# Cartilage catabolism in arthritis: factors that influence homeostasis

# Andrew D. Rowan

Preventing the destruction of articular cartilage has long been a goal in the treatment of arthritic diseases, in which a combination of cytokines and growth factors affect the catabolic state of cells within the joint. Normal tissue turnover can be viewed as a balance between degradation and synthesis of the macromolecules that constitute the extracellular matrix. This process is tightly regulated such that highly degradative proteinases are controlled at several levels, including synthesis and secretion, activation and inhibition. Tissue destruction occurs when proteinase-mediated degradation exceeds synthesis, and this is markedly influenced by cytokines and growth factors that stimulate matrix synthesis as well as the production of proteinases and/or their endogenous inhibitors. This review outlines current knowledge of factors that influence cartilage biology, with particular emphasis on chondrocytes and synovial fibroblasts. Recent findings from the delivery of cytokines to affected tissues are also summarised, and the potential impact these observations might have on new therapies for arthritic diseases is discussed.

The goal of generating new and improved treatments for arthritic diseases of the joints has not yet been achieved, although progress is being made. Destruction of articular cartilage is the hallmark of many of these disabling conditions, and an imbalance of pro-inflammatory cytokines over their anti-inflammatory counterparts promotes the disease process. Therefore, an understanding of the effects of the various cytokines and growth factors present during the disease process on the cells and tissues in the joint will greatly assist the development of new therapies. Since tissue destruction is considered the 'end-point' of the disease process, knowledge of the enzymes involved in this destruction is also paramount in new drug design. This review highlights the effects of cytokines and growth factors on cartilage chondrocytes and synovial fibroblasts with respect to cartilage matrix synthesis and repair, and its degradation.

# Articular cartilage

Articular cartilage provides a friction-free surface on which the bones of a joint articulate during motion. The joints in the lower body have to support the body weight even at rest. To fulfil these tasks, articular cartilage must be able to withstand the compressive forces that the joint experiences. Two major macromolecules help to provide this function: proteoglycan and

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collagen. Proteoglycan, of which aggrecan is the predominant monomer, has a high sulphated glycosaminoglycan content that attracts water molecules and hence allows the tissue to swell and resist compressive forces (Ref. 1). These proteoglycans are held in the tissue within a fibrous network of triple-helical type II collagen fibrils, which are crosslinked and provide cartilage with its tensile strength. Together, these macromolecules make up ~25% of the wet weight of cartilage, with water accounting for most of the remainder. Other molecules such as minor collagens (types VI, IX, X and XI) (Ref. 2), biglycan, decorin, laminin, tenascin and fibromodulin all assist in maintaining the tissue, through spatially organising components of the extracellular matrix (ECM) (Refs 1, 2, 3). One component of special note is cartilage oligomeric protein (COMP), which is a member of the thrombospondin family of proteins. COMP is present in high amounts in articular cartilage and its breakdown products have been suggested to be a marker for disease. A role for the matrix metalloproteinases (MMPs), as well as serine proteinases, has been demonstrated in this breakdown (Ref. 4). Although its true function remains unclear, COMP is known to interact with collagen, and given its inter-territorial localisation in mature cartilage, a structural role seems probable. This is further supported by the fact that mutations in COMP give rise to structural defects.

Cartilage receives nutrients from the synovial fluid, which is synthesised by the fibroblasts in the synovial membrane. It is a dynamic tissue that is constantly being remodelled as old macromolecules of the ECM are replaced with new. This matrix is maintained and replaced by the resident chondrocytes, which only sparsely populate the tissue. Adequate maintenance of cartilage is essential for chondrocyte viability and, conversely, loss of chondrocytes adversely affects the tissue. Such loss, via apoptosis, is known to be induced by nitric oxide (NO), which itself is produced by pro-inflammatory stimuli such as interleukin 1 (IL-1) and tumour necrosis factor  $\alpha$  (TNF- $\alpha$ ) (Ref. 5). These observations extend to osteoarthritis (OA) cartilage, where evidence of apoptosis has been found in abnormal tissue, and a significant correlation between apoptotic cell numbers and proteoglycan depletion (with elevated NO levels) has been shown (Ref. 5); activation of the Fas receptor is thought to be the apoptotic trigger (see Ref. 5 and references therein). A feedback loop is therefore created whereby a healthy matrix supports viable chondrocytes and vice versa and, by contrast, loss of chondrocytes through apoptosis favours deterioration of the ECM.

#### Arthritic disease

Rheumatoid arthritis (RA) is the most common inflammatory arthropathy, having a disease prevalence of 0.5–1.0%. It has a peak age of onset in the sixth decade and is three times more common in women. Although OA is generally more benign than RA, clinical OA of the knee and hips affects 10–20% of the over 65s, and again is more common in women. The combined costs to the UK NHS Executive are £560 million, although the overall cost in the UK is in excess of £1 billion (see Ref. 6 and references therein).

The term RA was coined in 1859 by Sir Alfred Garrod. He was the first to distinguish gout from RA, although OA was still included in the RA heading. The term OA was subsequently used in 1888, but it was not until 1907 that Garrod's son finally made the modern-day distinction between RA and OA. As clinical diagnosis has progressed, specific criteria have begun to be assigned to these diseases, although RA and OA are often still referred to as an inflammatory autoimmune disease and an erosive disease associated with old age, respectively. This classification is obviously over-simplified and some patients might present in the clinic with a combination of both diseases. Controversy also exists as to the precise nature of the initiating factor(s) underlying these diseases, as well as the enzymes that play a major role in the destructive process. A vast amount of research has been undertaken to try to unravel some of these fundamental questions but, although many advances have been made, they remain relatively unanswered. This review focuses on RA but, where possible, information on OA is also documented for comparison.

## **Rheumatoid arthritis**

RA is a complex disease and, although the initiating trigger is poorly understood, the disease is considered to have an autoimmune basis. Antigens suggested to be the target of autoimmunity include type II collagen, heat shock proteins and gp39 (Ref. 7). A genetic predisposition to RA has been demonstrated, with an increased susceptibility locus being

linked to expression of specific HLA-DR4 subtypes (Ref. 8) (see later section entitled 'Other factors'). One widely accepted hypothesis has been that RA is a disease driven primarily by the synovium, which undergoes proliferation to generate the 'pannus' that then invades articular cartilage and promotes its destruction (Ref. 9). At this stage of disease, the synovitis may subside only to reappear later; this cycle of inflammation followed by remission is typical of many RA patients, in whom damage to the cartilage is thought to occur during inflammatory episodes.

Maintenance of the invasive pannus tissue is an integral part of disease progression. The potent angiogenic cytokine vascular endothelial growth factor (VEGF) appears to play a role in this process (Ref. 10). Invasion is reduced by anti-TNF- $\alpha$  therapy; this might result from blockade of a cytokine cascade or a reduction in leukocyte trafficking into the joint (Ref. 11). Synovial invasion of the cartilage is associated with the plasminogen activation system (Ref. 12), and is also a phase in disease when synovial fluid neutrophils and chondrocytes within the cartilage might begin to respond to the predominantly pro-inflammatory stimuli (Ref. 13).

Recent evidence has indicated that tissue destruction and inflammation might not in fact be coupled (Ref. 14), and therefore that sustained degradation of the cartilage ECM might be a later consequence of the inflammation via stimulation of the resident chondrocytes. A variety of pro-inflammatory stimuli are known to induce chemokine production, including monocyte chemoattractant protein 1 (MCP-1), in chondrocytes, which might provide a mechanism by which cartilage actively promotes disease progression (Ref. 15). It is also unclear why activation of synovial fibroblasts persists even after anti-inflammatory therapy. One possibility is that the synovial membrane is gradually repopulated with immature mesenchymal and bone marrow cells with altered properties. This hypothesis is supported by work that has shown elevated expression of the human embryonic growth factor homologues of wingless and frizzled in RA fibroblasts compared with OA and normal cells, suggesting that the synovium is indeed repopulated with immature cells (Ref. 16). Alternatively, existing cells might undergo (de)differentiation in some way such that they then express these embryonic markers.

# Osteoarthritis

OA falls into two categories: primary OA occurs in middle-aged to elderly patients where an active disease process is often presumed to be a consequence of joint 'wear and tear'; secondary OA occurs at any age as a result of trauma or disease. In both situations, the central feature is loss of articular cartilage and a reduced capacity for repair; the chondrocytes themselves appear to be the driving force behind these deficiencies. A focal lesion in the cartilage might lead to abnormal loading of the surrounding chondrocytes, which, in turn, respond by promoting a cascade of slow but persistent degradation of cartilage, ultimately leading to loss of joint function. The cyclical disease course of OA has also been proposed to be the result of sequential cytokine stimulation followed by a feedback inhibition of autocrine cytokine and cytokine receptor production, which affects collagenase synthesis (Ref. 17). Although not considered an inflammatory disease, orthopaedic surgeons often comment on the marked synovial infiltration seen during joint replacement surgery of OA patients, further increasing the evidence for the involvement of pro-inflammatory cytokines such as IL-1 and TNF-α (Ref. 18).

## Matrix metabolism

Degradation and synthesis of cartilage macromolecules under normal physiological conditions is kept in equilibrium and can therefore be viewed as a balance (see Fig. 1). Chondrocytes and synovial cells respond to a variety of cytokines and growth factors that stimulate the production of destructive proteinases. All four major classes of proteolytic enzymes (aspartic, cysteine, serine and metallo) are involved in normal turnover and pathological destruction, and the pathway that predominates will alter depending on the resorptive circumstances (Ref. 19). These pathways are not mutually exclusive and it is highly probable that total degradation of matrix components involves several pathways and classes of proteinases.

