

Phase-specific developmental and reproductive strategies in the desert locust

K. Maeno^{1,2} and S. Tanaka^{1*}

¹Laboratory of Insect Life Cycles and Physiology, National Institute of Agrobiological Sciences at Ohwashi, Tsukuba, Ibaraki 305-8634, Japan; ²Graduate School of Science and Technology, Kobe University, Kobe, Hyogo 657-8501, Japan

Abstract

Locusts modify developmental and reproductive traits over successive generations depending on the population density. A trade-off between developmental rate and body size and between progeny size and number is often observed in organisms. In this study, we present evidence that this rule is evaded by desert locusts, *Schistocerca gregaria* Forskål, which often undergo outbreaks. Under isolated conditions, large hatchlings, typical of the gregarious forms, grow faster but emerge as larger adults than do small hatchlings typical of the solitary forms, except for some individuals of the latter group that undergo extra molting. Under crowded conditions, large and small hatchlings grow at a similar rate, but the former become larger adults than the latter. Small hatchlings show a trade-off between development time and body size at maturation, but this constraint is avoided by large hatchlings. Phase-specific, as well as body size-dependent, differences are also detected in reproductive performance. As adult body size increases, females of a solitary line produce more but slightly smaller eggs, whereas those of a gregarious line produce more and larger eggs. Total egg mass per pod is larger in gregarious forms than in solitary forms. A trade-off between egg size and number is shown by a solitary line but not by a gregarious line that produces relatively large eggs with similar numbers of eggs per pod. These results suggest that phase transformation involves not just a shift of resource allocation but also an enhanced capability expressed in response to crowding.

Keywords: locusts, maternal effects, phase polyphenism, progeny size, *Schistocerca gregaria*, trade-off

(Accepted 21 December 2007)

Introduction

Locusts often undergo outbreaks that last over several generations (Lecoq, 2005; Huis *et al.*, 2007). The desert locust, *Schistocerca gregaria* Forskål, and the migratory locust, *Locusta migratoria*, show density-dependent phase polyphenism in behavioural, morphological and physiological

characteristics (Faure, 1932; Uvarov, 1966, 1977; Pener, 1991; Heifetz & Applebaum, 1995; Pener & Yerushalmi, 1998). Although the behavioural aspect has been studied intensively (Simpson *et al.*, 1999; Seidelmann & Ferenz, 2002; Ferenz and Seidelmann, 2003; Tanaka & Zhu, 2003; Hassanali *et al.*, 2005), the morphological and developmental changes are also important in explaining locust outbreaks because some of these characteristics are directly related to population growth. In this study, we present evidence suggesting that solitary and gregarious locusts of *S. gregaria* show different patterns of nymphal development and reproduction in response to crowding conditions. That is, solitary

*Author for correspondence
Fax: +81-29-838-6110
E-mail: stanaka@affrc.go.jp

(isolation-reared) nymphs are constrained by a trade-off between developmental time and the final body size attained, whereas gregarious (crowd-reared) nymphs evade this constraint and grow faster or as fast as solitary ones without getting smaller as adults. In this study, we produced hatchlings of various body sizes, ranging from small to large, typical for solitary and gregarious forms, respectively, and reared them under either isolated or crowded conditions to determine the effects of hatchling body size on nymphal growth and adult body size. Few studies have manipulated progeny size and investigated the influence of juvenile size on growth rates in arthropods (Fox & Czesak, 2000).

In studies to compare reproductive performance, locusts are often kept in small cages individually or in large cages as a group. Although the differences in the reproductive potential between crowded and isolated locusts are well documented, the conclusions are not always consistent. Certain factors, such as competition for food and egg-laying space, relating more to the experimental methods than to real phase characteristics, may contribute to these differences (Pener, 1991). In this study, we minimized such differences by rearing all mated females individually in cages of the same size with either two males (crowded conditions) or with no males (isolated conditions). By collecting eggs from females with known body size, we analyzed the relationships between adult body size and fecundity in terms of egg size, egg number and egg biomass per egg pod between solitary and gregarious lines.

Materials and methods

Insects and rearing conditions

The *S. gregaria* colony used in the present study has been described (Tanaka & Yagi, 1997). The rearing method was described elsewhere (Maeno & Tanaka, 2007). Briefly, nymphs and adults were reared at $32 \pm 1^\circ\text{C}$ under a light-dark 16:8 h photoperiod and 40–70% relative humidity in a well-ventilated room. Locusts were kept either in a group of 100 individuals in a large cage ($42 \times 22 \times 42$ cm; crowded conditions) or in isolation in small cages ($28 \times 15 \times 28$ cm; isolated conditions). They were supplied with fresh leaves of orchard grass, cabbage and wheat bran. A gregarious line had been maintained at a density of *ca.* 100 individuals for more than 20 generations, and a solitary line was established from the gregarious colony by rearing nymphs and adults individually in small cages, except for a short period for mating (Maeno & Tanaka, 2007). Plastic cups (diameter, 9 cm; height, 5 cm) filled with clean moist sand were placed in cages to collect egg pods. All experiments were carried out with 3rd and 4th solitary generations and with >20th gregarious generations. The grass used was raised by the Field Management Section of NIAS at Ohwashi.

