

## Original Article

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# Three new meiobenthic species from a submarine cave in Japan: *Echinoderes gama*, *E. kajiharai* and *E. uozumii* (Kinorhyncha: Cyclorhagida)

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## Abstract

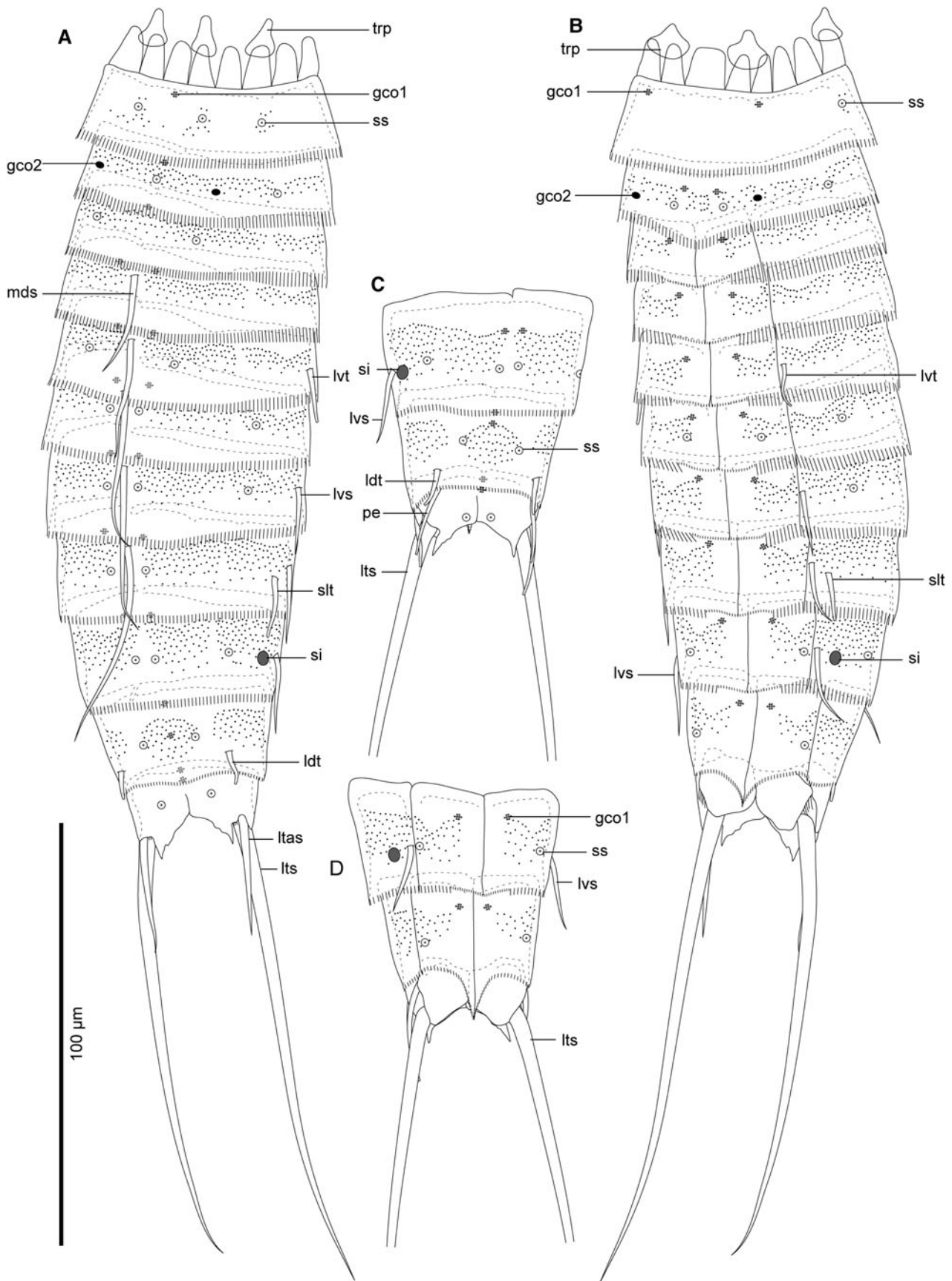
Three new species of echinoderid kinorhynchs are described from Daidokutsu, a submarine cave in Ryukyu Islands, Japan. *Echinoderes gama* sp. nov. is characterized by the presence of middorsal acicular spines on segments 4–8; lateroventral acicular spines on segments 7–9; lateroventral tubes on segment 5; sublateral tubes on segment 8; laterodorsal tubes on segment 10; and type-2 gland cell outlets in subdorsal and lateroventral position on segment 2. *Echinoderes kajiharai* sp. nov. is defined by the presence of middorsal acicular spines on segments 4, 6, 8; lateral accessory acicular spines on segment 9; lateroventral acicular spines on segments 6–8; lateroventral tubes on segments 2 and 5; midlateral tubes on segment 10; and type-2 gland cell outlets in laterodorsal position on segments 2 and 5, and subdorsal position on segments 8 and 9. *Echinoderes uozumii* sp. nov. is characterized by the presence of middorsal acicular spines on segments 4 and 6; lateroventral acicular spines on segments 6–9; lateroventral tubes on segments 2 and 5; sublateral tubes on segment 8; laterodorsal tubes on segment 10; type-2 gland cell outlets in subdorsal and lateral accessory position on segment 2; and blunt, short pectinate fringe teeth of primary pectinate fringe on segment 1. In addition, the *Echinoderes multiporus* species group including *E. kajiharai* sp. nov., and the *Echinoderes bispinosus* species group including *E. uozumii* sp. nov. are established. Furthermore, the distribution of the two species groups and the origin of *Echinoderes* species in Daidokutsu are discussed.

## Introduction

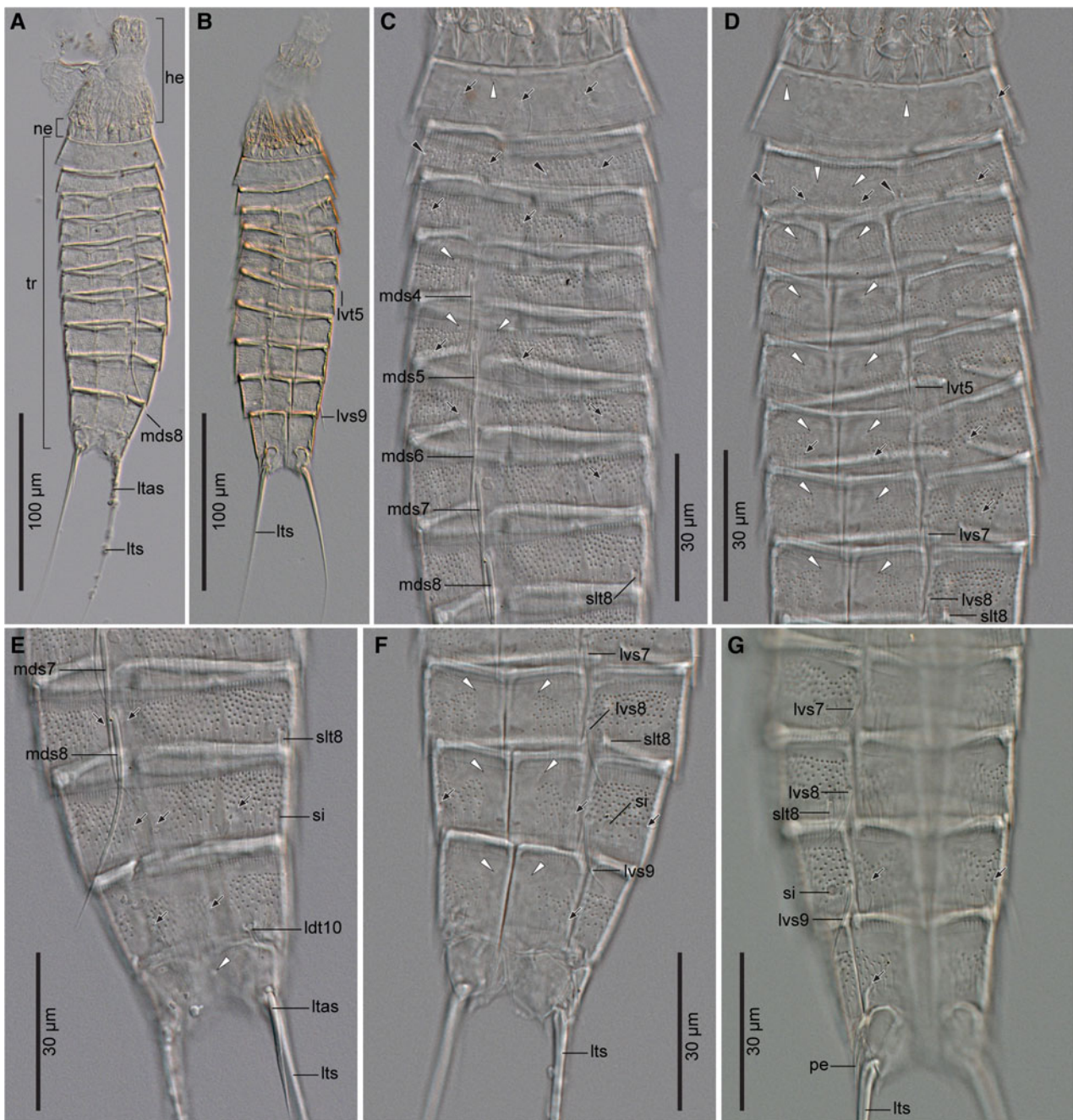
The species of the phylum Kinorhyncha (mud dragons) are exclusively meiobenthic, occur worldwide from tropical to polar regions and inhabit various marine environments such as sandy beaches, muddy sea floor, seamounts, deep sea and submarine caves (Sørensen & Pardos, 2008; Neuhaus, 2013; Yamasaki, 2016; Sørensen *et al.*, 2018; Yamasaki *et al.*, 2019). After the rearrangement of the classification system based on the recent molecular and morphological phylogenetic analyses, 11 families are recognized in the phylum (Sørensen *et al.*, 2015). Among the families, Echinoderidae represents the most speciose family, comprised of five genera and about 150 species (Sørensen *et al.*, 2015; Sánchez *et al.*, 2019; Varney *et al.*, 2019; Yamasaki *et al.*, 2019; Cepeda *et al.*, 2019a, 2019b, 2019c). Kinorhynch families, genera and some species-groups comprised of closely related species are distributed worldwide (Neuhaus, 2013; Randsø *et al.*, 2019). On the other hand, species are mostly reported from single localities and only a few species have broad distribution patterns, i.e. *Echinoderes pterus* Yamasaki *et al.*, 2018a recorded from the Arctic Sea, Atlantic Ocean and Mediterranean Sea and *Echinoderes unispinosus* Yamasaki *et al.*, 2018b from the Atlantic Ocean, Mediterranean Sea and East Pacific Ocean (Sørensen *et al.*, 2018; Yamasaki *et al.*, 2018a, 2018b, 2019). This general pattern of species being restricted to a confined geographic area is probably due to their low dispersal ability, or preference for a specific habitat (Artois *et al.*, 2011; Sánchez *et al.*, 2011, 2012); it should be said, however, that the scarcity of investigations could have prevented an unveiling of an ampler distribution for some species.

Although kinorhynchs inhabit various marine environments, investigations of cave species have been scarcely carried out. Until now, kinorhynchs were found from 12 caves in the north Caribbean Sea, Canary Islands (Atlantic Ocean), western to middle Mediterranean Sea, Atolls of the Maldives (Indian Ocean), southern Japan (Pacific Ocean) and eastern Australia (Coral Sea) (Sánchez & Martínez, 2019). Of the specimens found in these caves, some were identified as putative taxa and others reported as belonging to the following eight nominal species: *Centroderes barbanigra* Neuhaus *et al.*, 2014; *Echinoderes cavernus* Sørensen *et al.*, 2000; *Echinoderes dujardinii* Claparède, 1863; *Leiocanthus bretti* Sánchez & Martínez, 2019; *Meristoderes macracanthus* Herranz *et al.*, 2012; *Pycnophyes kukulkani* Sánchez & Martínez,





**Fig. 1.** *Echinoderes gama* sp. nov., camera lucida drawings. (A, B) holotype, female (ICHUM 5976), whole animal, dorsal and ventral view, respectively; (C, D) paratype male (ICHUM 5977), segments 9–11, dorsal and ventral view, respectively. Abbreviations: gco1/2, type-1/2 gland cell outlet; ldt, laterodorsal tube; ltas, lateral terminal accessory spine; lts, lateral terminal spine; lvs, lateroventral acicular spine; lvt, lateroventral tube; mds, middorsal acicular spine; pe, penile spine; si, sieve plate; slt, sublateral tube; ss, sensory spot; trp, trichoscalid plate.



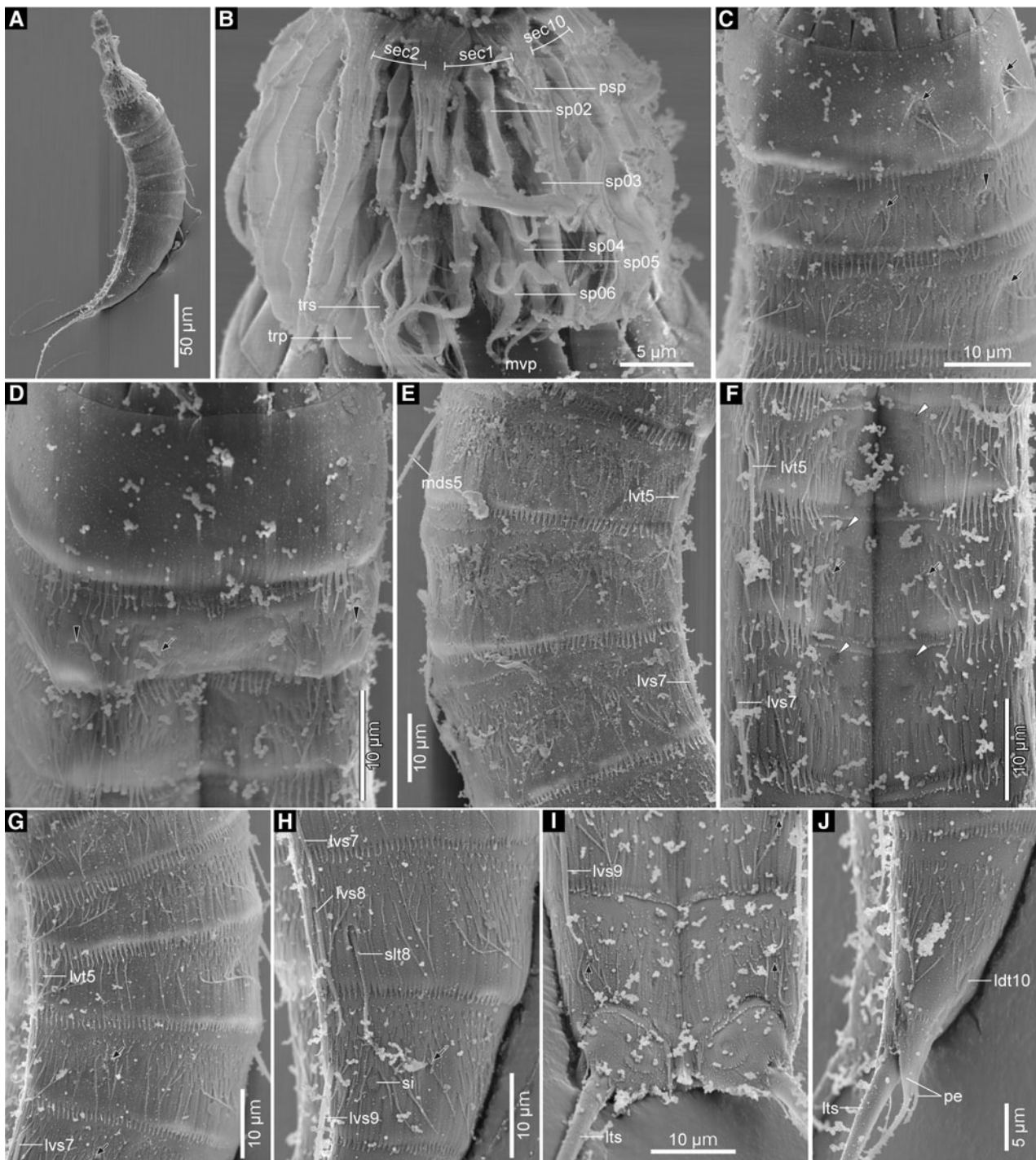
**Fig. 2.** *Echinoderes gama* sp. nov., Nomarski photomicrographs. (A, C–F) holotype, female (ICHUM 5976); (B, G) paratype, male (ICHUM 5977). (A) head, neck and segments 1–11, dorsal view; (B) head, neck and segments 1–11, ventral view; (C), neck and segments 1–8, dorsal view; (D), neck and segments 1–8, ventral view; (E) segments 7–11, dorsal view; (F) segments 7–11, ventral view; (G) segments 7–11, ventral view. Black arrows, white arrowheads and black arrowheads indicate sensory spots, type-1 gland cell outlets and type-2 gland cell outlets, respectively. Numbers after abbreviations indicate corresponding segment number. Abbreviations: he, head; ldt, laterodorsal tube; lts, lateral terminal spine; lvs, lateroventral acicular spine; lvt, lateroventral tube; mds, middorsal acicular spine; ne, neck; pe, penile spine; si, sieve plate; slt, sublateral tube; tr, trunk.