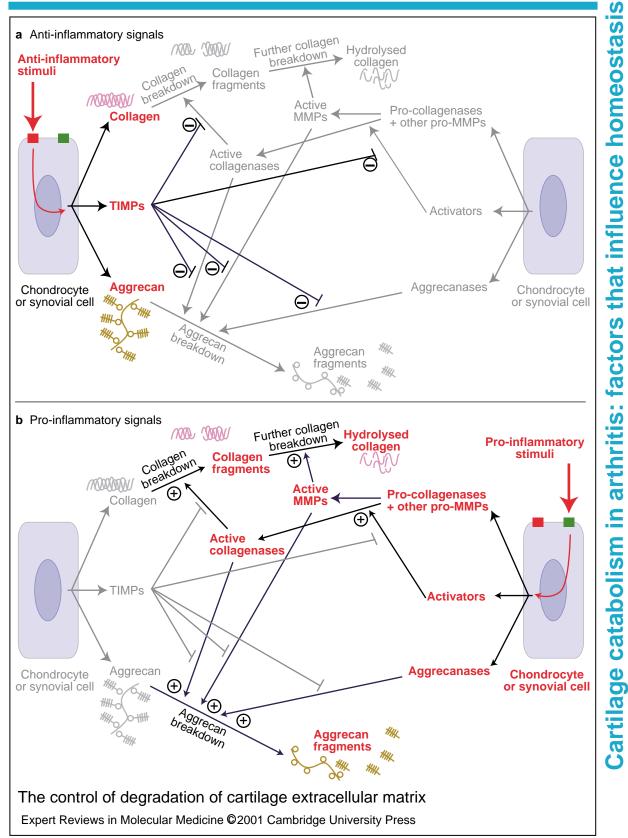
MMPs are a family of neutral zinc endoproteinases that collectively degrade all the components of the ECM (Ref. 20), and have received considerable attention with respect to arthritic tissue destruction because their expression correlates strongly with collagen degradation (Refs 21, 22), although this is not always the case (Ref. 23). The MMP family

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**Figure 1. The control of degradation of cartilage extracellular matrix.** Chondrocytes maintain articular cartilage by replacing degraded components via local synthesis, which is balanced with degradation to prevent over-deposition of matrix. In arthritic diseases, this equilibrium is shifted towards degradation. Chondrocytes and synovial cells are stimulated by both (a) anti-inflammatory and (b) pro-inflammatory cytokines, as well as mechanical stress, and cell–cell and cell–matrix contacts, via a variety of cell-surface receptors. These stimuli are transferred to the nucleus via intracellular signalling and mechanotransduction pathways, resulting in activation of gene transcription. Synthesis and secretion of matrix components, including aggrecan and collagen occurs, as well as enzymes such as aggrecanases (e.g. ADAMTS-5), pro-matrix metalloproteinases (pro-MMPs) (including procollagenases) and activating enzymes (e.g. MMP-3, membrane-type MMPs and plasmin). Aggrecanases promote rapid aggrecan loss, and this might be inhibited by tissue inhibitor of metalloproteinase 3 (TIMP-3). Induction of TIMPs, and other inhibitors, via anti-inflammatory agents can block some activating enzymes that otherwise convert pro-enzymes to their active forms. These activating enzymes might in turn also require activation. Once the level of active MMPs exceeds the local supply of TIMPs, they can promote specific cleavage of triple-helical collagen via collagenases, as well as further nonspecific collagen hydrolysis by other MMPs; aggrecan degradation by MMPs might also occur (fig001arn).

contains at least three collagenases [interstitial collagenase / collagenase 1 (MMP-1), neutrophil collagenase (MMP-8), and collagenase 3 (MMP-13)] that can degrade fibrillar collagen (Ref. 20). MMPs are controlled at several levels (Ref. 24), including inhibition by a family of endogenous inhibitors called the tissue inhibitors of metalloproteinases (TIMPs) (Refs 24, 25, 26). MMP inhibition has been the subject of much research in recent years by the pharmaceutical industry in an attempt to find a highly potent, specific and bioavailable inhibitor (Ref. 26). Other metalloproteinases, most notably of the ADAM (for 'a disintegrin and metalloproteinase') family of proteinases, are also expressed in cartilage (Ref. 27), although their roles in tissue maintenance and tissue destruction are still unclear. In particular, a subset of this family known as the ADAMTS (ADAM with thrombospondin motifs) proteins contains members that have recently been found to specifically cleave aggrecan at the site considered to be pathologically relevant (Refs 28, 29, 30). At least one of these enzymes (ADAMTS-5) is also expressed in the synovium (Ref. 31). Although these metalloproteinases are not MMPs, recent evidence indicates that TIMP-3 is a potent inhibitor (Ref. 32).

Cysteine proteinases (cathepsins B and L, which can degrade cartilage and bone) are expressed in RA synovial lining (Ref. 33), and have been implicated in cartilage degradation (Ref. 34). Cathepsin K has received recent attention because it is now thought to be strongly associated with pathological bone and cartilage resorption (Refs 35, 36). Serine proteinases, particularly those associated with the plasmin cascade, have also been implicated in tissue destruction (Refs 12, 37).

The role of MMPs in the pathological destruction of cartilage is promoted by various pro-inflammatory cytokines that perturb the balance between synthesis and degradation of ECM components to favour matrix breakdown (see below and also Fig. 1). Proteoglycan (i.e. aggrecan) loss is a rapid event following proinflammatory stimulation but it can be readily replaced once the stimulus is removed. Collagen is more resistant to degradation but is much more difficult to replace (see Ref. 24 and references therein). However, a different situation arises during normal cartilage metabolism (the socalled physiological 'steady-state'). Under these circumstances, degradation of collagen, and indeed probably proteoglycan, occurs within the lysosomal system following phagocytosis, a mechanism shown to occur in fibroblasts (Ref. 38). Although phagocytosis has not been described in chondrocytes, CD44-mediated endocytosis and breakdown of hyaluronate has been reported (Ref. 39), suggesting differences in mechanisms of ECM breakdown among mesenchymal cells. Nevertheless, an equilibrium exists whereby matrix turnover is tightly regulated; any disturbance of the various degradative pathways that prevail might lead to uncontrolled matrix destruction.

# Cytokines and cytokine interactions

Cytokines and growth factors have been subdivided into those with pro-inflammatory and those with anti-inflammatory effects; in arthritis, these generally exhibit either catabolic or anabolic effects on cartilage, respectively. Considerable attention has been focused on IL-1 and TNF- $\alpha$ – two pro-inflammatory cytokines that have been regarded as pivotal mediators in inflammatory

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diseases such as RA. However, several other cytokines present in the milieu might also be associated with active disease. The most abundant include IL-1, -6, -8, -10, -11, -13, -17 and -18, TNF- $\alpha$ , granulocyte–macrophage colony-stimulating factor (GM-CSF) and transforming growth factor  $\beta$  (TGF- $\beta$ ). Less-detectable mediators include IL-2, -3, -12 and -15, interferon  $\gamma$  (IFN- $\gamma$ ), oncostatin M (OSM) and leukaemia inhibitory factor (LIF). IL-4 is rarely detected.

It is important to remember that cytokines are capable of acting in synergy, whereby the combined effects of two or more cytokines far exceeds the effects they exert alone. This issue cannot be underestimated, and might occur inadvertently under certain conditions. For example, insulin-like growth factor 1 (IGF-1) has been shown to synergise with TGF- $\beta$  to induce matrix synthesis (Ref. 40); TGF- $\beta$  has also been shown to augment prostaglandin E, (PGE,) production in cultured chondrocytes when in the presence of serum (Ref. 41), a result that is most likely explained by the presence of IGF-1 in serum. Hence, culture conditions can significantly alter the responses to cytokines in vitro and careful consideration is required to ensure culture conditions are well defined.

Several antagonist factors might reduce the effects of a given cytokine on cartilage. For example, the effects of IL-1 can be reduced by IL-1 receptor antagonist (IL-1RA), which is a soluble factor that binds IL-1 and prevents it from binding to its cell-surface receptor. This might represent the mechanism of action of the chondroprotective cytokine IL-4, which is known to upregulate IL-1RA (Ref. 42). Many cells also express the IL-1 'decoy' receptor (IL-1RII). It is thought that binding of IL-1 by this receptor does not lead to signal transduction, although this has been challenged recently (Ref. 43) and will require further investigation in the context of experimental data from animal models. There are also various other soluble receptors, which can act as agonists and antagonists [e.g. soluble IL-6 receptor (sIL-6R) and soluble TNF- $\alpha$  receptor (sTNFR), respectively]. These soluble factors are important to consider because they can arise from tissues remote from the cartilage but might have a profound effect on the chondrocytes.

#### **Pro-inflammatory cytokines**

A hierarchy of pro-inflammatory cytokines has been proposed to exist in arthritic diseases, expert reviews

whereby IL-1 is the pivotal cytokine at early and late stages of disease and TNF- $\alpha$  is involved in disease onset (Ref. 44). Strong arguments have been proposed for IL-1 as the primary mediator in light of the IL-1 $\beta$ -knockout mouse, which lacks chronic erosive arthritis (Ref. 45). However, other cytokines have also been proposed to be key players, such as IL-15 (Ref. 46). In addition, recent evidence suggests that the T-cell-specific cytokine IL-17 might be the key mediator because it is produced very early in the disease process and is known to induce the production of other pro-inflammatory cytokines (Ref. 47). It is most probable that more than one of these proinflammatory cytokines are present in an arthritic joint (e.g. Ref. 48) and that they have varying effects on the disease process, including inhibition of matrix synthesis (e.g. Refs 49, 50), upregulation of matrix-degrading enzymes (e.g. Refs 50, 51, 52, 53, 54), and induction of other pro-inflammatory agents (e.g. Refs 17, 46, 50, 52, 54, 55, 56). This concept of multiple cytokines, perhaps acting synergistically, in arthritic disease has prompted the suggestion that fully effective anti-cytokine treatment will most probably be achieved using a combination therapy (Ref. 57).

