

Habitat preference and seasonal variability of epifaunal assemblages associated with macroalgal beds on the Central Red Sea coast, Saudi Arabia

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Macroalgal communities of coastal ecosystems play a key role in maintaining overall coastal biodiversity. In this study, habitat preference (macroalgal host preference) and temporal changes (season) of epifaunal communities associated to five macroalgal species such as Padina, Sargassum, Ulva, Acanthophora and Gracilaria were observed at seasonal intervals (summer and winter) for 2 years from May 2012 to March 2014 on the Jeddah coastal waters of the Red Sea. Major epifaunal groups observed included polychaetes, amphipods, isopods, gastropods, mussels and crabs. Polychaetes were represented by 10 species followed by amphipods with five species. The abundance of gastropods, polychaetes and mussels showed significant variation between the macroalgae irrespective of morphology. Generally, the abundance of these taxa were high on Padina and low on Sargassum. Significant seasonal changes were observed on the abundance of polychaetes, amphipods and isopods. The abundance of some epifaunal groups showed a significant relationship with environmental parameters such as surface water temperature, salinity, pH and dissolved oxygen content.

Keywords: Coastal biodiversity, epifauna, epibiosis, seaweeds, biogenic habitats, Red Sea, marine invertebrates

Submitted 16 February 2015; accepted 15 September 2015; first published online 15 October 2015

INTRODUCTION

Some marine organisms can act as biogenic habitats (Roberts *et al.*, 2008) in the coastal ecosystems and act as refugia for number of associated organisms. Biogenic habitats or habitat-forming organisms perform various functions and have a major effect on the structure of shallow water communities (Stachowicz, 2001; Cacabelos *et al.*, 2010). For example, marine macroalgae play an important role in coastal ecosystems by providing shelter and food for many organisms, increasing the space for settlement (Wikström & Kautsky, 2004; Cacabelos *et al.*, 2010), and protecting the organisms from wave action, heat and desiccation (Moore, 1978; Hicks, 1980). Marine invertebrates such as polychaetes, amphipods, isopods, gastropods and mussels are frequently associated with macroalgal communities (Christie *et al.*, 1998; Cunha *et al.*, 2013). These epifaunal communities may be an important food source for juvenile fishes, which are also abundant in macroalgal beds (Bray & Ebeling, 1975; Jones, 1988).

The epibenthic communities associated with macroalgae show changes in abundance and composition in relation to temporal and spatial scales due to various physical and biological factors (Caine, 1991; Taylor, 1998). Changes in environmental factors, competition, predation and recruitment are the

main factors that influence the abundance of epifauna (Jones & Thornber, 2010; Cacabelos *et al.*, 2010). Variations in physical conditions such as shore heights play a significant role in structuring the epifaunal abundance on macroalgae (Cacabelos *et al.*, 2010). Several previous studies reported the variability of epifaunal abundance and diversity in relation to macroalgal species (Taylor & Cole, 1994; Vazquez-Luis *et al.*, 2008). For example, Cacabelos *et al.* (2010) reported that the identity of host algae was an important factor in structuring the epifaunal assemblages. The differences in abundance of epifauna between different macroalgal species inhabiting the same area may be due to factors such as longevity, cell-wall structure, presence of algal epiphytes, production of secondary metabolites and structure of the host plant (Jacobi & Langevin, 1996; Steinberg *et al.*, 1998; Viejo, 1999; Le Lann *et al.*, 2008; Gestoso *et al.*, 2010). As most of the epifaunal species depend on the algae for their food, the cell-wall structure of the macroalgae (internal properties) may contribute much to the distribution patterns of epifauna (Cacabelos *et al.*, 2010). The internal properties vary across macroalgae resulting in differences in food value and palatability (Hawkins & Hartnoll, 1983; Bates, 2009). In addition, the defence mechanism of the macroalgae will also influence the epifaunal abundance. Besides internal properties, the external morphology and epiphytic load of the macroalgae may also play key roles in controlling epifaunal assemblages. The complex and highly branched macroalgae are reported to provide more space than simple forms (Duffy & Hay, 1991). The role of epiphytic load on the distribution of epifauna depends on strategies of the faunal groups to

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exploit the food sources. This was evidenced from the study by Bologna & Heck (1999), which highlighted the trophic role of epiphytes.

As coastal ecosystems are under the strong impact of pollution and climate change-related stressors (Duarte, 2002), there is a need to assess the changes in diversity and distribution of marine organisms associated with macroalgae and seagrasses. Knowledge about the diversity and distribution patterns of benthic populations in a particular region is also needed for valid estimates of population shifts due to human activities (Maurer *et al.*, 1979). The diversity and community dynamics of epifauna associated with macroalgae have been studied in coastal ecosystems around the world (Aoki, 1988; Taylor & Cole, 1994; Christie *et al.*, 2003; Fredriksen *et al.*, 2005; Sathesh & Wesley, 2006; Cacabelos *et al.*, 2010; Jones & Thornber, 2010; Cunha *et al.*, 2013). The main objective of the present study was to determine the abundance of invertebrates associated with common macroalgal species inhabiting the coastal ecosystem of the Central Red Sea region in relation to sampling season and macroalgal species. In particular, we posed the following questions: Do the dominant epifaunal communities show any preference to macroalgal host? Do the abundance of the epifaunal communities vary between different seasons in the Saudi Arabian Red Sea coast? The observed results may improve our knowledge on the factors structuring the epibenthic invertebrates associated with macroalgae on coastal ecosystems.

MATERIALS AND METHODS

Sampling of epifauna

The study was conducted at Jeddah coast (21°19'09"N 39°06'07"E) of the Central Red Sea region of Saudi Arabia. Seasons at Central Red Sea region can be classified into autumn (mid September–mid December), winter (mid December–mid March), spring (mid March–mid May) and summer (mid May–mid September). Five macroalgae abundantly available in the Jeddah coastal waters, *Sargassum*, *Gracilaria*, *Padina*, *Acanthophora* and *Ulva*, were selected for this study. The study focused on the same macroalgal species (based on morphological observations) in each genus throughout the duration. The sampling was conducted in winter and summer seasons for a period of 2 years covering from May 2012 to March 2014. Three samples were collected in each season (from three different stations within a 50 km radius) and the mean \pm SD values for those samples ($N = 3$) were considered. Epifauna were collected by detaching a small portion of each macroalgal front from the macroalgae bed by placing a polythene bag over it. During the sampling maximum care has been taken to avoid disturbance to the macroalgae in the natural habitat. The collected macroalgal samples were kept in polythene bags with fresh seawater and transported to the laboratory. In the laboratory, the algal samples were washed in a trough filled with seawater and drops of formalin to remove the epifauna (Taylor & Cole, 1994). The water from the trough was sieved through a 0.5 mm mesh to recover the marine invertebrates (Gestoso *et al.*, 2010). From the filtrate, the invertebrates were separated and preserved in absolute ethanol for further analysis. After epifaunal removal, each macroalgal sample was wiped using a tissue paper to remove the water on the surface and then

weighed to determine the wet weight. Epifaunal invertebrates were separated into lowest possible taxon wherever possible using standard manuals of marine invertebrates. The epifaunal assemblages were subjected to analyses including total abundance and number of taxa on each macroalga species. The abundance of invertebrate taxa on each macroalga species were counted manually under a stereomicroscope and presented as number of individuals per 100 g of macroalgal wet weight (100 g AW⁻¹).

Environmental parameters

Environmental parameters including surface water temperature, pH, salinity, dissolved oxygen content and concentration of nitrite, nitrate and phosphate were measured in the coastal waters during the sampling of epifauna usually between 9 and 10 a.m. Water temperature, salinity, pH and dissolved oxygen content were measured *in situ*. The salinity was measured using a refractometer and the other parameters were measured with the help of a portable dissolved oxygen probe (Hanna) and pH meter (Hanna). For the estimation of nutrients, water samples were collected in plastic bottles, kept in a coolbox and brought to the laboratory. In the laboratory, the samples were filtered and analysed for phosphate, nitrate and nitrite. All these parameters were analysed according to the standard methods compiled by Venugopalan & Paulpandian (1989).

