

## Short Communication

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# Efficacy of artemisinin–naphthoquine phosphate against *Schistosoma haematobium* adult flukes: dose–effect relationship and tegumental alterations

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## Abstract

*Schistosoma haematobium* and *Schistosoma mansoni* infections have broadly overlapping geographical distributions. Praziquantel is the only treatment for human schistosomiasis, so drug tolerance and/or resistance are major concerns. Artemisinin–naphthoquine phosphate (CO-ArNp), an artemisinin-based combination therapy endorsed by the World Health Organization as a gold standard therapy for malaria, has also been identified as a promising treatment for *S. mansoni*. In this *in vitro* study, we tested the effect of 1–40 µg/ml CO-ArNp on *S. haematobium* worms, and inspected tegumental changes by using scanning electron microscopy (SEM), aiming to determine if this combination therapy has a broad-spectrum antischistosomal activity. Incubation of *S. haematobium* adults with 20 or 30 µg/ml CO-ArNp caused 100% mortality of worms within 72 or 48 h, respectively. SEM examination showed extensive tegumental alterations such as oedema, constriction, shortening and loss of spines, fissuring, sloughing and perforation, resulting in exposure of the underlying basal lamina, mainly in treated male schistosomes. Besides the well-established potent efficacy, bio-availability, tolerability and safety of the antimalarial artemisinin–naphthoquine phosphate combined therapy, these results may also suggest its possible utilization as a new broad-spectrum antischistosomal agent.

## Introduction

Schistosomiasis is a challenging disease worldwide, with at least 92% of the 207 million estimated cases occurring in the African endemic zones (WHO, 2017). Infection with the neglected human schistosome, *Schistosoma haematobium* (Rinaldi *et al.*, 2011), causes urogenital schistosomiasis, a chronic morbid disease that starts during early life (King, 2010; Shiff *et al.*, 2010; Terer *et al.*, 2013). Control of schistosomiasis is mainly by preventive chemotherapy, whereby school-aged children and other high-risk groups are given praziquantel (PZQ) without previous diagnosis (Utzinger *et al.*, 2011). The extensive use of PZQ in the absence of other alternatives needs careful monitoring to identify early signs of therapeutic failure (Doenhoff *et al.*, 2009). Accordingly, the World Health Organization (WHO) has realized the exigency of detecting new alternatives to PZQ.

Both *Plasmodium* and schistosome parasites degrade the ingested haemoglobin of the host into amino acids and the potentially toxic haem that is subsequently discarded as haemozoin (Oliveira *et al.*, 2005), thus an effective antimalarial drug targeting haem would have a harmful effect against schistosomes (Xiao *et al.*, 2011). Drugs targeting haem are of interest, as free haem does not exist in non-infected individuals (Portela *et al.*, 2012). Interestingly, the antimalarial drugs artemisinin and its derivatives (artemether, arteether, artesunate, etc.) have shown antischistosomal activities (Utzinger *et al.*, 2007). Moreover, Xiao *et al.* (2011) reported promising *in vivo* synergistic effects of mefloquine combined with low or moderate effective doses of artemisinins, indicating that artemisinins combined with other antimalarial drugs may have potential application against schistosomiasis.

The WHO has recommended artemisinin-based combination therapies (ACTs, artemisinins + other antimalarial drugs) as the most effective treatment for malaria (WHO, 2016). Furthermore, previous studies have revealed their potent antischistosomal activity (Boulanger *et al.*, 2007; Mohamed *et al.*, 2009; Keiser *et al.*, 2010). The compound naphthoquine phosphate tablet (CO-ArNp), a single oral dose ACT, is composed of artemisinin and naphthoquine phosphate (tetra-aminoquinoline) (Benjamin *et al.*, 2012). The notable *in vivo* and *in vitro* effects of the combined artemisinin–naphthoquine phosphate therapy against *Schistosoma mansoni* have been reported (El-Beshbishi *et al.*, 2013, 2015). These results prompted us to examine whether a comparable efficacy can be detected against *S. haematobium*, in which case the treatment could

be used as a broad-spectrum antischistosomal therapy in overlapping schistosomiasis endemic foci.

In this paper, we report for the first time the *in vitro* activity of artemisinin–naphthoquine phosphate against adult *S. haematobium*, as well as the morphological and tegumental changes observed by scanning electron microscopy (SEM) as indices of the biological activity of schistosomes (Moraes *et al.*, 2011).