Large amounts of NO are produced by chondrocytes activated by pro-inflammatory cytokines (see Refs 5, 58 and references therein). Many studies have shown that NO is at least partly responsible for IL-1-induced effects such as suppressed collagen synthesis. NO production is a consistent feature of arthritic cartilage and synovium and has been associated with matrix degradation and chondrocyte apoptosis. Indeed, inhibitors of NO synthase, which produces NO, help reduce synovial inflammation and result in decreased destruction of cartilage and bone in experimental models of arthritis (Ref. 58).

In general, pro-inflammatory cytokines are potent mediators and exert their maximal effects on cells at relatively low concentrations. This is an important point, especially when considering cell culture experiments in comparison with active disease where, for example, synovial invasion occurs. The localised concentration of a given cytokine at this invasion front might be considerably higher than in the remaining cartilage, and therefore sufficient to affect the local chondrocyte population. Marked differences in cell numbers often occur when comparing cell and tissue culture experiments, such that responses observed in cell culture at a given

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time point might not always be seen in tissue culture.

#### Anti-inflammatory cytokines

The anti-inflammatory cytokines typically counteract the effects of their pro-inflammatory counterparts and often promote the synthesis of ECM components. Many of these cytokines favour the differentiation of T cells into T helper 2 (Th2) cells rather than the pro-inflammatory Th1 cells. Differentiation of T cells along the Th2 lineage is generally thought to be a more protective mode of the inflammatory response, owing to their antiinflammatory 'phenotype'. The effects of antiinflammatory cytokines include the promotion of matrix synthesis and repair (e.g. Refs 40, 59, 60, 61, 62, 63), induction of protective enzyme inhibitors such as TIMPs (e.g. Refs 64, 65, 66), downregulation of destructive enzymes (e.g. Refs 53, 56, 58, 59, 60), and reduction in the levels of pro-inflammatory cytokines (e.g. Refs 40, 44, 67).

TGF- $\beta$  has a major role in chondroprotective mechanisms and is known to promote matrix synthesis. Cartilage contains vast stores of TGF- $\beta$ , as much as 300–500 ng g<sup>-1</sup> of cartilage, with most of it in the latent, inactive form that requires proteolytic processing (Ref. 68). Various hypotheses have tried to explain the physiological significance of this large excess, and although the mechanism of TGF- $\beta$  autocrine regulation is still not understood, the presence of such reserves strongly implies a role in cartilage metabolism and a key role for TGF- $\beta$  activation. A recent report, however, has indicated that TGF- $\beta$  might also cause synovial hyperplasia and osteophyte formation (Ref. 69).

Also present at elevated levels in cartilage is IGF-1, which appears to have a role in stimulating chondrocyte anabolic activity. This growth factor is sequestered from its cell-surface receptor in the ECM by IGF-binding proteins (IGFBPs) and fibronectin (Ref. 70); this complex system is termed the IGF-1 axis and tightly regulates the bioavailability of IGF-1. IGFBP-3 expression appears to increase with age, paralleling an agerelated decline in matrix synthesis, and is also overexpressed in OA cartilage where it could cause metabolic disturbances (Ref. 70). The local increase in matrix synthesis following injury could result from damage-induced IGF-1 release from IGFBP pools. An age-related failure in this system could thus contribute to degenerative disease, and gives some credence to the belief that OA is due

to the 'wear and tear' of old age. The protective effects of IGF-1 have been further demonstrated using adenoviral vectors that specifically result in its overexpression (see below).

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Recent evidence indicates that some of the cytokines that have been described as antiinflammatory might not always be so. Members of the IL-6 cytokine family, including IL-6 and OSM, are known to induce TIMP production from chondrocytes and fibroblasts, as well as increase matrix synthesis. These abilities would favour a chondroprotective role for these cytokines, although recent evidence now indicates that this is not always the case (Refs 51, 54, 71). Rather, when in combination with IL-1 these cytokines induced a marked catabolic effect by promoting matrix degradation. A protective role for IL-4 has been described in arthritis (Refs 72, 73), although it is also implicated in asthma.

#### *Links between chondrocytes and matrix*

Interactions between chondrocytes and the ECM help to regulate many biological processes important to cartilage homeostasis and repair, including cell attachment, differentiation, growth and survival. The integrin family of cell-surface receptors plays an important role in mediating cell-matrix interactions. Several integrins are expressed on chondrocytes and these serve as receptors for collagens type II and VI ( $\alpha_1\beta_1, \alpha_2\beta_1$ ,  $\alpha_{10}\beta_1$ ), vitronectin and osteopontin ( $\alpha_v\beta_3$ ), laminin  $(\alpha_{5}\beta_{1})$  and fibronectin  $(\alpha_{5}\beta_{1})$  (Ref. 74). Integrins might be important transducers of mechanical stimuli because they provide a link between the cytoskeleton and the ECM. Such links are critical, for example, for relaying shear-stress-induced signals (Ref. 75). Expression of integrins can be regulated by IGF-1 and TGF- $\beta$  (see Ref. 74 and references therein). Cross-talk between the integrin signalling mechanisms has recently been demonstrated and is thought to play a role in cartilage homeostasis (Ref. 76). Indeed, an  $\alpha_{v}\beta_{2}$  antagonist has been shown to reduce inflammation and improve joint integrity in an experimental arthritis model (Ref. 77). Moreover, deregulation of this integrin network might be important in disease progression.

Cell-surface receptor binding is the primary means by which cytokines and growth factors mediate their effects upon cells. In many cases, very limited amounts of cytokine are required to elicit a response. A maximal response to IL-1 in chondrocytes can be achieved when as few as 40 cell-surface receptors bind ligand (Ref. 78). Observations such as this clearly suggest that modulation of the number of cell-surface receptors would not have a marked effect on cellular responses. However, where cell-surface receptor expression is limiting, then an increase in expression might alter the responsiveness of a given cell type. This is evident for the TNFRs (which are expressed as two different forms: TNFR55 and TNFR75); normal chondrocytes express mainly TNFR55, but pro-inflammatory cytokines such as TNF- $\alpha$  induce the expression of TNFR75 (Ref. 79). In OA cartilage, differences in TNFR distribution have been found and a role for TNF- $\alpha$  in focal cartilage loss has been reported (Ref. 80); this role is further supported by evidence that levels of TNF- $\alpha$  and TNF- $\alpha$ -converting enzyme (TACE or ADAM-17) are elevated in OA cartilage, suggesting a paracrine/autocrine loop (Ref. 81). A role for CD44, the hyaluronan receptor, on chondrocytes has been demonstrated in the maintenance of cartilage homeostasis (Ref. 82); in particular, this might have implications for aggrecan turnover.

Chondrocytes are able to respond to a wide variety of stimuli, suggesting that they have an extensive repertoire of receptors. When a specific receptor is absent, responsiveness might be conferred by the presence of a soluble receptor. For example, sIL-6R confers IL-6 responsiveness to chondrocytes (Ref. 83), which normally possess gp130, the common signalling subunit for IL-6type cytokines (Ref. 54). Membrane IL-6R lacks the cytoplasmic signalling domain and hence does not contribute to signal transduction other than by binding to gp130 once it has bound IL-6, as is the case for sIL-6R. Such findings might be relevant to RA since sIL-6R is elevated in RA synovial fluids (Ref. 84) and is known to be produced by the synovium.

# Other factors

Several studies have indicated a small but consistent increased risk of arthritic disease in siblings of affected individuals (Ref. 85), although identical twins show little similarity in the timing of disease onset. However, twin studies do support the notion that genetic factors contribute to increased disease risk: monozygotic and dizygotic twins have a disease concordance for RA of 12% and 4%, respectively, which is well above the background disease prevalence of 0.5–1.0% (see Ref. 85 and references cited therein). Polymorphisms in several genes associated with destructive joint diseases have been identified that might contribute to disease pathogenesis, such as an IL-6 polymorphism associated with juvenile chronic arthritis (Ref. 86). Ongoing studies of a polymorphism in the MMP-1 promoter (already shown to correlate with tumour invasion in cancer patients) might shed new light on the role of this proteinase in arthritic disease.

Autoimmunity is also thought to be a dominant factor in the pathogenesis of RA, and autoantibodies to cartilage-specific components have been detected in serum samples of RA patients (Ref. 87). Complement activation occurs adjacent to the cartilage surface, where abundant co-deposits of immunoglobulin and activated complement components such as C1s are present. This component has several novel functions (see Ref. 88), and is thought to participate in the pathogenesis of RA through its collagenolytic activity (Ref. 89), in addition to its role in the complement cascade. A genetic predisposition for RA has been identified in the major histocompatibility complex (MHC) that forms the human leukocyte antigen (HLA) types (see Ref. 8 and references therein). Interactions between antigenic peptide and an HLA molecule determine the immune response, and the HLA molecules associated with RA encode a sequence of amino acids that constitute the 'shared RA epitope'. This epitope has been shown to indicate an increased susceptibility for more-aggressive, erosive RA. Several models have been proposed for how this epitope could lead to antigen-specific T-cell-mediated autoimmunity (see Ref. 8 and references therein).

# Gene regulation in cartilage metabolism

Gene regulation through cytokine stimulation is mediated via specific signal-transduction pathways, an area of research that has prompted the search for novel therapeutic targets for many diseases, including inflammatory arthritis. Several transcription factor families, including AP-1, NF-kB and STAT, have been implicated in gene regulation in inflammation (see Ref. 90 and references cited therein). Moreover, the activity of these transcription factors is regulated by mitogen-activated protein kinase (MAPK) pathways. Cross-talk and/or interplay between these pathways might contribute to increased transcription of some catabolic genes (e.g. Ref. 91).