2019; *Pycnophyes* cf. *zelinka* Southern, 1914; and *Ryuguderis iejimaensis* Yamasaki, 2016 (summarized in Sánchez & Martínez, 2019).

Daidokutsu is one of the most investigated submarine caves in the world. The cave is located on the east coast of Iejima Island, Ryukyu Islands, Japan. The cave entrance opens at ~19 m depth on the reef slope, and the cave inside is about 40 m long, gradually deepening to ~30 m depth at its maximum (Kitamura *et al.*, 2007; Omori *et al.*, 2010; Yamamoto *et al.*, 2010). According to Omori *et al.* (2010), the cave has been completely submerged for 8000 years, under the influence of the sea-level rising after the last glacial period. The floor of the cave is covered by a layer of muddy sediment and many animals of various taxa have

been found from the cave, for example, Tardigrada (Fujimoto, 2015; Fujimoto *et al.*, 2017; Fujimoto & Jimi, 2020), Ostracoda (Tabuki & Hanai, 1999; Chiu *et al.*, 2017), Mysida (Hanamura & Kase, 2001) and Mollusca (Kase & Hayami, 1992). The kinorhynch *Ryuguderis iejimaensis* has recently been described from this cave (Yamasaki, 2016).

In this study, three new *Echinoderes* species from Daidokutsu, as the second to fourth species of Kinorhyncha from this cave, are described. They also represent the ninth to eleventh record of kinorhynch species from the cave environment. We furthermore discuss the distribution patterns of two species groups in *Echinoderes* established in this study, and the origin and endemism of echinoderid kinorhynchs in the cave environment.



**Fig. 3.** *Echinoderes gama* sp. nov., males, scanning electron micrographs. (A) segments 1–11, lateral view (left side); (B) head, ventral view; (C), segments 1–3, lateral view (left side); (D), segments 1–3, ventral view; (E) segments 5–7, lateral view (right side); (F) segments 5–7, ventral view; (G) segments 3–7, lateral view (left side); (H) segments 8 and 9 (left side); (I) segments 9–11, ventral view; (J) segments 10 and 11, lateral view (left side). Black arrows point to sensory spots and white/black arrowheads mark type-1/2 gland cell outlets. Numbers after abbreviations indicate corresponding sector/ring of head part or segment number of trunk part. Abbreviations: ldt, laterodorsal tube; lts, lateral terminal spine; lvs, lateroventral acicular spine; lvt, lateroventral tube; mds, middorsal acicular spine;.mvp, mid-ventral placid; pe, penile spine; psp, primary spinoscalid; sec, sector; si, sieve plate; slt, sublateral tube; sp, spinoscalid; trp, trichoscalid plate; trs, trichoscalid.

### Materials and methods

An 8-litre mud sediment sample was collected on 25 April 2015 by scuba at 28–30 m depth in the Daidokutsu submarine cave, Iejima Island, Okinawa, Japan (26°43'29"N 127°49'52"E). Meiofaunal organisms including kinorhynchs were extracted from the sample using the decantation method and the flotation method with Ludox® HS-40 (Higgins & Thiel, 1988; Giere, 2009) and preserved in 10% buffered formalin. The preserved organisms

were subsequently transferred to 70% ethanol and sorted under a stereomicroscope.

Kinorhynch specimens for light microscopy (LM) were dehydrated in glycerol and mounted individually in Fluoromount G® between two cover slips (a 22 mm × 24 mm square cover slip and a circle cover slip 15 mm in diameter) attached to a plastic H-S slide (Shirayama *et al.*, 1993). The LM specimens were observed with either an Olympus BX51 microscope or a Zeiss

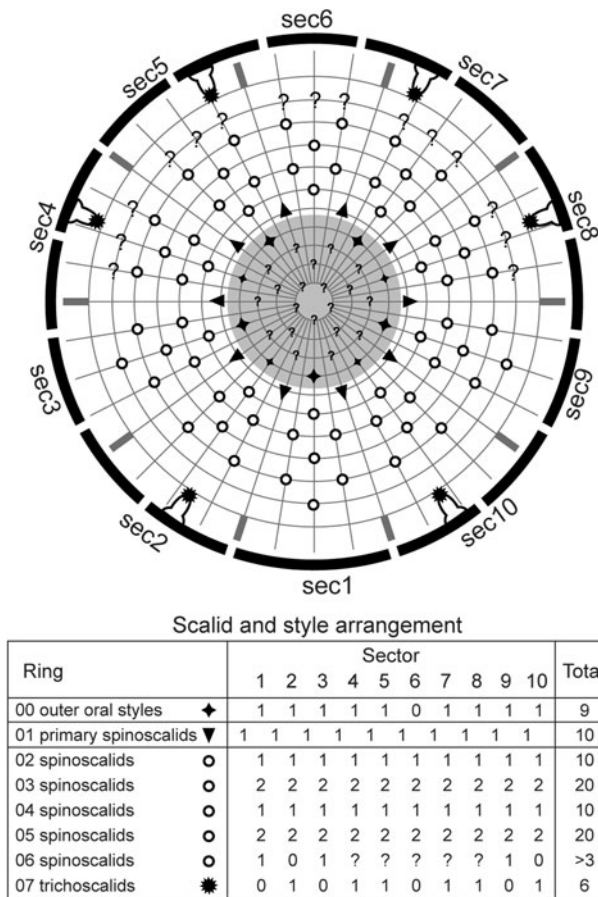


Fig. 4. *Echinoderes gama* sp. nov. and *Echinoderes kajiharai* sp. nov., polar diagram of mouth cone, introvert and placids. Grey shaded area shows mouth cone and bold bent bars symbolize placids. The table lists the scalid arrangement by sector.

Axioskop 50 microscope. A camera lucida equipped on the latter microscope was used to measure morphometrics of all the LM specimens and to make drafts for line art illustrations. The lengths of penile spines were not measured because the spines are so transparent that the correct lengths of them are difficult to measure, and also these lengths are not used as a taxonomic character in *Echinoderes* with the same reason. Final line art illustrations were drawn with Adobe Illustrator CS6 based on the drafts. Specimens were photographed with a Zeiss AxioCam MRC5 mounted on a Zeiss AxioPlan 2 MOT and an Axio Vs40 v.4.8.2.0 software.

Specimens for scanning electron microscopy (SEM) were transferred from ethanol to distilled water through a graded series of ethanol, post-fixed with 1% OsO<sub>4</sub> in 0.05 M phosphate buffer (pH = 7.3) with 0.3 M sodium chloride and 0.05% sodium azide for 2.5 h, dehydrated through a graded series of ethanol, critical-point dried with a BalTec CPD 030, mounted on aluminium stubs, sputter-coated with gold-palladium with a Polaron SC 7640, and observed with a Zeiss EVO LS 10 scanning electron microscope.

The map was drawn with the Generic Mapping Tools (GMT) (Wessel *et al.*, 2013), and subsequently modified with Adobe Illustrator CS6.

The terminology follows Neuhaus (2013). The classification system follows Sørensen *et al.* (2015). All specimens have been deposited in the invertebrate collection of the Hokkaido University Museum, Hokkaido University, Sapporo, Japan (catalogue numbers ICHUM 5976–5992).

Results

Systematics

Class CYCLORHAGIDA Zelinka, 1896  
 Order ECHINORHAGATA Sørensen *et al.*, 2015  
 Family ECHINODERIDAE Zelinka, 1894  
 Genus *Echinoderes* Claparède, 1863

*Echinoderes gama* sp. nov.

(Figures 1–4, Tables 1 & 2)

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Type material

Holotype: female, mounted in Fluoromount G® (ICHUM 5976), collected by HY and SF, at 28–30 m depth in Daidokutsu submarine cave, Iejima Island, Okinawa, Japan (26°43'29"N 127°49'52"E), on 25 April 2015. Paratypes: four males and two females, mounted in Fluoromount G® (ICHUM 5977–5982), collection data as holotype.

Additional material examined

Three males mounted on aluminium stubs for SEM observations, collection data as holotype.

Diagnosis

*Echinoderes* with middorsal acicular spines on segments 4–8; lateroventral acicular spines on segments 7–9; lateroventral tubes on segment 5; sublateral tubes on segment 8; laterodorsal tubes on segment 10; type-2 gland cell outlets in subdorsal and lateroventral position on segment 2.

Etymology

The species name comes from *gama* ('cave'), a word from the Okinawan local language, referring to the species' type locality.

Description

Adult with head, neck and 11 trunk segments (Figures 1A, B, 2A, B & 3A). Measurements and positions of cuticular structures (sensory spots, gland cell outlets, spines, tubes and sieve plates) summarized in Tables 1 & 2, respectively.

Head consisting of retractable mouth cone and introvert. Arrangement of oral styles, scalids and placids summarized in Figure 4. Mouth cone with inner oral styles and one ring of nine outer oral styles. Detailed arrangement of inner oral styles not examined. Each outer oral style consists of triangular distal and rectangular basal parts. Introvert composed of six rings of spinoscalids and one ring of trichoscalids (Figures 3B & 4). Ring 01 with 10 primary spinoscalids, rings 02 and 04 with 10 spinoscalids, and rings 03 and 05 with 20 spinoscalids. Each spinoscalid of rings 01–05 with basal sheath and end piece. Rings 06 not examined in detail, but with at least six spinoscalids. Six trichoscalids attached with trichoscalid plate in sectors 2, 4, 5, 7, 8 and 10.

Neck with 16 placids (Figures 1A, B, 2A–D, 3B & 4). Midventral placid broadest. Remaining placids similar in size. Two ventral and four dorsal trichoscalid plates present.

Segment 1 consisting of complete cuticular ring (Figure 1A, B). This and following 10 segments with pachycyclus at anterior margin of each segment. Sensory spots in subdorsal and laterodorsal position (Figures 1A, B, 2C, D & 3C). Each sensory spot composed of one central pore and numerous micropapillae. Type-1 gland cell outlets present in middorsal and lateroventral position (Figures 1A, B & 2C, D). Cuticular hairs arising from perforation sites: most hairs located around sensory spots, and others sporadically on remaining dorsal area (Figures 1A, 2C & 3C). No cuticular hairs on ventral side (Figures 1B, 2D & 3D). Posterior part of

**Table 1.** Measurements for adult *Echinoderes gama* sp. nov. (in micrometres or as percentage)

Character	Total				Females				Males			
	<i>N</i>	Range	Mean	SD	<i>N</i>	Range	Mean	SD	<i>N</i>	Range	Mean	SD
TL	7	177–190	182.2	4.90	3	180–190	183.9	5.25	4	177–188	180.9	4.95
MSW-8	6	33–35	34.1	0.91	3	33–35	34.3	1.28	3	33–34	33.8	0.46
MSW-8/TL	6	18.3–19.2%	18.6%	0.37%	3	18.3–19.2%	18.7	0.50	3	18.3–18.8%	18.6	0.28
SW-10	6	29–32	30.3	1.61	3	29–32	31.2	1.75	3	29–31	29.4	1.01
SW-10/TL	6	15.8–17.6%	16.5%	0.65%	3	16.2–17.6%	16.9	0.69	3	15.8–16.3%	16.1	0.30
S1	6	21–24	22.1	0.99	3	21–23	21.9	0.96	3	21–24	22.4	1.17
S2	6	19–23	20.3	1.39	3	19–21	19.8	1.14	3	19–23	20.8	1.67
S3	6	17–18	17.4	0.57	3	17–18	17.4	0.71	3	17–18	17.3	0.53
S4	6	17–19	17.5	0.90	3	17–19	17.6	1.22	3	17–18	17.4	0.71
S5	6	16–19	17.5	1.39	3	16–19	17.6	1.85	3	16–19	17.4	1.16
S6	6	17–20	18.4	1.15	3	18–20	18.7	1.07	3	17–19	18.2	1.41
S7	6	18–21	19.3	1.12	3	19–20	19.6	0.27	3	18–21	19.0	1.67
S8	6	20–24	21.7	1.38	3	21–22	22.0	0.61	3	20–24	21.4	2.02
S9	6	22–25	23.4	0.90	3	23–24	23.6	0.61	3	22–25	23.2	1.22
S10	6	28–31	29.8	1.31	3	28–31	29.6	1.97	3	30–31	30.1	0.46
S11	6	17–20	18.8	1.12	3	19–20	19.3	0.96	3	17–19	18.2	1.17
MD4 (ac)	7	18–21	19.5	1.21	3	18–20	19.4	1.22	4	18–21	19.6	1.38
MD5 (ac)	7	28–34	31.3	2.15	3	28–34	31.2	2.79	4	29–33	31.4	2.01
MD6 (ac)	7	34–43	38.3	3.03	3	34–40	37.0	3.04	4	36–43	39.2	3.08
MD7 (ac)	7	35–45	42.6	3.96	3	35–45	41.9	5.62	4	39–45	43.2	3.06
MD8 (ac)	6	49–62	56.8	4.39	3	49–57	53.0	5.57	4	55–62	58.7	2.74
LV5 (tu)	7	9–15	11.8	1.81	3	9–15	11.5	2.99	4	12–13	12.0	0.65
LV7 (ac)	7	13–17	14.6	1.34	3	14–16	15.0	0.94	4	13–17	14.3	1.67
LV8 (ac)	7	14–18	15.8	1.18	3	15–18	16.1	1.32	4	14–17	15.6	1.22
SL8 (tu)	7	12–13	12.7	0.64	3	12–13	12.3	0.40	4	12–13	13.0	0.65
LV9 (ac)	7	17–19	18.3	0.70	3	18–19	18.5	0.80	4	17–19	18.2	0.69
LD10 (tu)	7	7–18	12.5	4.45	3	7–8	7.9	0.46	4	15–18	16.0	1.40
LTS	7	113–125	120.6	3.69	3	113–120	117.7	3.72	4	120–125	122.7	1.89
LTAS	3	30–32	30.6	1.22	3	30–32	30.6	1.22	n.a.	n.a.	n.a.	n.a.
LTS/TL	7	62.8–70.6%	66.2%	2.91%	3	62.8–66.2%	64.0	1.92	4	65.4–70.6%	67.9%	2.45%
LATS/TL	3	16.3–16.8%	16.6%	0.28%	3	16.3–16.8%	16.6	0.28	n.a.	n.a.	n.a.	n.a.