Data analyses

Univariate diversity indices such as Shannon–Weiner (H'), Pielou's evenness (J') and species richness (Margalef, d) were calculated using PRIMER 6. The abundance pattern of polychaetes, amphipods, isopods, gastropods and bivalves on macroalgae was expressed by cluster analysis (single linkage) based on Bray–Curtis similarity index using BioDiversity Pro (McAleece *et al.*, 1997). Changes in the abundance of epifauna associated with macroalgae were analysed using a three-way ANOVA with year, season and macroalgal type as factors (STATISTICA software). The diversity indices were subjected to two-way ANOVA with season and macroalgal type as factors (Year was found to be not significant in a three-way ANOVA of abundance data and hence not included in this model.) Simple correlation coefficient was used to find out the relationship between environmental factors and epifaunal abundance by pooling the density of epifauna from all the macroalgae for each season.

RESULTS

The environmental parameters observed from the coastal waters during the study period are given Table 1. A total of 25 invertebrate species were commonly observed during the study period. Among these, polychaetes were the dominant group with 10 species, followed by amphipods with five species. Other important groups observed include gastropods (three species), isopods (three species), crabs (two species) and bivalves (two species) (Table 2). Abundance of epifaunal groups such as polychaetes, gastropods and bivalves showed a significant ($P < 0.05$) difference between macroalgae (Table 3). The total number of organisms associated with the five macroalgal genera during summer and winter seasons is presented in Figure 1. The density of epifauna

Table 1. Surface water environmental parameters of Jeddah coastal waters during the study period (mean \pm SD).

	Summer 2012	Winter 2012–2013	Summer 2013	Winter 2013–2014
Salinity	42 \pm 0.80	39 \pm 1.30	40 \pm 2.0	39 \pm 0.9
pH	8.4 \pm 0.03	8.1 \pm 0.05	8.3 \pm 0.02	8.2 \pm 0.07
Temperature ($^{\circ}$ C)	37.5 \pm 0.20	36 \pm 1.03	37.8 \pm 0.19	35.4 \pm 0.45
Dissolved oxygen content (mg l ⁻¹)	5.9 \pm 0.80	5.0 \pm 0.42	5.82 \pm 0.35	5.3 \pm 0.32
Nitrite (μ g l ⁻¹)	0.39 \pm 0.07	0.51 \pm 0.02	0.50 \pm 0.07	0.12 \pm 0.03
Nitrate (μ g l ⁻¹)	0.52 \pm 0.15	0.63 \pm 0.05	0.96 \pm 0.21	1.02 \pm 0.17
Phosphate (μ g l ⁻¹)	0.8 \pm 0.12	0.59 \pm 0.09	1.31 \pm 0.08	0.89 \pm 0.11

associated to *Padina* varied from 40 to 247 individuals 100 g AW⁻¹. Higher abundance was observed during summer 2013 and lower in winter 2013–14. On *Sargassum*, a total of 177 organisms were observed during summer 2013 and 39 individuals 100 g AW⁻¹ in winter 2012–13. The total number of epifaunal communities of *Ulva* varied between 88 and 32 individuals 100 g AW⁻¹. The abundance was low during winter 2012–13 and high in summer 2012. The number of organisms found associated with *Acanthophora* was 103 during summer 2013 and 64

individuals 100 g AW⁻¹ in winter 2013–14. On *Gracilaria*, 172 organisms 100 g AW⁻¹ were observed during summer 2013 and 27 individuals 100 g AW⁻¹ in winter 2013–14. In general, the total abundance of epibenthic communities on macroalgae was low during winter. The observed seasonal trend in total epifaunal abundance was mainly due to the variations exhibited by the polychaetes and amphipods. This was evidenced from the observed significant seasonal variation ($P < 0.05$) in the abundance of amphipods and isopods (three-way ANOVA, Table 3).

Table 2. Invertebrate taxa associated with seaweeds on the Jeddah coast of the central Red Sea (+ indicates the presence on a particular seaweed).

Epifauna	Macroalgae				
	<i>Padina</i>	<i>Sargassum</i>	<i>Ulva</i>	<i>Acanthophora</i>	<i>Gracilaria</i>
Polychaeta					
Family: Nereididae					
<i>Perinereis</i> sp.	+	+	+		+
<i>Platynereis dumerilli</i>	+		+		+
<i>Pseudonereis</i>	+		+	+	+
Nereidid 1	+				+
Nereidid 2		+			
Family: Syllidae					
<i>Syllis variegata</i>	+	+	+	+	+
<i>Haplosyllis spongicola</i>	+		+		+
Syllid sp1			+		+
Syllid sp2	+				
Family: Terebellidae					
<i>Terebella</i> sp.					+
Amphipoda					
Family: Caprellidae					
Family: Ampithoidae					
<i>Peramphithoe</i> sp.	+	+	+	+	+
Family: Lysianassidae					
<i>Orchomenella franklini</i>	+	+	+	+	+
Family: Gammaridae					
<i>Gammarus</i> sp.	+	+	+	+	+
Family: Corophiidae					
<i>Corophium</i> sp.	+		+	+	+
Isopoda					
Family: Anthuridae					
<i>Mesanthura</i> sp.	+	+	+	+	+
Family: Sphaeromatidae					
<i>Cymodoce</i> sp 1	+	+	+	+	+
<i>Cymodoce</i> sp 2	+	+	+		+
Decapoda					
Family: Majidae					
Family: Xanthidae					
Bivalvia					
Family: Mytilidae					
<i>Perna</i> sp.	+	+	+	+	+
<i>Modiolus</i> sp.					+

Table 3. Three-way ANOVA (analysis of variance) of abundance of epifauna on different seaweeds. Year (2012–13 and 2013–14), season (summer and winter) and seaweeds (*Padina*, *Sargassum*, *Ulva*, *Acanthophora* and *Gracilaria*) were selected as factors.

	df	F	P	df	F	P
	Polychaetes			Amphipods		
Year	1	1.132	0.306	1	1.184	0.296
Season	1	2.716	0.019*	1	19.469	0.0007**
Seaweed	4	3.260	0.017*	4	0.948	0.467
Error	13			13		
Total	20			20		
	Isopods			Gastropods		
Year	1	0.551	0.471	1	0.161	0.694
Season	1	6.160	0.027*	1	2.716	0.123
Seaweed	4	1.521	0.253	4	3.260	0.046*
Error	13			13		
Total	20			20		
	Bivalves			Crabs		
Year	1	0.0601	0.81	1	0.475	0.502
Season	1	1.7304	0.211	1	1.939	0.187
Seaweed	4	6.1739	0.005*	4	1.337	0.308
Error	13			13		
Total	20			20		

* $P < 0.5$, ** $P < 0.001$.

Shannon–Weiner diversity index (H') of the invertebrate fauna varied between 2.03 and 2.31 on *Padina*, 1.72 and 1.88 on *Sargassum*, 1.56 and 2.42 on *Ulva*, 1.88 and 2.28 on *Acanthophora*, and from 1.76 to 2.32 on *Gracilaria* (Figure 2). Evenness index (J') of the epifauna associated with macroalgae varied between 0.6 and 0.89 (Figure 3). The Margalef's species richness index is given in Figure 4. The species richness (d) of the invertebrates associated with macroalgae was high on *Gracilaria* (3.68) and low on *Sargassum* (2). Two-way ANOVA showed significant seasonal and macroalgal variation in species richness index (Table 4). Shannon–Weiner diversity index and evenness index did not show significant variation in relation to season and macroalgae.