This level of complexity is compounded when combinations of cytokines appear to have synergistic effects on gene transcription, and this can be problematic when trying to reproduce in vivo effects in vitro.

The advent of potent synthetic inhibitors of factors involved in a variety of signal-transduction pathways has begun to allow a more detailed analysis of the specific pathways that are involved in the regulation of genes considered to be important in the disease process. This applies not only to the degradative enzymes such as the collagenases but also to their inhibitors such as the TIMPs (e.g. Ref. 92). Furthermore, the identification of endogenous regulators of transcription factors such as those for STAT proteins has also allowed more detailed analyses of specific pathways that are utilised in the transcriptional activation of certain genes (e.g. Ref. 93).

# Genetic approaches to understanding arthritis

Although pro-inflammatory cytokines and the MMP family of enzymes have been strongly implicated in the pathological destruction of cartilage, considerable controversy still exists as to the identity of a specific cytokine(s) and MMP(s) that are involved in the disease process. In particular, the identity of the collagenase responsible for the destructive turnover of cartilage collagen has been much debated since collagenase 3 (MMP-13) was discovered (Ref. 94). This finding led to a reappraisal of the observations made using murine models of arthritis because rodents appeared to lack the homologue of human collagenase 1 (MMP-1); such a homologue has, however, now been described (Ref. 95). In humans, it is likely that all three collagenases are elevated at some stage in the disease process but that their regulation is not co-ordinated. To date, few studies have assessed multiple MMP expression in diseased tissues, although those that have clearly identify MMP-1 and MMP-13 as being strongly associated with RA synovitis (e.g. Ref. 96).

# **Transgenic studies**

To address some of these issues, transgenic animals have been developed to either overexpress or 'knock-out' expression of MMPs and various cytokines to identify their impact (Table 1). A role for both IL-1 and TNF- $\alpha$  has been demonstrated in these studies. Several MMP-transgenic mice have been developed and interestingly no MMP knockout has been lethal; indeed, several of the knockouts have phenotypes similar to the wild-type animals, suggesting a degree of redundancy within the MMP system. However, one transgenic study has suggested that MMP activation might be a key step in cartilage erosion (Ref. 97).

The major findings relevant to arthritic disease include the following. First, aggrecan cleavage at the 'aggrecanase' site still occurred in stromelysin 1 (MMP-3) knockout mice (Ref. Infl 98), clearly implicating the recently identified aggrecanase (ADAMTS) enzymes (Refs 28, 29, 52) in the specific cleavage of aggrecan; furthermore, in less-aggressive models of arthritis, fewer erosions were found in the MMP-3 knockout mice, suggesting a role for MMP-3 in collagenase activation. Second, the MMP-9 knockout had no obvious phenotypic defects, although there was a delay in long bone growth (Ref. 99). Perturbed vascularisation and ossification of the growth plate appeared to account for the MMP-9 knockout effects, indicating an important developmental role for this enzyme. Third, the MT1-MMP (MMP-14) knockout mice developed dwarfism, osteopaenia and a spontaneous arthritis (Ref. 100), and had severe defects in skeletal development and angiogenesis (Ref. 101). The shortening of bones in these mice is a consequence of decreased chondrocyte proliferation in the proliferative zone of the growth plates. Defective vascular invasion of cartilage promotes enlargement of the hypertrophic zones of the growth plate, which delays the formation of secondary ossification centers. Activation of latent gelatinase A (MMP-2) was also deficient, suggesting that MMP-14 is essential for its activation in vivo. It is unclear whether a nutritional defect might be the cause of some of these observations.

Other genes relevant to cartilage that have been targeted in knockout studies include ADAM-17 and ADAMTS-1, although experiments to assess the effects on cartilage remain to be reported. Loss of ADAM-17, the enzyme responsible for releasing TNF- $\alpha$  from the cell surface, results in mice with respiratory distress and abnormal lung development, implicating this enzyme in lung morphogenesis (Ref. 102). ADAMTS-1-null mice have growth retardation and impaired female fertility (Ref. 103).

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Gene	Genotype <sup>a</sup>	Findings	Refs
L-1α	0	Full signs of destructive arthritis	112
L-1β	К	Normal inflammatory but defective acute-phase responses; resistance to CIA and protection against cartilage destruction in streptococcal-cell-wall arthritis	113, 114
ΓNF-α	0	Chronic arthritis (inhibited by antibodies to IL-1RI)	108
ſNFRI	К	Milder CIA at a reduced incidence; however, once the joint is affected, the disease progresses to the same end-stage as in wild-type mice	115
L-6	К	Inflammatory cell infiltration in knee joints diminishes, but cartilage proteoglycan loss is enhanced during the early acute phase	116
		Only a transient inflammation during AIA, and no chronic synovitis in zymosan-induced arthritis	117
EGF	0	Growth retardation and reduced IGFBP3 levels; accumulation of pre-hypertrophic chondrocytes in the growth plate and abnormal osteoblast proliferation	118
FN-γR	К	CIA occurs earlier than in wild-type mice	119
FcR	К	Joint swelling reduced in AIA although sustained inflammation evident; chondrocyte death and matrix erosion absent	97
		Inflammation and cartilage destruction prevented	120

Abbreviations: AIA, antigen-induced arthritis; CIA, collagen-induced arthritis; EGF, epidermal growth factor; FcR, Fc receptor; IFN- $\gamma$ R, interferon  $\gamma$  receptor; IGFBP-3, insulin-like growth factor binding protein 3; IL-1, interleukin 1; IL-6, interleukin 6; IL-1RI, type I IL-1 receptor; TNF- $\alpha$ , tumour necrosis factor  $\alpha$ ; TNFRI, TNF- $\alpha$  receptor I (TNFR55).

# Gene therapy in arthritis

Another approach that has begun to receive much attention recently is that of adenoviral overexpression of (anti-inflammatory) cytokine genes in the joint to study the effects of individual cytokines on cartilage homeostasis and to ameliorate disease (Table 2). The viruses are attenuated so as to be infectious but unable to replicate. Recombinant adeno-associated viral (rAAV) vectors might be advantageous for in vivo gene therapy for arthritis because of their ability to transduce both fibroblasts and chondrocytes (Ref. 104). These studies have confirmed the protective nature of TGF-β and IGF-1, which has prompted the notion that such vectors could be used in the targeted repair of cartilage injury (Refs 59, 60). However, caution should also be used

when interpreting such studies with a view to treating human disease. Another study using TGF- $\beta$  recently reported synovial hyperplasia and osteophyte formation (Ref. 69) – findings that cast some doubt on the use of this growth factor as a reparative agent for cartilage damage.

# **Clinical implications**

While researchers have been trying to understand the factors and mechanisms underlying the disease processes, clinicians have been equally busy treating an ever increasing number of patients. This increase in number is partly due to improvements in general medical care, changes in lifestyles and the fact that many populations are increasing in age. Together, these factors are having considerable impact on the long-term

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10010 2.1		s-transfected mediators on experimental arthritis (tab002	
Factor	Cell/tissue targeted	<sup>a</sup> Effect on matrix components	Refs
BMP-2	Chondrocyte	Stimulated proteoglycan synthesis even following IL-1 stimulation	62
Plasmin inhibitor	RA synovial fibroblast	Reduced cartilage matrix degradation; reduced cartilage invasion	12
c-Fos	Chondrocyte <sup>b</sup>	Decreased proteoglycan synthesis and TIMP-1 expression, with an increase in MMP-3; decreased type II collagen expression with elevated MMP-1	121
OSM	Synovium	Increased synovial cell proliferation, with pannus-like appearance and infiltration of mononuclear cells; increased matrix deposition	71
TGF-β1	Chondrocyte	Increased matrix synthesis while maintaining type II collagen phenotype	59
		Restored proteoglycan synthesis following IL-1 stimulation	62
	Cartilage	Increased proteoglycan and collagen synthesis	60
	Synovium	Synovial hyperplasia and chondroosteophyte formation at the cartilage-syonvium junction	69
IGF-1	Cartilage	Induced proteoglycan synthesis but did not have anti- inflammatory or chondroprotective effects	122
		Induced matrix synthesis; maintained long-term chondrocyte phenotype in culture	61
	Chondrocyte	Stimulated proteoglycan type II and collagen synthesis; restored proteoglycan synthesis following IL-1 stimulation	62
iNOS	Chondrocyte	Reduced IGF-1; stimulated proteoglycan synthesis	123
IL-4	Chondrocyte	Prevented chondrocyte death and cartilage erosion; enhanced proteoglycan synthesis and reduced MMP expression; reduced IL-1 $\beta$ and nitric oxide production from synovium; inflammation unaffected	124
	Joint	CIA model: reduced disease prevalence and paw swelling; attenuated synovitis and delayed disease onset	125
IL-4, IL-13	Fibroblast (xenograft)	Significantly reduced disease severity	126
ΙκΒα	Synovial fibroblast	Blocked IL-6, IL-8 and MMP production	127
	Chondrocyte	Blocked IL-6, IL-8 and MMP production	127
CTGF	Chondrocyte	Stimulated expression of aggrecan and collagen types II and X	63
IL-1RII	Keratinocyte graft⁵	CIA model: reduced clinical and histological parameters of disease; reduced IL-6 expression	111

<sup>a</sup> Although a specific cell or tissue is described in the studies, the virus is likely to primarily infect synovial cells. However, specific differences between studies might be a result of a more localised infection (e.g. cartilage versus synovium).

<sup>b</sup> This was a stably transfected line.