Hatchling groups

Body size and colour at hatching are closely correlated; for example, hatchlings with darker body colouration are heavier (Hunter-Jones, 1958; Tanaka & Maeno, 2006, 2008). We used body colour to divide hatchlings into five groups of different body sizes. To obtain hatchlings of various sizes, each of 30 20-day-old sexually mature isolation-reared females was paired with a sexually mature male for various

lengths of time ranging from ten hours to ten days (Maeno & Tanaka, 2007). Females kept longer with a male tended to produce hatchlings with more extensive black patterns (Maeno & Tanaka, 2008). Hatchling body colouration was scored on the day of hatching using five colour grades ranging from entirely green to heavily black, as described by Maeno & Tanaka (2007). In this study, these five categories were called hatchling groups, or HGs. Hatchlings of each HG to be reared in isolation had been weighed individually. The mean and SD were 13.0 ± 1.6 mg ($n=50$), 14.8 ± 1.3 mg ($n=50$), 15.8 ± 0.9 mg ($n=50$), 17.2 ± 0.9 mg ($n=50$) and 19.8 ± 1.3 mg ($n=50$) in HGs 1, 2, 3, 4 and 5, respectively.

Determination of the number of nymphal stadia

S. gregaria normally passes through five or six nymphal stadia (Rao & Gupta, 1939; Maeno *et al.*, 2004). Two methods were used to determine the number of nymphal stadia. In one method, the number of nymphal stadia was counted by checking the nymphs every day for ecdysis. In the other method, the eye stripes in the adult stage were counted. Adults with five nymphal stadia have six eye stripes, whereas those with six nymphal stadia have seven (Rao & Gupta, 1939). Under crowded conditions, it was difficult to follow the history of ecdyses for each individual, so only the second method was used.

Measurements of adult body weight

Within 24 h after the final molt, adults that had not started feeding were weighed to give an estimate of adult size. The weight gain per day was determined by dividing adult body weight by the number of days required for nymphal development. In this case, the hatchling body weight (7–25 mg) was not subtracted from the adult body weight (*ca.* 1000–2600 mg) because it was small and the differences among individuals were negligible.

Egg pod collection and measurements of egg size and egg number

Egg pods were collected from female adults of a solitary line, as described above. Females of a gregarious line were weighed at adult emergence and marked individually with white paint (Pentel, EZL31-W, Japan). They were kept together with males in a group of about 100 individuals in a large cage during the first 12 days of adult life. To obtain egg pods from individual females with known body size, females were removed from the large cage and held individually in small cages with two sexually mature males. In *S. gregaria*, pairing of a female with a single male induces crowding effects on the progeny that are as strong as rearing her with many males (Hunter-Jones, 1958). Egg pods collected were incubated at $32 \pm 1^\circ\text{C}$. Egg length was measured using an ocular micrometer installed in a microscope two days after oviposition. A total of ten eggs were randomly chosen from each egg pod and placed on a piece of moist filter paper (9 cm dia.) to avoid desiccation before measurements. The number of eggs per egg pod was also counted at that time. In our unpublished observations, egg weight (y , mg) was highly correlated with egg length (x , mm) ($y = 2.09x - 7.288$; $r = 0.939$, $n = 200$, $P < 0.001$). Thus, this equation was used to estimate egg weight from egg

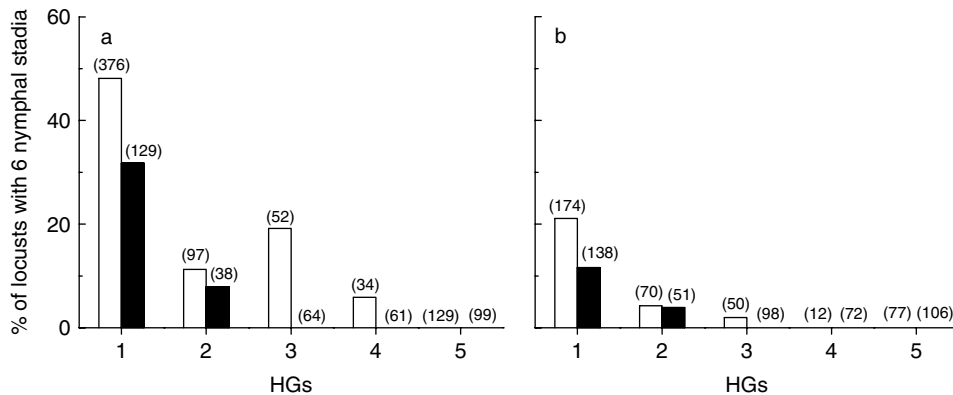


Fig. 1. Proportions of individuals with six nymphal stadia in different hatchling size groups (HGs) of *Schistocerca gregaria* reared under (□) isolated conditions or (■) crowded conditions. (a) females, (b) males. Numbers on bars indicate *n*.

length. We adopted this method because eggs were often coated with sand particles glued with egg foam and removing them without damaging the eggs was very time consuming. After measurements, eggs were returned to moist sand and incubated at the same temperature until hatching. Only egg pods deposited after 25 days of adult emergence were used.