Polychaete communities associated with the macroalgae mainly belonged to the families of Nereididae and Syllidae.

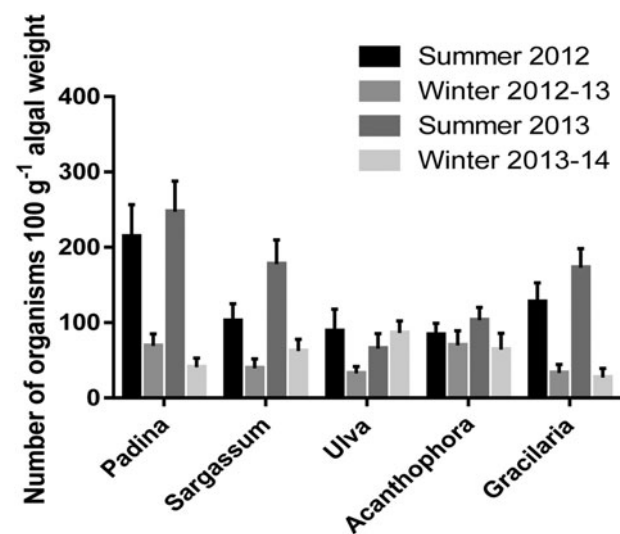


Fig. 1. Total number of organisms associated with different seaweeds inhabiting on the Jeddah coast. Error bars indicate standard deviation ($N = 3$).

A total of 10 species were observed from the macroalgae and *Syllis variegata*, *Perinereis cultifera* and *Platynereis dumerilii* were the abundant species. The abundance patterns of *Syllis variegata* and *Perinereis cultifera* are presented in Figure 6. Analysis of macroalgae showed that seven species of polychaetes were observed from *Padina* and their maximum density (15 ind. 100 g AW^{-1}) was observed during summer 2012 (Figure 5). The polychaete fauna associated with *Sargassum* consists of three species and the abundance was high during summer 2013–14 (five ind. 100 g AW^{-1}). The polychaete community associated to *Ulva* consisted of five species and the density was high during summer 2012 (15 ind. 100 g AW^{-1}). Three polychaete species (one syllid and two nereidids) were observed from *Acanthophora* and a maximum of seven individuals 100 g AW^{-1} was observed during winter 2012–13. The polychaete community associated with *Gracilaria* consists of nine species and the abundance was high in summer 2012 (17 ind. 100 g AW^{-1}). The abundance of polychaetes showed significant variation between macroalgae and seasons (Table 3).

Amphipods observed in this study include Caprellids, *Gammarus* sp., *Corophium* sp., *Peramphithoe* sp., and *Orchomenella franklini* (Table 2). The abundance of amphipods on *Padina* was high (191 ind. 100 g AW^{-1}) during summer 2013 and low (12 ind. 100 g AW^{-1}) in winter 2013–14 (Figure 5). On *Sargassum*, amphipods abundance was also high during summer 2013 (140 ind. 100 g AW^{-1}) and low during winter 2012–13 (22 ind. 100 g AW^{-1}). The total number of amphipods associated with *Ulva* varied between 22 (winter 2012–13) and 78 (summer 2013) ind. 100 g AW^{-1} . The amphipods associated with *Acanthophora* showed a maximum density of 75 ind. 100 g AW^{-1} during summer 2013 and minimum of 29 ind. 100 g AW^{-1} in winter 2013–14. Amphipod abundance on *Gracilaria* varied between 11 (winter 2013–14) and 140 (summer 2013) ind. 100 g AW^{-1} . *Peramphithoe* sp., and *Orchomenella franklini* were the dominant species in all the macroalgae (Figure 6). Three-way ANOVA showed a highly significant variation ($P < 0.001$) in the abundance of amphipods between summer

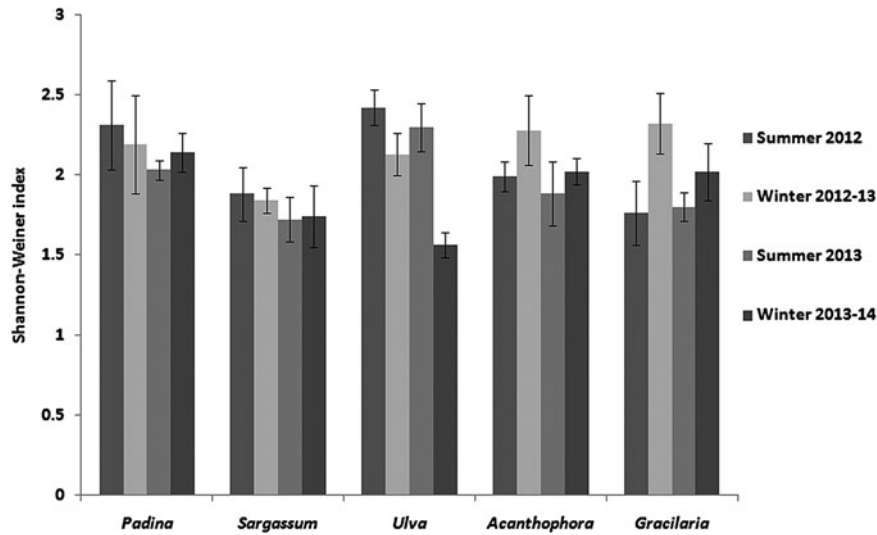


Fig. 2. Shannon–Weiner diversity index (H') of epifauna associated with seaweeds.

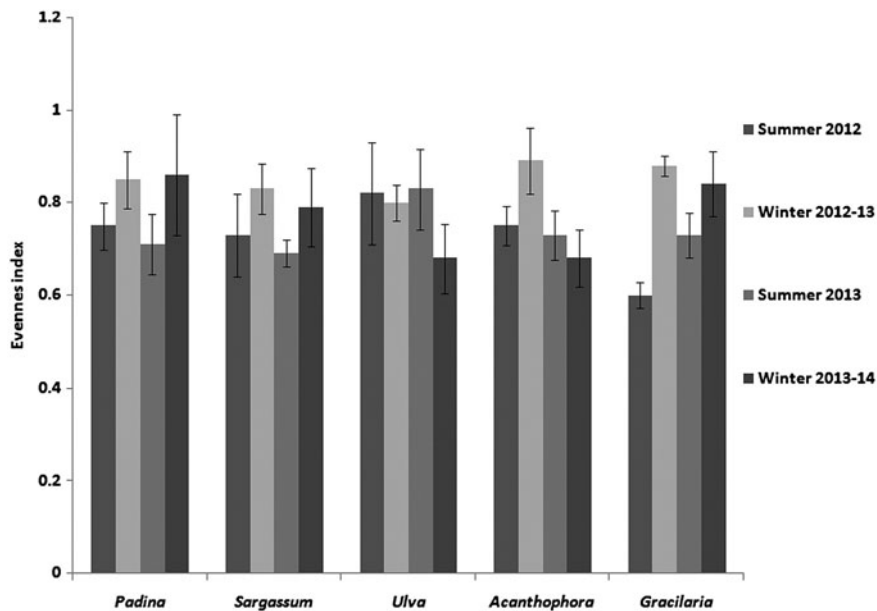


Fig. 3. The evenness index (Pielou's J') of the epifauna associated with seaweeds during the study period.

and winter seasons (Table 3). However, the abundance pattern between macroalgal species did not show significant variation.

