

Seed mass and shape are related to persistence in a sandy soil in northern China

Ling-Ping Zhao¹, Gao-Lin Wu^{1,2} and Ji-Min Cheng^{1,2*}

¹College of Animal Science and Technology of Northwest A & F University, Yangling, Shaanxi 712100, China; ²State Key Laboratory of Soil Erosion and Dryland Farming on the Loess Plateau of Institute of Soil and Water Conservation of Chinese Academy of Sciences & Ministry of Soil Resources, Yangling, Shaanxi 712100, China

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Abstract

Whether and how seed mass and shape are related to seed persistence in soil is a controversial topic in plant ecology. There is little information on this relationship in sandy habitats. We tested whether or not this pattern is retained among 141 species within a range of growth forms, from a wide range of families, in the Horqin sandy land of northern China, which belongs to the inland sand ecosystem. We collected information on seed mass, shape and persistence of these species in this region from published articles. Seed mass and shape were found to be significantly and negatively correlated to persistence in the soil in the Chinese flora, in the same way as in most other floras examined, but not in the same way as the Israeli coastal sand dune flora. The value of seed mass and shape as good predictors of persistence in the soil could be applied to the Horqin sandy land. Although many factors affect the relationship between seed mass, shape and persistence, ease of burial appears to be the main potential interpretation in the Horqin sandy land.

Keywords: persistence, sandy soil, seed mass, seed shape, soil seed-bank

Introduction

Thompson and Grime (1979) first mentioned that plant species with small and rounded seeds tend to persist longer than those with large and elongated or flat seeds in British floras. However, their results on the relationship between seed mass, shape and persistence were unrecognized by other seed ecologists for a long

time, until Thompson *et al.* (1993) conducted a study on the relationship between seed size, shape and persistence and proposed that there is a tendency for species with persistent seed-banks to have smaller seeds than those with transient seed-banks. This question then became a focus of interest of seed ecologists. This relationship has been found in a range of floras, such as grasslands in north-western Europe (Thompson *et al.*, 1993; Bakker *et al.*, 1996; McDonald *et al.*, 1996; Bekker *et al.*, 1998), grasslands in central Argentina (Funes *et al.*, 1999), and a wide range of habitats in Italy (Cerabolini *et al.*, 2003). As a characteristic of a species, persistence means the ability of the species to persist in the soil seed-bank (Csontos and Tamás, 2003; Fenner and Thompson, 2005; Yu *et al.*, 2007). It has become customary for seed banks to be simply divided into transient and persistent types (Thompson and Grime, 1979). A soil seed-bank is classified as persistent if seeds remain viable for more than 1 year. The persistent seed-banks play an important role in the conservation and restoration of destroyed or degraded plant communities, since the persistence in the soil of plant species would be a 'memory' of the original plant community (Bakker *et al.*, 1996). Therefore, the seed persistence characteristic of plant species is also essential to predict restoration success (Bossuyt *et al.*, 2005).

The basis of this relationship proposed by Thompson *et al.* (1993) is ease of burial, since small and rounded seeds are buried more easily and incorporated more quickly into the soil than large and flattened or elongated seeds; hence they have much less chance of germination and more easily escape post-dispersal predation. However, this relationship does not seem to be universal in all floras and vegetation types. Leishman and Westoby (1998) found no apparent relationships between seed mass, shape and persistence across 83 species from semiarid woodlands in Australia, which cast some doubt on the generality of the pattern formalized by Thompson *et al.* (1993). Some studies on the relationship of seed mass

*Correspondence
Fax: + 86 29 87012210
Email: gyzcjm@ms.iswc.ac.cn

and seed persistence in soil have been done in lowland forest (Moles *et al.*, 2000), Iranian grassland, scrub and woodland (Thompson *et al.*, 2001), Mediterranean grass and scrubland (Peco *et al.*, 2003), and Israeli coastal sand dune flora (Yu *et al.*, 2007). Most of these studies reported that seed mass was negatively related to seed persistence in soil. Conversely, Yu *et al.* (2007) found that species with bigger seeds appeared to persist longer in the soil relative to those with smaller seeds across 54 species. However, seed shape was found to be a good predictor of seed persistence (Schwienbacher *et al.*, 2010). There is little information on the relationship of seed shape and seed persistence in soil, especially in sandy grassland communities. Therefore, the question of whether and how seed mass and shape are associated with persistence in the soil should be further studied, preferably in arid or semi-arid ecosystems.