Statistics

Data for developmental and reproductive traits were mainly compared by a *t*-test or ANOVA using Stat View, version 6 (SAS Institute, Cary, North Carolina, USA). The number of eggs that correlated with adult body size was also analyzed by ANCOVA when it was appropriate. Ratios of egg biomass to adult body size were analyzed by the Mann-Whitney *U* test.

Results

Nymphal development and density

Desert locusts undergo either five or six nymphal stadia, as mentioned above. First, we examined the effects of hatchling body size and rearing density on this trait because it affects the duration of nymphal development. Hatchlings categorized into five different hatchling size groups (HGs) were reared under either isolated or crowded conditions. HG 1 consists of smallest hatchlings typical of solitary forms, whereas HG 5 comprises largest ones typical of gregarious forms. The incidence of nymphs exhibiting six stadia depended on HG, rearing density and sex (fig. 1). The highest incidence was obtained when females derived from HG 1 (smallest hatchlings) were reared under isolated conditions. No nymphs exhibiting six stadia appeared from HG 5 (largest hatchlings) in females or from HGs 4 and 5 in males. The incidence of nymphs exhibiting six stadia within HG 1 was higher under isolated conditions than under crowded conditions in either females ($\chi^2=10.429$; $df=1$; $P<0.001$) or males ($\chi^2=5.310$; $df=1$; $P<0.05$). The duration of nymphal development was significantly shorter in locusts with five nymphal stadia (mean \pm SD: 30.0 ± 2.2 days, $n=175$ for females; and 29.2 ± 2.2 days, $n=152$ for males) than in locusts with six nymphal stadia

(mean \pm SD: 33.6 ± 2.5 days, $n=170$ for females; $t=-13.969$, $df=343$, $P<0.001$; 33.8 ± 2.3 days, $n=44$ for males; $t=-11.965$, $df=194$, $P<0.001$). Within HG 1, adult body weight was significantly smaller in individuals with five nymphal stadia (mean \pm SD: 1947 ± 142 mg, $n=175$ for females; and 1394 ± 99 mg, $n=152$ for males) than in those with six nymphal stadia (mean \pm SD: 2166 ± 182 mg, $n=170$ for females; $t=-12.490$, $df=343$, $P<0.001$; 1508 ± 107 mg, $n=44$ for males; $t=-6.577$, $df=194$, $P<0.001$). Because the developmental performance in each HG was influenced by the number of nymphal stadia, the following analyses on nymphal development were conducted mainly for locusts with five nymphal stadia.

Rearing density influenced the duration of nymphal development (fig. 2a,b). In all HGs, nymphal development was faster under crowded conditions than under isolated conditions. Under isolated conditions, nymphs tended to grow slightly faster, as hatchlings were bigger (fig. 2a,b), and a negative correlation was found between individual hatchling body weight and developmental time for both sexes (data not shown: $r=-0.211$, $n=439$, $P<0.001$ for females; $r=-0.154$, $n=312$, $P<0.01$ for males). Under crowded conditions, developmental time did not vary with HG for either sex ($P>0.05$ each). However, hatchling body size influenced adult body weight under both isolated and crowded conditions; larger hatchlings attained a larger body weight at adult emergence (fig. 2c,d). Interestingly, rearing density resulted in differences in adult body weight in small hatchlings only. Daily weight gain during the nymphal stage was consistently greater under crowded conditions than under isolated conditions (fig. 2e,f), although the differences for HGs 1 and 2 in females were not significant ($P>0.05$; fig. 2e). In both sexes, larger hatchlings showed greater daily weight gain. Figure 3 summarizes the relationships between developmental rate (the inverse of the number of days required for nymphal development) and adult body weight for different HGs. For females (fig. 3a), a negative relationship indicating a trade-off was found in the smallest three HGs. For males (fig. 3b), a negative relationship was found only in HG 1. These results indicate that small hatchlings grow faster under crowded conditions than under isolated conditions at the expense of the final body size. Larger hatchlings also grew faster under crowded conditions than