Isopods such as *Mesanthura* sp. and *Cirolina* sp., were commonly associated with the macroalgae (Table 2). Density of isopods associated with *Padina* was high during summer 2012 (28 ind. 100 g AW⁻¹) and low in winter 2013–14 (three ind. 100 g AW⁻¹). The abundance of isopods on *Sargassum* varied between nine (winter 2012–13) and 27 (summer 2013) ind. 100 g AW⁻¹. Abundance of isopods on *Ulva* varied between 1.6 (winter 2013–14) and 8 (summer 2013) ind. 100 g AW⁻¹. On *Acanthophora*, a higher density of 15 isopods 100 g AW⁻¹ was observed during summer 2013 and a lower density of eight ind. 100 g AW⁻¹ in summer 2012. Isopods abundance on *Gracilaria* was low (three ind. 100 g AW⁻¹) during winter 2012–13 and high (13 ind. 100 g AW⁻¹) in summer 2013

(Figure 5). Three-way ANOVA revealed a significant seasonal variation in the abundance of isopods associated with macroalgae but did not show variation between macroalgae (Table 3).

Another important group associated with macroalgae was the gastropods represented by 3 species (two periwinkles, *Littorina* sp. and *Thais* sp.). The density of gastropods associated with *Padina* varied between two and 20 ind. 100 g AW⁻¹ (Figure 5). Gastropod abundance on other macroalgal species was very low with a maximum of six ind. 100 g AW⁻¹ on *Ulva* and *Acanthophora*. Gastropod density on *Sargassum* and *Gracilaria* showed a maximum of four and two individuals 100 g AW⁻¹ respectively. Three-way ANOVA showed a significant variation on the density of gastropods between macroalgal species but the seasonal variation was not significant (Table 3).

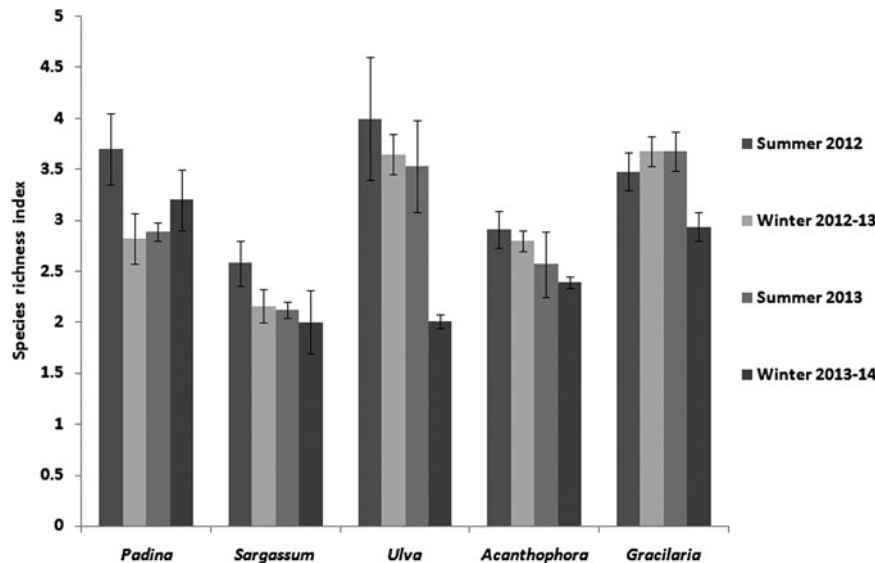


Fig. 4. The Margalef's species richness (d) of the invertebrates associated with seaweeds.

Table 4. Two-way ANOVA (analysis of variance) of Shannon–Weiner diversity index (H'), evenness index (J') and species richness index (d) of epifauna associated with seaweeds. Season (summer and winter) and seaweeds (*Padina*, *Sargassum*, *Ulva*, *Acanthophora* and *Gracilaria*) were used as factors.

Source of variation	df	F	P
Shannon–Weiner index			
Seaweed	4	1.7113	0.211
Season	3	1.4297	0.282
Error	12		
Total	19		
Evenness index			
Seaweed	4	0.1485	0.96
Season	3	2.6087	0.09
Error	12		
Total	19		
Species richness index			
Seaweed	4	6.8738	0.004*
Season	3	3.8827	0.037*
Error	12		
Total	19		

* $P < 0.05$.

Two species of mussels (bivalves) were observed from the macroalgae and they are abundantly noticed on *Padina* and *Acanthophora* (Figure 5). A maximum of 19 ind. 100 g AW⁻¹ were observed on *Padina* during winter 2012–13. On *Acanthophora*, the density of bivalves varied between 0.6 (summer 2012) and 17 (winter 2013–14) ind. 100 g AW⁻¹. Mussel density on *Ulva* showed a maximum of four ind. 100 g AW⁻¹ and a minimum of one ind. 100 g AW⁻¹. The other two macroalgae, *Sargassum* and *Gracilaria* harboured only few mussel individuals during the study period. Three-way ANOVA showed that the density mussels varied significantly between macroalgae (Table 3). Crabs mainly belonging to the families Xanthidae and Majidae were observed from the macroalgae occasionally (Figure 5).

Correlation analysis showed significant positive relationship between surface water temperature and density of

gastropods, amphipods and isopods (Table 5). The abundance of polychaetes showed a significant positive correlation with salinity and pH. Correlation analysis also revealed that the abundance of bivalves was negatively correlated with salinity, pH and dissolved oxygen content (Table 5). Cluster analysis dendrograms based on Bray–Curtis similarity index for polychaetes, amphipods, isopods, gastropods and bivalves are given in Figure 7A–E. The cluster analysis plots showed that the epifaunal community structure was influenced by the macroalgal species. For polychaetes, the abundance on macroalgae showed two distinct groups (*Ulva*, *Gracilaria* and *Padina* formed one group and the other group consisted of *Sargassum* and *Acanthophora*). The abundance of amphipods and isopods on *Ulva* differed from other macroalgal species (Figure 7B, C). Cluster analysis of gastropod density on *Padina* clearly varied from other macroalgal species (Figure 7D). Abundance of bivalves showed two groups in cluster analysis with *Padina* and *Acanthophora* forming one group and the remaining macroalgal species were in the other group (Figure 7E).

DISCUSSION

Understanding the ecology, general distribution and diversity of epifaunal community associated with macrophytes in the coastal ecosystems will help to predict the changes in benthic communities due to climate change-driven stressors and anthropogenic impacts. In the present study, the invertebrate communities associated with macroalgae were dominated by polychaetes followed by amphipods, isopods and gastropods. Polychaetes associated with macroalgae consisted of 10 species and were dominated by the families Nereididae and Syllidae throughout the study period. Most of the taxa were identified up to genus level or family level. Previous studies on faunal communities associated with macroalgae were mainly focused on one algal species or distribution of particular epifaunal group on different macroalgae. In this study, invertebrate taxa associated with five macroalgal species were analysed in two different seasons.