It remains unclear which relationship pattern applies in sandy grassland communities, and what is the main mechanism underlying the pattern. Therefore, it seems clear that this topic should be examined in sandy floras of China, to contribute further information to the debate. The re-analysis work in this paper will play an important role in the conservation and restoration of the species in soil seed-banks in sandy grassland communities.

Data sources and methods

The flora in the study region includes species with a wide range of families and almost all life forms. Species are herbaceous (forbs, grasses and sedges) and shrubs, belonging to a range of ecological types including steppe, meadow, psammophytes and weeds. The grassland region covers an area of approximately 43,000 km², and the elevation is 180–650 m. The region is characterized by a semi-arid continental monsoonal climate. Average annual rainfall is 343 mm (1978–2003), most of which falls during the summer months. Nearly 70–80% of the rainfall concentrates in the July–September period, and rainfall in spring (March–May) accounts for only 12% of annual precipitation. The average annual temperature is 6.5°C, and the coldest and hottest monthly mean temperatures are –13.1°C in January and 23.7°C in July. The prevailing winds during this period were generally north-westerly and westerly, but there is high inter-annual variation in wind regimes (Zhang *et al.*, 2004). Wind has an important effect on the dynamics of vegetation. Annual mean wind velocity ranges from 3.0 to 4.4 m s⁻¹, and the number of gale days (> 16 m s⁻¹) is 21–80 per year. The major wind, occurring in March–May, is north-west, and the secondary wind, occurring in June–September, is south-east. The soils are sandy, loose in texture and

are very susceptible to wind erosion (Li *et al.*, 2003). The surface sand deposits in this region are 20–120 m thick. The landscape is characterized by mobile, semi-stabilized and stabilized dunes.

All the data on seed size and shape (see Table 1) of all species in this paper were taken from the bibliography and literature (Liu *et al.*, 2003, 2004; Yan *et al.*, 2004) which has aggregated from three pre-existing published Horqin steppe species databases on seed mass and seed shape (Liu *et al.*, 2003, 2004; Yan *et al.*, 2004). A total of 141 species (116 genera, 38 families) was contained in the Horqin sandy land area (118°35′–123°30′E, 42°41′–45°45′N) in eastern Inner Mongolia province, China. From all of these, data were obtained on seeds and indehiscent, single-seeded fruits. Five replicates were employed, and 100 air-dried diaspores per replicate to quantify mass. They measured the diaspore with all its permanent attached structures. Most grass caryopses were measured with the persistent lemmas and awns, and the achenes of *Asteraceae* with pappus. The shape of diaspores index was captured by the variance of three dimensions (length, width, height), following the method put forward by Thompson *et al.* (1993). In this way, the shape becomes dimensionless and can vary between 0 and 0.33. A variance close to zero represents perfectly spherical seeds, while the variance increases to a maximum of 0.33 for elongated or flat seeds.

The database of relevant information on soil seed persistence was gathered from the China National Knowledge Infrastructure database for papers published in journals in Chinese, and Elsevier Science Direct database for papers published in journals in English, between 1981 and 2010. Literature data were treated according to the protocol by Thompson *et al.* (1997). The information on persistence was derived from the references (e.g. Zhao and Bai, 2001; Zhao and Li, 2003a, b; Jiang *et al.*, 2004; Zhao *et al.*, 2004; Li *et al.*, 2007; Yan *et al.*, 2007; Zhai *et al.*, 2009). Data from these studies were used to classify species as persistent (seeds persist in the soil for more than 1 year) or transient (seeds persist in the soil for less than 1 year) (Thompson, 1993; Thompson *et al.*, 1998; Csontos and Tamás, 2003; Fenner and Thompson, 2005). If a species is detected to survive in the soil for less than 1 year in artificial burial experiments, it is probably safe to say that this species has a transient seed-bank in the soil by the method of Bakker *et al.* (1996). Species persisting for more than 1 year in artificial burial experiments but surviving in naturally buried experiments for less than 1 year are considered to be species with transient seed-banks (Thompson, 1993; Csontos and Tamás, 2003; Fenner and Thompson, 2005), because artificial burial would circumvent the natural processes of seed burial and has a high probability of overestimating persistence of seeds in the soil (Thompson *et al.*, 2003). Other valuable but indirect evidence can be achieved by