(ac), acicular spine; LD, laterodorsal tube; LTAS, lateral terminal accessory spine; LTS, lateral terminal spine; LV, lateroventral spine/tube; MD, middorsal spine; MSW, maximum sternal width; S, segment length; SL, sublateral tube; SW, standard width; TL, trunk length; (tu), tube.

**Table 2.** Summary of locations of cuticular structures and appendages in *Echinoderes gama* sp. nov.

Position segment	MD	PD	SD	LD	ML	SL	LA	LV	VL	VM
1	gco1		ss	ss				gco1		
2	gco1, ss		gco2		ss			gco2		gco1, ss
3	gco1		ss							gco1
4	ac	gco1								gco1
5	ac	gco1	ss					tu		gco1
6	ac	gco1, ss			ss					gco1, ss
7	ac	gco1, ss			ss			ac		gco1
8	ac	gco1, ss				tu		ac		gco1
9		gco1, ss		ss		si		ac	ss	gco1
10	gco1, gco1		ss	tu					ss	gco1
11	gco1, gco1		ss				pe × 3 (m), ltas (f)	lts		

ac, acicular spine; (f), female condition of sexually dimorphic character; gco1/2, type-1/2 gland cell outlet; LA, lateral accessory; LD, laterodorsal; ltas, lateral terminal accessory spine; lts, lateral terminal spine; LV, lateroventral; (m), male condition of sexually dimorphic character; MD, middorsal; ML, midlateral; pe, penile spine; PD, paradorsal; SD, subdorsal; si, sieve plate; SL, sublateral; ss, sensory spot; tu, tube; VL, ventrolateral; VM, ventromedial.

this and following nine segments with primary pectinate fringe (Figures 1A, B & 3C, D). Pectinate fringe teeth on this segment long and slender.

Segment 2 with complete cuticular ring as segment 1 (Figure 1A, B). Middorsal, midlateral, ventromedial sensory spots present (Figures 1A, B, 2C, D & 3C, D). Each sensory spot of this and following segments consisting of one central pore and fewer micropapillae than those on segment 1. Type-1 gland cell outlets present in middorsal and ventromedial positions (Figures 1A, B & 2D). Type-2 gland cell outlets in subdorsal and lateroventral positions (Figures 1A, B, 2C, D & 3C, D). Primary pectinate fringe as on preceding segment.

Segment 3 and following eight segments consisting of one tergal and two sternal plates (Figure 1A, B). Sensory spots present in subdorsal position (Figures 1A, 2C & 3C). Type-1 gland cell outlets in middorsal and ventromedial position (Figures 1A, B & 2D). Pectinate fringe teeth of primary pectinate fringe on this and following six segments as those on preceding segments, except for ventromedial area with conspicuously shorter teeth (Figures 1B & 3F).

Segment 4 with middorsal acicular spine (Figures 1A & 2C). Type-1 gland cell outlets present in paradorsal and ventromedial positions (Figures 1A, B & 2C, D).

Segment 5 with middorsal acicular spine and lateroventral tubes (Figures 1A, B, 2B–D & 3E–G). Sensory spots in subdorsal position (Figures 1A, & 2C). Type-1 gland cell outlets in paradorsal and ventromedial positions (Figures 1A, B & 2C, D).

Segment 6 with middorsal acicular spine (Figures 1A & 2C). Sensory spots present in paradorsal, midlateral and ventromedial positions (Figures 1A, B, 2C, D & 3F, G). Type-1 gland cell outlets in paradorsal and ventromedial positions (Figures 1A, B & 2D).

Segment 7 with middorsal and lateroventral acicular spines (Figures 1A, B, 2C–G & 3E–H). Sensory spots present in paradorsal and midlateral positions (Figures 1A, B, 2C, D & 3G). Type-1 gland cell outlets in paradorsal and ventromedial positions (Figures 1A, B & 2D).

Segment 8 with middorsal and lateroventral acicular spines, and sublateral tubes (Figures 1A, B, 2A, C–G & 3H). Sensory spots present in paradorsal position (Figures 1A & 2E). Type-1 gland cell outlets in paradorsal and ventromedial positions (Figures 1A, B & 2D, F).

Segment 9 with lateroventral acicular spines (Figures 1, 2B, F, G & 3H, I). Paradorsal, laterodorsal and ventrolateral sensory

spots present (Figures 1, 2E–G & 3H, I). Type-1 gland cell outlets in paradorsal and ventromedial positions (Figures 1 & 2F). Small sieve plates present in sublateral position (Figures 1, 2E–G & 3H).

Segment 10 with laterodorsal tubes (Figures 1A, C, 2E & 3J). Laterodorsal tubes in males longer than those in females (Table 1). Subdorsal and ventrolateral sensory spots present (Figures 1, 2E–G & 3I). Two middorsal type-1 gland cell outlets aligned in tandem (Figure 1A, C). Additional pair of type-1 gland cell outlets present in ventromedial position (Figures 1B & 2F). Pectinate fringe teeth of primary pectinate fringe thinner and shorter than those on preceding segment (Figures 1 & 3I).

Segment 11 with lateral terminal spines (Figures 1, 2A, B, E–G & 3I, J). Three pairs of penile spines present in males: dorsal and ventral pairs long and tube-like, whereas middle one stout and triangular-shaped (Figures 1C, 2G & 3J). Lateral terminal accessory spines present in females (Figures 1A & 2A, E). Subdorsal sensory spots present (Figure 1A, C). Type-1 gland cell outlet present middorsally (Figures 1A, C & 2E). Posterior edge of tergal plate protruding subdorsally, forming short pointed tergal extensions (Figure 1). Posterior edges of sternal plates rounded (Figures 1B, D, 2F & 3I).

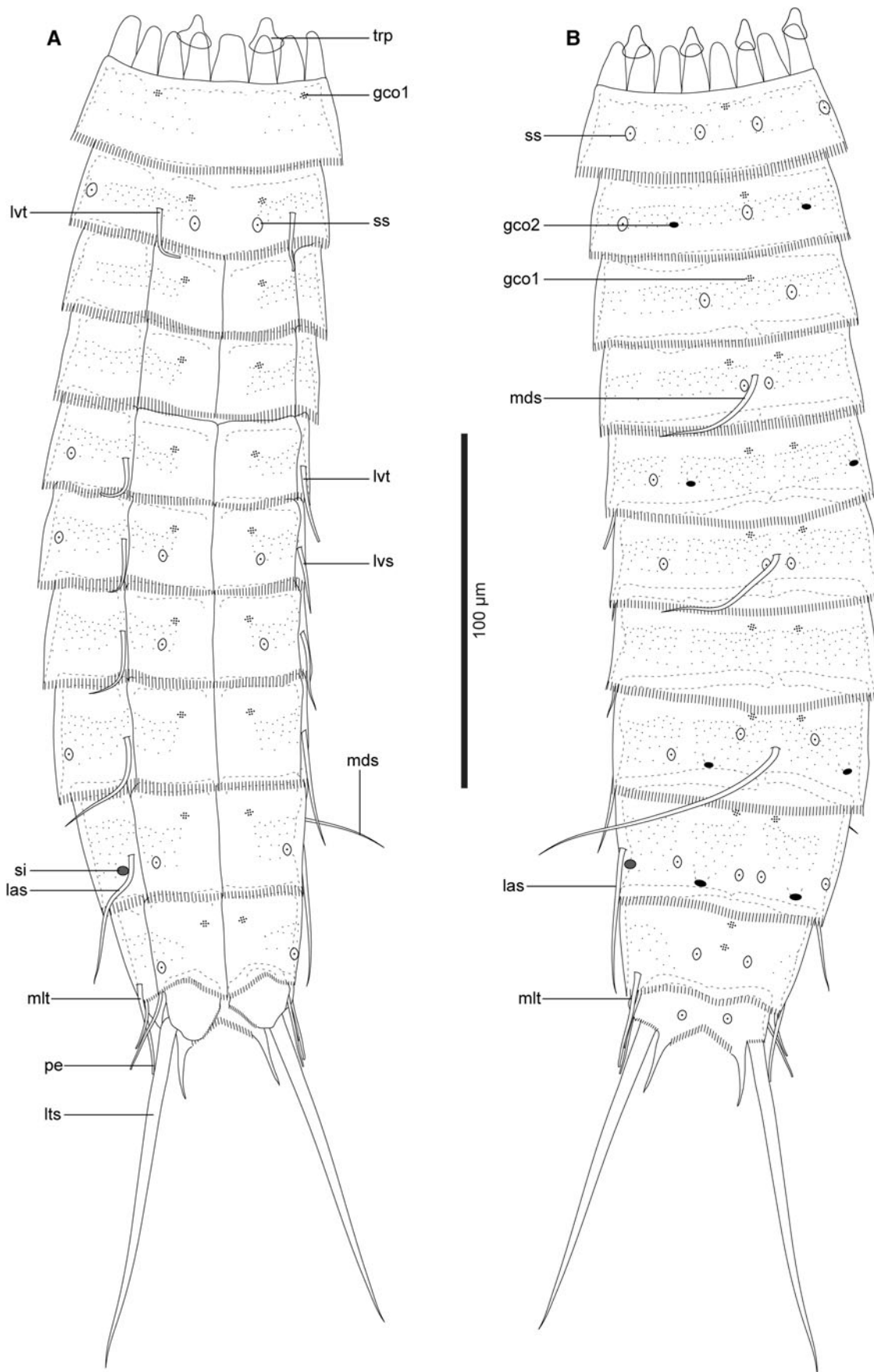
### Remarks

Although the presence of the middorsal acicular spines on segments 4–8 is the most common middorsal-spine pattern in the genus shared by 59 of 127 species, *E. gama* sp. nov. differs from them in the arrangement of the lateroventral acicular spines. Most of the 59 species have lateroventral acicular spines on segments 6–9, whereas only three species lack lateroventral acicular spines on segment 6, namely *Echinoderes eximus* Higgins & Kristensen, 1988, *Echinoderes lusitanicus* Neves *et al.*, 2016, and *Echinoderes tchefouensis* Lou, 1934 (Higgins & Kristensen, 1988; Sørensen *et al.*, 2012; Neves *et al.*, 2016; Grzelak & Sørensen, 2018). Nevertheless, *E. gama* sp. nov. can be easily distinguished from them in the presence of the lateroventral acicular spines on segment 7, which are absent in the three species.

*Echinoderes kajiharai* sp. nov.

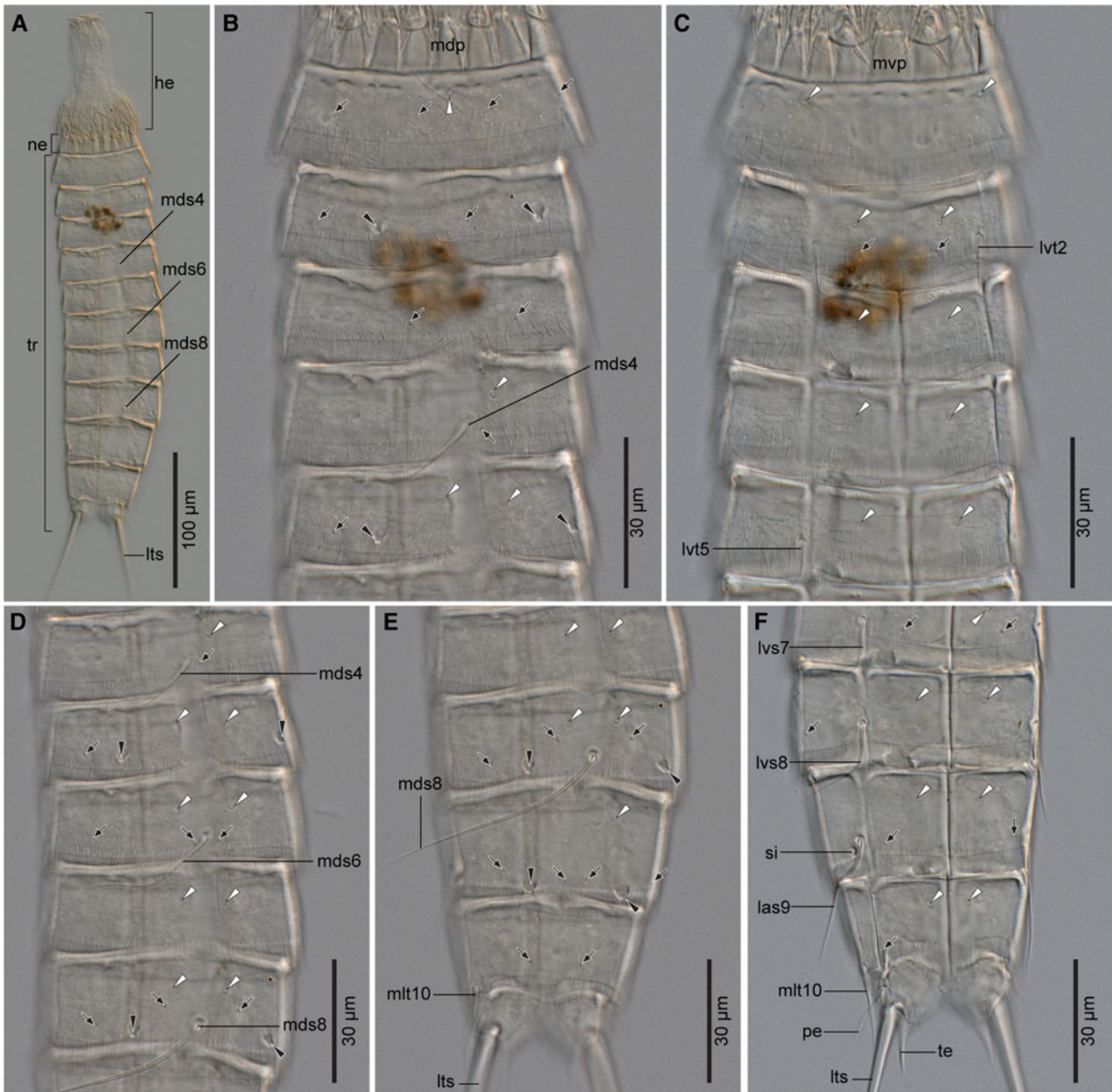
(Figures 5–8, Tables 3 & 4)

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**Fig. 5.** *Echinoderes kajiharai* sp. nov., camera lucida drawings. (A, B) holotype, male (ICHUM 5983), whole animal, ventral and dorsal view, respectively. Abbreviations: gco1/2, type-1/2 gland cell outlet; las, lateral accessory acicular spine; lts, lateral terminal spine; lvs, lateroventral acicular spine; lvt, lateroventral tube; mds, middorsal acicular spine; mlt, midlateral tube; pe, penile spine; si, sieve plate; ss, sensory spot; trp, trichoscalid plate.





**Fig. 6.** *Echinoderes kajiharai* sp. nov., Nomarski photomicrographs, holotype, male (ICHUM 5983). (A) head, neck and segments 1–11, dorsal view; (B) neck and segments 1–5, dorsal view; (C) neck and segments 1–5, ventral view; (D) segments 4–8, dorsal view; (E) segments 7–11, dorsal view; (F) segments 7–11, ventral view. Black arrows, white arrowheads and black arrowheads indicate sensory spots, type-1 gland cell outlets, and type-2 gland cell outlets, respectively. Numbers after abbreviations indicate corresponding segment number. Abbreviations: he, head; las, lateral accessory acicular spine; lts, lateral terminal spine; lvs, lateroventral acicular spine; lvt, lateroventral tube; mdp, middorsal placid; mds, middorsal acicular spine; mlt, midlateral tube;.mvp, midventral placid; ne, neck; pe, penile spine; si, sieve plate; te, tergal extension; tr, trunk.