Abbreviations: BMP-2, bone morphogenic protein 2; CIA, collagen-induced arthritis; CTGF, connective tissue growth factor; IGF-1, insulin-like growth factor 1;  $I\kappa B\alpha$ , inhibitor of nuclear factor kappa B; IL-1, interleukin 1; IL-1RII, type II IL-1 receptor; IL-4, interleukin 4; IL-13, interleukin 13; iNOS, inducible nitric oxide synthase; MMP, matrix metalloproteinase; OSM, oncostatin M; RA, rheumatoid arthritis; TIMP, tissue inhibitor of metalloproteinase; TGF- $\beta$ , transforming growth factor  $\beta$ .

Unfortunately, these new treatments are expensive. The medical services could find themselves in the predicament of being able to offer a good chance of stopping disease progression in some patients (but not all), but probably at the expense of treating a much larger number of patients who would obtain some benefit from the more traditional treatments already available. This situation therefore suggests that there is still a need for smallmolecule drugs that prevent joint destruction. Financial constraints aside, researchers and clinicians alike continue to strive towards being able to offer improved treatment for these chronic and disabling diseases.

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# References

- 1 Muir, H. (1995) The chondrocyte, architect of cartilage. Biomechanics, structure, function and molecular biology of cartilage matrix macromolecules. Bioessays 17, 1039-1048, PubMed ID: 96244356
- 2 Temenoff, J.S. and Mikos, A.G. (2000) Review: tissue engineering for regeneration of articular cartilage. Biomaterials 21, 431-440, PubMed ID: 20137272
- 3 Buckwalter, J.A. and Mankin, H.J. (1997) Articular cartilage. Part I: Tissue design and chondrocyte–matrix interactions. J Bone Joint Surg 79A, 600-611
- 4 Ganu, V. et al. (1998) Inhibition of interleukin-1alpha-induced cartilage oligomeric matrix protein degradation in bovine articular cartilage by matrix metalloproteinase inhibitors: potential role for matrix metalloproteinases in the generation of cartilage oligomeric matrix protein fragments in arthritic synovial fluid. Arthritis Rheum 41, 2143-2151, PubMed ID: 99086418
- 5 Lotz, M., Hashimoto, S. and Kuhn, K. (1999) Mechanisms of chondrocyte apoptosis.

management of arthritis sufferers, not least financially.

During the past ten years, several synthetic MMP inhibitors have been developed for the treatment of arthritic disease (see Ref. 26). Enthusiasm was increased when the first fulllength crystal structure of a collagenase (MMP-1) was determined (Ref. 105) to assist such developments. However, despite considerable improvements in bioavailability and avoidance of gut modification, MMP inhibitors have generally not been very successful, because of lack of efficacy. Furthermore, adverse side effects of these inhibitors, presumably due to inappropriate inhibition of metalloproteinases other than those thought to be involved in the disease process, have also been reported. Metalloproteinases, including the MMPs, have many normal physiological roles and considerable debate has arisen as to whether a broad-spectrum inhibitor or a highly specific inhibitor should be used (Refs 26, 106). This is an important and fundamental issue and is biased by how data are interpreted. Some MMP inhibitors have been used with limited success in cancer treatment, but none has yet been used successfully to treat arthritic patients. Whether a specific enzyme responsible for tissue destruction in RA or OA will be identified is also a highly debated point, but is of considerable importance in minimising side effects.

One of the major landmarks in the treatment of RA has been anti-TNF- $\alpha$  therapies; in studies of such therapies ~70% of patients showed some benefit (Ref. 107). However, the remaining 30% of apparent non-responders presumably have a disease that is mediated by another proinflammatory agent, assumed by many to be IL-1. This is supported by data from a TNF- $\alpha$ overexpressing transgenic mouse model, where treatment with antibodies to IL-1RI completely prevented the otherwise spontaneous arthritis (Ref. 108). A combinational therapy with both anti-TNF- $\alpha$  and anti-IL-1 has therefore been proposed (Ref. 57). This is further supported by the finding that elevated levels of the natural antiinflammatory agent IL-1RA can occur in RA (Ref. 109). Furthermore, treatment with IL-1RA has been shown to be efficacious against arthritic joint disease (Ref. 110), and the decoy receptor IL-1RII blocks experimental arthritis (Ref. 111). These observations might therefore indicate the most promising avenues to pursue towards providing a markedly better prognosis for patients with

Osteoarthritis Cartilage 7, 389-391, PubMed ID: 99350802

- 6 Scott, D.L. et al. (1998) The clinical management of rheumatoid arthritis and osteoarthritis: strategies for improving clinical effectiveness. Br J Rheumatol 37, 546-554, PubMed ID: 98313104
- 7 Panayi, G.S. (1999) Targeting of cells involved in the pathogenesis of rheumatoid arthritis. Rheumatology (Oxford) 38 Suppl 2, 8-10, PubMed ID: 20111960
- 8 Nepom, G.T. and Nepom, B. (1998) Genetics of the major histocompatibility complex in rheumatoid arthritis. In Rheumatology (2nd edn) (Klippel, J.H. and Dieppe, P.A., eds), pp. 5.7.1-5.7.12, Mosby, London
- 9 Konttinen, Y.T. et al. (1998) New collagenolytic enzymes/cascade identified at the pannus-hard tissue junction in rheumatoid arthritis: destruction from above. Matrix Biol 17, 585-601, PubMed ID: 99120527
- 10 Miotla, J. et al. (2000) Treatment with soluble VEGF receptor reduces disease severity in murine collagen-induced arthritis. Lab Invest 80, 1195-1205, PubMed ID: 20405043
- 11 Paleolog, E. (1997) Target effector role of vascular endothelium in the inflammatory response: insights from the clinical trial of anti-TNF alpha antibody in rheumatoid arthritis. Mol Pathol 50, 225-233, PubMed ID: 98159346
- 12 van der Laan, W.H. et al. (2000) Cartilage degradation and invasion by rheumatoid synovial fibroblasts is inhibited by gene transfer of a cell surface-targeted plasmin inhibitor. Arthritis Rheum 43, 1710-1718, PubMed ID: 20397776
- 13 Dodge, G.R. and Poole, A.R. (1989) Immunohistochemical detection and immunochemical analysis of type II collagen degradation in human normal, rheumatoid, and osteoarthritic articular cartilages and in explants of bovine articular cartilage cultured with interleukin 1. J Clin Invest 83, 647-661, PubMed ID: 89109582
- 14 van den Berg, W.B. (1998) Joint inflammation and cartilage destruction may occur uncoupled. Springer Semin Immunopathol 20, 149-164, PubMed ID: 99053094
- 15 Villiger, P.M., Terkeltaub, R. and Lotz, M. (1992) Monocyte chemoattractant protein-1 (MCP-1) expression in human articular cartilage. Induction by peptide regulatory factors and

differential effects of dexamethasone and retinoic acid. J Clin Invest 90, 488-496, PubMed ID: 92355784

- 16 Sen, M. et al. (2000) Expression and function of wingless and frizzled homologs in rheumatoid arthritis. Proc Natl Acad Sci U S A 97, 2791-2796, PubMed ID: 20183967
- 17 Shlopov, B.V., Gumanovskaya, M.L. and Hasty, K.A. (2000) Autocrine regulation of collagenase 3 (matrix metalloproteinase 13) during osteoarthritis. Arthritis Rheum 43, 195-205, PubMed ID: 20106510
- 18 Goldring, M.B. (2000) Osteoarthritis and cartilage: the role of cytokines. Curr Rheumatol Rep 2, 459-465, PubMed ID: 21064118
- 19 Buttle, D.J., Bramwell, H. and Hollander, A.P. (1995) Proteolytic mechanisms of cartilage breakdown: a target for arthritis therapy. J Clin Pathol 48M, M167-M177
- 20 Woessner, J.F., Jr and Nagase, H. (2000) Matrix Metalloproteinases and TIMPs, Oxford University Press, Oxford, UK
- 21 Murphy, G. and Reynolds, J.J. (1985) Current views of collagen degradation. Progress towards understanding the resorption of connective tissues. BioEssays 2, 55-60
- 22 Kozaci, L.D., Buttle, D.J. and Hollander, A.P. (1997) Degradation of type II collagen, but not proteoglycan, correlates with matrix metalloproteinase activity in cartilage explant cultures. Arthritis Rheum 40, 164-174, PubMed ID: 97161355
- 23 Price, J.S. et al. (1999) Retinoic acid-induced type II collagen degradation does not correlate with matrix metalloproteinase activity in cartilage explant cultures. Arthritis Rheum 42, 137-147, PubMed ID: 99116752
- 24 Cawston, T. (1998) Matrix metalloproteinases and TIMPs: properties and implications for the rheumatic diseases. Mol Med Today 4, 130-137, PubMed ID: 98236409
- 25 Brew, K., Dinakarpandian, D. and Nagase, H. (2000) Tissue inhibitors of metalloproteinases: evolution, structure and function. Biochim Biophys Acta 1477, 267-283, PubMed ID: 20175276
- 26 Clark, I.M., Rowan, A.D. and Cawston, T.E. (2000) Matrix metalloproteinase inhibitors in the treatment of arthritis. Curr Opin Antiinflammatory Immunomodulatory Drugs 2, 16-25
- 27 McKie, N. et al. (1997) Expression of members of a novel membrane linked metalloproteinase

Accession information: (01)00320-9a.pdf (short code: txt001arn); 5 July 2001 ISSN 1462-3994 ©2001 Cambridge University Press

family (ADAM) in human articular chondrocytes. Biochem Biophys Res Commun 230, 335-339, PubMed ID: 97168971