## Materials and methods

### Compounds and reagents

Compound naphthoquine phosphate tablets (artemisinin : naphthoquine phosphate at a ratio of 3 : 1) were a gift from Xiamen WinWord Industrial Co., Ltd (Xiamen, China). Praziquantel was obtained from Sigma-Aldrich (USA). All cell culture grade reagents were obtained from Gibco (Thermo Fisher Scientific, USA) except dimethyl sulfoxide (DMSO), which was obtained from CARLO ERBA Reagents (France). Foetal bovine serum (FBS) was obtained from Lonza Biologics (Switzerland).

### Animals and parasites

The life cycle of *S. haematobium* Egyptian strain was maintained in *Bulinus truncatus* and male Syrian golden hamsters (*Mesocricetus auratus*) at the Schistosome Biological Supply Centre, Theodor Bilharz Research Institute, Imbaba, Giza, Egypt. Briefly, 60–80 g golden hamsters were infected by percutaneous route with 350–400 freshly shed cercariae of *S. haematobium*. The hamsters were kept in controlled environmental conditions (20–22°C, 12/12 h dark/light cycle), with Rodent Blox and water *ad libitum*.

After 12 weeks of infection, adult *S. haematobium* worms were collected from golden hamsters by aseptic perfusion of the portal and mesenteric veins, with sterile saline–sodium citrate (Duvall & DeWitt, 1967). Worms were then transferred to Roswell Park Memorial Institute medium (RPMI-1640) and washed several times. Viable, contractile worms showing intact tegument, as observed by light microscopy, were used in this study.

### *In vitro* artemisinin–naphthoquine phosphate treatment of *Schistosoma haematobium*

Adult *S. haematobium* worms were maintained in sterile, flat-bottom Corning Costar 24-well plates (Sigma-Aldrich), using 3–4 pairs of worms per well. The culture medium in each well contained 2 ml RPMI-1640 supplemented with L-glutamine + 20% FBS (heat-inactivated) + 160 µg/ml gentamycin + 300 IU/ml penicillin + 300 µg/ml streptomycin (Ramirez *et al.*, 2007). *Schistosoma haematobium* cultures were incubated with CO-ArNp at concentrations of 1–40 µg/ml. Pairs of *S. haematobium* adults were cultured in medium without and medium with 0.8% DMSO (the highest concentration of solvent used) as negative controls, or exposed to 10 µg/ml PZQ as positive controls. All assays were performed at least twice over two separate studies. Cultures were incubated at 37°C in 5% CO<sub>2</sub> atmosphere for 72 h. Worms were observed, under a stereomicroscope every 24 h for changes in general appearance, motor activity and mortality rate.

Treatment was considered lethal when no worm motility was observed during 2 minutes of examination, after agitation of the plate to ensure high parasitic visibility. Accordingly, the mortality rates, LC<sub>50</sub> and LC<sub>90</sub> of CO-ArNp and PZQ against *S.*

*haematobium* adult worms were calculated by using the statistical program SPSS v. 20 (IBM, Armonk, USA).

### Scanning electron microscopic examination

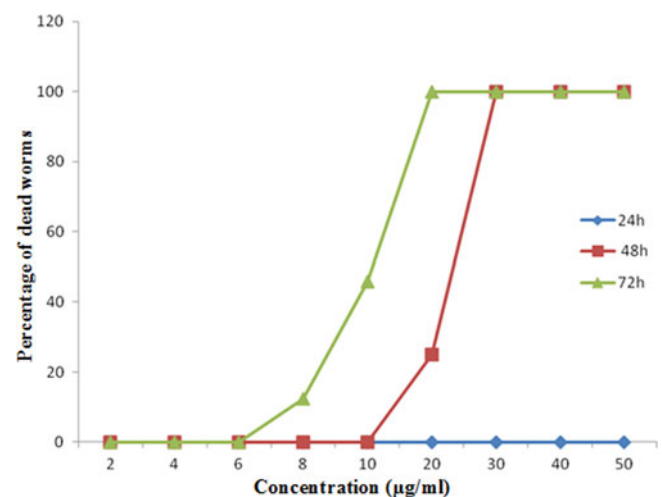
A total of eight pairs of *S. haematobium* worms exposed to 20 µg/ml CO-ArNp for 48 h, as well as unexposed worms (negative controls), were sampled and washed twice with PBS, then fixed in 2.5% glutaraldehyde, examined and photographed using SEM (Inspect S, FEI Company, Holland) (Shaw & Erasmus, 1987).