**Type material**

Holotype: Male, mounted in Fluoromount G® (ICHUM 5983), collected by HY and SF, at 28–30 m depth in Daidokutsu submarine cave, Iejima Island, Okinawa, Japan (26°43'29"N 127°49'52"E), on 25 April 2015. Paratypes: four males, mounted in Fluoromount G® (ICHUM 5984–5987), collection data as holotype.

**Additional material examined**

Two males and one female mounted on aluminium stubs for SEM observations, collection data as holotype.

**Diagnosis**

*Echinoderes* with middorsal acicular spines on segments 4, 6 and 8; lateral accessory acicular spines on segment 9; lateroventral acicular spines on segments 6–8; lateroventral tubes on segments 2

and 5; midlateral tubes on segment 10; type-2 gland cell outlets in laterodorsal position on segments 2 and 5, and subdorsal position on segments 8 and 9.

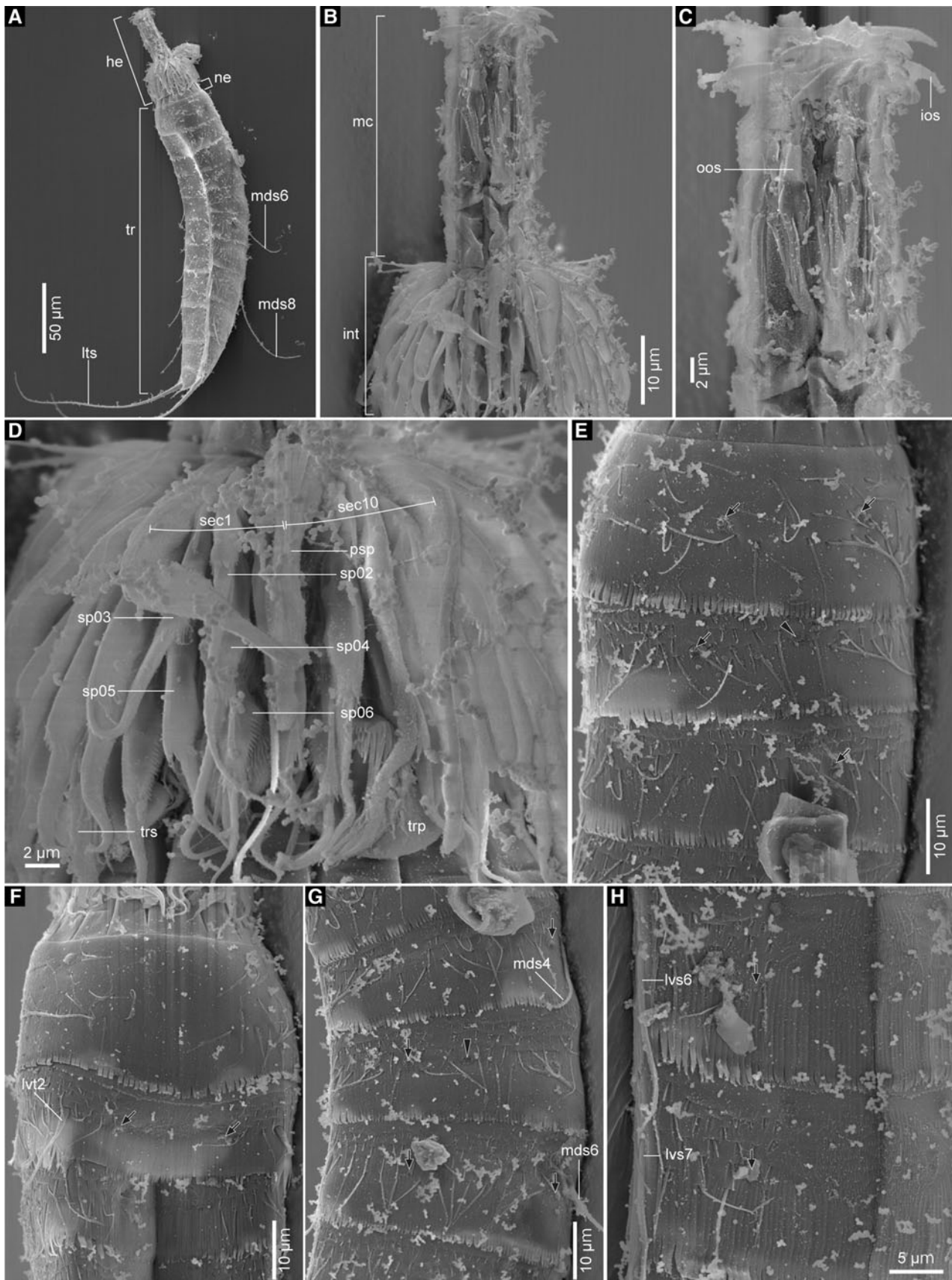
**Etymology**

The species epithet is named after Dr Hiroshi Kajihara, a Japanese taxonomist, for his great contributions to taxonomy, as well as for always supporting and encouraging the first author's research.

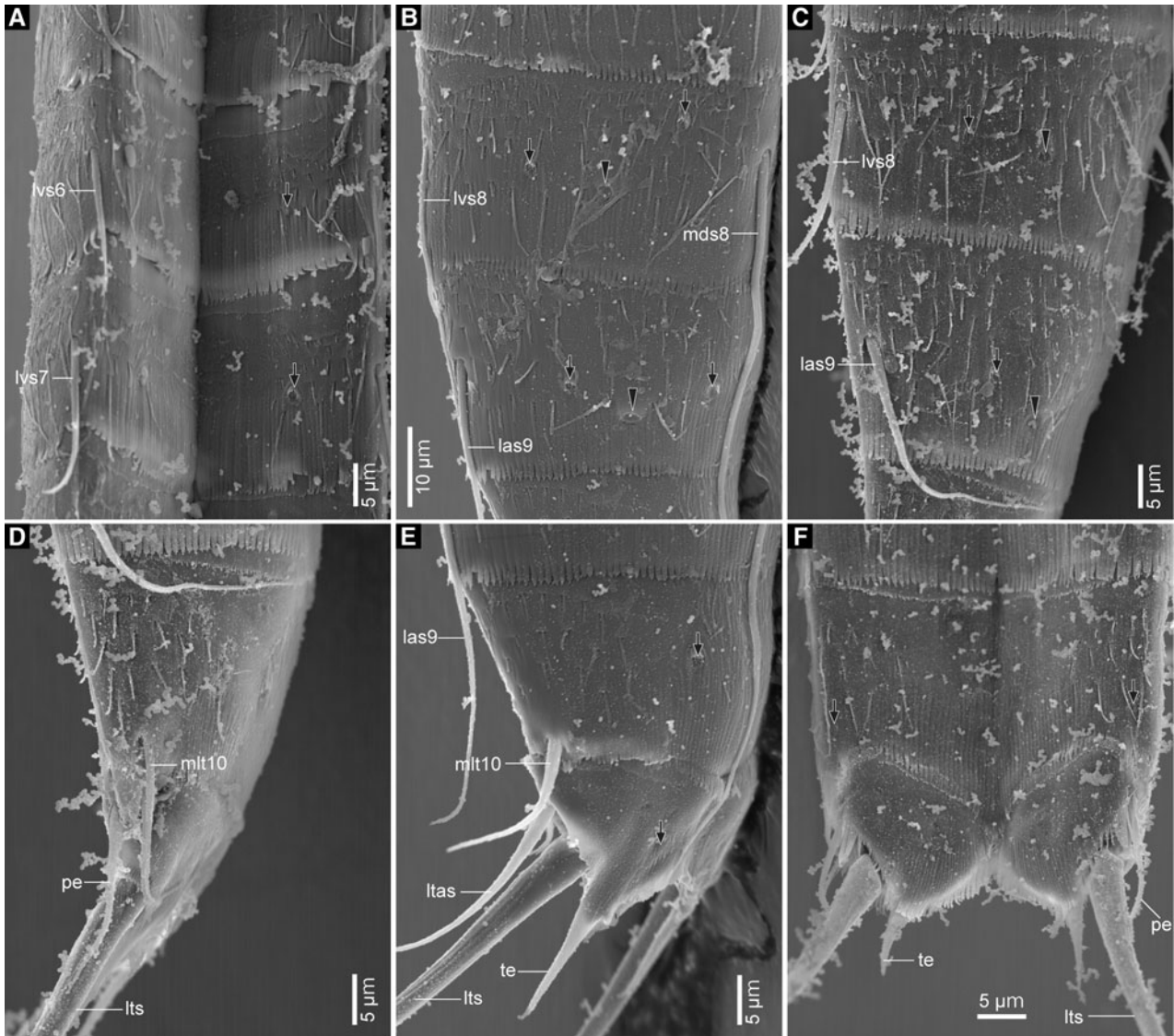
**Description**

Adult with head, neck and 11 trunk segments (Figures 5, 6A & 7A). Measurements presented in Table 3. Positions of cuticular structures (sensory spots, gland cell outlets, spines, tubes and sieve plates) summarized in Table 4.

Head consisting of retractable mouth cone and introvert (Figure 7B). Arrangement of oral styles, scalids and placids



**Fig. 7.** *Echinoderes kajiharai* sp. nov., males, scanning electron micrographs. (A) head, neck and segments 1–11, lateroventral view (left side); (B) head, lateral view (left side); (C) mouth cone, lateral view (left side); (D) introvert, lateroventral view (left side); (E) segments 1–3, laterodorsal view (left side); (F) segments 1–3, ventral view; (G) segments 4–6, laterodorsal view (left side); (H) left sternal plates on segments 6 and 7. Black arrows point to sensory spots and black arrowheads mark type-2 gland cell outlets. Numbers after abbreviations indicate corresponding sector/ring of head part or segment number of trunk part. Abbreviations: he, head; int, introvert; ios, inner oral style; lts, lateral terminal spine; lvs, lateroventral acicular spine; lvt, lateroventral tube; mc, mouth cone; mds, middorsal acicular spine; ne, neck; oos, outer oral style; psp, primary spinoscalid; sec, sector; sp, spinoscalid; tr, trunk; trp, trichoscalid plate; trs, trichoscalid.



**Fig. 8.** *Echinoderes kajiharai* sp. nov., males (A, C, D, F) and female (B, E), scanning electron micrographs. (A) segments 6 and 7, ventral view; (B) segments 8 and 9, lateral view (left side); (C) segments 8 and 9, lateral view (left side); (D) segments 10 and 11, lateral view (left side); (E) segments 10 and 11, lateral view (left side); (F) segments 10 and 11, ventral view. Black arrows point to sensory spots and black arrowheads mark type-2 gland cell outlets. Numbers after abbreviations indicate segment number. Abbreviations: ltas, lateral terminal accessory spine; lts, lateral terminal spine; las, lateral accessory acicular spine; lvs, lateroventral acicular spine; mds, middorsal acicular spine; mlt, midlateral tube; pe, penile spine; te, tergal extension.

shown in Figure 4. Mouth cone with inner oral styles and one ring of nine outer oral styles (Figure 7C). Detailed arrangement of inner oral styles not examined. Each outer oral style consists of triangular distal and rectangular basal parts (Figure 7B, C). Introvert composed of six rings of spinoscalids and one ring of trichoscalids (Figure 7D). Ring 01 with 10 primary spinoscalids, rings 02 and 04 with 10 spinoscalids, and rings 03 and 05 with 20 spinoscalids. Each spinoscalid of rings 01–05 with basal sheath and end piece. Rings 06 not examined in detail, but with at least six spinoscalids. Six trichoscalids attached with trichoscalid plate in sectors 2, 4, 5, 7, 8 and 10.

Neck with 16 placids (Figures 5 & 6B, C), of which midventral one broadest, and remaining ones of similar size. Two ventral and four dorsal trichoscalid plates (Figure 5).

Segment 1 consisting of complete cuticular ring (Figures 5 & 6B, C). This and following 10 segments with thick pachycyclus at anterior margin of each segment. Sensory spots in subdorsal and midlateral positions (Figures 5B, 6B & 7E). Each sensory spot composed of one central pore and numerous micropapillae. Type-1 gland cell outlets present in middorsal and lateral

accessory positions (Figures 5 & 6B, C). Cuticular hairs arising from perforation sites covering entire segment except for ventromedial area on this and following nine segments (Figures 5, 6B, C & 7E, F). Posterior part of this segment and following nine segments with primary pectinate fringe (Figures 5 & 7E, F).

Segment 2 with complete cuticular ring as segment 1 (Figures 5 & 6B, C). Lateroventral tubes present (Figures 5A, 6C & 7F). Sensory spots in middorsal, midlateral and ventromedial positions (Figures 5, 6B, C & 7E, F). Type-1 gland cell outlets present in middorsal and ventromedial positions (Figures 5 & 6C). Type-2 gland cell outlets located laterodorsally (Figures 5B, 6B & 7E).

Segment 3 and following eight segments consisting of one tergal and two sternal plates (Figure 5). Sensory spots present in subdorsal position (Figures 5B, 6B & 7E). Type-1 gland cell outlets as on preceding segment.

Segment 4 with middorsal acicular spine (Figures 5B, 6A, B, D & 7G). Paradorsal sensory spots beside base of middorsal acicular spine (Figures 5B, 6B, D & 7G). Type-1 gland cell outlets present in subdorsal and ventromedial positions (Figures 5 & 6B–D).

**Table 3.** Measurements for adult *Echinoderes kajiharai* sp. nov. (in micrometres or as percentage)

Character	N	Range	Mean	SD
TL	5	284–305	291.4	9.75
MSW-8	5	48–50	48.5	1.15
MSW-8/TL	5	16–17%	16.7%	0.41%
SW-10	5	41–42	41.3	0.56
SW-10/TL	5	14–15%	14.2%	0.42%
S1	5	28–33	30.7	2.00
S2	5	24–27	25.7	1.37
S3	5	20–25	23.1	1.83
S4	5	25–27	25.8	1.00
S5	5	25–27	25.5	1.09
S6	5	27–29	28.2	0.95
S7	5	27–34	29.4	2.59
S8	5	29–35	32.3	2.41
S9	5	33–36	34.6	1.28
S10	5	32–39	35.5	2.87
S11	5	29–34	31.1	1.71
MD4 (ac)	5	31–36	34.6	1.93
MD6 (ac)	5	37–45	39.9	3.54
MD8 (ac)	5	74–78	76.9	1.70
LV2 (tu)	5	15–20	18.2	2.09
LV5 (tu)	5	13–19	17.2	2.61
LV6 (ac)	5	16–24	19.9	2.76
LV7 (ac)	5	19–27	23.2	3.65
LV8 (ac)	5	25–36	30.7	4.40
LA9 (ac)	5	37–45	41.3	3.68
ML10 (tu)	5	16–26	22.6	4.13
LTS	5	167–191	183.6	10.12
LTS/TL	5	59–67%	63.1%	3.79%

ac, acicular spine; LA, lateral accessory spine; LTS, lateral terminal spine; LV, lateroventral spine/tube; MD, middorsal spine; ML, midlateral tube; MSW, maximum sternal width; S, segment length; SW, standard width; TL, trunk length; tu, tube.

Segment 5 with lateroventral tubes (Figures 5A & 6C). Sensory spots situated midlaterally (Figures 5, 6B, D & 7G). Type-1 gland cell outlets as on preceding segment. Type-2 gland cell outlet present on laterodorsal position (Figures 5B, 6B, D & 7G).

Segment 6 with middorsal and lateroventral acicular spines (Figures 5, 6A, D, 7A, G, H & 8A). Sensory spots present in paradorsal, midlateral and ventromedial positions (Figures 5, 6D, 7G, H & 8A). Type-1 gland cell outlets present as on segment 4.

Segment 7 with lateroventral acicular spines (Figures 5A, 6F, 7H & 8A). Sensory spots in ventromedial position (Figures 5A, 6F, 7H & 8A). Type-1 gland cell outlets as on segment 4.