- 28 Abbaszade, I. et al. (1999) Cloning and characterization of ADAMTS11, an aggrecanase from the ADAMTS family. J Biol Chem 274, 23443-23450, PubMed ID: 99367476
- 29 Tortorella, M.D. et al. (1999) Purification and cloning of aggrecanase-1: a member of the ADAMTS family of proteins. Science 284, 1664-1666, PubMed ID: 99286303
- 30 Kuno, K. et al. (2000) ADAMTS-1 cleaves a cartilage proteoglycan, aggrecan. FEBS Lett 478, 241-245, PubMed ID: 20389568
- 31 Vankemmelbeke, M.N. et al. (2001) Expression and activity of ADAMTS-5 in synovium. Eur J Biochem 268, 1259-1268, PubMed ID: 21153230
- 32 Kashiwagi, M. et al. (2001) Timp-3 is a potent inhibitor of aggrecanase 1 (adam-ts4) and aggrecanase 2 (adam-ts5). J Biol Chem 276, 12501-12504, PubMed ID: 21201070
- 33 Keyszer, G. et al. (1998) Differential expression of cathepsins B and L compared with matrix metalloproteinases and their respective inhibitors in rheumatoid arthritis and osteoarthritis: a parallel investigation by semiquantitative reverse transcriptase-polymerase chain reaction and immunohistochemistry. Arthritis Rheum 41, 1378-1387, PubMed ID: 98368333
- 34 Creemers, L.B. et al. (1998) Participation of intracellular cysteine proteinases, in particular cathepsin B, in degradation of collagen in periosteal tissue explants. Matrix Biol 16, 575-584, PubMed ID: 98228081
- 35 Kafienah, W. et al. (1998) Human cathepsin K cleaves native type I and II collagens at the Nterminal end of the triple helix. Biochem J 331, 727-732, PubMed ID: 98228208
- 36 Saftig, P. et al. (2000) Functions of cathepsin K in bone resorption. Lessons from cathepsin K deficient mice. Adv Exp Med Biol 477, 293-303, PubMed ID: 20308303
- 37 Bryson, H. et al. (1998) A serine proteinase inactivator inhibits chondrocyte-mediated cartilage proteoglycan breakdown occurring in response to proinflammatory cytokines. Arch Biochem Biophys 355, 15-25, PubMed ID: 98313240
- 38 Everts, V. et al. (1996) Phagocytosis and intracellular digestion of collagen, its role in turnover and remodelling. Histochem J 28, 229-245, PubMed ID: 96358050

- 39 Culty, M., Nguyen, H.A. and Underhill, C.B. (1992) The hyaluronan receptor (CD44) participates in the uptake and degradation of hyaluronan. J Cell Biol 116, 1055-1062, PubMed ID: 92129427
- 40 Yaeger, P.C. et al. (1997) Synergistic action of transforming growth factor-beta and insulin-like growth factor-I induces expression of type II collagen and aggrecan genes in adult human articular chondrocytes. Exp Cell Res 237, 318-325, PubMed ID: 98096240
- 41 Fawthrop, F.W. et al. (1997) Effects of transforming growth factor beta on the production of prostaglandin E and caseinase activity of unstimulated and interleukin 1stimulated human articular chondrocytes in culture. Br J Rheumatol 36, 729-734, PubMed ID: 97398976
- 42 Ohmori, Y., Smith, M.F., Jr. and Hamilton, T.A. (1996) IL-4-induced expression of the IL-1 receptor antagonist gene is mediated by STAT6. J Immunol 157, 2058-2065, PubMed ID: 96355010
- 43 Chou, H.H. et al. (2000) Induction of intracellular interleukin-1 beta signals via type II interleukin-1 receptor in human gingival fibroblasts. J Dent Res 79, 1683-1688, PubMed ID: 20476115
- 44 Goldring, M.B. (1999) The role of cytokines as inflammatory mediators in osteoarthritis: lessons from animal models. Connect Tissue Res 40, 1-11, PubMed ID: 20231415
- 45 van den Berg, W.B. (2000) Arguments for interleukin 1 as a target in chronic arthritis. Ann Rheum Dis 59 Suppl 1, i81-84, PubMed ID: 20507133
- 46 McInnes, I.B. and Liew, F.Y. (1998) Interleukin 15: a proinflammatory role in rheumatoid arthritis synovitis. Immunol Today 19, 75-79, PubMed ID: 98170502
- 47 Jovanovic, D.V. et al. (1998) IL-17 stimulates the production and expression of proinflammatory cytokines, IL-beta and TNF-alpha, by human macrophages. J Immunol 160, 3513-3521, PubMed ID: 98189800
- 48 Leistad, L., Ostensen, M. and Faxvaag, A. (1998) Detection of cytokine mRNA in human, articular cartilage from patients with rheumatoid arthritis and osteoarthritis by reverse transcriptase- polymerase chain reaction. Scand J Rheumatol 27, 61-67, PubMed ID: 98165726
- 49 Studer, R.K. et al. (1999) Inhibition of
- Accession information: (01)00320-9a.pdf (short code: txt001arn); 5 July 2001 ISSN 1462-3994 ©2001 Cambridge University Press

transforming growth factor beta production by nitric oxide-treated chondrocytes: implications for matrix synthesis. Arthritis Rheum 42, 248-257, PubMed ID: 99148785

- 50 Olee, T. et al. (1999) IL-18 is produced by articular chondrocytes and induces proinflammatory and catabolic responses. J Immunol 162, 1096-1100, PubMed ID: 99113776
- 51 Cawston, T.E. et al. (1998) The role of oncostatin M in animal and human connective tissue collagen turnover and its localization within the rheumatoid joint. Arthritis Rheum 41, 1760-1771, PubMed ID: 98449666
- 52 Caterson, B. et al. (2000) Mechanisms involved in cartilage proteoglycan catabolism. Matrix Biol 19, 333-344, PubMed ID: 20420447
- 53 Isomaki, P. and Punnonen, J. (1997) Pro- and anti-inflammatory cytokines in rheumatoid arthritis. Ann Med 29, 499-507, PubMed ID: 98221008
- 54 Rowan, A.D. et al. (2001) Synergistic effects of gp130 binding cytokines in combination with interleukin-1 on cartilage collagen breakdown. Arthritis Rheum 44, 1620-1632, PubMed ID: 21358266
- 55 Henrotin, Y.E. et al. (1996) Effects of exogenous IL-1 beta, TNF alpha, IL-6, IL-8 and LIF on cytokine production by human articular chondrocytes. Osteoarthritis Cartilage 4, 163-173, PubMed ID: 97050489
- 56 Pulsatelli, L. et al. (1999) Chemokine production by human chondrocytes. J Rheumatol 26, 1992-2001, PubMed ID: 99421425
- 57 van den Berg, W.B., Joosten, L.A. and van de Loo, F.A. (1999) TNF alpha and IL-1 beta are separate targets in chronic arthritis. Clin Exp Rheumatol 17, S105-114, PubMed ID: 20056793
- 58 Lotz, M. (1999) The role of nitric oxide in articular cartilage damage. Rheum Dis Clin North Am 25, 269-282, PubMed ID: 99284905
- 59 Shuler, F.D. et al. (2000) Increased matrix synthesis following adenoviral transfer of a transforming growth factor beta1 gene into articular chondrocytes. J Orthop Res 18, 585-592, PubMed ID: 20505680
- 60 Goto, H. et al. (2000) Gene therapy for meniscal injury: enhanced synthesis of proteoglycan and collagen by meniscal cells transduced with a TGFbeta(1)gene. Osteoarthritis Cartilage 8, 266-271, PubMed ID: 20366445
- 61 Nixon, A.J. et al. (2000) Insulinlike growth

factor-I gene therapy applications for cartilage repair. Clin Orthop S201-213, PubMed ID: 20492914

- 62 Smith, P. et al. (2000) Genetic enhancement of matrix synthesis by articular chondrocytes: comparison of different growth factor genes in the presence and absence of interleukin-1. Arthritis Rheum 43, 1156-1164, PubMed ID: 20275271
- 63 Nakanishi, T. et al. (2000) Effects of CTGF/ Hcs24, a product of a hypertrophic chondrocytespecific gene, on the proliferation and differentiation of chondrocytes in culture. Endocrinology 141, 264-273, PubMed ID: 20080284
- 64 Fischer, D.C. et al. (1999) Induction of alpha1antitrypsin synthesis in human articular chondrocytes by interleukin-6-type cytokines: evidence for a local acute-phase response in the joint. Arthritis Rheum 42, 1936-1945, PubMed ID: 99441968
- 65 Li, W.Q. and Zafarullah, M. (1998) Oncostatin M up-regulates tissue inhibitor of metalloproteinases-3 gene expression in articular chondrocytes via de novo transcription, protein synthesis, and tyrosine kinase-and mitogen-activated protein kinase-dependent mechanisms. J Immunol 161, 5000-5007, PubMed ID: 99008568
- 66 Hui, W., Rowan, A.D. and Cawston, T. (2001) Insulin-like growth factor 1 blocks collagen release and down regulates matrix metalloproteinase-1, -3, -8, and -13 mRNA expression in bovine nasal cartilage stimulated with oncostatin M in combination with interleukin 1alpha. Ann Rheum Dis 60, 254-261, PubMed ID: 21091711
- 67 Smeets, T.J. et al. (2000) The effects of interferon-beta treatment of synovial inflammation and expression of metalloproteinases in patients with rheumatoid arthritis. Arthritis Rheum 43, 270-274, PubMed ID: 20155532
- 68 Morales, T.I. (1994) Transforming growth factor-beta and insulin-like growth factor-1 restore proteoglycan metabolism of bovine articular cartilage after depletion by retinoic acid. Arch Biochem Biophys 315, 190-198, PubMed ID: 95070163
- 69 Bakker, A.C. et al. (2001) Overexpression of active TGF-beta-1 in the murine knee joint: evidence for synovial-layer-dependent chondroosteophyte formation. Osteoarthritis Cartilage 9,