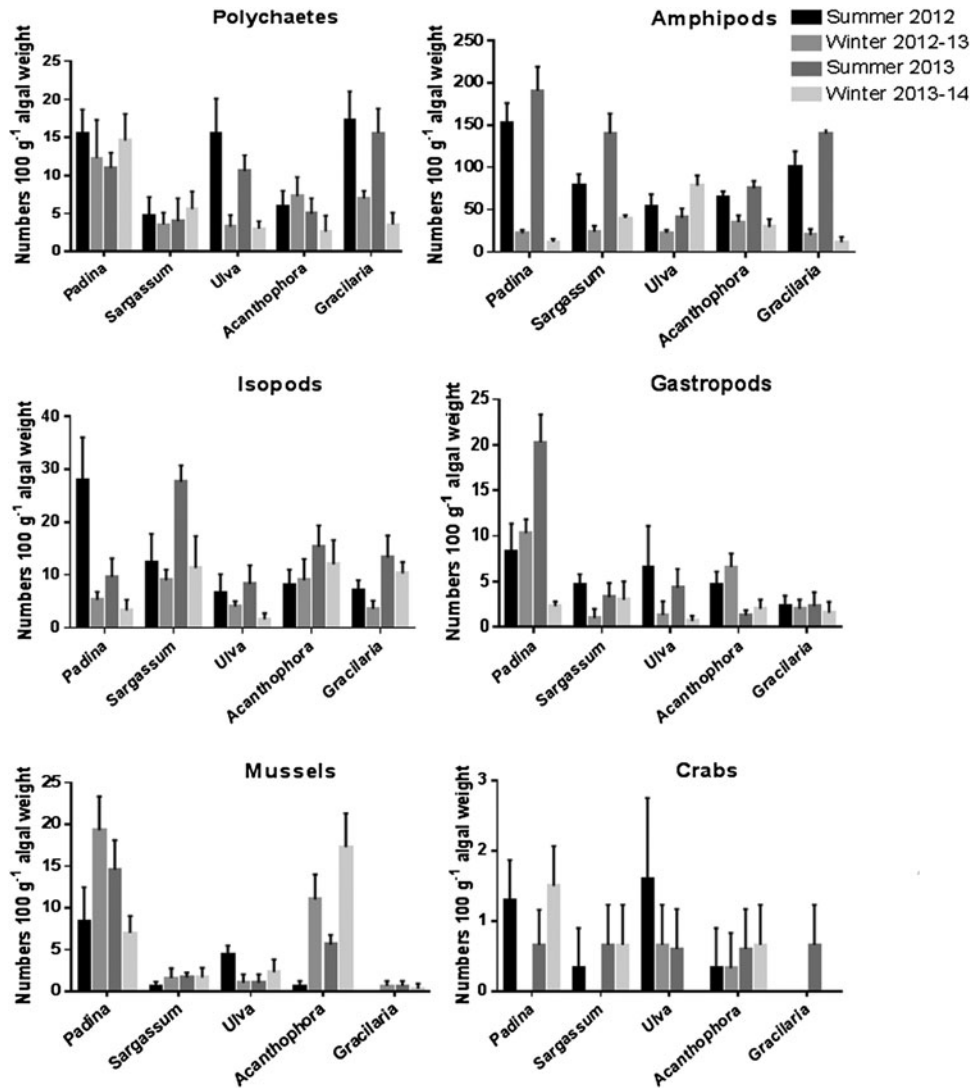


Fig. 5. Abundance of invertebrate groups studied, i.e. polychaetes, amphipods, isopods, gastropods, mussels and crabs on different seaweeds during summer and winter seasons. Error bars extend to standard deviation ($n = 3$).

Polychaetes are important groups which play a key role in macrobenthic secondary production in coastal ecosystems (Paiva, 1993) and form an important part in the food chain. Other taxa such as gastropods and amphipods are often reported as dominant groups on phytal assemblages in coastal waters (Tararam & Wakabara, 1981). Polychaetes, amphipods and isopods showed significant seasonal variation in their densities on macroalgae. Many epifaunal taxa, particularly amphipods and isopods, are considered as demersal zooplankton; they spend part of the time in the marine waters normally during the night and resettle on various substrata before the daybreak (Hobson & Chess, 1976). Hence, the density of these epifaunal communities may depend on their resettling capability on the macroalgae. While seasonal fluctuations in environmental factors are often cited as important factors controlling the abundance of benthic communities in coastal ecosystems, the Red Sea is a distinctive region in the tropics due to its partial isolation from the open ocean (Shaikh *et al.*, 1986), high evaporation rate and lower precipitation. The environmental parameters such as surface water temperature,

salinity, pH and dissolved oxygen content observed during this study indicate small differences between summer and winter seasons. The observed values were within the range reported for Red Sea by the previous researchers. For example, water temperature in Red Sea ranged between 25.5 and 31°C but higher values were observed in shallow and coastal areas (Edwards & Head, 1987) and mean salinity may exceed 40 psu (Al-Farawati, 2010). The correlation analysis revealed that water temperature appears to influence the abundance of gastropods, amphipods and isopods. Also, the abundance of polychaetes showed a positive correlation with salinity while bivalves showed a negative relationship. Other important factors such as pH and dissolved oxygen content of the coastal waters also exert some influence on epifaunal abundance as shown from the correlation analysis. The observed significant relationship between abundance of epifauna and environmental parameters such as temperature, salinity, pH and dissolved oxygen content is of particular interest due to the fact that the Red Sea is considered to be fragile and vulnerable to global warming (Cantin *et al.*, 2010; Raitos *et al.*, 2011).

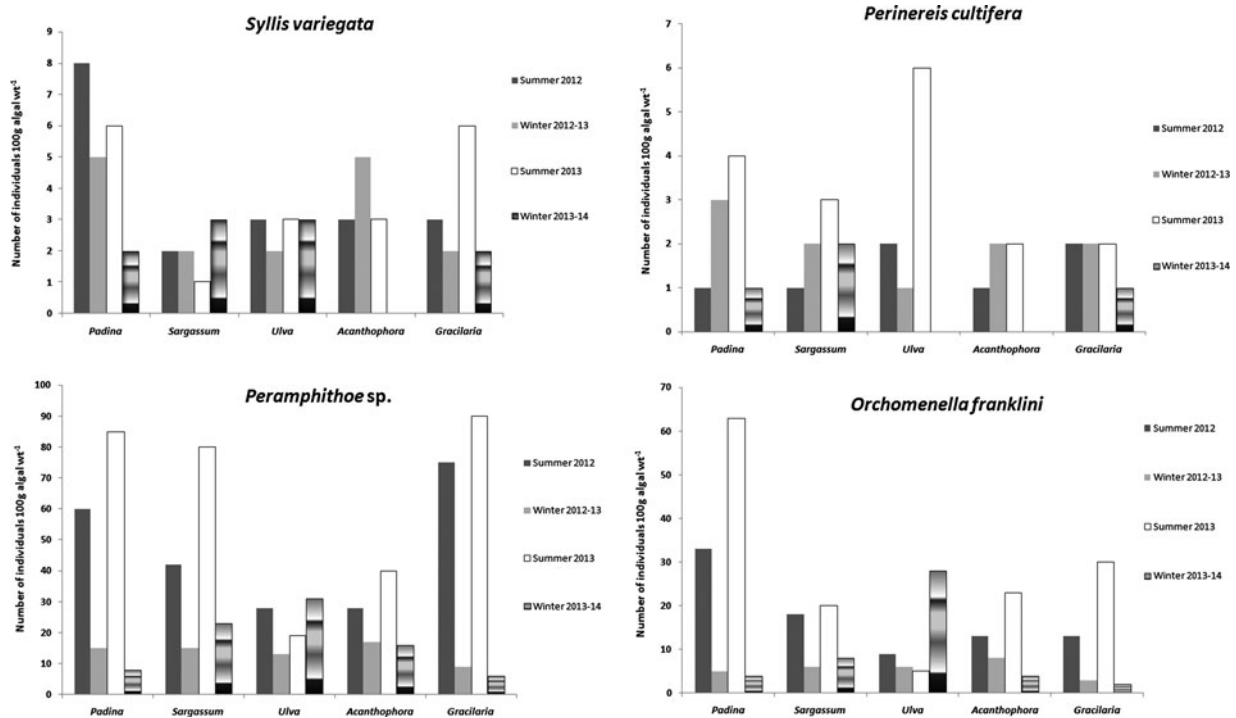


Fig. 6. Abundance pattern of dominant epifaunal invertebrate species *Syllis variegata* (Polychaete), *Perinereis cultifera* (Polychaete), *Peramphithoe* sp. (amphipod) and *Orchomenella franklini* (amphipod) associated with seaweeds.

Table 5. Correlation coefficient (r) between environmental parameters and abundance of epifauna on macroalgae. The density epifaunal groups on all the five macroalgal species were pooled for each season.