Table 1. Seed mass (mg), seed shape (variance of diaspore dimensions) and persistence in the soil for 91 species in the Horqin sandy land, northern China

Species	Family	Life form	Seed mass (mg)	Seed shape	Persistence	Data source
<i>Adenophora tetraphylla</i>	Campanulaceae	P	0.19	0.091	p	Artificial
<i>Agriophyllum squarrosum</i>	Chenopodiaceae	A	1.52	0.074	p	Artificial
<i>Agropyron cristatum</i>	Poaceae	P	2.04	0.156	t	Artificial
<i>Allium ramosum</i>	Liliaceae	P	1.09	0.070	p	Artificial
<i>Amaranthus retroflexus</i>	Amaranthaceae	A	0.49	0.024	p	Natural
<i>Anemarrhena asphodeloides</i>	Liliaceae	P	3.91	0.126	t	Artificial
<i>Aristida adscensionis</i>	Poaceae	A	0.98	0.207	t	Artificial
<i>Artemisia frigida</i>	Asteraceae	P	0.15	0.088	p	Natural
<i>Artemisia gmelinii</i>	Asteraceae	P	0.20	0.082	p	Natural
<i>Artemisia halodendron</i>	Asteraceae	SS	0.51	0.121	p	Artificial
<i>Artemisia scoparia</i>	Asteraceae	AB	0.05	0.076	p	Natural
<i>Artemisia sieversiana</i>	Asteraceae	AB	0.24	0.078	t	Artificial
<i>Artemisia wudanica</i>	Asteraceae	SS	0.38	0.093	p	Artificial
<i>Arundinella hirta</i>	Poaceae	P	0.58	0.101	p	Natural
<i>Asparagus brachyphyllus</i>	Liliaceae	P	17.14	0.019	t	Artificial
<i>Atraphaxis manshurica</i>	Polygonaceae	S	3.23	0.077	p	Natural
<i>Bassia dasyphylla</i>	Chenopodiaceae	A	0.74	0.152	p	Artificial
<i>Bolboschoenus compactus</i>	Cyperaceae	P	3.00	0.088	p	Natural
<i>Calamagrostis epigeios</i>	Poaceae	P	0.20	0.157	p	Natural
<i>Caragana microphylla</i>	Leguminosae	S	33.24	0.047	t	Artificial
<i>Carduus nutans</i>	Asteraceae	B	14.86	0.159	t	Natural
<i>Chenopodium acuminatum</i>	Chenopodiaceae	A	0.40	0.045	p	Artificial
<i>Chenopodium aristatum</i>	Chenopodiaceae	A	0.09	0.052	p	Natural
<i>Chenopodium glaucum</i>	Chenopodiaceae	A	0.37	0.034	p	Natural
<i>Chloris virgata</i>	Poaceae	A	0.57	0.133	p	Artificial
<i>Cleistogenes squarrosa</i>	Poaceae	P	1.28	0.204	p	Natural
<i>Corispermum candelabrum</i>	Chenopodiaceae	A	2.22	0.112	p	Artificial
<i>Cynanchum sibiricum</i>	Asclepiadaceae	P	7.87	0.140	t	Artificial
<i>Datura stramonium</i>	Solanaceae	A	6.29	0.059	t	Natural
<i>Digitaria ciliaris</i>	Poaceae	A	1.08	0.127	t	Artificial
<i>Echinochloa hispidula</i>	Poaceae	A	1.80	0.060	p	Artificial
<i>Echinops gmelinii</i>	Asteraceae	A	6.27	0.144	p	Artificial
<i>Elymus dahuricus</i>	Poaceae	P	3.93	0.172	t	Artificial
<i>Erigeron acer</i>	Asteraceae	B	0.14	0.102	p	Artificial
<i>Erodium stephanianum</i>	Geraniaceae	A	2.78	0.141	p	Artificial
<i>Eupatorium japonicum</i>	Asteraceae	P	0.32	0.118	p	Artificial
<i>Euphorbia humifusa</i>	Euphorbiaceae	A	0.37	0.040	p	Artificial
<i>Galium verum</i>	Rubiaceae	P	0.62	0.020	t	Artificial
<i>Glycine soja</i>	Leguminosae	A	8.60	0.037	p	Natural
<i>Hedysarum fruticosum</i>	Leguminosae	SS	12.64	0.062	t	Artificial
<i>Hemarthria compressa</i>	Poaceae	P	2.08	0.155	p	Natural
<i>Hemerocallis minor</i>	Liliaceae	P	9.69	0.042	t	Artificial
<i>Hibiscus trionum</i>	Malvaceae	A	3.66	0.027	p	Artificial
<i>Hyoscyamus niger</i>	Solanaceae	B	0.58	0.060	p	Natural
<i>Inula britannica</i>	Asteraceae	P	0.12	0.129	p	Natural
<i>Iris dichotoma</i>	Iridaceae	P	3.74	0.086	p	Artificial
<i>Kochia scoparia</i>	Chenopodiaceae	A	0.20	0.047	p	Artificial
<i>Lactuca indica</i>	Asteraceae	AB	1.12	0.126	t	Artificial
<i>Lactuca tatarica</i>	Asteraceae	P	0.63	0.137	p	Natural
<i>Lappula myosotis</i>	Boraginaceae	A	1.93	0.064	p	Artificial
<i>Leonurus japonicus</i>	Lamiaceae	AB	2.08	0.080	p	Artificial
<i>Lespedeza daurica</i>	Leguminosae	SS	2.34	0.071	p	Artificial
<i>Leymus chinensis</i>	Poaceae	P	2.71	0.171	t	Artificial
<i>Linaria vulgaris</i>	Scrophulariaceae	P	0.17	0.128	t	Artificial
<i>Linum stelleroides</i>	Linaceae	AB	0.48	0.104	p	Artificial
<i>Medicago sativa</i>	Leguminosae	P	2.26	0.060	p	Natural