Segment 8 with middorsal and lateroventral acicular spines (Figures 5, 6A, D–F, 7A & 8B, C). Sensory spots in subdorsal and laterodorsal positions (Figures 5, 6D–F & 8B, C). Type-1 gland cell outlets as on segment 4. Subdorsal type-2 gland cell outlet present (Figures 5B, 6D, E & 8B, C).

Segment 9 with lateral accessory acicular spines (Figures 5, 6F & 8B, C, E), of which position may not be fully clear when specimen observed in improper orientation. Paradorsal, laterodorsal and ventrolateral sensory spots present (Figures 5, 6E, F & 8B,

C). Type-1 gland cell outlets as on segment 4. Type-2 gland cell outlets present subdorsally (Figures 5B, 6E & 8B, C). Small sieve plates present in sublateral position (Figures 5 & 6F).

Segment 10 with midlateral tubes (Figures 5, 6E, F & 8D, E). Sensory spots in subdorsal and ventrolateral positions (Figures 5, 6E, F & 8E, F). Two type-1 gland cell outlets in mid-dorsal position (Figure 5B). Additional pair of type-1 gland cell outlets present ventromedially (Figures 5A & 6F).

Segment 11 with lateral terminal spines (Figures 5, 6A, E, F, 7A & 8D–F). Two pairs of penile spines present in males, and pair of lateral terminal accessory spines in females (Figures 5A, 6F & 8D–F). Subdorsal sensory spots present (Figures 5B & 8E). Posterior edge of tergal plate protruding subdorsally, forming long pointed tergal extensions (Figures 5, 6F & 8E, F). Posterior edges of sternal plates rounded (Figures 5A, 6F & 8F).

#### Remarks

Among the congeners, *E. kajiharai* sp. nov. is the most similar to *Echinoderes multiporus* Yamasaki et al., 2018b and *Echinoderes schwieringae* Yamasaki et al., 2019 by sharing the presence of (1) middorsal acicular spines on segments 4, 6, 8, (2) lateroventral/ventrolateral tubes on segments 2 and 5, (3) lateral accessory/lateroventral acicular spines on segments 6–9, (4) midlateral tubes on segment 10, and (5) subdorsal/laterodorsal type-2 gland cell outlets at least on segments 2, 5, 8 and 9. The combination of these similarities suggests that they might be closely related to each other, representing the *Echinoderes multiporus* species group, as with the *Echinoderes coulli* species group and the *Echinoderes spinifurca* species group (see e.g. Sørensen et al., 2018; Randsø et al., 2019; Varney et al., 2019). The presence/absence of female-specific ventrolateral/ventromedial papillae in *E. kajiharai* sp. nov. could not be confirmed due to the lack of female specimens for LM observation. However, the presence of the papillae both in *E. multiporus* and *E. schwieringae* suggests that the female papillae are probably present in *E. kajiharai* sp. nov. as well, making it another diagnostic feature for the *E. multiporus* species group.

The members of the *E. multiporus* species group can be distinguished from each other by the position and the number of the type-2 gland cell outlets. *Echinoderes multiporus* has the highest number of the outlets, namely in the subdorsal position on segment 2 and the laterodorsal position on segments 4–9 (Yamasaki et al., 2018b). *Echinoderes schwieringae* possesses the subdorsal outlets on segments 2 and 4, and the laterodorsal outlets on segments 5 and 7–9 (Yamasaki et al., 2019). *Echinoderes kajiharai* sp. nov., on the other hand, has the lowest number of the outlets, i.e. laterodorsally on segments 2 and 5, and subdorsally on segments 8 and 9.

*Echinoderes kajiharai* sp. nov., further differs from the other two species of the *E. multiporus* species group in the position of acicular spines on segment 9. The spines on segment 9 in *E. kajiharai* sp. nov. are positioned more laterally than those on the preceding segments, locating them in lateral accessory position, whereas in *E. multiporus* and *E. schwieringae*, the spines on segment 9 are in the same lateroventral position as those on the preceding segments.

*Echinoderes uozumii* sp. nov.

(Figures 9–13, Tables 5 & 6)

<http://zoobank.org/3FAC9678-E067-45CB-AB98-ADF634367E97>

#### Type material

Holotype: Female, mounted in Fluoromount G<sup>®</sup> (ICHUM 5988), collected by HY and SF, at 28–30 m depth in Daidokutsu submarine cave, Iejima Island, Okinawa, Japan (26°43'29"N 127°49'52"E), on

**Table 4.** Summary of locations of cuticular structures and appendages in *Echinoderes kajiharai* sp. nov.

Position segment	MD	PD	SD	LD	ML	SL	LA	LV	VL	VM
1	gco1		ss		ss		gco1			
2	gco1, ss			gco2	ss			tu		gco1, ss
3	gco1		ss							gco1
4	ac	ss	gco1							gco1
5			gco1	gco2	ss			tu		gco1
6	ac	ss	gco1		ss			ac		gco1, ss
7			gco1					ac		gco1, ss
8	ac		gco1, ss, gco2	ss				ac		gco1
9		ss	gco1, gco2	ss		si	ac		ss	gco1
10	gco1, gco1		ss		tu				ss	gco1
11			ss				pe × 2 (m), ltas (f)	lts		

ac, acicular spine; (f), female condition of sexually dimorphic character; gco1/2, type-1/2 gland cell outlet; LA, lateral accessory; LD, laterodorsal; ltas, lateral terminal accessory spine; lts, lateral terminal spine; LV, lateroventral; (m), male condition of sexually dimorphic character; MD, middorsal; ML, midlateral; pe, penile spine; PD, paradorsal; SD, subdorsal; si, sieve plate; SL, sublateral; ss, sensory spot; tu, tube; VL, ventrolateral; VM, ventromedial.

25 April 2015. Paratypes: one male and three females, mounted in Fluoromount G® (ICHUM 5989–5992), collection data as holotype.

#### Additional material examined

One male and seven females mounted on aluminium stubs for SEM observations, collection data as holotype.

#### Diagnosis

*Echinoderes* with middorsal acicular spines on segments 4 and 6; lateroventral acicular spines on segments 6–9; lateroventral tubes on segments 2 and 5; sublateral tubes on segment 8; laterodorsal tubes on segment 10; type-2 gland cell outlets in subdorsal and lateral accessory positions on segment 2; pectinate fringe teeth of primary pectinate fringe on segment 1 with blunt tip, and conspicuously shorter than those on following segments.

#### Etymology

The species is named after Ryosuke Uozumi for his contribution to the ecological study of kinorhynchs in Uozumi *et al.* (2018).

#### Description

Adult with head, neck and 11 trunk segments (Figures 9A, B, 10A & 11A). Measurements shown in Table 5. Positions of cuticular structures (sensory spots, gland cell outlets, spines, tubes and sieve plates) summarized in Table 6.

Head consisting of retractable mouth cone and introvert (Figure 11A–D). Arrangement of oral styles and scalds summarized in Figure 12. Mouth cone with inner and outer oral styles (Figure 11B, C). Detail of inner oral styles not observed. Nine outer oral styles present in one ring. Each outer oral style consists of triangular distal and rectangular basal part (Figure 11C). Introvert composed of six rings of spinoscalids and one ring of trichoscalids (Figure 11D). Each spinoscalid of ring 01–05 consisting of basal sheath and distal end piece. Ring 01 with 10 primary spinoscalids, with longest distal end pieces among all spinoscalids. Rings 02 and 04 with 10 spinoscalids; rings 03 and 05 with 20 spinoscalids. Rings 06 with at least eight shortest spinoscalids. Spinoscalids of ring 06 in sectors 5 and 7 not examined. Six trichoscalids attached with trichoscalid plate in sectors 2, 4, 5, 7, 8 and 10. Each trichoscalid covered with hairs.

Neck with 16 placids (Figures 9A, B & 10B, C). Midventral placid broadest. Remaining placids similar in size. Two trichoscalid plates present ventrally and four dorsally.

Segment 1 consisting of complete cuticular ring (Figures 9A, B & 10B, C). This and following 10 segments with thick pachycyclus at anterior margin of each segment. Sensory spots in subdorsal, laterodorsal and ventromedial positions (Figures 9A, B, 10B, C & 11E–G). Each sensory spot on this and following nine segments composed of one central pore and numerous micropapillae, and surrounded by one to three cuticular hairs. Type-1 gland cell outlets present in middorsal and lateroventral positions (Figures 9A, B & 10B, C). Cuticular hairs arising from perforation sites sporadically covering this segment (Figures 9A, B, 10B, C & 11E–G). Posterior part of this segment with primary pectinate fringe, with short and blunt pectinate fringe teeth (Figures 9A, B & 11E–G).

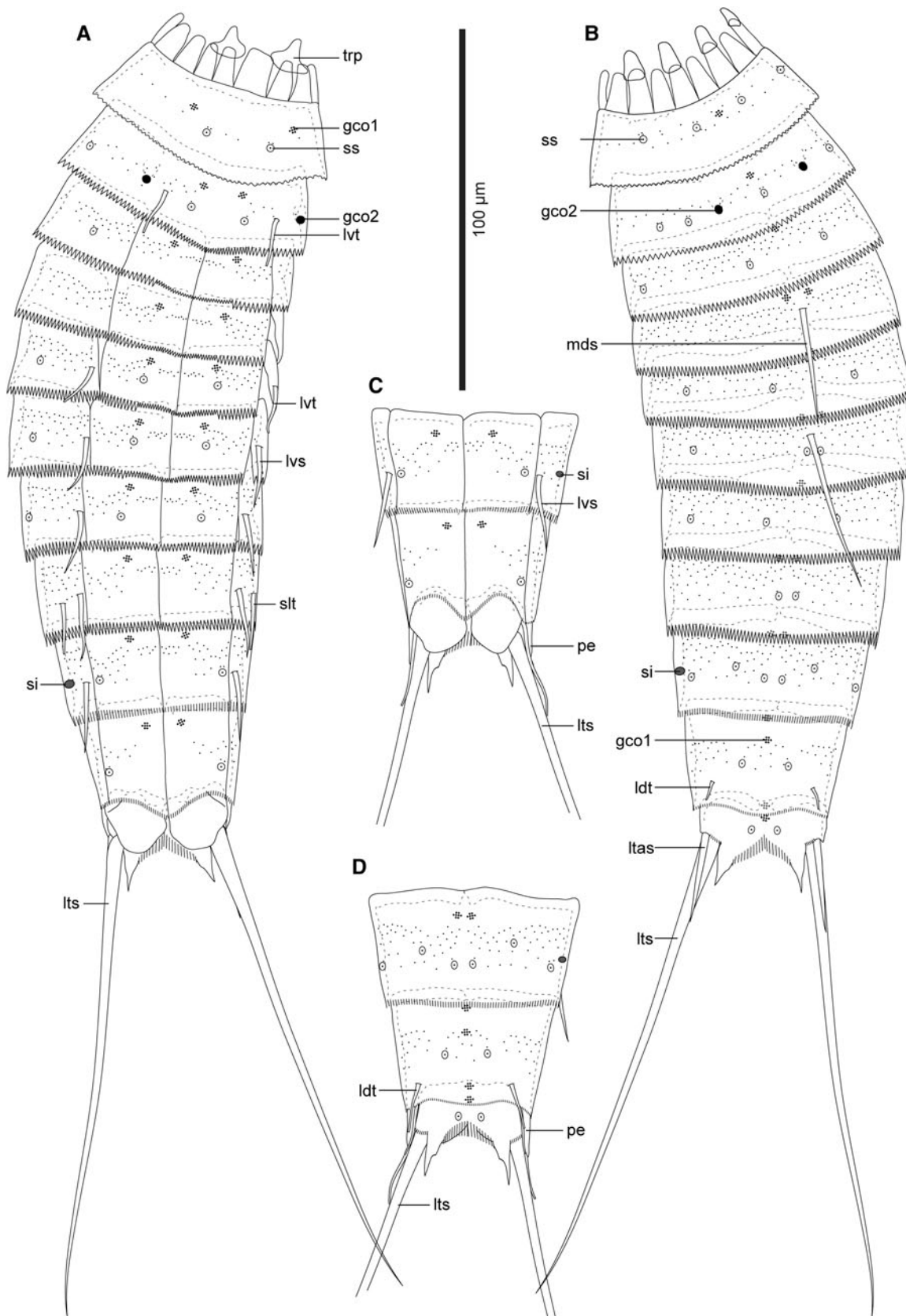
Segment 2 with complete cuticular ring as segment 1 (Figures 9A, B & 10B, C). Lateroventral tubes present (Figures 9A & 11G). Sensory spots present: one in middorsal, two pairs in laterodorsal, and one pair in ventromedial positions (Figures 9A, B, 10B, C & 11E–G). Type-1 gland cell outlets present in middorsal and ventromedial positions (Figures 9A, B & 1C). Type-2 gland cell outlets in subdorsal and lateral accessory positions (Figures 9A, B, 10A–C & 11E–G). Cuticular hairs as in segment 1. Primary pectinate fringe on this and following eight with medium-length pectinate fringe teeth (Figures 9A, B & 11E–G).

Segment 3 and following eight segments consisting of one tergal and two sternal plates (Figures 9A, B & 10B, C). Sensory spots present in subdorsal and midlateral positions (Figures 9A, B, 10B, C, 11E–G & 13A, B). Type-1 gland cell outlets as on preceding segment. Cuticular hairs on this and following seven segments more densely aligned than those on preceding segments (Figures 9A, B, 10A, B, 11E–G & 13A, B).

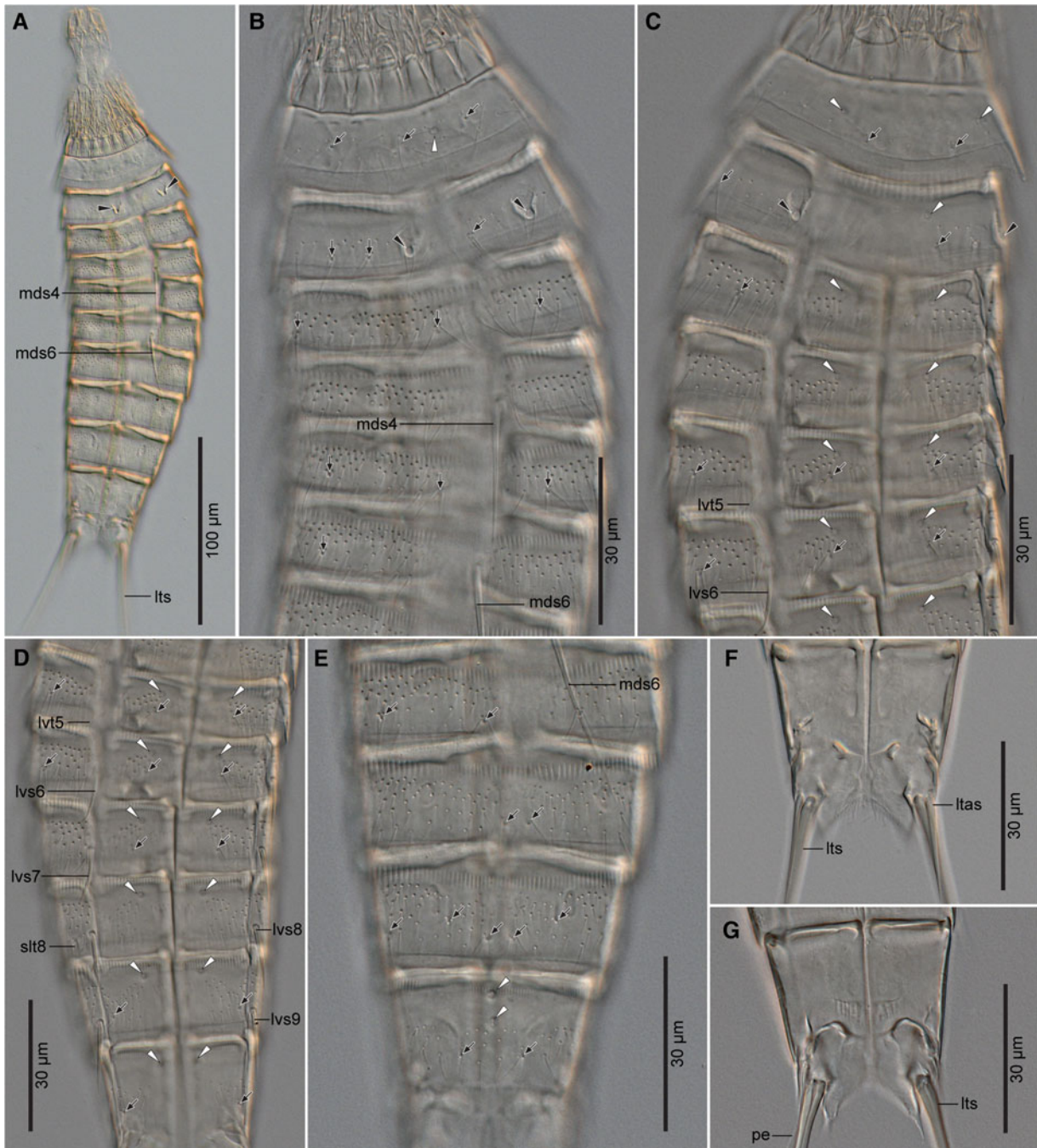
Segment 4 with middorsal acicular spine (Figures 9B, 10A, B, 11E, F, 13A, B). Type-1 gland cell outlets present in paradorsal and ventromedial positions (Figures 9A, B & 10C).