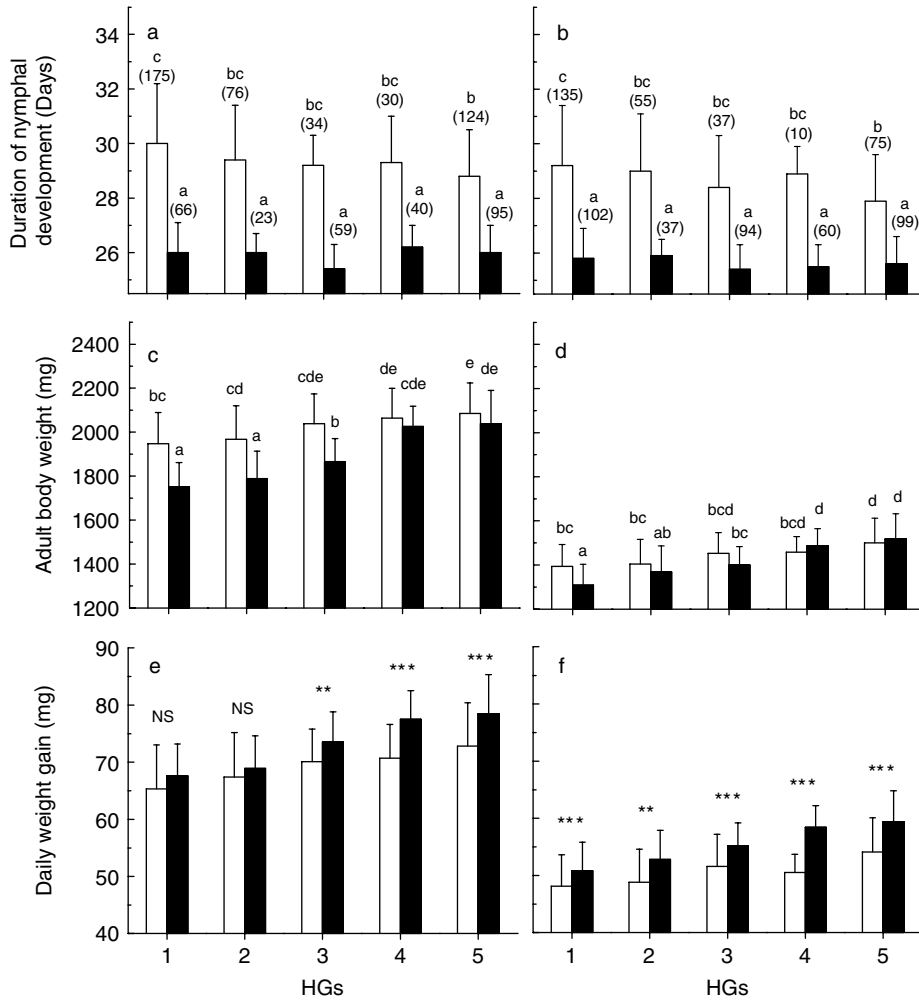


Fig. 2. Effects of hatching body size on various developmental traits in *Schistocerca gregaria*. Duration of nymphal development (a) and (b), adult body weight (c) and (d) and daily weight gain (e) and (f) in locusts of different HGs reared under (□) isolated conditions or (■) crowded conditions. (a), (c) and (e), females; (b), (d) and (f), males. Means with one side of SD are presented. Numbers in parentheses in panel (a) and (b) indicate *n*. Different letters in each panel indicate significant differences at $P < 0.05$ by Scheffé's test. Daily weight gain was tested by Mann-Whitney *U* test: NS, not significant; **, $P < 0.01$; ***, $P < 0.001$.

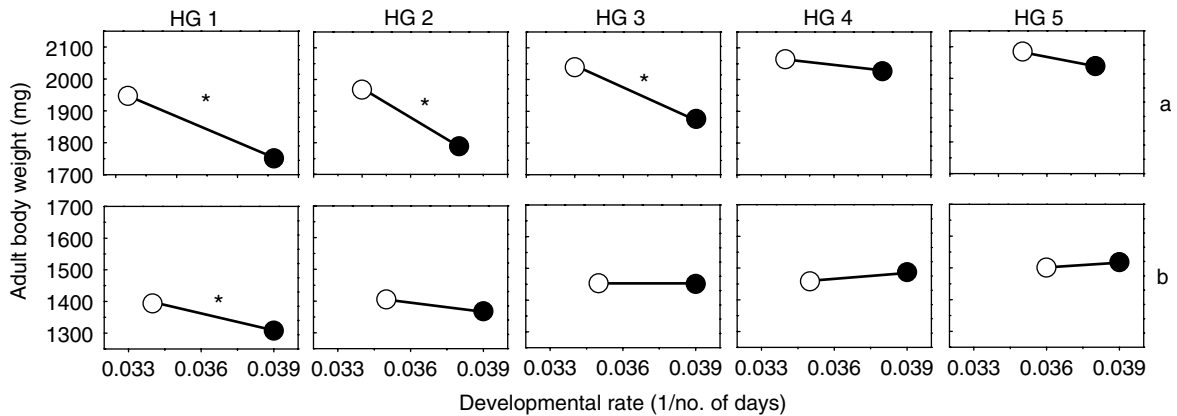


Fig. 3. Relationships between developmental rate of nymphs and adult body weight in different HGs of *Schistocerca gregaria* reared under (○) isolated conditions or (●) crowded conditions. Data are based on results in fig. 2. (a) females, (b) males. Asterisks indicate significant differences in both adult body weight (by *t*-test) and developmental rate (by Mann-Whitney *U* test) between the two groups at $P < 0.05$. Sample sizes are given in fig. 2.

