, J.A., Farrand, W.H., Herkenhoff, K.E., Johnson, J.R., Klingelhöfer, G., Knoll, A.H., McLennan, S.M., McSween, H.Y., Morris, R.V., Rice, J.W., Rieder, R. & Soderblom, L.A. (2004). In Situ Evidence for an Ancient Aqueous Environment at Meridiani Planum, Mars. *Science* **306**(5702), 1709–1714.
- Stoker, C., Clarke, J., Direito, S., Blake, D., Martin, K., Zavaleta, J. & Foing, B.H. (2011). Mineralogical, chemical, organic and microbial properties of subsurface soil cores from mars desert research station, a phyllosilicate and sulfate mars analog site for MSL. *Int. J. Astrobiol.* 10, 269–289.
- Stoker, C., Foing, B., Zavaleta, J. & Clark, J. (2009). Drilling on the Moon and Mars: human exploration simulation experiments. EPSC Abstr. 4, EPSC2009-659.
- Stoker, C.R., Zavaleta, J., Bell, M., Direto, S., Foing, B., Blake, D. & Kim, S. (2010). Drilling on the Moon and Mars: developing the science approach for subsurface exploration with human crews. LPI Contribution No. 41, p. 2697.
- Thiel, C., Ehrenfreund, P., Foing, B., Pletser, V. & Ullrich, O. (2011). PCRbased analysis of microbial communities during the EuroGeoMars campaign at Mars Desert Research Station, Utah. *Int. J. Astrobiol.* 10, 177–190.
- Thiel, C., Wills, D. & Foing, B.H. (2009). PCR-based detection of microbial communities during the EuroGeoMars MDRS campaign. EPSC Abstr. 4, EPSC2009-660.
- Vondrak, R., Keller, J., Chin, G. & Garvin, J. (2010). Lunar Reconnaissance Orbiter (LRO): Observations for lunar exploration and science. *Space Sci. Rev.* 150(1–4), 7–22.
- Wendt, L., Mahapatra, P., Gross, C., Borst, A., Foing, B.H. & Exogeolab Team, Eurogeomars Team. (2009). Raman investigations of the EuroGeoMars Campaign. EPSC Abstr. 4, EPSC2009-457.