	Gastropods	Polychaetes	Amphipods	Bivalves	Isopods
Salinity	0.552	0.971*	0.643	-0.963*	0.636
pH	0.512	0.900*	0.811	-0.985*	0.825
Temp	0.938*	0.869	0.945*	-0.726	0.922*
DO	0.631	0.865	0.927*	-0.917*	0.938*
Nitrite	0.845	0.386	0.384	-0.023	0.320
Nitrate	-0.338	-0.550	0.046	0.354	0.092
Phosphate	0.443	0.227	0.787	-0.288	0.809

* $P < 0.05$.

The changes in diversity and abundance of benthic communities in relation to different spatial and temporal scales are also due to the result of ecological processes such as recruitment and disturbance (Thrush & Dayton, 2002). Recruitment will also depend on a number of factors such as reproduction, hydrological conditions, biotic factors, etc. It is important to note that the reproductive cycles of most of the taxa observed in this study are not known in Saudi Arabian Red Sea region. However, recent studies by Al-Aidaros & Satheesh (2014) reported the pronounced seasonality (active breeding in spring and summer) in reproductive cycle of barnacles in the central Red Sea region. We assume that other species may also follow a seasonal reproductive cycle and that this could be a possible reason for observed seasonal fluctuations. Previous studies on benthic and plankton communities of Red Sea also reported seasonal variations in abundance (El-Sherbiny *et al.*, 2007; Belal & Ghobashy, 2014).

Food availability and predation are other possible factors that may influence the epifaunal abundance. The seasonal

variation in species density and diversity observed in the present study was possibly due to the limitation of epifaunal populations by primary production of plant material (Edgar, 1991) and food source of epifauna. For example, those epifaunal organisms that graze directly upon the macroalgae are likely to be influenced by seasonal fluctuations in food quality. The epifaunal species observed in this study, such as the amphipod *Amphithoe* sp., and the polychaetes *Syllis* species and *Platynereis dumerilli*, were reported to graze the algae (Cunha *et al.*, 2013). Another possible reason will be seasonal predation pressure (Aoki, 1988) from predators especially crabs and fishes, which influences the epifauna population dynamics.

The present study showed that the total epifaunal abundance was higher on *Padina* than other macroalgae, mainly *Sargassum* and *Gracilaria*. The influence of macroalgae morphology on the abundance of epifaunal assemblages was previously observed by Stoner & Lewis (1985). Generally, shape and structural complexity of the macroalgae are the key factors that determine the abundance and community composition of the associated epifauna (Cacabelos *et al.*, 2010). Hence, macroalgae with more structural complexity may harbour more epifauna than those with low structural complexity as they provide a greater surface area (Taylor & Cole, 1994; Chemello & Milazzo, 2002). While no attempt has been made in this study to analyse the morphological characters of the macroalgae, the data reported in the literature were used for comparison. The macroalgal genera selected in this study may be classified (modified from Holmlund *et al.*, 1990) into highly branched forms (*Gracilaria* and *Acanthophora*), sheet-like forms (*Padina* and *Ulva*) and intermediate forms i.e. between highly branched and sheet like forms (*Sargassum*). The genus *Sargassum* is reported as structurally more complex in Fucales by various studies (Leite *et al.*, 2007). Holmlund *et al.* (1990) observed that amphipods were more susceptible to predation by fish on highly branched (more complex) algae

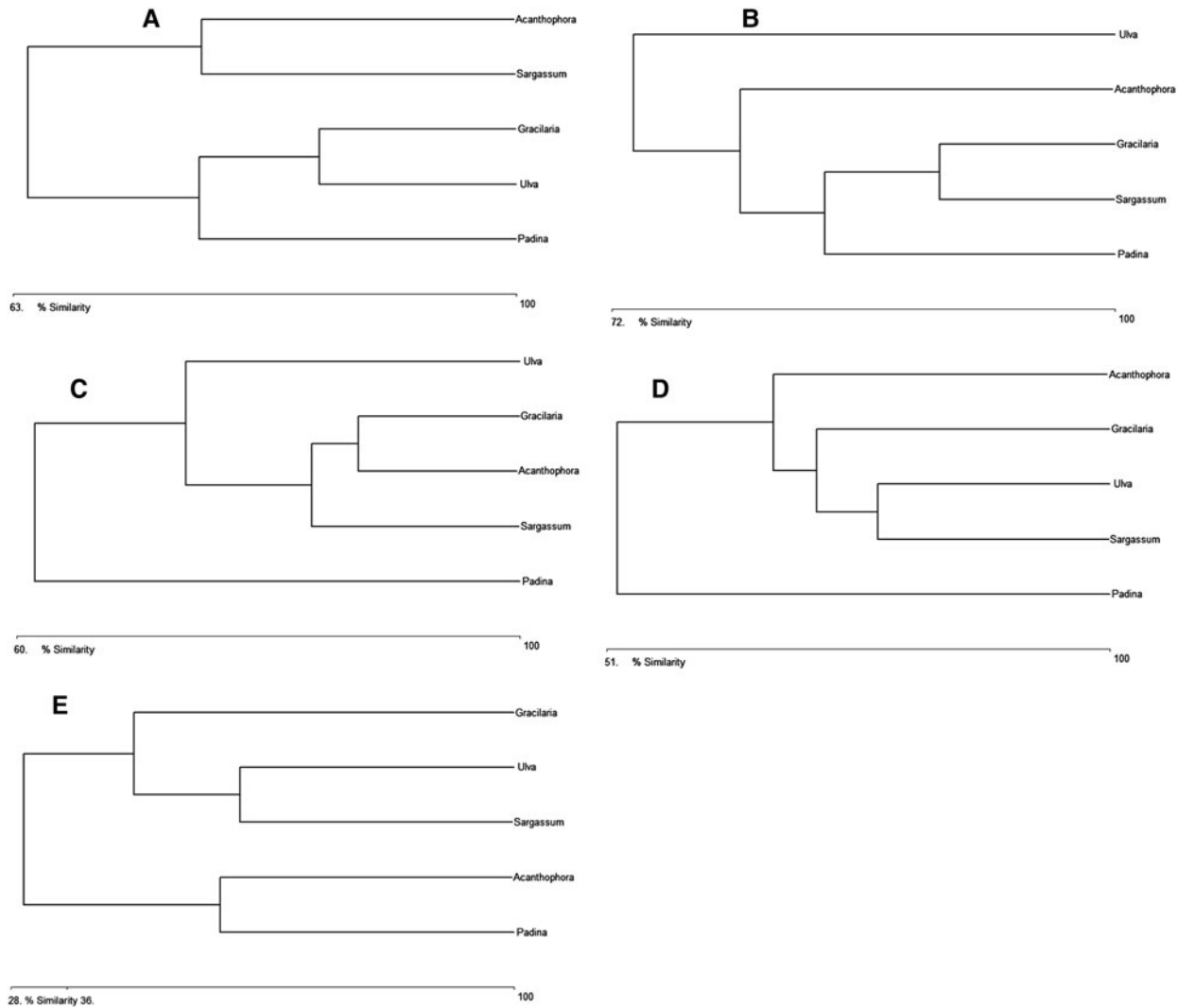


Fig. 7. Cluster analysis based on Bray–Curtis similarity index for polychaetes, amphipods, isopods, gastropods and bivalves associated with macroalgae (A. Polychaetes; B. Amphipods; C. Isopods; D. Gastropods; E. Bivalves).

than those associated with simple sheet-like morphology such as *Padina* and *Ulva*. Amphipod abundance patterns observed in this study showed significant seasonal variation rather than between macroalgal species. Abundance pattern of gastropods and bivalves which showed significant variation between different macroalgae was low on *Sargassum* and *Gracilaria*. The number of polychaetes which showed significant seasonal variation as well as difference between macroalgae was also low on *Sargassum*. While there was no observed clear pattern on abundance of epifaunal groups in relation to macroalgal morphology, *Sargassum*, which is also considered to be structurally complex, harboured low numbers of common epifaunal groups such as polychaetes, gastropods and bivalves. This observation was in contradiction to earlier studies which reported a higher abundance of epifauna on *Sargassum* species (Satheesh & Wesley, 2006; Leninraj *et al.*, 2007).