## Results and discussion

### Effect of artemisinin–naphthoquine phosphate on the viability of *Schistosoma haematobium* adult worms

Over the 72 h, all *S. haematobium* worms cultured in RPMI-1640 drug-free medium revealed distinctive wavy as well as peristaltic motion, with maintenance of male–female mating. Worms incubated with PZQ showed immediate muscle contraction, without separation of worms, and all worms were dead 48 h after incubation. Interestingly, incubation of worms with the different concentrations (1–40 µg/ml) of CO-ArNp led to dissociation of *S. haematobium* pairs and their irreversible debilitation within 24 h; also, all schistosomes displayed only minimal activity, with slow convulsions and abnormal contractions, which were more prominent in female worms that died earlier than males, while male worms appeared shrunken and markedly more coiled than females, and all worms were dead after incubation with 20 and 30 µg/ml for 72 and 48 h, respectively (fig. 1). The LC<sub>50</sub> and LC<sub>90</sub> of CO-ArNp recorded 72 h post incubation were 11.1 µg/ml and 18.5 µg/ml, while those of PZQ were 0.2 µg/ml and 0.3 µg/ml, respectively, as shown in table 1.

In comparison, treatment of *S. mansoni* worms with 20 and 40 µg/ml of the same drug combination killed all parasites at the same time points, but males were more afflicted and died earlier than females (El-Beshbishi *et al.*, 2015). In an *in vitro* study performed by Eissa *et al.* (2011), the authors declared that



**Fig. 1.** *In vitro* effect of artemisinin–naphthoquine phosphate at various concentrations on the mortality of adult *Schistosoma haematobium* under culture conditions. The data were generated from at least two independent studies. Note that the bars show the range of percentage of dead worms in the experiments, and the intervals on the x-axis are not equal.

**Table 1.** *In vitro* schistosomicidal activity of artemisinin–naphthoquine phosphate combination and a reference drug (praziquantel) on adult *Schistosoma haematobium* worms after three days' exposure.

Drug	LC <sub>50</sub> (µg/ml)	LC <sub>90</sub> (µg/ml)
Artemisinin–naphthoquine phosphate	11.1	18.5
Praziquantel (reference drug)	0.2	0.3

miltefosine killed all worms after five days of incubation with 10 µg/ml, and the drug was more active on *S. haematobium* than on *S. mansoni* adult worms. Indeed, *S. haematobium* and *S. mansoni*, two major human schistosomes, have different sensitivities to particular antischistosomal drugs, which may be attributed to specific intrinsic biochemical processes (Camacho & Agnew, 1995), and depend on the enzyme(s) involved (Pica-Mattoccia *et al.*, 1997). *Schistosoma haematobium* adults exposed to 20 µg/ml artemether for 72 h, showed no apparent changes (Xiao *et al.*, 2001). Our results may be attributed to drug–drug interaction between naphthoquine phosphate and artemisinin exerting a more toxic effect on the worms than using artemisinins alone. Artemisinins interact with haemin and other iron systems, producing toxic compounds and free radicals, which cause oxidative damage of the parasite (Xiao *et al.*, 2001). Additionally, it is well known that 4-aminoquinoline derivatives interfere with haemozoin formation, hence the accumulated free haem kills the parasite through oxidative stress (Corrêa Soares *et al.*, 2009).

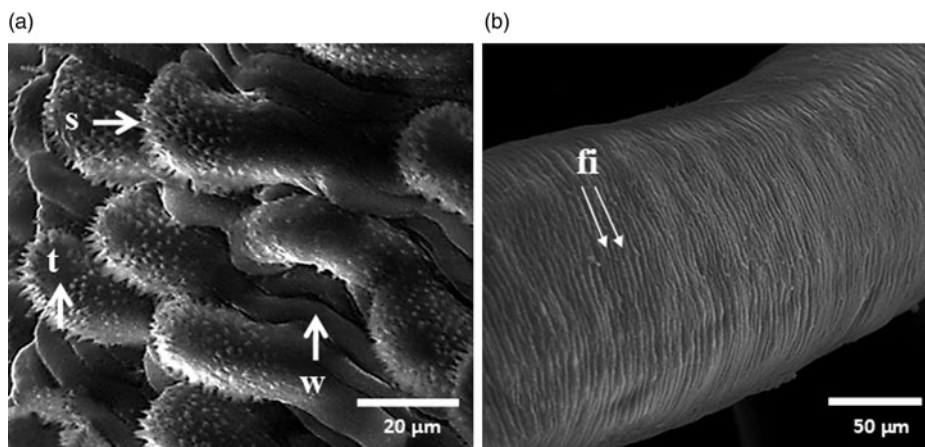
#### Ultrastructure analysis of artemisinin–naphthoquine phosphate-induced surface damage in *Schistosoma haematobium* adult worms