Table 1. *Continued*

Species	Family	Life form	Seed mass (mg)	Seed shape	Persistence	Data source
<i>Melilotus suaveolens</i>	Leguminosae	AB	1.60	0.061	p	Natural
<i>Pappophorum boreale</i>	Poaceae	A	0.55	0.109	t	Artificial
<i>Pennisetum centrasiaticum</i>	Poaceae	P	0.93	0.159	t	Artificial
<i>Periploca sepium</i>	Asclepiadaceae	S	12.62	0.133	t	Natural
<i>Peucedanum terebinthaceum</i>	Umbelliferae	P	2.60	0.105	p	Artificial
<i>Phragmites communis</i>	Poaceae	P	0.32	0.167	t	Artificial
<i>Plantago maritima</i>	Plantaginaceae	P	0.54	0.099	p	Artificial
<i>Polygonum divaricatum</i>	Polygonaceae	P	8.45	0.090	t	Artificial
<i>Polygonum thunbergii</i>	Polygonaceae	A	0.94	0.007	t	Natural
<i>Portulaca oleracea</i>	Portulacaceae	A	0.15	0.026	p	Natural
<i>Potentilla chinensis</i>	Rosaceae	P	0.42	0.031	p	Natural
<i>Potentilla discolor</i>	Rosaceae	P	0.07	0.048	p	Natural
<i>Ranunculus japonicus</i>	Ranunculaceae	P	0.99	0.100	t	Artificial
<i>Salix gordejewii</i>	Salicaceae	S	0.30	0.096	t	Artificial
<i>Salsola ruthenica</i>	Chenopodiaceae	A	0.60	0.039	p	Natural
<i>Sanguisorba officinalis</i>	Rosaceae	P	1.79	0.031	t	Natural
<i>Saussurea glomerata</i>	Asteraceae	P	0.64	0.145	p	Artificial
<i>Scorzonera albicaulis</i>	Asteraceae	P	3.19	0.184	t	Artificial
<i>Senecio argunensis</i>	Asteraceae	P	0.73	0.144	t	Artificial
<i>Setaria glauca</i>	Asteraceae	A	3.30	0.047	t	Artificial
<i>Setaria viridis</i>	Poaceae	A	0.56	0.089	t	Artificial
<i>Silene jensiseensis</i>	Caryophyllaceae	P	0.15	0.007	t	Natural
<i>Sonchus brachyotus</i>	Asteraceae	P	0.40	0.142	t	Artificial
<i>Sophora flavescens</i>	Leguminosae	P	46.78	0.022	t	Artificial
<i>Spodiopogon sibiricus</i>	Poaceae	P	1.09	0.150	t	Artificial
<i>Stellera chamaejasme</i>	Thymelaeaceae	P	1.16	0.055	p	Artificial
<i>Stipa krylovii</i>	Poaceae	P	13.30	0.198	t	Artificial
<i>Suaeda corniculata</i>	Chenopodiaceae	A	0.89	0.019	p	Artificial
<i>Taraxacum mongolicum</i>	Asteraceae	P	0.87	0.145	t	Artificial
<i>Thalictrum simplex</i>	Ranunculaceae	P	0.73	0.081	p	Artificial
<i>Thermopsis lanceolata</i>	Leguminosae	P	15.90	0.035	p	Natural
<i>Thymus serpyllum</i>	Lamiaceae	SS	0.28	0.088	t	Natural
<i>Tribulus terrestris</i>	Zygophyllaceae	A	130.76	0.044	t	Artificial
<i>Trigonella korshinskyi</i>	Leguminosae	A	3.69	0.062	p	Natural
<i>Xanthium sibiricum</i>	Asteraceae	A	77.89	0.086	t	Artificial