Timing of the sample collection, tidal heights and epiphytic load of the macroalgae have been reported as more important factors determining the epifaunal abundance than structural complexity of the host (Cacabelos *et al.*, 2010). In the present study, samples were collected at the same time in each sampling regime (9–10 a.m.). So the bias due to

sampling time or month was minimized in our study and the density of epifauna did not show significant variation in relation to sampling year. The sampling was also conducted in the same site throughout the study period and all the macroalgal species were collected within the same shore height. The epiphytic load of the macroalgal species may be one of the possible important factors for the observed variability as studies from various coastal regions suggest that the invertebrates associated with macroalgae are taxonomically and morphologically diverse, and show a range of tropic habits. These epibenthic invertebrates may consist of filter feeders (Caine, 1977), grazers that depend on epiphytic algae (Brawley & Fei, 1987), those organisms which eat detritus (Zimmerman *et al.*, 1979), predators which eat other epifauna (Roland, 1978), or herbivores which consume the host alga (Duffy, 1990). As most of the epifaunal groups directly graze upon the macroalgae or the epiphytes for their food source, it is important to consider whether the epiphytic load will differ between macroalgal species. Gestoso *et al.* (2010) confirmed that epiphytic algal density and composition varied between two macroalgal species and influenced the abundance of associated epifauna. A study by

Bologna & Heck (1999) also highlighted the importance of epiphytes in a higher abundance of bivalves associated with seagrass.

The defence measures of the host plant against the epifauna might be another possible reason for the variation in epifaunal assemblages observed in the present study. To prevent colonization by epibionts, macroalgae have widespread means of direct physical protective measures such as release of mucus, peeling of outer teguments, moulting etc. (Wahl, 1989). Among the five macroalgae studied, different species of *Sargassum* are known to produce polyphenols which showed inhibitory activity against invertebrate larvae (Lau & Qian, 1997). Considering the low abundance of epifauna on *Sargassum*, the possible role of a chemical defence mechanism requires consideration. Previous studies mainly on antifouling activities of different macroalgae showed varying levels of activities (Fusetani, 2004). As the chemical defence may vary over spatial and possibly temporal scales further studies may offer more details on this aspect.

In conclusion, our study confirmed that patterns of abundance of epifaunal communities living on macroalgae in the coastal ecosystems are not driven by a single factor but controlled by a combination of factors across temporal scales. The abundance of epifaunal groups such as polychaetes, gastropods and mussels was high on *Padina* and low on *Sargassum*. This variation in the abundance between macroalgal species was possibly not associated with morphology or structural complexity of macroalgae as noted by previous investigations (e.g. Taylor & Cole, 1994). The observed seasonal variation in abundance of some epifaunal groups also attains significance due to the prevailing environmental conditions in the Red Sea. As coastal ecosystems are under the influence of multiple stressors, the high diversity of epifaunal invertebrates associated with macroalgae indicate the need for more studies to predict the factors responsible for the observed variations.

ACKNOWLEDGEMENTS

We thank Mr. Salman A. Al-Ahmadi for his help during field trips.

FINANCIAL SUPPORT

This project was funded by the Deanship of Scientific Research (DSR), King Abdulaziz University (grant no. 473/150/1434). The authors, therefore, acknowledge with thanks DSR technical and financial support.

REFERENCES

- Al-Aidaros A.M. and Satheesh S.** (2014) Larval development and settlement of the barnacle *Amphibalanus amphitrite* from the Red Sea: influence of nauplii hatching season. *Oceanological and Hydrobiological Studies* 43, 17–177.
- Al-Farawati R.** (2010) Environmental conditions of the coastal waters of Southern Corinche, Jeddah, Eastern Red Sea: physico-chemical approach. *Australian Journal of Basic and Applied Sciences* 4, 3324–3337.
- Aoki M.** (1988) Factors affecting population fluctuations of caprellid amphipods inhabiting *Sargassum patens* bed (preliminary report). *Benthos Research* 32, 42–49.
- Bates C.R.** (2009) Host taxonomic relatedness and functional-group affiliation as predictors of seaweed–invertebrate epifaunal associations. *Marine Ecology Progress Series* 387, 125–136.
- Belal A.A. and Ghobashy M.A.F.A.** (2014) Macrobenthic invertebrates in the intertidal zone at both sides of the Suez Gulf. *International Journal of Marine Science* 4, 1–11.
- Bologna P.A.X. and Heck K.L. Jr** (1999) Macrofaunal associations with seagrass epiphytes. Relative importance of trophic and structural characteristics. *Journal of Experimental Marine Biology and Ecology* 242, 21–39.
- Bray R.N. and Ebeling A.W.** (1975) Food, activity, and habitat of three ‘picker-type’ microcarnivorous fishes in the kelp forests off Santa Barbara, California. *Fishery Bulletin* 73, 815–829.
- Brawley S. H. and Fei X.G.** (1987) Studies of mesoherbivory in aquaria and in an unbarricaded mariculture farm on the Chinese coast. *Journal of Phycology* 23, 614–623.
- Cacabelos E., Olabarria C., Incera M., Jesús S. and Troncoso J.S.** (2010) Effects of habitat structure and tidal height on epifaunal assemblages associated with macroalgae. *Estuarine Coastal and Shelf Science* 89, 43–52.
- Caine E.A.** (1977) Feeding mechanisms and possible resource partitioning of the Caprellidae (Crustacea : Amphipoda) from Puget Sound, USA. *Marine Biology* 42, 331–336.
- Caine E.A.** (1991) Caprellid amphipods: fast food for the reproductively active. *Journal of Experimental Marine Biology and Ecology* 148, 27–33.
- Cantin N.E., Cohen A.L., Karnauskas K.B., Tarrant A.M. and McCorkle D.C.** (2010) Ocean warming slows coral growth in the central Red Sea. *Science* 329, 322–325.
- Chemello R. and Milazzo M.** (2002) Effect of algal architecture on associated fauna: some evidence from phytal molluscs. *Marine Biology* 140, 981–990.
- Christie H., Fredriksen S. and Rind E.** (1998) Regrowth of kelp and colonization of epiphyte and fauna community after kelp trawling at the coast of Norway. *Hydrobiologia* 375/376, 49–58.
- Christie H., Jorgensen N.M., Norderhaug K.M. and Waage-Nielsen E.** (2003) Species distribution and habitat exploration of fauna associated with the kelp (*Laminaria hyperborea*) along the Norwegian coast. *Journal of the Marine Biological Association of the United Kingdom* 83, 687–699.
- Cunha T.J., Güth A. Z., Bromberg S. and Sumida P.Y.G.** (2013) Macrofauna associated with the brown algae *Dictyota* spp. (Phaeophyceae, Dictyotaceae) in the Sebastião Gomes Reef and Abrolhos Archipelago, Bahia, Brazil. *Continental Shelf Research* 70, 140–149.
- Duarte C.M.** (2002) The future of seagrass meadows. *Environmental Conservation* 29, 192–206.
- Duffy J.E. and Hay M.E.** (1991) Food and shelter as determinants of food choice by an herbivorous marine amphipod. *Ecology* 72, 1286–1298.
- Duffy J.M.** (1990) Amphipods on seaweeds: partners or pests? *Oecologia* 83, 267–276.
- Edgar G.J.** (1991) Distribution patterns of mobile epifauna associated with rope fibre habitats within the Bathurst Harbour estuary, South-western Tasmania. *Estuarine Coastal and Shelf Science* 33, 589–604.
- Edwards A.J. and Head S.M.** (1987) *Key environments: Red Sea*. Oxford: Pergamon Press.
- El-Sherbiny M.M., Hanafy M.H. and Aamer M.A.** (2007) Monthly variation in abundance and species composition of the epipelagic