Segment 5 with lateroventral tubes (Figures 9A, 10C, D & 13C). Sensory spots in subdorsal, midlateral and ventromedial positions (Figures 9A, B, 10B–D & 13A–C). Type-1 gland cell outlets as in segment 2.

Segment 6 with middorsal and lateroventral acicular spines (Figures 9A, B, 10A–E & 13C–E). Sensory spots present in



**Fig. 9.** *Echinoderes uozumii* sp. nov., camera lucida drawings. (A, B) holotype, female (ICHUM 5988), whole animal, ventral and dorsal view, respectively; (C, D) paratype male (ICHUM 5989), segments 9–11, ventral and dorsal view, respectively. Abbreviations: gco1/2, type-1/2 gland cell outlet; ldt, laterodorsal tube; lts, lateral terminal accessory spine; lts, lateral terminal spine; lvs, lateroventral acicular spine; lvt, lateroventral tube; mds, middorsal acicular spine; pe, penile spine; si, sieve plate; slt, sublateral tube; ss, sensory spot; trp, trichoscalid plate.



**Fig. 10.** *Echinoderes uozumii* sp. nov., Nomarski photomicrographs. (A–F) holotype, female (ICHUM 5988); (G) paratype, male (ICHUM 5989). (A) head, neck and segments 1–11, dorsal view; (B) neck and segments 1–6, dorsal view; (C) neck and segments 1–6, ventral view; (D) segments 5–10, ventral view; (E) segments 7–10, dorsal view; (F) segments 10 and 11, ventral view; (G) segments 10 and 11, ventral view. Black arrows, white arrowheads, and black arrowheads indicate sensory spots, type-1 gland cell outlets, and type-2 gland cell outlets, respectively. Numbers after abbreviations indicate corresponding segment number. Abbreviations: ltas, lateral terminal accessory spine; lts, lateral terminal spine; lvs, lateroventral acicular spine; lvt, lateroventral tube; mds, middorsal acicular spine; pe, penile spine; stl, sublateral tube.

paradorsal, midlateral and ventromedial positions (Figures 9A, B, 10B–D & 13C–E). Type-1 gland cell outlets present as on segment 4.

Segment 7 with lateroventral acicular spines (Figures 9A, 10D & 13C, F). Sensory spots in subdorsal, midlateral and ventromedial positions (Figures 9A, B, 10D, E & 13C–F). Type-1 gland cell outlets as on segment 2.

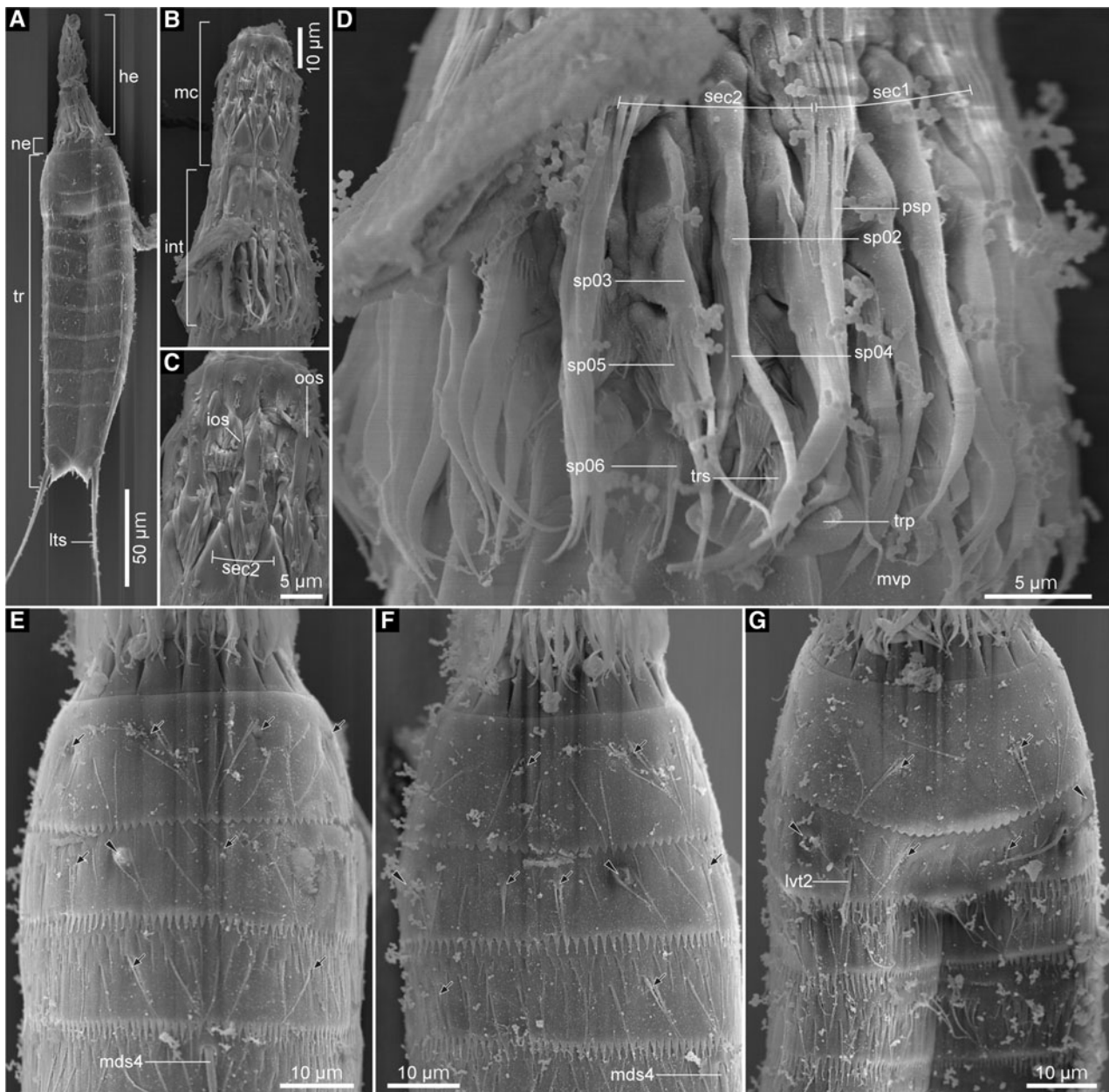
Segment 8 with sublateral tubes and lateroventral acicular spines (Figures 9A, 10D & 13E–G). Sensory spots in paradorsal position (Figures 9B, 10E & 13D, E). Type-1 gland cell outlets as on segment 4.

Segment 9 with lateroventral acicular spines (Figures 9A, C, 10D & 13F, G). Paradorsal, subdorsal, midlateral and ventrolateral

sensory spots present (Figures 9, 10D, E & 13F, G). Type-1 gland cell outlets as on segment 4. Small sieve plates present in sublateral position (Figures 9 & 13F).

Segment 10 with laterodorsal tubes (Figures 9B, D & 13H). Length of laterodorsal tubes in males longer than those in females (Table 5). Subdorsal and ventrolateral sensory spots present (Figures 9, 10D, E & 13G, H). Two middorsal type-1 gland cell outlets aligned in tandem (Figures 9B, D & 10E). Additional pair of type-1 gland cell outlets present in ventromedial position (Figures 9A, C & 10D).

Segment 11 with lateral terminal spines (Figures 9, 10A, F, G, 11A & 13H–J). Three pairs of penile spines present in males



**Fig. 11.** *Echinoderes uozumii* sp. nov., male (B–D) and females (A, E–G), scanning electron micrographs. (A) head, neck and segments 1–11, ventral view; (B) head, ventral view; (C) mouth cone, ventral view; (D) introvert, ventral view; (E) neck and segments 1–4, dorsal view; (F) neck and segments 1–4, laterodorsal view (left side); (G) neck and segments 1–4, ventral view. Black arrows and arrowheads mark sensory spots and type-2 gland cell outlets, respectively. Numbers after abbreviations indicate corresponding sector/ring of head part or segment number of trunk part. Abbreviations: he, head; int, introvert; ios, inner oral style; lts, lateral terminal spine; lvt, lateroventral tube; mc, mouth cone; mds, middorsal acicular spine; mvp, midventral placid; ne, neck; oos, outer oral style; psp, primary spinoscalid; sec, sector; sp, spinoscalid; tr, trunk; trp, trichoscalid plate; trs, trichoscalid.

(Figures 9C, D, 10G & 13J). Dorsal and ventral penile spines long and tube-like, whereas middle ones stout and triangular-shaped. One pair of lateral terminal accessory spines present in females (Figures 9A, B, 10F & 13H, I). Paradorsal sensory spots present (Figures 9B, D & 13H). Two type-1 gland cell outlets present mid-dorsally (Figure 9B, D). Posterior edge of tergal plate forming triangular tergal extensions (Figures 9 & 13H–J). Posterior edges of sternal plates rounded.

#### Remarks

The presence of the middorsal acicular spines only on segments 4 and 6 is an uncommon feature within *Echinoderes*. This trait is shared only by *E. uozumii* sp. nov., *Echinoderes astridae* Sørensen, 2014, and *Echinoderes bispinosus* Higgins, 1982 (Higgins, 1982; Sørensen, 2014). Among the three species, *E.*

*uozumii* sp. nov. and *E. astridae* differ from *E. bispinosus* by the presence of sublateral tubes on segment 8. However, *E. uozumii* sp. nov. and *E. astridae* are identical in the spine/tube pattern, except for the presence/absence of laterodorsal tubes on segment 10. The arrangement of the type-2 gland cell outlets is also identical for the two species. The most prominent difference between the two species is the primary pectinate fringe on segment 1. In *E. uozumii* sp. nov., the pectinate fringe teeth of primary pectinate fringe on segment 1 are short and blunt, obviously differing in shape in comparison with those on following segments. While in *Echinoderes astridae*, the pectinate fringe teeth on segment 1 are medium-length and sharp-angled, which are similar in shape to those on following segments (Sørensen, 2014). Additional differences are (1) the presence/absence of some sensory spots (*E. uozumii* sp. nov. has one or two more pairs of



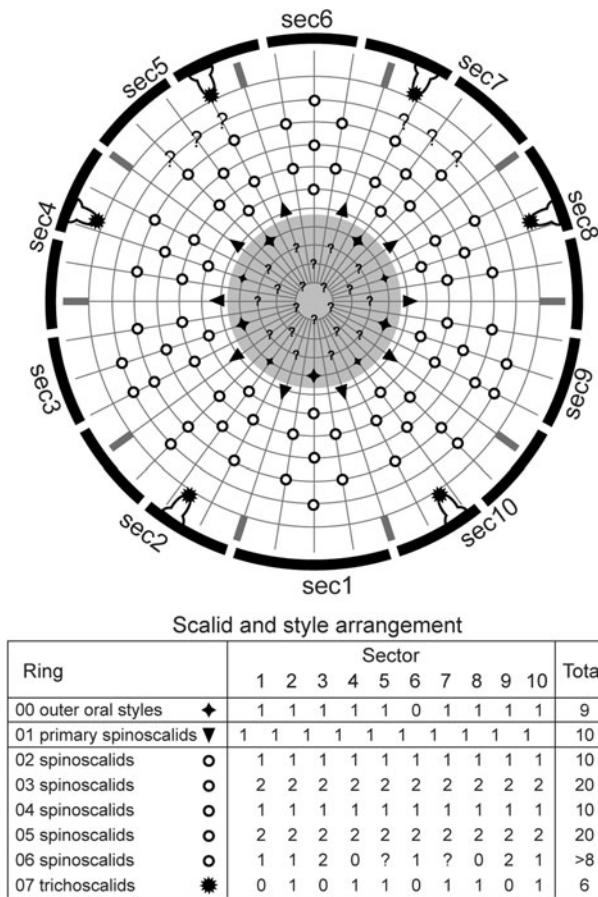


Fig. 12. Polar diagram of mouth cone, introvert and placids in *Echinoderes uozumii* sp. nov. Grey shaded area shows mouth cone and bold bent bars symbolize placids. The table lists the scalid arrangement by sector.

sensory spots on segments 2, 3, 5, 9–11 than *E. astridae*), (2) trunk length (202–229 μm in *E. uozumii* sp. nov., whereas 276–312 μm in *E. astridae*) and (3) relative length of lateral terminal spine (142–146 μm representing 63–70% of trunk length in *E. uozumii* sp. nov., whereas 127–132 μm representing 42–46% of trunk length in *E. astridae*) (Sørensen, 2014).

Two additional species resemble *E. uozumii* sp. nov. in having the middorsal acicular spines on segments 4 and 6: *Echinoderes* aff. *bispinosus* reported from the coast of Turkey by Sönmez et al. (2016) and *Echinoderes* sp. 1 from Singapore by Sørensen et al. (2016). Although the former was identified as ‘*E. aff. bispinosus*’ in the original record by Sönmez et al. (2016), Sørensen et al. (2016) suggested ‘*E. aff. bispinosus*’ is similar to *E. bispinosus* but would represent an undescribed species. In comparison with the brief descriptions provided for the two species, *E. uozumii* sp. nov. differs from the two at least in the presence of the type-2 gland cell outlets in subdorsal and lateral accessory positions on segment 2 (Sönmez et al., 2016; Sørensen et al., 2016). In addition, *E. uozumii* sp. nov. differs from *Echinoderes* sp. 1 in Sørensen et al. (2016) in having lateroventral acicular spines on segment 7 (Sørensen et al., 2016).

All the above-mentioned species, including three known, one new and two undescribed species, share several morphological features: the presence of (1) middorsal acicular spines on segments 4 and 6, (2) lateroventral/ventrolateral tubes on segments 2 and 5 and (3) lateroventral acicular spines at least on segments 6, 8 and 9. These similarities suggest the close relationship of the three species, composing the *Echinoderes bispinosus* species group.