Life form classes: A, annual herbs; AB, annual-biennial herbs; B, biennial herbs; P, perennial herbs; S, shrubs; SS, semi-shrubs. Persistence classes: p, seeds persist in the soil for more than 1 year; t, seeds persist in the soil for less than 1 year. Artificial means persistence data collected from artificial burial experiments; natural means persistence data collected from soil cores (germination or extraction). Nomenclature follows Liu (1985, 1987, 1992) and Liu *et al.* (1996) for Chinese species.

sampling soil cores and using seed germination or seed extraction methods to identify species. We adopted several conservative criteria proposed by Thompson (1993) and Thompson *et al.* (1997) to distinguish species with persistent seed-banks from those with transient seed-banks. These criteria were only applied to naturally buried seed data of the references.

The relationship between seed mass and seed shape was analysed by Pearson correlation analysis. For the difference in seed mass and shape between transient and persistent species, we used the *t*-test to evaluate after seed mass was logarithmically transformed to achieve normality. For all the above analyses, we employed the software

package SPSS Version 16.0 package (SPSS Inc., Chicago, Illinois, USA).

Results

In the dataset 52 species of this area were classified as having persistent seed-banks, 39 were classified as having transient seed-banks, and 50 could not be determined, mainly owing to conflicting or insufficient information. Fifty-seven per cent of species had persistent seed-banks in this region, which is higher than in a Mediterranean sand dune community (Yu *et al.*, 2007). Seed mass ranged from 0.01 mg (*Orobancha coerulescens*) (not shown in Table 1) to 130.76 mg

(*Tribulus terrestris*), almost exactly the same as that in the Mediterranean sand dune community, but narrower than that recorded in New Zealand (Moles *et al.*, 2000) or Iran (Thompson *et al.*, 2001). Seed shape ranged from 0.007 (*Silene jensseensis*) to 0.207 (*Aristida adscensionis*), which was narrower than other studies on dry Mediterranean sand dune communities (Table 1).

Log seed mass was non-significantly negatively related to deviations from isodiametric seed shape ($r = -0.097$, $P = 0.255$) for 141 species. Species with persistent seeds had significantly smaller seeds than those with transient seeds (one-tailed t -test = 3.39, $n = 91$, $P < 0.01$), and species with persistent seeds yielded significantly more spherical shape than those with transient seeds (one-tailed t -test = 1.94, $n = 91$, $P = 0.028$). The relationship between seed mass, shape and persistence is shown in Fig. 1. The 91 species in this area were widely scattered across the mass–shape space. Forty-one species with persistent seed-banks had seed mass <3 mg and seed shape <0.14, and accounted for 78.8% of all persistent species. However, there was no obvious threshold of seed mass (3 mg) and seed shape (0.14) as established by Thompson *et al.* (1993) to distinguish persistent from transient species.