The characteristics of the tegument of untreated worms (fig. 2a, b) were compared with those exposed to CO-ArNp. Male and female worms incubated with 20 µg/ml CO-ArNp for 48 h showed surface membrane ultrastructural damage (figs 3 and 4). Tegumental changes were more prominent and outspread in male than female schistosomes. Generally, all inspected male worms showed extensive oedema of the tegument, with thinning of tegumental ridges. There were also pronounced changes in the tubercles in the form of reduction in size and number, with shortening and loss of spines leaving a smooth surface (fig. 3a).

Erosion of the tegument in several areas of the body, cleft formation with exposure of the subtegumental tissue (fig. 3b), exfoliation of the tegument and roughness of the surface (fig. 3c), as well as extreme distortion of the tegumental folds, with concomitant collapse and deformity of the tubercles, were encountered (fig. 3c, d). Moreover, *in vitro* incubated worms with CO-ArNp showed longitudinal muscle contraction, and the worms were bent, shortened and deformed, with marked corrugation at the anterior end (fig. 3e). Besides, the gynaecophoric canal revealed contraction deformity at the edges and formation of protuberance in the tegument (fig. 3f). Disfigurement of the ventral sucker was also noticed in the form of marked oedema in > 50% of treated worms (fig. 3g), while marked contraction and deformity, with erosion and vesicle formation, were seen in the remaining males, as well as evident shrinkage and wrinkling of the area between the oral and ventral suckers (fig. 3h). Xiao *et al.* (2000) declared that such damage to the suckers may predispose schistosomes to loss of power to adhere to the vasculature, hence making uptake of nutrients more difficult. In turn, this may affect the worms' maturity, fecundity and ability to deposit eggs – the major contributory factor to schistosomiasis morbidity.

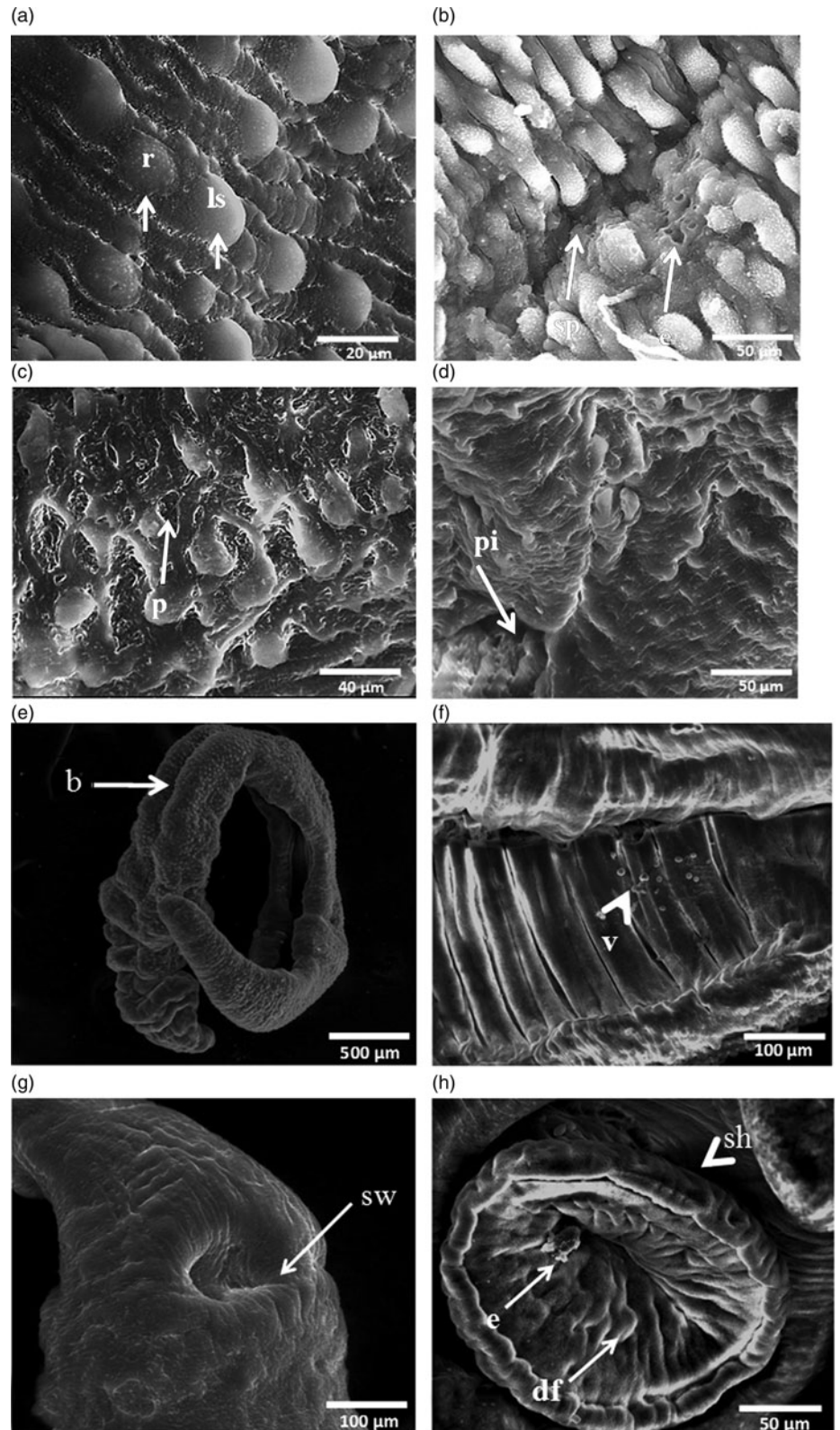
Treatment with the same dose of CO-ArNp and for the same time point revealed less extensive tegumental changes in females, as the damage was localized to the mid-body. Tegumental swelling with absence of normal arranged fissures was evident in all female worms examined; besides, constriction and collapse of the tegument with furrow formation and vesiculation (fig. 4a). In addition, the tegumental surface of some worms (< 50%) was distorted, exfoliated (fig. 4b) and eroded, leaving profound fissures with exposure of the subtegumental tissues (fig. 4c).