## Discussion

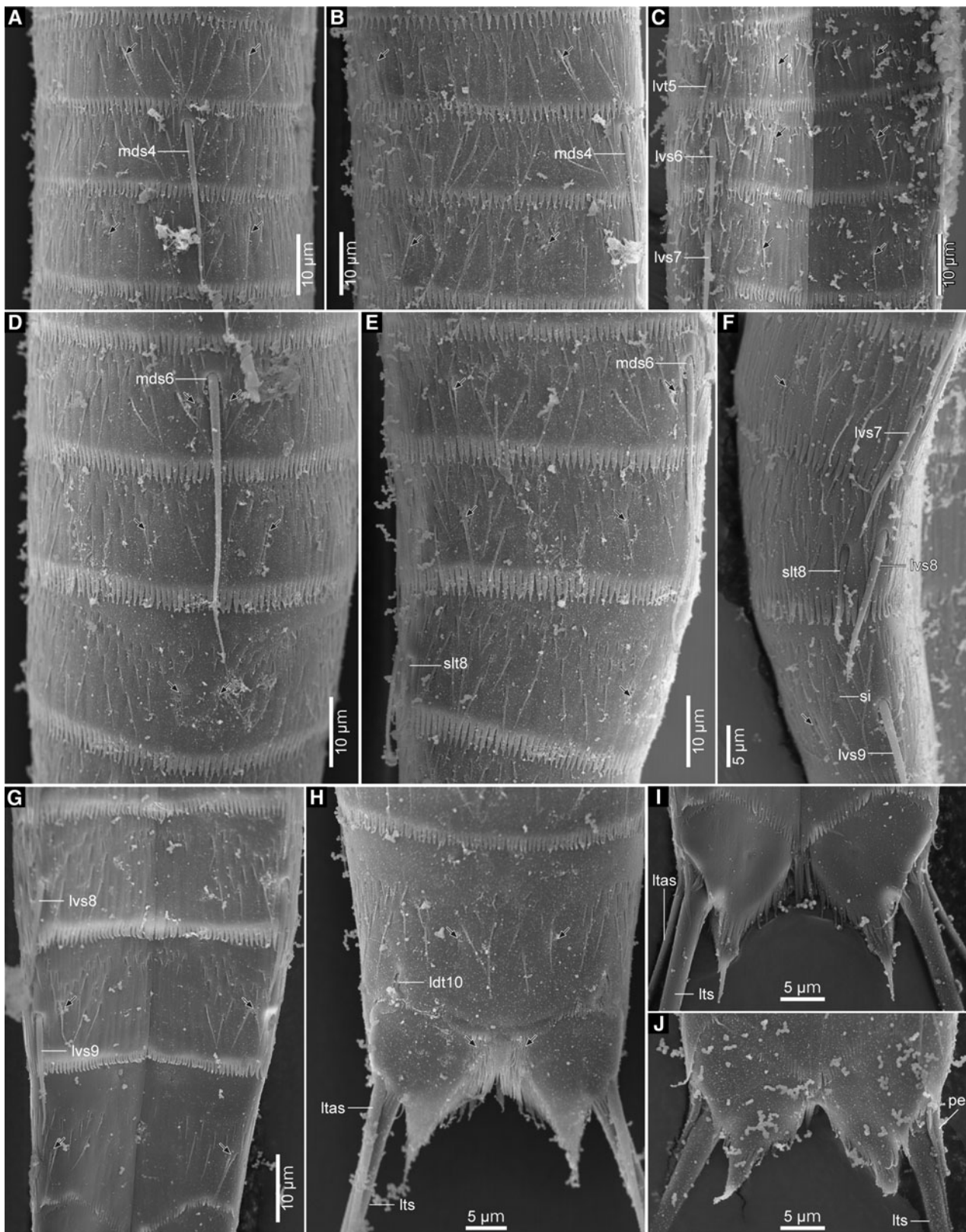
### Wide geographic distributions of *E. multiporus* and *E. bispinosus* species groups

Despite their low motility and lack of a planktonic larval stage, many meiofauna are distributed quite widely and this phenomenon is referred as the ‘meiofauna paradox’ (Higgins & Thiel, 1988; Giere, 2009; Cerca et al., 2018). Recent investigations have revealed that the meiofauna paradox is present in Kinorhyncha at species-group or species level (Neuhaus & Sørensen, 2013; Herranz & Leander, 2016; Yamasaki et al., 2018a, 2019; Randsø et al., 2019). This is also the case for the *E. multiporus* species group and the *E. bispinosus* species group (Figure 14).

With regard to the *E. multiporus* species group, *E. multiporus* and *E. schwieringae* have been recorded from seamounts: the former was from the summit to slope of Eratosthenes Seamount in the east Mediterranean Sea and from the slope of Senghor Seamount in the North-east Atlantic Ocean (Yamasaki et al., 2018b, 2019); the latter was found on the summit of Senghor Seamount in the North-east Atlantic Ocean (Yamasaki et al., 2019). *Echinoderes kajiharai* sp. nov., on the other hand, was from the Daidokutsu submarine cave in the North-west Pacific Ocean (this study), so the distribution of the species group seems to be even wider than was previously supposed.

Concerning the *E. bispinosus* species group, all the congeners inhabit shallow waters in low-latitude regions: *E. bispinosus* was described from Bermuda, North Atlantic Ocean (Higgins, 1982); *E. astridae* was described from the Brazilian coast, South-west Atlantic Ocean (Sørensen, 2014), and more recently reported in the Hispaniola Island in the Caribbean Sea (Cepeda et al., 2019c); ‘*Echinoderes* aff. *bispinosus*’ was reported from a Turkish beach, east Mediterranean Sea (Sönmez et al., 2016); *Echinoderes* sp. 1 in Sørensen et al. (2016) was from the Singapore coast, South China Sea (Sørensen et al., 2016); and *E. uozumii* sp. nov. from the Daidokutsu submarine cave, North-west Pacific Ocean (this study). The last finding expands the species group’s distribution range to the East Asian region.

Recent findings of widely distributed species/species groups of Kinorhyncha have drawn interest to how these wide distribution patterns were formed. Until now, three mechanisms have been proposed. The first mechanism is the dispersal by oceanic currents (Neuhaus & Sørensen, 2013; Yamasaki et al., 2014; Ishii et al., 2016). This would occur for kinorhynchs in association with suspension in the water column (Yamasaki et al., 2014), drifting by regulating its hydrophilic mucus secretion (Ishii et al., 2016), and rafting on other drifting organisms and substrates (Yamasaki et al., 2014). The second mechanism is dispersal effected by human activity (Herranz & Leander, 2016). An intertidal kinorhynch, *Echinoderes ohtsukai* Yamasaki & Kajihara, 2012, was supposedly introduced to British Columbia, Canada, associated with oyster cultivation (Herranz & Leander, 2016). The third mechanism suggests that past tectonic drifting produced the present-day wide distribution of kinorhynchs (Randsø et al., 2019). The wide distribution pattern of *Echinoderes coulli* species group is considered to have formed under the effect of plate tectonics since the Devonian period (Randsø et al., 2019). In the case of the *E. multiporus* species group, at least the second mechanism of artificial invasion can be rejected, because there has been no fishery cultivation in their habitats, i.e. seamounts and a submarine cave, as well as the considerably small impact of human activity. Whereas for the *E. bispinosus* species group, none of the three mechanisms can be ruled out. Further analyses of their phylogenetic relationships within the group are required to support or reject the remaining mechanisms.



**Fig. 13.** *Echinoderes uozumii* sp. nov., male (J) and females (A–I), scanning electron micrographs. (A) segments 3–5, dorsal view; (B) segments 35–, laterodorsal view (left side); (C) segments 5–7, ventral view; (D) segments 6–8, dorsal view; (E) segments 6–8, laterodorsal view (left side); (F) segments 7–9, lateroventral view (right side); (G) segments 8–10, ventral view; (H) segments 10 and 11, dorsal view; (I) segment 11, ventral view; (J) segment 11, ventral view. Black arrows point to sensory spots. Numbers after abbreviations indicate corresponding segment number. Abbreviations: ldt, laterodorsal tube; ltas, lateral terminal accessory spine; lts, lateral terminal spine; lvs, lateroventral acicular spine; lvt, lateroventral tube; mds, middorsal acicular spine; pe, penile spine; si, sieve plate; slt, sublateral tube.

#### *Origin and endemism of echinoderid fauna in Daidokutsu*

Animals living in submarine caves often show high endemism and unique features in fauna, morphology and phylogenetic

relationship (Moldovan *et al.*, 2018). This endemism and uniqueness have been caused by an isolation of the cave environment from open waters, and attracted researchers to study their origin,

**Table 5.** Measurements for adult *Echinoderes uozumii* sp. nov. (in micrometres or as percentage)

Character	Total				Female				Male	
	N	Range	Mean	SD	N	Range	Mean	SD	N	
TL	5	202–229	217.6	9.88	4	202–229	217.6	11.41	1	218
MSW-8	5	42–44	43.1	1.18	4	42–44	42.7	1.03	1	44
MSW-8/TL	5	19–21%	19.8%	0.84%	4	19–21%	19.7%	0.89%	1	20%
SW-10	5	34–38	36.4	1.56	4	34–38	36.1	1.65	1	38
SW-10/TL	5	16–17%	16.7%	0.32%	4	16–17%	16.6%	0.18%	1	17%
S1	5	23–28	25.0	1.94	4	23–28	25.2	2.22	1	25
S2	5	23–25	24.0	1.11	4	23–25	24.3	0.96	1	23
S3	5	18–20	19.3	0.77	4	18–20	19.3	0.88	1	19
S4	5	18–20	18.5	0.96	4	18–20	18.4	1.09	1	19
S5	5	18–23	20.3	1.89	4	18–23	20.4	2.17	1	20
S6	5	19–23	21.0	1.25	4	19–23	21.2	1.38	1	20
S7	5	21–24	23.1	1.11	4	21–24	23.2	1.25	1	23
S8	5	24–27	25.5	1.18	4	24–27	25.7	1.22	1	25
S9	5	26–30	27.0	1.49	4	26–30	27.1	1.71	1	27
S10	5	32–37	34.0	1.59	4	33–37	34.4	1.53	1	32
S11	5	23–30	25.9	2.60	4	23–27	25.0	1.81	1	30
MD4 (ac)	4	30–32	30.6	1.00	3	30–32	30.6	1.22	1	31
MD6 (ac)	5	42–49	45.4	2.80	4	42–49	44.8	2.86	1	48
LV2 (tu)	5	12–13	12.8	0.51	4	12–13	12.8	0.58	1	13
LV5 (tu)	5	12–14	12.8	0.92	4	12–13	12.6	0.81	1	14
LV6 (ac)	5	15–18	16.9	1.21	4	15–18	17.0	1.33	1	16
LV7 (ac)	5	17–20	18.3	1.33	4	17–20	18.4	1.53	1	18
LV8 (ac)	5	18–21	19.4	1.37	4	18–20	18.9	0.95	1	21
SL8 (tu)	5	13–16	13.9	1.27	4	13–16	13.8	1.43	1	14
LV9 (ac)	5	20–22	21.0	1.07	4	20–22	20.7	0.95	1	22
LD10 (tu)	5	4–14	7.1	3.87	4	4–6	5.4	0.79	1	14
LTS	5	142–146	143.8	1.51	4	142–145	143.3	1.11	1	146
LTAS	4	26–29	27.7	1.16	4	26–29	27.7	1.16	n.a.	n.a.
LTS/TL	5	63–70%	66.2%	2.77%	4	63–70%	66.0%	3.15%	1	67%
LTAS/TL	4	12–13%	12.7%	0.49%	4	12–13%	12.7	0.49	n.a.	n.a.

ac, acicular spine; LD, laterodorsal; LTAS, lateral terminal accessory spine; LTS, lateral terminal spine; LV, lateroventral spine/tube; MD, middorsal spine; MSW, maximum sternal width; S, segment length; SL, sublateral tube; SW, standard width; TL, trunk length; tu, tube.

adaptation and evolution in the cave environment (Moldovan *et al.*, 2018). Nevertheless, investigations of kinorhynchs in caves have been scarcely conducted. So far, only eight species are known from the cave environment, three of which represent the species of Echinoderidae, i.e. *E. cavernus*, *E. dujardinii* and *M. macracanthus* (Riedl, 1966; Sørensen *et al.*, 2000; Dal Zotto & Todaro, 2016). Among the three species, only *E. cavernus* is exclusively known from the cave environment, whereas the other two have been reported both inside and outside caves (Sánchez & Martínez, 2019). Based on these echinoderid records from caves made so far, it would be rather plausible to regard most cave-dwelling representatives of *Echinoderes* as not endemic to the cave environment.

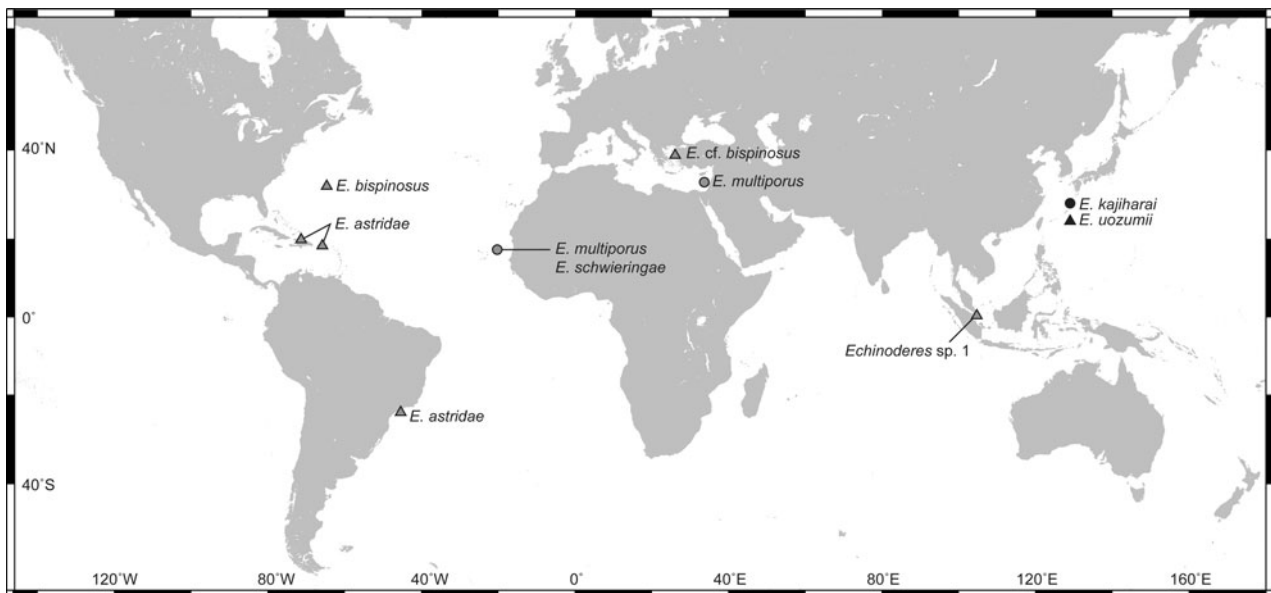
The three species described in this study, *E. gama* sp. nov., *E. kajiharai* sp. nov. and *E. uozumii* sp. nov., have been found only from Daidokutsu. However, the scarce knowledge of kinorhynch

fauna outside the cave does not allow us to reject the possibility that they inhabit other neighbouring environments outside of Daidokutsu. Because Daidokutsu is a young submarine cave, which submerged ~8000 years ago (Omori *et al.*, 2010), the three kinorhynchs arguably invaded the cave relatively recently from the neighbouring waters, for instance from muddy bottom (similar sediment type to the cave) of open waters, under overhangs, or even other caves in the Ryukyu Islands. Considering their recent invasion to Daidokutsu, they may still inhabit such environments outside Daidokutsu also. It also can be assumed that *E. kajiharai* sp. nov. and *E. uozumii* sp. nov. can survive in non-cave environments, because all the other congeners of the *E. multiporus* and the *E. bispinosus* species groups inhabit non-cave environments. For now, we cannot conclude whether the three new species exclusively inhabit caves. Further investigations in the neighbouring environment of Daidokutsu, and

**Table 6.** Summary of locations of cuticular structures and appendages in *Echinoderes uozumii* sp. nov.

Position segment	MD	PD	SD	LD	ML	SL	LA	LV	VL	VM
1	gco1		ss	ss				gco1		ss
2	gco1, ss		gco2	ss, ss			gco2	tu		gco1, ss
3	gco1		ss		ss					gco1
4	ac	gco1								gco1
5	gco1		ss		ss			tu		gco1, ss
6	ac	gco1, ss			ss			ac		gco1, ss
7	gco1		ss		ss			ac		gco1, ss
8		gco1, ss				tu		ac		gco1
9		gco1, ss	ss		ss	si		ac	ss	gco1
10	gco1, gco1		ss	tu					ss	gco1
11	gco1, gco1	ss					pe × 3 (m), ltas (f)	lts		

ac, acicular spine; (f), female condition of sexually dimorphic character; gco1/2, type-1/2 gland cell outlet; LA, lateral accessory; LD, laterodorsal; ltas, lateral terminal accessory spine; lts, lateral terminal spine; LV, lateroventral; (m), male condition of sexually dimorphic character; MD, middorsal; ML, midlateral; pe, penile spine; PD, paradorsal; SD, subdorsal; si, sieve plate; SL, sublateral; ss, sensory spot; tu, tube; VL, ventrolateral; VM, ventromedial.