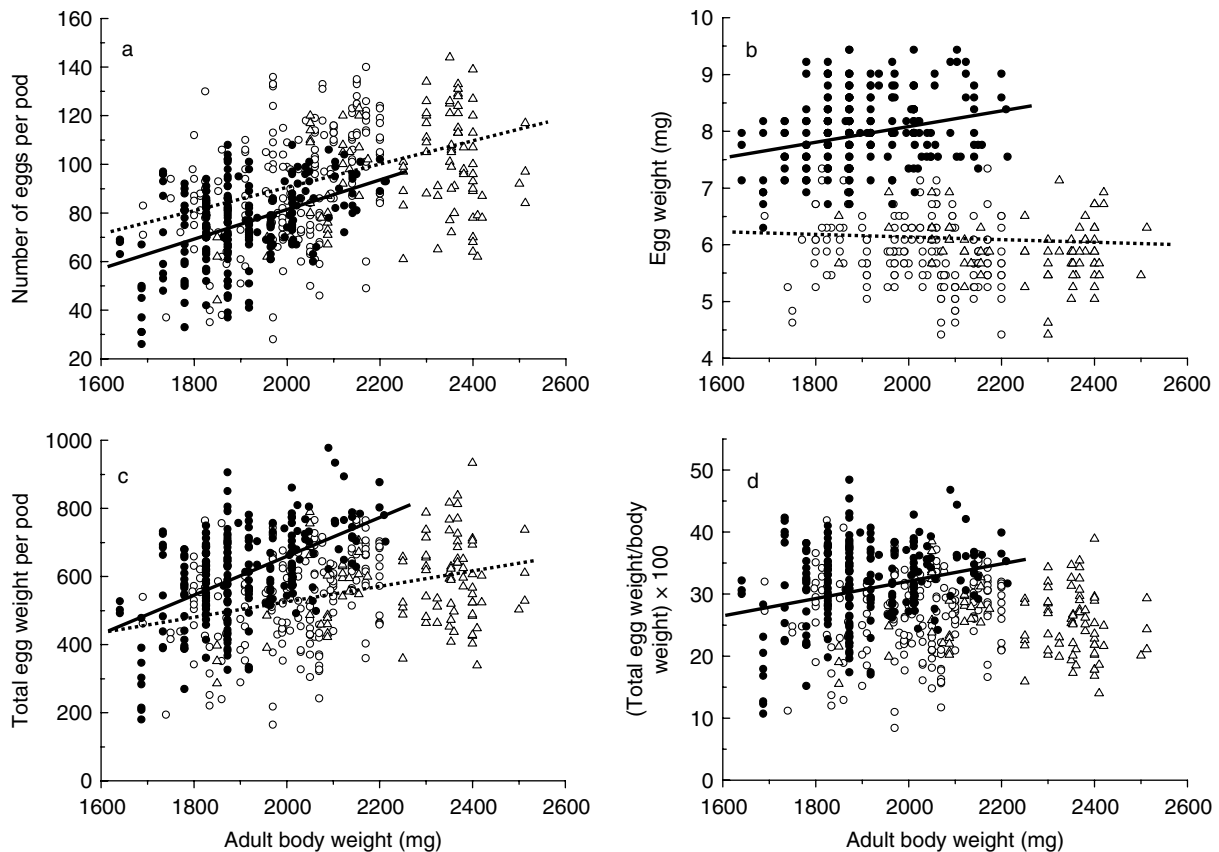


Fig. 4. Relationships between body weight at adult emergence and various reproductive traits in female adults of a solitary and a gregarious line of *Schistocerca gregaria*. (a) the number of eggs per pod, (b) egg weight, (c) total egg weight per pod, and (d) total egg weight relative to adult body weight. Eggs produced by solitary adults with (○) five or (△) six nymphal stadia are shown separately, together with those produced by (●) gregarious adults. Regression lines for the gregarious line are drawn by black lines, and those for the solitary line are dotted lines. $n=193$ and $n=103$ for solitary individuals with five and six nymphal stadia, respectively, and $n=234$ for gregarious individuals.

under isolated conditions, but without becoming smaller as adults.

Reproductive traits and density

Female adults from a solitary (isolation-reared) and a gregarious (crowd-reared) line were housed individually or with two male adults, and the number of eggs, individual egg weight and total egg weight per egg pod were determined (fig. 4). For the solitary line, adults with five and six nymphal stadia are presented separately, because adult body weight was significantly smaller in the former, as mentioned above. The number of eggs per egg pod increased with adult body weight in both lines (fig. 4a; $P<0.001$). ANCOVA with adult body weight as the covariate indicated significant differences in the number of eggs between the two groups of solitary adults ($F_{1,294}=7.22$, $P<0.01$), as well as between the two lines ($F_{1,527}=23.69$, $P<0.001$). Egg weight varied positively with adult body weight in the gregarious line (fig. 4b; $r=0.269$, $n=234$, $P<0.001$) but negatively in the solitary line ($r=-0.128$, $n=296$, $P<0.05$). Mean egg weight was significantly greater in the gregarious line (mean \pm SD: 8.02 ± 0.63 mg) than in the solitary one (5.89 ± 0.54 mg, $t=41.66$, $df=528$, $P<0.001$).