- zooplankton off Sharm El-Sheikh, Northern Red Sea. *Research Journal of Environmental Sciences* 1, 200–210.
- Fredriksen S., Christie H. and Saethre B.A.** (2005) Species richness in macroalgae and macrofauna assemblages on *Fucus serratus* L. (Phaeophyceae) and *Zostera marina* L. (Angiospermae) in Skagerrak, Norway. *Marine Biology Research* 1, 2–19.
- Fusetani N.** (2004) Biofouling and antifouling. *Natural Product Reports* 21, 94–104.
- Gestoso I., Olabarria C. and Troncoso J.S.** (2010) Variability of epifaunal assemblages associated with native and invasive macroalgae. *Marine and Freshwater Research* 61, 724–731.
- Hawkins S.J. and Hartnoll R.G.** (1983) Grazing of intertidal algae by marine invertebrates. *Oceanography and Marine Biology: An Annual Review* 21, 195–282.
- Hicks G.R.F.** (1980) Structure of phytal harpacticoid copepod assemblages and the influence of habitat complexity and turbidity. *Journal of Experimental Marine Biology and Ecology* 44, 157–192.
- Hobson E.S. and Chess J.R.** (1976) Trophic interactions among fishes and zooplankters near shore at Santa Catalina Island, California. *Fishery Bulletin* 74, 567–598.
- Holmlund M.B., Peterson C.H. and Hay M.E.** (1990) Does algal morphology affect amphipod susceptibility to fish predation? *Journal of Experimental Marine Biology and Ecology* 139, 65–83.
- Jacobi C.M. and Langevin R.** (1996) Habitat geometry of benthic substrata: effects on arrival and settlement of mobile epifauna. *Journal of Experimental Marine Biology and Ecology* 206, 39–54.
- Jones E. and Thornber C.S.** (2010) Effects of habitat-modifying invasive macroalgae on epiphytic algal communities. *Marine Ecology Progress Series* 400, 87–100.
- Jones G.P.** (1988) Ecology of rocky reef fish of north-eastern New Zealand a review. *New Zealand Journal of Marine and Freshwater Research* 22, 445–462.
- Lau S.C.K. and Qian P.Y.** (1997) Phlorotannins and related compounds as larval settlement inhibitors of the tube-building polychaete *Hydroides elegans*. *Marine Ecology Progress Series* 159, 219–227.
- Leninraj Y., Satheesh S. and Wesley S.G.** (2007) Biodiversity of epifauna associated with *Sargassum* sp. around Kudankulam Nuclear Power Project, East coast of India. In Director – Zoological Survey of India (ed.) *National symposium on conservation and valuation of marine biodiversity*. Kolkata: Zoological Survey of India Publications Division, pp. 219–230.
- Le Lann K., Jegou C. and Stiger-Pouvreau V.** (2008) Effect of different conditioning treatments on total phenolic content and antioxidant activities in two Sargassacean species: comparison of the frondose *Sargassum muticum* (Yendo) Fensholt and the cylindrical *Bifurcaria bifurcata* R. Ross. *Phycological Research* 56, 238–245.
- Leite F.P.P., Tanaka M.O. and Gebara R.S.** (2007) Structural variation in the brown alga *Sargassum cymosum* and its effects on associated amphipod assemblages. *Brazilian Journal of Biology* 67, 215–221.
- Maurer D.L., Leathem W., Kinner P. and Tinsman J.** (1979) Seasonal fluctuations in coastal benthic invertebrate assemblages. *Estuarine and Coastal Marine Science* 8, 181–193.
- McAleece N., Gage J.D.G., Lamshead P.J.D. and Paterson G.L.J.** (1997) BioDiversity. Professional statistics analysis software. <http://www.sams.ac.uk/peter-lamont/biodiversity-pro#sthash.Yl8MCAFl.dpuf>
- Moore P.G.** (1978) Turbidity and kelp holdfast. Amphipoda. I. Wales and S.W. England. *Journal of Experimental Marine Biology and Ecology* 32, 53–96.
- Paiva P.C.** (1993) Trophic structure of a shelf polychaete taxocoenosis in southern Brazil. *Cahiers de Biologie Marine* 35, 39–55.
- Raitsos D.E., Hoteit I., Prihartono P.K., Chronis T., Triantafyllou G. and Abualnaja Y.** (2011) Abrupt warming of the Red Sea. *Geophysical Research Letters* 38, L14601.
- Roberts D.A., Johnston E.L. and Poore A.G.B.** (2008) Contamination of marine biogenic habitats and effects upon associated epifauna. *Marine Pollution Bulletin* 56, 1057–1065.
- Roland W.** (1978) Feeding behaviour of the kelp clingfish *Rimicola muscarum* residing on the kelp *Macrocystis integrifolia*. *Canadian Journal of Zoology* 56, 711–712.
- Satheesh S. and Wesley S.G.** (2006) Polychaetes associated with the brown algae *Sargassum* spp. in Kudankulam waters, East coast of India. *Ecology Environment and Conservation* 13, 321–324.
- Shaikh E.A., Roff J.C. and Dowidar N.M.** (1986) Phytoplankton ecology and production in the Red Sea off Jiddah, Saudi Arabia. *Marine Biology* 92, 405–416.
- Stachowicz J.J.** (2001) Mutualism, facilitation, and the structure of ecological communities. *Bioscience* 51, 235–246.
- Steinberg P.D., DeNys R. and Kjelleberg S.** (1998) Chemical inhibition of epibiota by Australian seaweeds. *Biofouling* 12, 227–244.
- Stoner A.W. and Lewis F.G.** (1985) The influence of quantitative and qualitative aspects of habitat complexity in tropical seagrass meadows. *Journal of Experimental Marine Biology and Ecology* 94, 19–40.
- Tararam A.S. and Wakabara Y.** (1981) The mobile fauna especially Gammaridea of *Sargassum cymosum*. *Marine Ecology Progress Series* 5, 157–163.
- Taylor R.B.** (1998) Seasonal variation in assemblages of mobile epifauna inhabiting three subtidal brown seaweeds in northeastern New Zealand. *Hydrobiologia* 361, 25–35.
- Taylor R.B. and Cole R.G.** (1994) Mobile epifauna on subtidal brown seaweeds in northeastern New Zealand. *Marine Ecology Progress Series* 115, 271–282.
- Thrush S.F. and Dayton P.K.** (2002) Disturbance to marine benthic habitats by trawling and dredging: implications for marine biodiversity. *Annual Review of Ecology, Evolution and Systematics* 33, 449–473.
- Vazquez-Luis M., Sanchez-Jerez P. and Bayle-Sempere J.T.** (2008) Changes in amphipod (Crustacea) assemblages associated with shallow-water algal habitats invaded by *Caulerpa racemosa* var. *cylin-dracea* in the western Mediterranean Sea. *Marine Environmental Research* 65, 416–426.
- Venugopalan V.K. and Paulpandian A.L.** (1989) *Methods in hydrobiology*. Parangipettai: C.A.S. in Marine Biology, Annamalai University.
- Viejo R.M.** (1999) Mobile epifauna inhabiting the invasive *Sargassum muticum* and two local seaweeds in northern Spain. *Aquatic Botany* 64, 131–149.
- Wahl M.** (1989) Marine epibiosis. 1. Fouling and antifouling; some basic aspects. *Marine Ecology Progress Series* 58, 175–189.
- Wikström S.A. and Kautsky L.** (2004) Invasion of a habitat-forming seaweed: effects on associated biota. *Biological Invasions* 6, 141–150.
- and
- Zimmerman R., Gibson R. and Harrington J.** (1979) Herbivory and detritivory among gammaridean amphipods from a Florida seagrass community. *Marine Biology* 54, 41–47.

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