Accession information: (01)00320-9a.pdf (short code: txt001arn); 5 July 2001 ISSN 1462-3994 ©2001 Cambridge University Press

in molecular medic

128-136, PubMed ID: 21138040

- 70 Martin, J.A. and Buckwalter, J.A. (2000) The role of chondrocyte-matrix interactions in maintaining and repairing articular cartilage. Biorheology 37, 129-140, PubMed ID: 20369886
- 71 Langdon, C. et al. (2000) Murine oncostatin M stimulates mouse synovial fibroblasts in vitro and induces inflammation and destruction in mouse joints in vivo. Am J Pathol 157, 1187-1196, PubMed ID: 20476205
- 72 Cleaver, C.S., Rowan, A.D. and Cawston, T.E. (2001) Interleukin 13 blocks the release of collagen from bovine nasal cartilage treated with proinflammatory cytokines. Ann Rheum Dis 60, 150-157, PubMed ID: 21068302
- 73 Cawston, T.E. et al. (1996) Interleukin-4 blocks the release of collagen fragments from bovine nasal cartilage treated with cytokines. Biochim Biophys Acta 1314, 226-232, PubMed ID: 97136899
- 74 Loeser, R.F. (2000) Chondrocyte integrin expression and function. Biorheology 37, 109-116, PubMed ID: 20369884
- 75 Jalali, S. et al. (2001) Integrin-mediated mechanotransduction requires its dynamic interaction with specific extracellular matrix (ECM) ligands. Proc Natl Acad Sci U S A 98, 1042-1046, PubMed ID: 21107667
- 76 Attur, M.G. et al. (2000) Functional genomic analysis in arthritis-affected cartilage: yinyang regulation of inflammatory mediators by alpha 5 beta 1 and alpha V beta 3 integrins. J Immunol 164, 2684-2691, PubMed ID: 20143864
- 77 Badger, A.M. et al. (2001) Disease-modifying activity of SB 273005, an orally active, nonpeptide alphavbeta3 (vitronectin receptor) antagonist, in rat adjuvant-induced arthritis. Arthritis Rheum 44, 128-137, PubMed ID: 21079685
- 78 Martel-Pelletier, J. et al. (1992) The interleukin-1 receptor in normal and osteoarthritic human articular chondrocytes. Identification as the type I receptor and analysis of binding kinetics and biologic function. Arthritis Rheum 35, 530-540, PubMed ID: 92247049
- 79 Alsalameh, S. et al. (1999) Preferential expression of tumor necrosis factor receptor 55 (TNF-R55) on human articular chondrocytes: selective transcriptional upregulation of TNF-R75 by proinflammatory cytokines interleukin 1beta, tumor necrosis factor-alpha, and basis fibroblast growth factor. J Rheumatol 26, 645-653, PubMed ID: 99188563

- 80 Westacott, C.I. et al. (2000) Tumor necrosis factor alpha can contribute to focal loss of cartilage in osteoarthritis. Osteoarthritis Cartilage 8, 213-221, PubMed ID: 20268130
- 81 Amin, A.R. (1999) Regulation of tumor necrosis factor-alpha and tumor necrosis factor converting enzyme in human osteoarthritis. Osteoarthritis Cartilage 7, 392-394, PubMed ID: 99350803
- 82 Chow, G. et al. (1998) Antisense inhibition of chondrocyte CD44 expression leading to cartilage chondrolysis. Arthritis Rheum 41, 1411-1419, PubMed ID: 98368337
- 83 Silacci, P. et al. (1998) Interleukin (IL)-6 and its soluble receptor induce TIMP-1 expression in synoviocytes and chondrocytes, and block IL-1induced collagenolytic activity. J Biol Chem 273, 13625-13629, PubMed ID: 98256284
- 84 Robak, T. et al. (1998) Serum levels of interleukin-6 type cytokines and soluble interleukin-6 receptor in patients with rheumatoid arthritis. Mediators Inflamm 7, 347-353, PubMed ID: 99098439
- 85 MacGregor, A.J. and Silman, A.J. (1998) Classification and epidemiology in rheumatoid arthritis. In Rheumatology (2nd edn) (Klippel, J.H. and Dieppe, P.A., eds), pp. 5.2.1-5.2.6, Mosby, London
- 86 Woo, P. (2000) Cytokine polymorphisms and inflammation. Clin Exp Rheumatol 18, 767-771, PubMed ID: 21020348
- 87 Clague, R.B. et al. (1983) Serum antibodies to type II collagen in rheumatoid arthritis: comparison of 6 immunological methods and clinical features. Ann Rheum Dis 42, 537-544, PubMed ID: 84022742
- 88 Nakagawa, K. et al. (1999) Complement C1s activation in degenerating articular cartilage of rheumatoid arthritis patients: immunohistochemical studies with an active form specific antibody. Ann Rheum Dis 58, 175-181, PubMed ID: 99293250
- 89 Yamaguchi, K. et al. (1990) Degradation of type I and II collagen by human activated C1-s. FEBS Lett 268, 206-208, PubMed ID: 90346138
- 90 Firestein, G.S. and Manning, A.M. (1999) Signal transduction and transcription factors in rheumatic disease. Arthritis Rheum 42, 609-621, PubMed ID: 99226856
- 91 Catterall, J.B. et al. Synergistic induction of matrix metalloproteinase-1 by interleukin-1alpha and oncostatin M in human chondrocytes involves signal transducer and activator of

prphisms and atol 18, 767-771, antibodies to arthritis: 1 methods and is 42, 537-544, uplement C1s ular cartilage of with an active eum Dis 58, 175radation of type I ated C1-s. FEBS 00346138 M. (1999) Signal factors in eum 42, 609-621, nduction of nterleukin-1alpha ondrocytes activator of arn); 5 July 2001

Accession information: (01)00320-9a.pdf (short code: txt001arn); 5 July 2001 ISSN 1462-3994 ©2001 Cambridge University Press

in molecular medicir

transcription and activator of protein-1 transcription factors via a novel mechanism. Arthritis Rheum 44 (in press)

- 92 Clark, I.M. et al. (1997) Transcriptional activity of the human tissue inhibitor of metalloproteinases 1 (TIMP-1) gene in fibroblasts involves elements in the promoter, exon 1 and intron 1. Biochem J 324, 611-617, PubMed ID: 97307877
- 93 Starr, R. et al. (1997) A family of cytokineinducible inhibitors of signalling. Nature 387, 917-921, PubMed ID: 97345633
- 94 Freije, J.M. et al. (1994) Molecular cloning and expression of collagenase-3, a novel human matrix metalloproteinase produced by breast carcinomas. J Biol Chem 269, 16766-16773, PubMed ID: 94266894
- 95 Balbin, M. et al. (2001) Identification and enzymatic characterization of two diverging murine counterparts of human interstitial collagenase (MMP-1) expressed at sites of embryo implantation. J Biol Chem 276, 10253-10262, PubMed ID: 21167837
- 96 Konttinen, Y.T. et al. (1999) Analysis of 16 different matrix metalloproteinases (MMP-1 to MMP-20) in the synovial membrane: different profiles in trauma and rheumatoid arthritis. Ann Rheum Dis 58, 691-697, PubMed ID: 20001911
- 97 van Lent, P.L. et al. (2000) Role of Fc receptor gamma chain in inflammation and cartilage damage during experimental antigen-induced arthritis. Arthritis Rheum 43, 740-752, PubMed ID: 20226985
- 98 Mudgett, J.S. et al. (1998) Susceptibility of stromelysin 1-deficient mice to collageninduced arthritis and cartilage destruction. Arthritis Rheum 41, 110-121, PubMed ID: 98093965
- 99 Vu, T.H. et al. (1998) MMP-9/gelatinase B is a key regulator of growth plate angiogenesis and apoptosis of hypertrophic chondrocytes. Cell 93, 411-422, PubMed ID: 98250057
- 100 Holmbeck, K. et al. (1999) MT1-MMP-deficient mice develop dwarfism, osteopenia, arthritis, and connective tissue disease due to inadequate collagen turnover. Cell 99, 81-92, PubMed ID: 99449306
- 101 Zhou, Z. et al. (2000) Impaired endochondral ossification and angiogenesis in mice deficient in membrane-type matrix metalloproteinase I. Proc Natl Acad Sci U S A 97, 4052-4057, PubMed ID: 20226056

- 102 Zhao, J. et al. (2001) Pulmonary hypoplasia in mice lacking tumor necrosis factor-alpha converting enzyme indicates an indispensable role for cell surface protein shedding during embryonic lung branching morphogenesis. Dev Biol 232, 204-218, PubMed ID: 21153176
- 103 Shindo, T. et al. (2000) ADAMTS-1: a metalloproteinase-disintegrin essential for normal growth, fertility, and organ morphology and function. J Clin Invest 105, 1345-1352, PubMed ID: 20273926
- 104 Goater, J. et al. (2000) Empirical advantages of adeno associated viral vectors in vivo gene therapy for arthritis. J Rheumatol 27, 983-989, PubMed ID: 20242921
- 105 Li, J. et al. (1995) Structure of full-length porcine synovial collagenase reveals a C-terminal domain containing a calcium-linked, four-bladed beta-propeller. Structure 3, 541-549, PubMed ID: 96173003
- 106 Cawston, T.E. and Rowan, A. (1998) Prevention of cartilage breakdown by matrix metalloproteinase inhibition—a realistic therapeutic target? Br J Rheumatol 37, 353-356, PubMed ID: 98281180
- 107 Brennan, F.M. et al. (1997) Reduction of serum matrix metalloproteinase 1 and matrix metalloproteinase 3 in rheumatoid arthritis patients following anti- tumour necrosis factoralpha (cA2) therapy. Br J Rheumatol 36, 643-650, PubMed ID: 97379874
- 108 Probert, L. et al. (1995) The type I interleukin-1 receptor acts in series with tumor necrosis factor (TNF) to induce arthritis in TNF-transgenic mice. Eur J Immunol 25, 1794-1797, PubMed ID: 95339901
- 109 Jouvenne, P. et al. (1998) Elevated levels of soluble interleukin-1 receptor type II and interleukin-1 receptor antagonist in patients with chronic arthritis: correlations with markers of inflammation and joint destruction. Arthritis Rheum 41, 1083-1089, PubMed ID: 98288707
- 110 Bresnihan, B. et al. (1998) Treatment of rheumatoid arthritis with recombinant human interleukin-1 receptor antagonist. Arthritis Rheum 41, 2196-2204, PubMed ID: 99086423
- 111 Bessis, N. et al. (2000) The type II decoy receptor of IL-1 inhibits murine collagen-induced arthritis. Eur J Immunol 30, 867-875, PubMed ID: 20203444
- 112 Niki, Y. et al. (1998) Membrane-associated IL-1 contributes to chronic synovitis in human IL-1RA