No significant difference was found in egg weight between the two groups within the solitary line ($P>0.05$). Total egg biomass per pod, calculated based on egg weight and the number of eggs per pod, was positively correlated to adult body weight in the two lines (fig. 4c). It was significantly larger in the gregarious line (604.6 ± 134.9 mg, $n=234$) than in the solitary line (554.1 ± 13.2 mg, $n=296$, $t=4.358$, $df=528$, $P<0.001$). However, no significant difference was found in this trait between the solitary females with five nymphal stadia (538.8 ± 131.9 mg, $n=193$) and those with six nymphal stadia (582.8 ± 122.6 mg, $n=103$, ANCOVA $F_{1,294}=2.267$, $P>0.05$) after adult body size was adjusted. Body weight-specific egg production (total egg mass/adult body weight) was relatively constant over a wide range of adult body weights in the solitary line (fig. 4d; $r=-0.031$, $n=296$, $P>0.05$) but rapidly increased with adult body weight in the gregarious line ($r=0.253$, $n=234$, $P<0.001$).

Trade-off between egg size and number

Figure 5 illustrates the relationship between egg weight and number per pod produced by adults of a solitary and a gregarious line. The overall correlation involving

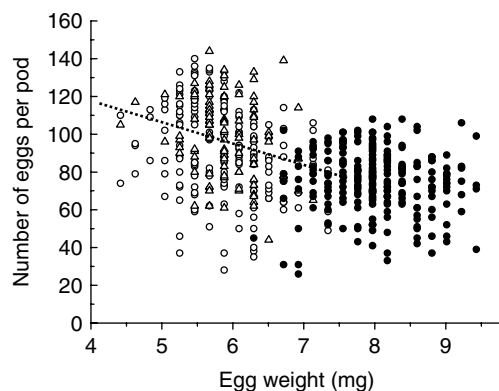


Fig. 5. Relationships between the weight and number of eggs per pod laid by solitary (isolation-reared) adults with (○) five or (△) six nymphal stadia and by (●) gregarious (crowd-reared) adults of *Schistocerca gregaria*. A negative correlation is found only in the solitary line (dotted line). $n = 296$ in the solitary line and $n = 234$ in the gregarious line.

all eggs produced by the two lines was highly significant ($r = -0.456$, $n = 530$, $P < 0.001$). In the solitary line alone, the negative correlation was less obvious but still significant for egg pods produced by all adults ($r = -0.273$, $n = 296$, $P < 0.001$) and those produced by adults with five nymphal stadia ($r = -0.286$, $n = 193$; $P < 0.001$) or six nymphal stadia ($r = -0.265$, $n = 103$, $P < 0.01$). Unexpectedly, the corresponding correlation was not statistically significant for the gregarious line ($P > 0.05$).

Discussion

In *S. gregaria*, the density experienced by females as adults determines the progeny size (Faure, 1932; Chauvin, 1941; Hunter-Jones, 1958); solitary females produce small green hatchlings and gregarious females large black hatchlings. The phase-dependent hatchling body colouration has been claimed to be determined after egg deposition by a water-soluble pheromonal factor produced by the accessory gland of the female parent (McCaffery *et al.*, 1998; Simpson *et al.*, 1999; Hägele *et al.*, 2000; Simpson & Miller, 2007). However, recent studies have failed to reproduce this result and have concluded that hatchling body colour, as well as body size, are pre-determined in the ovary of the mother (Tanaka & Maeno, 2006, 2008). The variation in progeny size caused by the maternal effect influences the number of nymphal stadia (Hunter-Jones, 1958) and, thus, the development and body size at maturation, as demonstrated in this study. We investigated developmental performance of various sizes of *S. gregaria* hatchlings under isolated and crowded conditions. We confirmed that extra molting occurs only in small hatchlings (Hunter-Jones, 1958) and found that the incidence of extra molting also depends on the rearing density of the nymphs and the sex.

Because the number of nymphal stadia influences the duration of nymphal development and body size at maturation, our analysis of developmental traits was conducted mainly for locusts with five nymphal stadia. We found that, irrespective of hatchling body size, nymphal

development was consistently faster under crowded conditions than under isolated conditions. Although locusts have received much attention (Faure, 1932; Uvarov, 1966), few reliable data are available about phase-dependent differences in duration of nymphal development and the information is not consistent. In *S. gregaria* and *L. migratoria*, nymphal growth has been reported to be faster in crowd-reared locusts than in those reared in isolation in some studies (Kennedy, 1956; Uvarov, 1966, 1977; Pener, 1991; Heifetz & Applebaum, 1995), whereas the reverse conclusion has been reported in other studies (Staal, 1961; Applebaum & Heifetz, 1999). Most studies were conducted without considering the variation in the number of nymphal stadia. Crowding, particularly in late-stadium nymphs, can easily cause a shortage of food, and special care is required to avoid a secondary effect of crowding. In the present study, we analyzed locusts with five or six nymphal stadia separately and changed the grass twice a day for late-stadium nymphs to ensure that they had food *ad libitum* throughout nymphal life. Other factors influencing nymphal development and reproductive performance include temperature, humidity and food (Uvarov, 1966). In the present study, variation in these conditions was minimized by rearing locusts in a similar way except for rearing density.