**Fig. 14.** Map of locations of all records of the *E. multiporus* species group (circles) (Yamasaki et al., 2018b, 2019; this study) and the *E. bispinosus* species group (triangles) (Higgins, 1982; Sørensen, 2014; Sönmez et al., 2016; Sørensen et al., 2016; Cepeda et al., 2019c; this study).

phylogenetic analyses of the *E. multiporus* and the *E. bispinosus* species groups will shed light on their endemism and migration process into the cave.

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## References

Artois T, Fontaneto D, Hummon WD, McInnes SJ, Todaro MA, Sørensen MV and Zullini A (2011) Ubiquity of microscopic animals? Evidence from the morphological approach in species identification. In Fontaneto D (ed.),

*Biogeography of Microscopic Organisms: Is Everything Small Everywhere?* Cambridge: Cambridge University Press, pp. 244–283.

Cepeda D, Álvarez-Castillo L, Hermoso-Salazar M, Sánchez N, Gómez S and Pardos F (2019a) Four new species of Kinorhyncha from the Gulf of California, eastern Pacific Ocean. *Zoologischer Anzeiger* **282**, 140–160.

Cepeda D, Pardos F and Sánchez N (2019b) Kinorhyncha from the Caribbean, with the description of two new species from Puerto Rico and Barbados. *Zoologischer Anzeiger* **282**, 127–139.

Cepeda D, Sánchez N and Pardos F (2019c) First extensive account of the phylum Kinorhyncha from Haiti and the Dominican Republic (Caribbean Sea), with the description of four new species. *Marine Biodiversity* **49**, 2281–2309.

Cerca J, Purschke G and Struck TH (2018) Marine connectivity dynamics: clarifying cosmopolitan distributions of marine interstitial invertebrates and the meiofauna paradox. *Marine Biology* **165**, 123.

Chiu W, Yasuhara M, Iwatani H, Kitamura A and Fujita K (2017) Response of subtropical submarine-cave ecosystem to Holocene cave development and Asian monsoon variability. *Paleobiology* **43**, 425–434.

Claparède ARE (1863) Zur Kenntnis der Gattung *Echinoderes* Duj. In *Beobachtungen über Anatomie und Entwicklungsgeschichte wirbelloser*

- Thiere an der Küste von Normandie angestellt*, vol. 119. Leipzig: Verlag von Wilhelm Engelmann.
- Dal Zotto M and Todaro MA** (2016) Kinorhyncha from Italy, a revision of the current checklist and an account of the recent investigations. *Zoologischer Anzeiger* **265**, 90–107.
- Fujimoto S** (2015) *Quisarcus Yasumurai* gen. et sp. nov. (Arthrotardigrada: Halechiniscidae) from a submarine cave, off Iejima, Ryukyu Islands, Japan. *Zootaxa* **3948**, 145–450.
- Fujimoto S and Jimi N** (2020) A new marine tardigrade genus and species (Arthrotardigrada, Styraconyxidae) with unique pockets on the legs. *Zoosystematics and Evolution* **96**, 115–122.
- Fujimoto S, Jørgensen A and Hansen JG** (2017) A molecular approach to arthrotardigrade phylogeny (Heterotardigrada, Tardigrada). *Zoologica Scripta* **46**, 496–505.
- Giere O** (2009) *Meiobenthology. The Microscopic Motile Fauna of Aquatic Sediments*, 2nd Edn. Berlin: Springer Verlag.
- Grzelak K and Sørensen MV** (2018) New species of *Echinoderes* (Kinorhyncha: Cyclorhagida) from Spitsbergen, with additional information about known Arctic species. *Marine Biology Research* **14**, 113–147.
- Hanamura Y and Kase T** (2001) A new species of *Heteromysoides* (Mysidacea: Mysidae) from submarine caves of Okinawa, southwestern Japan, with a key to the world species. *Crustacean Research* **30**, 65–71.
- Herranz M and Leander BS** (2016) Redescription of *Echinoderes ohtsukai* Yamasaki and Kajihara, 2012 and *E. kozloffii* Higgins, 1977 from the north-eastern Pacific coast, including the first report of a potential invasive species of kinorhynch. *Zoologischer Anzeiger* **265**, 108–126.
- Herranz M, Thormar J, Benito J, Sánchez N and Pardos F** (2012) *Meristoderes* gen. nov., a new kinorhynch genus, with the description of two new species and their implications for echinoderid phylogeny (Kinorhyncha: Cyclorhagida, Echinoderidae). *Zoologischer Anzeiger* **251**, 161–179.
- Higgins RP** (1982) Three new species of Kinorhyncha from Bermuda. *Transactions of the American Microscopical Society* **101**, 305–316.
- Higgins RP and Kristensen RM** (1988) Kinorhyncha from Disko Island, West Greenland. *Smithsonian Contributions to Zoology* **458**, 1–55.
- Higgins RP and Thiel H** (1988) *Introduction to the Study of Meiofauna*. Washington, DC: Smithsonian Institution Press.
- Ishii D, Yamasaki H, Uozumi R and Hirose E** (2016) Does the kinorhynch have a hydrophobic body surface? Measurement of the wettability of a meiobenthic metazoan. *Royal Society Open Science* **3**, 160512.
- Kase T and Hayami I** (1992) Unique submarine cave mollusc fauna: composition, origin and adaptation. *Journal of Molluscan Studies* **58**, 446–449.
- Kitamura A, Hiramoto M, Kase T, Yamamoto N, Mariko A and Ohashi S** (2007) Changes in cavernicolous bivalve assemblages and environments within a submarine cave in the Okinawa Islands during the last 5000 years. *Paleontological Research* **11**, 163–182.
- Lou TH** (1934) Sur la présence d'un nouveau kinorhynque à Tchefou: *Echinoderes tchefouensis* sp. nov. *Contributions from the Institute of Zoology. National Academy of Peiping* **1**, 1–9.
- Moldovan OT, Kováč E and Stuart H** (2018) *Cave Ecology*. Cham: Springer.
- Neuhaus B** (2013) 5. Kinorhyncha (=Echinodera). In Schmidt-Rhaesa A (ed.), *Handbook of Zoology. Gastrotricha. Cycloneuralia and Gnathifera. Vol. 1. Nematomorpha. Priapulida. Kinorhyncha. Loricifera*. Berlin: Walter de Gruyter, pp. 177–348.
- Neuhaus B and Sørensen MV** (2013) Populations of *Campyloderes* sp. (Kinorhyncha, Cyclorhagida): one global species with significant morphological variation? *Zoologischer Anzeiger* **252**, 48–75.
- Neuhaus B, Pardos F, Sørensen MV and Higgins RP** (2014) New species of *Centroderes* (Kinorhyncha: Cyclorhagida) from the Northwest Atlantic Ocean, life cycle, and ground pattern of the genus. *Zootaxa* **3901**, 1–69.
- Neves RC, Sørensen MV and Herranz M** (2016) First account on kinorhynchs from Portugal, with the description of two new species: *Echinoderes lusitanicus* sp. nov. and *E. reicherti* sp. nov. *Marine Biology Research* **12**, 455–470.
- Omori A, Kitamura A, Fujita K, Honda K and Yamamoto N** (2010) Reconstruction of light conditions within a submarine cave during the past 7000 years based on the temporal and spatial distribution of algal symbiont-bearing large benthic foraminifers. *Palaeogeography, Palaeoclimatology, Palaeoecology* **292**, 443–452.
- Randsø PV, Yamasaki H, Bownes S, Herranz M, Di Domenico M, Qi GB and Sørensen MV** (2019) Phylogeny of the *Echinoderes coulli*-group (Kinorhyncha; Cyclorhagida, Echinoderidae) – a cosmopolitan species group trapped in the intertidal. *Invertebrate Systematics* **33**, 501–517.
- Riedl R** (1966) *Biologie der Meereshöhlen*. Hamburg: Verlag Paul Parey.
- Sánchez N and Martínez A** (2019) Dungeons and dragons: two new species and records of Kinorhyncha from anchialine cenotes and marine lava tubes. *Zoologischer Anzeiger* **282**, 161–175.
- Sánchez N, Pardos F, Herranz M and Benito J** (2011) *Pycnophyes dolichurus* sp. nov. and *P. aulacodes* sp. nov. (Kinorhyncha, Homalorhagida, Pycnophyidae), two new kinorhynchs from Spain with a reevaluation of homalorhagid taxonomic characters. *Helgolander Marine Research* **65**, 319–334.
- Sánchez N, Herranz M, Benito J and Pardos F** (2012) Kinorhyncha from the Iberian Peninsula: new data from the first intensive sampling campaigns. *Zootaxa* **3402**, 24–44.
- Sánchez N, Pardos F and Martínez Arbizu P** (2019) Deep-sea Kinorhyncha diversity of the polymetallic nodule fields at the Clarion-Clipperton Fracture Zone (CCZ). *Zoologischer Anzeiger* **282**, 88–105.
- Shirayama Y, Kaku T and Higgins RP** (1993) Double-sided microscopic observation of meiofauna using an HS-slide. *Benthos Research* **44**, 41–44.
- Sönmez S, Köroğul NÖ and Karaytuğ S** (2016) First record of the family Echinoderidae Zelinka, 1894 (Kinorhyncha: Cyclorhagida) from Turkish marine waters. *Biharean Biologist* **10**, 8–11.
- Sørensen MV** (2014) First account of echinoderid kinorhynchs from Brazil, with the description of three new species. *Marine Biodiversity* **44**, 251–274.
- Sørensen MV and Pardos F** (2008) Kinorhynch systematics and biology – an introduction to the study of kinorhynchs: inclusive identification keys to the genera. *Meiofauna Marina* **16**, 21–73.
- Sørensen MV, Jørgensen A and Boesgaard TM** (2000) A new *Echinoderes* (Kinorhyncha: Cyclorhagida) from a submarine cave in New South Wales, Australia. *Cahiers de Biologie Marine* **41**, 167–179.
- Sørensen MV, Rho HS, Min WG, Kim D and Chang CY** (2012) An exploration of *Echinoderes* (Kinorhyncha: Cyclorhagida) in Korean and neighboring waters with the description of four new species and a redescription of *E. tchefouensis* Lou, 1934. *Zootaxa* **3368**, 161–196.
- Sørensen MV, Dal Zotto M, Rho HS, Herranz M, Sánchez N, Pardos F and Yamasaki H** (2015) Phylogeny of Kinorhyncha based on morphology and two molecular loci. *PLoS ONE* **10**, e0133440.
- Sørensen MV, Gąsiorowski L, Randsø PV, Sánchez N and Neves RC** (2016) First report of kinorhynchs from Singapore, with the description of three new species. *Raffles Bulletin of Zoology* **64**, 3–27.
- Sørensen MV, Rohal M and Thistle D** (2018) Deep-sea Echinoderidae (Kinorhyncha: Cyclorhagida) from the Northwest Pacific. *European Journal of Taxonomy* **456**, 1–75.
- Southern R** (1914) Clare Island Survey. Nematelmia, Kinorhyncha, and Chaetognatha. *Proceedings of the Royal Irish Academy* **31**, 1–80.
- Tabuki R and Hanai T** (1999) A new sigillid ostracod from submarine caves of the Ryukyu Islands, Japan. *Palaeontology* **42**, 596–593.
- Uozumi R, Yamasaki H and Hirose E** (2018) Mangrove forests may serve as stable environments for the meiobenthic *Echinoderes komatsui* (Kinorhyncha: Cyclorhagida): distribution patterns and population dynamics in a subtropical estuary. *Marine Biology Research* **14**, 321–333.
- Varney RM, Funch P, Kocot KM and Sørensen MV** (2019) A new species of *Echinoderes* (Cyclorhagida: Echinoderidae) from the San Juan Islands, Washington State, USA, and insights into the kinorhynch transcriptome. *Zoologischer Anzeiger* **282**, 52–63.
- Wessel P, Smith WHF, Scharroo R, Luis J and Wobbe F** (2013) Generic mapping tools: improved version released. *Eos, Transactions American Geophysical Union* **94**, 409–410.
- Yamamoto N, Kitamura A, Irino T, Kase T and Ohashi S** (2010) Climatic and hydrologic variability in the East China Sea during the last 7000 years based on oxygen isotope records of the submarine cavernicolous micro-bivalve *Carditella iejimensis*. *Global and Planetary Change* **72**, 131–140.
- Yamasaki H** (2016) *Ryugoderes iejimensis*, a new genus and species of Campyloderidae (Xenosomata: Cyclorhagida: Kinorhyncha) from a submarine cave in the Ryukyu Islands, Japan. *Zoologischer Anzeiger* **265**, 69–79.
- Yamasaki H and Kajihara H** (2012) A new brackish-water species of *Echinoderes* (Kinorhyncha: Cyclorhagida) from the Seto Inland Sea, Japan. *Species Diversity* **17**, 109–118.
- Yamasaki H, Hiruta SF, Kajihara H and Dick MH** (2014) Two kinorhynch species (Cyclorhagida, Echinoderidae, *Echinoderes*) show different distribution patterns across Tsugaru Strait, northern Japan. *Zoological Science* **31**, 421–429.

- Yamasaki H, Grzelak K, Sørensen MV, Neuhaus B and George KH (2018a)** *Echinoderes pterus* sp. n. showing a geographically and bathymetrically wide distribution pattern on seamounts and on the deep-sea floor in the Arctic Ocean, Atlantic Ocean, and the Mediterranean Sea (Kinorhyncha, Cyclorhagida). *Zookeys* **771**, 15–40.
- Yamasaki H, Neuhaus B and George KH (2018b)** New species of *Echinoderes* (Kinorhyncha: Cyclorhagida) from Mediterranean seamounts and from the deep-sea floor in the Northeast Atlantic Ocean, including notes on two undescribed species. *Zootaxa* **4387**, 541–566.
- Yamasaki H, Neuhaus B and George KH (2019)** Echinoderid mud dragons (Cyclorhagida: Kinorhyncha) from Senghor Seamount (NE Atlantic Ocean) including general discussion of faunistic characters and distribution patterns of seamount kinorhynchs. *Zoologischer Anzeiger* **282**, 64–87.
- Zelinka C (1894)** Über die Organisation von Echinoderes. *Verhandlungen der Deutschen Zoologischen Gesellschaft* **4**, 46–49.
- Zelinka C (1896)** Demonstration der Tafeln der *Echinoderes*-Monographie. *Verhandlungen der Deutschen Zoologischen Gesellschaft* **6**, 197–199.