The tegument has a pivotal role in protection, immunomodulation, sensation, nutrient ingestion and synthesis of some proteins, osmoregulation, and excretion (Skelly & Wilson, 2014). It is noteworthy that alterations in the tegument structure deteriorate its functions and lead to worm death (Shaw & Erasmus, 1987). The results of the present study demonstrated the schistosomicidal capability of CO-ArNp, wherein *S. haematobium* females revealed faster and higher susceptibility to the combined regimen compared to males, although male worms showed the most prominent and extensive tegumental damage. Corroborating our results, Soliman & Ibrahim (2005) reported that both atorvastatin alone and its combination with medroxyprogesterone acetate administered to experimentally infected hamsters led to more



**Fig. 2.** Normal surface ultrastructure of *Schistosoma haematobium* adult worms (controls). (a) Normal dorsal tegument of a male worm (×5000); S: spines, t: tubercles, w: wrinkles. (b) Normal tegument of a female worm (×3000); fi: fissures.

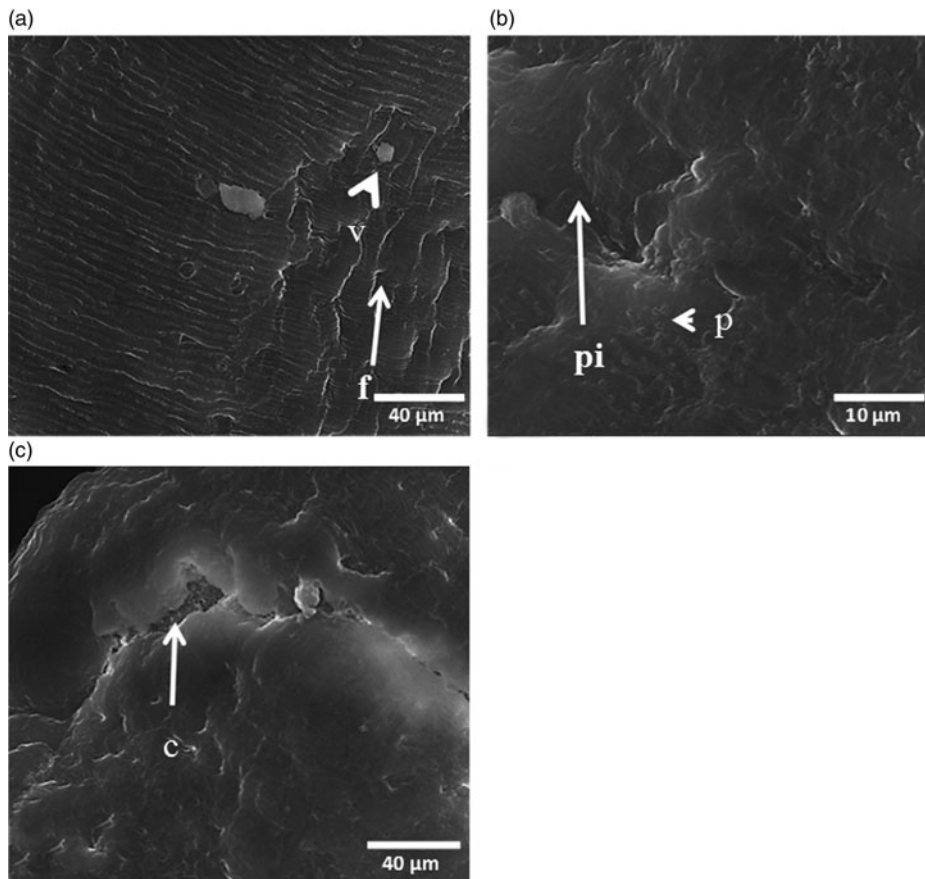




**Fig. 3.** SEM micrographs of adult *Schistosoma haematobium* male worms incubated with 20 µg/ml CO-ArNp for 48 h. (a) Tegumentary changes in a male worm, including reduction in size and number of tubercles, and loss of spines (×5000); r: tubercles reduced in size, ls: tubercles showing loss of spines. (b) Tegumental erosion and splitting (×2400); sp: splitting, e: erosion. (c) Tegumental peeling off, with deformity of the tubercles (×3000); p: peeling. (d) Extensive oedema of the tegument, with deep pitting, as well as marked atrophy of the tubercles (×1600); pi: pitting. (e) Male *S. haematobium*, showing contraction and deformity at the anterior end, with bending at the mid-body region (×270); b: binding. (f) Gynaecophoric canal with small vesicles and contraction deformity at the edges (×1000); v: vesicles. (g) Marked swelling of the ventral sucker and the tegument around it (×1200); sw: swelling. (h) Ventral sucker, showing contraction and deformity, erosion and vesicles, together with shrinkage of the area between suckers (×1600); df: deformity, e: erosion, sh: shrinkage.

marked tegumental damage in *S. haematobium* male worms than females. Similarly, Eraky *et al.* (2016) reported more pronounced tegumental alterations in male worms incubated with phytol than females exposed to the same dose and time period. Importantly, the differences in the severity and extent of morphological