Rapid development often results in smaller body size. This negative relationship, or trade-off, is well documented in various organisms (Stearns, 1992). In *S. gregaria*, rearing density affects developmental rate and adult body size. We found a trade-off between the two variables for relatively small hatchlings, which grow faster but emerge as smaller adults under crowded conditions than under isolated conditions. However, such a trade-off is not found in large hatchlings, which grow faster under crowded conditions than under isolated conditions without becoming smaller adults (fig. 3). This finding may suggest an important feature of this locust, which often undergoes outbreaks. In solitary forms, hatchlings are small and take a long time to mature (with low developmental rates) but can attain a large adult body size. At low population density at which food is less likely to be limiting, large adult body size, rather than rapid growth, may be more important in terms of fitness. Conversely, in gregarious forms, hatchlings are large and grow rapidly. These characteristics are likely to be adaptive under crowded conditions because large hatchlings are more tolerant to desiccation and fasting than small ones (Albrecht & Blackith, 1960), and rapid development would reduce the time of exposure to predators. Indeed, large hatchlings rarely undergo extra molting even if reared in isolation. With increased nymphal growth efficiency, gregarious locusts can accomplish both rapid growth and large adult body size by evading the trade-off by which solitary locusts are constrained.

A comparison of reproductive performance between a solitary and a gregarious line revealed another important feature of this locust. ANCOVA demonstrated that the number of eggs per pod depends on the body size of the female parent and confirmed that solitary females produce more eggs than gregarious ones (Uvarov, 1966). The present study also confirmed that gregarious locusts produce larger eggs than solitary locusts (Uvarov, 1966). Interestingly, with increased body size of the female parents, egg size tends to increase in gregarious forms, whereas it tends to decrease slightly in solitary forms.

A negative correlation between progeny size and female size is rare (Fox & Czesak, 2000). It is possible that under non-competitive conditions at low population density, selective pressure has favoured solitary females to increase the number of eggs at the expense of individual egg size. The production of smaller eggs by larger females in isolation-reared locusts might be an adaptive response because the environment in which large adults occur is likely to be more favorable for nymphal growth compared with an environment where small adults occur. This would effectively lead to a reduction in the amount of investment to each egg without lowering hatchling survival and a greater investment in egg production. At high population density, on the other hand, large body size in hatchlings is likely to impart increased fitness, as mentioned above. Because of such differences in selective pressure between the two phases, a trade-off between egg size and number, which is clearly shown when data for the two phases are combined, may become less obvious within the solitary forms and non-significant within the gregarious forms.

In conclusion, locusts reared in isolation or in groups with minimal stress from competition for food and egg-laying space exhibit phase-dependent and body size-dependent differences in various developmental and reproductive characteristics. Unlike the solitary forms in which development and reproduction are constrained by a trade-off, the gregarious forms have acquired capacities to grow faster without reducing the final body size and to produce more and larger eggs as the body size of the female parent increases. The latter is achieved by increasing the egg-developing capacity relative to body size. Crowding seems to serve as a stimulating signal for locusts to express a set of gregarious characteristics that contribute to rapid population growth during outbreaks.

Acknowledgements

The authors thank Ms Hiroko Ikeda, Ms Chieko Ito and Ms Masako Higuchi for laboratory assistance and Dr Toyomi Kotaki for helpful suggestions at NIAS. K.M. is grateful to Prof. Makio Takeda (Kobe University) for kind advice and encouragement. This study was partly supported by a JSPS Research Fellowship for young Scientists to K.M. and Kakenhi Funds of Japan to S.T. The authors wish to thank two anonymous reviewers who helped to improve the manuscript.