17

18

# expert reviews

transgenic mice. Arthritis Rheum 41, S212

- 113 Zheng, H. et al. (1995) Resistance to fever induction and impaired acute-phase response in interleukin-1 beta-deficient mice. Immunity 3, 9-19, PubMed ID: 95346514
- 114 van den Berg, W.B. (1997) Lessons for joint destruction from animal models. Curr Opin Rheumatol 9, 221-228, PubMed ID: 97348206
- 115 Mori, L. et al. (1996) Attenuation of collageninduced arthritis in 55-kDa TNF receptor type 1 (TNFR1)-IgG1-treated and TNFR1-deficient mice. J Immunol 157, 3178-3182, PubMed ID: 96413269
- 116 van de Loo, F.A. et al. (1997) Interleukin-6 reduces cartilage destruction during experimental arthritis. A study in interleukin-6deficient mice. Am J Pathol 151, 177-191, PubMed ID: 97356308
- 117 de Hooge, A.S. et al. (2000) Involvement of IL-6, apart from its role in immunity, in mediating a chronic response during experimental arthritis. Am J Pathol 157, 2081-2091, PubMed ID: 20560568
- 118 Chan, S.Y. and Wong, R.W. (2000) Expression of epidermal growth factor in transgenic mice causes growth retardation. J Biol Chem 275, 38693-38698, PubMed ID: 20556317
- 119 Manoury-Schwartz, B. et al. (1997) High susceptibility to collagen-induced arthritis in mice lacking IFN- gamma receptors. J Immunol 158, 5501-5506, PubMed ID: 97307631
- 120 Blom, A.B. et al. (2000) Fc gamma R expression on macrophages is related to severity and chronicity of synovial inflammation and cartilage destruction during experimental immune-complex-mediated arthritis (ICA). Arthritis Res 2, 489-503, PubMed ID: 21062458

- 121 Tsuji, M. et al. (2000) The possible role of c-fos expression in rheumatoid cartilage destruction. J Rheumatol 27, 1606-1621, PubMed ID: 20370525
- 122 Mi, Z. et al. (2000) Adenovirus-mediated gene transfer of insulin-like growth factor 1 stimulates proteoglycan synthesis in rabbit joints. Arthritis Rheum 43, 2563-2570, PubMed ID: 20534252
- 123 Studer, R.K. et al. (2000) Nitric oxide inhibits chondrocyte response to IGF-I: inhibition of IGF- IRbeta tyrosine phosphorylation. Am J Physiol Cell Physiol 279, C961-969, PubMed ID: 20459423
- 124 Lubberts, E. et al. (1999) Adenoviral vectormediated overexpression of IL-4 in the knee joint of mice with collagen-induced arthritis prevents cartilage destruction. J Immunol 163, 4546-4556, PubMed ID: 99441379
- 125 Cottard, V. et al. (2000) Adeno-associated virusmediated delivery of IL-4 prevents collageninduced arthritis. Gene Ther 7, 1930-1939, PubMed ID: 21010713
- 126 Bessis, N. et al. (1999) Encapsulation in hollow fibres of xenogeneic cells engineered to secrete IL-4 or IL-13 ameliorates murine collageninduced arthritis (CIA). Clin Exp Immunol 117, 376-382, PubMed ID: 99373644
- 127 Bondeson, J. et al. (2000) Effective adenoviral transfer of IkappaBalpha into human fibroblasts and chondrosarcoma cells reveals that the induction of matrix metalloproteinases and proinflammatory cytokines is nuclear factorkappaB dependent. J Rheumatol 27, 2078-2089, PubMed ID: 20443624
- 128 Rawlings, N.D. and Barrett, A.J. (2000) MEROPS: the peptidase database. Nucleic Acids Res 28, 323-325, PubMed ID: 20063295

Further reading, resources and contacts	
Newcastle Rheumatology: information about the research projects in rheumatology in Newcastle, UK	
http://www.ncl.ac.uk/rheumatology	
Arthritis Research Campaign: the major funding organisation for rheumatological research in the UK	
http://www.arc.org.uk	
Arthritis & Rheumatism: the highest-rated, peer-reviewed rheumatological journal	
http://www.interscience.wiley.com/jpages/0004-3591/	
Arthritis Research: a relatively new research journal available online	
http://www.arthritis-research.com	
Cytokine and Growth Factor Reviews: publishes excellent articles, available on the web	
http://www.elsevier.com/locate/cytogfr	
General sites about arthritis and various treatment regimens	
http://www.arthritis.about.com/health/arthritis http://www.healthtalk.com http://www.arthritisinsight.com	
The MEROPS database (Ref. 128) has helped to reform the nomenclature of proteinases. Many have more than one trivial name; in addition, some cannot be conveniently separated by their catalytic activity and are thus not neatly covered by the International Union of Biochemistry and Molecular Biology (IUBMB) enzyme nomenclature system. It is now strongly recommended that researchers use the MEROPS identifier when discussing a particular proteinase to avoid possible confusion. This database is constantly being updated as new proteinases are described.	eteesened consultation to dt castering alta dtae alta and to
http://www.merops.co.uk	
An excellent and easy-to-read rheumatological textbook for medical students:	
Athanasou, N.A. (1999) Colour Atlas of Bone, Joint and Soft Tissue Pathology, Oxford University Press, Oxford, UK.	
A useful overview of the MAPKs that are utilised by many pro-inflammatory cytokines:	
Chang, L. and Karin, M. (2001) Mammalian MAP kinase signalling cascades. Nature 410, 37-40, PubMed ID: 11242034	
The following papers/reviews provide an excellent reference point for:	
Bone loss in rheumatoid disease	
Lane, N.E. and Goldring, S.R. (1998) Bone loss in rheumatoid arthritis: what role does inflammation play? J Rheumatol 25, 1251-1253, PubMed ID: 98339373	0.000
Gravallese, E.M. and Goldring, S.R. (2000) Cellular mechanisms and the role of cytokines in bone erosions in rheumatoid arthritis. Arthritis Rheum 43, 2143-2151, PubMed ID: 20489573	C
Catabolic events in rheumatoid disease	
Birkedal-Hansen, H. (1995) Proteolytic remodeling of extracellular matrix. Curr Opin Cell Biol 7, 728-735,	
PubMed ID: 96107563 (Continued on next page)	
	   1
Accession information: (01)00320-9a.pdf (short code: txt001arn); 5 July 2001	

ISSN 1462-3994 ©2001 Cambridge University Press

nolecular medicine

- Henrotin, Y. and Reginster, J.Y. (1999) Anabolic events in osteoarthritis. Osteoarthritis Cartilage 7, 310-312, PubMed ID: 99309166
- Goldring, M.B. (2000) The role of the chondrocyte in osteoarthritis. Arthritis Rheum 43, 1916-1926, PubMed ID: 20466269
- Firestein, G.S, Panayi, G.S. and Wollheim, F.A. (2000) Rheumatoid Arthritis: New Frontiers in Pathogenesis and Treatment, Oxford University Press, Oxford, UK
- Tak, P.P. and Bresnihan, B. (2000) The pathogenesis and prevention of joint damage in rheumatoid arthritis: advances from synovial biopsy and tissue analysis. Arthritis Rheum 43, 2619-2633, PubMed ID: 21017429
- Muller-Ladner, U. and Nishioka, K. (2000) p53 in rheumatoid arthritis: friend or foe? Arthritis Res 2, 175-178, PubMed ID: 21062428
- Tak, P.P. and Firestein, G.S. (2001) NF-kappaB: a key role in inflammatory diseases. J Clin Invest 107, 7-11, PubMed ID: 20576565
- Evans, C.H. et al. (2001) Future of adenoviruses in the gene therapy of arthritis. Arthritis Res 3, 142-146, PubMed ID: 21196423
- · Lessons from animal models of arthritic disease
- Goldring, M.B. (1999) The role of cytokines as inflammatory mediators in osteoarthritis: lessons from animal models. Connect Tissue Res 40, 1-11, PubMed ID: 20231415
- van den Berg, W.B. (1997) Lessons for joint destruction from animal models. Curr Opin Rheumatol 9, 221-228, PubMed ID: 97348206
- The aggrecanases

Kaushal, G.P. and Shah, S.V. (2000) The new kids on the block: ADAMTSs, potentially multifunctional metalloproteinases of the ADAM family. J Clin Invest 105, 1335-1337, PubMed ID: 20273923

Tang, B.L. (2001) ADAMTS: a novel family of extracellular matrix proteases. Int J Biochem Cell Biol 33, 33-44, PubMed ID: 21109804

# Features associated with this article

#### Figure

Figure 1. The control of degradation of cartilage extracellular matrix (fig001arn).

#### Tables

Table 1. Effects of cytokine transgenes on experimental arthritis (tab001arn).

Table 2. Effects of adenovirus-transfected mediators on experimental arthritis (tab002arn).

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