changes, and death rates and time, caused by Co-ArNp may be ascribed to the ability of the drug to attack different targets in both genders, based on the structural, physical and biochemical variations between schistosome males and females (Shaw & Erasmus, 1987). Drugs attacking schistosomes in a sex-dependent



**Fig. 4.** SEM micrographs of adult *Schistosoma haematobium* female worms incubated with 20 µg/ml CO-ArNp for 48 h. (a) Tegumental changes in the mid-body, including loss of the normally arranged furrows in some parts, with contraction of the tegument as well as furrows and vesicle formation ( $\times 3000$ ); f: furrows, v: vesicle. (b) Marked oedema and pitting of the tegument, as well as peeling off ( $\times 1000$ ); pi: pitting, p: peeling. (c) Swollen female tegument with deep crake formation, exposing the subtegumental tissue ( $\times 3000$ ); c: crake.

pattern could control disease morbidity and limit the spread of infection by decreasing the number of eggs released.

Considering the results of this study, artemisinin–naphthoquine phosphate exerts *in vitro* effects on *S. haematobium* adult worms, as it affects the morphology, tegumental surface and viability of the exposed worms. Therefore, this work opens up perspectives for future research on this drug *in vivo* to disclose its mechanism(s) of action.

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**Conflict of interest.** None.

**Ethical standards.** All study procedures were approved by Mansoura Faculty of Medicine Institutional Research Board (MFM-IRB approval number: R/16.05.38), Mansoura University, Mansoura, Egypt, and Theodor Bilharz Research Institute (TBRI), Imbaba, Giza, Egypt, and complied with the relevant international animal ethics regulations for the care and use of experimental animals.

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