References

- Albrecht, O.F. & Blackith, R.E. (1960) Poids et délai de survie des larves nouveau-nées chez les acridiens migrants. Données physiologiques. *Comptes Rendus Academy Science, Paris* **250**, 3388–3390.
- Applebaum, S.W. & Heifetz, Y. (1999) Density-dependent physiological phase in insects. *Annual Review of Entomology* **44**, 317–341.
- Chauvin, M.R. (1941) Sur le grégarisme du criquet pèlerin (*Schistocerca gregaria* Forsk.). *Comptes Rendus Academy Science, Paris* **212**, 175–177.
- Faure, J.C. (1932) The phases of locusts in South Africa. *Bulletin of Entomological Research* **23**, 293–405.
- Ferenz, H.-J. & Seidelmann, K. (2003) Pheromones in relation to aggregation and reproduction in desert locusts. *Physiological Entomology* **28**, 11–18.
- Fox, C.W. & Czesak, M.E. (2000) Evolutionary ecology of progeny size in arthropods. *Annual Review of Entomology* **45**, 341–369.
- Hägele, B.F., Oag, V., Bouaïchi, A., McCaffery, A.R. & Simpson, S.J. (2000) The role of female accessory glands in maternal inheritance of phase in the desert locust *Schistocerca gregaria*. *Journal of Insect Physiology* **46**, 275–280.
- Hassanali, A., Njagi, P.G.N. & Bashir, M.O. (2005) Chemical ecology of locusts and related acridids. *Annual Review of Entomology* **50**, 223–245.
- Heifetz, Y. & Applebaum, S.W. (1995) Density-dependent physiological phase in a non-migratory grasshopper *Aiolopus thalassinus*. *Entomologia Experimentalis et Applicata* **77**, 251–262.
- Huis, A.V., Cressman, K. & Magor, J.I. (2007) Preventing desert locust plagues: optimizing management interventions. *Entomologia Experimentalis et Applicata* **122**, 191–214.
- Hunter-Jones, P. (1958) Laboratory studies on the inheritance of phase characters in locusts. *Anti-Locust Bulletin* **29**, 1–32.
- Kennedy, J.S. (1956) Phase transformation in locust biology. *Biological Reviews* **31**, 349–370.
- Lecoq, M. (2005) Desert Locust management: from ecology to anthropology. *Journal of Orthoptera Research* **14**, 179–186.
- Maeno, K. & Tanaka, S. (2007) Effects of hatchling body colour and rearing density on body colouration in last stadium nymphs of the desert locust, *Schistocerca gregaria* (Forskål) (Orthoptera: Acrididae). *Physiological Entomology* **32**, 87–94.
- Maeno, K. & Tanaka, S. (2008) Maternal effects on progeny size, number and body color in the desert locust, *Schistocerca gregaria*: density- and reproductive cycle-dependent variation. *Journal of Insect Physiology*, in press (doi:10.1016/j.jinsphys.2008.04.010).
- Maeno, K., Gotoh, T. & Tanaka, S. (2004) Phase-related morphological changes induced by [His⁷]-corazonin in two species of locusts, *Schistocerca gregaria* and *Locusta migratoria* (Orthoptera: Acrididae). *Bulletin of Entomological Research* **94**, 349–357.
- McCaffery, A.R., Simpson, S.J., Islam, M.S. & Roessingh, P. (1998) A gregarizing factor present in the egg pod foam of the desert locust *Schistocerca gregaria*. *Journal of Experimental Biology* **201**, 347–363.
- Pener, M.P. (1991) Locust phase polymorphism and its endocrine relations. *Advances in Insect Physiology* **23**, 1–79.
- Pener, M.P. & Yerushalmi, Y. (1998) The physiology of locust phase polymorphism: an update. *Journal of Insect Physiology* **44**, 365–377.
- Rao, Y.R. & Gupta, R.L. (1939) Some notes on eye-stripes in Acrididae. *Indian Journal of Agricultural Sciences* **9**, 727–729.
- Seidelmann, K. & Ferenz, H.J. (2002) Courtship-inhibition pheromone in desert locusts, *Schistocerca gregaria*. *Journal of Insect Physiology* **48**, 991–996.
- Simpson, S.J. & Miller, G.A. (2007) Maternal effects on phase characteristics in the desert locust, *Schistocerca gregaria*: A review of current understanding. *Journal of Insect Physiology*, **53**, 869–876.
- Simpson, S.J., McCaffery, A.R. & Hägele, B.F. (1999) A behavioural analysis of phase change in the desert locust.

- Biological Reviews of the Cambridge Philosophical Society* **74**, 461–480.
- Staal, G.B.** (1961) *Studies on the Physiology of Phase Induction in Locusta migratoria migratorioides R. & F.* 125 pp. Wageningen, The Netherlands, H. Veenman & Zonen N.V.
- Stearns, S.C.** (1992) *The Evolution of Life Histories*. 249 pp. Oxford, Oxford University Press.
- Tanaka, S. & Maeno, K.** (2006) Phase-related body-color polyphenism in hatchlings of the desert locust, *Schistocerca gregaria*: re-examination of the maternal and crowding effects. *Journal of Insect Physiology* **52**, 1054–1061.
- Tanaka, S. & Maeno, K.** (2008) Maternal effects on progeny body size and color in the desert locust, *Schistocerca gregaria*: Examination of a current view. *Journal of Insect Physiology* **54**, 612–618.
- Tanaka, S. & Yagi, S.** (1997) Evidence for the involvement of a neuropeptide in the control of body color in the desert locust, *Schistocerca gregaria*. *Japanese Journal of Entomology* **65**, 447–457.
- Tanaka, S. & Zhu, D.-H.** (2003) Phase-related differences in mating strategy of a locust. *Annals of Entomological Society of America* **96**, 498–502.
- Uvarov, P.** (1966) *Grasshoppers and Locusts, Vol. 1.* 481 pp. Cambridge, Cambridge University Press.
- Uvarov, P.** (1977) *Grasshoppers and Locusts, Vol. 2.* 613 pp. London, Centre for Overseas Pest Research.