Discussion

Our results show that seed mass and seed shape are both significantly and negatively related to seed persistence in the soil. In other words, small and rounded seeds tend to persist longer in the soil than

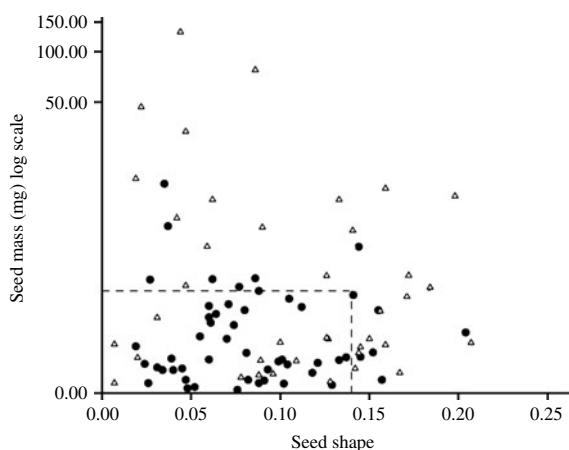


Figure 1. Relationship between seed mass (log scale), shape and persistence for 91 species in the Horqin sandy land, northern China. ●, Species with persistent seed-banks; Δ, species with transient seed-banks. The dashed line denotes the threshold of seed mass (3 mg) and seed shape (0.14) established by Thompson *et al.* (1993) to distinguish persistent from transient species.

large and elongated or flattened ones. The results further confirmed the pattern formalized by Thompson *et al.* (1993), and are consistent with results in other floras around the world, such as Argentina (Funes *et al.*, 1999), Iran (Thompson *et al.*, 2001) and Italy (Cerabolini *et al.*, 2003), but different from the results of Australian (Leishman and Westoby, 1998), New Zealand (Moles *et al.*, 2000) and Israeli floras (Yu *et al.*, 2007).

The results show that seed mass influenced persistence in the soil more than seed shape, that is to say, seed mass is a better predictor of persistence in soil relative to seed shape in the Horqin sandy land, in north-eastern China. However, there is no clear threshold, established by Thompson *et al.* (1993), for species to predict persistence, as detected by other authors (Cerabolini *et al.*, 2003; Peco *et al.*, 2003). The relationship between seed mass, shape and persistence identified in Horqin sandy land may be less clear (or even absent) in floras with a high proportion of legumes. Legume seeds typically have a greater seed mass, are well known to require more time to break the testa, and persist in the soil longer. In the Israeli coastal sand dune flora, Yu *et al.* (2007) detected no obvious relationship between seed mass, shape and persistence, and indicated that this result was probably due to the composition and structure of the vegetation, since legume seeds accounted for 76.0% of the total persistent soil seed-banks in this ecosystem. Especially in the Australian flora, the main reason why there is no close relationship between seed mass, shape and persistence is that persistence is wrongly inferred from dormancy, since there is no close relationship between seed dormancy and the formation or the accumulation of a persistent seed-bank (Thompson *et al.*, 2003; Honda, 2008). However, our study contained species with a range of growth forms from a wide range of families (species of *Asteraceae* and *Poaceae* were dominant in this study site). In this ecosystem, *Leguminosae* only accounted for 6.3% of all species, and legume seeds were 11.5% of the total persistent soil seed-bank. The percentage was much lower relative to the Mediterranean coastal sand dune ecosystem.

Why do seed mass and shape negatively and significantly relate to persistence in the Horqin sandy land? Ease of seed burial, proposed by Thompson *et al.* (1993) is likely to be the main underlying mechanism of patterns observed in this region. The Horqin sandy land is an area of aeolian sandy soil (texture coarse and loosely structured), and seeds seem to be more easily buried in sand ecosystems than in others. So, seeds may indeed be more easily buried in open, loose, sandy soils and have a deep burial position, which restrains seed germination, seedling emergence and establishment (X.W. Zhu *et al.*, 2004; Li *et al.*, 2006; Su *et al.*, 2007; Ma and Liu, 2008; Y.J. Zhu *et al.*, 2009), and then transform into a persistent seed-bank. In addition,

it is often assumed that small seeds have a lower germination rate, seed emergence percentage and require a longer time for germination relative to large seeds, thus they are more likely to persist longer in the soil if they become buried, mainly because small seeds have a greater light requirement for germination (Milberg *et al.*, 2000).

In summary, not only seed mass, but also seed shape, is a key trait in determining seed fate and seed persistence in a sandy soil. Seed mass and seed shape could be robust indicators to predict the ability of seed persistence in soil for a species in this region. Although many factors affect the relationships between seed mass, shape and persistence in soil seed-banks, as mentioned above, ease of burial appears to be the main potential interpretation.

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